Electronic Supplementary Information

Diversity-Oriented Synthesis of Peptide-Boronic Acids by a Versatile Building-Block Approach

Stefan P. A. Hinkes, Severin Kämmerer and Christian D. P. Klein* Institute of Pharmacy and Molecular Biotechnology (IPMB), Medicinal Chemistry, Heidelberg University, Im Neuenheimer Feld 364, 69120 Heidelberg.

Table of contents

General information	S3
Synthesis of <i>N</i> -hydroxyphthalimide (NHPI) esters	S6
General procedure	S6
<i>N</i> -hydroxyphthalimide (NHPI) esters	S6
Synthesis of α-aminopinacolyl boronates	S20
General procedure	S20
α-Aminopinacolyl boronates	S21
Synthesis of α-aminoboronic acids	
Conditions A: MeCN/0.1 N HCl (1:1, v/v)	
Conditions B: MeCN/0.1 N HCl (9:1, v/v)	
Conditions C: MeCN/phosphate buffer pH 7 (1:1, v/v)	
Determination of loading efficiency onto 1-glycerol polystyrene resin	S34
α-Aminoboronic acids	S34
Solid-phase synthesis of peptide-boronic acids	S47
General procedure	S47
Peptide-boronic acids	S48
NMR spectra of NHPI esters	
NMR spectra of α-aminopinacolyl boronates	S106
NMR spectra of α-aminoboronic acids	S147
NMR spectra of peptide-boronic acids	S188
References	S211

General information

Reagent	Indicated Purity	Supplier
Acetonitrile HPLC grade (ACN)	≥ 99.9%	VWR Chemicals
1-[Bis(dimethylamino)methylene]-1 <i>H</i> -1,2,3- triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate (HATU)	99%	Carbolution
Bis(pinacolato)diboron (B 2 pin 2)	99%	Carbolution
(1-Cyano-2-ethoxy-2-oxoethylidenaminooxy)- dimethylamino-morpholino-carbenium hexafluorophosphate (COMU)	98%	Carbolution
Cyclohexane	≥ 99.5%	Honeywell
<i>N,N</i> '-Dicyclohexylcarbodiimide (DCC)	99%	Merck/Aldrich
Diethanolamine (DEA)	98%	Merck/Aldrich
Diethylether (DEE), stabilised with BHT	≥ 99.8%	Merck/Aldrich
<i>N</i> , <i>N</i> '-Diisopropylcarbodiimide (DIC)	98%	Carbolution
4-(Dimethylamino)pyridine (DMAP)	≥99%	Merck/Aldrich
<i>N,N</i> -Dimethylformamide (DMF)	≥ 99.8%	Honeywell
<i>N</i> -Ethyldiisopropylamine (DIPEA)	99%	Alfa Aesar
Hydrochloric acid (1 N)		Honeywell
<i>N</i> -Hydroxyphthalimide (NHPI)	≥99%	TCI
Magnesium bromide diethyl etherate	99%	Merck/Aldrich
Methanol HPLC Grade	≥ 99.8%	Fisher Scientific
Methylboronic acid	98%	Carbolution
Methylene chloride (DCM)	≥ 99.9%	Merck/Aldrich
Nickel-(II)-chloride hexahydrate	≥ 98%	Honeywell
Piperidine	99%	Merck/Aldrich
Phosphate buffer pH 7.0		Honeywell
Tetrahydrofuran anhydrous (THF), stabilised with BHT	≥99.5%	TCI
Trifluoroacetic acid (TFA)	99%	Alfa Aesar
Triisopropylsilane (TIPS)	≥98%	TCI
2,4,6-Trimethylpyridine (TMP)	99%	Merck/Aldrich

The following reagents and solvents were used:

Fmoc-protected amino acids were purchased from Carbolution, Merck/Aldrich, TCI, Alfa Aesar and Orpegen Peptide Chemicals and were used without further purification. Reactions were performed in ambient atmosphere with oven-dried glassware unless otherwise stated. Anhydrous solvents were purchased as such and stored under inert gas and molecular sieves. Ethyl acetate and deionised water were distilled prior to use.

For solid-phase synthesis, polypropylene reaction vessels equipped with polyethylene frits (Carl Roth) were used. Reaction vessels were continuously shaken at ambient temperature using an IKA "KS 130 basic" orbital shaker. 1-Glycerol polystyrene resin (loading capacity: 0.60 mmol/g) was purchased from Iris Biotech.

Reported yields of boronic acids are calculated based on the free acid form without consideration of anhydride formation unless otherwise stated. Depending on the product's tendency to form anhydrides, e.g. boroxines or intramolecular cyclisation products, actual yields are expected to be slightly higher.

Flash column chromatography was done using an "Isolera One" system (Biotage) with silica gel (40 – 63 μ m, 60 Å, Material Harvest). Purifications were usually achieved by gradient elution with binary mixtures of cyclohexane and ethyl acetate and UV detection at 254 nm and 280 nm.

MS-ESI analysis was performed on a Bruker "micrOTOF-Q II" mass spectrometer. For highresolution mass spectra, sodium formate was used as calibrating reagent. In some cases, a solution of diethanolamine in acetonitrile (10 μ L, c = 10 mM) was added to the MS samples of boronic acids to facilitate the formation of cationic species and therefore the detection in positive mode.

NMR spectra were recorded on 300 MHz or 500 MHz Varian instruments. Samples were dissolved in CDCl₃, acetone- d_6 , DMSO- d_6 , CD₃CN, CD₃OD and D₂O or mixtures thereof. To reverse anhydride formation that impedes NMR analysis of boronic acids, small amounts of D₂O were usually added to the samples of boronic acids. Chemical shifts are reported in ppm relative to TMS using the solvent residual peaks of the deuterated species as an internal standard. Solvent residual shifts for ¹H and ¹³C NMR nuclei were taken from the literature (CDCl₃: 7.26 ppm and 77.16 ppm; acetone- d_6 : 2.05 ppm and 29.84 ppm; DMSO- d_6 : 2.50 ppm and 39.50 ppm; CD₃OD: 3.31 ppm and 49.00 ppm; CD₃CN: 1.94 ppm and 1.32 ppm).¹ Coupling constants are given in Hertz (Hz). Multiplicity is reported as observed, with the following abbreviations used: s (singlet), d (doublet), t (triplet), q (quadruplet), dd (doublet of doublet), dt (doublet of triplet), td (triplet of doublet), m (multiplet). Two-dimensional NMR

experiments (COSY, HSQC, HMBC) were performed in some cases to facilitate assignments, e.g. of diastereomeric mixtures. In proton NMR, acidic protons (e.g. Fmoc-N*H*) were not observed after addition of D₂O due to solvent exchange processes. ¹³C NMR were measured as "attached proton test" (APT) experiments for facilitated assignments. The carbon attached to boron was not observed due to the quadrupolar relaxation of ¹⁰B and ¹¹B nuclei. ¹¹B spectra were measured for representative compounds at 160 MHz using BF₃·OEt₂ as an external standard. Signals of boron nuclei in pinacolyl boronates and free boronic acids were found in the range of 25-35 ppm, regardless of their substitution pattern. Therefore, the respective spectra are not shown.

Final peptides were purified by RP-HPLC, if indicated, using an "ÄKTApurifier" (GE Healthcare, Germany) with a precolumn (Reprospher 100 C18-DE, 5 μ m, 30×16 mm, Dr. Maisch, Germany) and a main column (Reprospher 100 C18-DE, 5 μ m, 125×16 mm, Dr. Maisch, Germany), Purification was achieved using gradient elution (solvent A: H₂O + 0.1% TFA; solvent B: ACN + 0.1% TFA or MeOH + 0.1% TFA) with UV detection at 214, 254 and 280 nm. Product-containing fractions were lyophilised with an "Alpha 1-2 LDplus" freeze-drier (Martin Christ, Germany).

Synthesis of N-hydroxyphthalimide (NHPI) esters



General procedure

NHPI esters were prepared by a literature-known method with slight modifications.¹

A round-bottom flask was charged with the respective Fmoc-protected amino acid (1, 1.0 eq), N-hydroxyphthalimide (NHPI, 1.0 eq) and N,N-dimethylaminopyridine (DMAP, 0.1 eq) were added. After the addition of methylene chloride (~ 5 mL/mmol), the mixture was cooled to 0°C and N,N'-diisopropylcarbodiimide (DIC, 1.1 eq) was added dropwise. The reaction was stirred for 1 h at 0°C, allowed to warm up to ambient temperature and stirred for 2-16 hours. The resulting precipitate was filtered and washed with additional methylene chloride. The filtrate was concentrated *in vacuo* and purified by flash chromatography using silica and cyclohexane/ethyl acetate gradients.

NOTE: Nucleophilic solvents, e. g. alcohols, should be avoided for NMR and MS analysis due to the enhanced reactivity of the activated esters. MS samples were therefore prepared using acetonitrile, NMR samples were prepared using CDCl₃ or CD₃CN.

N-hydroxyphthalimide (NHPI) esters

Fmoc-Gly-ONHPI (2a)



On 1.0 mmol scale, Fmoc-Gly-ONHPI (**2a**) was obtained as a colorless foam (363.2 mg, 0.821 mmol, 82%).

The spectroscopic data is consistent with the literature.²

¹H NMR (300 MHz, CDCl₃) δ 7.95 – 7.85 (m, 2H), 7.84 – 7.78 (m, 2H), 7.76 (d, *J* = 7.6 Hz, 2H), 7.59 (d, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.3 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.34 (t, *J* = 5.6 Hz, 1H), 4.53 – 4.32 (m, 4H), 4.24 (t, *J* = 7.1 Hz, 1H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.0, 161.5, 156.1, 143.8, 141.4, 135.1, 128.9, 127.9, 127.2, 125.2, 124.3, 120.1, 67.7, 47.2, 40.8 ppm.

HRMS (ESI, m/z): Calcd for $C_{25}H_{18}N_2O_6Na$ [M + Na]⁺ 465.1057; found 465.1061.

Fmoc-Ala-ONHPI (2b)



On 1.0 mmol scale, Fmoc-Ala-ONHPI (**2b**) was obtained as a colorless solid (375.6 mg, 0.82 mmol, 82%).

¹H NMR (300 MHz, CDCl₃) δ 7.94 – 7.85 (m, 2H), 7.84 – 7.72 (m, 4H), 7.61 (d, *J* = 7.5 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.3 Hz, 2H), 5.31 (d, *J* = 8.9 Hz, 1H), 4.94 – 4.75 (m, 1H), 4.55 – 4.34 (m, 2H), 4.25 (t, *J* = 6.7 Hz, 1H), 1.68 (d, *J* = 7.3 Hz, 3H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.8, 161.6, 155.5, 143.9, 143.7, 141.4, 135.0, 128.9, 127.8, 127.2, 125.2, 124.2, 120.1, 67.4, 48.3, 47.2, 18.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{26}H_{20}N_2O_6Na [M + Na]^+ 479.1214$; found 479.1205.

Fmoc-Val-ONHPI (2c)



On 1.0 mmol scale, Fmoc-Val-ONHPI (2c) was obtained as a colorless solid (439.0 mg, 0.906 mmol, 91%).

¹H NMR (300 MHz, CDCl₃) δ 7.94 – 7.84 (m, 2H), 7.84 – 7.73 (m, 4H), 7.61 (d, *J* = 7.3 Hz, 2H), 7.40 (t, *J* = 7.3 Hz, 2H), 7.32 (t, *J* = 7.3 Hz, 2H), 5.33 (d, *J* = 9.5 Hz, 1H), 4.76 (dd, *J* = 9.3, 4.7 Hz, 1H), 4.47 (d, *J* = 6.7 Hz, 2H), 4.25 (t, *J* = 7.1 Hz, 1H), 2.51 – 2.30 (m, 1H), 1.19 – 1.00 (m, 6H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.8, 161.6, 156.0, 144.0, 143.7, 141.4, 135.0, 128.9, 127.9, 127.2, 125.2, 124.2, 120.1, 67.4, 57.7, 47.3, 31.9, 18.9, 17.5 ppm.

HRMS (ESI, m/z): Calcd for $C_{28}H_{24}N_2O_6Na [M + Na]^+ 507.1527$; found 507.1538.

Fmoc-Leu-ONHPI (2d)



On 1.0 mmol scale, Fmoc-Leu-ONHPI (**2d**) was obtained as a colorless foam (439.7 mg, 0.882 mmol, 88%).

The spectroscopic data is consistent with the literature.³

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.84 (m, 2H), 7.83 – 7.72 (m, 4H), 7.61 (d, *J* = 7.2 Hz, 2H), 7.40 (t, *J* = 7.3 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.20 (d, *J* = 8.7 Hz, 1H), 4.90 – 4.77 (m, 1H), 4.47 (d, *J* = 6.7 Hz, 2H), 4.25 (t, *J* = 6.6 Hz, 1H), 1.98 – 1.82 (m, 2H), 1.82 – 1.69 (m, 1H), 1.03 (d, *J* = 5.7 Hz, 6H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.8, 161.6, 155.7, 144.0, 143.7, 141.5, 135.0, 129.0, 127.8, 127.2, 125.2, 124.2, 120.1, 67.4, 51.1, 47.3, 41.9, 24.9, 23.0, 21.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{29}H_{26}N_2O_6Na [M + Na]^+ 521.1683$; found 521.1684.

Fmoc-Ile-ONHPI (2e)



On 1.0 mmol scale, Fmoc-Ile-ONHPI (2e) was obtained as a colorless oil (463.5 mg, 0.930 mmol, 93%).

¹H NMR (300 MHz, CDCl₃) δ 7.89 – 7.80 (m, 2H), 7.79 – 7.68 (m, 4H), 7.61 (d, *J* = 7.0 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 5.62 – 5.41 (m, 1H), 4.81 (dd, *J* = 9.1,

5.1 Hz, 1H), 4.46 (d, *J* = 6.8 Hz, 2H), 4.23 (t, *J* = 6.3 Hz, 1H), 2.21 – 2.00 (m, 1H), 1.77 – 1.59 (m, 1H), 1.41 – 1.27 (m, 1H), 1.12 (d, *J* = 6.8 Hz, 3H), 1.04 (t, *J* = 7.4 Hz, 3H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.7, 161.5, 155.9, 143.9, 143.6, 141.3, 134.9, 128.8, 127.8, 127.2, 125.1, 124.0, 120.0, 67.3, 57.1, 47.2, 38.3, 24.9, 15.2, 11.6 ppm.

HRMS (ESI, m/z): Calcd for $C_{29}H_{26}N_2O_6Na \ [M + Na]^+ 521.1683$; found 521.1684.

Fmoc-Pro-ONHPI (2f)



On 1.0 mmol scale, Fmoc-Pro-ONHPI (**2f**) was obtained as a colorless foam (437.8 mg, 0.907 mmol, 91%) as a mixture of rotamers.

The spectroscopic data is consistent with the literature.³

¹H NMR (300 MHz, CDCl₃) δ 7.86 – 7.56 (m, 8H), 7.46 – 7.27 (m, 4H), 4.89 – 4.76 (m, 1H), 4.71 – 4.34 (m, 2H), 4.32 – 4.17 (m, 1H), 3.81 – 3.65 (m, 1H), 3.65 – 3.47 (m, 1H), 2.62 – 2.32 (m, 2H), 2.24 – 1.95 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.4, 169.1, 161.5, 154.5, 154.0, 144.7, 143.9, 143.7, 143.5, 141.2, 141.2, 141.1, 141.1, 134.7, 134.7, 128.7, 128.6, 127.6, 127.5, 127.1, 127.0, 125.5, 125.1, 125.0, 123.8, 119.9, 119.8, 68.4, 67.6, 57.3, 57.2, 47.1, 46.9, 46.4, 31.4, 30.2, 24.4, 23.5 ppm.

HRMS (ESI, m/z): Calcd for $C_{28}H_{22}N_2O_6Na [M + Na]^+ 505.1370$; found 505.1365.

Fmoc-Met-ONHPI (2g)



On 1.0 mmol scale, Fmoc-Met-ONHPI (**2g**) was obtained as a colorless solid (429.8 mg, 0.832 mmol, 83%).

The spectroscopic data is consistent with the literature.³

¹H NMR (300 MHz, CDCl₃) δ 7.88 – 7.79 (m, 2H), 7.79 – 7.69 (m, 4H), 7.65 – 7.53 (m, 2H), 7.37 (t, *J* = 7.4 Hz, 2H), 7.33 – 7.23 (m, 2H), 5.79 (d, *J* = 8.4 Hz, 1H), 5.01 (q, *J* = 7.7 Hz, 1H), 4.45 (d, *J* = 6.8 Hz, 2H), 4.21 (t, *J* = 6.6 Hz, 1H), 2.79 – 2.59 (m, 2H), 2.44 – 2.17 (m, 2H), 2.14 (s, 3H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.9, 161.4, 155.6, 143.8, 143.5, 141.2, 134.9, 128.7, 127.7, 127.1, 125.0, 124.0, 119.9, 67.2, 51.7, 47.1, 31.9, 29.5, 15.4 ppm.

HRMS (ESI, m/z): Calcd for $C_{28}H_{24}N_2O_6SNa \ [M + Na]^+ 539.1247$; found 539.1237.

Fmoc-Phe-ONHPI (2h)



On 1.0 mmol scale, Fmoc-Phe-ONHPI (**2h**) was obtained as a colorless solid (414.1 mg, 0.778 mmol, 78%).

¹H NMR (300 MHz, CDCl₃) δ 7.95 – 7.85 (m, 2H), 7.85 – 7.71 (m, 4H), 7.56 (t, *J* = 6.6 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.37 – 7.26 (m, 7H), 5.27 – 5.08 (m, 2H), 4.53 – 4.33 (m, 2H), 4.21 (t, *J* = 6.8 Hz, 1H), 3.43 – 3.25 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.5, 161.5, 155.4, 143.9, 143.7, 141.4, 135.0, 134.5, 129.9, 128.9, 128.9, 127.9, 127.7, 127.2, 125.22, 125.17, 124.2, 120.1, 67.4, 53.1, 47.2, 38.3 ppm.
HRMS (ESI, m/z): Calcd for C₃₂H₂₄N₂O₆Na [M + Na]⁺ 555.1527; found 555.1520.

Fmoc-Asn(Trt)-ONHPI (2i)



On 1.0 mmol scale, Fmoc-Asn(Trt)-ONHPI (2i) was obtained as a colorless solid (591.8 mg, 0.798 mmol, 80%).

¹H NMR (300 MHz, CDCl₃) δ 7.88 – 7.80 (m, 2H), 7.80 – 7.70 (m, 4H), 7.63 – 7.54 (m, 2H), 7.38 (t, *J* = 7.3 Hz, 2H), 7.33 – 7.15 (m, 17H), 7.08 (s, 1H), 6.51 (d, *J* = 8.2 Hz, 1H), 5.04 (dt, *J* = 9.1, 5.0 Hz, 1H), 4.45 – 4.26 (m, 2H), 4.20 (t, *J* = 7.3 Hz, 1H), 3.29 – 2.90 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.3, 168.2, 161.4, 156.0, 144.3, 143.9, 143.8, 141.3, 134.9, 128.9, 128.9, 128.1, 127.7, 127.2, 125.4, 125.3, 124.1, 120.0, 71.3, 67.6, 49.8, 47.1, 38.3 ppm. HRMS (ESI, m/z): Calcd for C₄₆H₃₅N₃O₇Na [M + Na]⁺ 764.2367; found 764.2330.

Fmoc-Gln(Trt)-ONHPI (2j)



On 1.0 mmol scale, Fmoc-Gln(Trt)-ONHPI (**2j**) was obtained as a colorless solid (610.0 mg, 0.807 mmol, 81%).

¹H NMR (300 MHz, CDCl₃) δ 7.91 – 7.83 (m, 2H), 7.83 – 7.77 (m, 2H), 7.74 (d, J = 7.4 Hz, 2H), 7.58 (d, J = 7.3 Hz, 2H), 7.38 (t, J = 7.4 Hz, 2H), 7.33 – 7.17 (m, 17H), 7.03 (s, 1H), 5.88 (d, J = 7.5 Hz, 1H), 4.86 – 4.71 (m, 1H), 4.51 – 4.30 (m, 2H), 4.23 (t, J = 6.8 Hz, 1H), 2.72 – 2.47 (m, 2H), 2.46 – 2.17 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 170.9, 169.0, 161.7, 155.9, 144.6, 144.0, 143.8, 141.4, 135.0, 128.9, 128.8, 128.1, 127.8, 127.3, 127.2, 125.4, 125.3, 124.2, 120.0, 70.9, 67.4, 52.4, 47.3, 32.8, 27.6 ppm.

HRMS (ESI, m/z): Calcd for $C_{47}H_{37}N_3O_7Na$ [M + Na]⁺ 778.2524; found 778.2518.

Fmoc-Asp(tBu)-ONHPI (2k)



On 1.0 mmol scale, Fmoc-Asp(tBu)-ONHPI (**2k**) was obtained as a pale yellow oil (499.2 mg, 0.897 mmol, 90%).

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.86 (m, 2H), 7.83 – 7.72 (m, 4H), 7.64 – 7.56 (m, 2H), 7.39 (t, *J* = 7.3 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.99 (d, *J* = 9.4 Hz, 1H), 5.10 (dt, *J* = 9.4, 4.7 Hz, 1H), 4.54 – 4.32 (m, 2H), 4.26 (t, *J* = 7.2 Hz, 1H), 3.21 – 2.85 (m, 2H), 1.52 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.3, 167.9, 161.4, 155.8, 143.9, 143.7, 141.4, 135.0, 129.0, 127.9, 127.2, 125.3, 125.3, 124.2, 120.1, 82.9, 67.7, 49.3, 47.2, 37.9, 28.2 ppm.

HRMS (ESI, m/z): Calcd for $C_{31}H_{28}N_2O_8Na$ [M + Na]⁺ 579.1738; found 579.1728.

Fmoc-Glu(tBu)-ONHPI (2l)



On 1.0 mmol scale, Fmoc-Glu(tBu)-ONHPI (21) was obtained as a colorless oil (349.0 mg, 0.612 mmol, 61%).

¹H NMR (300 MHz, CDCl₃) δ 7.86 – 7.79 (m, 2H), 7.78 – 7.68 (m, 4H), 7.65 – 7.55 (m, 2H), 7.37 (t, *J* = 7.4 Hz, 2H), 7.29 (t, *J* = 7.4 Hz, 2H), 5.92 (d, *J* = 8.3 Hz, 1H), 4.94 – 4.79 (m, 1H), 4.52 – 4.30 (m, 2H), 4.22 (t, *J* = 6.9 Hz, 1H), 2.60 – 2.46 (m, 2H), 2.46 – 2.32 (m, 1H), 2.31 – 2.16 (m, 1H), 1.48 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 172.0, 168.9, 161.4, 155.8, 143.8, 143.6, 141.2, 134.8, 128.7, 127.7, 127.1, 125.1, 124.0, 119.9, 81.2, 67.3, 52.1, 47.1, 31.2, 28.1, 27.4 ppm.

HRMS (ESI, m/z): Calcd for C₃₂H₃₀N₂O₈Na [M + Na]⁺ 593.1894; found 593.1906.

Fmoc-Ser(tBu)-ONHPI (2m)



On 1.0 mmol scale, Fmoc-Ser(tBu)-ONHPI (2m) was obtained as a colorless oil (495.8 mg, 0.938 mmol, 94%).

¹H NMR (300 MHz, CDCl₃) δ 7.92 – 7.83 (m, 2H), 7.83 – 7.73 (m, 4H), 7.62 (dd, *J* = 7.1, 3.3 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 5.76 (d, *J* = 9.1 Hz, 1H), 4.98 (dt, *J* = 8.8, 2.5 Hz, 1H), 4.53 – 4.34 (m, 2H), 4.27 (t, *J* = 7.3 Hz, 1H), 4.05 (dd, *J* = 8.9, 2.7 Hz, 1H), 3.74 (dd, *J* = 9.0, 3.2 Hz, 1H), 1.28 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.6, 161.5, 155.9, 144.0, 143.8, 141.4, 134.9, 129.0, 127.8, 127.2, 125.3, 124.1, 120.1, 74.2, 67.6, 62.0, 53.6, 47.2, 27.4 ppm.

HRMS (ESI, m/z): Calcd for C₃₀H₂₈N₂O₇Na [M + Na]⁺ 551.1789; found 551.1774.

Fmoc-Thr(tBu)-ONHPI (2n)



On 1.0 mmol scale, Fmoc-Thr(tBu)-ONHPI (**2n**) was obtained as a colorless foam (528.3 mg, 0.974 mmol, 97%).

¹H NMR (300 MHz, CDCl₃) δ 7.87 (dd, J = 5.4, 3.1 Hz, 2H), 7.82 – 7.71 (m, 4H), 7.70 – 7.57 (m, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.32 (t, J = 7.3 Hz, 2H), 5.74 (d, J = 9.5 Hz, 1H), 4.75 – 4.68 (m, 1H), 4.53 – 4.37 (m, 3H), 4.29 (t, J = 7.1 Hz, 1H), 1.34 (d, J = 6.2 Hz, 3H), 1.28 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.9, 161.4, 156.4, 144.0, 143.8, 141.4, 134.8, 128.9, 127.8, 127.2, 125.3, 124.0, 120.0, 74.9, 67.5, 67.4, 58.8, 47.3, 28.7, 20.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{31}H_{30}N_2O_7Na \ [M + Na]^+ 565.1945$; found 565.1943.

Fmoc-Orn(Boc)-ONHPI (20)



On 1.0 mmol scale, Fmoc-Orn(Boc)-ONHPI (**20**) was obtained as a colorless foam (498.1 mg, 0.831 mmol, 83%).

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.85 (m, 2H), 7.84 – 7.73 (m, 4H), 7.62 (d, *J* = 7.6 Hz, 2H), 7.40 (t, *J* = 7.3 Hz, 2H), 7.32 (t, *J* = 7.1 Hz, 2H), 5.69 – 5.47 (m, 1H), 4.90 – 4.77 (m, 1H), 4.76 – 4.58 (m, 1H), 4.47 (d, *J* = 6.8 Hz, 2H), 4.25 (t, *J* = 5.6 Hz, 0H), 3.32 – 3.12 (m, 2H), 2.20 – 1.87 (m, 2H), 1.78 – 1.62 (m, 2H), 1.46 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.1, 161.5, 156.3, 155.8, 143.9, 143.7, 141.3, 134.9, 128.8, 127.8, 127.2, 125.2, 124.1, 120.0, 79.4, 67.3, 52.4, 47.2, 39.8, 29.7, 28.5, 25.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{33}H_{33}N_3O_8Na$ [M + Na]⁺ 622.2160; found 622.2131.

Fmoc-Lys(Boc)-ONHPI (2p)



On 1.0 mmol scale, Fmoc-Lys(Boc)-ONHPI (**2p**) was obtained as a colorless solid (516.0 mg, 0.841 mmol, 84%).

¹H NMR (300 MHz, CDCl₃) δ 7.92 – 7.83 (m, 2H), 7.82 – 7.72 (m, 4H), 7.61 (d, *J* = 7.4 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.57 (d, *J* = 7.7 Hz, 1H), 4.87 – 4.67 (m,

2H), 4.53 – 4.31 (m, 2H), 4.24 (t, *J* = 7.0 Hz, 1H), 3.29 – 3.03 (m, 2H), 2.14 – 1.83 (m, 2H), 1.66 – 1.49 (m, 4H), 1.44 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.3, 161.6, 156.3, 155.8, 143.9, 143.7, 141.4, 135.0, 128.9, 127.8, 127.2, 125.2, 124.2, 120.1, 79.3, 67.4, 52.4, 47.2, 39.9, 32.1, 29.6, 28.6, 22.0 ppm.
HRMS (ESI, m/z): Calcd for C₃₄H₃₅N₃O₈Na [M + Na]⁺ 636.2316; found 636.2307.

Fmoc-Arg(Boc)₂-ONHPI (2q)



NOTE: Other arginine protecting groups (-Pbf, -Mtr, -Tos, -NO₂) have also been investigated. However, the corresponding NHPI esters could not be isolated due to the highly favored δ lactam formation. This side reaction was likely suppressed in the present example by the steric hindrance of the dual Boc protection. However, it is expected that the described compound is not suitable for long-term storage.

On 0.5 mmol scale, Fmoc-Arg(Boc)₂-ONHPI (**2q**) was obtained as a colorless foam (310.7 mg, 0.419 mmol, 84%).

¹H NMR (300 MHz CDCl₃) δ 11.51 (s, 1H), 8.42 (t, *J* = 4.9 Hz, 1H), 7.91 – 7.80 (m, 2H), 7.80 – 7.69 (m, 4H), 7.68 – 7.55 (m, 2H), 7.37 (t, *J* = 7.4 Hz, 2H), 7.33 – 7.26 (m, 2H), 6.26 (d, *J* = 8.5 Hz, 1H), 4.93 – 4.75 (m, 1H), 4.44 (d, *J* = 7.1 Hz, 2H), 4.23 (t, *J* = 6.7 Hz, 1H), 3.73 – 3.50 (m, 1H), 3.48 – 3.31 (m, 1H), 2.20 – 1.90 (m, 2H), 1.88 – 1.70 (m, 2H), 1.48 (s, 18H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 169.0, 163.4, 161.5, 156.5, 155.9, 153.3, 143.9, 143.7, 141.3, 134.9, 128.8, 127.7, 127.1, 125.3, 125.2, 124.0, 120.0, 83.2, 79.4, 67.2, 52.6, 47.2, 40.1, 29.0, 28.3, 28.1, 25.8 ppm.

HRMS (ESI, m/z): Calcd for $C_{39}H_{43}N_5O_{10}Na$ [M + Na]⁺ 764.2902; found 764.2880.

Fmoc-Tyr(tBu)-ONHPI (2r)



On 1.0 mmol scale, Fmoc-Tyr(tBu)-ONHPI (**2r**) was obtained as a pale yellow oil (491.8 mg, 0.813 mmol, 81%).

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.85 (m, 2H), 7.82 – 7.72 (m, 4H), 7.61 – 7.52 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 7.20 (d, *J* = 8.2 Hz, 2H), 6.96 (d, *J* = 8.4 Hz, 2H), 5.27 (d, *J* = 8.8 Hz, 1H), 5.17 – 5.05 (m, 1H), 4.51 – 4.28 (m, 2H), 4.21 (t, *J* = 6.9 Hz, 1H), 3.40 – 3.19 (m, 2H), 1.34 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.5, 161.5, 155.4, 155.0, 143.9, 143.7, 141.4, 135.0, 130.3, 129.2, 128.9, 127.8, 127.2, 125.2, 125.2, 124.4, 124.2, 120.1, 78.6, 67.3, 53.2, 47.2, 37.6, 29.0 ppm.

HRMS (ESI, m/z): Calcd for $C_{36}H_{32}N_2O_7Na \ [M + Na]^+ 627.2102$; found 627.2100.

Fmoc-Trp(Boc)-ONHPI (2s)



On 1.0 mmol scale, Fmoc-Trp(Boc)-ONHPI (**2s**) was obtained as a colorless foam (469.2 mg, 0.739 mmol, 74%).

¹H NMR (300 MHz, CDCl₃) δ 8.23 (d, *J* = 7.9 Hz, 1H), 7.91 – 7.78 (m, 3H), 7.79 – 7.69 (m, 4H), 7.64 (d, *J* = 7.7 Hz, 1H), 7.58 – 7.46 (m, 2H), 7.44 – 7.34 (m, 3H), 7.33 – 7.22 (m, 3H), 5.64 (d, *J* = 8.6 Hz, 1H), 5.34 – 5.26 (m, 1H), 4.48 – 4.33 (m, 2H), 4.25 – 4.17 (m, 1H), 3.65 – 3.39 (m, 2H), 1.71 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.5, 161.3, 155.5, 149.5, 143.7, 143.5, 141.2, 135.5, 134.8, 130.5, 128.7, 127.7, 127.1, 125.1, 124.6, 123.9, 122.8, 119.9, 118.6, 115.4, 113.4, 83.6, 67.4, 52.9, 46.9, 28.1, 27.9 ppm.

HRMS (ESI, m/z): Calcd for C₃₉H₃₃N₃O₈Na [M + Na]⁺ 694.2160; found 694.2147.

Fmoc-His(Boc)-ONHPI (2t)



On 1.0 mmol scale, Fmoc-His(Boc)-ONHPI (**2**t) was obtained as a white foam (522.0 mg, 0.838 mmol, 84%).

¹H NMR (300 MHz, CDCl₃) δ 8.09 (s, 1H), 7.88 – 7.79 (m, 2H), 7.79 – 7.70 (m, 4H), 7.66 – 7.57 (m, 2H), 7.49 (s, 1H), 7.37 (t, *J* = 7.3 Hz, 2H), 7.33 – 7.25 (m, 2H), 6.66 (d, *J* = 8.2 Hz, 1H), 5.20 – 5.06 (m, 1H), 4.45 – 4.31 (m, 2H), 4.25 (t, *J* = 7.3 Hz, 1H), 3.44 – 3.21 (m, 2H), 1.62 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 168.3, 161.4, 155.9, 146.8, 143.9, 143.8, 141.3, 137.1, 134.8, 134.0, 128.9, 127.7, 127.1, 125.4, 125.3, 124.0, 120.0, 116.1, 85.8, 67.6, 52.7, 47.1, 29.8, 27.9 ppm.

HRMS (ESI, m/z): Calcd for C₃₄H₃₀N₄O₈Na [M + Na]⁺ 645.1956; found 645.1962.

Fmoc-His(Trt)-ONHPI (2u)



On 0.5 mmol scale, Fmoc-His(Trt)-ONHPI (**2u**) was obtained as a colorless solid (160.6 mg, 0.210 mmol, 42%).

NOTE: Deactivated silica gel (100% $SiO_2/35\%$ H₂O) was used for chromatographic purification.

¹H NMR (300 MHz, CD₃CN) δ 7.86 – 7.70 (m, 6H), 7.58 (d, *J* = 7.2 Hz, 2H), 7.41 (s, 1H), 7.34 – 7.16 (m, 15H), 7.12 – 7.02 (m, 6H), 6.93 (s, 1H), 5.01 – 4.75 (m, 1H), 4.31 (d, *J* = 6.6 Hz, 2H), 4.15 (t, *J* = 6.5 Hz, 1H), 3.25 – 3.08 (m, 2H) ppm.

¹³C NMR (75 MHz, CD₃CN) δ 169.8, 162.5, 156.7, 144.8, 143.2, 142.0, 136.1, 135.9, 135.0, 130.4, 129.4, 129.0, 128.9, 128.6, 128.0, 126.1, 124.8, 121.7, 120.9, 76.2, 67.6, 53.8, 47.8, 30.1 ppm.

HRMS (ESI, m/z): Calcd for $C_{48}H_{37}N_4O_6$ [M + H]⁺ 765.2708; found 765.2702.

Fmoc-Cys(Mob)-ONHPI (2v)



On 1.0 mmol scale, Fmoc-Cys(Mob)-ONHPI (**2v**) was obtained as a colorless solid (518.3 mg, 0.852 mmol, 85%).

¹H NMR (300 MHz, CDCl₃) δ 7.93 – 7.82 (m, 2H), 7.82 – 7.71 (m, 4H), 7.61 (d, *J* = 7.3 Hz, 2H), 7.40 (t, *J* = 7.2 Hz, 2H), 7.35 – 7.23 (m, 4H), 6.89 – 6.81 (m, 2H), 5.66 (d, *J* = 8.5 Hz, 1H), 5.07 – 4.95 (m, 1H), 4.46 (d, *J* = 6.9 Hz, 2H), 4.26 (t, *J* = 6.9 Hz, 1H), 3.81 (s, 2H), 3.76 (s, 3H), 3.18 – 2.93 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.9, 161.4, 158.9, 155.6, 143.8, 143.7, 141.3, 135.0, 130.2, 129.3, 128.8, 127.8, 127.2, 125.2, 124.2, 120.1, 114.2, 67.6, 55.3, 52.1, 47.1, 36.2, 33.7 ppm.
HRMS (ESI, m/z): Calcd for C₃₄H₂₈N₂O₇SNa [M + Na]⁺ 631.1509; found 631.1504.

Fmoc-Cys(StBu)-ONHPI (2w)



On 1.0 mmol scale, Fmoc-Cys(StBu)-ONHPI (**2w**) was obtained as a colorless solid (383.7 mg, 0.665 mmol, 67%).

¹H NMR (300 MHz, CDCl₃) δ 7.91 – 7.83 (m, 2H), 7.80 – 7.72 (m, 4H), 7.63 (d, J = 7.1 Hz, 2H), 7.39 (t, J = 7.3 Hz, 2H), 7.31 (t, J = 7.2 Hz, 2H), 5.80 (d, J = 8.3 Hz, 1H), 5.23 – 5.10 (m, 1H), 4.52 – 4.36 (m, 2H), 4.27 (t, J = 7.0 Hz, 1H), 3.50 – 3.17 (m, 2H), 1.39 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.8, 161.4, 155.6, 143.8, 143.7, 141.3, 134.9, 128.8, 127.8, 127.2, 125.3, 124.1, 120.0, 67.7, 52.4, 48.6, 47.1, 42.1, 29.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{30}H_{28}N_2O_6S_2Na$ [M + Na]⁺ 599.1281; found 599.1293.

Fmoc-Cys(Trt)-ONHPI (2x)



On 1.0 mmol scale, Fmoc-Cys(Trt)-ONHPI (**2x**) was obtained as a colorless foam (467.0 mg, 0.639 mmol, 64%).

¹H NMR (300 MHz, CDCl₃) δ 7.91 – 7.83 (m, 2H), 7.82 – 7.72 (m, 4H), 7.60 (d, J = 7.2 Hz, 2H), 7.49 (d, J = 7.4 Hz, 6H), 7.40 (t, J = 7.4 Hz, 2H), 7.36 – 7.18 (m, 11H), 5.14 (d, J = 8.2 Hz, 1H), 4.58 – 4.30 (m, 3H), 4.25 (t, J = 7.0 Hz, 1H), 2.97 – 2.77 (m, 2H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 167.6, 161.3, 155.4, 144.2, 143.9, 143.7, 141.4, 134.9, 129.7, 128.9, 128.3, 127.8, 127.2, 127.2, 125.3, 124.1, 120.1, 67.8, 67.4, 51.7, 47.2, 33.7 ppm.

HRMS (ESI, m/z): Calcd for $C_{45}H_{34}N_2O_6SNa [M + Na]^+ 753.2030$; found 753.2034.

Synthesis of α-aminopinacolyl boronates



General procedure

Pinacolyl boronates were prepared by a modified, literature-known procedure.⁴ Most tested α amino pinacolyl boronates lacked stability even on deactivated silica or alumina columns. Thus, the workup was optimised, allowing to circumvent chromatography. It was found that stabilised 2-iodoxybenzoic acid (sIBX)⁵ oxidatively destroys B₂pin₂ while leaving the C–B bond of α amino pinacolyl boronates unharmed under the tested conditions.

A round-bottom flask was charged with the corresponding NHPI ester (1.0 eq) and MgBr₂·OEt₂ (1.5 eq). After evacuating and flushing the flask with nitrogen for three times, a suspension of NiCl₂·6 H₂O (0.1 eq) and 4,4'-dimethoxy-2,2'-bipyridine (0.13 eq) in stabilised THF (4 mL/mmol), prepared 24 hours prior to use, was added. The suspension was stirred for 10 minutes at room temperature and then cooled to 0°C. Afterwards, a suspension of pre-complexed [B₂pin₂Me]Li (3.0 eq) in stabilised THF (5 mL/mmol), prepared one hour prior to use, was added in one portion. The resulting mixture was stirred for one hour at 0°C and for one additional hour at ambient temperature. The suspension was diluted with ether (~50 mL/mmol) and quickly filtered through a short pad of celite and silica gel. After washing with additional ether (~100 mL/mmol), the filtrate was concentrated *in vacuo* to obtain the desired product containing B₂pin₂ and phthalimide as impurities.

Workup procedure (Route 1)

The crude product was dissolved in ethyl acetate (~50 mL/g of crude product). Stabilised 2iodoxybenzoic acid⁵ (sIBX, 6.0 eq) was added and the mixture was stirred for two hours at 40°C. The solvent was removed under reduced pressure, the residue was redissolved in methylene chloride and washed three times with aqueous K₂CO₃ solution (w = 10%) and once with saturated aqueous NaCl solution. The organic phase was dried over anhydrous MgSO₄ and dried *in vacuo* to obtain the corresponding pinacolyl boronate **3**.

Workup procedure (Route 2)

The crude product was dissolved in methylene chloride and washed three times with aqueous K_2CO_3 solution (w = 10%) and once with saturated aqueous NaCl solution. The organic phase was dried over anhydrous MgSO₄ and dried *in vacuo* to obtain the corresponding pinacolyl boronate **3** as a crude containing B_2pin_2 contaminants.

NOTE: The organic phase should be washed thoroughly with K_2CO_3 solution to completely remove IBX stabilisers that are present in the material.

α-Aminopinacolyl boronates

Fmoc-Gly-Bpin (3a)

H O H

Route 1: On 0.681 mmol scale, Fmoc-Gly-Bpin (**3a**) was obtained as a pale yellow oil (117.5 mg, 0.310 mmol, 46%).

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.4 Hz, 2H), 7.62 (d, *J* = 7.3 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.31 (td, *J* = 7.4, 1.2 Hz, 2H), 4.99 – 4.88 (m, 1H), 4.39 (d, *J* = 7.0 Hz, 2H), 4.23 (t, *J* = 6.9 Hz, 1H), 2.90 (d, *J* = 4.4 Hz, 2H), 1.29 (s, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.5, 144.2, 141.3, 127.7, 127.1, 125.2, 120.0, 84.4, 66.9, 47.4, 24.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{22}H_{26}BNO_4Na [M + Na]^+ 402.1851$; found 402.1847.

Fmoc-Ala-Bpin ((±)-3b)

Fmoc^N B O

Route 1: On 0.656 mmol scale, Fmoc-Ala-Bpin ((\pm)-**3b**) was obtained as a colorless oil (150.6 mg, 0.383 mmol, 58%).

Route 2: On 0.486 mmol scale using compound **2b**, Fmoc-Ala-Bpin ((\pm)-**3b**) was obtained as a pale yellow crude material (254.5 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.62 (d, *J* = 7.4 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (td, *J* = 7.4, 1.2 Hz, 2H), 5.02 (d, *J* = 5.8 Hz, 1H), 4.46 – 4.28 (m, 2H), 4.23 (t, *J* = 7.1 Hz, 1H), 3.29 – 3.15 (m, 1H), 1.27 – 1.23 (m, 15H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 156.9, 144.1, 144.0, 141.3, 127.6, 127.0, 125.2, 119.9, 84.1, 66.7, 47.3, 25.0, 16.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{23}H_{28}BNO_4Na \ [M + Na]^+ 416.2008$; found 416.2005.

Fmoc-Val-Bpin ((±)-3c)



Route 1: On 0.887 mmol scale, Fmoc-Val-Bpin ((\pm)-**3c**) was obtained as a colorless oil (247.2 mg, 0.587 mmol, 66%).

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.65 – 7.58 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.04 (d, *J* = 7.3 Hz, 1H), 4.46 – 4.29 (m, 2H), 4.25 (t, *J* = 7.2 Hz, 1H), 3.19 (dd, *J* = 7.3, 5.2 Hz, 1H), 2.08 – 1.90 (m, 1H), 1.29 – 1.23 (m, 13H), 1.02 – 0.80 (m, 6H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.1, 144.3, 144.2, 141.4, 127.7, 127.1, 125.3, 120.1, 84.2, 66.8, 47.5, 30.6, 25.1, 24.9, 20.0, 19.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{25}H_{32}BNO_4Na [M + Na]^+ 444.2321$; found 444.2342.

Fmoc-Leu-Bpin ((±)-3d)

Fmoc H B O

Route 1: On 0.772 mmol scale, Fmoc-Leu-Bpin ((\pm)-**3d**) was obtained as a pale yellow oil (234.6 mg, 0.539 mmol, 70%).

¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, *J* = 7.4 Hz, 2H), 7.66 – 7.56 (m, 2H), 7.38 (t, *J* = 7.4 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 2H), 4.97 (d, *J* = 6.5 Hz, 1H), 4.47 – 4.29 (m, 2H), 4.24 (t, *J* = 7.0 Hz, 1H), 3.26 (q, *J* = 7.1 Hz, 1H), 1.77 – 1.59 (m, 1H), 1.56 – 1.39 (m, 2H), 1.27 (s, 12H), 1.00 – 0.84 (m, 6H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 156.9, 144.2, 144.1, 141.3, 127.6, 127.0, 125.2, 119.9, 84.0, 66.7, 47.4, 40.4, 25.3, 24.9, 24.8, 23.2, 22.3 ppm.

HRMS (ESI, m/z): Calcd for C₂₆H₃₄BNO₄ [M + Na]⁺ 458.2478; found 458.2471.

Fmoc-Ile-Bpin (3e)



Route 1: On 0.712 mmol scale, Fmoc-Ile-Bpin (**3e**) was obtained as a pale yellow oil (160.9 mg, 0.370 mmol, 52%) as a mixture of diastereomers.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.65 – 7.58 (m, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 5.09 (d, *J* = 6.9 Hz, 0.5H, diastereomer), 5.04 (d, *J* = 8.2 Hz, 0.5H, diastereomer), 4.46 – 4.29 (m, 2H), 4.25 (t, *J* = 7.2 Hz, 1H), 3.36 (dd, *J* = 7.7, 4.2 Hz, 0.5H, diastereomer), 3.29 (dd, *J* = 7.3, 5.3 Hz, 0.5H, diastereomer), 1.84 – 1.65 (m, 1H), 1.62 – 1.39 (m, 2H), 1.26 (s, 12H), 1.17 – 1.03 (m, 1H), 0.98 – 0.83 (m, 6H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ (157.1 and 156.9, diastereomers), 144.3, 144.2, 141.4, 127.7, 127.1, 125.3, 120.0, 84.2, 66.7, 47.5, 37.4, 27.0, 25.1, 24.9, (16.8 and 16.7, diastereomers), (12.3 and 12.1, diastereomers) ppm.

HRMS (ESI, m/z): Calcd for $C_{26}H_{34}BNO_4Na [M + Na]^+ 458.2478$; found 458.2467.

Fmoc-Pro-Bpin ((±)-3f)



Route 1: On 0.908 mmol scale, Fmoc-Pro-Bpin ((\pm) -**3f**) was obtained as a colorless solid (231.3 mg, 0.552 mmol, 61%) as a mixture of rotamers.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, J = 7.4 Hz, 2H), 7.71 – 7.59 (m, 2H), 7.45 – 7.35 (m, 2H), 7.35 – 7.27 (m, 2H), 4.69 – 4.00 (m, 3H), 3.63 – 3.42 (m, 2H), 3.42 – 3.32 (m, 0.5H, rotamer), 3.13 (dd, J = 9.7, 6.6 Hz, 0.5H, rotamer), 2.11 – 1.72 (m, 4H), 1.32 – 1.15 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 155.2, 144.9, 144.43, 144.41, 141.40, 141.36, 127.8, 127.7, 127.2, 127.1, 125.6, 125.5, 120.0, 83.9, 83.8, 67.4, 67.3, 47.7, 47.5, 47.3, 46.2, 29.1, 28.1, 27.2, 25.8, 25.01, 24.96, 24.7, 24.6 ppm.

HRMS (ESI, m/z): Calcd for $C_{25}H_{30}BNO_4Na [M + Na]^+ 442.2164$; found 442.2155.

Fmoc-Met-Bpin ((±)-3g)



Route 2: On 0.784 mmol scale, Fmoc-Met-Bpin ((\pm)-**3**g) was obtained as a pale yellow crude material (350.7 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

NOTE: For spectroscopic analysis, a small amount of Fmoc-Met-B(OH)₂ ((\pm)-4g) was esterified with pinacol (1.0 eq) in methylene chloride for one hour. After drying *in vacuo*, the title compound (\pm)-3g was obtained as a colorless solid.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.4 Hz, 2H), 7.64 – 7.57 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.10 (d, *J* = 6.5 Hz, 1H), 4.51 – 4.28 (m, 2H), 4.23 (t, *J* = 6.7 Hz, 1H), 3.36 – 3.25 (m, 1H), 2.61 – 2.43 (m, 2H), 2.08 (s, 3H), 2.06 – 1.96 (m, 1H), 1.94 – 1.79 (m, 1H), 1.27 (s, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.0, 144.3, 144.1, 141.5, 127.8, 127.2, 125.3, 120.1, 84.4, 66.8, 47.5, 31.5, 30.2, 25.0, 24.9, 15.5 ppm.

HRMS (ESI, m/z): Calcd for $C_{25}H_{32}BNO_4SNa [M + Na]^+ 476.2042$; found 476.2048.

Fmoc-Phe-Bpin ((±)-3h)



Route 1: On 0.783 mmol scale, Fmoc-Phe-Bpin $((\pm)$ -**3h**) was obtained as a colorless solid (168.6 mg, 0.359 mmol, 46%).

¹H NMR (300 MHz, CDCl₃) δ 7.78 (d, *J* = 7.4 Hz, 2H), 7.62 (d, *J* = 7.4 Hz, 2H), 7.42 (t, *J* = 7.3 Hz, 2H), 7.37 – 7.15 (m, 7H), 4.98 (d, *J* = 6.2 Hz, 1H), 4.55 – 4.30 (m, 2H), 4.24 (t, *J* = 6.8 Hz, 1H), 3.46 (q, *J* = 6.3 Hz, 1H), 3.09 – 2.84 (m, 1H), 1.30 – 1.20 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.0, 144.1, 144.0, 141.4, 139.5, 129.5, 128.4, 127.7, 127.1, 126.4, 125.2, 120.0, 84.2, 66.8, 47.4, 36.8, 25.0, 24.8 ppm.

HRMS (ESI, m/z): Calcd for C₂₉H₃₂BNO₄ [M + Na]⁺ 492.2322; found 492.2313.

Fmoc-Asn(Trt)-Bpin ((±)-3i)



Route 1: On 0.800 mmol scale, Fmoc-Asn(Trt)-Bpin ((±)-**3i**) was obtained as a colorless solid (204.6 mg, 0.301 mmol, 38%).

¹H NMR (300 MHz, CDCl₃) δ 7.81 – 7.70 (m, 2H), 7.66 – 7.54 (m, 2H), 7.46 – 7.35 (m, 2H), 7.34 – 7.13 (m, 17H, overlapping with the solvent signal), 6.63 (s, 1H), 5.53 (d, *J* = 7.7 Hz, 1H), 4.47 – 4.26 (m, 2H), 4.21 (t, *J* = 7.3 Hz, 1H), 3.39 – 3.26 (m, 1H), 2.87 – 2.66 (m, 2H), 1.27 (s, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.3, 144.5, 144.1, 144.0, 141.3, 128.7, 128.0, 127.6, 127.6, 127.1, 125.3, 119.9, 83.7, 71.0, 66.9, 47.3, 39.1, 24.8, 24.7 ppm.

HRMS (ESI, m/z): Calcd for C₄₃H₄₃BN₂O₅Na [M + Na]⁺ 701.3165; found 701.3152.

Fmoc-Gln(Trt)-Bpin ((±)-3j)



Route 1: On 0.780 mmol scale, Fmoc-Gln(Trt)-Bpin ((\pm)-**3j**) was obtained as a colorless solid (231.4 mg, 0.334 mmol, 43%).

¹H NMR (300 MHz, CDCl₃) δ δ 7.77 (d, *J* = 7.6 Hz, 2H), 7.64 (d, *J* = 7.4 Hz, 2H), 7.47 – 7.36 (m, 2H), 7.35 – 7.22 (m, 17H), 7.12 (s, 1H), 5.25 (d, *J* = 6.8 Hz, 1H), 4.50 – 4.34 (m, 2H), 4.26 (t, *J* = 6.9 Hz, 1H), 3.29 – 3.15 (m, 1H), 2.50 – 2.27 (m, 2H), 2.11 – 1.78 (m, 2H), 1.31 – 1.26 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 172.2, 157.6, 144.8, 144.1, 143.9, 141.3, 128.8, 127.9, 127.7, 127.1, 126.9, 125.2, 123.5, 120.0, 84.2, 70.6, 66.8, 47.4, 34.9, 27.9, 24.9, 24.9 ppm.

HRMS (ESI, m/z): Calcd for C₄₄H₄₅BN₂O₅Na [M + Na]⁺ 715.3321; found 715.3303.

Fmoc-Asp(tBu)-Bpin ((±)-3k)



Route 1: On 0.897 mmol scale, Fmoc-Asp(tBu)-Bpin ((\pm) -3k) was obtained as a colorless oil (230.3 mg, 0.467 mmol, 52%).

¹H NMR (300 MHz, CDCl₃) δ 7.74 (d, *J* = 7.5 Hz, 2H), 7.59 (t, *J* = 6.5 Hz, 2H), 7.37 (t, *J* = 7.4 Hz, 2H), 7.33 – 7.25 (m, 2H), 5.53 (d, *J* = 7.0 Hz, 1H), 4.41 – 4.26 (m, 2H), 4.21 (t, *J* = 7.2 Hz, 1H), 3.44 – 3.33 (m, 1H), 2.78 – 2.53 (m, 2H), 1.45 (s, 9H), 1.30 – 1.25 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 172.9, 156.9, 144.1, 143.9, 141.2, 127.6, 127.0, 125.2, 119.9, 84.3, 80.9, 66.9, 47.2, 37.4, 28.2, 24.9, 24.7 ppm.

HRMS (ESI, m/z): Calcd for $C_{28}H_{36}BNO_6Na \ [M + Na]^+ 516.2533$; found 516.2529.

Fmoc-Glu(tBu)-Bpin ((±)-3l)



Route 1: On 0.587 mmol scale, Fmoc-Glu(tBu)-Bpin ((\pm) -**3l**) was obtained as a pale yellow oil (192.0 mg, 0.378 mmol, 64%).

¹H NMR (300 MHz, CDCl₃) δ 7.75 (d, *J* = 7.4 Hz, 2H), 7.60 (dd, *J* = 7.2, 2.3 Hz, 2H), 7.38 (t, *J* = 7.3 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 2H), 5.18 (d, *J* = 6.4 Hz, 1H), 4.43 – 4.28 (m, 2H), 4.22 (t, *J* = 7.0 Hz, 1H), 3.18 (q, *J* = 6.3 Hz, 1H), 2.40 – 2.21 (m, 2H), 2.03 – 1.74 (m, 2H), 1.44 (s, 9H), 1.28 – 1.25 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 173.1, 157.1, 144.2, 144.0, 141.3, 127.7, 127.1, 125.2, 120.0, 84.2, 80.3, 66.9, 47.3, 33.1, 28.2, 26.8, 24.9, 24.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{29}H_{38}BNO_6Na \ [M + Na]^+ 530.2690$; found 530.2683.

Fmoc-Ser(tBu)-Bpin ((±)-3m)



Route 1: On 0.727 mmol scale, Fmoc-Ser(tBu)-Bpin ((\pm) -**3m**) was obtained as a colorless oil (182.7 mg, 0.393 mmol, 54%).

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.4 Hz, 2H), 7.67 – 7.57 (m, 2H), 7.40 (t, *J* = 7.2 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.43 (d, *J* = 6.3 Hz, 1H), 4.45 – 4.29 (m, 2H), 4.25 (t, *J* = 7.1 Hz, 1H), 3.57 (d, *J* = 4.1 Hz, 2H), 3.40 – 3.30 (m, 1H), 1.29 (s, 12H), 1.17 (s, 9H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.3, 144.2, 144.1, 141.4, 127.7, 127.1, 125.3, 120.0, 84.1, 72.9, 67.0, 63.0, 47.4, 27.6, 24.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{27}H_{36}BNO_5Na \ [M + Na]^+ 488.2584$; found 488.2608.

Fmoc-Thr(tBu)-Bpin (3n)



Route 1: On 0.967 mmol scale, Fmoc-Thr(tBu)-Bpin (**3n**) was obtained as a pale yellow oil (295.2 mg, 0.616 mmol, 64%) as a mixture of diastereomers.

Route 2: On 0.471 mmol scale, Fmoc-Thr(tBu)-Bpin (**3n**) was obtained as a pale yellow crude material (316.7 mg, containing B_2pin_2 as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.68 – 7.56 (m, 2H), 7.39 (t, *J* = 7.3 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.44 (d, *J* = 7.2 Hz, 0.5H, diastereomer), 5.37 (d, *J* = 4.4 Hz, 0.5H, diastereomer), 4.52 – 4.17 (m, 3H), 3.98 – 3.84 (m, 1H), 3.21 (dd, *J* = 7.0, 1.7 Hz, 0.5H, diastereomer), 2.88 (dd, *J* = 8.5, 4.6 Hz, 0.5H, diastereomer), 1.29 – 1.21 (m, 21H), 1.17 (d, *J* = 6.0 Hz, 3H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ (157.5 and 157.1, diastereomers), 144.1, 141.3, 127.7, 127.6, 127.1, 127.1, 125.3, 125.2, 120.1, 120.0, 84.1, 83.8, 73.9, 73.2, 70.3, 68.0, 67.3, 66.8, 47.4, 47.3, 28.9, 28.7, 25.1, 24.9, 24.8, 21.3, 20.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{28}H_{38}BNO_5Na [M + Na]^+ 502.2740$; found 502.2760.

Fmoc-Orn(Boc)-Bpin ((±)-30)



Route 1: On 0.747 mmol scale, Fmoc-Orn(Boc)-Bpin ((±)-**30**) was obtained as a colorless solid (178.4 mg, 0.332 mmol, 45%).

Route 2: On 0.830 mmol scale, Fmoc-Orn(Boc)-Bpin ((\pm)-**30**) was obtained as a pale yellow crude material (475 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 7.5 Hz, 2H), 7.61 (d, *J* = 6.9 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.05 (d, *J* = 7.0 Hz, 1H), 4.70 – 4.57 (m, 1H), 4.48 – 4.28 (m, 2H), 4.22 (t, *J* = 6.9 Hz, 1H), 3.25 – 2.99 (m, 3H), 1.77 – 1.48 (m, 4H), 1.44 (s, 9H), 1.29 – 1.26 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.0, 156.1, 144.2, 144.0, 141.4, 127.7, 127.1, 125.2, 125.1, 120.0, 84.3, 79.1, 66.7, 47.4, 40.5, 28.7, 28.5, 27.3, 25.1, 24.9 ppm.

HRMS (ESI, m/z): Calcd for C₃₀H₄₁BN₂O₆Na [M + Na]⁺ 559.2955; found 559.2952.

Fmoc-Lys(Boc)-Bpin ((±)-3p)



Route 1: On 0.840 mmol scale, Fmoc-Lys(Boc)-Bpin ((±)-**3p**) was obtained as a pale yellow oil (143.2 mg, 0.428 mmol, 51%).

Route 2: On 0.845 mmol scale, Fmoc-Lys(Boc)-Bpin ((\pm)-**3p**) was obtained as a pale yellow crude material (466 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 7.74 (d, J = 7.4 Hz, 2H), 7.62 – 7.55 (m, 2H), 7.37 (t, J = 7.4 Hz, 2H), 7.29 (t, J = 7.4 Hz, 2H), 5.10 (d, J = 5.7 Hz, 1H), 4.78 – 4.58 (m, 1H), 4.36 (t, J = 7.1

Hz, 2H), 4.21 (t, *J* = 7.0 Hz, 1H), 3.20 (q, *J* = 7.1, 6.6 Hz, 1H), 3.14 – 2.96 (m, 2H), 1.74 – 1.32 (m, 15H), 1.25 (s, 12H) ppm.

¹³C NMR (75 MHz CDCl₃) δ 157.0, 156.1, 144.3, 144.2, 141.4, 127.7, 127.1, 125.3, 120.1, 84.3, 79.1, 66.8, 47.5, 40.6, 31.2, 29.9, 28.6, 25.0, 24.9, 24.0 ppm.

HRMS (ESI, m/z): Calcd for C₃₁H₄₃BN₂O₆ [M + Na]⁺ 573.3112; found 573.3121.

Fmoc-Arg(Boc₂)-Bpin ((±)-3q)



Route 1: On 0.674 mmol scale, Fmoc-Arg(Boc₂)-Bpin ((\pm)-**3q**) was obtained as a colorless oil (213.6 mg, 0.315 mmol, 47%).

Route 2: On 0.419 mmol scale, Fmoc-Arg(Boc₂)-Bpin ((\pm)-**3q**) was obtained as a pale yellow crude material (243 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 11.49 (s, 1H), 8.34 (t, *J* = 5.3 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.61 (d, *J* = 7.3 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 5.12 (d, *J* = 6.5 Hz, 1H), 4.48 – 4.29 (m, 2H), 4.23 (t, *J* = 7.0 Hz, 1H), 3.40 (q, *J* = 6.8 Hz, 2H), 3.27 – 3.16 (m, 1H), 1.82 – 1.55 (m, 4H), 1.51 – 1.46 (m, 18H), 1.26 (s, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 163.7, 157.0, 156.3, 153.4, 144.3, 144.1, 141.4, 127.7, 127.1, 125.35, 125.29, 120.0, 84.3, 83.1, 79.3, 66.8, 47.5, 40.9, 28.7, 28.4, 28.2, 26.5, 25.0, 24.9 ppm. HRMS (ESI, m/z): Calcd for C₃₆H₅₂BN₄O₈ [M + H]⁺ 679.3879; found 679.3866.

Fmoc-Tyr(tBu)-Bpin ((±)-3r)



Route 1: On 0.482 mmol scale, Fmoc-Tyr(tBu)-Bpin ((\pm)-**3r**) was obtained as a yellow oil (147.2 mg, 0.272 mmol, 56%).

¹H NMR (300 MHz, CDCl₃) δ 7.77 (d, *J* = 7.6 Hz, 2H), 7.61 (d, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.2 Hz, 2H), 7.31 (t, *J* = 7.5 Hz, 2H), 7.04 (d, *J* = 8.3 Hz, 2H), 6.88 (d, *J* = 8.3 Hz, 2H), 4.92 (d, *J* = 6.2 Hz, 1H), 4.54 – 4.30 (m, 2H), 4.23 (t, *J* = 6.6 Hz, 1H), 3.43 (q, *J* = 6.3 Hz, 1H), 3.03 – 2.64 (m, 2H), 1.32 (s, 9H), 1.25 – 1.16 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 156.9, 153.9, 144.3, 144.1, 141.5, 134.4, 130.0, 127.8, 127.2, 125.3, 125.2, 124.2, 120.1, 84.3, 78.4, 66.8, 47.5, 36.2, 29.0, 25.2, 24.9 ppm.

HRMS (ESI, m/z): Calcd for $C_{33}H_{40}BNO_5Na [M + Na]^+ 564.2898$; found 564.2908.

Fmoc-Trp(Boc)-Bpin ((±)-3s)



Route 1: On 0.710 mmol scale, Fmoc-Trp(Boc)-Bpin ((\pm)-**3s**) was obtained as a colorless oil (144.7 mg, 0.238 mmol, 34%).

¹H NMR (300 MHz, CDCl₃) δ 8.13 (d, *J* = 7.9 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 2H), 7.63 – 7.52 (m, 3H), 7.46 (s, 1H), 7.43 – 7.27 (m, 5H), 7.19 (t, *J* = 7.5 Hz, 1H), 5.07 (d, *J* = 5.9 Hz, 1H), 4.50 – 4.29 (m, 2H), 4.22 (t, *J* = 7.3 Hz, 1H), 3.61 – 3.49 (m, 1H), 3.23 – 2.94 (m, 2H), 1.65 (s, 9H), 1.26 – 1.19 (m, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.2, 149.8, 144.2, 144.1, 141.4, 135.6, 131.1, 127.8, 127.2, 125.4, 124.5, 123.7, 122.6, 120.0, 119.5, 118.5, 115.3, 84.4, 83.5, 67.2, 47.4, 28.4, 26.3, 25.0, 24.9 ppm.

HRMS (ESI, m/z): Calcd for C₃₆H₄₁BN₂O₆Na [M + Na]⁺ 631.2956; found 631.2962.

Fmoc-His(Boc)-Bpin ((±)-3t)



Route 1: On 0.819 mmol scale, Fmoc-His(Boc)-Bpin ((\pm)-**3t**) was obtained as a colorless oil (222.0 mg, 0.397 mmol, 48%).

Route 2: On 0.434 mmol scale, Fmoc-His(Boc)-Bpin ((\pm)-**3t**) was obtained as a pale yellow crude material (196.5 mg, containing B₂pin₂ as impurity) that was directly used in the next step.

¹H NMR (300 MHz, CDCl₃) δ 8.02 (s, 1H), 7.75 (d, *J* = 7.5 Hz, 2H), 7.61 – 7.54 (m, 2H), 7.38 (t, *J* = 7.3 Hz, 2H), 7.32 – 7.27 (m, 2H), 7.12 (s, 1H), 5.48 (d, *J* = 5.9 Hz, 1H), 4.41 – 4.27 (m, 2H), 4.26 – 4.19 (m, 1H), 3.45 (q, *J* = 6.0, 5.4 Hz, 1H), 3.02 – 2.79 (m, 2H), 1.59 (s, 9H), 1.28 (s, 12H) ppm.

¹³C NMR (75 MHz, CDCl₃) δ 157.2, 147.1, 144.2, 141.8, 141.4, 136.3, 127.7, 127.1, 125.4, 120.0, 113.9, 85.6, 83.9, 77.2, 67.0, 47.4, 29.2, 28.0, 25.0, 24.9 ppm.

HRMS (ESI, m/z): Calcd for C₃₁H₃₈BN₃O₆Na [M + Na]⁺ 582.2751; found 582.2760.

Synthesis of a-aminoboronic acids



Fmoc-protected α -aminoboronic acids were obtained by modifying our recently published method.⁶ The procedure was optimised for sensitive substrates.

Conditions A: MeCN/0.1 N HCl (1:1, v/v)

The respective Fmoc-protected pinacolyl boronate **3** (1.0 eq) was dissolved in acetonitrile (~10 mL/mmol). Methylboronic acid (5.0 eq) and dilute aqueous HCl (0.1 N, ~10 mL/mmol) were added and the resulting mixture was stirred at ambient temperature overnight. The solution was diluted with acetonitrile and deionised water, and volatiles were removed by freeze-drying to obtain the corresponding Fmoc- α -aminoboronic acid **4**.

Conditions B: MeCN/0.1 N HCl (9:1, v/v)

The respective Fmoc-protected pinacolyl boronate **3** (1.0 eq) was dissolved in acetonitrile (~10 mL/mmol). Methylboronic acid (10.0 eq) and dilute aqueous HCl (0.1 N, ~1 mL/mmol) were added and the resulting mixture was stirred at ambient temperature for two hours. The solution was diluted with acetonitrile and deionised water, and volatiles were removed by freeze-drying to obtain the corresponding Fmoc- α -aminoboronic acid **4**.

Conditions C: MeCN/phosphate buffer pH 7 (1:1, v/v)

The respective Fmoc-protected pinacolyl boronate **3** (1.0 eq) was dissolved in acetonitrile (~10 mL/mmol) and phosphate buffer pH 7.0 (~10 mL/mmol) After the addition of MeB(OH)₂ (10.0 eq), the resulting mixture was stirred at room temperature for two hours and concentrated *in vacuo*. The residue was redissolved in methylene chloride and deionised water, the aqueous phase was extracted twice with methylene chloride. The combined organic phases were washed with brine, dried over anhydrous MgSO₄ and concentrated *in vacuo* to obtain the corresponding Fmoc- α -aminoboronic acid **4**.

NOTE: Most Fmoc-protected α -aminoboronic acids could be stored at -20° C for several weeks without appreciable deterioration. However, it is expected that the compounds are prone to disproportionation and decomposition. Long-term storage should therefore be avoided.

Determination of loading efficiency onto 1-glycerol polystyrene resin

1-Glycerol polystyrene resin (100.0 mg, $B_{max.} = 0.60 \text{ mmol/g}$, 0.06 mmol) was weighed into a 5 mL polypropylene reaction vessel equipped with a polyethylene frit. The respective *N*-Fmoc- α -aminoboronic acid **4** (0.072 mmol, 1.2 eq) was dissolved in methylene chloride (1.5 mL) and added as a solution to the resin. The reaction vessel was shaken on an orbital shaker for two hours, before the resin was washed thoroughly with methylene chloride (9x) to remove unbound material. Cleavage was performed by treating the resin with a solution of DCM/MeOH/H₂O (5:4:1, v/v/v) for 3x30 min and collecting the cleavage fractions. The combined solutions were concentrated under reduced pressure to remove DCM and MeOH. The resulting suspension was diluted with acetonitrile and freeze-dried to constant weight. The loading efficiency is expressed by the molar fraction of freeze-dried compound and the theoretical scale of 0.06 mmol.

a-Aminoboronic acids

Fmoc-Gly-B(OH)₂ (4a)

Fmoc^{-N}, B, OH

On 0.148 mmol scale, conditions A were used to obtain Fmoc-Gly-B(OH)_2 (4a) as a colorless oil (41.3 mg, 0.139 mmol, 94%).

Loading efficiency: 66%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.82 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 7.5 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.36 – 7.30 (m, 2H), 4.30 (d, *J* = 7.0 Hz, 2H), 4.20 (t, *J* = 6.9 Hz, 1H), 2.61 (s, 2H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.6, 145.2, 142.1, 128.7, 128.2, 126.2, 121.0, 67.2, 48.1 ppm.

HRMS (ESI, m/z): Calcd for $C_{16}H_{16}BNO_4Na \ [M + Na]^+ 320.1067$; found 320.1072.

Fmoc-Ala-B(OH)₂ ((±)-4b)



Route 1: Conditions A were used with Fmoc-Ala-Bpin ((\pm)-**3b**) on 0.252 mmol scale to obtain the title compound (\pm)-**4b** as a colorless solid (73.7 mg, 0.237 mmol, 94%).

Route 2: Crude (\pm)-**3b** (254.5 mg, 0.486 mmol based on **2b**) was transesterified using conditions A. After stirring at ambient temperature overnight, volatiles were removed *in vacuo*. The residue was redissolved in methanol and concentrated *in vacuo* twice, before it was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-**4b** (80.1 mg, 0.257 mmol, 53%, 2 steps) as a colorless solid.

Loading efficiency: 81%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.6 Hz, 2H), 7.65 (d, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 4.33 – 4.24 (m, 2H), 4.20 (t, *J* = 6.8 Hz, 1H), 2.93 (q, *J* = 7.4 Hz, 1H), 1.11 (d, *J* = 7.5 Hz, 3H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.1, 145.21, 145.17, 142.1, 128.7, 128.1, 126.2, 121.0, 67.1, 48.1, 17.1 ppm.

HRMS (ESI, m/z): Calcd for $C_{17}H_{18}BNO_4Na \ [M + Na]^+ 334.1224$; found 334.1234.

Fmoc-Val-B(OH)₂ ((±)-4c)



On 0.384 mmol scale, conditions A were used to obtain Fmoc-Val-B(OH)₂ ((\pm)-4c) as a colorless solid (125.1 mg, 0.369 mmol, 96%).

Loading efficiency: 46%

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 7.84 (d, *J* = 7.6 Hz, 2H), 7.69 (d, *J* = 7.4 Hz, 2H), 7.43 (t, *J* = 7.6 Hz, 2H), 7.35 (t, *J* = 7.4 Hz, 2H), 4.42 – 4.18 (m, 3H), 2.82 (d, *J* = 6.3 Hz, 1H), 1.94 – 1.81 (m, 1H), 0.89 (d, *J* = 6.8 Hz, 6H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.4, 145.3, 145.2, 142.1, 128.7, 128.1, 126.2, 121.0, 67.1, 48.2, 30.7, 20.7, 20.3 ppm.

HRMS (ESI, m/z): Calcd for C₂₃H₃₀BN₂O₄ [DEA adduct + H]⁺ 409.2297; found 409.2302.

Fmoc-Leu-B(OH)₂ ((±)-4d)



On 0.104 mmol scale, conditions A were used to obtain Fmoc-Leu-B(OH)₂ ((\pm)-4d) as a colorless solid (33.6 mg, 0.095 mmol, 91%).

Loading efficiency: 60%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, J = 7.5 Hz, 2H), 7.65 (d, J = 7.5 Hz, 2H), 7.40 (t, J = 7.4 Hz, 2H), 7.32 (t, J = 7.4 Hz, 2H), 4.33 – 4.24 (m, 2H), 4.20 (t, J = 7.0 Hz, 1H), 2.95 (dd, J = 9.7, 5.2 Hz, 1H), 1.63 – 1.50 (m, 1H), 1.46 – 1.27 (m, 2H), 0.92 – 0.82 (m, 6H) ppm. ¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.1, 145.3, 145.2, 142.1, 128.7, 128.1, 126.2, 121.0, 67.0, 48.2, 40.9, 26.0, 23.6, 22.2 ppm.

HRMS (ESI, m/z): Calcd for $C_{20}H_{24}BNO_4Na [M + Na]^+ 376.1694$; found 376.1696.

Fmoc-Ile-B(OH)₂ (4e)



On 0.349 mmol scale, conditions A were used to obtain $\text{Fmoc-Ile-B}(\text{OH})_2$ (**4e**) as a colorless solid (115.1 mg, 0.326 mmol, 93%) as a mixture of diastereomers.

Loading efficiency: 64%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.82 (d, *J* = 7.5 Hz, 2H), 7.70 – 7.62 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 4.35 – 4.24 (m, 2H), 4.21 (t, *J* = 6.8 Hz, 1H), 3.02 (d, *J* = 4.9 Hz, 0H), 1.68 – 1.56 (m, 1H), 1.48 – 1.31 (m, 1H), 1.15 – 1.03 (m, 1H), 0.91 – 0.79 (m, 6H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.3, 145.3, 145.2, 142.1, 128.7, 128.1, 126.2, 121.0, 67.1, 48.2, (37.6 and 37.5, diastereomers), (27.9 and 27.4, diastereomers), (17.3 and 17.1, diastereomers), (12.4 and 12.0, diastereomers) ppm.

HRMS (ESI, m/z): Calcd for C₂₀H₂₄BNO₄Na [M + Na]⁺ 376.1694; found 376.1697.
Fmoc-Pro-B(OH)₂ ((±)-4f)



On 0.284 mmol scale, conditions A were used to obtain Fmoc-Pro-B(OH)_2 ((±)-4f) as a colorless solid (93.2 mg, 0.276 mmol, 97%) as a mixture of rotamers.

Loading efficiency: 60%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.84 – 7.78 (m, 2H), 7.72 – 7.63 (m, 2H), 7.43 – 7.36 (m, 2H), 7.36 – 7.29 (m, 2H), 4.38 – 4.18 (m, 3H), 3.46 – 3.33 (m, 1H), 3.34 – 3.23 (m, 1H), 3.15 – 3.10 (m, 0.5H, rotamer), 2.96 – 2.89 (m, 0.5H, rotamer), 2.13 – 1.96 (m, 1H), 1.92 – 1.84 (m, 1H), 1.82 – 1.63 (m, 2H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 156.4, 156.2, 145.5, 145.3, 145.2, 145.0, 142.1, 142.0, 128.7, 128.2, 126.5, 126.3, 126.2, 121.0, 118.7, 68.2, 67.8, 48.2, 48.1, 48.1, 47.3, 29.9, 28.9, 27.3, 26.5 ppm.

HRMS (ESI, m/z): Calcd for $C_{19}H_{20}BNO_4Na [M + Na]^+$ 360.1381; found 360.1392.

Fmoc-Met-B(OH)₂ ((±)-4g)



Route 2: Crude (\pm)-**3g** (350.7 mg, 0.784 mmol based on **2g**) was transesterified using conditions A. After stirring at ambient temperature overnight, volatiles were removed *in vacuo*. The residue was redissolved in methanol and concentrated *in vacuo* twice, before it was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-**4g** (107.8 mg, 0.290 mmol, 37%, 2 steps) as a colorless solid.

Loading efficiency: 43%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.83 (d, *J* = 7.6 Hz, 2H), 7.66 (d, *J* = 7.5 Hz, 2H), 7.41 (t, *J* = 7.3 Hz, 2H), 7.33 (td, *J* = 7.5, 1.1 Hz, 2H), 4.36 – 4.27 (m, 2H), 4.21 (t, *J* = 6.8 Hz, 1H), 3.04 – 2.96 (m, 1H), 2.53 – 2.39 (m, 2H), 2.03 (s, 3H), 1.88 – 1.69 (m, 2H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 145.3, 145.2, 142.1, 128.7, 128.1, 126.2, 121.0, 67.0,
48.2, 32.2, 31.4, 15.3 ppm. (carbamate not observed)

HRMS (ESI, m/z): Calcd for C₁₉H₂₂BNO₄SNa [M + Na]⁺ 394.1258; found 394.1260.

Fmoc-Phe-B(OH)₂ ((±)-4h)



On 0.133 mmol scale, conditions A were used to obtain Fmoc-Phe-B(OH)₂ ((\pm)-**4h**) as a colorless solid (46.9 mg, 0.121 mmol, 91%).

Loading efficiency: 46%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.60 (d, *J* = 7.5 Hz, 2H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.35 - 7.29 (m, 2H), 7.22 (t, *J* = 7.3 Hz, 2H), 7.19 - 7.09 (m, 3H), 4.33 - 4.23 (m, 2H), 4.16 (t, *J* = 6.8 Hz, 1H), 3.16 (dd, *J* = 8.6, 5.4 Hz, 1H), 2.93 - 2.83 (m, 1H), 2.78 - 2.69 (m, 1H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.0, 145.1, 142.1, 141.1, 130.1, 129.2, 128.7, 128.1, 127.0, 126.2, 121.0, 67.0, 48.1, 37.4 ppm.

HRMS (ESI, m/z): Calcd for $C_{23}H_{22}BNO_4Na \ [M + Na]^+ 410.1538$; found 410.1551.

Fmoc-Asn(Trt)-B(OH)₂ ((±)-4i)



On 0.309 mmol scale, conditions A were used to obtain $\text{Fmoc-Asn}(\text{Trt})-B(\text{OH})_2$ ((±)-4i) as a colorless solid (155.1 mg, 0.260 mmol, 84%).

Loading efficiency: 44%

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 7.84 – 7.75 (m, 2H), 7.64 (d, *J* = 7.5 Hz, 2H), 7.38 (t, *J* = 7.7 Hz, 2H), 7.31 – 7.14 (m, 17H), 4.36 – 4.28 (m, 2H), 4.20 (t, *J* = 6.9 Hz, 1H), 3.18 – 3.10 (m, 1H), 2.71 – 2.51 (m, 2H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 173.6, 158.1, 145.6, 145.2, 145.1, 142.1, 129.6, 128.7, 128.7, 128.2, 127.8, 126.2, 121.0, 71.1, 67.2, 48.1, 39.4 ppm.

HRMS (ESI, m/z): Calcd for $C_{37}H_{33}BN_2O_5Na [M + Na]^+ 410.1538$; found 410.1532.

Fmoc-Gln(Trt)-B(OH)₂ ((±)-4j)



On 0.330 mmol scale, conditions A were used to obtain $\text{Fmoc-Gln}(\text{Trt})-B(\text{OH})_2$ ((±)-4j) as a colorless solid (190.8 mg, 0.313 mmol, 95%).

Loading efficiency: 62%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 7.5 Hz, 2H), 7.39 (t, *J* = 7.5 Hz, 2H), 7.31 (t, *J* = 7.3 Hz, 2H), 7.29 – 7.16 (m, 16H), 4.37 – 4.25 (m, 2H), 4.21 (t, *J* = 6.9 Hz, 1H), 2.89 (dd, *J* = 8.9, 5.3 Hz, 1H), 2.39 – 2.19 (m, 2H), 1.85 – 1.72 (m, 1H), 1.72 – 1.59 (m, 1H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 174.3, 158.3, 145.7, 145.2, 142.1, 129.6, 128.7, 128.7, 128.2, 127.7, 126.2, 121.0, 70.8, 67.1, 48.2, 35.1, 28.2 ppm.

HRMS (ESI, m/z): Calcd for $C_{38}H_{35}BN_2O_5Na [M + Na]^+ 633.2538$; found 633.2543.

Fmoc-Asp(tBu)-B(OH)₂ ((±)-4k)



On 0.197 mmol scale, conditions B were used to obtain $\text{Fmoc-Asp}(tBu)-B(OH)_2$ ((±)-4k) as a colorless solid (77.0 mg, 0.187 mmol, 95%).

Loading efficiency: 71%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.62 (d, *J* = 7.5 Hz, 2H), 7.40 (t, *J* = 7.2 Hz, 2H), 7.35 - 7.28 (m, 2H), 4.34 - 4.24 (m, 2H), 4.20 (t, *J* = 6.8 Hz, 1H), 3.06 (dd, *J* = 6.8, 2.5 Hz, 0H), 2.65 - 2.43 (m, 2H), 1.39 (s, 9H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 173.6, 157.9, 145.2, 145.1, 142.1, 128.7, 128.1, 126.2, 121.0, 81.8, 67.2, 48.1, 38.3, 28.3 ppm.

HRMS (ESI, m/z): Calcd for $C_{22}H_{26}BNO_6Na [M + Na]^+ 434.1749$; found 434.1751.

Fmoc-Glu(tBu)-B(OH)₂ ((±)-4l)



On 0.381 mmol scale, conditions B were used to obtain $\text{Fmoc-Glu}(t\text{Bu})-B(OH)_2$ ((±)-4l) as a colorless solid (147.6 mg, 0.347 mmol, 91%).

Loading efficiency: 84%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 7.1 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.5 Hz, 2H), 4.34 – 4.25 (m, 2H), 4.20 (t, *J* = 7.0 Hz, 1H), 2.91 (dd, *J* = 8.3, 5.2 Hz, 1H), 2.22 – 2.16 (m, 2H), 1.87 – 1.76 (m, 1H), 1.73 – 1.62 (m, 1H), 1.40 (s, 9H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 174.4, 158.2, 145.2, 145.2, 142.1, 128.7, 128.1, 126.2, 121.0, 81.1, 67.1, 48.2, 33.8, 28.3, 27.3 ppm.

HRMS (ESI, m/z): Calcd for $C_{23}H_{28}BNO_6Na [M + Na]^+ 448.1906$; found 448.1914.

Fmoc-Ser(**tBu**)-**B**(**OH**)₂((±)-4m)



On 0.399 mmol scale, conditions B were used to obtain $\text{Fmoc-Ser}(tBu)-B(OH)_2$ ((±)-4m) as a colorless solid (145.8 mg, 0.380 mmol, 95%).

Loading efficiency: 67%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.9 Hz, 2H), 4.34 – 4.26 (m, 2H), 4.20 (t, *J* = 6.9 Hz, 1H), 3.52 – 3.42 (m, 2H), 3.05 – 3.00 (m, 1H), 1.12 (s, 9H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.2, 145.2, 145.1, 142.1, 128.7, 128.1, 126.2, 121.0, 118.7, 74.0, 67.2, 63.2, 48.1, 27.7 ppm.

HRMS (ESI, m/z): Calcd for $C_{21}H_{26}BNO_5Na [M + Na]^+ 406.1800$; found 406.1792.

Fmoc-Thr(tBu)-B(OH)₂ (4n)



On 0.617 mmol scale, conditions C were used to obtain $\text{Fmoc-Thr}(tBu)-B(OH)_2$ (**4n**) as a colorless solid (180.4 mg, 0.454 mmol, 74%) as a mixture of diastereomers.

Route 2: Crude **3n** (316.7 mg, 0.471 mmol based on **2n**) was transesterified using conditions C. After stirring at ambient temperature for two hours, volatiles were removed *in vacuo*. The residue was redissolved in methylene chloride and water and the aqueous phase was extracted twice with methylene chloride. The combined organic phases were washed with brine, dried over anhydrous MgSO₄ and concentrated *in vacuo*. The residue was dissolved in water/acetonitrile and lyophilised to obtain compound **4n** (103.5 mg, 0.261 mmol, 55%, 2 steps) as a colorless solid.

Loading efficiency: 66%

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 4.43 – 4.25 (m, 2H), 4.20 (t, *J* = 6.6 Hz, 1H), 3.99 – 3.87 (m, 1H), 3.10 (d, *J* = 5.8 Hz, 1H), 1.18 (s, 9H), 1.02 (d, *J* = 6.2 Hz, 3H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.1, 145.3, 145.0, 142.1, 128.7, 128.1, 126.2, 126.2, 121.0, 75.6, 68.4, 67.1, 48.2, 28.5, 20.4 ppm.

HRMS (ESI, m/z): Calcd for $C_{22}H_{28}BNO_5Na [M + Na]^+ 420.1957$; found 420.1970.

Fmoc-Orn(Boc)-B(OH)₂ ((±)-40)



Route 1: On 0.559 mmol scale, conditions B were used to obtain $\text{Fmoc-Orn(Boc)-B(OH)}_2$ ((±)-40) as a colorless solid (235.0 mg, 0.517 mmol, 93%).

Route 2: Crude compound (\pm)-**30** (475 mg, 0.830 mmol based on **20**) was transesterified using conditions B. After stirring at ambient temperature for two hours, volatiles were removed *in vacuo*. The residue was redissolved in methanol and concentrated *in vacuo* twice, before it was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-**40** (146.8 mg, 0.323 mmol, 39%, 2 steps) as a colorless solid.

Loading efficiency: 84%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.6 Hz, 2H), 7.68 – 7.62 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.35 – 7.29 (m, 2H), 4.34 – 4.23 (m, 2H), 4.23 – 4.17 (m, 1H), 3.02 – 2.87 (m, 3H), 1.68 – 1.26 (m, 13H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.2, 157.5, 145.2, 145.1, 142.1, 128.7, 128.1, 126.2, 121.0, 79.5, 67.1, 48.1, 40.8, 29.1, 28.7, 28.2 ppm.

HRMS (ESI, m/z): Calcd for C₂₄H₃₁BN₂O₆Na [M + Na]⁺ 477.2172; found 477.2177.

Fmoc-Lys(Boc)-B(OH)₂ ((±)-4p)



Route 1: On 0.252 mmol scale, conditions B were used to obtain $\text{Fmoc-Lys(Boc)-B(OH)}_2$ ((±)-**4p**) as a colorless solid (111.9 mg, 0.239 mmol, 95%).

Route 2: Crude (\pm)-**3p** (466 mg, 0.845 mmol based on **2p**) was transesterified using conditions B. After stirring at ambient temperature for two hours, volatiles were removed *in vacuo*. The residue was redissolved in methanol and concentrated *in vacuo* twice, before it was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-**4p** (141.1 mg, 0.301 mmol, 36%, 2 steps) as a colorless solid.

Loading efficiency: 66%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.67 – 7.62 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 2H), 7.32 (t, *J* = 7.9 Hz, 2H), 4.34 – 4.24 (m, 2H), 4.20 (t, *J* = 6.9 Hz, 1H), 3.00 – 2.82 (m, 3H), 1.68 – 1.39 (m, 4H), 1.36 (s, 9H), 1.30 – 1.19 (m, 2H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.2, 157.5, 145.2, 145.1, 142.1, 128.7, 128.2, 126.2, 121.0, 79.4, 67.1, 48.1, 40.8, 31.5, 30.5, 28.7, 24.8 ppm.

HRMS (ESI, m/z): Calcd for C₂₅H₃₃BN₂O₆Na [M + Na]⁺ 491.2328; found 491.2339.

Fmoc-Arg(Boc)₂-B(OH)₂ ((±)-4q)



Route 1: Conditions C were used with Fmoc-Arg(Boc)_2 -Bpin ((±)-**3q**) on 0.250 mmol scale to obtain the title compound (±)-**4q** (84.7 mg, 0.142 mmol, 57%) as a colorless oil.

Route 2: Crude (\pm)-**3q** (243 mg, 0.419 mmol based on **2q**) was transesterified using conditions C. After stirring at ambient temperature for two hours, volatiles were removed *in vacuo*. The residue was redissolved in methylene chloride and water, the aqueous phase was extracted twice with methylene chloride. The combined organic phases were washed with brine, dried over anhydrous MgSO₄ and concentrated *in vacuo*. The residue was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-**4q** (56.3 mg, 0.094 mmol, 23%, 2 steps) as a colorless solid.

Loading efficiency: 54%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.5 Hz, 2H), 7.65 (d, *J* = 6.7 Hz, 2H), 7.39 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 4.35 – 4.25 (m, 2H), 4.20 (t, *J* = 6.9 Hz, 1H), 3.32 – 3.23 (m, 2H), 2.95 – 2.91 (m, 1H), 1.59 – 1.46 (m, 4H), 1.45 (s, 9H), 1.40 (s, 9H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 164.4, 158.2, 156.9, 153.8, 145.2, 145.1, 142.1, 128.7, 128.1, 126.2, 121.0, 84.2, 79.8, 67.1, 48.2, 41.2, 28.9, 28.5, 28.2, 27.4 ppm.

HRMS (ESI, m/z): Calcd for C₃₀H₄₂BN₄O₈ [M + H]⁺ 597.3096; found 597.3100.

Fmoc-Tyr(tBu)-B(OH)₂ ((±)-4r)



On 0.255 mmol scale, conditions B were used to obtain $\text{Fmoc-Tyr}(tBu)-B(OH)_2$ ((±)-4r) as a pale yellow solid (115.8 mg, 0.252 mmol, 99%).

Loading efficiency: 26%

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 7.81 (d, *J* = 7.4 Hz, 2H), 7.61 (d, *J* = 7.2 Hz, 2H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.32 (t, *J* = 7.4 Hz, 2H), 7.01 (d, *J* = 8.3 Hz, 2H), 6.81 (d, *J* = 8.3 Hz, 2H), 4.34 - 4.19 (m, 2H), 4.14 (t, *J* = 6.6 Hz, 1H), 3.13 (dd, *J* = 9.0, 5.5 Hz, 1H), 2.91 - 2.62 (m, 2H), 1.22 (s, 9H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 154.4, 149.3, 145.2, 142.1, 136.1, 130.6, 128.7, 128.1, 126.2, 124.8, 121.0, 79.0, 67.0, 48.1, 36.8, 29.0 ppm.

HRMS (ESI, m/z): Calcd for $C_{27}H_{30}BNO_5Na [M + Na]^+ 482.2114$; found 482.2107.

Fmoc-Trp(Boc)-B(OH)₂ ((±)-4s)



On 0.238 mmol scale, conditions B were used to obtain $\text{Fmoc-Trp}(\text{Boc})-B(\text{OH})_2$ ((±)-4s) as a pale yellow solid (114.5 mg, 0.218 mmol, 91%).

Loading efficiency: 49%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 8.04 (d, *J* = 8.2 Hz, 1H), 7.78 (d, *J* = 7.6 Hz, 2H), 7.59 (d, *J* = 7.8 Hz, 1H), 7.53 (t, *J* = 7.8 Hz, 2H), 7.45 (s, 1H), 7.36 (t, *J* = 7.5 Hz, 2H), 7.31 – 7.16 (m, 4H), 4.26 – 4.18 (m, 2H), 4.13 (t, *J* = 7.1 Hz, 1H), 3.29 (dd, *J* = 8.8, 5.3 Hz, 1H), 3.05 – 2.86 (m, 2H), 1.53 (s, 9H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 158.1, 150.8, 145.1, 142.0, 136.3, 131.9, 128.7, 128.1, 126.2, 126.2, 125.2, 124.5, 123.4, 120.9, 120.3, 119.8, 115.9, 84.6, 67.2, 48.1, 28.3, 26.8 ppm.
HRMS (ESI, m/z): Calcd for C₃₀H₃₁BN₂O₆Na [M + Na]⁺ 549.2173; found 549.2175.

Fmoc-His(Boc)-B(OH)₂ ((±)-4t)



Route 1: On 0.351 mmol scale, conditions C were used to obtain $\text{Fmoc-His(Boc)-B(OH)}_2$ ((±)-4t) as a colorless solid (81.2 mg, 0.170 mmol, 48%).

Route 2: Crude (\pm)-**3t** (196.5 mg, 0.434 mmol based on **2t**) was transesterified using conditions C. After stirring at ambient temperature for two hours, volatiles were removed *in vacuo*. The residue was redissolved in methylene chloride and water, the aqueous phase was extracted twice with methylene chloride. The combined organic phases were washed with brine, dried over anhydrous MgSO₄ and concentrated *in vacuo*. The residue was dissolved in water/acetonitrile and lyophilised to obtain compound (\pm)-4t (62.3 mg, 0.131 mmol, 30%, 2 steps) as a colorless solid.

Loading efficiency: 70%

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 8.05 (s, 1H), 7.79 (d, *J* = 7.1 Hz, 2H), 7.61 – 7.53 (m, 2H), 7.38 (t, *J* = 7.1 Hz, 2H), 7.33 – 7.25 (m, 2H), 7.19 (s, 1H), 4.28 – 4.17 (m, 2H), 4.16 – 4.10 (m, 1H), 3.20 – 3.16 (m, 1H), 2.85 – 2.66 (m, 2H), 1.47 (s, 9H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 158.1, 148.0, 145.1, 142.0, 141.8, 137.9, 128.7, 128.1, 126.2, 126.2, 120.9, 115.3, 86.7, 67.2, 48.0, 30.0, 27.9 ppm.

HRMS (ESI, m/z): Calcd for C₂₅H₂₈BN₃O₆Na [M + Na]⁺ 500.1968; found 500.1959.

Solid-phase synthesis of peptide-boronic acids



General procedure

The standard protocol was elaborated adopting a literature-known method with slight modifications.⁷ The use of THF was generally avoided in this work, because oxidised byproducts, namely *N*-acyl hemiaminals, were observed in some attempts. DMF was stored over molecular sieves (4Å) to avoid premature cleavage from the resin.

A 5 mL PP reaction vessel equipped with a PE frit was charged with 1-glycerol polystyrene resin (1.0 eq, max. loading: B = 0.6 mmol/g). The respective Fmoc-protected α -aminoboronic acid 4 (1.2 eq) was dissolved in DCM (~ 1 mL/50 µmol) and added to the resin. The suspension was continuously shaken at ambient temperature for two hours and the resin was washed (6× DCM, 6× DMF). Subsequently, the desired sequence was assembled by alternating Fmoc deprotection and amide coupling steps.

Fmoc deprotection was achieved by treating the resin with 10% piperidine solution in DMF for 10 min + 5 min. Afterwards, the resin was washed thoroughly (6× DMF, 3× DCM, 3× DMF). For amide coupling, the corresponding carboxylic acid (3.0 eq), HATU (3.0 eq) and DIPEA (4.0 eq) were dissolved in DMF (1 mL) and added to the reaction vessel. After shaking for two hours, the resin was washed (3× DMF, 3× DCM, 3× DMF).

NOTE: When residues were coupled that are prone to racemisation, e. g. phenylglycine derivatives, COMU and TMP were used as coupling reagents instead of HATU and DIPEA to avoid epimer formation, as it was described recently.⁸

After the final coupling step, the resin was washed ($6 \times DMF$, $9 \times DCM$). Cleavage from solid support was achieved under complete conservation of side-chain protecting groups using

DCM/MeOH/H₂O (5:4:1, v/v) for 3×30 min. The combined cleavage solutions were collected, DCM and MeOH were removed under reduced pressure. The resulting suspension was diluted with MeCN and lyophilised. Final compounds were obtained as diastereomers if more than one stereocentre was present.

NOTE: The loading step could also be performed overnight. The synthesis of longer sequences could be paused overnight. In that case, the resin was washed ($3 \times DMF$, $6 \times DCM$) after amide coupling and dried in a desiccator overnight. The resin was swollen in DCM for 30 min, washed ($3 \times DMF$) and the synthesis was continued with Fmoc deprotection.

Peptide-boronic acids

Bortezomib (5a)



On 0.12 mmol scale, bortezomib (**5a**) was obtained as a colorless solid after HPLC purification (16.1 mg, 0.042 mmol, 35%) as a mixture of diastereomers.

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 9.17 (s, 1H), 8.76 (s, 1H), 8.60 (s, 1H), 7.32 – 7.14 (m, 5H), 4.82 – 4.68 (m, 1H), 3.31 – 3.01 (m, 3H), 1.52 – 1.13 (m, 3H), 0.87 – 0.74 (m, 6H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 171.9, 164.2, 148.8, 145.1, 144.6, 144.4, 137.8, 130.4, 129.4, 127.8, (55.3 and 54.9, diastereomers), (40.3 and 40.0, diastereomers), (38.9 and 38.7, diastereomers), (26.0 and 25.9, diastereomers), 23.6, (22.0 and 21.9, diastereomers).

HRMS (ESI, m/z): Calcd for C₁₉H₂₅BN₄O₄Na [M + Na]⁺ 407.1864; found 407.1870.

Ixazomib ((±)-5b)



On 0.09 mmol scale, ixazomib ((\pm)-**5b**) was obtained after HPLC purification as a colorless solid (18.15 mg, 0.050 mmol, 56%).

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.55 (s, 1H), 7.45 (s, 2H), 3.94 (s, 2H), 3.06 (dd, *J* = 9.6, 5.4 Hz, 1H), 1.66 – 1.54 (m, 1H), 1.50 – 1.40 (m, 1H), 1.38 – 1.30 (m, 1H), 0.91 – 0.82 (m, 6H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 170.3, 167.4, 137.8, 133.5, 132.5, 132.2, 130.2, 130.0, 43.5, 40.4, 26.1, 23.6, 22.1 ppm.

HRMS (ESI, m/z): Calcd for C₁₄H₁₉BCl₂N₂O₄Na [M + Na]⁺ 383.0710; found 383.0712.

Bz-Ala-Ile-B(OH)₂ (5c)



On 0.06 mmol scale, Bz-Ala-Ile-B(OH)₂ (5c) was obtained as a colorless solid after lyophilisation (7.23 mg, 0.024 mmol, 40%) as a mixture of diastereomers.

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.82 (d, *J* = 8.2 Hz, 2H), 7.54 (t, *J* = 7.4 Hz, 1H), 7.46 (t, *J* = 7.0 Hz, 2H), 4.54 – 4.46 (m, 1H), 3.13 (d, *J* = 5.4 Hz, 0.5H, diastereomer), 2.86 (d, *J* = 7.1 Hz, 0.5H, diastereomer), 1.68 – 1.57 (m, 1H), 1.40 (d, *J* = 7.1 Hz, 3H), 1.38 – 1.28 (m, 1H), 1.15 – 0.99 (m, 1H), 0.88 – 0.79 (m, 6H) ppm.

 13 C NMR (126 MHz, CD₃CN/D₂O 9:1) δ (174.80 and 174.78, diastereomers), (169.0 and 168.9, diastereomers), (134.7 and 134.6, diastereomers), (132.84 and 132.83, diastereomers), 129.5, 128.3, (50.6 and 50.4, diastereomers), (36.9 and 36.7, diastereomers), (27.9 and 27.4, diastereomers), (18.3 and 17.9, diastereomers), (17.20 and 17.16, diastereomers), (12.3 and 11.8, diastereomers) ppm.

HRMS (ESI, m/z): Calcd for $C_{15}H_{23}BN_2O_4Na [M + Na]^+ 329.1646$; found 329.1670.

Bz-Asp(tBu)-Ile-B(OH)₂ (5d)



On 0.06 mmol scale, Bz-Asp(tBu)-Ile-B(OH)₂ (**5d**) was obtained as a colorless solid after lyophilisation (11.20 mg, 0.028 mmol, 47%) as a mixture of diastereomers.

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.84 – 7.75 (m, 2H), 7.55 (t, *J* = 7.1 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 2H), 4.89 – 4.80 (m, 1H), 3.14 (d, *J* = 5.2 Hz, 0.5H, diastereomer), 2.98 (d, *J* = 6.7 Hz, 0.5H, diastereomer), 2.87 – 2.64 (m, 2H), 1.67 – 1.55 (m, 1H), 1.39 – 1.36 (m, 9H), 1.35 – 1.28 (m, 1H), 1.13 – 0.99 (m, 1H), 0.86 – 0.78 (m, 6H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 172.1, 171.4, 168.9, 134.5, 133.0, 129.6, 128.3, 82.3, (51.5 and 51.3, diastereomers), (38.3 and 37.8, diastereomers), (36.94 and 36.86, diastereomers), 28.2, (27.9 and 27.3, diastereomers), (17.3 and 17.1, diastereomers), (12.3 and 11.9, diastereomers) ppm.

HRMS (ESI, m/z): Calcd for C₂₀H₃₁BN₂O₆Na [M + Na]⁺ 429.2171; found 429.2192.

Bz-Asp-Ile-B(OH)₂ (5e)



To a solution of **5d** in methylene chloride (3 mL), 2 mLtrifluoroacetic acid were added dropwise at 0°C under constant stirring and the solution was allowed to warm to room temperature. Upon completion, the solution was coevaporated with acetonitrile and the remaining residue was dissolved in acetonitrile/water and lyophilised. The crude product (9.39 mg) was purified by HPLC to obtain Bz-Asp-Ile-B(OH)₂ (**5e**) as a colorless solid (3.29 mg, 0.009 mmol, 34%).

¹H NMR (500 MHz, CD₃CN/D₂O 9:1) δ 7.81 – 7.75 (m, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.46 (t, *J* = 7.6 Hz, 2H), 4.92 – 4.81 (m, 1H), 3.07 (d, *J* = 5.5 Hz, 0.5H, diastereomer), 2.96 – 2.86 (m, 1.5H, diastereomer), 2.84 – 2.75 (m, 1H), 1.65 – 1.54 (m, 1H), 1.43 – 1.25 (m, 1H), 1.12 – 0.98 (m, 1H), 0.88 – 0.73 (m, 6H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 9:1) δ 174.2, 173.9, 172.6, 169.4, 134.3, 133.1, 129.6, (128.35 and 128.33, diastereomers), 51.2, 50.9, 36.7, 36.7, 36.6, 36.2, 27.9, 27.3, 17.2, 17.1, 12.2, 11.8 ppm.

HRMS (ESI, m/z): Calcd for $C_{16}H_{23}BN_2O_6Na [M + Na]^+ 373.1544$; found 373.1542.

Bz(2,5-di-chloro)-Ser(tBu)-Gln(Trt)-B(OH)2 (5f)



On 0.06 mmol scale, **5f** was obtained after lyophilisation as a colorless solid (19.73 mg, 0.028 mmol, 47%).

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 7.56 – 7.50 (m, 1H), 7.46 – 7.40 (m, 2H), 7.30 – 7.14 (m, 15H), 4.51 (q, *J* = 4.7 Hz, 1H), 3.79 – 3.70 (m, 1H), 3.67 – 3.56 (m, 1H), 3.11 – 2.96 (m, 1H), 2.45 – 2.19 (m, 2H), 1.89 – 1.63 (m, 2H), 1.15 (s, 9H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ (174.25 and 174.20, diastereomers), (171.33 and 171.25, diastereomers), (166.88 and 166.86, diastereomers), 145.8, 137.7, 133.5, 132.5, 132.2, 130.2, 130.1, 129.6, 128.6, 127.7, (74.8 and 74.7, diastereomers), 70.8, 62.3, (55.4 and 55.3, diastereomers) 35.3, (27.9 and 27.8, diastereomers) 27.6 ppm.

HRMS (ESI, m/z): Calcd for $C_{37}H_{40}BCl_2N_3O_6Na [M + Na]^+$ 726.2286; found 726.2292.

Pyrazinoyl-Ser(tBu)-Glu(tBu)-B(OH)₂ (5g)



On 0.06 mmol scale, **5g** was obtained after lyophilisation as a colorless solid (16.59 mg, 0.037 mmol, 61%).

¹H NMR (300 MHz, CD₃CN/D₂O 9:1) δ 9.26 – 9.21 (m, 1H), 8.79 (d, *J* = 2.5 Hz, 1H), 8.68 – 8.62 (m, 1H), 4.58 – 4.49 (m, 1H), 3.87 – 3.77 (m, 1H), 3.68 – 3.56 (m, 1H), 3.11 – 2.94 (m, 1H), 2.34 – 2.10 (m, 2H), 1.88 – 1.63 (m, 2H), 1.43 – 1.31 (m, 9H), 1.17 (s, 9H) ppm.

¹³C NMR (75 MHz, CD₃CN/D₂O 9:1) δ 174.4, 171.3, 164.4, 148.9, 145.1, 144.7, 144.5, 81.0, 74.9, 62.6, 54.4, 33.9, 28.3, 27.6, 26.8, 26.6 ppm.

HRMS (ESI, m/z): Calcd for C₂₀H₃₃BN₄O₇Na [M + Na]⁺ 475.2338; found 475.2347.

Pyrazinoyl-Ser-Glu-B(OH)₂ (5h)



On 0.037 mmol scale, the compound was synthesised from 5g by dissolving it in a solution of TFA in DCM (40%, v/v) and stirring at room temperature for 2 hours. After HPLC purification and lyophilisation, 5h was obtained as a pale yellow solid as a mixture of anhydrides and diastereomers (9.14 mg, 0.027 mmol, 73%).

¹H NMR (500 MHz, CD₃OD/D₂O 9:1) δ 9.31 (s, 1H), 8.87 (s, 1H), 8.75 (s, 1H), 4.95 – 4.89 (m, 1H), 4.18 – 3.83 (m, 2H), 2.98 – 2.88 (m, 0.5H, diastereomer), 2.65 (q, *J* = 7.2 Hz, 0.5H, diastereomer), 2.44 – 2.37 (m, 1H), 2.36 – 2.23 (m, 1H), 1.89 – 1.67 (m, 2H) ppm.

¹³C NMR (126 MHz, CD₃OD/D₂O 9:1) δ 177.0, 175.9, 165.8, 165.5, 149.0, 145.7, 144.9, 62.3 62.2, 61.8, 61.7, 53.8, 53.6, 53.5, 32.6, 29.1, 29.1, 27.0, 25.5 ppm.

HRMS (ESI, m/z): Calcd for $C_{12}H_{15}BN_4O_6Na [M + Na]^+ 345.0990$; found 345.0970.

Bz-Phg-Lys(Boc)-B(OH)₂ (5i)



On 0.06 mmol scale, Bz-Phg-Lys(Boc)-B(OH)₂ (**5i**) was obtained as a colorless solid (13.67 mg, 0.028 mmol, 47%) that was directly used in the next step.

HRMS (ESI, m/z): Calcd for C₂₅H₃₄BN₃O₆Na [M + Na]⁺ 506.2437; found 506.2458.

Bz-Phg-Lys-B(OH)₂ \times TFA (5j)



Compound **5i** (12.50 mg, 0.026 mmol, 1.0 eq) was stirred in a solution of acetonitrile/TFA/H₂O (7:2:1, v/v, 2 mL) at ambient temperature for 2 hours. After HPLC purification and lyophilisation, Bz-Phg-Lys-B(OH)₂ × TFA (**5j**) was obtained as a colorless solid (5.72 mg, 0.015 mmol, 57%) as a mixture of diastereomers.

¹H NMR (500 MHz, CD₃CN/D₂O 5:1) δ 7.83 (d, *J* = 7.2 Hz, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.52 – 7.43 (m, 4H), 7.42 – 7.32 (m, 3H), 5.59 – 5.53 (m, 1H), 2.98 (dd, *J* = 8.6, 5.9 Hz, 1H), 2.86 – 2.69 (m, 2H), 1.63 – 1.38 (m, 4H), 1.32 – 1.06 (m, 2H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 5:1) δ (172.1 and 172.0, diastereomers), 168.8 and 168.7, diastereomers), (138.5 and 138.3, diastereomers), 134.4, 133.0, 129.8, 129.6, 129.4, 128.8, 128.7, (128.5 and 128.4, diastereomers), (58.73 and 58.65, diastereomers), (40.3 and 40.2, diastereomers), (30.55 and 30.52, diastereomers), (27.3 and 27.2, diastereomers), (24.3 and 24.1, diastereomers) ppm.

HRMS (ESI, m/z): Calcd for $C_{20}H_{25}BN_3O_3$ [M – H₂O + H]⁺ 366.1987; found 366.1989.

Bz-Phe(4-NHBoc)-Lys(Boc)-B(OH)₂ (5k)



On 0.06 mmol scale, Bz-Phe(4-NHBoc)-Lys(Boc)-B(OH)₂ (**5**k) was obtained as a colorless solid (21.32 mg, 0.035 mmol, 58%) that was directly used in the next step.

HRMS (ESI, m/z): Calcd for C₂₅H₃₄BN₃O₆Na [M + Na]⁺ 635.3228; found 635.3238.

Bz-Phe(4-NH₂)-Lys-B(OH)₂ \times 2 TFA (5l)



Compound **5k** (20.50 mg, 0.033 mmol, 1.0 eq) was stirred in a solution of acetonitrile/TFA/H₂O (7:2:1, v/v, 2 mL) at ambient temperature for 2 hours. After HPLC purification and lyophilisation, Bz-Phe(4-NH₂)-Lys-B(OH)₂ × 2 TFA (**5l**) was obtained as a pale yellow solid (12.96 mg, 0.020 mmol, 61%) as a mixture of diastereomers.

¹H NMR (500 MHz, CD₃CN/D₂O 2:1) δ 7.72 (d, *J* = 7.3 Hz, 2H), 7.57 – 7.49 (m, 1H), 7.44 (t, *J* = 7.8 Hz, 2H), 7.41 – 7.33 (m, 2H), 7.28 (d, *J* = 6.3 Hz, 2H), 4.81 – 4.66 (m, 1H), 3.30 – 2.67 (m, 5H), 1.63 – 1.33 (m, 3H), 1.33 – 0.88 (m, 3H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 2:1) δ (173.5 and 173.4, diastereomers), (169.5 and 169.4, diastereomers), 139.1, 134.3, 133.1, 131.9, 130.1, 129.6, 128.4, 124.1, (55.5 and 55.3, diastereomers) 40.2, 37.7, (30.5 and 30.3, diastereomers), 27.3, 24.3 ppm.

HRMS (ESI, m/z): Calcd for $C_{21}H_{28}BN_4O_3$ [M – H₂O + H]⁺ 395.2253; found 395.2258.

Bz(4-Ph)-Aib-Tyr(tBu)-B(OH)₂ (5m)



On 0.06 mmol scale, Bz(4-Ph)-Aib-Tyr(tBu)-B(OH)₂ (**5m**) was obtained as colorless solid (16.6 mg, 0.033 mmol, 55%) that was directly used in the next step.

HRMS (ESI, m/z): Calcd for C₂₉H₃₅BN₂O₅Na [M + Na]⁺ 525.2536; found 525.2531.

Bz(4-Ph)-Aib-Tyr-B(OH)₂ (5n)



Compound **5m** (16.5 mg, 0.033 mmol, 1.0 eq) was stirred in a solution of acetonitrile/TFA/H₂O (7:2:1, v/v, 2 mL) at ambient temperature for 2 hours. After HPLC purification and lyophilisation, Bz(4-Ph)-Aib-Tyr-B(OH)₂ (**5n**) was obtained as a colorless solid (5.52 mg, 0.012 mmol, 38%) containing small fractions of anhydrides.

¹H NMR (500 MHz, CD₃CN/D₂O 3:1) δ 7.86 (d, *J* = 8.4 Hz, 2H), 7.73 (d, *J* = 8.5 Hz, 2H), 7.69 (d, *J* = 7.3 Hz, 2H), 7.48 (t, *J* = 7.6 Hz, 2H), 7.40 (t, *J* = 7.3 Hz, 1H), 6.96 (d, *J* = 8.4 Hz, 2H), 6.58 (d, *J* = 8.4 Hz, 2H), 2.92 (dd, *J* = 10.4, 4.8 Hz, 1H), 2.75 (dd, *J* = 14.1, 4.9 Hz, 1H), 2.58 (dd, *J* = 13.9, 10.5 Hz, 1H), 1.47 (s, 3H), 1.43 (s, 3H) ppm.

¹³C NMR (126 MHz, CD₃CN/D₂O 3:1) δ 178.0, 168.8, 155.8, 145.2, 140.6, 133.4, 132.5, 131.0, 130.1, 129.22, 129.20, 128.1, 127.9, 115.9, 56.7, 36.1, 25.7, 25.0 ppm (Aib carbonyl at 178.0 ppm identified *via* HMBC).

HRMS (ESI, m/z): Calcd for $C_{25}H_{26}BN_2O_4$ [M – H₂O + H]⁺ 429.1985; found 429.1984.

NMR spectra of NHPI esters


































































































NMR spectra of α-aminopinacolyl boronates















































































NMR spectra of α -aminoboronic acids
















































































NMR spectra of peptide-boronic acids













































References

- J. Cornella, J. T. Edwards, T. Qin, S. Kawamura, J. Wang, C.-M. Pan, R. Gianatassio, M. Schmidt, M. D. Eastgate and P. S. Baran, *Journal of the American Chemical Society*, 2016, **138**, 2174-2177.
- 2. J. Ma, J. Lin, L. Zhao, K. Harms, M. Marsch, X. Xie and E. Meggers, *Angewandte Chemie International Edition*, 2018, **57**, 11193-11197.
- J. Wang, H. Lundberg, S. Asai, P. Martín-Acosta, J. S. Chen, S. Brown, W. Farrell, R.
 G. Dushin, C. J. O'Donnell, A. S. Ratnayake, P. Richardson, Z. Liu, T. Qin, D. G.
 Blackmond and P. S. Baran, *Proceedings of the National Academy of Sciences*, 2018, 115, E6404-E6410.
- C. Li, J. Wang, L. M. Barton, S. Yu, M. Tian, D. S. Peters, M. Kumar, A. W. Yu, K. A. Johnson, A. K. Chatterjee, M. Yan and P. S. Baran, *Science (New York)*, 2017, 356, eaam7355.
- A. Ozanne, L. Pouységu, D. Depernet, B. François and S. Quideau, *Organic Letters*, 2003, 5, 2903-2906.
- 6. S. P. A. Hinkes and C. D. P. Klein, Organic Letters, 2019, 21, 3048-3052.
- M. A. M. Behnam, T. R. Sundermann and C. D. Klein, *Organic Letters*, 2016, 18, 2016-2019.
- 8. C. Liang, M. A. M. Behnam, T. R. Sundermann and C. D. Klein, *Tetrahedron Letters*, 2017, **58**, 2325-2329.