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# Site-to-Site Peptide Transport on a Molecular Platform Using a Small-Molecule Robotic Arm

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## **1. General Experimental**

Unless stated otherwise, reagents were obtained from commercial sources and used without purification. Reactions were carried out in anhydrous solvents and under an  $N_2$  atmosphere. Anhydrous solvents were obtained by passing the solvent through an activated alumina column on a Phoenix SDS (solvent drying system; JC Meyer Solvent Systems, CA, USA). EZ-1 and EZ-2 were synthesized according to our previously reported procedure.<sup>[1]</sup> Pivaloylphenylalanine and Pivaloylalanine were prepared according to known literature procedures.<sup>[2]</sup> <sup>1</sup>H NMR spectra were recorded on a Bruker Avance III instrument with an Oxford AS600 magnet equipped with a cryoprobe [5mm CPDCH <sup>13</sup>C-<sup>1</sup>H/D] (600 MHz). Chemical shifts are reported in parts per million (ppm) from high to low frequency using the residual solvent peak as the internal reference ( $CDCI_3 = 7.26$  ppm,  $C_2D_2CI_4$ = 6.00 ppm, CD<sub>2</sub>Cl<sub>2</sub> = 5.32 ppm, CD<sub>3</sub>CN = 1.94 ppm, and (CD<sub>3</sub>)<sub>2</sub>SO = 2.50 ppm).<sup>[3,4]</sup> All <sup>1</sup>H resonances are reported to the nearest 0.01 ppm. The multiplicity of  $^{1}$ H signals are indicated as: s = singlet; d = doublet; t = triplet; q = quartet; quint = quintet; sept = septet; m = multiplet; br = broad; or combinations of thereof. Coupling constants (J) are quoted in Hz and reported to the nearest 0.1 Hz. Where appropriate, averages of the signals from peaks displaying multiplicity were used to calculate the value of the coupling constant. <sup>13</sup>C NMR spectra were recorded on the same spectrometer with the central resonance of the solvent peak as the internal reference (CDCl<sub>3</sub> = 77.16 ppm, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub> = 73.00 ppm, CD<sub>2</sub>Cl<sub>2</sub> = 54.00 ppm, CD<sub>3</sub>CN = 118.26 ppm and (CD<sub>3</sub>)<sub>2</sub>SO = 39.52 ppm).<sup>[3,4]</sup> All <sup>13</sup>C resonances are reported to the nearest 0.1 ppm, except in cases to aid the differentiation of closely resolved signals (which are reported to the nearest 0.01 ppm). DEPT, COSY, HSQC and HMBC experiments were used to aid structural determination and spectral assignment. Where necessary, 1D NOESY and 2D NOESY or ROESY spectra were used to aid the assignment of <sup>1</sup>H spectra. *Cis* and *trans* acyl hydrazone rotamers where assigned according to the characteristic <sup>13</sup>C shift of the amide carbon (with a lower field shift for the cis conformer). Fully characterized compounds were chromatographically homogeneous. Flash column chromatography was carried out using Silica 60 Å (particle size 40–63 μm, Sigma Aldrich, UK) as the stationary phase. Preparative TLC was performed using either PLC 20×20 cm, 60 F<sub>254</sub> Preparative plates (Merck) or Silica Gel GF 20  $\times$  20 cm, U<sub>254</sub> Preparative plates (Analtech) of various thicknesses. TLC was performed on precoated silica gel plates (0.25 mm thick, 60 F<sub>254</sub>, Merck, Germany) and visualized using both short and long wave ultraviolet light in combination with standard laboratory stains (basic potassium permanganate, acidic ammonium molybdate and ninhydrin). Low resolution ESI mass spectrometry was performed with a Thermo Scientific LCQ Fleet Ion Trap Mass Spectrometer or an Agilent Technologies 1200 LC system with either an Agilent 6130 single quadrupole MS detector or an Advion Expression LCMS single quadrupole MS detector. High-resolution mass spectrometry was carried out by the EPSRC National Mass Spectrometry Service Centre (Swansea, UK) or by staff at the Mass Spectrometry Service, School of Chemistry, The University of Manchester. Melting points (M.p.) were determined using a Büchi M-565 apparatus and are uncorrected.

## 2. Experimental Procedures

## 2.1 Synthesis of EZ-9

## 2.1.1 Synthetic Scheme



**Scheme 1.** Synthesis of *Z*-**9** and *E*-**9**. Reagents and conditions: (i) Ph<sub>3</sub>COH, TFA, r.t., 75 min, 82%. (ii) PivCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, -10 °C to r.t., 1.5 h, 45%. (iii) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O, MeOH, Δ, 67 h, 89%. (iv) *p*-anisaldehyde, AcOH, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 24 h, 95%. (v) AgNO<sub>3</sub>, CHCl<sub>3</sub>, MeOH, r.t., 30 mins, then *EZ*-**2** in CHCl<sub>3</sub>, r.t., 3 h, 79%. (vi) TFA, CHCl<sub>3</sub>, r.t., 17 h, 53%. (vii) TFA, CHCl<sub>3</sub>, r.t., 41 h, 64%.

#### 2.1.2 Synthetic Procedures and Characterisation Data

#### Synthesis of **S1**



*L*-Cysteine ethyl ester hydrochloride (7.45 g, 40.1 mmol, 1.1 eq.) and triphenylmethanol (9.50 g, 36.5 mmol, 1 eq.) were dissolved in TFA (25 mL). The orange solution was stirred at room temperature for 75 minutes and the solvent was removed under reduced pressure. The resulting brown oil was dissolved in  $CH_2Cl_2$  (200 mL) and water (200 mL) was added upon which the brown organics turned

colourless. The mixture was neutralised by careful portion-wise addition of K<sub>2</sub>CO<sub>3</sub> (*ca.* 15 g). The organic layer was separated, dried over MgSO<sub>4</sub> and concentrated under reduced pressure to a light purple oil. The crude oil was triturated with *n*-hexane which was then decanted to give the title compound as a yellow oil (12.9 g, 33.0 mmol, 82%). <sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.37 – 7.29 (m, 12H, H<sub>6</sub>+ H<sub>7</sub>), 7.27 – 7.21 (m, 3H, H<sub>8</sub>), 4.02 (q, *J* = 7.1 Hz, 2H, H<sub>3</sub>), 3.14 (t, *J* = 6.4 Hz, 1H, H<sub>2</sub>), 2.36 (dd, *J* = 11.9, 6.2 Hz, 1H, H<sub>5</sub>), 2.28 (dd, *J* = 11.9, 6.7 Hz, 1H, H<sub>5</sub>'), 1.82 (s, 2H, H<sub>1</sub>), 1.13 (t, *J* = 7.1 Hz, 3H, H<sub>4</sub>). <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  173.7, 144.4, 129.1, 128.0, 126.7, 65.9, 60.2, 53.7, 36.4, 14.1. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>24</sub>H<sub>26</sub>N<sub>1</sub>O<sub>2</sub>S: 392.1679, found 392.1673 [M+H]<sup>+</sup>. [*a*]<sup>20</sup><sub>D</sub> +80.0 (*c* = 1.00, MeOH).

#### Synthesis of S2



Pivaloyl chloride (130  $\mu$ L, 1.06 mmol) was added dropwise to a solution of **S1** (338 mg, 0.862 mmol) and triethylamine (130  $\mu$ L, 1.51 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (3 mL) at -10 °C under nitrogen atmosphere. The reaction mixture was warmed to room temperature and stirred for 1.5 hours. The reaction was quenched with H<sub>2</sub>O and the layers separated. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the

combined organic layers were dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. Flash column chromatography (SiO<sub>2</sub>, EtOAc/hexane, 20%  $\rightarrow$  70%) afforded **S2** as a colourless oil (184 mg, 0.387 mmol, 45%). <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.38 (d, *J* = 7.3 Hz, 6H, H<sub>7</sub>), 7.28 (t, *J* = 7.3 Hz, 6H, H<sub>8</sub>), 7.21 (t, *J* = 7.3 Hz, 3H, H<sub>9</sub>), 6.29 (d, *J* = 7.6 Hz, 1H, H<sub>2</sub>), 4.59 (dt, *J* = 7.6, 5.2 Hz, 1H, H<sub>3</sub>), 4.17 (q, *J* = 7.1 Hz, 2H, H<sub>4</sub>), 2.62 – 2.56 (m, 2H, H<sub>6</sub>), 1.24 (t, *J* = 7.1 Hz, 3H, H<sub>7</sub>), 1.21 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  178.0, 170.7, 144.3, 129.4, 128.0, 126.8, 66.5, 61.7, 50.9, 38.7, 34.0, 27.4, 14.1. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>29</sub>H<sub>33</sub>NO<sub>3</sub>SNa: 498.2073, found 498.2070 [M+Na]<sup>+</sup>.

#### Synthesis of S3



Hydrazine hydrate (45.0  $\mu$ L, 0.926 mmol) was added to a solution of **S2** (153 mg, 0.322 mmol) in MeOH (2 mL) and the reaction was stirred at reflux for 67 hours. The crude mixture was concentrated under reduced pressure and purified by flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 0%  $\rightarrow$  4%) to afford **S3** as

colourless solid (133 mg, 0.288 mmol, 89%). **M.p.** 88 °C. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.42 (d, *J* = 7.2 Hz, 6H, H<sub>7</sub>), 7.30 (t, *J* = 7.2 Hz, 6H, H<sub>8</sub>), 7.23 (t, *J* = 7.2 Hz, 3H, H<sub>9</sub>), 7.18 (br, s, 1H, H<sub>4</sub>), 6.03 (d, *J* = 7.3 Hz, 1H, H<sub>2</sub>), 3.98 (q, *J* = 6.9 Hz, 1H, H<sub>3</sub>), 3.77 (br, s, 2H, H<sub>5</sub>), 2.68 – 2.53 (m, 2H, H<sub>6</sub>), 1.15 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  178.7, 170.9, 144.2, 129.5, 128.1, 126.9, 67.1, 50.7, 38.7, 33.4, 27.4. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>27</sub>H<sub>31</sub>N<sub>3</sub>O<sub>2</sub>SNa: 484.2029, found 484.2015 [M+Na]<sup>+</sup>.

#### Synthesis of 3



*p*-Anisaldehyde (49 µL, 0.400 mmol) was added to a solution of **S3** (92.3 mg, 0.200 mmol) and acetic acid (2 drops) in  $CH_2CI_2$  (1.2 mL). The reaction mixture was stirred at room temperature for 24 hours. The crude mixture was concentrated under reduced pressure and purified by flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>CI<sub>2</sub>, 0%  $\rightarrow$  2%) to afford **3** as a pale yellow solid (110 mg, 0.190 mmol, 95%). **M.p.** 105 °C. <sup>1</sup>**H NMR** (600 MHz,

CDCl<sub>3</sub>) ~3:2 *cis:trans* hydrazone amide rotamers. *Trans*-**3**  $\delta$  9.58 (s, 1H, H<sub>4</sub>), 7.93 (s, 1H, H<sub>5</sub>), 7.63 (d, *J* = 8.8 Hz, 2H, H<sub>6</sub>), 7.43 (d, *J* = 7.1 Hz, 6H, H<sub>10</sub>), 7.28 (t, *J* = 7.1 Hz, 6H, H<sub>11</sub>), 7.21 (t, *J* = 7.1 Hz, 3H, H<sub>12</sub>), 6.87 (d, *J* = 8.8 Hz, 2H, H<sub>7</sub>), 6.16 (d, *J* = 7.5 Hz, 1H, H<sub>2</sub>), 4.05 – 4.01 (m, 1H, H<sub>3</sub>), 3.82 (s, 3H, H<sub>8</sub>), 2.74 – 2.65 (m, 2H, H<sub>9</sub>), 1.17 (s, 9H, H<sub>1</sub>). *Cis*-**3**  $\delta$  8.66 (s, 1H, H<sub>4</sub>), 7.55 (s, 1H, H<sub>5</sub>), 7.45 (d, *J* = 8.8 Hz, 2H, H<sub>6</sub>), 7.35 (d, *J* = 7.4 Hz, 6H, H<sub>10</sub>), 7.15 (t, *J* = 7.4 Hz, 6H, H<sub>11</sub>), 7.09 (t, *J* = 7.3 Hz, 3H, H<sub>12</sub>), 6.90 (d, *J* = 8.8 Hz, 2H, H<sub>7</sub>), 6.72 (d, *J* = 7.9 Hz, 1H, H<sub>2</sub>), 5.49 – 5.46 (m, 1H, H<sub>3</sub>), 3.87 (s, 3H, H<sub>8</sub>), 2.93 – 2.90 (m, 1H, H<sub>9</sub>), 2.56 – 2.53 (m, 1H, H<sub>9</sub>'), 1.27 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  179.6, 178.0, 171.1, 166.7, 161.7, 161.6, 148.7, 144.6, 144.5, 129.7, 129.6, 129.2, 128.2, 128.0, 127.0, 126.8, 126.2, 125.9, 114.3, 114.2, 67.3, 66.4, 55.6, 55.5, 51.7, 49.5, 39.0, 38.9, 34.0, 32.9, 27.7, 27.5. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>35</sub>H<sub>37</sub>N<sub>3</sub>O<sub>3</sub>SNa: 602.2448, found 602.2418 [M+Na]<sup>+</sup>.

Synthesis of EZ-6



AgNO<sub>3</sub> (2.3 mg, 14  $\mu$ mol) was added to a solution of **3** (6.5 mg, 11  $\mu$ mol) in a 2:1 CHCl<sub>3</sub>:MeOH mixture (300  $\mu$ L), the mixture was stirred at room temperature for 30 minutes. A solution of *EZ*-**2** (18.3 mg, 19  $\mu$ mol) in CHCl<sub>3</sub> (50  $\mu$ L) was added and the mixture was stirred at room temperature for 3 hours. The reaction mixture was

diluted with CHCl<sub>3</sub>, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.2% Et<sub>3</sub>N), 10%) afforded *EZ*-**6** 

as an orange solid (10.6 mg, 8.75  $\mu$ mol, 79%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~62:38 *E:Z* pyridyl-hydrazone isomers and mixture of acyl-hydrazone rotamers. Major isomer reported (*E*-**7**)  $\delta$  15.82 (s, 1H, H<sub>22</sub>), 10.16 – 10.14 (m, 1H, H<sub>1</sub>), 9.16 (m, 1H, H<sub>27</sub>), 9.05 – 8.98 (m, 1H, H<sub>10</sub>), 8.98 – 8.85 (m, 1H, H<sub>39</sub>), 8.36 – 8.29 (m, 1H, H<sub>25</sub>), 8.24 – 7.36 (m, 25H, H<sub>3</sub>+ H<sub>2</sub>+ H<sub>4</sub>+ H<sub>5</sub>+ H<sub>6</sub>+ H<sub>8</sub>+ H<sub>9</sub>+ H<sub>11</sub>+ H<sub>12</sub>+ H<sub>13</sub>+ H<sub>15</sub>+ H<sub>16</sub>+ H<sub>17</sub>+ H<sub>18</sub>+ H<sub>19</sub>+ H<sub>20</sub>+ H<sub>21</sub>+ H<sub>26</sub>+ H<sub>28</sub>+ H<sub>29</sub>+ H<sub>30</sub>+ H<sub>31</sub>+ H<sub>40</sub>+ H<sub>41</sub>), 6.90 (d, *J* = 8.3 Hz, 2H, H<sub>42</sub>), 6.79 (m, 1H, H<sub>37</sub>), 5.78 – 5.62 (m, 1H, H<sub>36</sub>), 4.47 (q, *J* = 7.2 Hz, 2H, H<sub>23</sub>), 4.18 – 4.09 (m, 2H, H<sub>32</sub>), 3.81 (s, 3H, H<sub>43</sub>), 3.42 – 3.41 (m, 1H, H<sub>35</sub>), 3.21 – 3.19 (m, 1H, H<sub>35'</sub>), 2.99 – 2.95 (m, 2H, H<sub>34</sub>), 2.50 (s, 6H, H<sub>7</sub>+ H<sub>14</sub>), 2.27 – 2.18 (m, 2H, H<sub>33</sub>), 1.48 (t, *J* = 7.2 Hz, 3H, H<sub>24</sub>), 1.26 (m, 9H, H<sub>38</sub>). **LRMS** (ESI+) 1211.44 (35%, [M+H]<sup>+</sup>), 1257.46 (100%, [M+OC<sub>2</sub>H<sub>6</sub>+H]<sup>+</sup>).

#### Synthesis of E-9



*EZ*-**6** (9.8 mg, 8.1  $\mu$ mol) and trifluoroacetic acid (44  $\mu$ L, 0.568 mmol) were dissolved in CHCl<sub>3</sub> (8.1 mL, 1.0 mM) and stirred at room temperature for 17 hours. Triethylamine (160  $\mu$ L, 1.15 mmol) was added and the reaction mixture was washed with a saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was washed with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic layers were dried over

Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.2% Et<sub>3</sub>N), 20%) afforded E-9 (4.6 mg, 4.3 µmol, 53%) and Z-9 (1.5 mg, 1.4 µmol, 17%) as yellow solids. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) *E*-9 δ 16.28 (s, 1H, H<sub>22</sub>), 9.21 (d, *J* = 2.4 Hz, 1H, H<sub>27</sub>), 9.05 (d, J = 2.2 Hz, 1H, H<sub>10</sub>), 9.03 (s, 1H, H<sub>39</sub>), 8.32 (d, J = 8.5 Hz, 1H, H<sub>25</sub>), 8.22 - 8.21 (m, 3H, H<sub>2</sub>+ H<sub>11</sub>+ H<sub>18</sub>), 8.10 (dd, J = 8.6, 2.5 Hz, 1H, H<sub>26</sub>), 8.04 (d, J = 2.2 Hz, 1H, H<sub>13</sub>), 7.98 (dt, J = 7.9, 1.4 Hz, 1H, H<sub>19</sub>), 7.90 (dt, J = 7.6, 1.4 Hz, 1H, H<sub>5</sub>), 7.84 (s, 1H, H<sub>1</sub>), 7.72 (d, J = 7.9 Hz, 1H, H<sub>3</sub>), 7.66 (t, J = 7.5 Hz, 1H, H<sub>20</sub>),  $7.64 - 7.59 (m, 4H, H_4 + H_6 + H_{15} + H_8), 7.56 - 7.50 (m, 3H, H_9 + H_{16} + H_{21}), 7.47 (d, J = 1.7 Hz, 1H, H_{12}), 7.41 (d, J = 1.$  $(t, J = 7.9 Hz, 1H, H_{30}), 7.36 - 7.34 (m, 2H, H_{28} + H_{17}), 7.25 (d, J = 6.8 Hz, 1H, H_{31}), 6.95 (dd, J = 8.2, 2.4)$ Hz, 1H, H<sub>29</sub>), 6.87 (d, J = 7.7 Hz, 1H, H<sub>37</sub>), 5.74 – 5.70 (m, 1H, H<sub>36</sub>), 4.45 (q, J = 7.1 Hz, 2H, H<sub>23</sub>), 4.16 – 4.12 (m, 2H, H<sub>32</sub>), 3.58 (dd, J = 14.2, 4.2 Hz, 1H, H<sub>35</sub>), 3.26 (dd, J = 14.2, 6.9 Hz, 1H, H<sub>35</sub>'), 3.01 – 2.88 (m, 2H, H<sub>34</sub>), 2.51 (s, 6H, H<sub>7</sub>+ H<sub>14</sub>), 2.31 – 2.21 (m, 2H, H<sub>33</sub>), 1.46 (t, J = 7.1 Hz, 3H, H<sub>24</sub>), 1.29 (s, 9H, H<sub>38</sub>). <sup>13</sup>C **NMR** (151 MHz, CDCl<sub>3</sub>) *E*-**9** δ 178.5, 171.3, 165.9, 159.6, 151.3, 150.4, 145.3, 144.9, 142.0, 141.8, 141.4, 140.8, 139.4, 139.0, 138.4, 137.7, 137.0, 136.7, 136.6, 136.3, 135.2, 134.8, 134.7, 134.6, 133.6, 133.2, 131.0, 130.8, 130.5, 129.9, 129.7, 129.5, 129.4, 129.3, 129.2, 128.9, 128.4, 128.2, 127.8, 125.4, 124.7, 123.9, 120.5, 119.8, 115.1, 112.1, 112.0, 66.2, 61.3, 51.5, 39.6, 39.0, 34.8, 32.1, 28.6, 27.6, 21.0, 14.6. HRMS (NSI<sup>+</sup>): Calc. for C<sub>63</sub>H<sub>58</sub>DN<sub>7</sub>O<sub>6</sub>S<sub>2</sub>H: 1075.4104, found 1075.4100 [M+H]<sup>+</sup>.

#### Synthesis of Z-9



*EZ*-**6** (9.8 mg, 8.1  $\mu$ mol) and trifluoroacetic acid (2 $\mu$ L, 26.1  $\mu$ mol) were dissolved in CHCl<sub>3</sub> (3.3 mL) and stirred at room temperature for 41 hours. Triethylamine (8  $\mu$ L, 57.5 1  $\mu$ mol) was added and the reaction mixture was washed with a saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was washed with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>,

filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.2% Et<sub>3</sub>N), 20%) afforded Z-9 as a yellow solid (5.6 mg, 5.2 μmol, 64%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~63:37 cis:trans hydrazone amide rotamers.  $\delta$  13.63 (s, 1H, H<sub>22cis</sub>), 13.38 (s, 1H, H<sub>22trans</sub>), 10.24 (s, 1H, H<sub>1</sub>), 10.13 (s, 1H, H<sub>39trans</sub>), 9.01 (d, J = 2.2 Hz, 1H, H<sub>10</sub>), 8.99 (s, 1H, H<sub>39cis</sub>), 8.88 (s, 1H, H<sub>27</sub>), 8.27 - 8.14 (m, 3H, H<sub>2</sub>+ H<sub>11</sub>+ H<sub>26</sub>), 8.06 - 7.85 (m, 6H, H<sub>3</sub>+ H<sub>5</sub>+ H<sub>13</sub>+ H<sub>18</sub>+ H<sub>19</sub> + H<sub>25</sub>), 7.70 - 7.61 (m, 4H,  $H_{4}+H_{6}+H_{15}+H_{20}$ , 7.57 – 7.43 (m, 6H,  $H_{8}+H_{9}+H_{16}+H_{17}+H_{21}+H_{31}$ ), 7.40 – 7.35 (m, 2H,  $H_{12}+H_{30}$ ), 7.22 (t, J = 1.8 Hz, 1H, H<sub>28trans</sub>), 7.16 (t, J = 1.8 Hz, 1H, H<sub>28cis</sub>), 7.08 (d, J = 7.7 Hz, 1H, H<sub>37cis</sub>), 6.95 (m, 1H, H<sub>29</sub>), 6.87 (d, J = 7.7 Hz, 1H, H<sub>37trans</sub>), 5.74 – 5.70 (m, 1H, H<sub>36cis</sub>), 4.79 (q, J = 6.5 Hz, 1H, H<sub>36trans</sub>), 4.45 (m, 2H, H<sub>23</sub>), 4.16 – 4.12 (m, 2H, H<sub>32</sub>), 3.46 – 3.31 (m, 2H, H<sub>35</sub>), 3.01– 2.88 (m, 2H, H<sub>34</sub>), 2.51 (m, 6H, H<sub>7</sub>+ H<sub>14</sub>), 2.31 – 2.21 (m, 2H, H<sub>33</sub>), 1.46 (t, J = 7.1 Hz, 3H, H<sub>24</sub>), 1.29 (s, 9H, H<sub>38</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 192.5, 178.1, 171.1, 159.6, 159.4, 149.7, 149.6, 142.3, 142.0, 141.7, 141.6, 141.3, 141.1, 140.2, 140.0, 139.7, 138.9, 138.1, 137.1, 136.8, 136.5, 136.1, 135.8, 135.7, 135.5, 135.4, 135.3, 134.0, 133.4, 133.2, 131.1, 130.5, 130.4, 130.0, 129.8, 129.6, 129.3, 129.1, 128.2, 128.1, 128.0, 126.1, 125.1, 125.0, 124.1, 123.3, 119.5, 119.3, 114.7, 114.0, 113.6, 112.7, 135.8, 129.2, 128.6, 127.2, 70.7, 66.3, 65.8, 63.3, 61.7, 54.8, 52.3, 51.1, 50.1, 45.5, 43.2, 41.9, 40.7, 39.4, 39.2, 39.0, 36.5, 35.3, 35.0, 34.1, 33.7, 33.0, 28.5, 27.6, 22.8, 20.9, 14.5, 14.3. HRMS (NSI<sup>+</sup>): Calc. for C<sub>63</sub>H<sub>58</sub>DN<sub>7</sub>O<sub>6</sub>S<sub>2</sub>H: 1075.4104, found 1075.4078 [M+H]<sup>+</sup>.

## 2.2 Synthesis of EZ-10

#### 2.2.1 Synthetic Scheme



**Scheme 2.** Synthesis of *Z*-**10** and *E*-**10**. Reagents and conditions: (i) **S1**, HOBt·H<sub>2</sub>O, EDCl·HCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 17 h, 99%. (ii) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O, MeOH, Δ, 17 h, 65%. (iii) *p*-anisaldehyde, AcOH, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 18 h, 72%. (iv) AgNO<sub>3</sub>, CHCl<sub>3</sub>, MeOH, r.t., 40 mins, then *EZ*-**2** in CHCl<sub>3</sub>, r.t., 3 h, 69%. (v) TFA, CHCl<sub>3</sub>, r.t., 16 h, 70%. (vi) TFA, CHCl<sub>3</sub>, r.t., 65 h, 38%.

#### 2.2.2 Synthetic Procedures and Characterisation Data

#### Synthesis of S4



To a solution of **S1** (430 mg, 1.10 mmol) in  $CH_2Cl_2$  (20 mL) was added triethylamine (170 µL, 1.22 mmol), HOBt·H<sub>2</sub>O (166 mg, 1.23 mmol), EDCI·HCl (240 mg, 1.25 mmol) and pivaloylphenylalanine (299 mg, 1.20 mmol). The reaction mixture was stirred at room temperature for 17 hours. The reaction mixture was diluted with  $CH_2Cl_2$ , washed with a saturated aqueous solution of NaHCO<sub>3</sub>, a 1 M aqueous solution of HCl, and brine. The organic layer was

dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. Flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>,  $0\% \rightarrow 5\%$ ) afforded **S4** as a colourless solid (679 mg, 1.09 mmol, 99%). **M.p.** 61 °C. <sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) ~1:1 mixture of rotamers.  $\delta$  8.47–8.41 (m, 1H, H<sub>4</sub>), 7.38 (d, *J* = 8.6 Hz, 1H, H<sub>2</sub>), 7.36 – 7.19 (m, 19H, H<sub>13</sub>+ H<sub>14</sub>+ H<sub>15</sub>+ H<sub>9</sub>+ H<sub>10</sub>), 7.17 – 7.14 (m, 1H, H<sub>11</sub>), 4.59 – 4.54 (m, 1H, H<sub>3</sub>), 4.16 – 4.12 (m, 1H, H<sub>5</sub>), 4.05 – 3.99 (m, 2H, H<sub>6</sub>), 3.04 – 2.98 (m, 1H, H<sub>8</sub>), 2.89 – 2.84 (m, 1H, H<sub>8'</sub>), 2.57 – 2.52 (m, 1H, H<sub>12</sub>), 2.42 – 2.36 (m, 1H, H<sub>12'</sub>), 1.12 – 1.09 (m, 3H, H<sub>7</sub>), 0.97 – 0.96 (m, 9H, H<sub>1</sub>). <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  177.1, 171.8, 171.6, 170.1, 144.1, 138.0, 129.4, 129.1, 128.2, 127.9, 127.2, 126.9, 126.2, 66.4, 66.3 61.0, 60.9, 53.8, 53.6, 51.8, 51.3, 38.0, 37.0, 33.3, 32.8, 27.2, 14.0. HRMS (NSI<sup>+</sup>): Calc. for C<sub>38</sub>H<sub>42</sub>N<sub>2</sub>O<sub>4</sub>SNa: 645.2757, found 645.2732 [M+Na]<sup>+</sup>.

Synthesis of S5



Hydrazine hydrate (15.0 µL, 0.309 mmol) was added to a solution of **S4** (156 mg, 0.250 mmol) in MeOH (1 mL) and the reaction mixture was stirred at reflux for 17 hours. The reaction mixture was concentrated under reduced pressure and purified by flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>,  $0\% \rightarrow 4\%$ ) to afford **S5** as a colourless solid (98.5 mg, 0.162 mmol, 65%).<sup>1</sup>H **NMR** (600 MHz, CDCl<sub>3</sub>) ~1:2 *cis:trans* mixture of hydrazone amide rotamers.

Major rotamer reported.  $\delta$  7.79 (s, 2H, H<sub>6</sub>), 7.38 (d, *J* = 8.3 Hz, 6H, H<sub>13</sub>), 7.29 – 7.11 (m, 14H, H<sub>14</sub>+ H<sub>15</sub>+ H<sub>9</sub>+ H<sub>10</sub>+ H<sub>11</sub>), 6.85 (d, *J* = 7.0 Hz, 1H, H<sub>4</sub>), 6.31 (d, *J* = 7.1 Hz, 1H, H<sub>2</sub>), 4.54 (q, *J* = 7.0 Hz, 1H, H<sub>3</sub>), 4.02 (q, *J* = 7.3, 1H, H<sub>5</sub>), 3.76 (s, 3H, H<sub>7</sub>), 3.04 (dd, *J* = 14.0, 7.0 Hz, 1H, H<sub>8</sub>), 2.95 (dd, *J* = 13.8, 7.2 Hz, 1H, H<sub>8'</sub>), 2.59 (dd, *J* = 13.0, 7.6 Hz, 1H, H<sub>12</sub>), 2.53 (dd, *J* = 13.3, 5.9 Hz, 1H, H<sub>12'</sub>), 1.07 (s, 9H, H<sub>1</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  179.1, 171.2, 170.0, 144.3, 136.1, 129.5, 129.3, 128.6, 128.0, 127.0, 126.9, 67.2, 54.4, 51.2, 38.6, 37.9, 33.3, 27.3, 18.4. HRMS (NSI<sup>+</sup>): Calc. for C<sub>36</sub>H<sub>40</sub>N<sub>4</sub>O<sub>3</sub>SNa: 631.2713, found 631.2736 [M+Na]<sup>+</sup>.

#### Synthesis of 4



**S5** (98.5 mg, 0.162 mmol), *p*-anisaldehyde (24  $\mu$ L, 0.200 mmol) and acetic acid (2 drops) were combined in CH<sub>2</sub>Cl<sub>2</sub> (1.6 mL) and stirred at room temperature for 18 hours. The reaction mixture was concentrated under reduced pressure and purified by flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 0%  $\rightarrow$  2.5%) to afford **4** as a colourless solid (85.1 mg, 0.117 mmol, 72%). Preparative thin layer

chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub>, 20% × 2) was used to separate the different diastereoisomers. **M.p.** 212 °C. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) ~43:57 *cis:trans* hydrazone amide rotamers. *Trans*-4  $\delta$  9.96 (s, 1H, H<sub>6</sub>), 8.29 (s, 1H, H<sub>7</sub>), 7.66 (d, *J* = 6.7 Hz, 2H, H<sub>8</sub>), 7.41 – 7.17 (m, 19H, H<sub>16</sub>+ H<sub>17</sub>+ H<sub>18</sub>+ H<sub>12</sub>+ H<sub>13</sub>), 7.13 – 7.10 (m, 1H, H<sub>14</sub>), 6.90–6.87 (m, 2H, H<sub>9</sub>), 5.99 (d, *J* = 4.9 Hz, 1H, H<sub>2</sub>), 5.93 (d, *J* = 8.2 Hz, 1H, H<sub>4</sub>), 4.37 – 4.33 (m, 1H, H<sub>5</sub>), 4.06 – 4.02 (m, 1H, H<sub>3</sub>), 3.82 (s, 3H, H<sub>10</sub>), 3.10 – 3.07 (m, 1H, H<sub>11</sub>), 2.98 – 2.90 (m, 3H, H<sub>15</sub>+H<sub>11</sub>), 2.63 – 2.60 (m, 1H, H<sub>15</sub>), 1.08 (s, 9H, H<sub>1</sub>). *Cis*-4  $\delta$ 9.42 (s, 1H, H<sub>6</sub>), 7.64 (s, 1H, H<sub>7</sub>), 7.46 (d, *J* = 8.8 Hz, 1H, H<sub>8</sub>), 7.41 – 7.17 (m, 19H, H<sub>16</sub>+ H<sub>17</sub>+ H<sub>18</sub>+ H<sub>12</sub>+ H<sub>13</sub>), 7.14 (d, *J* = 7.3 Hz, 1H, H<sub>4</sub>), 7.13 – 7.10 (m, 1H, H<sub>14</sub>), 6.90 – 6.87 (m, 2H, H<sub>9</sub>), 6.29 (d, *J* = 8.1 Hz, 1H, H<sub>2</sub>), 5.46 – 5.43 (m, 1H, H<sub>5</sub>), 5.10 – 5.06 (m, 1H, H<sub>3</sub>), 3.86 (s, 3H, H<sub>10</sub>), 3.24 – 3.20 (m, 1H, H<sub>11</sub>), 3.05 – 3.01 (m, 1H, H<sub>11</sub>), 2.98 – 2.90 (m, 1H, H<sub>15</sub>), 2.48 – 2.45 (m, 1H, H<sub>15</sub>), 1.08 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  180.2, 178.4, 171.2, 165.8, 161.5, 149.3, 114.2, 67.4, 66.7, 56.1, 55.5, 53.4, 51.8, 49.7, 38.8, 36.8, 34.1, 33.4, 29.9, 27.5, 27.4. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>44</sub>H<sub>46</sub>N<sub>4</sub>O<sub>4</sub>SNa: 749.3132, found 749.3119 [M+Na]<sup>+</sup>.

#### Synthesis of EZ-7

AgNO<sub>3</sub> (4.3 mg, 25 µmol) was added to a solution of 4 (15.4 mg, 21.2 µmol) in a 2:1 CHCl<sub>3</sub>:MeOH



mixture (600  $\mu$ L) and the mixture was stirred at room temperature for 40 minutes, a solution of *EZ*-**2** (31.3 mg, 31.8  $\mu$ mol) in CHCl<sub>3</sub> (200  $\mu$ L) was added and the reaction was stirred at room temperature for 3 hours. The reaction mixture was diluted with CHCl<sub>3</sub>, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered

and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.1% Et<sub>3</sub>N), 30%) afforded *EZ*-**7** as a yellow solid (20.0 mg, 14.7  $\mu$ mol, 69%). <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) ~62:38 *E:Z* pyridyl-hydrazone isomers and mixture of acyl-hydrazone rotamers.  $\delta$  15.82 – 15.78 (m, 1H), 13.48 – 13.41 (m, 1H), 10.35 – 10.31 (m, 1H), 10.15 – 10.05 (m, 2H), 9.16 – 9.10 (m, 1H), 9.05 –

8.96 (m, 2H), 8.87 – 8.85 (m, 1H), 8.37 (d, J = 6.6 Hz, 1H), 8.31 – 8.27 (m, 1H), 8.23 – 8.07 (m, 5H), 8.05 – 8.00 (m, 1H), 7.99 – 7.85 (m, 5H), 7.84 – 7.74 (m, 3H), 7.69 – 7.53 (m, 14H), 7.51 – 7.29 (m, 11H), 7.22 – 7.12 (m, 5H), 6.98 – 6.81 (m, 5H), 6.41 – 6.33 (m, 1H), 6.21 – 6.12 (m, 1H), 5.74 (q, J = 6.0 Hz, 1H), 5.50 – 5.33 (m, 1H), 5.18 (quint., J = 6.6 Hz, 1H), 4.92 – 4.85 (m, 1H), 4.53 (q, J = 7.2 Hz, 1H), 4.43 (q, J = 7.2 Hz, 2H), 4.24 – 4.18 (m, 1H), 4.16 – 4.06 (m, 3H), 3.83 – 3.74 (m, 5H), 3.43 – 3.35 (m, 6H), 3.34 – 3.28 (m, 1H), 3.25 – 3.21 (m, 1H), 3.20 – 3.14 (m, 2H), 3.09 – 2.96 (m, 3H), 2.95 – 2.85 (m, 4H), 2.52 – 2.42 (m, 9H), 2.25 – 2.16 (m, 4H), 1.47 – 1.41 (m, 5H), 1.25 (s, 5H), 1.17 – 1.14 (m, 7H), 1.13 – 1.08 (m, 2H), 1.07 – 1.05 (m, 6H). **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>80</sub>H<sub>75</sub>DN<sub>8</sub>O<sub>9</sub>S<sub>2</sub>H: 1358.5312; found 1358.5290 [M+H]<sup>+</sup>.

#### Synthesis of E-10



*EZ*-**7** (10.0 mg, 7.35  $\mu$ mol) and trifluoroacetic acid (40  $\mu$ L, 0.515 mmol) were dissolved in CHCl<sub>3</sub> (7.4 mL) and stirred at room temperature for 16 h. Triethylamine (143  $\mu$ L, 1.03 mmol) was added and the mixture was washed with a

saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.1% Et<sub>3</sub>N), 30%) afforded E-10 as a yellow solid (6.3 mg, 5.2 μmol, 70%) and Z-10 as a yellow solid (2.0 mg, 1.6 μmol, 22%). <sup>1</sup>H NMR (600 MHz, Chloroform-d) ~87:13 cis:trans hydrazone amide rotamers, major rotamer reported δ 16.34 (s, 1H, H<sub>22</sub>), 10.53 (s, br, 1H, H<sub>38</sub>), 9.26 (d, J = 2.4 Hz, 1H, H<sub>27</sub>), 9.11 (d, J = 2.1 Hz, 1H, H<sub>10</sub>), 8.37 – 8.34 (m, 2H, H<sub>25</sub>+ H<sub>2</sub>), 8.28 - 8.21 (m, 2H, H<sub>11</sub>+ H<sub>18</sub>), 8.14 (dd, J = 8.6, 2.5 Hz, 1H, H<sub>26</sub>), 8.10 - 8.04 (m, 2H, H<sub>13</sub>+ H<sub>1</sub>), 8.01 (d, J = 7.5 Hz, 1H, H<sub>19</sub>), 7.93 (dt, J = 7.5 Hz, 1H, H<sub>5</sub>), 7.76 – 7.65 (m, 7H, H<sub>4</sub>+ H<sub>6</sub>+ H<sub>8</sub>+ H<sub>15</sub>+ H<sub>16</sub>+ H<sub>20</sub>+ H<sub>37</sub>), 7.64 – 7.62 (m, 1H, H<sub>17</sub>), 7.60 – 7.57 (m, 2H, H<sub>3</sub>+ H<sub>21</sub>), 7.53 – 7.48 (m, 2H, H<sub>12</sub>+ H<sub>9</sub>), 7.46 (t, *J* = 7.8 Hz, 1H, H<sub>30</sub>), 7.41 (s, 1H, H<sub>28</sub>), 7.33 – 7.30 (m, 4H, H<sub>31</sub>+ H<sub>43</sub>+ H<sub>45</sub>), 7.27 – 7.24 (m, 2H, H<sub>44</sub>), 7.00 (dd, J = 8.2, 2.0 Hz, 1H, H<sub>29</sub>), 6.30 (d, J = 8.3 Hz, 1H, H<sub>40</sub>), 5.84 (dt, J = 7.2, 6.4 Hz, 1H, H<sub>36</sub>), 5.25 (dt, J = 8.1, 6.8 Hz, 1H, H<sub>39</sub>), 4.48 (q, J = 7.2 Hz, 2H, H<sub>23</sub>), 4.27 – 4.11 (m, 2H, H<sub>32</sub>), 3.44 (dd, J = 13.3, 5.7 Hz, 1H, H<sub>35</sub>), 3.32 (dd, J = 14.0, 6.1 Hz, 1H, H<sub>42</sub>), 3.17 (dd, J = 13.3, 5.7 Hz, 1H, H<sub>35</sub>), 3.06 (dd, J = 14.0, 6.1 Hz, 1H, H<sub>42</sub>'), 3.02 – 2.92 (m, 2H, H<sub>34</sub>), 2.57 (s, 3H, H<sub>7</sub>), 2.54 (s, 3H, H<sub>14</sub>), 2.36 – 2.32 (m, 2H, H<sub>33</sub>), 1.50 (t, *J* = 7.2 Hz, 3H, H<sub>24</sub>), 1.05 (s, 9H, H<sub>41</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 178.7, 171.2, 171.1, 165.8, 159.6, 159.6, 151.1, 150.4, 145.9, 145.2, 141.9, 141.7, 141.6, 141.3, 140.6, 139.2, 138.9, 138.3, 137.4, 136.9, 136.6, 136.5, 136.4, 136.2, 135.1, 134.7, 134.5, 134.1, 133.1, 130.9, 130.7, 130.4, 129.9, 129.6, 129.5, 129.2, 129.1, 129.0, 128.8, 128.5, 128.3, 128.1, 127.7, 126.9, 125.4, 124.6, 123.8, 120.3, 119.9, 114.7, 112.3, 111.8, 66.3, 61.2, 53.4, 49.9, 40.1, 39.1, 38.6, 34.5, 31.9, 29.7, 29.6, 28.7, 27.4, 27.3, 27.3, 27.2, 22.7, 20.9, 20.9, 14.5, 14.2. **HRMS** (ESI<sup>+</sup>): Calc. for C<sub>72</sub>H<sub>67</sub>DN<sub>8</sub>O<sub>7</sub>S<sub>2</sub>H: 1222.4788; found 1222.4751 [M+H]<sup>+</sup>.

#### Synthesis of Z-10



*EZ*-**7** (10.0 mg, 7.35  $\mu$ mol) and trifluoroacetic acid (2  $\mu$ L, 22  $\mu$ mol) were dissolved in CHCl<sub>3</sub> (3.0 mL) and stirred at room temperature for 65 hours. Triethylamine (6  $\mu$ L, 44  $\mu$ mol) was added and the mixture was washed with a saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the combined

organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.1% Et<sub>3</sub>N), 30%) afforded Z-10 as a yellow solid (3.4 mg, 2.8  $\mu$ mol, 38%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~22:78 *cis:trans* hydrazone amide rotamers.  $\delta$  13.65 (s, 1H,  $H_{22cis}$ ), 13.44 (s, 1H,  $H_{22trans}$ ), 10.59 (s, 1H,  $H_{38trans}$ ), 10.16 (s, 1H,  $H_1$ ), 9.01 (d, J = 2.1 Hz, 1H,  $H_{10cis}$ ), 8.99 (d, J = 2.1 Hz, 1H, H<sub>10trans</sub>), 8.93 (m, 2H, H<sub>27</sub>+ H<sub>38cis</sub>), 8.46 (t, J = 1.8 Hz, 1H, H<sub>18</sub>), 8.22 (m, 1H, H<sub>2</sub>), 8.18 (d, J = 2.2 Hz, 1H, H<sub>11</sub>), 8.09 (dd, J = 8.2, 2.3 Hz, 1H, H<sub>26</sub>), 7.99 - 7.92 (m, 6H, H<sub>3</sub>+ H<sub>5</sub>+ H<sub>13</sub>+ H<sub>19</sub>+ H<sub>21</sub>+  $H_{25}$ ), 7.73 – 7.67 (m, 4H,  $H_6$ +  $H_4$ +  $H_{20}$ +  $H_8$ ), 7.66 – 7.62 (m, 2H,  $H_{15}$ +  $H_{16}$ ), 7.56 (d, J = 7.8 Hz, 1H,  $H_{37}$ ), 7.53 – 7.47 (m, 3H, H<sub>12</sub>+ H<sub>17</sub>+ H<sub>9</sub>), 7.43 – 7.39 (m, 2H, H<sub>29</sub>+ H<sub>30</sub>), 7.37 – 7.34 (m, 2H, H<sub>43</sub>), 7.31 (d, J = 7.8 Hz, 1H, H<sub>45</sub>), 7.28 - 7.23 (m, 3H, H<sub>31</sub>+ H<sub>44</sub>), 6.94 (dd, J = 9.8, 8.1 Hz, 1H, H<sub>28</sub>), 6.36 (d, J = 8.7 Hz, 1H, H<sub>40trans</sub>), 6.21 (d, J = 4.7 Hz, 1H, H<sub>36trans</sub>), 6.19 (d, J = 8.7 Hz, 1H, H<sub>40cis</sub>), 5.76 (m, 1H, H<sub>36cis</sub>), 5.12 (m, 1H, H<sub>39cis</sub>), 4.95 (dt, J = 9.4, 5.1 Hz, 1H, H<sub>39trans</sub>), 4.62 – 4.49 (m, 2H, H<sub>23</sub>), 4.23 – 4.14 (m, 2H, H<sub>32</sub>), 3.56 (dd, J = 13.5, 5.2 Hz, 1H, H<sub>42</sub>), 3.22 (dd, J = 13.1, 8.4 Hz, 1H, H<sub>35</sub>), 3.15 (dd, J = 13.1, 7.7 Hz, 1H, H<sub>35'</sub>), 2.97 -2.89 (m, 2H, H<sub>34</sub>), 2.86 (dd, J = 13.5, 4.8 Hz, 1H, H<sub>42</sub>), 2.50 (s, 3H, H<sub>7</sub>), 2.47 (s, 3H, H<sub>14</sub>), 2.31 – 2.15 (m, 2H, H<sub>33</sub>), 1.50 (t, J = 7.2 Hz, 3H, H<sub>24</sub>), 1.27 (s, 9H, H<sub>41</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 192.4, 180.7, 171.1, 166.1, 163.2, 159.4, 154.2, 149.4, 146.9, 141.9, 141.6, 141.4, 140.9, 140.1, 139.6, 138.5, 138.1, 137.0, 136.7, 135.9, 135.6, 135.4, 135.2, 134.8, 134.5, 133.1, 130.9, 130.1, 130.0, 129.8, 129.6, 129.5, 129.4, 129.1, 129.0, 128.9, 128.8, 128.6, 128.4, 128.1, 128.0, 127.9, 127.6, 125.1, 125.0, 124.7, 123.9, 123.2, 119.8, 119.3, 113.9, 113.8, 112.7, 66.8, 61.6, 61.6, 57.6, 50.6, 38.7, 38.6, 38.2, 36.7, 35.2, 34.4, 31.9, 29.7, 29.6, 29.4, 29.2, 27.4, 27.3, 22.7, 20.8, 14.3, 14.2, 14.1. HRMS (ESI<sup>+</sup>): Calc. for C<sub>72</sub>H<sub>67</sub>DN<sub>8</sub>O<sub>7</sub>S<sub>2</sub>H: 1222.4788; found 1222.4742 [M+H]+.

## 2.3 Synthesis of EZ-11

## 2.3.1 Synthetic Scheme



**Scheme 3.** Synthesis of *Z*-**11** and *E*-**11**. Reagents and conditions: (i) HO-Ala-Piv, HOBt·H<sub>2</sub>O, EDCl·HCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 18 h, 97%. (ii) LiOH, H<sub>2</sub>O, THF, r.t., 2 h, 95%. (iii) **S1**, HOBt·H<sub>2</sub>O, EDCl·HCl, CH<sub>2</sub>Cl<sub>2</sub>, r.t., 22 h, 91%. (iv) NH<sub>2</sub>NH<sub>2</sub>·H<sub>2</sub>O, MeOH, Δ, 22 h, 50%. (v) *p*-anisaldehyde, AcOH, CH<sub>2</sub>Cl<sub>2</sub>, 30 °C, 23 h, 99%. (vi) AgNO<sub>3</sub>, CHCl<sub>3</sub>, MeOH, r.t., 60 mins, then *EZ*-**2** in CHCl<sub>3</sub>, r.t., 27 h, 63%. (vii) TFA, CHCl<sub>3</sub>, r.t., 23 h, 22%. (viii) TFA, CHCl<sub>3</sub>, r.t., 47 h, 43%.

#### 2.3.2 Synthetic Procedures and Characterisation Data

#### Synthesis of S6



To a solution of L-phenylalanine methyl ester hydrochloride (654 mg, 3.03 mmol) in  $CH_2Cl_2$  (60 mL) was added triethylamine (500  $\mu$ L, 3.60 mmol,) HOBt·H<sub>2</sub>O (489 mg, 3.62 mmol), EDCI·HCl (700 mg, 3.65 mmol) and

Pivaloylalanine (624 mg, 3.60 mmol). The reaction mixture was stirred at room temperature for 18 hours. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with a saturated aqueous solution of NaHCO<sub>3</sub> and brine. The organic layer was dried over MgSO<sub>4</sub>, filtered and concentrated under reduced pressure. Flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>,  $3\% \rightarrow 6\%$ ) afforded **S6** as a colourless solid (980 mg, 2.93 mmol, 97%). **M.p.** 69 °C. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.28 (t, *J* = 7.3 Hz, 2H, H<sub>9</sub>), 7.23 (t, *J* = 7.3 Hz, 1H, H<sub>10</sub>), 7.11 (d, *J* = 7.3 Hz, 2H, H<sub>8</sub>), 6.74 (d, *J* = 7.7 Hz, 1H, H<sub>2</sub>), 6.23 (d, *J* = 7.1 Hz, 1H, H<sub>4</sub>), 4.81 (q, *J* = 6.7 Hz, 1H, H<sub>5</sub>), 4.48 (quint, *J* = 7.1 Hz, 1H, H<sub>3</sub>), 3.71 (s, 3H, H<sub>6</sub>), 3.15 (dd, *J* = 13.9, 5.8 Hz, 1H, H<sub>7</sub>), 3.07 (dd, *J* = 14.0, 6.6 Hz, 1H, H<sub>7</sub>), 1.34 (d, *J* = 7.0 Hz, 3H, H<sub>11</sub>), 1.17 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  178.3, 172.2, 171.7, 135.8, 129.2, 128.6, 127.2, 53.4, 52.4, 48.5, 38.6, 37.8, 27.4, 18.4. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>18</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub>Na: 357.1785, found 357.1767 [M+Na]<sup>+</sup>.

#### Synthesis of S7



**S6** (670 mg, 2.00 mmol) was dissolved in a 3:1 THF:LiOH (1 M) mixture (16 mL) and stirred at room temperature for 2 h. The reaction mixture was acidified with 1 M HCl (4.0 mL) and diluted with EtOAc. The two layers were separated and the aqueous layer extracted with EtOAc. The combined organic layers

were washed with brine, dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>,  $0\% \rightarrow 5\%$ ) afforded **S7** as a colourless solid (609 mg, 1.90 mmol, 95%). **M.p.** 98 °C. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.25 (t, *J* = 7.3 Hz, 2H, H<sub>9</sub>), 7.20 (t, *J* = 7.3 Hz, 1H, H<sub>10</sub>), 7.13 (d, *J* = 7.3 Hz, 2H, H<sub>8</sub>), 7.10 (d, *J* = 7.6 Hz, 1H, H<sub>2</sub>), 6.48 (d, *J* = 7.6 Hz, 1H, H<sub>4</sub>), 4.79 (q, *J* = 6.6 Hz, 1H, H<sub>5</sub>), 4.56 (quint, *J* = 7.1 Hz, 1H, H<sub>3</sub>), 3.17 (dd, *J* = 14.0, 5.6 Hz, 1H, H<sub>7</sub>), 2.99 (dd, *J* = 14.0, 6.7 Hz, 1H, H<sub>7</sub>), 1.29 (d, *J* = 6.9 Hz, 3H, H<sub>11</sub>), 1.17 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  178.9, 173.6, 172.3, 136.0, 129.4, 128.5, 127.0, 53.5, 48.6, 38.7, 37.6, 27.3, 18.3. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>17</sub>H<sub>24</sub>N<sub>2</sub>O<sub>4</sub>H: 321.1809, found 321.1798 [M+H]<sup>+</sup>.

#### Synthesis of S8



To a solution of **S1** (392 mg, 1.00 mmol) in  $CH_2CI_2$  (20 mL) was added HOBt·H<sub>2</sub>O (162 mg, 1.20 mmol), EDCI·HCI (230 mg, 1.20 mmol) and **S7** (384 mg, 1.20 mmol). The reaction mixture was stirred at room temperature for 22 hours. The reaction mixture was diluted with  $CH_2CI_2$ , washed with a saturated aqueous solution of NaHCO<sub>3</sub>, a 1 M solution of HCl, and brine. The organic layer was dried over MgSO<sub>4</sub>,

filtered and concentrated under reduced pressure. Flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 9%) afforded **S8** as a pale yellow solid (632 mg, 0.911 mmol, 91%). **M.p.** 104 °C. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (d, *J* = 7.3 Hz, 6H, H<sub>11</sub>), 7.27 (t, *J* = 7.4 Hz, 6H, H<sub>12</sub>), 7.22 – 7.20 (m, 7H, H<sub>13</sub>+ H<sub>15</sub>+ H<sub>16</sub>), 7.14 – 7.11 (m, 1H, H<sub>17</sub>), 6.74 (d, *J* = 5.6 Hz, 1H, H<sub>4</sub>), 6.33 (d, *J* = 7.0 Hz, 1H, H<sub>5</sub>), 6.17 (d, *J* = 6.7 Hz, 1H, H<sub>2</sub>), 4.61 (q, *J* = 6.9 Hz, 1H, H<sub>5</sub>), 4.44 – 4.39 (m, 2H, H<sub>7</sub>+ H<sub>3</sub>), 4.17 – 4.11 (m, 2H, H<sub>8</sub>), 3.09 – 3.01 (m, 1H, H<sub>5</sub>), 2.65 (dd, *J* = 12.6, 6.2 Hz, 1H, H<sub>14</sub>), 2.53 (dd, *J* = 12.5, 5.0 Hz, 1H, H<sub>14</sub>'), 1.31 (d, *J* = 7.0 Hz, 3H, H<sub>18</sub>), 1.24 (t, *J* = 7.1 Hz, 3H, H<sub>9</sub>), 1.14 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  178.5, 172.1, 170.2, 169.8, 144.2, 136.2, 129.5, 129.4, 128.7, 128.0, 127.0, 126.9, 66.9, 61.8, 54.1, 51.5, 48.8, 38.6, 38.0, 33.5, 27.4, 18.3, 14.1. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>41</sub>H<sub>47</sub>N<sub>3</sub>O<sub>5</sub>SH: 694.3309, found 694.3280 [M+H]<sup>+</sup>.

Synthesis of S9



Hydrazine hydrate (42  $\mu$ L, 0.864 mmol) was added to a solution of **S8** (173 mg, 0.249 mmol) in MeOH (3 mL) and the reaction mixture was stirred at reflux for 22 hours. The reaction mixture was allowed to cool to room temperature to precipitate the product which was isolated by filtration as a colourless solid (84.5 mg, 0.124 mmol, 50%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 (s, br, 1H, H<sub>8</sub>), 7.38 (d, *J* = 7.2 Hz, 6H, H<sub>11</sub>), 7.28

(t, J = 7.6 Hz, 6H, H<sub>12</sub>), 7.23 – 7.16 (m, 6H, H<sub>13</sub>+ H<sub>16</sub>+ H<sub>17</sub>), 7.12 (d, J = 7.7 Hz, 2H, H<sub>15</sub>), 6.75 (d, J = 6.7 Hz, 1H, H<sub>4</sub>), 6.61 (d, J = 8.3 Hz, 1H, H<sub>6</sub>), 6.06 (d, J = 5.3 Hz, 1H, H<sub>2</sub>), 4.52 (q, J = 6.6 Hz, 1H, H<sub>5</sub>), 4.29 – 4.22 (m, 2H, H<sub>7</sub>+ H<sub>3</sub>), 3.74 (s, br, 2H, H<sub>9</sub>), 3.16 (dd, J = 14.1, 6.1 Hz, 1H, H<sub>10</sub>), 2.92 (dd, J = 14.0, 6.6 Hz, 1H, H<sub>10'</sub>), 2.73 (dd, J = 13.1, 8.7 Hz, 1H, H<sub>14</sub>), 2.65 (dd, J = 13.1, 5.1 Hz, 1H, H<sub>14</sub>), 1.30 (d, J = 7.0 Hz, 3H, H<sub>18</sub>), 1.09 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  179.7, 172.9, 170.5, 169.9, 144.4, 135.6, 129.6, 129.3, 129.0, 128.0, 127.3, 126.9, 67.2, 54.4, 51.8, 50.0, 38.5, 37.3, 33.4, 27.3, 17.8. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>39</sub>H<sub>45</sub>N<sub>5</sub>O<sub>4</sub>SH: 680.3265, found 680.3246 [M+H]<sup>+</sup>.

#### Synthesis of 5



**S9** (75.0 mg, 0.110 mmo), *p*-anisaldehyde (27  $\mu$ L, 0.220 mmol) and acetic acid (2 drops) were combined in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) and the reaction mixture was stirred at 30 °C for 23 hours. The reaction mixture was concentrated under reduced pressure and purified by flash column chromatography (SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 0%  $\rightarrow$  3%) to afford **5** as a colourless film (87.1

mg, 0.109 mmol, 99%). <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) ~32:68 *cis:trans* hydrazone amide rotamers. *Trans*-**5** δ 10.13 (s, 1H, H<sub>8</sub>), 8.18 (s, 1H, H<sub>9</sub>), 7.66 (d, *J* = 8.8 Hz, 2H, H<sub>10</sub>), 7.38 (d, *J* = 7.6 Hz, 6H, H<sub>14</sub>), 7.28 – 7.07 (m, 14H, H<sub>15</sub>+ H<sub>16</sub>+ H<sub>18</sub>+ H<sub>19</sub>+ H<sub>20</sub>), 7.01 (d, *J* = 7.9 Hz, 1H, H<sub>4</sub>), 6.88 (d, *J* = 8.8 Hz, 2H, H<sub>11</sub>), 6.80 – 6.77 (m, 1H, H<sub>6</sub>), 6.20 (s, br, 1H, H<sub>2</sub>), 4.59 – 4.56 (m, 2H, H<sub>17</sub>), 4.50 – 4.45 (m, 1H, H<sub>7</sub>), 4.35 (s, br, 1H, H<sub>3</sub>), 3.82 (s, 3H, H<sub>12</sub>), 3.07 – 2.95 (m, 1H, H<sub>5</sub>), 2.79 – 2.73 (m, 2H, H<sub>13</sub>), 1.28 (d, *J* = 7.1 Hz, 3H, H<sub>21</sub>), 1.11 (s, 9H, H<sub>1</sub>). *Cis*-**5** δ 9.27 (s, 1H, H<sub>8</sub>), 7.62 (s, 1H, H<sub>9</sub>), 7.46 (d, *J* = 8.8 Hz, 2H, H<sub>10</sub>), 7.31 (d, *J* = 7.3 Hz, 6H, H<sub>14</sub>), 7.28 – 7.07 (m, 14H, H<sub>15</sub>+ H<sub>16</sub>+ H<sub>18</sub>+ H<sub>19</sub>+ H<sub>20</sub>), 7.01 (d, *J* = 7.9 Hz, 1H, H<sub>4</sub>), 6.90 (d, *J* = 8.7 Hz, 2H, H<sub>11</sub>), 6.80 – 6.77 (m, 1H, H<sub>6</sub>), 6.27 (d, *J* = 7.3 Hz, 1H, H<sub>2</sub>), 5.37 – 5.34 (m, 1H, H<sub>7</sub>), 4.73 – 4.69 (m, 2H, H<sub>17</sub>), 4.50 – 4.45 (m, 1H, H<sub>3</sub>), 3.87 (s, 3H, H<sub>12</sub>), 3.14 – 3.11 (m, 1H, H<sub>5</sub>), 3.07 – 2.95 (m, 1H, H<sub>13</sub>), 2.61 – 2.58 (m, 1H, H<sub>13</sub>), 1.31 (d, *J* = 7.0 Hz, 3H, H<sub>21</sub>), 1.15 (s, 9H, H<sub>1</sub>). <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>) δ 179.6, 178.4, 173.1, 172.5, 170.8, 170.4, 166.0, 161.6, 149.1, 145.1, 144.6, 144.5, 136.5, 135.8, 129.7, 129.6, 129.5, 129.2, 129.1, 128.2, 128.0, 127.4, 127.1, 127.0, 126.8, 126.7, 126.1, 114.4, 114.2, 67.3, 66.8, 55.6, 55.5, 54.8, 52.2, 50.0, 48.8, 38.7, 38.5, 36.8, 33.9, 33.7, 27.6, 27.5, 18.7, 18.2. **HRMS** (NSI<sup>+</sup>): Calc. for C<sub>47</sub>H<sub>51</sub>N<sub>5</sub>O<sub>5</sub>SNa: 820.3503, found 820.3476 [M+Na]<sup>+</sup>.

Synthesis of EZ-8



AgNO<sub>3</sub> (6.1 mg, 36.0  $\mu$ mol) was added to a solution of **5** (23.9 mg, 30.0  $\mu$ mol) in 3:1 CHCl<sub>3</sub>:MeOH mixture (800  $\mu$ L) and the mixture was stirred at room temperature for 1 hour. A solution of *EZ*-**2** (44.3 mg, 45.0  $\mu$ mol) in CHCl<sub>3</sub> (450  $\mu$ L) was added and the reaction was stirred at room temperature for 27 hours. The reaction mixture was diluted with CHCl<sub>3</sub>, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated

under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.1% Et<sub>3</sub>N), 40%) afforded *EZ*-**8** as a yellow solid (27.0 mg, 18.9  $\mu$ mol, 63%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~70:30 *E:Z* pyridyl-hydrazone isomers and mixture of acyl-hydrazone rotamers.  $\delta$  16.23 (s, 1H), 13.47 (s, br, 1H), 10.15

(s, 1H), 10.03 (s, br, 1H), 9.18 – 9.14 (m, 1H), 9.05 – 8.97 (m, 2H), 8.34 – 8.28 (m, 2H), 8.24 – 8.15 (m, 4H), 8.12 – 8.10 (m, 1H), 8.08 – 8.03 (dt, *J* = 8.7, 1.8 Hz, 1H), 8.00 – 7.90 (m, 5H), 7.84 – 7.78 (m, 2H), 7.74 – 7.56 (m, 16H), 7.55 – 7.40 (m, 12H), 7.34 – 7.30 (m, 4H), 7.27 – 7.18 (m, 9H), 7.02 – 6.89 (m, 6H), 6.36 (s, br, 1H), 5.93 (dd, *J* = 15.4, 2.8 Hz, 1H), 5.08 – 5.03 (m, 1H), 5.06 (t, *J* = 9.9 Hz, 1H), 4.62 (q, *J* = 6.2 Hz, 1H), 4.59 – 4.54 (m, 1H), 4.46 (q, *J* = 6.8 Hz, 2H), 4.23 – 4.02 (m, 6H), 3.84 (d, *J* = 2.5 Hz, 4H), 3.54 (dt, *J* = 13.8, 3.5 Hz, 1H), 3.43 – 3.38 (m, 13H), 3.20 – 3.06 (m, 4H), 3.01 – 2.97 (m, 3H), 2.87 (s, 5H), 2.54 – 2.46 (m, 10H), 2.40 (t, *J* = 8.1 Hz, 3H), 2.30 – 2.23 (m, 3H), 2.04 (quint, *J* = 8.1 Hz, 4H), 1.48 (t, *J* = 6.9 Hz, 3H), 1.04 – 1.02 (m, 11H).

#### Synthesis of E-11



*EZ*-**8** (8.0 mg, 5.6  $\mu$ mol) and trifluoroacetic acid (30  $\mu$ L, 0.392 mmol) were dissolved in CHCl<sub>3</sub> (5.6 mL) and stirred at room temperature for 23 hours. Triethylamine (109  $\mu$ L, 0.784 mmol) was added and the mixture

was washed with a saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.2% Et<sub>3</sub>N), 50%) afforded E-11 as a yellow solid (1.6 mg, 1.2 µmol, 22%) and Z-11 as a yellow solid (1.2 mg, 0.93 µmol, 17%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~88:12 *cis:trans* hydrazone amide rotamers, major rotamer reported. δ 16.28 (s, 1H, H<sub>22</sub>), 9.93 (s, 1H, H<sub>38</sub>), 9.20 (s, 1H, H<sub>27</sub>), 9.05 (s, 1H, H<sub>10</sub>), 8.31 (d, J = 8.4 Hz, 1H, H<sub>25</sub>), 8.25 (s, 1H, H<sub>11</sub>), 8.22 (s, 2H, H<sub>2</sub>+ H<sub>18</sub>), 8.09 (d, J = 9.6 Hz, 1H, H<sub>26</sub>), 8.03 (d, J = 1.3 Hz, 1H, H<sub>13</sub>), 7.99 (s, 1H,  $H_1$ ), 7.97 (d, J = 7.8 Hz, 1H,  $H_{19}$ ), 7.91 (d, J = 7.2 Hz, 1H,  $H_5$ ), 7.72 (d, J = 7.2 Hz, 1H,  $H_3$ ), 7.70 – 7.66 (m, 3H, H<sub>6</sub>+ H<sub>8</sub>+ H<sub>15</sub>), 7.63 – 7.62 (m, 1H, H<sub>20</sub>), 7.59 – 7.53 (m, 4H, H<sub>4</sub>+ H<sub>9</sub>+ H<sub>16</sub>+ H<sub>17</sub>), 7.49 (d, J = 1.3 Hz, 1H,  $H_{12}$ ), 7.46 – 7.37 (m, 4H,  $H_{21}$ +  $H_{30}$ +  $H_{46}$ ), 7.28 – 7.26 (m, 5H,  $H_{28}$ +  $H_{31}$ +  $H_{45}$ +  $H_{47}$ ), 7.25 – 7.24 (m, 1H,  $H_{37}$ ), 7.22 – 7.21 (m, 1H, H<sub>40</sub>), 6.92 (d, J = 7.2 Hz, 1H, H<sub>29</sub>), 6.13 (d, J = 7.8 Hz, 1H, H<sub>42</sub>), 5.73 (q, J = 6.6 Hz, 1H, H<sub>36</sub>), 4.84 (q, J = 7.2 Hz, 1H, H<sub>39</sub>), 4.49 (quint., J = 6.6 Hz, 1H, H<sub>41</sub>), 4.44 (q, J = 6.6 Hz, 1H, H<sub>23</sub>), 4.18 -4.12 (m, 2H, H<sub>32</sub>), 3.56 (dd, J = 13.2, 4.2 Hz, 1H, H<sub>35</sub>), 3.25 – 3.17 (m, 2H, H<sub>35</sub>'+ H<sub>44</sub>), 3.16 – 3.06 (m, 1H, H<sub>44</sub>'), 3.10 – 2.85 (m, 2H, H<sub>34</sub>), 2.52 (s, 3H, H<sub>7</sub>), 2.51 (s, 3H, H<sub>14</sub>), 2.33 – 2.21 (m, 2H, H<sub>33</sub>), 1.46 (t, *J* = 7.2 Hz, 3H, H<sub>24</sub>), 1.32 (d, J = 7.2 Hz, 3H, H<sub>48</sub>), 1.14 (s, 9H, H<sub>43</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 178.6, 172.3, 170.9, 170.5, 165.8, 159.5, 151.1, 150.3, 149.9, 148.4, 145.8, 145.2, 142.9, 141.9, 141.8, 141.7, 141.3, 140.6, 139.2, 138.9, 138.3, 137.5, 136.9, 136.5, 136.5, 136.4, 136.2, 135.1, 134.7, 134.6, 134.5, 133.7, 133.1, 130.9, 130.7, 130.4, 129.8, 129.6, 129.4, 129.3, 129.2, 128.8, 128.6, 128.3, 128.1, 127.7, 126.9, 126.1, 125.3, 125.2, 124.5, 123.7, 122.6, 120.4, 119.6, 115.1, 111.9, 66.2, 61.2, 54.7, 51.0, 48.7, 45.7, 38.6, 38.3, 34.6, 31.9, 31.6, 30.3, 30.1, 29.7, 29.4, 28.6, 27.4, 27.2, 22.7, 20.9, 18.1, 14.51. **HRMS** (ESI<sup>+</sup>): Calc. for  $C_{75}H_{72}DN_9O_8S_2Na$ : 1315.4978, found 1315.3048 [M+Na]<sup>+</sup>. **HRMS** (ESI<sup>-</sup>): Calc. for  $C_{75}H_{71}DN_9O_8S_2$ : 1291.5014, found 1291.3281 [M-H]<sup>-</sup>.

#### Synthesis of Z-11



*EZ*-**8** (8.0 mg, 5.6  $\mu$ mol) and trifluoroacetic acid (1  $\mu$ L, 17  $\mu$ mol) were dissolved in CHCl<sub>3</sub> (2.2 mL) and stirred at room temperature for 47 hours. Triethylamine (5  $\mu$ L, 34  $\mu$ mol) was added and the mixture was washed with a saturated aqueous solution of NaHCO<sub>3</sub>. The aqueous layer was extracted

with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. Preparative thin layer chromatography (SiO<sub>2</sub>, EtOAc/CH<sub>2</sub>Cl<sub>2</sub> (0.2% Et<sub>3</sub>N), 50%) afforded Z-11 as a yellow solid (3.1 mg, 2.4 µmol, 43%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) ~10:90 cis:trans hydrazone amide rotamers, major rotamer reported  $\delta$  13.28 (s, 1H, H<sub>22</sub>), 10.26 (s, 1H, H<sub>38</sub>), 10.13 (s, 1H, H<sub>1</sub>), 8.98 (d, J = 1.9 Hz, 1H, H<sub>10</sub>), 8.94 (d, J = 1.5 Hz, 1H, H<sub>27</sub>), 8.47 (t, J= 1.8 Hz, 1H, H<sub>18</sub>), 8.22 (t, J = 1.8 Hz, 1H, H<sub>2</sub>), 8.19 (d, J = 1.9 Hz, 1H, H<sub>11</sub>), 8.17 (dd, J = 8.2, 1.5 Hz, H<sub>26</sub>), 8.01 (d, J = 8.2 Hz, 1H, H<sub>25</sub>), 7.99 - 7.97 (m, 2H, H<sub>3</sub>+ H<sub>5</sub>), 7.90 (m, 1H, H<sub>19</sub>), 7.71 - 7.66 (m, 3H, H<sub>4</sub>+ H<sub>13</sub>+ H<sub>20</sub>), 7.64 (dd, J = 8.0, 1.6 Hz, 1H, H<sub>21</sub>), 7.60 (s, 1H, H<sub>6</sub>), 7.55 – 7.45 (m, 5H, H<sub>8</sub>+ H<sub>9</sub>+ H<sub>15</sub>+ H<sub>29</sub>+ H<sub>31</sub>), 7.43 – 7.37 (m, 4H, H<sub>12</sub>+ H<sub>16</sub>+  $H_{17}+H_{47}$ , 7.34 – 7.31 (m, 3H,  $H_{46}+H_{30}$ ), 7.23 (d, J = 7.2 Hz, 2H,  $H_{45}$ ), 6.92 (dd, J = 7.2, 1.8 Hz, 1H,  $H_{28}$ ), 6.32 (s, 1H, H<sub>40</sub>), 5.93 (s, 1H, H<sub>42</sub>), 5.14 (ddd, J = 10.8, 8.4, 2.4 Hz, 1H, H<sub>36</sub>), 4.64 (q, J = 6.0 Hz, 1H, H<sub>39</sub>),  $4.57 (q, J = 7.2 Hz, 2H, H_{23}), 4.19 - 4.08 (m, 2H, H_{32}), 4.07 - 4.00 (m, 1H, H_{41}), 3.67 - 3.59 (m, 1H, H_{35}),$ 3.48 (dd, J = 14.4, 4.8 Hz, 1H, H<sub>44</sub>), 3.15 (dd, J = 13.2, 10.8 Hz, 1H, H<sub>35'</sub>), 3.08 - 3.01 (m, 2H, H<sub>34</sub>+ H<sub>44'</sub>), 2.95 (ddd, J = 13.2, 10.2, 5.4 Hz, 1H, H<sub>34'</sub>), 2.48 (s, 3H, H<sub>7</sub>), 2.47 (s, 3H, H<sub>14</sub>), 2.37 - 2.29 (m, 1H, H<sub>33</sub>), 2.26 – 2.19 (m, 1H, H<sub>33</sub>'), 1.50 – 1.43 (m, 6H, H<sub>24</sub>+ H<sub>48</sub>), 0.99 (s, 9H, H<sub>43</sub>). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 192.5, 181.4, 174.2, 171.2, 167.4, 163.3, 159.6, 149.5, 142.1, 141.8, 141.4, 140.8, 140.5, 139.7, 138.2, 137.1, 136.8, 135.9, 135.7, 135.5, 135.3, 135.2, 134.7, 133.2, 131.1, 130.3, 130.2, 130.2, 129.7, 129.6, 129.5, 129.0, 128.9, 128.8, 128.6, 128.3, 128.1, 127.9, 125.4, 125.1, 119.5, 114.4, 70.7, 66.5, 63.3, 61.8, 54.8, 52.2, 51.9, 38.9, 38.7, 36.1, 34.8, 33.6, 32.9, 32.1, 29.9, 29.5, 27.3, 25.9, 24.9, 22.8, 21.1, 20.9, 17.3, 14.4. HRMS (ESI<sup>+</sup>): Calc. for C<sub>75</sub>H<sub>72</sub>DN<sub>9</sub>O<sub>8</sub>S<sub>2</sub>Na: 1315.4978, found 1315.3048 [M+Na]<sup>+</sup>. HRMS (ESI<sup>-</sup>): Calc. for C<sub>75</sub>H<sub>71</sub>DN<sub>9</sub>O<sub>8</sub>S<sub>2</sub>: 1291.5014, found 1291.3281 [M-H]<sup>-</sup>.

## 3. Effect of Acid Stoichiometry on E-1/Z-1 Interconversion

## **3.1 Titration Procedure**

To a solution of *E*-**1**/*Z*-**1** (1.22 mg, 1.25  $\mu$ mol) in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub> (500  $\mu$ L, 2.5 mM) in a standard NMR tube was added trifluoroacetic acid and the sample was left to equilibrate at r.t. Triethylamine was then added and the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water. The layers were separated and washed with a saturated aqueous solution of ammonium chloride, water, a saturated aqueous solution of sodium bicarbonate, and brine. The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was analysed by <sup>1</sup>H NMR in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub> and compared to authentic samples of *E*-**1** and *Z*-**1** to determine the conversion of the operation (based on the integration of the hydrazone ester peaks).

## 3.2 Results



**Figure 1**. a. Graph showing the effect of acid stoichiometry on the *E*-**1**/*Z*-**1** interconversion (margin of error ±2%). b. *E*-**1** titration results. c. *Z*-**1** titration results.

## 4. Effect of Solvent on E-1/Z-1 Interconversion

## **4.1 Titration Procedure**

To a solution of Z-1 (1.22 mg, 1.25  $\mu$ mol) in the adequate solvent (500  $\mu$ L, 2.5 mM) in a standard NMR tube was added trifluoroacetic acid (8  $\mu$ L, 80 eq.) and the sample was left to equilibrate at r.t. Triethylamine (16  $\mu$ L, 100 eq.) was then added and the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water. The layers were separated and washed with a saturated aqueous solution of ammonium chloride, water, a saturated aqueous solution of sodium bicarbonate, and brine. The

organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was analysed by <sup>1</sup>H NMR in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub> and compared to authentic samples of *E*-**1** and *Z*-**1** to determine the conversion of the operation (based on the integration of the hydrazone ester peaks).

## 4.2 Characterisation of Z-1-H<sub>3</sub><sup>3+</sup>-Left and Z-1-H<sub>3</sub><sup>3+</sup>-Right



#### 4.2.1 Z-1-H<sub>3</sub><sup>3+</sup>-Left in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1)

**Figure 2**. Partial <sup>1</sup>H NMR (600 MHz, 295 K,  $C_2D_2Cl_4:CD_3CN$  (2:1)) spectra of the transformation and intermediates observed upon the addition of 80 equivalents of TFA to *Z*-**1** in  $C_2D_2Cl_4:CD_3CN$  (as an example of polar solvent mixture). a) *Z*-**1** (mixture of acyl hydrazone rotamers); b) *Z*-**1**-H<sub>3</sub><sup>3+</sup>-left 10 mins after the addition; c) *Z*-**1**-H<sub>3</sub><sup>3+</sup>-left 12 h after the addition; d) sample after neutralisation and work-up (*Z*-**1**). Signals from traces of residual solvents are shown in grey. Dashed lines connect resonances indicative of the configuration of the hydrazone switch (H<sub>2</sub>, H<sub>3</sub> and H<sub>4</sub>) and the position of the cargo (H<sub>1</sub> and H<sub>5</sub>).

## <u>2D ROESY spectrum of Z-1-H<sub>3</sub><sup>3+</sup>-left in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1)</u>



Figure 3. 2D ROESY NMR (600 MHz, 295 K, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1)) spectrum of *Z*-1-H<sub>3</sub><sup>3+</sup>-left.



#### 4.2.1 Z-1-H<sub>3</sub><sup>3+</sup>-Right in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:C<sub>6</sub>D<sub>5</sub>CD<sub>3</sub> (1:2)

**Figure 4.** Partial <sup>1</sup>H NMR (600 MHz, 295 K,  $C_2D_2Cl_4:C_6D_5CD_3$  (1:2)) spectra of the transformation and intermediates observed upon the addition of 80 equivalents of TFA to *Z*-**1** in  $C_2D_2Cl_4:C_6D_5CD_3$  (as an example of unpolar solvent mixture). a) *Z*-**1** (mixture of acyl hydrazone rotamers); b) *Z*-**1**-H<sub>3</sub><sup>3+</sup>-left: *Z*-**1**-H<sub>3</sub><sup>3+</sup>-right (81:19) 12 mins after the addition; c) *Z*-**1**-H<sub>3</sub><sup>3+</sup>-left: *Z*-**1**-H<sub>3</sub><sup>3+</sup>-right (45:55) 1 h after the addition; d) *Z*-**1**-H<sub>3</sub><sup>3+</sup>-left: *Z*-**1**-H<sub>3</sub><sup>3+</sup>-right (17:83) 5 h after the addition; e) sample after neutralisation and work-up (*E*-**1**:*Z*-**1** 83:17). Signals from traces of residual solvents are shown in grey. Dashed lines connect resonances indicative of the configuration of the hydrazone switch (H<sub>2</sub>, H<sub>3</sub> and H<sub>4</sub>) and the position of the cargo (H<sub>1</sub> and H<sub>5</sub>).

#### <u>2D ROESY spectrum of Z-1-H<sub>3</sub><sup>3+</sup>-right in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:C<sub>6</sub>D<sub>5</sub>CD<sub>3</sub> (2:1)</u>



Figure 5. 2D ROESY NMR (600 MHz, 295 K, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:C<sub>6</sub>D<sub>5</sub>CD<sub>3</sub> (1:2)) spectrum of Z-1-H<sub>3</sub><sup>3+</sup>-right

## 5. Optimised Backward Transport (E-1 to Z-1)

## **5.1 General Procedure**

To a solution of *E*-**1** (1.22 mg, 1.25 µmol) in the C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1, 500 µL, 2.5 mM) in a standard NMR tube was added trifluoroacetic acid (8 µL, 80 eq.) and the sample was left to equilibrate at r.t. Triethylamine (16 µL, 100 eq.) was then added and the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water. The layers were separated; the organic layer was washed with saturated aqueous ammonium chloride solution, water, saturated aqueous sodium bicarbonate solution, and brine. The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was analysed by <sup>1</sup>H NMR in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub> and compared to authentic samples of *E*-**1** and *Z*-**1** to determine the conversion of the operation (based on the integration of the hydrazone ester peaks).

#### 5.2 <sup>1</sup>H NMR Spectra



**Figure 6.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1)) spectra of the conversion of *E*-1 to *Z*-1 with 80 equivalents of TFA. a) *E*-1; b) *Z*-1-H<sub>3</sub><sup>3+</sup>-right:*E*-1 (82:18) 10 mins after the addition; c) *Z*-1-H<sub>3</sub><sup>3+</sup>-left: *Z*-1-H<sub>3</sub><sup>3+</sup>-right (18:82) 1 h after the addition; d) *Z*-1-H<sub>3</sub><sup>3+</sup>-left: *Z*-1-H<sub>3</sub><sup>3+</sup>-left: *Z*-1-H<sub>3</sub><sup>3+</sup>-right (72:28) 5 h after the addition; e) *Z*-1-H<sub>3</sub><sup>3+</sup>-left: *Z*-1-H<sub>3</sub><sup>3+</sup>-right (>98:2) 8 h after the addition; f) sample after neutralisation and work-up (*E*-1:*Z*-1 >2:98). Signals from traces of residual solvents are shown in grey. Dashed lines connect resonances indicative of the configuration of the hydrazone switch (H<sub>2</sub>, H<sub>3</sub> and H<sub>4</sub>) and the position of the cargo (H<sub>1</sub> and H<sub>5</sub>).

## 6. Site-to-Site Transport of Peptide Derivatives 9, 10 and 11

## 6.1 Forward Transport

## 6.1.1 General Procedure

To a solution of Z-9-11 (1.125  $\mu$ mol) in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>/C<sub>6</sub>D<sub>5</sub>CD<sub>3</sub> (1:2, 450  $\mu$ L, 2.5 mM) in a standard NMR tube was added trifluoroacetic acid (7  $\mu$ L, 90  $\mu$ mol, 80 eq.) and the sample (final volume: 450  $\mu$ L, 2.5 mM) was left to equilibrate at r.t.. Triethylamine (16  $\mu$ L, 113  $\mu$ mol, 100 eq.) was then added and the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water. The layers were separated; the organic layer was washed with saturated aqueous ammonium chloride solution, water, saturated aqueous sodium bicarbonate solution, and brine. The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was analysed by <sup>1</sup>H NMR in CDCl<sub>3</sub> and compared to authentic samples of *Z*-9-11 and *E*-9-11 to determine the conversion of the operation.



# **Figure 7.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *Z*-**9** to *E*-**9** using the general procedure described above (**6.1.1**). a) *Z*-**9**; b) sample after neutralisation and work-up (*E*-**9**:*Z*-**9** 66:34). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.



**Figure 8.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *Z*-**10** to *E*-**10** using the general procedure described above (**6.1.1**). a) *Z*-**10**; b) sample after neutralisation and work-up (*E*-**10**:*Z*-**10** 65:35). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.



**Figure 9.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *Z*-**11** to *E*-**11** using the general procedure described above (**6.1.1**). a) *Z*-**11**; b) sample after neutralisation and work-up (*E*-**11**:*Z*-**11** 65:35). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.

## 6.2 Backward Transport

#### 6.2.1 General Procedure

To a solution of *E*-**9-11** (1.125  $\mu$ mol) in C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>:CD<sub>3</sub>CN (2:1, 450  $\mu$ L, 2.5 mM) in a standard NMR tube was added trifluoroacetic acid (7  $\mu$ L, 90  $\mu$ mol, 80 eq.) and the sample was left to equilibrate at r.t. Triethylamine (16  $\mu$ L, 113  $\mu$ mol, 100 eq.) was then added and the reaction mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> and water. The layers were separated; the organic layer was washed with saturated aqueous ammonium chloride solution, water, saturated aqueous sodium bicarbonate solution, and brine. The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The residue was analysed by <sup>1</sup>H NMR in CDCl<sub>3</sub> and compared to authentic samples of *Z*-**9-11** and *E*-**9-11** to determine the conversion of the operation.



#### 6.2.2 <sup>1</sup>H NMR Spectra

**Figure 10.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *E*-**9** to *Z*-**9** using the general procedure described above (**6.2.1**). a) *E*-**9**; b) sample after neutralisation and work-up (*E*-**9**:*Z*-**9** 2:98). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.



**Figure 11.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *E*-**10** to *Z*-**10** using the general procedure described above (**6.2.1**). a) *E*-**10**; b) sample after neutralisation and work-up (*E*-**10**:*Z*-**10** >2:98). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.



**Figure 12.** Partial <sup>1</sup>H NMR (600 MHz, 295 K, CDCl<sub>3</sub>) spectra of the conversion of *E*-**11** to *Z*-**11** using the general procedure described above (**6.2.1**). a) *E*-**11**; b) sample after neutralisation and work-up (*E*-**11**:*Z*-**11** >2:98). Signals from traces of residual solvents and organic salts are shown in grey. Dashed lines connect resonances belonging to the same proton.

## 7. Spectroscopic Data



Spectrum 1. <sup>1</sup>H NMR (600 MHz, DMSO- $d_6$ ) of S1



**Spectrum 2.** <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>) of **S1** 







Spectrum 4.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl<sub>3</sub>) of S2





Spectrum 6.  $^{\rm 13}C$  NMR (151 MHz, CDCl\_3) of S3



Spectrum 8.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl<sub>3</sub>) of 3











Spectrum 16.  $^1\text{H}$  NMR (600 MHz, CDCl\_3) of S5



Spectrum 17.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl<sub>3</sub>) of S5



Spectrum 19.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl\_3) of 4



Spectrum 20.  $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>) of <code>EZ-7</code>







Spectrum 26. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of S6

 110 100 δ (ppm)

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Spectrum 27. <sup>1</sup>H NMR (600 MHz,  $CDCl_3$ ) of S7





Spectrum 29. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of S8



Spectrum 30.  $^{\rm 13}C$  NMR (151 MHz, CDCl\_3) of S8



Spectrum 32.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl<sub>3</sub>) of S9



Spectrum 34.  $^{\rm 13}{\rm C}$  NMR (151 MHz, CDCl<sub>3</sub>) of 5



Spectrum 35. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of EZ-8





## 8. References

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