## **Supporting Information**

# **Cleavable and Tunable Cysteine-Specific Arylation**

# **Modification by Aryl Thioethers**

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## **Table of Contents**

1.	General information	S1
2.	LC-MS and preparative HPLC information	S1
3.	Solid phase peptide synthesis (SPPS) and purification	S3
5.	Optimization of reaction conditions for cysteine arylation	S5
6.	Chemoselectivity determination of arylation modification	S6
7.	Synthesis and characterization of aryl thioethers	S7
8.	Arylation of cysteine-containing peptides/proteins	S15
9.	Stability evaluation of modified peptide	S60
10.	Regeneration activity evaluation of modified peptides	S61
11.	Evaluation of Reaction Kinetics	\$89
12.	BSA modification by $2v$	S92
13.	Secondary labeling of modified peptide	\$94
14.	Spectra of new compounds	\$95
15.	Reference	.S125

#### 1. General information

#### 1.1 Reagents

Commercial available reagents and solvents were purchased from Energy Chemical, J&K Chemical, Innochem or Bidepharm and all these reagents were used directly without further purification unless otherwise noted. RP-HPLC solvents were purchased as HPLC grade from Energy Chemical.

Fmoc-amino acids for solid phase peptide synthesis (SPPS) were used with the following side-chain protection: Fmoc-Arg(Pbf)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asp(*t*Bu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Glu(*t*Bu)-OH, Fmoc-His(Trt)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Ser(*t*Bu)-OH, FmocThr(*t*Bu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Tyr(*t*Bu)-OH.

#### 1.2 Instruments

NMR spectra were recorded on a Bruker AVANCE AV 400 or 500 instruments and all NMR experiments were reported in units, parts per million (ppm), using TMS as internal reference. Data for <sup>1</sup>H NMR are recorded as follows: chemical shift ( $\delta$ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet, q = quartet, dd = doublet of doublets, dt = doublet of triplets, td = triplet of doublets, and br = broad signal), Coupling constants (*J*) were reported as Hertz (Hz). HRMS spectra were recorded with a LCMS-IT-TOF mass spectrometer, equipped with an ESI source. LC-MS spectra were performed on an Agilent Technologies 1260 Infinity II HPLC system, connected to an Agilent G6125B LC/MSD, equipped with an Agilent ZORBAX SB-C18 column, 2.1 × 50 mm, 1.8 µm). Preparative RP-HPLC were performed on Hanbon Sci. &Tech. Newstyle HPLC systems equipped with NP7000 serials pump, NU3000 serials UV/VIS detector and DM-A dynamic mixer, using the following column: Dubhe C18 D18122010.25 column, 12 nm, 10 µm, 20 x 250 mm. TLC analysis was visualized by fluorescence quenching under UV light (254 nm and 365 nm), or developing the plates with I<sub>2</sub>, phosphomolybdic acid (PMA), permanganate. Flash chromatography purifications were performed on silica gel GF254 (200-300 mesh) purchased from Qingdao Haiyang Chemical.

#### 2. LC-MS and preparative HPLC information

#### 2.1 LC-MS analysis

LC-MS measurements were performed on an Agilent Technologies 1260 Infinity II HPLC system with a G7129A 1260 Vislsampler, a G7111B 1260 Quat Pump and a G7114A 1260 VWD detector, connected to an Agilent G6125B LC/MSD, equipped with an Agilent ZORBAX SB-C18 column,  $2.1 \times 50$  mm,  $1.8 \mu$ m). Water (solvent A) and acetonitrile (solvent B), each containing 0.1% formic acid, were used as the mobile phase. Low-resolution mass spectrometric measurements were acquired using the following parameters: positive electrospray ionization (ESI), temperature of drying gas = 350 °C, flow rate of drying gas = 12 L/min, pressure of nebulizer gas = 60 psi, capillary voltage = 4000 V and fragmentor voltage = 70 V.

Following LC methods were used:

Method A (Column: Agilent ZORBAX SB-C18 column,  $2.1 \times 50$  mm,  $1.8 \mu$ m, flow rate 0.3 mL / min)

Time (min)	H <sub>2</sub> O (%)	Acetonitrile (%)
0	95	5
2	95	5
17	70	30
18	0	100
24	0	100

Method B (Column: Agilent ZORBAX SB-C18 column,  $2.1 \times 50$  mm,  $1.8 \mu$ m, flow rate 0.3 mL / min)

Time (min)	H₂O (%)	Acetonitrile (%)
0	100	0
2	100	0
17	70	30
18	0	100
24	0	100

Method C (Column: Agilent Poroshell SB-C18 column,  $3 \times 100$  mm,  $2.7 \mu$ m, flow rate 0.6 mL / min)

Time (min)	H <sub>2</sub> O (%)	Acetonitrile (%)
0	95	5
2	95	5
10	70	30
18	5	95
24	5	95

Method D (Column: Agilent Poroshell SB-C18 column,  $3 \times 100$  mm,  $2.7 \mu$ m, flow rate 0.6 mL / min)

Time (min)	H <sub>2</sub> O (%)	Acetonitrile (%)
0	95	5
2	95	5
17	70	30
18	0	100
24	0	100

All reported LC-MS yields were determined by integrating TIC spectra. The peak areas for all relevant peptide-containing species on the chromatogram were integrated using Agilent software package. The yields were determined as follows: %yield =  $S_{product}/S_{total}$ , where  $S_{product}$  is the peak area of the product and  $S_{total}$  is the peak area of combined peptide-containing species (product, starting material and byproduct).

#### 2.2 Preparative HPLC

Preparative RP-HPLC was performed on a Hanbon Sci.&Tech. Newstyle HPLC system with a NU3000 serials UV/VIS detector, a NP7000 serials pump, a DM-A dynamic mixer and a collector, coupled with a Dubhe C18 column (30 x 250 mm, 10  $\mu$ m). Water (solvent A) and acetonitrile (solvent B), each containing 0.1% TFA, were used as the mobile phase.

Method E (Coldmin. Duble C18 coldmin, 30 x 250 min, 10 µm, now rate 20 mE7 min)			
Time (min)	H <sub>2</sub> O (%)	Acetonitrile (%)	
0	95	5	
5	95	5	
35	60	40	
45	0	100	
60	0	100	

Following LC method was used: Method E (Column: Dubhe C18 column, 30 x 250 mm, 10 µm, flow rate 20 mL / min)

### 3. Solid phase peptide synthesis (SPPS) and purification

Peptide synthesis was carried out manually using standard Fmoc SPPS-chemistry and 2-Chloro-trityl chloride (2-CTC) resin (0.98 mmol/g resin, 0.2 mmol) or Rink amide resin (0.61 mmol/g resin, 0.2 mmol). Peptides **1m/1n** were made via Rink amide resin and other peptides were made via 2-chloro-trityl chloride resin.

Preloading 2-chloro-trityl chloride resin: 2-CTC resin was swollen in anhydrous DCM for 10 min, then washed with DCM ( $3 \times 5$  mL). A solution of Fmoc-AA-OH (4 equiv.) and DIPEA (8 equiv.) in anhydrous DCM (4 mL) was added to the resin and the suspension was shaken at room temperature for 2 hours. The resin was washed with DCM ( $3 \times 5$  mL), DMF ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL). The resin was capped with a solution of DCM/MeOH/DIPEA (17/2/1, v/v/v, 4 mL) for 30 min and washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL).

Preloading Rink amide resin: Rink amide resin was swollen in anhydrous DCM for 10 min, then washed with DCM ( $3 \times 5$  mL) and DMF ( $3 \times 5$  mL). Then the resin was treated with piperidine/DMF (1/4, v/v,  $2 \times 4$  mL,  $2 \times 5$  min) at room temperature and washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL). A solution of Fmoc-AA-OH (4 equiv.), HOBT (4 equiv.) and collidine (8 equiv.) in anhydrous DMF (4 mL) was added to the resin and the suspension was shaken at room temperature for 2 hours. The resin was washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL). The resin was capped with acetic anhydride/pyridine (1/9, v/v) (30 min) and washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL).

Fmoc-deprotection: The washed resin was treated with piperidine/DMF (1/4, v/v, 2 × 4 mL,  $2 \times 5$  min) at room temperature and then washed with DMF (3 × 5 mL), MeOH (3 × 5 mL) and DCM (3 × 5 mL).

Amino acid coupling and Fmoc-deprotection: A solution of protected amino acid (4 equiv.),

HATU (4 equiv.), HOBT (4 equiv.) and collidine (8 equiv.) in DMF (4 mL) was added to the resin. The reaction mixture was agitated at room temperature for 2 hours. After the reaction was completed, the resin-bound peptide was washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL). Then the washed resin was treated with piperidine/DMF (1/4, v/v, 2 × 4 mL, 2 × 5 min) at room temperature and then washed with DMF ( $3 \times 5$  mL), MeOH ( $3 \times 5$  mL) and DCM ( $3 \times 5$  mL).

Peptide cleavage and deprotection: Peptides were deprotected and cleaved from the resin under reducing conditions, by treatment with 2.5% v/v water, 2.5% v/v *i*-Pr<sub>3</sub>SiH, 2.5% v/v ethanethiol and 2.5% v/v thioanisole in neat trifluoroacetic acid (4 mL). The resulting mixture was shaken for 1 hours at room temperature. The resin was removed by filtration and washed with TFA (2 × 3 mL). The combined cleavage solutions were concentrated in vacum.

Peptide purification and analysis: Peptides were dissolved in water with a minimum amount of organic co-solvent (acetonitrile). Then purified by preparative RP-HPLC (Method D). Fractions containing the desired peptide were lyophilized. The purity was assessed by analyzing a peptide water solution by LC-MS. At the same time, low-resolution mass spectrometric measurements were also acquired.

# 5. Optimization of reaction conditions for cysteine arylation

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	H₂N–	А Г С А-соон SH	N N N H 2 (equiv.) buffer (x mL), pH (y % v/v DMSO)	H₂N- <b>A</b> (F) 3a		
	R= ई-Cl	ξ− <b>S−</b> {− <b>S</b> −	 √NO₂ §−S−	N O	§−s– <mark>×</mark>	
	2aa	2ab	2ac	2ad	2ae	
Entry	<b>2</b> (eq.)	buffer	рН	х	У	Yield (%) <sup>b</sup>
1	<b>2aa</b> (1.5 eq.)	100 mM PBS	8.0	0.5	10	n.d.
2	<b>2ab</b> (1.5 eq.)	100 mM PBS	8.0	0.5	10	n.d.
3	<b>2ac</b> (1.5 eq.)	100 mM PBS	8.0	0.5	10	n.d.
4	<b>2ad</b> (1.5 eq.)	100 mM PBS	8.0	0.5	10	43
5	<b>2ae</b> (1.5 eq.)	100 mM PBS	8.0	0.5	10	17
6	<b>2ad</b> (1.5 eq.)	100 mM HEPE	S 8.0	0.5	10	41
7	<b>2ad</b> (1.5 eq.)	100 mM Tris	8.0	0.5	10	47
8	<b>2ad</b> (1.5 eq.)	100 mM Tris	7.4	0.5	10	35
9	<b>2ad</b> (1.5 eq.)	100 mM Tris	7.6	0.5	10	39
10	<b>2ad</b> (1.5 eq.)	100 mM Tris	7.8	0.5	10	43
11	<b>2ad</b> (1.5 eq.)	100 mM Tris	8.2	0.5	10	47
12	<b>2ad</b> (1.5 eq.)	100 mM Tris	8.4	0.5	10	46
13	<b>2ad</b> (1.5 eq.)	200 mM Tris	8.0	0.5	10	46
14	<b>2ad</b> (2.0 eq.)	100 mM Tris	8.0	0.5	10	55
15	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	0.5	10	64 (85)
16	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	0.5	5	62 (87)
17	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	1.0	5	54 (82)
18	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	0.5	2	53 (79)
19	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	1.0	2	57 (89)
20	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	0.5	1	45 (80)
21	<b>2ad</b> (3.0 eq.)	100 mM Tris	8.0	1.0	1	59 (90 <i>,</i> 83°)

Table S1: Optimization of reaction conditions for cysteine arylation<sup>a</sup>

a) Reaction conditions: 1.0  $\mu$ mol **1a** and **2** (eq.) in non-degassed buffer at room tempeture for 1 h. b) Reported yields are LC-MS yields after 1 h (The yields in parentheses correspond to the yields after 5 h of reaction). c) Isolated yield.

# 6. Chemoselectivity determination of arylation modification



Figure S1. Comparison of 1D <sup>1</sup>H NMR spectra of peptides 1a and 3aa.



# 7. Synthesis and characterization of aryl thioethers



Compound **2aa** is commercial available. Compound **2ab**<sup>[1]</sup>, **2g**<sup>[2]</sup> were synthesized according to reported literature.

General procedure for 2ac-2k:



The starting chloro compound (S-1) in solvent was heated under reflux with aryl mercaptan (S-2, 1.5 eq.) in the presence of base (1.5 eq.), monitored by TLC until reaction completion. Cooled down to room temperature, concentrated in vacuo, and the residue was purified by silica gel (200–300 mesh) column chromatography to afford the desired compound.

General procedure for S-4, S-5, 2m, 2o-2q:



The starting compound (**2ad** or **2i**, 0.2 mmol) and  $K_2CO_3$  (1.2 eq.) were dissolved in DMF (2 mL). Then corresponding bromo compound (1.2 eq.) was added and the reaction mixture was stirred at room temperature until reaction completion. The suspension was diluted with EtOAc (20 mL) and washed successively with saturated aqueous NaHCO<sub>3</sub> (10 mL), water (10 mL) and brine (10 mL). The organic extract was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel (200–300 mesh) column chromatography to afford the desired compound.

General procedure for 2r, 2t-2z:



2-((1H-pyrazolo[3,4-d]pyrimidin-4-yl)thio)benzo[d]oxazole (**2i**, 0.2 mmol) and triphenylphosphine (PPh<sub>3</sub>, 1.5 eq.) were dissolved in dry THF (2 mL) under N<sub>2</sub> and cooled to 0 °C. The alcohol (1.5 eq.) was added immediately followed by dropwise addition of diisopropyl azodiformate (DIAD, 1.5 eq.). Stirring at room temperature or 60 °C until reaction completion. The reaction mixture was evaporated in vacuo and purified by silica gel (200–300 mesh)

column chromatography to afford the desired compound.



**6-((4-nitrophenyl)thio)-9H-purine (2ac):** According to the general procedure, using *i*-PrOH as solvent and  $(CH_3)_3COK$  as base, affording the **2ac** as a yellow solid. 86% yield. <sup>1</sup>H NMR (500 MHz, DMSO)  $\delta$  13.72 (s, 1H), 8.65 (s, 1H), 8.59 (s, 1H), 8.30 (d, *J* = 8.0 Hz, 2H), 7.94 (d, *J* = 8.0 Hz, 2H). <sup>13</sup>C NMR (125 MHz, DMSO)  $\delta$  152.08, 148.01, 137.25, 135.60, 124.37. HRMS (ESI) m/z calcd for C<sub>11</sub>H<sub>8</sub>N<sub>5</sub>O<sub>2</sub>S<sup>+</sup> [M+H]<sup>+</sup> 274.0393, found 270.0389.



**2-((9***H***-purin-6-yl)thio)benzo[***d***]oxazole (2ad):** According to the general procedure, using *i*-PrOH as solvent and  $(CH_3)_3COK$  as base, affording the **2ad** as a white solid. 86% yield. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  13.85 (s, 1H), 8.71 (s, 1H), 8.59 (s, 1H), 7.86 (dd, *J* = 7.2, 1.6 Hz, 1H), 7.79 (dd, *J* = 7.2, 1.6 Hz, 1H), 7.55 – 7.44 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  155.99, 152.62, 152.21, 145.74, 141.83, 126.84, 125.50, 120.50, 111.55.HRMS (ESI) m/z calcd for C<sub>12</sub>H<sub>8</sub>N<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 270.0444, found 270.0441.



**2-((9***H***-purin-6-yl)thio)benzo[***d***]thiazole (2ae):** According to the general procedure, using *i*-PrOH as solvent and (CH<sub>3</sub>)<sub>3</sub>COK as base, affording the **2ae** as a white solid. 76% yield. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  8.87 (s, 1H), 8.65 (s, 1H), 8.17 (d, *J* = 7.2 Hz, 1H), 8.04 (d, *J* = 7.6 Hz, 1H), 7.62 – 7.54 (m, 1H), 7.53 – 7.48 (m, 1H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  158.54, 151.74, 151.68, 145.56, 136.56, 126.97, 125.98, 122.73, 122.32. HRMS (ESI) m/z calcd for C<sub>12</sub>H<sub>8</sub>N<sub>5</sub>S<sub>2</sub><sup>+</sup> [M+H]<sup>+</sup> 286.0216, found 286.0209.



**2-((2-chloro-9H-purin-6-yl)thio)benzo[d]oxazole (2b):** According to the general procedure, using *i*-PrOH as solvent and  $(CH_3)_3COK$  as base, affording the **2b** as a white solid. 64% yield. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  14.02 (s, 1H), 8.60 (s, 1H), 7.91 – 7.85 (m, 1H), 7.84 – 7.79 (m, 1H), 7.58 – 7.44 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  155.19, 152.60, 152.08, 141.74, 127.03, 125.60, 120.61, 111.60. HRMS (ESI) m/z calcd for  $C_{12}H_7CIN_5OS^+$  [M+H]<sup>+</sup> 304.0054, found 304.0054.



**6-(benzo[d]oxazol-2-ylthio)-9H-purin-2-amine (2c):** According to the general procedure, using DMF as solvent and  $(CH_3)_3COK$  as base, affording the **2c** as a white solid. 50% yield. <sup>1</sup>H NMR (400 MHz, DMSO) δ 12.74 (s, 1H), 7.95 (s, 1H), 7.81 (dd, *J* = 6.8, 1.6 Hz, 1H), 7.74 (dd, *J* = 6.8, 1.6 Hz, 1H), 7.50 – 7.38 (m, 2H), 6.53 (s, 2H). <sup>13</sup>C NMR (100 MHz, DMSO) δ 160.27, 152.55, 141.91, 141.33, 126.50, 125.31, 120.29, 111.46. HRMS (ESI) m/z calcd for  $C_{12}H_9N_6OS^+$  [M+H]<sup>+</sup> 285.0553, found

285.0546.



**2-(pyrimidin-4-ylthio)benzo[***d***]oxazole (2d):** According to the general procedure, using EtOH as solvent and triethylamine as base, affording the **2d** as a white solid. 66% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.01 (s, 1H), 8.62 (d, *J* = 5.6 Hz, 1H), 7.84 – 7.72 (m, 2H), 7.62 – 7.53 (m, 1H), 7.45 – 7.36 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  165.89, 158.47, 156.84, 156.54, 152.01, 141.53, 126.13, 125.09, 120.18, 119.92, 110.74. HRMS (ESI) m/z calcd for C<sub>11</sub>H<sub>8</sub>N<sub>3</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 230.0383, found 230.0379.



**2-(quinazolin-4-ylthio)benzo**[*d*]**oxazole (2e):** According to the general procedure, using EtOH as solvent and KOH as base, affording the **2e** as a white solid. 85% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.92 (s, 1H), 8.12 (dd, *J* = 8.4, 0.4 Hz, 1H), 8.06 (d, *J* = 8.4 Hz, 1H), 7.99 – 7.91 (m, 1H), 7.86 – 7.81 (m, 1H), 7.74 – 7.67 (m, 1H), 7.63 – 7.57 (m, 1H), 7.49 – 7.37 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  167.63, 155.03, 153.88, 153.06, 149.17, 141.98, 134.68, 129.20, 128.41, 126.33, 124.91, 123.86, 123.54, 120.65, 110.96. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>10</sub>N<sub>3</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 280.0539, found 280.0529.



**2-((2-chloroquinazolin-4-yl)thio)benzo**[*d*]**oxazole (2f):** According to the general procedure, using THF as solvent and triethylamine as base, affording the **2f** as a white solid. 52% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.10 (d, *J* = 8.4 Hz, 1H), 8.00 – 7.92 (m, 2H), 7.84 (d, *J* = 7.2 Hz, 1H), 7.73 – 7.66 (m, 1H), 7.62 (d, *J* = 7.6 Hz, 1H), 7.51 – 7.39 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.24, 155.90, 153.85, 153.08, 150.67, 141.84, 135.78, 128.56, 128.39, 126.56, 125.02, 123.93, 121.72,

120.69, 111.00. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>9</sub>ClN<sub>3</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 314.0149, found 314.0135.



**2-((2-chloro-7-methyl-7***H***-purin-6-yl)thio)benzo[***d***]oxazole (2h): According to the general procedure, using DMF as solvent and KOH as base, affording the <b>2h** as a white solid. 30% yield. <sup>1</sup>H NMR (400 MHz, DMSO) δ 8.81 (s, 1H), 7.94 – 7.70 (m, 2H), 7.60 – 7.33 (m, 2H), 4.12 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO) δ 162.58, 156.29, 153.30, 152.49, 152.04, 148.04, 141.55, 126.86, 125.68, 125.00, 120.40, 111.51, 34.77. HRMS (ESI) m/z calcd for C<sub>13</sub>H<sub>9</sub>ClN<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 318.0211, found

318.0204.



**2-((1***H***-pyrazolo[3,4-***d***]pyrimidin-4-yl)thio)benzo[***d***]oxazole (2i): According to the general procedure, using THF as solvent and triethylamine as base, affording the <b>2i** as a white solid. 51% yield. <sup>1</sup>H NMR (400 MHz, DMSO) δ 14.41 (s, 1H), 8.74 (s, 1H), 8.11 (s, 1H), 7.91 (d, *J* = 7.6 Hz, 1H), 7.84 (d, *J* = 8.0 Hz, 1H), 7.67 – 7.37 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO) δ 155.15, 154.77, 152.61, 141.67, 127.32, 125.81, 120.78, 111.77. HRMS (ESI) m/z calcd for C<sub>12</sub>H<sub>8</sub>N<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 270.0444, found 270.0432.



**2-((6-chloro-1***H***-pyrazolo[3,4-***d***]pyrimidin-4-yl)thio)benzo[***d***] oxazole <b>(2j):** According to the general procedure, using THF as solvent and triethylamine as base, reacted in -20 °C affording the **2j** as a white solid. 24% yield. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  14.56 (s, 1H), 8.08 (s, 1H), 7.94 (d, *J* = 7.6 Hz, 1H), 7.88 (d, *J* = 8.0 Hz, 1H), 7.64 – 7.47 (m, 2H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  162.40, 155.77, 154.92, 154.33, 152.58, 141.55, 133.21, 127.54, 125.92, 120.90, 111.80, 111.00. HRMS (ESI) m/z calcd

for C<sub>12</sub>H<sub>7</sub>ClN<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 304.0054, found 304.0054.



**4-(benzo[***d***]oxazol-2-ylthio)-1***H*-**pyrazolo**[**3**,**4**-*d*]**pyrimidin-6-amine** (**2k**): According to the general procedure, using THF as solvent and triethylamine as base, affording the **2k** as a white solid. 48% yield. <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.20 (s, 1H), 7.88 (d, *J* = 7.6 Hz, 1H), 7.83 (d, *J* = 8.0 Hz, 1H), 7.58 – 7.45 (m, 2H), 7.30 (s, 1H), 6.95 (s, 2H). <sup>13</sup>C NMR (100 MHz, DMSO) δ 161.46, 159.54, 156.97, 156.13, 152.46, 141.64, 132.23, 127.15, 125.76, 120.64, 111.73, 105.90. HRMS (ESI) m/z calcd for C<sub>12</sub>H<sub>9</sub>N<sub>6</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 285.0553, found 285.0548.





**2-((9-(3-azidopropyl)-9H-purin-6-yl)thio)benzo[***d***]oxazole (2I): S-4** was synthesized according to the general procedure. Then **S-4** (69.3 mg, 1.8 mmol) and NaN<sub>3</sub> (1.1 eq) was stirred in DMF (2 mL) at room temperature until reaction completion. The suspension was diluted with EtOAc (20 mL) and washed successively with saturated aqueous NaHCO<sub>3</sub> (10 mL), water (10 mL) and brine (10 mL). The organic extract was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under

reduced pressure. The crude residue was purified by silica gel (200–300 mesh) column chromatography to afford the desired compound **2I** as a white solid. 38% yield for two steps. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.70 (s, 1H), 7.99 (s, 1H), 7.86 – 7.76 (m, 1H), 7.60 – 7.50 (m, 1H), 7.47 – 7.34 (m, 2H), 4.38 (t, *J* = 6.8 Hz, 2H), 3.36 (t, *J* = 6.4 Hz, 2H), 2.24 – 2.11 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  156.06, 155.76, 152.76, 152.22, 150.25, 144.36, 141.98, 131.78, 126.07, 124.77, 120.49, 110.84, 48.11, 41.43, 28.89. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>13</sub>N<sub>8</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 353.0928, found 353.0922.

#### 2-((9-(2-(2-methoxyethoxy)ethyl)-9H-purin-6-yl)thio) benzo[d]oxazole (2m): According to



the general procedure affording the **2m** as a white solid. 83% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.69 (s, 1H), 8.21 (s, 1H), 7.84 – 7.77 (m, 1H), 7.59 – 7.51 (m, 1H), 7.45 – 7.34 (m, 2H), 4.46 (t, *J* = 4.8 Hz, 2H), 3.84 (t, *J* = 4.8 Hz, 2H), 3.63 – 3.56 (m, 2H), 3.52 – 3.44 (m, 2H), 3.33 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  156.02, 155.38, 152.75, 151.99, 150.18, 145.67, 141.99, 131.62, 125.97, 124.71, 120.45, 110.80, 71.77, 70.57, 68.93, 59.01, 43.84. HRMS (ESI) m/z calcd for C<sub>17</sub>H<sub>18</sub>N<sub>5</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup> 372.1125, found

372.1109.



N-(3-(6-(benzo[d]oxazol-2-ylthio)-9H-purin-9-yl)propyl)-5-((3aS,4S,6aR)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide (2n): S-5 was synthesized according to the general procedure. Then S-5 (85.3 mg, 0.2 mmol) was dissolved in DCM (3 mL), added with TFA (1 mL) and the reactant stirred in room temperature for 1h. Reaction mixture concentrated under reduced pressure without further purification. The residue dissolved in DMF (2 mL), followed by adding with DIEA (3 eq), D-biotin (1.2 eq) and HATU (1.2 eq), the reactant stirred in room temperature for 2h. The reactant was diluted with water (20 mL) and extracted by DCM ( $3 \times 20$  mL). The combine organic phase was concentrated under reduced pressure and purified by silica gel (200-300 mesh) column chromatography to afford the desired compound **2n** as a white solid. 56% yield for three steps. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.66 (s, 1H), 8.25 (s, 1H), 7.81 – 7.71 (m, 1H), 7.59 – 7.51 (m, 1H), 7.45 – 7.33 (m, 2H), 7.23 (t, J = 5.6 Hz, 1H), 6.91 (s, 1H), 6.12 (s, 1H), 4.54 - 4.41 (m, 1H), 4.35 - 4.24 (m, 3H), 3.30 - 3.16 (m, 2H), 3.14 - 3.06 (m, 1H), 2.86 (dd, J = 12.8, 4.8 Hz, 1H), 2.67 (d, J = 12.8 Hz, 1H), 2.20 (t, J = 7.2 Hz, 2H), 2.14 -2.02 (m, 2H), 1.78 – 1.53 (m, 2H), 1.44 – 1.34 (m, 2H), 1.33 – 1.12 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 173.87, 164.39, 155.71, 155.50, 152.71, 151.93, 150.30, 145.36, 141.82, 131.69, 126.19, 124.87, 120.35, 110.91, 61.83, 60.17, 55.87, 41.88, 40.66, 36.11, 35.89, 29.93, 28.27, 28.03, 25.67. HRMS (ESI) m/z calcd for  $C_{25}H_{29}N_8O_3S_2^+$  [M+H]<sup>+</sup> 553.1799, found 553.1800.



**2-((1-allyl-1***H***-pyrazolo[3,4-***d***]pyrimidin-4-yl)thio)benzo[***d***] oxazole (2o): According to the general procedure affording the <b>2m** as a white solid. 63% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.74 (s, 1H), 7.93 – 7.78 (m, 1H), 7.65 (s, 1H), 7.62 – 7.54 (m, 1H), 7.52 – 7.34 (m, 2H), 6.25 – 5.94 (m, 1H), 5.33 – 5.16 (m, 2H), 5.12 – 5.03 (m, 2H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.83, 155.02, 154.29, 152.60, 151.87, 141.75, 131.70, 131.50, 126.63, 125.21, 120.73, 118.93, 112.58, 111.02, 49.80. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>12</sub>N<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 310.0757, found 310.0746.



**2-((1-(prop-2-yn-1-yl)-1H-pyrazolo[3,4-d]pyrimidin-4-yl)thio) benzo[d]oxazole (2p):** According to the general procedure affording the **2p** as a white solid. 60% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.77 (s, 1H), 7.84 (dd, *J* = 6.8, 2.4 Hz, 1H), 7.68 (s, 1H), 7.59 (dd, *J* = 6.8, 2.0 Hz, 1H), 7.51 - 7.40 (m, 2H), 5.27 (d, *J* = 2.8 Hz, 2H), 2.41 (t, *J* = 2.4 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  161.18, 154.81, 154.56, 152.61, 151.89, 141.71, 132.18, 126.73, 125.28, 120.77, 112.70, 111.07, 76.29, 73.78, 36.95. HRMS (ESI)

m/z calcd for  $C_{15}H_{10}N_5OS^+$  [M+H]<sup>+</sup> 308.0601, found 308.0597.



Ethyl 2-(4-(benzo[*d*]oxazol-2-ylthio)-1*H*-pyrazolo[3,4-*d*] pyrimidin-1yl)acetate (2q): According to the general procedure affording the 2q as a white solid. 69% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.74 (s, 1H), 7.88 – 7.81 (m, 1H), 7.72 (s, 1H), 7.63 – 7.56 (m, 1H), 7.49 – 7.38 (m, 2H), 5.24 (s, 2H), 4.24 (q, *J* = 7.2 Hz, 3H), 1.27 (t, *J* = 7.2 Hz, 4H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 166.98, 161.08, 154.84, 154.59, 152.85, 152.63, 141.75, 132.26, 126.66, 125.22, 120.75, 112.68, 111.05, 62.14, 48.44,

14.09. HRMS (ESI) m/z calcd for C<sub>16</sub>H<sub>14</sub>N<sub>5</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup> 356.0812, found 356.0803.



**2-((1-(3-bromopropyl)-1H-pyrazolo[3,4-d]pyrimidin-4-yl)thio) benzo[d]oxazole (2r):** According to the general procedure affording the **2r** as a white solid. 89% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.74 (s, 1H), 7.86 – 7.80 (m, 1H), 7.64 (s, 1H), 7.63 – 7.56 (m, 1H), 7.50 – 7.40 (m, 2H), 4.65 (t, *J* = 6.4 Hz, 2H), 3.39 (t, *J* = 6.4 Hz, 2H), 2.50 (p, *J* = 6.4 Hz, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  160.94, 154.95, 154.32, 152.60, 152.12, 141.72, 131.52, 126.69, 125.26, 120.76, 112.55, 111.05, 45.72,

32.24, 29.59. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>13</sub>BrN<sub>5</sub>OS<sup>+</sup> [M+H]<sup>+</sup> 390.0019, found 390.0017.



**2-((1-(3-azidopropyl)-1***H***-pyrazolo[3,4-***d***]pyrimidin-4-yl)thio) benzo[***d***]oxazole (2s): According to the general procedure affording the 2s as a white solid. 61% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) \delta 8.74 (s, 1H), 7.93 – 7.76 (m, 1H), 7.65 (s, 1H), 7.62 – 7.54 (m, 1H), 7.53 – 7.37 (m, 2H), 4.58 (t,** *J* **= 6.8 Hz, 2H), 3.34 (t,** *J* **= 6.8 Hz, 2H), 2.19 (p,** *J* **= 6.8 Hz, 1H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) \delta 160.92, 154.96, 154.32, 152.60, 152.09, 141.74, 131.48, 126.66, 125.24, 120.74, 112.53, 111.03, 48.54, 44.58, 28.79. HRMS (ESI) m/z calcd for C<sub>15</sub>H<sub>13</sub>N<sub>8</sub>O<sub>3</sub>S<sup>+</sup> [M+H]<sup>+</sup> 353.0928,** 

found 353.0925.



*tert*-Butyl (2-(4-(benzo[*d*]oxazol-2-ylthio)-1*H*-pyrazolo[3,4*d*]pyrimidin-1-yl)ethyl)carbamate (2t): According to the general procedure affording the 2t as a white solid. 70% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.72 (s, 1H), 7.90 – 7.79 (m, 1H), 7.65 (s, 1H), 7.62 – 7.56 (m, 1H), 7.50 – 7.40 (m, 2H), 4.97 (s, 1H), 4.80 – 4.46 (t, *J* = 5.2 Hz, 2H), 3.75 – 3.57 (m, 2H), 1.37 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 160.81, 155.65, 154.95, 154.29, 152.57, 152.45, 141.69, 131.58, 126.66, 125.23, 120.72, 112.54, 111.03, 79.62, 47.44, 40.16, 28.29 (s). HRMS (ESI) m/z calcd for  $C_{19}H_{21}N_6O_3S^+$  [M+H]<sup>+</sup> 413.1390, found 413.1374.



**2-((1-(2-(2-(2-ethoxyethoxy)ethoxy)ethyl)-1***H***-pyrazolo[3,4-***d***] pyrimidin-4-yl)thio)benzo[***d***]thiazole (2u): According to the general procedure affording the 2u as a white solid. 71% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) \delta 8.73 (s, 1H), 7.91 – 7.77 (m, 1H), 7.65 (s, 1H), 7.62 – 7.55 (m, 1H), 7.50 – 7.37 (m, 2H), 4.66 (t,** *J* **= 5.6 Hz, 2H), 3.98 (t,** *J* **= 5.6 Hz, 2H), 3.63 – 3.58 (m, 2H), 3.57 – 3.44 (m, 8H), 1.19 (t,** *J* **= 7.2 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) \delta 160.62, 155.04, 154.19, 152.59,** 

152.45, 141.73, 131.37, 126.62, 125.20, 120.72, 112.55, 111.03, 70.65, 70.50, 70.42, 69.75, 68.87, 66.62, 47.13, 15.16. HRMS (ESI) m/z calcd for  $C_{20}H_{24}N_5O_4S^+$  [M+H]<sup>+</sup> 430.1544, found 430.1542.



*N*-(2-(4-(benzo[*d*]oxazol-2-ylthio)-1*H*-pyrazolo[3,4-*d*] pyrimidin-1-yl)ethyl)-5-((3aS,4S,6aR)-2-oxohexahydro -1Hthieno[3,4-*d*]imidazol-4-yl)pentanamide (2v): According to the general procedure affording the 2v as a white solid. 39% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.73 (s, 1H), 7.88 – 7.82 (m, 1H), 7.68 (s, 1H), 7.64 – 7.56 (m, 1H), 7.53 – 7.39 (m, 2H), 6.72 (s, 1H), 6.56 (s, 1H), 6.19 (s, 1H), 4.65 – 4.48 (m, 3ZH), 4.43 – 4.36 (m, 1H), 3.29 – 3.05 (m, 3H), 2.93 (dd, *J* = 12.8, 4.4 Hz, 1H), 2.76 (d, *J* = 12.8 Hz, 1H), 2.36 – 2.22 (m, 2H), 2.17 – 2.06 (m, 2H),

1.84 – 1.63 (m, 2H), 1.55 – 1.39 (m, 2H), 1.36 – 1.25 (m, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  173.87, 164.69, 161.26, 154.77, 154.25, 152.61, 151.94, 141.62, 131.56, 126.80, 125.34, 120.72, 112.46, 111.09, 62.29, 60.69, 55.16, 44.79, 40.39, 36.39, 35.75, 28.94, 27.88, 27.80, 25.36. HRMS (ESI) m/z calcd for C<sub>25</sub>H<sub>29</sub>N<sub>8</sub>O<sub>3</sub>S<sub>2</sub><sup>+</sup> [M+H]<sup>+</sup> 553.1799, found 553.1803.



Allyl (S)-4-(4-(benzo[*d*]oxazol-2-ylthio)-1*H*-pyrazolo [3,4*d*]pyrimidin-1-yl)-2-((*tert*-butoxycarbonyl)amino) butanoate (2w): According to the general procedure affording the 2w as a white solid. 92% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.72 (s, 1H), 7.87 – 7.80 (m, 1H), 7.62 (s, 1H), 7.61 – 7.55 (m, 1H), 7.51 – 7.39 (m, 2H), 5.91 – 5.75 (m, 1H), 5.39 – 5.18 (m, 3H), 4.59 (t, *J* = 6.8 Hz, 2H), 4.53 – 4.46 (m, 2H), 4.44 – 4.33 (m, 1H), 2.60 – 2.46 (m, COOAIIyi 1H), 2.44 – 2.28 (m, 1H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.30, 160.80, 155.31, 154.97, 154.20, 152.57, 152.01,

141.71, 131.45, 131.30, 126.64, 125.21, 120.71, 119.02, 112.57, 111.01, 80.17, 66.14, 51.36, 43.83, 31.73, 28.28. HRMS (ESI) m/z calcd for  $C_{24}H_{27}N_6O_5S_2^+$  [M+H]<sup>+</sup> 511.1758, found 511.1746.

2-((6-(4-(2-(4-(benzo[*d*]oxazol-2-ylthio)-1*H*-pyrazolo[3,4-*d*]pyrimidin-1-yl)ethyl) piperazin-1yl)-2-methylpyrimidin-4-yl)amino)-*N*-(2-chloro-6-methylphenyl) thiazole-5-carboxamide (2x):



According to the general procedure affording the **2x** as a white solid. 58% yield. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$ 11.51 (s, 1H), 9.93 (s, 1H), 8.82 (s, 1H), 8.27 (s, 1H), 8.25 (s, 1H), 7.94 (d, *J* = 7.6 Hz, 1H), 7.88 (d, *J* = 7.6 Hz,

1H), 7.64 – 7.51 (m, 2H), 7.45 (d, J = 6.8 Hz, 1H), 7.38 – 7.26 (m, 2H), 6.06 (s, 1H), 4.66 (t, J = 5.6 Hz, 2H), 3.47 – 3.40 (m, 4H), 2.90 (t, J = 5.6 Hz, 2H), 2.55 (s, 4H), 2.44 (s, 3H), 2.29 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  165.61, 163.01, 162.77, 160.50, 160.39, 157.38, 155.00, 154.69, 152.62, 152.14, 141.67, 141.30, 139.29, 133.99, 132.90, 131.98, 129.50, 128.64, 127.48, 127.34, 126.17, 125.82, 120.80, 112.09, 111.76, 83.10, 56.71, 52.37, 44.83, 43.94, 26.04, 18.78. HRMS (ESI) m/z calcd for C<sub>34</sub>H<sub>31</sub>ClN<sub>12</sub>O<sub>2</sub>S<sub>2</sub><sup>+</sup> [M+H]<sup>+</sup> 739.1896, found 739.1925.

# 2-(2-(2-(2-(4-(benzo[d]oxazol-2-ylthio)-1*H*-pyrazolo[3,4-d]pyrimidin-1-yl)ethoxy) ethoxy)ethyl)-3',6'-bis(diethylamino)spiro[isoindoline-1,9'-xanthen]-3-one (2y):



According to the general procedure affording the **2x** as a pink solid. 52% yied. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.67 (s, 1H), 7.93 – 7.86 (m, 1H), 7.85 – 7.78 (m, 1H), 7.61 (s, 1H), 7.60 – 7.54 (m, 1H), 7.49 – 7.35 (m, 4ZH), 7.09 – 7.01 (m, 1H), 6.45 – 6.34 (m, 4H), 6.24 (dd, *J* =

8.8, 2.4 Hz, 2H), 4.59 (t, J = 5.7 Hz, 2H), 3.90 (t, J = 5.6 Hz, 2H), 3.46 – 3.41 (m, 2H), 3.36 – 3.24 (m, 12H), 3.11 (t, J = 7.2 Hz, 2H), 1.15 (t, J = 7.2 Hz, 12H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.28, 160.47, 155.18, 154.16, 153.78, 153.25, 152.59, 152.39, 148.77, 141.75, 132.36, 131.33, 130.95, 128.84, 127.95, 126.57, 125.16, 123.76, 122.76, 120.67, 112.60, 111.03, 108.09, 105.56, 97.82, 70.11, 69.94, 68.85, 67.84, 64.85, 47.09, 44.36, 39.27, 12.60. HRMS (ESI) m/z calcd for C<sub>46</sub>H<sub>49</sub>N<sub>8</sub>O<sub>5</sub>S<sup>+</sup> [M+H]<sup>+</sup> 824.3541, found 825.3559.

#### 8. Arylation of cysteine-containing peptides/proteins

General procedure for arylation of cysteine-containing peptides: 1.0  $\mu$ mol peptide (1) was dissolved in 990  $\mu$ L non-degassed Tris buffer (100 mM, pH 8.0). Then, aryl thioether (2, 3.0  $\mu$ mol) in DMSO was added. The resulting solution was vortexed and shaking at room temperature for 1h. After this time, the reaction was analyzed by HPLC-MS.



According to the general procedure for peptide synthesis affording **1a** as a white solid. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  12.64 (s, 1H), 8.60 (d, *J* = 8.0 Hz, 1H), 8.33 (d, *J* = 7.2 Hz, 1H), 8.27 (d, *J* = 8.0 Hz, 1H), 8.04 – 7.98 (m, 2H), 7.33 – 7.17 (m, 5H), 4.67 – 4.55 (m, 1H), 4.49 – 4.39 (m, 1H), 4.27 – 4.12 (m, 1H), 3.82 – 3.68 (m, 1H), 3.07 (dd, *J* = 14.0, 4.0 Hz, 1H), 2.88 – 2.65 (m, 3H),

2.25 (t, J = 8.4 Hz, 1H), 1.34 (d, J = 6.8 Hz, 3H), 1.29 (d, J = 7.2 Hz, 3H). <sup>13</sup>C NMR (100 MHz,

DMSO)  $\delta$  174.15, 171.34, 169.87, 169.82, 137.73, 129.56, 128.72, 127.04, 54.74, 52.20, 48.23, 48.10, 40.47, 37.56, 17.46, 17.36. HRMS (ESI) m/z calcd for C<sub>18</sub>H<sub>27</sub>N<sub>4</sub>O<sub>5</sub>S<sup>+</sup> [M+H]<sup>+</sup> 411.1697, found 411.1679.



According to the general procedure for arylation of cysteine containing peptides affording **3aa** as a white solid. <sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  8.71 (s, 1H), 8.60 (d, J = 8.0 Hz, 1H), 8.56 (d, J = 8.4 Hz, 1H), 8.46 (s, 1H), 8.33 (d, J = 7.2 Hz, 1H), 8.01 (d, J = 4.4 Hz, 2H), 7.33 – 7.11 (m, 5H), 4.75 – 4.64 (m, 1H), 4.63 – 4.54 (m, 1H), 4.30 – 4.15 (m, 1H), 3.90 (dd, J = 13.2, 4.4 Hz, 1H), 3.81 – 3.71 (m, 1H), 3.51 (dd, J = 13.2, 9.2 Hz, 1H), 3.06 (dd, J = 14.0, 4.0 Hz, 1H), 2.75 (dd, J = 14.0, 9.6 Hz, 1H), 1.32

(d, J = 4.8 Hz, 3H), 1.30 (d, J = 5.2 Hz, 3H). <sup>13</sup>C NMR (100 MHz, DMSO)  $\delta$  174.17, 171.27, 169.99, 169.73, 158.83, 158.48, 151.94, 143.85, 137.99, 129.61, 128.63, 126.89, 117.98, 54.72, 52.29, 48.43, 48.21, 37.70, 30.81, 17.68. HRMS (ESI) m/z calcd for C<sub>23</sub>H<sub>29</sub>N<sub>8</sub>O<sub>5</sub>S<sup>+</sup> [M+H]<sup>+</sup> 529.1976, found 529.1992.



Figure S3. LC-MS TIC curve of peptide 1a. Gradient used: Method A



**Figure S4.** ESI Mass spectrum of peptide **1a**. Calculated Mass [M+H]<sup>+</sup>: 411.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 411.1.



Figure S5. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S6. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S7.** ESI Mass spectrum of modified product **3aa**. Calculated Mass [M+H]<sup>+</sup>: 529.2; [M+2H]<sup>2+</sup>: 265.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 529.1; [M+2H]<sup>2+</sup>: 265.1.



**Figure S8.** ESI Mass spectrum of dimerized byproduct by peptide **1a**. Calculated Mass [M+H]<sup>+</sup>: 819.3; [M+2H]<sup>2+</sup>: 410.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 819.3; [M+2H]<sup>2+</sup>: 410.2.



Figure S9. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S10. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S11.** ESI Mass spectrum of modified product **3ab**. Calculated Mass [M+H]<sup>+</sup>: 563.2; [M+2H]<sup>2+</sup>: 282.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 563.1; [M+2H]<sup>2+</sup>: 282.2.



Figure S12. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S13. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S14.** ESI Mass spectrum of modified product **3ac**. Calculated Mass [M+H]<sup>+</sup>: 544.2; [M+2H]<sup>2+</sup>: 272.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 544.1; [M+2H]<sup>2+</sup>: 272.6.



Figure S15. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S16. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S17.** ESI Mass spectrum of modified product **3ad**. Calculated Mass [M+H]<sup>+</sup>: 489.2; [M+2H]<sup>2+</sup>: 245.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 489.1; [M+2H]<sup>2+</sup>: 245.2.



Figure S18. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S19. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S20.** ESI Mass spectrum of modified product **3ae**. Calculated Mass [M+H]<sup>+</sup>: 539.2; [M+2H]<sup>2+</sup>: 270.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 539.1; [M+2H]<sup>2+</sup>: 270.1.



Figure S21. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S22. LC-MS TIC curve of reaction mixture. Gradient used: Method A



Figure S23. ESI Mass spectrum of modified product **3af**. Calculated Mass [M+H]<sup>+</sup>: 573.2; [M+H]<sup>+</sup>: 573.1.



Figure S24. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S25. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S26.** ESI Mass spectrum of modified product **3ag**. Calculated Mass [M+H]<sup>+</sup>: 620.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 620.2.



Figure S27. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S28. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S29.** ESI Mass spectrum of modified product **3ah**. Calculated Mass [M+H]<sup>+</sup>: 577.2; [M+2H]<sup>2+</sup>: 289.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 577.1; [M+2H]<sup>2+</sup>: 289.1.



Figure S30. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S31. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S32.** ESI Mass spectrum of modified product **3ai**. Calculated Mass [M+H]<sup>+</sup>: 529.2; [M+2H]<sup>2+</sup>: 265.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 529.1; [M+2H]<sup>2+</sup>: 265.1.



Figure S33. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S34. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S35.** ESI Mass spectrum of modified product **3aj**. Calculated Mass [M+H]<sup>+</sup>: 563.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 563.1.



Figure S36. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S37. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S38.** ESI Mass spectrum of modified product **3ak**. Calculated Mass [M+H]<sup>+</sup>: 544.2; [M+2H]<sup>2+</sup>: 272.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 544.1; [M+2H]<sup>2+</sup>: 272.6.



Figure S39. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S40. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S41.** ESI Mass spectrum of modified product **3al**. Calculated Mass [M+H]<sup>+</sup>: 612.2; [M+2H]<sup>2+</sup>: 306.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 612.2; [M+2H]<sup>2+</sup>: 306.7.



Figure S42. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S43. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S44.** ESI Mass spectrum of modified product **3am**. Calculated Mass [M+H]<sup>+</sup>: 631.3; [M+2H]<sup>2+</sup>: 316.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 631.2; [M+2H]<sup>2+</sup>: 316.2.



Figure S45. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S46. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S47.** ESI Mass spectrum of modified product **3an**. Calculated Mass [M+H]<sup>+</sup>: 812.3; [M+2H]<sup>2+</sup>: 406.7; Mass Found (ESI+); [M+H]<sup>+</sup>: 812.3; [M+2H]<sup>2+</sup>: 406.7.



Figure S48. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S49. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S50.** ESI Mass spectrum of modified product **3ao**. Calculated Mass [M+H]<sup>+</sup>: 569.2; [M+2H]<sup>2+</sup>: 285.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 569.2; [M+2H]<sup>2+</sup>: 285.2.



Figure S51. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S52. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S53.** ESI Mass spectrum of modified product **3ap**. Calculated Mass [M+H]<sup>+</sup>: 567.2; [M+2H]<sup>2+</sup>: 284.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 567.1; [M+2H]<sup>2+</sup>: 284.1.



Figure S54. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S55. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S56.** ESI Mass spectrum of modified product **3aq**. Calculated Mass [M+H]<sup>+</sup>: 615.2; [M+2H]<sup>2+</sup>: 308.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 615.2; [M+2H]<sup>2+</sup>: 308.2.



Figure S57. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S58. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S59.** ESI Mass spectrum of modified product **3ar**. Calculated Mass [M+H]<sup>+</sup>: 649.2; [M+2H]<sup>2+</sup>: 325.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 649.1; [M+2H]<sup>2+</sup>: 325.1.



Figure S60. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S61. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S62.** ESI Mass spectrum of modified product **3as**. Calculated Mass [M+H]<sup>+</sup>: 612.2; [M+2H]<sup>2+</sup>: 306.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 612.2; [M+2H]<sup>2+</sup>: 306.7.


Figure S63. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S64. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S65.** ESI Mass spectrum of modified product **3at**. Calculated Mass [M+H]<sup>+</sup>: 672.3; Mass Found (ESI+); [M+H]<sup>+</sup>: 672.2.



Figure S66. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S67. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S68.** ESI Mass spectrum of modified product **3au**. Calculated Mass [M+H]<sup>+</sup>: 689.3; [M+2H]<sup>2+</sup>: 345.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 688.3; [M+2H]<sup>2+</sup>: 345.2.



Figure S69. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S70. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S71.** ESI Mass spectrum of modified product **3av**. Calculated Mass [M+H]<sup>+</sup>: 812.3; [M+2H]<sup>2+</sup>: 406.7; Mass Found (ESI+); [M+H]<sup>+</sup>: 812.2; [M+2H]<sup>2+</sup>: 406.7.



Figure S72. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method A



Figure S73. LC-MS TIC curve of reaction mixture. Gradient used: Method A



**Figure S74.** ESI Mass spectrum of modified product **3aw**. Calculated Mass [M+H]<sup>+</sup>: 770.3; Mass Found (ESI+); [M+H]<sup>+</sup>: 770.2.



Figure S75. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S76. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S77.** ESI Mass spectrum of modified product **3ax**. Calculated Mass [M+H]<sup>+</sup>: 998.3; [M+2H]<sup>2+</sup>: 499.7; [M+3H]<sup>3+</sup>: 333.5; Mass Found (ESI+); [M+H]<sup>+</sup>: 998.1; [M+2H]<sup>2+</sup>: 499.6; [M+3H]<sup>3+</sup>: 333.5.



Figure S78. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S79. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S80.** ESI Mass spectrum of modified product **3ay**. Calculated Mass [M+2H]<sup>2+</sup>: 542.8; [M+3H]<sup>3+</sup>: 362.2; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 542.7; [M+3H]<sup>3+</sup>: 362.3.



Figure S81. LC-MS TIC curve of peptide 1b. Gradient used: Method B



**Figure S82.** ESI Mass spectrum of peptide **1b**. Calculated Mass [M+H]<sup>+</sup>: 434.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 434.2.







**Figure S84.** ESI Mass spectrum of modified product **3bi**. Calculated Mass [M+H]<sup>+</sup>: 552.3; [M+2H]<sup>2+</sup>: 276.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 552.2; [M+2H]<sup>2+</sup>: 276.7.



Figure S85. LC-MS TIC curve of peptide 1c. Gradient used: Method B



**Figure S86.** ESI Mass spectrum of peptide **1c**. Calculated Mass [M+H]<sup>+</sup>: 462.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 462.2.







**Figure S88.** ESI Mass spectrum of modified product **3ci**. Calculated Mass [M+H]<sup>+</sup>: 580.3; [M+2H]<sup>2+</sup>: 290.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 580.2; [M+2H]<sup>2+</sup>: 290.7.



Figure S89. LC-MS TIC curve of peptide 1d. Gradient used: Method B



**Figure S90.** ESI Mass spectrum of peptide **1d**. Calculated Mass [M+H]<sup>+</sup>: 497.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 497.1.





**Figure S92.** ESI Mass spectrum of modified product **3di**. Calculated Mass [M+H]<sup>+</sup>: 615.2; [M+2H]<sup>2+</sup>: 308.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 615.1; [M+2H]<sup>2+</sup>: 308.2.





**Figure S94.** ESI Mass spectrum of peptide **1e**. Calculated Mass [M+H]<sup>+</sup>: 443.2; [M+2H]<sup>2+</sup>: 222.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 443.1; [M+2H]<sup>2+</sup>: 222.2.





**Figure S96.** ESI Mass spectrum of modified product **3ei**. Calculated Mass [M+H]<sup>+</sup>: 561.2; [M+2H]<sup>2+</sup>: 281.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 561.1; [M+2H]<sup>2+</sup>: 281.2.



**Figure S98.** ESI Mass spectrum of peptide **1f**. Calculated Mass [M+H]<sup>+</sup>: 393.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 393.1.

m/z



**Figure S100.** ESI Mass spectrum of modified product **3fi**. Calculated Mass [M+H]<sup>+</sup>: 511.2; [M+2H]<sup>2+</sup>: 256.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 511.2; [M+2H]<sup>2+</sup>: 256.2.



**Figure S102.** ESI Mass spectrum of peptide **1g**. Calculated Mass [M+H]<sup>+</sup>: 427.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 427.1.



**Figure S104.** ESI Mass spectrum of modified product **3gi**. Calculated Mass [M+H]<sup>+</sup>: 545.2; [M+2H]<sup>2+</sup>: 273.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 545.1; [M+2H]<sup>2+</sup>: 273.2.



Figure S105. LC-MS TIC curve of peptide 1h. Gradient used: Method B



**Figure S106.** ESI Mass spectrum of peptide **1h**. Calculated Mass [M+H]<sup>+</sup>: 464.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 464.1.





**Figure S108.** ESI Mass spectrum of modified product **3hi**. Calculated Mass [M+H]<sup>+</sup>: 582.2; [M+2H]<sup>2+</sup>: 291.6; Mass Found (ESI+); [M+H]<sup>+</sup>: 582.2; [M+2H]<sup>2+</sup>: 291.7.



Figure S109. LC-MS TIC curve of peptide 1i. Gradient used: Method B



**Figure S110.** ESI Mass spectrum of peptide **1i**. Calculated Mass [M+H]<sup>+</sup>: 649.3; [M+2H]<sup>2+</sup>: 325.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 649.3; [M+2H]<sup>2+</sup>: 325.2.





**Figure S112.** ESI Mass spectrum of modified product **3ii**. Calculated Mass [M+H]<sup>+</sup>: 767.4; [M+2H]<sup>2+</sup>: 384.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 767.3; [M+2H]<sup>2+</sup>: 384.2.



Figure S113. LC-MS TIC curve of peptide 1j. Gradient used: Method B



**Figure S114.** ESI Mass spectrum of peptide **1***j*. Calculated Mass [M+H]<sup>+</sup>: 728.4; [M+2H]<sup>2+</sup>: 364.7; Mass Found (ESI+); [M+H]<sup>+</sup>: 728.3; [M+2H]<sup>2+</sup>: 364.8.



Figure S115. LC-MS TIC curve of reaction mixture. Gradient used: Method B



**Figure S116.** ESI Mass spectrum of modified product **3ji**. Calculated Mass [M+H]<sup>+</sup>: 846.4; [M+2H]<sup>2+</sup>: 423.7; [M+3H]<sup>3+</sup>: 282.8; Mass Found (ESI+); [M+H]<sup>+</sup>: 846.3; [M+2H]<sup>2+</sup>: 423.7; [M+3H]<sup>3+</sup>: 282.9.





**Figure S118.** ESI Mass spectrum of peptide **1k**. Calculated Mass [M+H]<sup>+</sup>: 979.4; [M+2H]<sup>2+</sup>: 490.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 979.3; [M+2H]<sup>2+</sup>: 490.2.



**Figure S120.** ESI Mass spectrum of modified product **3ki**. Calculated Mass [M+H]<sup>+</sup>: 1097.4; [M+2H]<sup>2+</sup>: 549.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 1097.3; [M+2H]<sup>2+</sup>: 549.3.



Figure S121. LC-MS TIC curve of peptide 1I. Gradient used: Method C



**Figure S122.** ESI Mass spectrum of peptide **1**I. Calculated Mass [M+H]<sup>+</sup>: 1011.5; [M+2H]<sup>2+</sup>: 506.2; [M+3H]<sup>3+</sup>: 337.8; Mass Found (ESI+); [M+H]<sup>+</sup>: 1011.3; [M+2H]<sup>2+</sup>: 506.2; [M+3H]<sup>3+</sup>: 338.0.



Figure S123. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S124. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S125.** ESI Mass spectrum of modified product **3li**. Calculated Mass [M+H]<sup>+</sup>: 1129.5; [M+2H]<sup>2+</sup>: 565.3; [M+3H]<sup>3+</sup>: 377.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 1130.1; [M+2H]<sup>2+</sup>: 565.3; [M+3H]<sup>3+</sup>: 377.3.



Figure S126. LC-MS TIC curve of peptide 11'. Gradient used: Method D



**Figure S127.** ESI Mass spectrum of peptide **1***I*'. Calculated Mass [M+H]<sup>+</sup>: 979.5; [M+2H]<sup>2+</sup>: 490.3; [M+3H]<sup>3+</sup>: 327.3; Mass Found (ESI+); [M+H]<sup>+</sup>: 980.3; [M+2H]<sup>2+</sup>: 490.3; [M+3H]<sup>3+</sup>: 327.2.



Figure S128. LC-MS TIC curve of reaction mixture. Gradient used: Method D



**Figure S129.** LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S130. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S131.** ESI Mass spectrum of modified product **3ls**. Calculated Mass [M+H]<sup>+</sup>: 1212.5; [M+2H]<sup>2+</sup>: 606.8; [M+3H]<sup>3+</sup>: 404.9; Mass Found (ESI+); [M+H]<sup>+</sup>: 1212.2; [M+2H]<sup>2+</sup>: 606.8; [M+3H]<sup>3+</sup>: 404.9.



**Figure S132.** LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S133. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S134.** ESI Mass spectrum of modified product **3lv**. Calculated Mass [M+2H]<sup>2+</sup>: 706.8; [M+3H]<sup>3+</sup>: 471.6; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 706.8; [M+3H]<sup>3+</sup>: 471.6.



Figure S135. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S136. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S137.** ESI Mass spectrum of modified product **3la**. Calculated Mass [M+H]<sup>+</sup>: 1129.5; [M+2H]<sup>2+</sup>: 565.3; [M+3H]<sup>3+</sup>: 377.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 1130.2; [M+2H]<sup>2+</sup>: 565.3; [M+3H]<sup>3+</sup>: 377.3.



Figure S138. LC-MS TIC curve of control reaction (without peptide). Gradient used: Method C



Figure S139. LC-MS TIC curve of reaction mixture. Gradient used: Method C



**Figure S140.** ESI Mass spectrum of modified product **3In**. Calculated Mass [M+2H]<sup>2+</sup>: 706.8; [M+3H]<sup>3+</sup>: 471.6; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 706.8; [M+3H]<sup>3+</sup>: 471.6.



**Figure S141.** LC-MS TIC curve of control reaction (without peptide). Gradient used: Method D



Figure S142. LC-MS TIC curve of peptide 1m. Gradient used: Method D



**Figure S143.** ESI Mass spectrum of peptide **1m**. Calculated Mass [M+2H]<sup>2+</sup>: 752.9; [M+3H]<sup>3+</sup>: 502.2; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 753.1; [M+3H]<sup>3+</sup>: 502.5.



Figure S144. LC-MS TIC curve of reaction mixture. Gradient used: Method D



**Figure S145.** ESI Mass spectrum of modified product **3mi**. Calculated Mass [M+2H]<sup>2+</sup>: 811.9; [M+3H]<sup>3+</sup>: 541.6; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 812.2; [M+3H]<sup>3+</sup>: 541.8.



Figure S146. LC-MS TIC curve of peptide 1n. Gradient used: Method D



**Figure S147.** ESI Mass spectrum of peptide **1n**. Calculated Mass [M+2H]<sup>2+</sup>: 787.4; [M+3H]<sup>3+</sup>: 525.3; [M+4H]<sup>4+</sup>: 394.2; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 787.6; [M+3H]<sup>3+</sup>: 525.4; [M+4H]<sup>4+</sup>: 394.4.



Figure S148. LC-MS TIC curve of reaction mixture. Gradient used: Method D



**Figure S149.** ESI Mass spectrum of modified peptide **1ni**. Calculated Mass [M+2H]<sup>2+</sup>: 846.4; [M+3H]<sup>3+</sup>: 564.6; [M+4H]<sup>4+</sup>: 423.7; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 846.7; [M+3H]<sup>3+</sup>: 546.9; [M+4H]<sup>4+</sup>: 423.9.

# 9. Stability evaluation of modified peptide

Peptide conjugates were pre-dissolved in water to afford the 1.11 mM stock solution.  $K_2CO_3$  and  $H_5IO_6$  were dissolved in water to afford 50 mM  $K_2CO_3$  and 5 mM  $H_5IO_6$  solutions. Glutathione (GSH) and 2-mercaptoethanol ( $\beta$ ME) were dissolved in Tris buffer (100 mM, pH 8.0) to afford external thiol test solutions.

## Acidic conditions (pH 4.0 AcOH)

Peptide conjugates were dissolved in pH 4.0 AcOH buffeer to afford the 1 mM peptide solutions. Then these samples left at room temperature and analyzed by LC-MS at corresponding time.

## Basic conditions (5 mM K<sub>2</sub>CO<sub>3</sub>)

Corresponding cysteine conjugates (1.11 mM; 450  $\mu$ L) and K<sub>2</sub>CO<sub>3</sub> solution (50  $\mu$ L, 50 mM in H<sub>2</sub>O) were combined in a plastic Eppendorf and left at room temperature and analyzed by LC-MS at corresponding time.

#### Oxidation conditions (5 mM H<sub>5</sub>IO<sub>6</sub>)

Corresponding cysteine conjugates (1.11 mM; 450  $\mu$ L) and H<sub>5</sub>IO<sub>6</sub> solution (50  $\mu$ L, 50 mM in H<sub>2</sub>O) were combined in a plastic Eppendorf and left at room temperature and analyzed by LC-MS at corresponding time.

#### External thiol nucleophiles (GSH and βME)

Peptide conjugates were dissolved in GSH (100 mM in Tris) orβME (100 mM in Tris) to afford the 1 mM peptide solutions. Then these samples left at room temperature and analyzed by LC-MS at corresponding time.



Figure S150. Stability of modified peptides 3li, 3ls, 3lv, 3la and 3ln.

# 10. Regeneration activity evaluation of modified peptides

General procedure: Firstly, according the general procedure for arylation of cysteinecontaining peptides to afford the modified peptide **3**. After reaction finished, 1 mL 100 mM mercaptoethanol ( $\beta$ ME) or glutathione (GSH) Tris (100 mM, pH 8.0) solution was added to reaction mixture respectively, then monitored by LC-MS to determine the ratio of **3** and **1a** in peptide species. %ratio = S<sub>desired</sub>/S<sub>total</sub>, where S<sub>desired</sub> is the peak area of **3** or **1a** and S<sub>total</sub> is the peak area of combined peptide-containing species (**3**, **1a** and byproduct). The blue curve represent the ratio of **3** and the orange represent **1a**.

























Figure 151. Regeneration activity evaluation of modified peptides.



Figure S152. LC-MS TIC curve of control reaction for the whole process with 2ad and  $\beta$ ME (without peptide). Gradient used: Method D



S63



#### ESI Mass spectrum of peak B





ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S153.** LC-MS TIC curve of reaction mixture of **3aa** and corresponding ESI mass spectrum after added βME for 8h. Gradient used: Method D



**Figure S154.** LC-MS TIC curve of control reaction for the whole process with **2ad** and GSH (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C



ESI Mass spectrum of peak E

Figure S155. LC-MS TIC curve of reaction mixture of 3aa and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D



Figure S156. LC-MS TIC curve of control reaction for the whole process with 2b and  $\beta$ ME (without peptide). Gradient used: Method D





ESI Mass spectrum of peak C ESI Mass spectrum of peak D **Figure S157.** LC-MS TIC curve of reaction mixture of **3ab** and corresponding ESI mass spectrum after added βME for 8h. Gradient used: Method D



**Figure S158.** LC-MS TIC curve of control reaction for the whole process with **2b** and GSH (without peptide). Gradient used: Method D





ESI Mass spectrum of peak C ESI Mass spectrum of peak D Figure S159. LC-MS TIC curve of reaction mixture of **3ab** and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D



**Figure S160.** LC-MS TIC curve of control reaction for the whole process with **2e** and  $\beta$ ME (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C



ESI Mass spectrum of peak D ESI Mass spectrum of peak E Figure S161. LC-MS TIC curve of reaction mixture of 3ae and corresponding ESI mass spectrum after added βME for 2h. Gradient used: Method D



Figure S162. LC-MS TIC curve of control reaction for the whole process with 2e and GSH (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A

ESI Mass spectrum of peak B ESI Mass spectrum of peak C



ESI Mass spectrum of peak D ESI Mass spectrum of peak E Figure S163. LC-MS TIC curve of reaction mixture of **3ae** and corresponding ESI mass spectrum after added GSH for 3h. Gradient used: Method D



**Figure S164.** LC-MS TIC curve of control reaction for the whole process with **2g** and  $\beta$ ME (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A

ESI Mass spectrum of peak B

ESI Mass spectrum of peak C



ESI Mass spectrum of peak D ESI Mass spectrum of peak E **Figure S165.** LC-MS TIC curve of reaction mixture of **3ag** and corresponding ESI mass spectrum after added βME for 1h. Gradient used: Method D



**Figure S166.** LC-MS TIC curve of control reaction for the whole process with **2g** and GSH (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A

ESI Mass spectrum of peak B



ESI Mass spectrum of peak D ESI Mass spectrum of peak E

**Figure S167.** LC-MS TIC curve of reaction mixture of **3ag** and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D



**Figure S168.** LC-MS TIC curve of control reaction for the whole process with **2h** and  $\beta$ ME (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C


ESI Mass spectrum of peak D ESI Mass spectrum of peak E **Figure S169.** LC-MS TIC curve of reaction mixture of **3ah** and corresponding ESI mass spectrum after added βME for 1h. Gradient used: Method D



**Figure S170.** LC-MS TIC curve of control reaction for the whole process with **2h** and GSH (without peptide). Gradient used: Method D



ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C Figure S171. LC-MS TIC curve of reaction mixture of **3ah** and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D



**Figure S172.** LC-MS TIC curve of control reaction for the whole process with **2i** and  $\beta$ ME (without peptide). Gradient used: Method D



ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S173.** LC-MS TIC curve of reaction mixture of **3ai** and corresponding ESI mass spectrum after added βME for 8h. Gradient used: Method D



**Figure S174.** LC-MS TIC curve of control reaction for the whole process with **2i** and GSH (without peptide). Gradient used: Method D



**Figure S175.** LC-MS TIC curve of reaction mixture of **3ai** and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D

![](_page_75_Figure_0.jpeg)

**Figure S176.** LC-MS TIC curve of control reaction for the whole process with **2j** and  $\beta$ ME (without peptide). Gradient used: Method D

![](_page_75_Figure_2.jpeg)

ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C **Figure S177.** LC-MS TIC curve of reaction mixture of **3aj** and corresponding ESI mass spectrum after added βME for 1h. Gradient used: Method D

![](_page_75_Figure_4.jpeg)

**Figure S178.** LC-MS TIC curve of control reaction for the whole process with **2j** and GSH (without peptide). Gradient used: Method D

![](_page_76_Figure_0.jpeg)

ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S179.** LC-MS TIC curve of reaction mixture of **3aj** and corresponding ESI mass spectrum after added GSH for 8h. Gradient used: Method D

![](_page_76_Figure_4.jpeg)

**Figure S180.** LC-MS TIC curve of control reaction (without peptide) for the first step with **2j**. Gradient used: Method D

![](_page_77_Figure_0.jpeg)

**Figure S181.** LC-MS TIC curve of control reaction (without peptide) for the first step with **2ad**. Gradient used: Method D

![](_page_77_Figure_2.jpeg)

**Figure S182.** LC-MS TIC curve of control reaction for the whole process with **2j** and  $\beta$ ME (without peptide). Gradient used: Method D

![](_page_77_Figure_4.jpeg)

**Figure S183.** LC-MS TIC curve of control reaction for the whole process with **2ad** and  $\beta$ ME (without peptide). Gradient used: Method D

![](_page_77_Figure_6.jpeg)

Figure S184. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_78_Figure_0.jpeg)

**Figure S185.** ESI Mass spectrum of modified product **3di**. Calculated Mass [M+H]<sup>+</sup>: 649.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 649.2.

![](_page_78_Figure_2.jpeg)

ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S186.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_79_Figure_0.jpeg)

Figure S187. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_79_Figure_2.jpeg)

**Figure S188.** ESI Mass spectrum of modified product **3da**. Calculated Mass [M+H]<sup>+</sup>: 615.2; [M+2H]<sup>2+</sup>: 308.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 615.3; [M+2H]<sup>2+</sup>: 308.3.

![](_page_79_Figure_4.jpeg)

![](_page_80_Figure_0.jpeg)

ESI Mass spectrum of peak C ESI Mass spectrum of peak D **Figure S189.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_80_Figure_2.jpeg)

Figure S190. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_80_Figure_4.jpeg)

**Figure S191.** ESI Mass spectrum of modified product **3fj**. Calculated Mass [M+H]<sup>+</sup>: 545.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 545.2.

![](_page_80_Figure_6.jpeg)

![](_page_81_Figure_0.jpeg)

![](_page_81_Figure_1.jpeg)

![](_page_81_Figure_2.jpeg)

m/z

ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

Figure S192. LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_81_Figure_6.jpeg)

![](_page_81_Figure_7.jpeg)

![](_page_81_Figure_8.jpeg)

Figure S194. ESI Mass spectrum of modified product 3fa. Calculated Mass [M+H]<sup>+</sup>: 511.2;

![](_page_82_Figure_0.jpeg)

![](_page_82_Figure_1.jpeg)

ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S195.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_82_Figure_5.jpeg)

Figure S196. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_83_Figure_0.jpeg)

**Figure S197.** ESI Mass spectrum of modified product **3jj**. Calculated Mass [M+H]<sup>+</sup>: 880.3; [M+2H]<sup>2+</sup>: 440.7; [M+3H]<sup>3+</sup>: 294.1; Mass Found (ESI+); [M+H]<sup>+</sup>: 880.4; [M+2H]<sup>2+</sup>: 440.8; [M+3H]<sup>3+</sup>: 294.3.

![](_page_83_Figure_2.jpeg)

ESI Mass spectrum of peak C

ESI Mass spectrum of peak D

**Figure S198.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_84_Figure_0.jpeg)

Figure S199. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_84_Figure_2.jpeg)

**Figure S200.** ESI Mass spectrum of modified product **3ja**. Calculated Mass [M+H]<sup>+</sup>: 846.4; [M+2H]<sup>2+</sup>: 423.7; [M+3H]<sup>3+</sup>: 282.8; Mass Found (ESI+); [M+H]<sup>+</sup>: 846.4; [M+2H]<sup>2+</sup>: 423.8; [M+3H]<sup>3+</sup>: 283.0.

![](_page_84_Figure_4.jpeg)

ESI Mass spectrum of peak A

ESI Mass spectrum of peak B

![](_page_85_Figure_0.jpeg)

ESI Mass spectrum of peak C ESI Mass spectrum of peak D **Figure S201.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_85_Figure_2.jpeg)

Figure S202. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_85_Figure_4.jpeg)

**Figure S203.** ESI Mass spectrum of modified product **3kj**. Calculated Mass [M+H]<sup>+</sup>: 1131.4; [M+2H]<sup>2+</sup>: 566.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 1131.4; [M+2H]<sup>2+</sup>: 566.3.

![](_page_85_Figure_6.jpeg)

![](_page_86_Figure_0.jpeg)

ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C **Figure S204.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_86_Figure_2.jpeg)

Figure S205. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_86_Figure_4.jpeg)

**Figure S206.** ESI Mass spectrum of modified product **3ka**. Calculated Mass [M+H]<sup>+</sup>: 1097.4; [M+2H]<sup>2+</sup>: 549.2; Mass Found (ESI+); [M+H]<sup>+</sup>: 1097.4; [M+2H]<sup>2+</sup>: 549.4.

![](_page_86_Figure_6.jpeg)

![](_page_87_Figure_0.jpeg)

ESI Mass spectrum of peak D

Figure S207. LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_87_Figure_3.jpeg)

Figure S208. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

![](_page_88_Figure_0.jpeg)

**Figure S209.** ESI Mass spectrum of modified product **3mj**. Calculated Mass [M+2H]<sup>2+</sup>: 829.3; [M+3H]<sup>3+</sup>: 553.2; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 829.2; [M+3H]<sup>3+</sup>: 553.4.

![](_page_88_Figure_2.jpeg)

ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C **Figure S210.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_88_Figure_4.jpeg)

![](_page_89_Figure_0.jpeg)

Figure S212. LC-MS TIC curve of reaction mixture for the first step. Gradient used: Method D

**Figure S213.** ESI Mass spectrum of modified product **3ma**. Calculated Mass [M+2H]<sup>2+</sup>: 811.9; [M+3H]<sup>3+</sup>: 541.6; [M+4H]<sup>4+</sup>: 406.7; Mass Found (ESI+); [M+2H]<sup>2+</sup>: 812.0; [M+3H]<sup>3+</sup>: 541.8; [M+4H]<sup>4+</sup>: 406.7.

![](_page_89_Figure_3.jpeg)

ESI Mass spectrum of peak A ESI Mass spectrum of peak B ESI Mass spectrum of peak C **Figure S214.** LC-MS TIC curve of reaction mixture for the whole process and corresponding ESI mass spectrum. Gradient used: Method D

### **11.Evaluation of Reaction Kinetics**

![](_page_89_Figure_6.jpeg)

![](_page_90_Figure_0.jpeg)

Figure S215. Time-course analysis of Cys arylation between 1a and 2.

Table S2. Competiti	ion experime	ents			
H₂N- <b>А́Р́СА</b> -СООН SH 1a		I → NH <sub>2</sub> S-6 F → F F → F F → F S-8	100 mM Tris, pH 8.0 → 5% v/v DMSO r.t. 5min	$H_{2}N - A F C A - COOH H$ $S$ $3aj Cl N H$ $H_{2}N - A F C A - COOH H$ $S$ $S-10 0 N$	$H_2N$ $A$ $F$ $C$ $A$ $-COOH$ S-9 $H_2$ $H_2N$ $A$ $F$ $C$ $A$ $-COOH$ S $F$ $F$ $FS-11 F F F$
Entry		Condition		Results	
1		<b>2j</b> (3eq.), <b>S-6</b> (3eq.)		<b>3aj : S-9</b> = 99:1	
2		<b>2j</b> (3eq.), <b>S-7</b> (3eq.)		<b>3aj : S-10</b> = 5:95	
3		<b>2j</b> (3eq.), <b>S-8</b> (3eq.)		<b>3aj : S-11</b> > 99:1	

General procedure for competition experiments: 1.0  $\mu$ mol peptide (1a) was dissolved in 950  $\mu$ L non-degassed Tris buffer (100 mM, pH 8.0). Then, 3.0  $\mu$ mol aryl thioether 2j (reach reaction completion less than 30s) and 3.0  $\mu$ mol S-6 (or S-7, or S-8) in 50  $\mu$ L DMSO was added. The

resulting solution was vortexed and shaking at room temperature for 5 min. After this time, the reaction was analyzed by HPLC-MS.

![](_page_91_Figure_1.jpeg)

ESI Mass spectrum of S-9

ESI Mass spectrum of 3aj

**Figure S216.** LC-MS TIC curve of reaction mixture modified by **2j** and **S-6** and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_91_Figure_5.jpeg)

ESI Mass spectrum of S-10 ESI Mass spectrum of 3aj

**Figure S217.** LC-MS TIC curve of reaction mixture modified by **2j** and **S-7** and corresponding ESI mass spectrum. Gradient used: Method D

![](_page_92_Figure_0.jpeg)

#### ESI Mass spectrum of 3aj

**Figure S217.** LC-MS TIC curve of reaction mixture modified by **2j** and **S-8** and corresponding ESI mass spectrum. Gradient used: Method D

### 12.BSA modification by 2v

Bovine serum albumin (BSA) was purchased from Sigma and used without further purification. BSA is composed of 583 amino acids and contains one free cysteine Cys34 (35 Cys residues in total, and 34 cysteine residues are in the form of disulfide bonds) and 59 lysines. Calculated M.W. (average): 66429.98 Da

Sequence:

DTHKSEIAHRFKDLGEEHFKGLVLIAFSQYLQQC<sup>34</sup>PFDEHVKLVNELTEFAKTCVADESHAGCEKSLHTLF GDELCKVASLRETYGDMADCCEKQEPERNECFLSHKDDSPDLPKLKPDPNTLCDEFKADEKKFWGKYLYEI ARRHPYFYAPELLYYANKYNGVFQECCQAEDKGACLLPKIETMREKVLTSSARQRLRCASIQKFGERALKA WSVARLSQKFPKAEFVEVTKLVTDLTKVHKECCHGDLLECADDRADLAKYICDNQDTISSKLKECCDKPLLE KSHCIAEVEKDAIPENLPPLTADFAEDKDVCKNYQEAKDAFLGSFLYEYSRRHPEYAVSVLLRLAKEYEATLEE CCAKDDPHACYSTVFDKLKHLVDEPQNLIKQNCDQFEKLGEYGFQNALIVRYTRKVPQVSTPTLVEVSRSL GKVGTRCCTKPESERMPCTEDYLSLILNRLCVLHEKTPVSEKVTKCCTESLVNRRPCFSALTPDETYVPKAFD EKLFTFHADICTLPDTEKQIKKQTALVELLKHKPKATEEQLKTVMENFVAFVDKCCAADDKEACFAVEGPKL VV STQTALA

![](_page_92_Figure_7.jpeg)

In a 1.5 mL eppendorf tube, bovine serum albumin (BSA) solution in 100 mM PBS buffer (pH = 7.2) (10 nmmol, 0.9 mL), 2v (100 equiv in DMSO) was added to BSA solution and shaking at room temperature for 5 h. The sample was lyophilized and analyzed by electrospray ionization mass spectrometry (ESI-MS). The lyophilized protein samples were first dissolved in Milli-Q water and buffer-exchanged with 10 mM ammonium acetate (AA) at a concentration of 50  $\mu$ M using Amicon centrifugal filters (Merck Millipore, Darmstadt, Germany) with a molecular weight cutoff of 30 k. For denatured nanoESI-MS analysis, the desalted protein samples were further diluted with 49.5/49.5/1 (v/v/v) methanol/water/formic acid mixture solution to a concentration of 3  $\mu$ M. BSA protein aliquots (3  $\mu$ L) were analyzed using a quadrupole ion mobility time-of-flight mass spectrometer (Synapt G2-Si HDMS, Waters, UK) in the positive ion mode according to the previously described method.<sup>[4]</sup> Briefly, BSA protein ions were generated by nano-ESI from a homemade borosilicate capillary emitter (1.0 mm o.d./0.58 mm i.d., Sutter Instruments, Novato, CA, USA) with a tip i.d. of  $\sim$ 1  $\mu$ m pulled using a P-97 puller (Sutter Instruments, Novato, CA, USA). The detailed instrumental parameters were as follows: capillary voltage 0.7 kV, sampling cone 100 V, extraction cone 80 V, and source temperature 30 °C.

![](_page_93_Figure_1.jpeg)

Figure S219. Ion series of BSA modification by 2v

![](_page_93_Figure_3.jpeg)

Figure S220. ESI-MS spectra of native BSA and modified BSA-2v protein.

### 13.Secondary labeling of modified peptide

![](_page_94_Figure_1.jpeg)

Figure S221. Secondary labeling of modified peptide 3ss via click reaction

**3ss** (1.31 mg, 1 µmol, 1 equiv.) in water (480 µL) was added to the solution of **4** (20 equiv.) in *t*BuOH (500 µL). CuSO4·5H<sub>2</sub>O (5 mol%) and sodium ascorbate (10 mol%) in water (20 µL) were added under vigorously stirring. The mixture was stirred at room temperature for 12 hours under Ar atmosphere. Then the crude mixture was filtered, and analyzed by LC-MS.

![](_page_94_Figure_4.jpeg)

![](_page_94_Figure_5.jpeg)

**Figure S224**. ESI Mass spectrum of product **5**. Calculated Mass [M+2H]<sup>+</sup>: 813.3; [M+3H]<sup>3+</sup>: 542.6; [M+4H]<sup>4+</sup>: 407.2; Mass Found (ESI+); [M+2H]<sup>+</sup>: 813.5; [M+3H]<sup>3+</sup>: 542.7; [M+4H]<sup>4+</sup>: 407.3.

# 14. Spectra of new compounds

![](_page_95_Figure_1.jpeg)

![](_page_96_Figure_0.jpeg)

![](_page_97_Figure_0.jpeg)

![](_page_97_Figure_1.jpeg)

![](_page_97_Figure_2.jpeg)

![](_page_97_Figure_3.jpeg)

![](_page_98_Figure_0.jpeg)

![](_page_99_Figure_0.jpeg)

S99

![](_page_100_Figure_0.jpeg)

![](_page_100_Figure_1.jpeg)

![](_page_100_Figure_2.jpeg)

![](_page_101_Figure_0.jpeg)

![](_page_101_Figure_1.jpeg)

# 

![](_page_102_Figure_1.jpeg)

![](_page_102_Figure_2.jpeg)

![](_page_102_Figure_3.jpeg)

![](_page_102_Figure_4.jpeg)

![](_page_103_Figure_0.jpeg)

![](_page_104_Figure_0.jpeg)

![](_page_105_Figure_0.jpeg)

S105

![](_page_106_Figure_0.jpeg)

![](_page_107_Figure_0.jpeg)

![](_page_107_Figure_1.jpeg)

![](_page_107_Figure_2.jpeg)




























100 90 80 fl (ppm) 



2u





























## 15. Reference

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