

1 **Ammonia uptake by transmembrane pH gradient poly(isoprene)-block-poly(ethylene
2 glycol) polymersomes**
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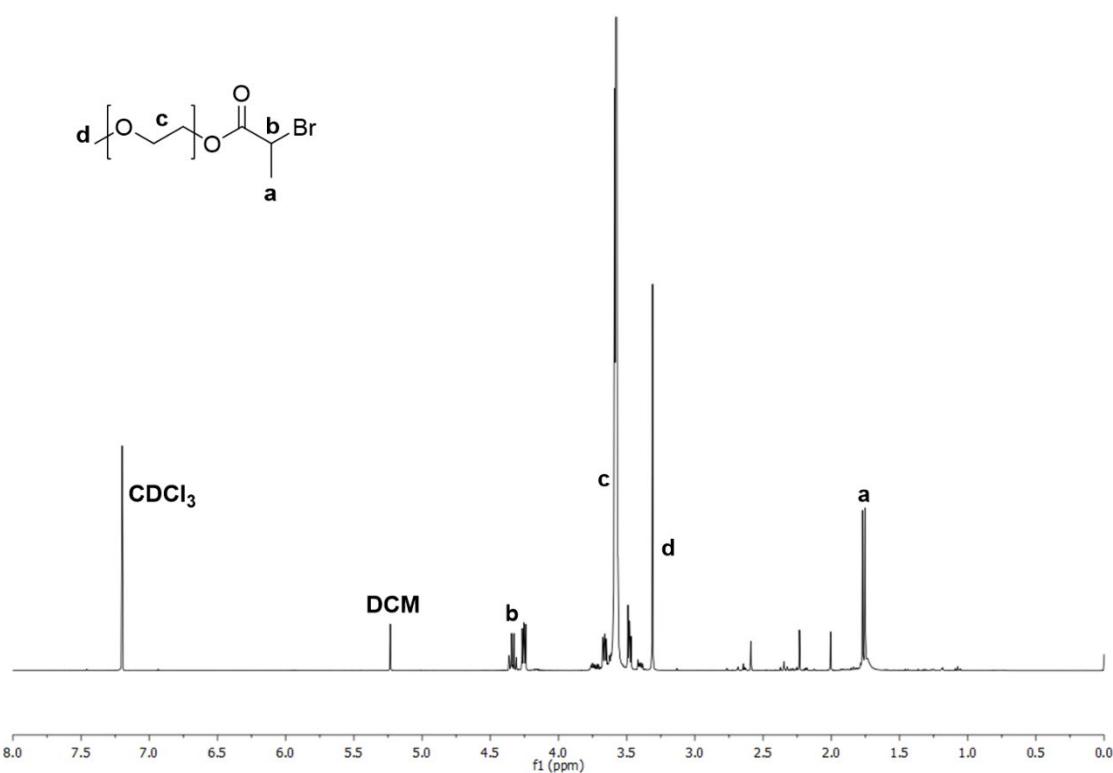
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10 **Supplementary Information**
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12 **Supplementary Figures**
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17 **Figure S1.** ¹H NMR spectrum of PEG₇₀₀-Br recorded in CDCl₃.
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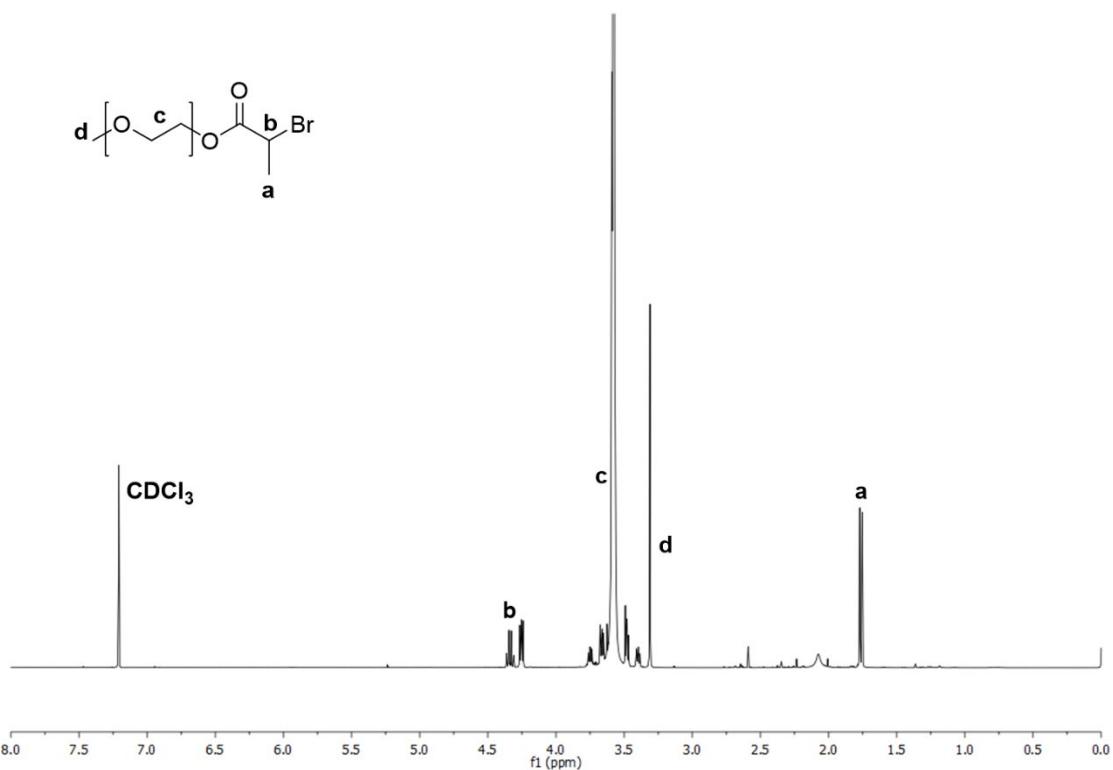
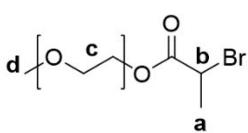


Figure S2. ¹H NMR spectrum of PEG₂₀₀₀-Br recorded in CDCl₃.

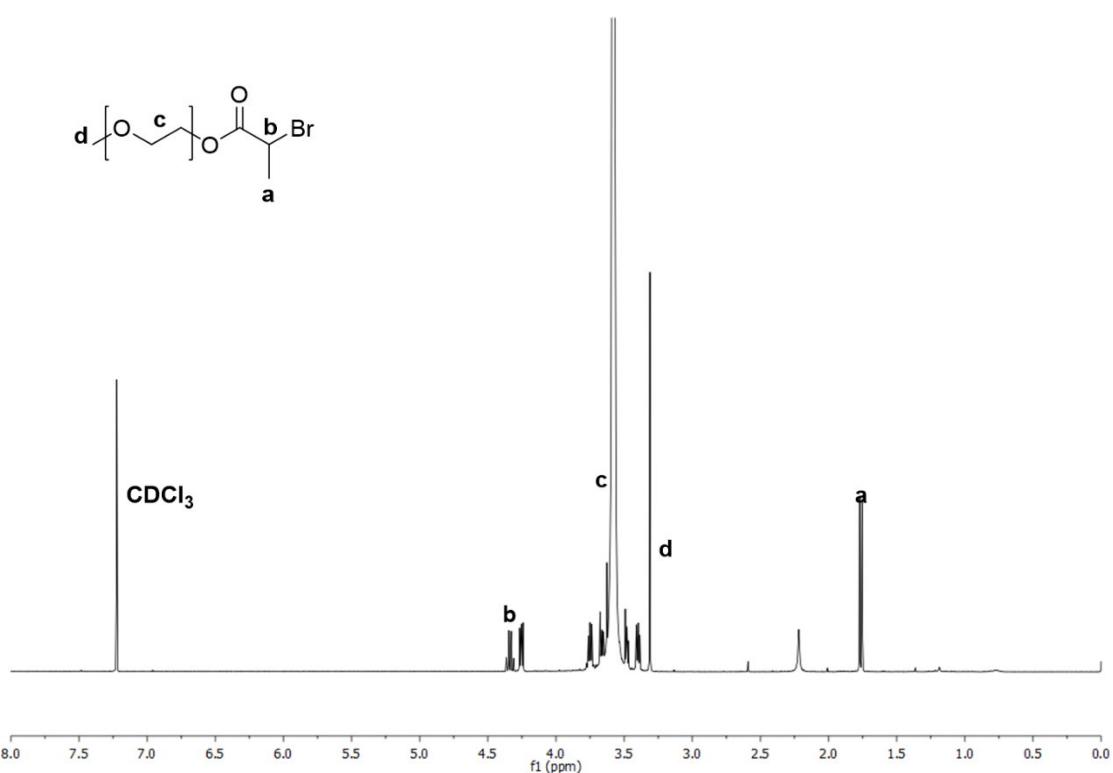
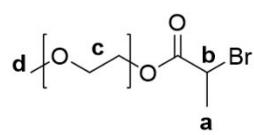


Figure S3. ¹H NMR spectrum of PEG₅₀₀₀-Br recorded in CDCl₃.

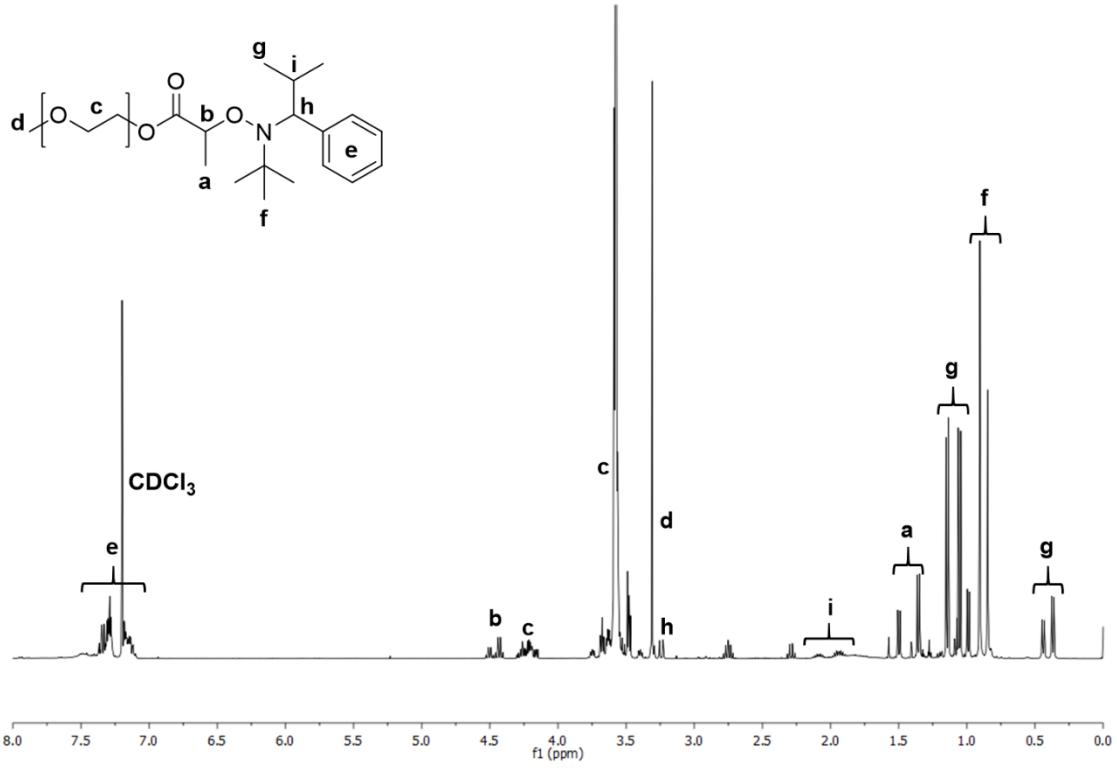


Figure S4. ¹H NMR spectrum of PEG₇₀₀-TIPNO recorded in CDCl₃.

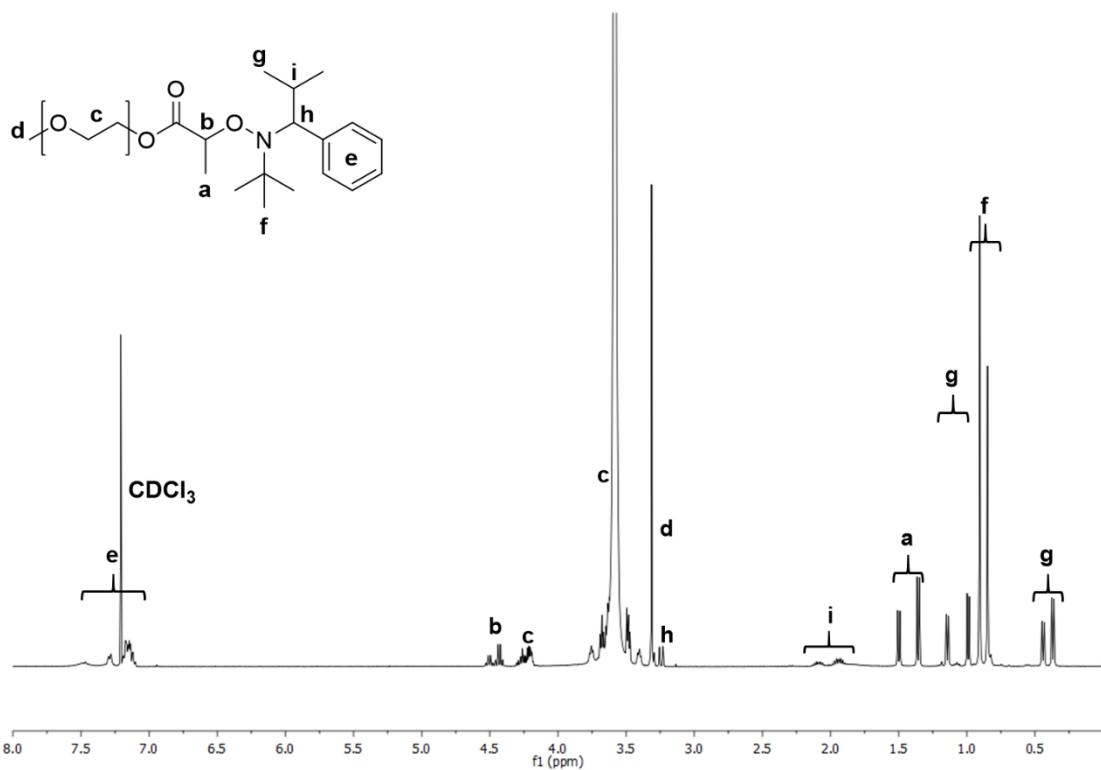
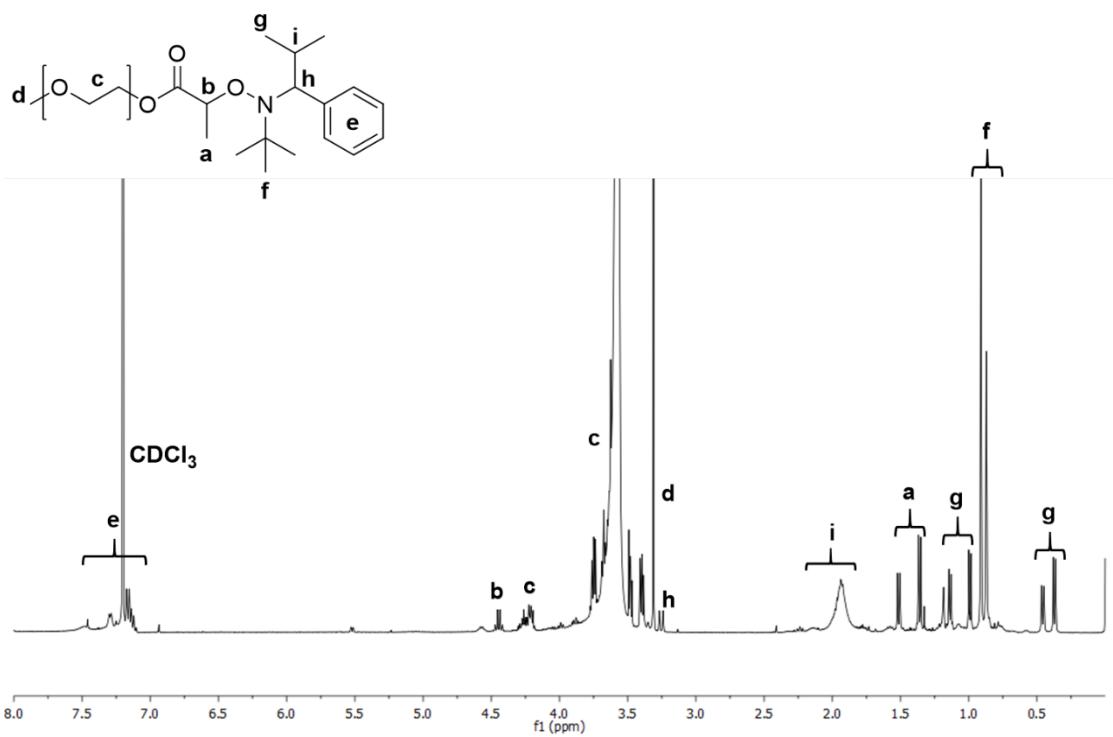
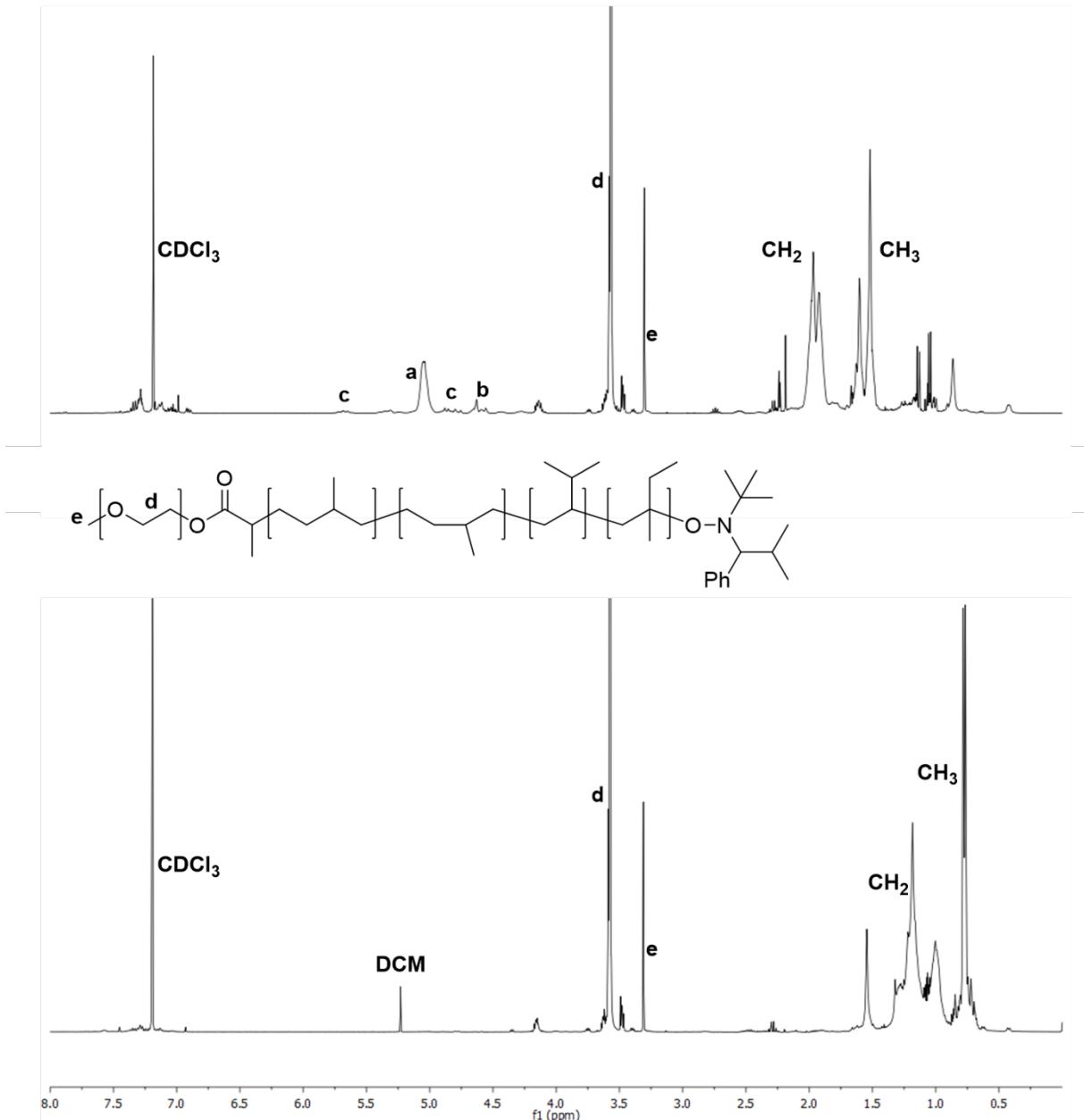
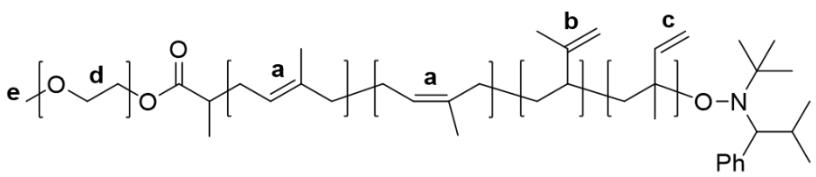


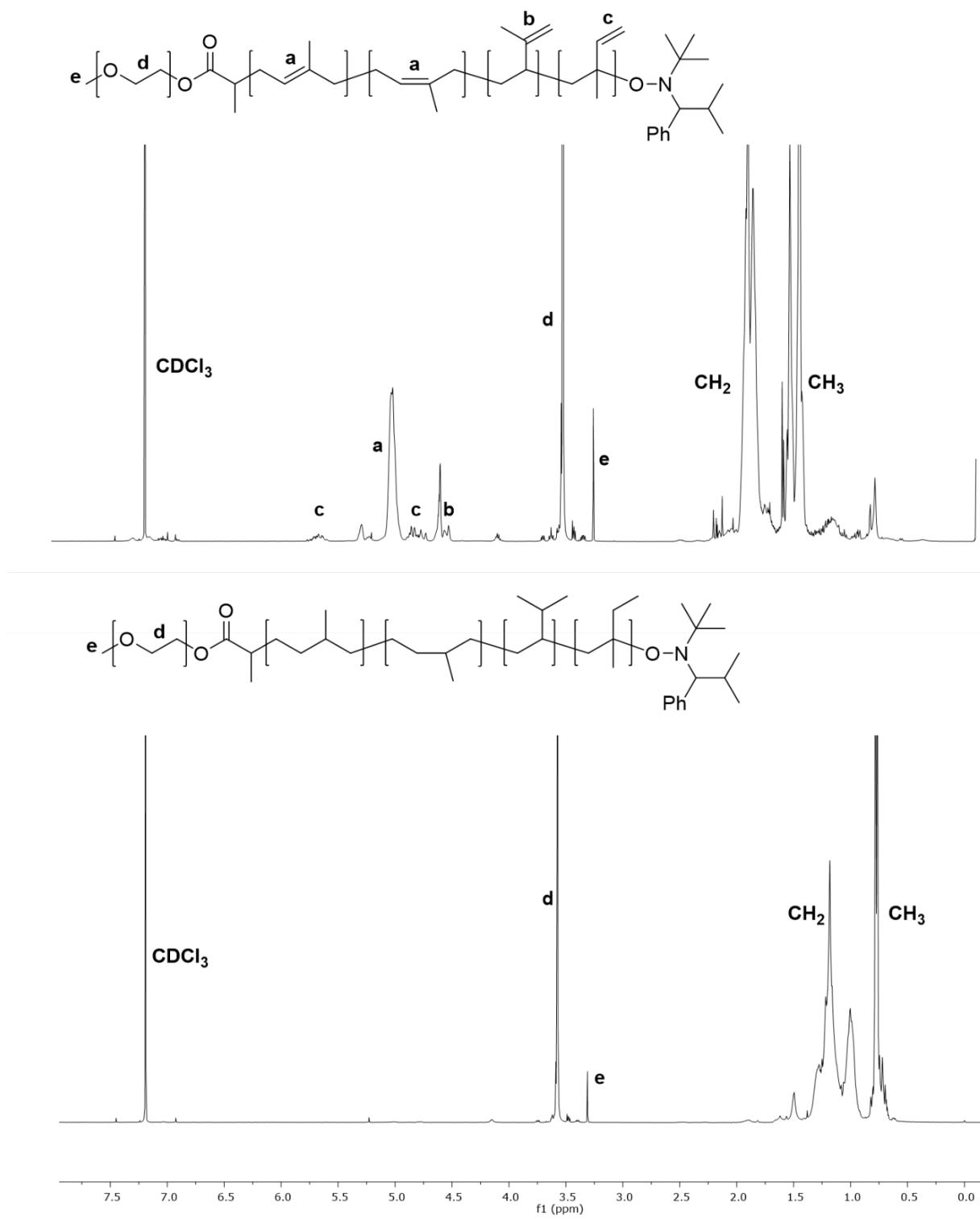
Figure S5. ¹H NMR spectrum of PEG₂₀₀₀-TIPNO recorded in CDCl₃.



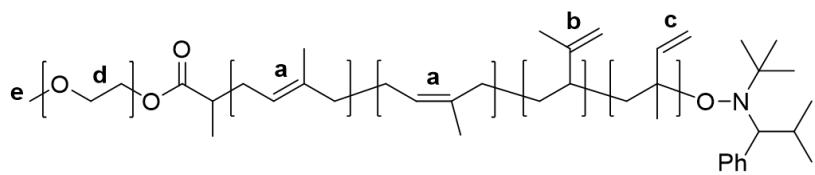
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2 **Figure S6.** ¹H NMR spectrum of PEG₅₀₀₀-TIPNO recorded in CDCl₃.



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 2 **Figure S7.** ^1H NMR spectra recorded in CDCl_3 of PI-*b*-PEG₇₀₀ polymer (PI/PEG (*w/w*) of 1.62)
 3 before (top) and after (bottom) the hydrogenation reaction.
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2 **Figure S8.** ^1H NMR spectra recorded in CDCl_3 of PI-*b*-PEG₂₀₀₀ polymer (PI/PEG (w/w) ratio
3 of 3.62) before (top) and after (bottom) the hydrogenation reaction.



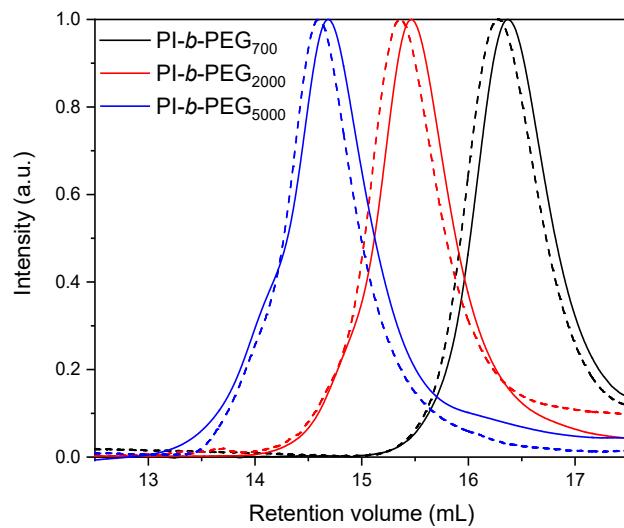


Figure S10. SEC traces of PI-*b*-PEG polymers of all three libraries before (straight lines) and after (dashed lines) hydrogenation. Recorded at 35 °C in THF and measured against PMMA standards.

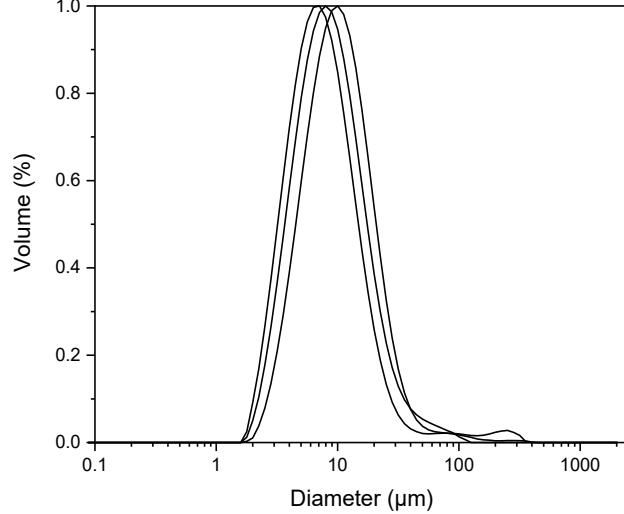


Figure S11. Size distribution of PI-*b*-PEG₂₀₀₀ polymer (PI/PEG (*w/w*) 1.99) by LD (*n* = 3).

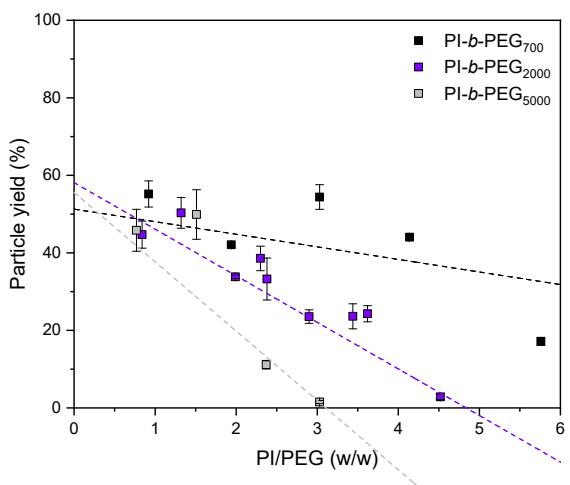


Figure S12. Particle yield of PI-*b*-PEG polymer assemblies with different PI/PEG (*w/w*) ratios. Particles were prepared using an emulsification process with an initial polymer feed of 30 mg (mean \pm SD, $n = 3$).

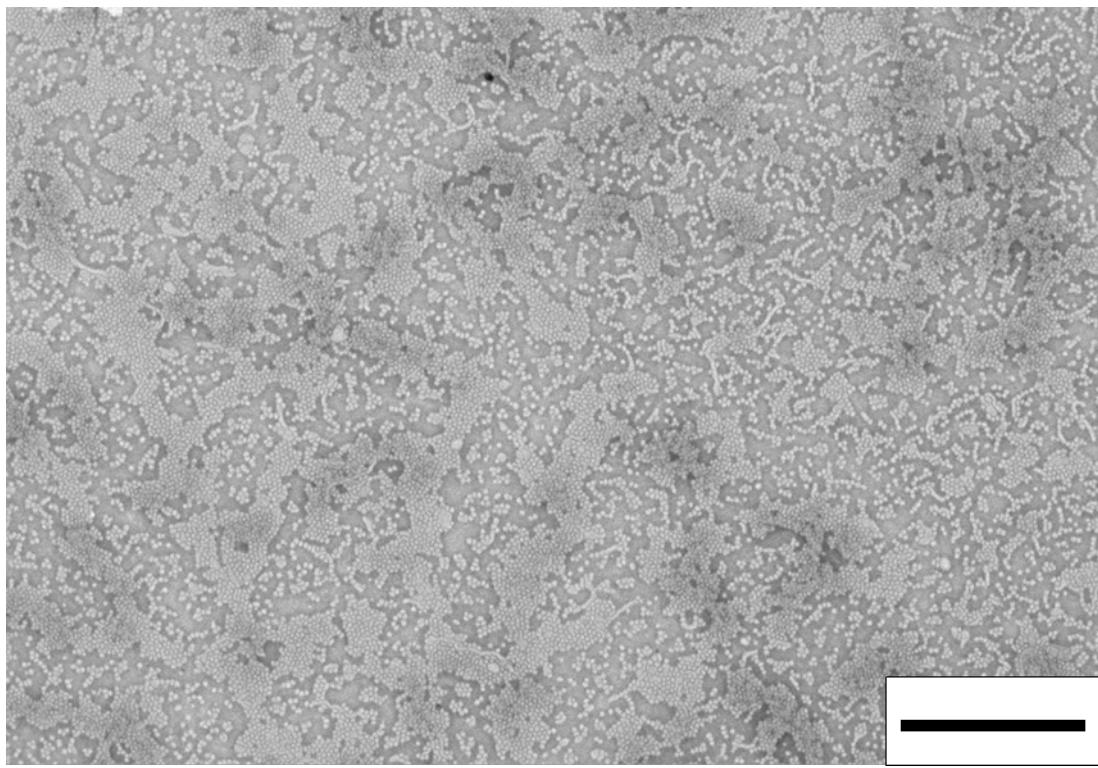
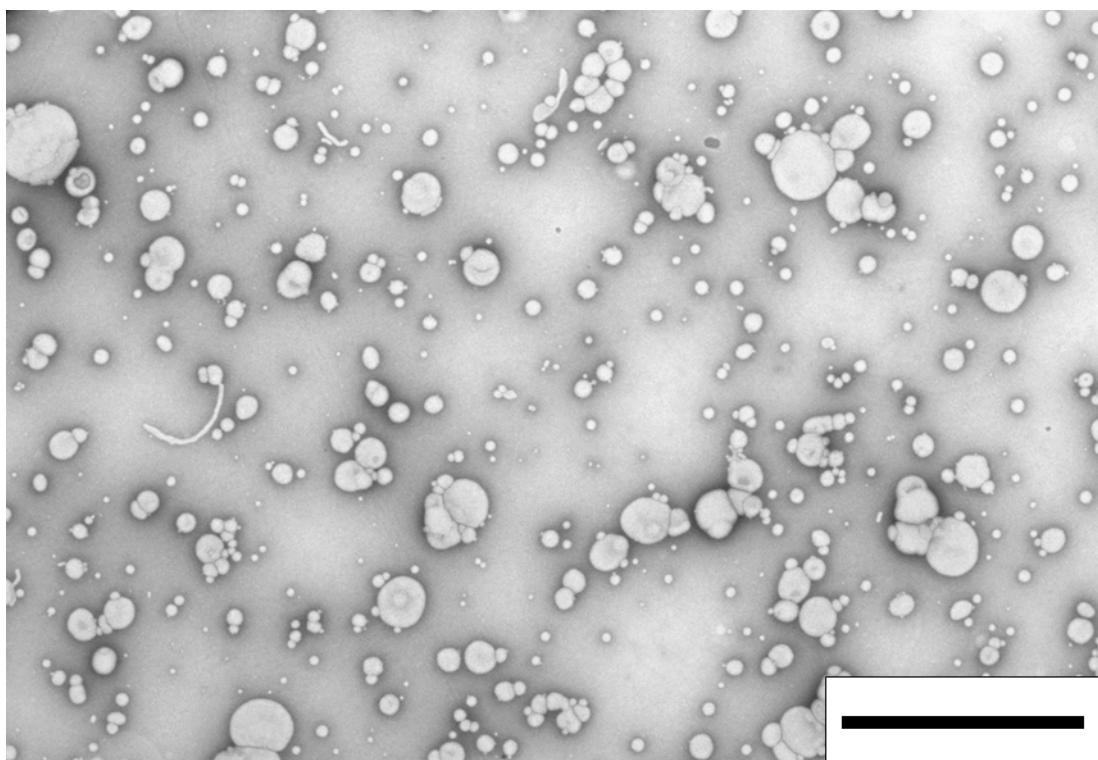
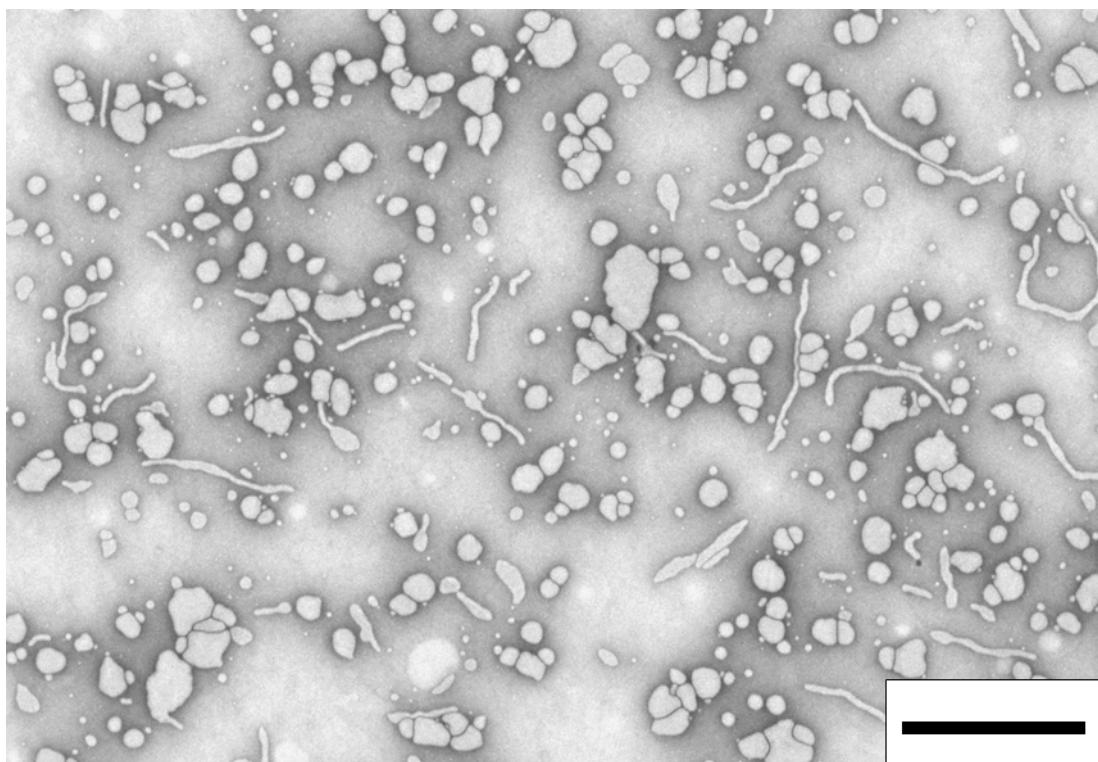
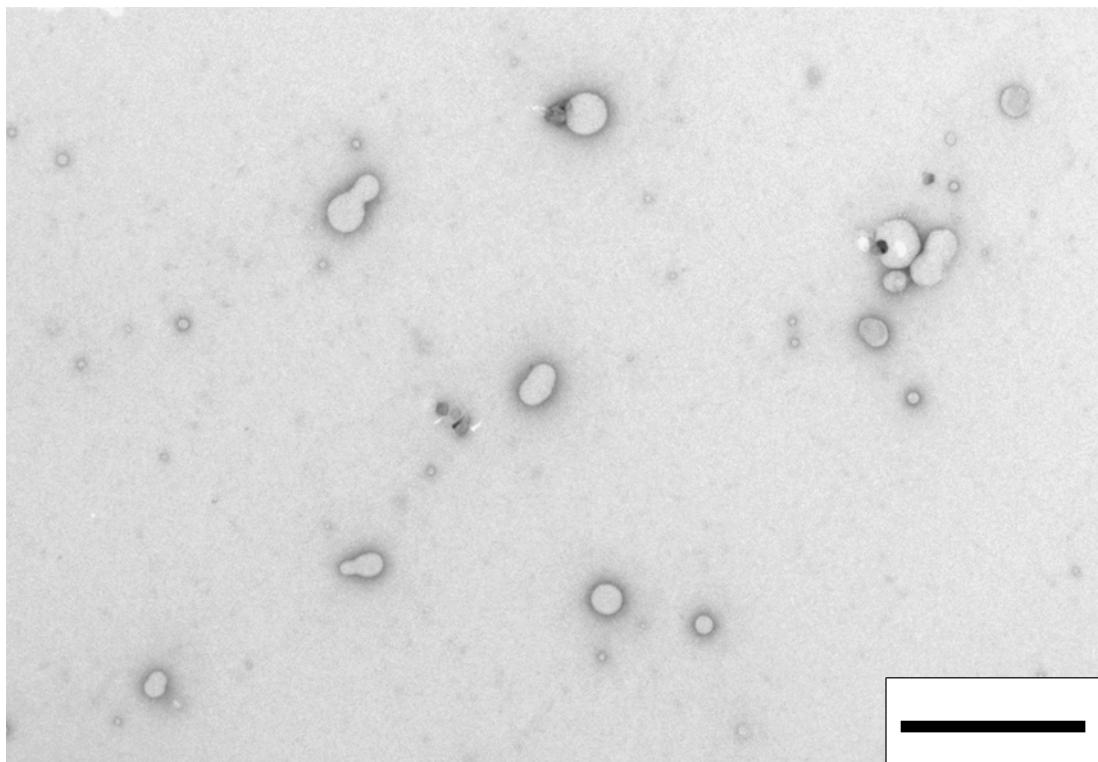
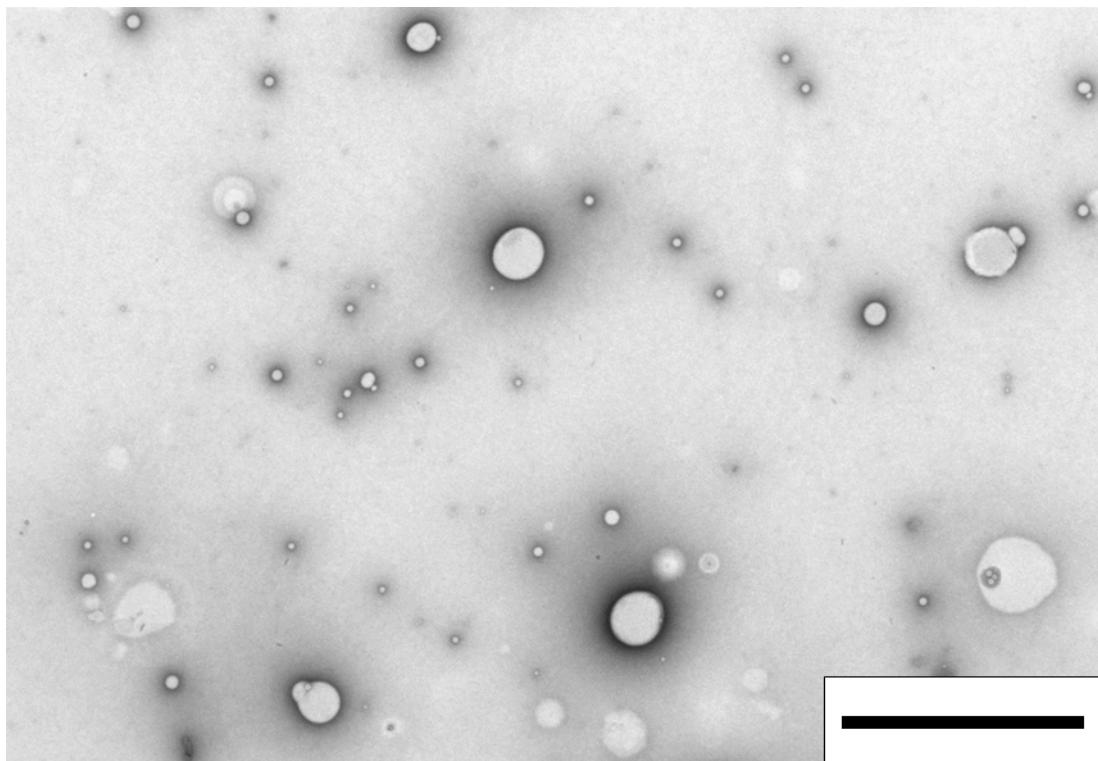


Figure S13. Representative TEM image of PI-*b*-PEG₂₀₀₀ (PI/PEG 0.84) after self-assembly in phosphate buffer at pH 6.8 (scale bar: 1000 nm).

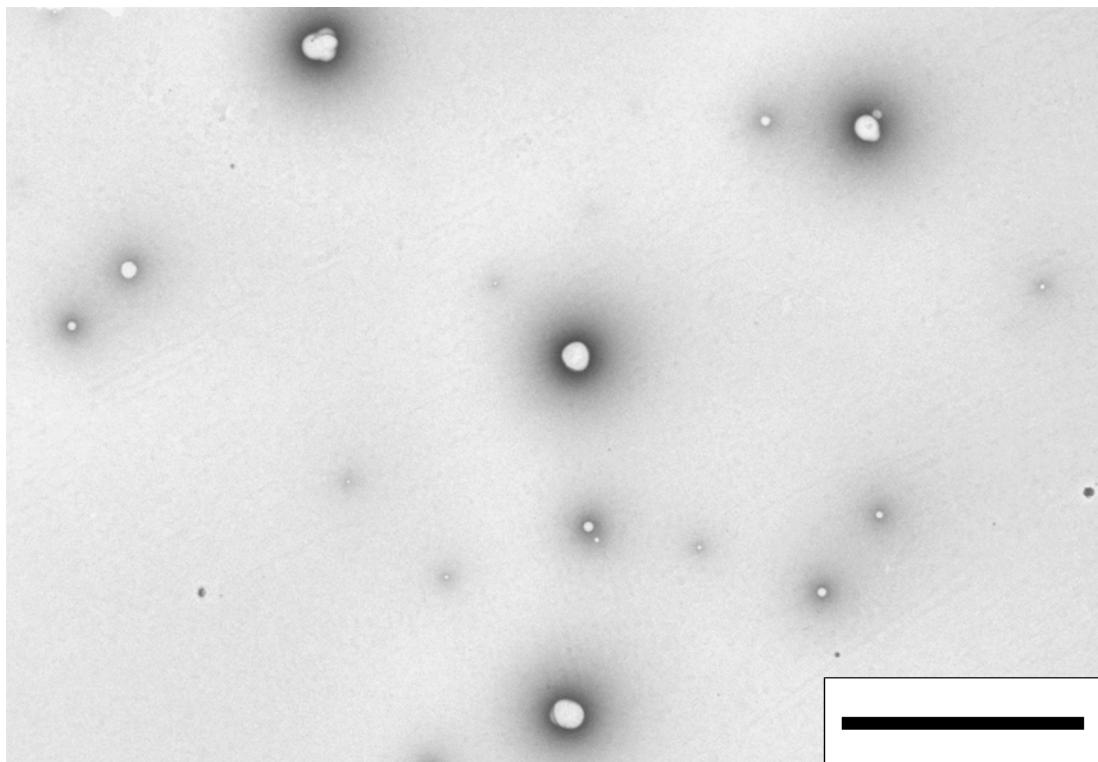




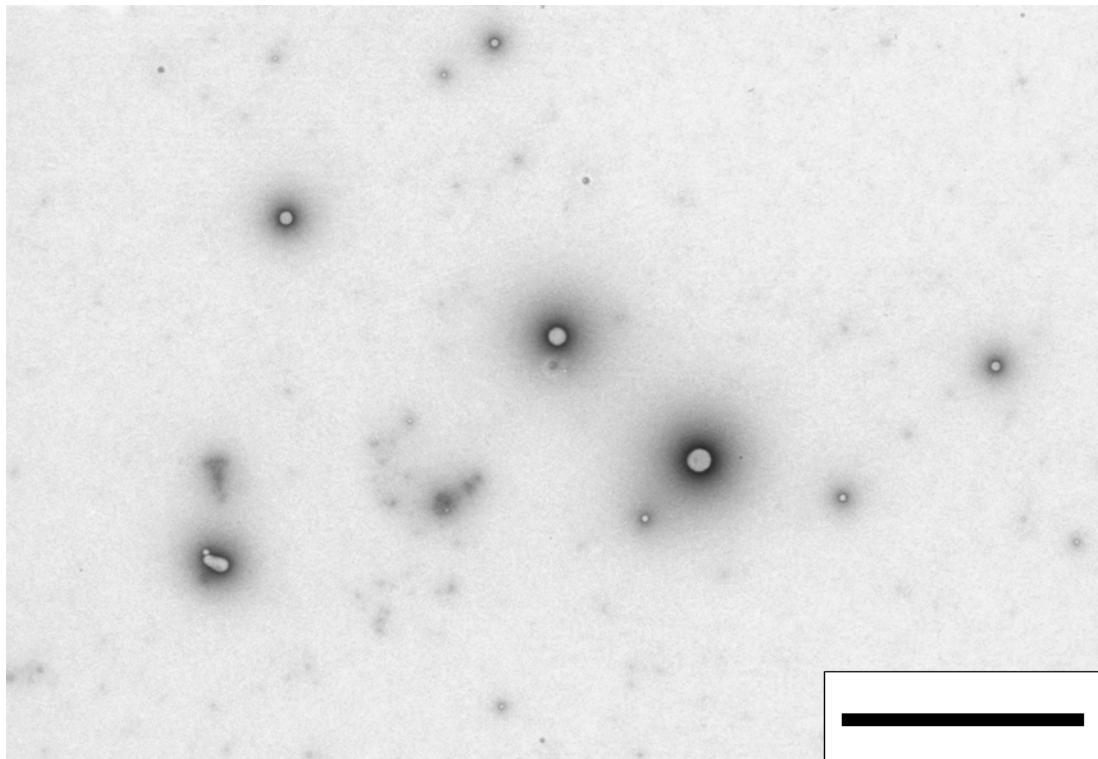
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2 **Figure S16.** Representative TEM image of PI-*b*-PEG₂₀₀₀ (PI/PEG 2.30) after self-assembly in
3 phosphate buffer at pH 6.8 (scale bar: 1000 nm).
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9 **Figure S17.** Representative TEM image of PI-*b*-PEG₂₀₀₀ (PI/PEG 2.90) after self-assembly in
10 phosphate buffer at pH 6.8 (scale bar: 2000 nm).
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2 **Figure S18.** Representative TEM image of PI-*b*-PEG₂₀₀₀ (PI/PEG 3.62) after self-assembly in
3 phosphate buffer at pH 6.8 (scale bar: 2000 nm).
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9 **Figure S19.** Representative TEM image of PI-*b*-PEG₂₀₀₀ (PI/PEG 4.52) after self-assembly in
10 phosphate buffer at pH 6.8 (scale bar: 2000 nm).
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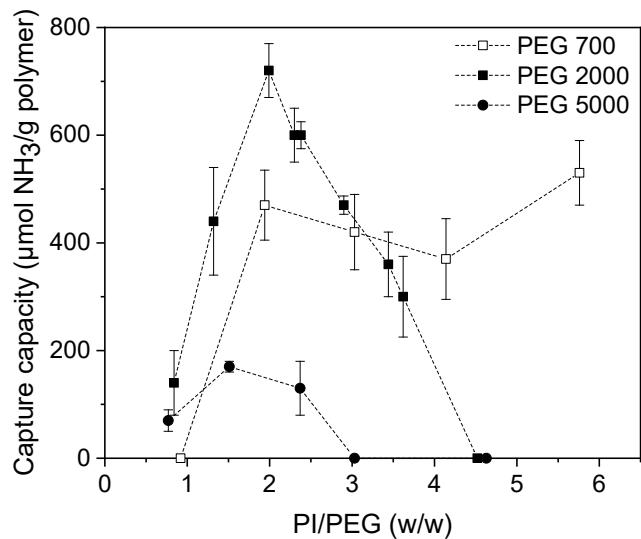


Figure S20. Ammonia capture capacity vs. PI/PEG (w/w) ratio after 24 h of incubation in phosphate buffer, pH 6.8 (mean \pm SD, n = 3).

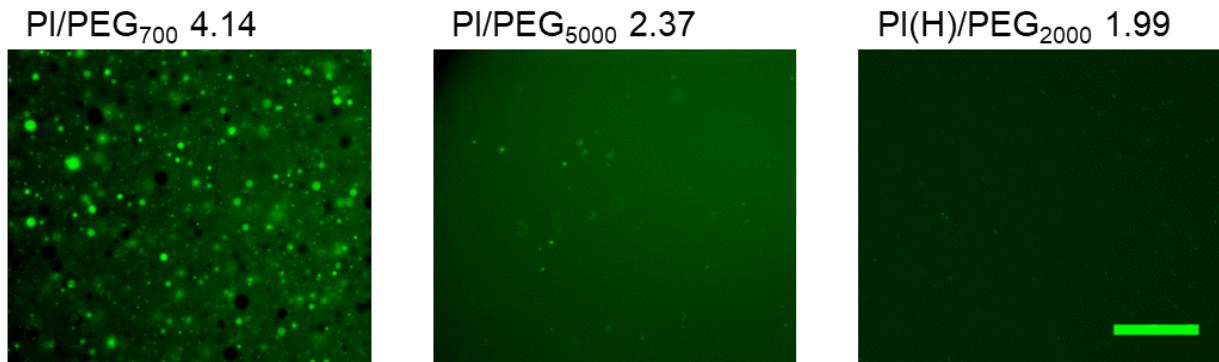


Figure S21. Representative fluorescence microscopy images of PI-*b*-PEG assemblies of PI-*b*-PEG₇₀₀, PI-*b*-PEG₅₀₀₀ and hydrogenated PI-*b*-PEG₂₀₀₀, loaded with pyranine (scale bar: 50 μm).

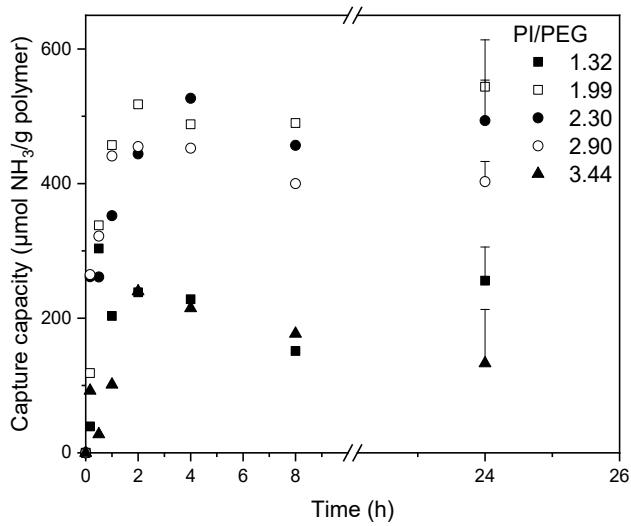


Figure S22. Ammonia capture by vesicles prepared with PI-*b*-PEG₂₀₀₀ polymers in simulated intestinal fluids over time (mean + SD, n = 3). Statistics were performed on the area under the capture capacity vs. time curve from 0 - 8 h (AUC_{0-8h}) (**Table S10**).



Figure S23. Image of crosslinked PI-*b*-PEG vesicles (PI/PEG 1.99) after resuspension in DCM.

1 **Supplementary tables**2
3 **Table S1.** Microstructure of PI-*b*-PEG amphiphiles synthesized in this study. Molar amount of
4 the different PI isomers present in the PI-*b*-PEG amphiphiles was calculated by ¹H NMR
5 spectroscopy. Data presented as % of total PI molar units.

Polymer	PI/PEG [w/w]	1,4-PI isomer [%]	1,2-PI isomer [%]	3,4-PI isomer [%]
PI- <i>b</i> -PEG ₇₀₀	0.92	89	4	7
	1.94	90	4	6
	3.03	88	4	8
	4.14	82	5	13
	5.76	81	5	14
PI- <i>b</i> -PEG ₂₀₀₀	0.84	90	4	6
	1.32	90	4	6
	1.99	86	4	10
	2.30	90	4	6
	2.38	90	4	6
	2.90	90	4	6
	3.44	89	4	7
	3.62	85	5	10
	4.52	90	4	6
	0.77	90	4	6
PI- <i>b</i> -PEG ₅₀₀₀	1.51	90	4	6
	2.37	89	4	7
	3.03	86	4	10
	4.63	88	4	8

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1 **Table S2.** Hydrogenated PI(H)-*b*-PEG amphiphiles and their respective polymersome
 2 ammonia capture capacity after 24 h incubation in phosphate buffer at pH 6.8. Polymersomes
 3 were produced using a sonication probe at an amplitude of 5 for 1 min.

	PI/PEG [w/w]	Yield [%] ^{a)}	Ammonia capture capacity [μmol NH ₃ /g polymer]
PI(H)- <i>b</i> -PEG ₇₀₀	1.62	76	-
	3.03	84	168
PI(H)- <i>b</i> -PEG ₂₀₀₀	0.84	100	-
	1.32	89	160
PI(H)- <i>b</i> -PEG ₅₀₀₀	1.99	91	85
	2.30	85	22
	3.62	94	*
	4.52	85	*
	0.77	100	*
	2.37	94	*
	3.03	97	*
	4.63	85	*

4 ^{a)}Obtained after hydrogenation reaction; - No uptake experiments performed; * No vesicles due to solubility issues
 5 of the PI(H)-*b*-PEG.
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1 **Table S3.** Ammonia capture capacities of PI-*b*-PEG polymersomes after 24 h (mean \pm SD, n =
 2 3).

	PI/PEG [w/w]	Ammonia capture capacity [$\mu\text{mol NH}_3/\text{g polymer}$] ^{a)}	Ammonia capture capacity [$\mu\text{mol NH}_3/\text{g polymer}$] ^{b)}
PI- <i>b</i> -PEG ₇₀₀	0.92	-	
	1.94	470 \pm 70	
	3.03	420 \pm 70	
	4.14	370 \pm 80	
	5.76	530 \pm 60	
PI- <i>b</i> -PEG ₂₀₀₀	0.84	140 \pm 60	-
	1.32	440 \pm 100	170 \pm 50
	1.99	720 \pm 50	540 \pm 10
	2.30	600 \pm 50	460 \pm 120
	2.38	600 \pm 25	
	2.90	470 \pm 20	410 \pm 30
	3.44	360 \pm 60	120 \pm 80
	3.62	300 \pm 80	
	4.52	-	
	0.77	70 \pm 20	
PI- <i>b</i> -PEG ₅₀₀₀	1.51	170 \pm 10	
	2.37	130 \pm 50	
	3.03	-	
	4.63	-	

3 ^{a)} Performed in phosphate buffer at pH 6.8; ^{b)} Performed in buffer supplemented with bile salt at pH 6.8; - No
 4 capture observed.

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1 **Table S4.** Statistical data of the *in vitro* ammonia capture experiments in phosphate buffer at
 2 pH 6.8 of PI-*b*-PEG₇₀₀ polymersomes. The area under the capture capacity *vs.* time curves from
 3 0-8 h (AUC_{0-8h}) were compared using a one-way ANOVA and subsequent Tukey's multiple
 4 comparisons test.

PI/PEG (<i>w/w</i>)	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
0.92 <i>vs.</i> 1.94	-4400	****	< 0.0001
0.92 <i>vs.</i> 3.03	-2700	****	< 0.0001
0.92 <i>vs.</i> 4.14	-2200	***	0.0001
0.92 <i>vs.</i> 5.76	-2600	****	< 0.0001
1.94 <i>vs.</i> 3.03	1700	**	0.0013
1.94 <i>vs.</i> 4.14	2200	***	0.0002
1.94 <i>vs.</i> 5.76	1800	***	0.0007
3.03 <i>vs.</i> 4.14	500	n.s.	0.4609
3.03 <i>vs.</i> 5.76	100	n.s.	0.993
4.14 <i>vs.</i> 5.76	-400	n.s.	0.6889

5 n.s.: not significant.

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8 **Table S5.** Statistical data of the *in vitro* ammonia capture experiments in phosphate buffer at
 9 pH 6.8 of PI-*b*-PEG₂₀₀₀ polymersomes. The area under the capture capacity *vs.* time curves
 10 from 0-8 h (AUC_{0-8h}) were compared using a one-way ANOVA and subsequent Tukey's
 11 multiple comparisons test.

PI/PEG (<i>w/w</i>)	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
0.84 <i>vs.</i> 1.32	-2900	****	< 0.0001
0.84 <i>vs.</i> 1.99	-4600	****	< 0.0001
0.84 <i>vs.</i> 2.30	-3300	****	< 0.0001
0.84 <i>vs.</i> 2.38	-3600	****	< 0.0001
0.84 <i>vs.</i> 2.90	-2500	****	< 0.0001
0.84 <i>vs.</i> 3.44	-1400	*	0.0411
0.84 <i>vs.</i> 3.62	-900	n.s.	0.2994
0.84 <i>vs.</i> 4.52	1200	n.s.	0.0897
1.32 <i>vs.</i> 1.99	-1600	*	0.0105
1.32 <i>vs.</i> 2.30	-400	n.s.	0.981
1.32 <i>vs.</i> 2.38	-600	n.s.	0.7522
1.32 <i>vs.</i> 2.90	400	n.s.	0.9666
1.32 <i>vs.</i> 3.44	1600	*	0.0121
1.32 <i>vs.</i> 3.62	2000	**	0.0012
1.32 <i>vs.</i> 4.52	4100	****	< 0.0001
1.99 <i>vs.</i> 2.30	1200	n.s.	0.0772

1.99 vs. 2.38	1000	n.s.	0.2515
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Continuation Table S5

1.99 vs. 2.90	2000	**	0.0011
1.99 vs. 3.44	3200	****	< 0.0001
1.99 vs. 3.62	3600	****	< 0.0001
1.99 vs. 4.52	5800	****	< 0.0001
2.30 vs. 2.38	-300	n.s.	0.9985
2.30 vs. 2.90	800	n.s.	0.5014
2.30 vs. 3.44	2000	**	0.0015
2.30 vs. 3.62	2400	***	0.0002
2.30 vs. 4.52	4500	****	< 0.0001
2.38 vs. 2.90	1000	n.s.	0.189
2.38 vs. 3.44	2200	***	0.0004
2.38 vs. 3.62	2600	****	< 0.0001
2.38 vs. 4.52	4800	****	< 0.0001
2.90 vs. 3.44	1200	n.s.	0.1059
2.90 vs. 3.62	1600	*	0.0117
2.90 vs. 4.52	3700	****	< 0.0001
3.44 vs. 4.52	400	n.s.	0.9641
3.44 vs. 3.62	2600	****	< 0.0001
3.62 vs. 4.52	2100	***	0.0006

1 n.s.: not significant.

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1 **Table S6.** Statistical data of the *in vitro* ammonia capture experiments in phosphate buffer at
 2 pH 6.8 of PI-*b*-PEG₅₀₀₀ polymersomes. The area under the capture capacity *vs.* time curves
 3 from 0-8 h (AUC_{0-8h}) were compared using a one-way ANOVA and subsequent Tukey's
 4 multiple comparisons test.

PI/PEG (<i>w/w</i>)	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
0.77 <i>vs.</i> 1.51	-600	n.s.	0.0548
0.77 <i>vs.</i> 2.37	-800	*	0.0158
0.77 <i>vs.</i> 3.03	400	n.s.	0.3742
0.77 <i>vs.</i> 4.63	400	n.s.	0.3742
1.51 <i>vs.</i> 2.37	-200	n.s.	0.9239
1.51 <i>vs.</i> 3.03	1000	**	0.0031
1.51 <i>vs.</i> 4.63	1000	**	0.0031
2.37 <i>vs.</i> 3.03	1100	**	0.001
2.37 <i>vs.</i> 4.63	1100	**	0.001
3.03 <i>vs.</i> 4.63	0	n.s.	> 0.9999

5 n.s.: not significant.

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9 **Table S7.** Statistical data of the *in vitro* ammonia capture experiments using PI-*b*-PEG
 10 polymersomes (PI/PEG 1.99). The area under the capture capacity *vs.* time curves from 0-8 h
 11 (AUC_{0-8h}) were compared using an unpaired t-test.

Experiment	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
Phosphate buffer pH 6.8 <i>vs.</i> bile salt buffer pH 6.8	-1900	**.	0.0027
No pH gradient <i>vs.</i> pH gradient in phosphate buffer	-5500	****	< 0.0001
Hydrogenated polymer <i>vs.</i> non-hydrogenated polymer in phosphate buffer	-5800	****	< 0.0001

12 n.s.: not significant.

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1 **Table S8.** Statistical data of the *in vitro* ammonia capture experiments using native (Nat) or
 2 crosslinked (CL) PI-*b*-PEG₂₀₀₀ polymersomes in phosphate buffer (Pho) or colon-mimicking
 3 environment (Meta) (PI/PEG 1.99, unless indicated otherwise). For multiple groups ammonia
 4 capture values obtained after 24 h were compared using a one-way ANOVA and subsequent
 5 Tukey's multiple comparisons test. In case of experiments with only two groups, ammonia
 6 capture values obtained after 24 h were compared using an unpaired t-test.

	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
Nat Meta vs. Nat Pho	-520	***	0.0005
PI/PEG 2.30, Nat Meta vs. Nat Pho	-410	**	0.0014
CL Pho vs. CL Meta	380	***	0.0003
CL Pho vs. Nat Pho	100	n.s.	0.2909
CL Pho vs. Nat Meta	620	****	< 0.0001
CL Meta vs. Nat Pho	-280	**	0.0026
CL Meta vs. Nat Meta	230	**	0.0087
Nat Pho vs. Nat Meta	520	****	< 0.0001

7 n.s.: not significant.

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9 **Table S9.** Statistical data of the *in vitro* ammonia capture experiments in phosphate buffer at
 10 pH 6.8 of crosslinked polymersomes (PI/PEG₂₀₀₀ 1.99) with different concentrations of
 11 crosslinker. Ammonia capture capacity values obtained after 24 h were compared using a one-
 12 way ANOVA and subsequent Tukey's multiple comparisons test.

Concentration of crosslinker [mM]	Mean difference [$\mu\text{mol g}^{-1} \text{h}^{-1}$]	Significance	p value
0 vs. 0.5	100	n.s.	0.4543
0 vs. 1	190	*	0.0385
0 vs. 2	320	***	0.0008
0 vs. 5	370	***	0.0002
0 vs. 10	340	***	0.0005
0.5 vs. 1	90	n.s.	0.5963
0.5 vs. 2	220	*	0.0177
0.5 vs. 5	260	**	0.0044
0.5 vs. 10	240	*	0.0101
1 vs. 2	130	n.s.	0.2485
1 vs. 5	180	n.s.	0.0666
1 vs. 10	150	n.s.	0.1509
2 vs. 5	50	n.s.	0.9537
2 vs. 10	20	n.s.	0.9993
5 vs. 10	-30	n.s.	0.9949

13 n.s.: not significant.

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2 **Table S10.** Statistical data of the *in vitro* ammonia capture experiments in simulated intestinal
3 fluids at pH 6.8 of PI-*b*-PEG₂₀₀₀ polymersomes. The area under the capture capacity vs. time
4 curves from 0-8 h (AUC_{0-8h}) were compared using a one-way ANOVA and subsequent Tukey's
5 multiple comparisons test.

PI/PEG (w/w)	Mean difference [μmol g ⁻¹ h ⁻¹]	Significance	p value
0.84 vs. 1.32	-2100	*	0.0345
0.84 vs. 1.99	-3800	***	0.0003
0.84 vs. 2.30	-3700	***	0.0004
0.84 vs. 2.90	-3400	***	0.001
0.84 vs. 3.44	-1500	n.s.	0.1884
1.32 vs. 1.99	-1700	n.s.	0.0934
1.32 vs. 2.30	-1600	n.s.	0.1435
1.32 vs. 2.90	-1300	n.s.	0.3097
1.32 vs. 3.44	600	n.s.	0.8936
1.99 vs. 2.30	200	n.s.	0.9998
1.99 vs. 2.90	500	n.s.	0.9635
1.99 vs. 3.44	2400	*	0.0162
2.30 vs. 2.90	300	n.s.	0.9941
2.30 vs. 3.44	2200	*	0.0256
2.90 vs. 3.44	1900	n.s.	0.0617

6 n.s.: not significant.