

Supplementary materials

Creating locally crowded environment with nanoclay hydrogel for cell-free biosynthesis

Xinjie Chen^{1,#}, Qi Sun^{1,#}, and Yuan Lu^{1,*}

¹Key Laboratory of Industrial Biocatalysis, Ministry of Education, Department of Chemical Engineering, Tsinghua University, Beijing 100084, China.

[#]These authors contributed equally to this work.

^{*}Corresponding author. Email: yuanlu@tsinghua.edu.cn (Y. L.).

Supplementary Table S1. Composition of plasmid labeling system

Components	Reaction system (50 μ L)
10 \times Labeling Buffer A	5 μ L
1 mg/mL DNA template	5 μ L
Label IT®Tracker Reagent	2.5 μ L
ddH ₂ O	37.5 μ L

Supplementary Table S2. Quantitative real-time PCR reaction components

Components	Reaction system (20 μ L)
Forward primer (10 μ M)	0.4 μ L
Reverse primer (10 μ M)	0.4 μ L
2 \times TransStart Top Green QPCR SuperMix	10 μ L
Passive Reference Dye (50 \times)	0.4 μ L
Nuclease-free Water	4 μ L
DNA template	4.8 μ L

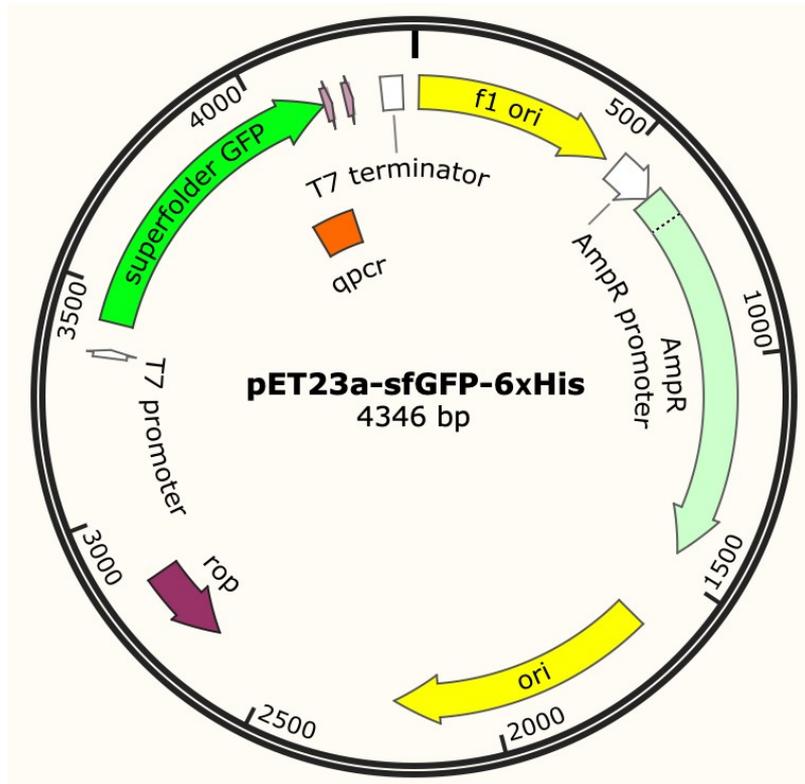
Supplementary Table S3. Quantitative real-time PCR reaction system

Steps	Temperature	Time
Stage1	94°C	30 s
	94°C	5 s
Stage2 (40×)	57°C	15 s
	72°C	31 s
	95°C	15 s
Stage3	60°C	60 s
	95°C	15 s

Supplementary Table S4. List of nanoclay solution concentration and CaCl₂

solution concentration

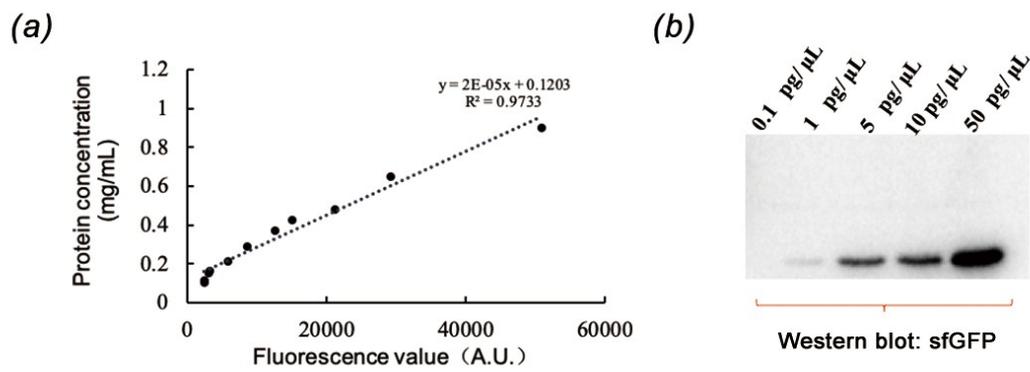
Nanoclay solution (mg/mL)	CaCl ₂ solution (mM)
2	10
4	20
6	30
8	40
10	50
12	60
14	70
16	80
18	90
20	100



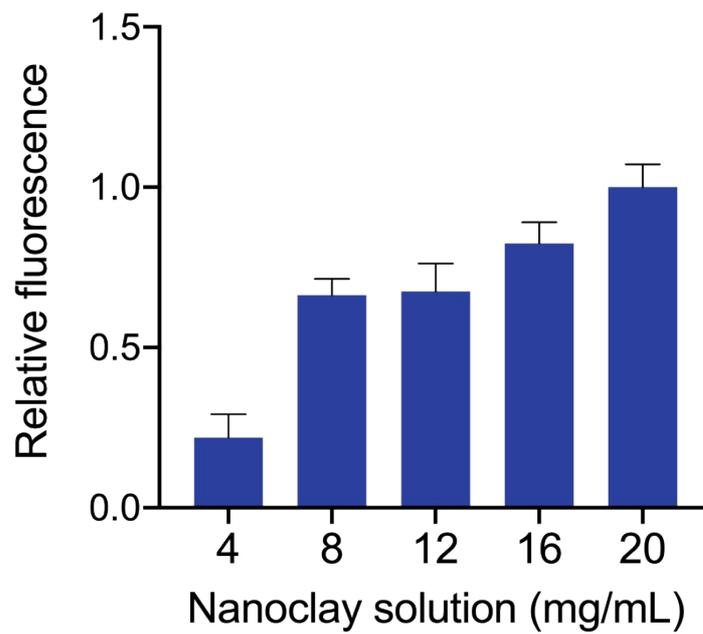
T7 promoter-sfGFP-6xHis-T7 terminator

TAATACGACTCACTATAGG GAGACCACAACGGTTTCCCTCTAGAAATAAT
 TTTGTTAACTTTAAGAAGGAGATACAT ATGCGTAAAGGCGAAGAGCT
 GTTCACTGGTGTTCGTCCTATTCTGGTGGAACTGGATGGTGTGATGTCAACGG
 TCATAAGTTTTCCGTGCGTGGCGAGGGTGAAGGTGACGCAACTAATGGTA
 AACTGACGCTGAAGTTCATCTGTACTACTGGTAAACTGCCGTACCTTGGC
 CGACTCTGGTAACGACGCTGACTTATGGTGTTCAGTGCTTTGCTCGTTATC
 CGGACCATATGAAGCAGCATGACTTCTTCAAGTCCGCCATGCCGGAAGGC
 TATGTGCAGGAACGCACGATTTCTTTAAGGATGACGGCACGTACAAAAC
 GCGTGCGGAAGTGAAATTTGAAGGCGATACCCTGGTAAACCGCATTGAGC
 TGAAAGGCATTGACTTTAAAGAAGACGGCAATATCCTGGGCCATAAGCTG
 GAATACAATTTTAACAGCCACAATGTTTACATCACCGCCGATAAACAAAA
 AAATGGCATTAAAGCGAATTTTAAAATTCGCCACAACGTGGAGGATGGCA
 GCGTGACGCTGGCTGATCACTACCAGCAAAACACTCCAATCGGTGATGGT
 CCTGTTCTGCTGCCAGACAATCACTATCTGAGCACGCAAAGCGTTCTGTCT
 AAAGATCCGAACGAGAAACGCGATCATATGGTTCTGCTGGAGTTCGTAAC
 CGCAGCGGGCATCACGCATGGTATGGATGAACTGTACAAA CATCACCATC
 ACCATCATTAAGTCGACAAGCTTTCGGCCGCACTCGAG CACCACCACCAC
 CACCAC TGAGATCCGGCTGCTAACAAAGCCCGAAAGGAAGCTGAGTTGGC
 TGCTGCCACCGCTGAGCAATAA CTAGCATAACCCCTGGGGCCTCTAAAC
 GGGTCTTGAGGGGTTTTTTG

Supplementary Figure S1. The pET23a-sfGFP-6242\252His plasmid map and the sequence information. The sfGFP was regulated under the bacteriophage derived T7 promoter and terminator in the pET-23a backbone. The last 150 bps of sfGFP were designed for the qPCR (Real-time Quantitative PCR).



Supplementary Figure S2. Fluorescence value could represent the protein expression level. (a) Standard curve of sfGFP. The fluorescence value of sfGFP increased linearly with the concentration of protein, which indicated that the protein expression level could be characterized by the fluorescence, and the influence of the environment could be ignored. (b) The results of Western blot. The gray-scale value increased linearly with the concentration of protein.



Supplementary Figure S3. The protein expression level with different concentrations of nanoclay solution. When the nanoclay solution concentration was 20 mg/mL, the fluorescence of sfGFP was set to 1.