

Supporting Information

Trifunctional modification of individual bacterial cells for magnet-assisted bioanode with high performance in microbial fuel cells

Yujing Jiang,^a Pingping Li,^a Yuanyuan Wang,^a Li-Ping Jiang,^a Rong-Bin Song,^{*ab}
Jian-Rong Zhang^{*ac} and Jun-Jie Zhu^{*a}

^a State Key Laboratory of Analytical Chemistry for Life Science, School of Chemistry and Chemical Engineering, Nanjing University, Nanjing 210023, P.R. China

E-mail: rbsong@nju.edu.cn; jrzhang@nju.edu.cn; jjzhu@nju.edu.cn

^b School of Ecology and Environment, Zhengzhou University, Zhengzhou 450001, P.R. China

E-mail: rbsong@zzu.edu.cn

^c School of Chemistry and Life Science, Nanjing University Jingling College, Nanjing 210089, P.R. China

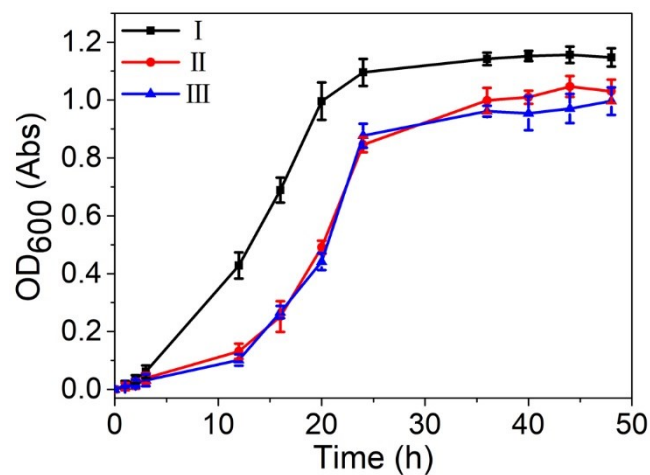


Fig. S1 Growth curves of native *S. oneidensis* MR-1 (curve I), *S. oneidensis* MR-1@Au (curve II) and *S. oneidensis* MR-1@Au@Fe₃O₄ (curve III). Error bars represent standard error (s.e.) determined by three independent experiments.

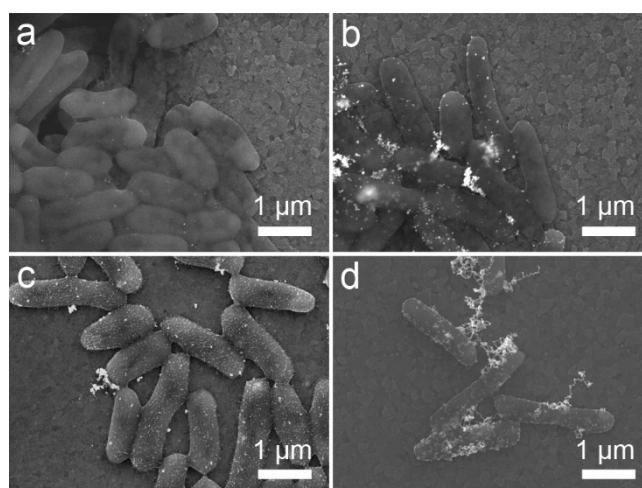


Fig. S2 SEM images of *S. oneidensis* MR-1@Au with different Au content. (a) 0.5 mM L⁻¹, (b) 1 mM L⁻¹, (c) 1.5 mM L⁻¹, (d) 2 mM L⁻¹.

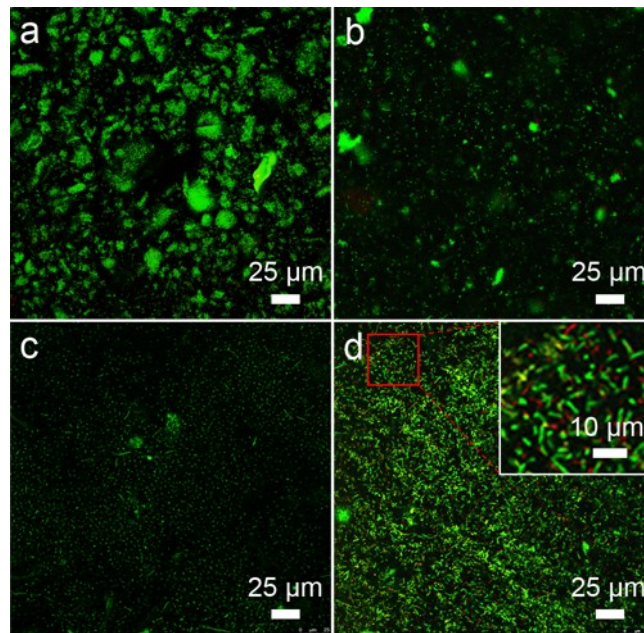


Fig. S3 CLSM images of *S. oneidensis* MR-1@Au with different Au content. (a) 0.5 mM L⁻¹, (b) 1 mM L⁻¹, (c) 1.5 mM L⁻¹, (d) 2 mM L⁻¹. The inset in image d is the corresponding high-magnification CLSM image.

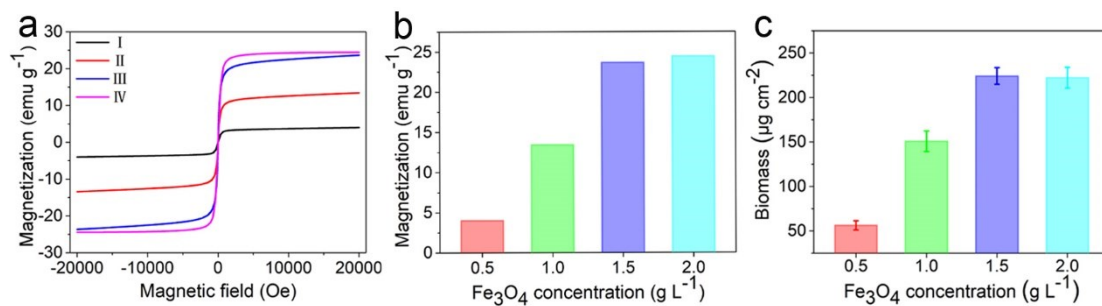


Fig. S4 The hysteresis loops (a), saturation magnetization (b) and biomass (c) of *S. oneidensis* MR-1@Au@Fe₃O₄ at different Fe₃O₄ concentration (curve I, 0.5 g L⁻¹; curve II, 1 g L⁻¹; curve III, 1.5 g L⁻¹; curve IV, 2 g L⁻¹).

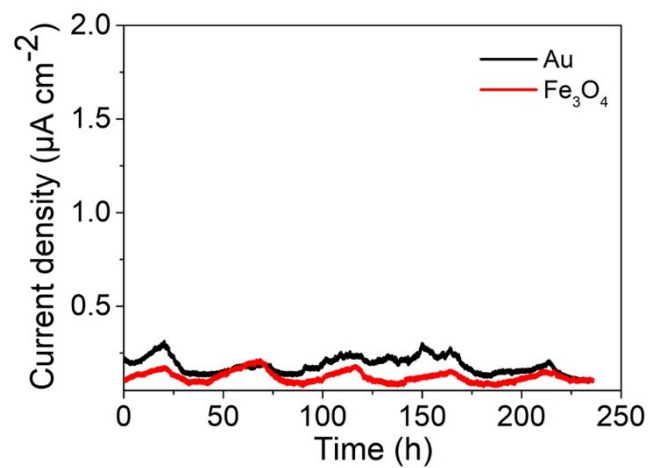


Fig. S5 Time profile of electricity generation of magnetic substrate electrode with Au and Fe_3O_4 .

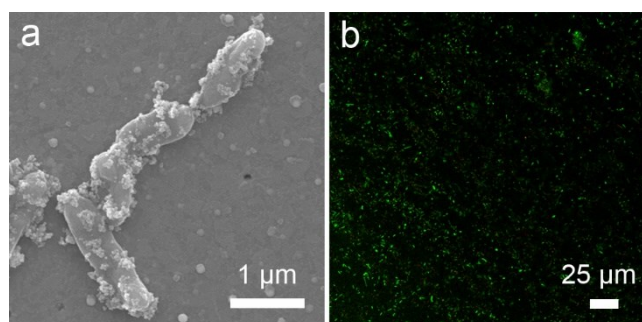


Fig. S6 SEM image (a) and CLSM image (b) of *S. oneidensis* MR-1@ Fe_3O_4 after modification.

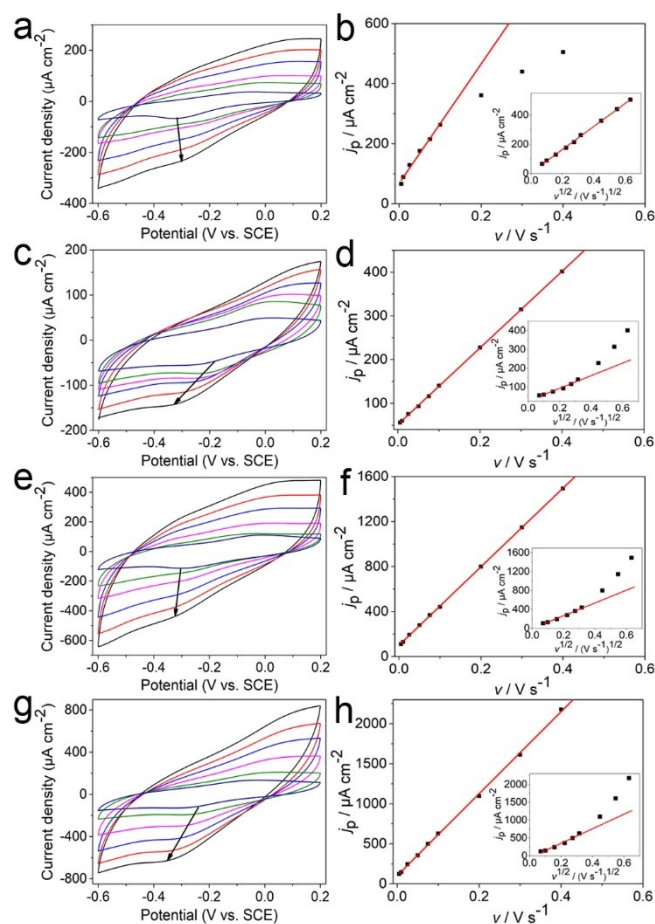


Fig. S7 Cyclic voltammograms of *S. oneidensis* MR-1 (a), *S. oneidensis* MR-1@Au (c), *S. oneidensis* MR-1@Fe₃O₄ (e) and *S. oneidensis* MR-1@Au@Fe₃O₄ (g) biofilms at different scan rates (arrows showed scan rates at 5, 10, 25, 50, 75 and 100 mV s^{-1} , respectively). Dependence of reduction current density (j_p) versus scan rate (ν) on *S. oneidensis* MR-1 (b), *S. oneidensis* MR-1@Au (d), *S. oneidensis* MR-1@Fe₃O₄ (f) and *S. oneidensis* MR-1@Au@Fe₃O₄ (h) biofilms, separately; Inset: linear dependence of j_p versus $\nu^{1/2}$.

Table S1. Comparison of the performance of previous MFCs using Au, Fe₃O₄ and their corresponding hybrids as anodes, as well as functionalized bacterial cells as bioanode.

Electrode substrates	Anode materials	Microbe type	Power density (mW m ⁻²)	Ref.
Graphite felt	Fe ₃ O ₄	Mixed bacteria	18.28	1
Carbon felt	MWCNT-Au-Pt/ osmium redox polymer	<i>Gluconobacter oxydans</i>	32.1	2
Carbon cloth	BioAu/MWCNT	Mixed bacteria	178.34 ± 4.79	3
Carbon paper	Au	Mixed bacteria	461.6	4
Stainless steel mesh	Activated carbon/ Fe ₃ O ₄	Mixed bacteria	809 ± 5	5
Carbon paper	Fe ₃ O ₄ /CNT	<i>Escherichia coli</i>	830	6
Carbon paper	Fe ₃ O ₄ /CNT	<i>Escherichia coli</i>	865	7
Carbon paper	Graphene/Fe ₃ O ₄	<i>Shewanella oneidensis</i>	891	8
Carbon paper	Au	Mixed bacteria	990	9
Stainless steel plates	Fe ₃ O ₄ /Fe ₂ O ₃	<i>Geobacter sulfurreducens</i>	1500	10
		Polydopamine-coated		
Carbon felt	/	<i>Shewanella xiamenensis</i>	452.8	11
		Polypyrrole-coated		
Carbon cloth	/	<i>Shewanella oneidensis</i>	1479	12
		Carbon dots-coated		
Carbon felt	/	<i>Shewanella oneidensis</i>	1697.9	13
		Au and Fe₃O₄-coated		
Carbon cloth	/	<i>Shewanella oneidensis</i>	1792	This work

References

1. B. Yu, Y. Li and L. Feng, *J. Hazard. Mater.*, 2019, **377**, 70-77.
2. S. Aslan, P. Ó Conghaile, D. Leech, L. Gorton, S. Timur and U. Anik, *ChemistrySelect*, 2017, **2**, 12034-12040.
3. X. Wu, X. Xiong, G. Owens, G. Brunetti, J. Zhou, X. Yong, X. Xie, L. Zhang, P. Wei and H. Jia, *Bioresour. Technol.*, 2018, **270**, 11-19.
4. F. A. a. Alatraktchi, Y. Zhang and I. Angelidaki, *Appl. Energy*, 2014, **116**, 216-222.
5. X. Peng, H. Yu, X. Wang, Q. Zhou, S. Zhang, L. Geng, J. Sun and Z. Cai, *Bioresour. Technol.*, 2012, **121**, 450-453.
6. I. H. Park, M. Christy, P. Kim and K. S. Nahm, *Biosens. Bioelectron.*, 2014, **58**, 75-80.
7. I. H. Park, P. Kim, G. Gnana kumar and K. S. Nahm, *Appl. Biochem. Biotechnol.*, 2016, **179**, 1170-1183.
8. R.-B. Song, C.-e. Zhao, P.-P. Gai, D. Guo, L.-P. Jiang, Q. Zhang, J.-R. Zhang and J.-J. Zhu, *Chem. Asian J.*, 2017, **12**, 308-313.
9. S. Mateo, P. Cañizares, M. A. Rodrigo and F. J. Fernandez-Morales, *Appl. Energy*, 2018, **225**, 52-59.
10. X. Long, X. Cao, C. Wang, S. Liu and X. Li, *J. Electroanal. Chem.*, 2019, **855**, 113497.
11. S.-R. Liu, L.-F. Cai, L.-Y. Wang, X.-F. Yi, Y.-J. Peng, N. He, X. Wu and Y.-P. Wang, *Chem. Commun.*, 2019, **55**, 10535-10538.
12. R.-B. Song, Y. Wu, Z.-Q. Lin, J. Xie, C. H. Tan, J. S. C. Loo, B. Cao, J.-R. Zhang, J.-J. Zhu and Q. Zhang, *Angew. Chem. Int. Ed.*, 2017, **56**, 10516-10520.
13. D. Guo, H.-F. Wei, R.-B. Song, J. Fu, X. Lu, R. Jelinek, Q. Min, J.-R. Zhang, Q. Zhang and J.-J. Zhu, *Nano Energy*, 2019, **63**, 103875.