Supplementary Materials

Iron-Doped Titanium Dioxide Hollow Nanospheres for Efficient Nitrogen Fixation and Zn-N₂ Aqueous Battery

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1.1 Characterization

Powder X-ray diffraction (PXRD) patterns were collected on a Bruker D8 Focus diffractometer with Cu- $K\alpha$ irradiation. The morphologies of products were investigated by scanning electron microscopy (SEM, JEOL JSF-7500 L, 5 kV) and transmission electron microscopy (TEM, JEOL JEM-2800, 200 kV). X-ray photoelectron spectroscopy (XPS) was performed on a Thermo Scientific ESCALAB 250Xi spectrometer using a monochromatic Al $K\alpha$ X-ray (1486.6 eV) as the exciting source.

1.2 Electrochemical measurement

Mild electrochemical nitrogen fixation experiment was performed in a H-type reaction cell on a WaveDrive 20 Bipotentiostat/Galvanostat (Pine Research Instrumentation, USA) using a threeelectrode system. The Pt, catalysts/carbon paper and Ag/AgCl rod (saturated KCl electrolyte) were used as the counter, working and reference electrodes, respectively. Calibrating all potentials according to the following equation:

$$E_{\rm RHE} = E_{\rm Ag/AgCl} + 0.059 \rm pH + E^{o}_{\rm Ag/AgCl}$$

Before electrochemical tests, the electrolyte (0.1 M Na₂SO₄) was saturated with N₂ for 0.5 h, where N₂ gas (\geq 99.999%) passed by the alkaline solution (pH=13 KOH aqueous solution) and acid solution (pH=1 H₂SO₄ aqueous solution) to remove any N contamination before purging into the electrolyte solution. To further ensure a pollution-free nitrogen reduction process, we pre-reduced catalysts for 3 h at -0.7 V vs. RHE under Ar to remove residual or adsorbed NO_x contaminants.^[1-3]

1.3 Determination of NH₃

1.3.1 Analysis of ammonia by the indophenol blue method^[4-6]:

Typically, 2 mL electrolyte was obtained from the cathodic reaction chamber and mixed with 2 mL NaOH solution (1 M) containing $C_7H_5NaO_3$ (5 wt%) and $Na_3C_6H_5O_7$ (5 wt%), 1 mL of NaClO (0.05 M) and 0.2 mL of $C_5FeN_6Na_2O$ (1 wt%) for 2 h. Absorbance measurements were performed at wavelength of 655 nm. The corresponding curve (y = 0.4595x + 0.0068, $R^2 = 0.999$) showed a good linear relation between absorbance value and NH₃ concentration through three times independent calibrations.

1.3.2 Analysis of ammonia by the Nessler's reagent method:

Typically, 5 mL electrolyte was taken from the reaction cell and added into 1 mL potassium sodium tartrate (KNaC₄H₄O₆, 0.2 M) solution and 1 mL of the Nessler's reagent. Hereafter, the solution was kept to stand for 25 min for full color processing. Finally, NH₃ concentration was detected using an UV-vis spectrophotometer at 420 nm wavelength. The fitting curve (y = 0.09x + 0.032, R² = 0.999) shows good linear relation of absorbance value with NH₃ concentration.

1.3.3 Analysis of ammonia by nuclear magnetic resonance (NMR) with ¹⁵N labelling^[4-6]:

The NRR measurements were conducted using either ${}^{15}N_2$ (99% isotopic purity, supplied by Beijing Yinuokai Technology Co., Ltd.) forisotopic labelling or normal ${}^{14}N_2$ for comparison. Each NRR test was performed at -0.7 V (vs. RHE) for 2.5 h. Then, 0.5 mL of above solution mixed with 0.05 mL of d6-DMSO was used for ¹H nuclear magnetic resonance (NMR) spectroscopy examination (Bruker AVANCE III 600 MHz). The corresponding absolute calibration curves were obtained by using the ${}^{15}NH_4Cl$ (99% isotopic purity, Shanghai Xinbo Chemical Technology Co., Ltd.) and ${}^{14}NH_4Cl$ aqueous solutions with a series of known concentrations as the standards.

1.4 Determination of N₂H₄^[4-6]:

Hydrazine quantification was obtained via the Watt and Chrisp method.^[4-6] Typically, 5 mL electrolyte was obtained from reaction cell and mixed with 5 mL color reagent (mixture of 5.99g p-C₉H₁₁NO, 30 mL HCl and 300 mL C₂H₅OH) for 10 min. Absorbance measurements were performed at wavelength of 455 nm. The fitting curve (y = 1.1317x + 0.0199, $R^2 = 0.9997$) exhibited a good linear relationship between the absorbance value and N₂H₄ concentration through three times independent calibrations.

1.5 Faradaic efficiency:

Ammonia formation rate was calculated using the following equation:

$$v_{\rm NH3} = \frac{V \times C_{\rm NH_3}}{17 \times A \times t}$$

FE was calculated according to the following equation:

$$FE = \frac{3F \times n_{NH_3}}{Q}$$

Where V is the volume of the electrolyte, C_{NH3} means NH₃ concentration, t is the reduction reaction time, A is the geometric area of the cathode, F is the Faraday constant, and Q is the quantity of applied electricity.

1.6 Zn-N₂ battery tests:

The Zn-N₂ battery performance tests were carried out on a homemade Zn-N₂ cell filled with of 0.1 M KOH, where a polished Zn plate was used as anode and carbonfiber paper supported catalyst as N₂ cathode. The discharge curve of Zn-N₂ cell was recorded on a Zahner IM6eX electrochemical workstation.

1.7 In situ FTIR measurement:

The electrochemical *in situ* Fourier transform infrared (*in situ* FTIR) spectroscopy was collected using a Nexus 870 spectrometer (Nicolet) equipped with a liquid-nitrogen-cooled MCT-A detector and an EverGlo IR source. *In situ* FTIR spectra were recorded by means of time-resolved FTIR spectroscopy procedures. The resulting spectra were reported as the relative change in reflectivity and calculated as follows:

$$\frac{\Delta R}{R} = \frac{R(E_s) - R(E_R)}{R(E_R)}$$

where $R(E_S)$ and $R(E_R)$ are the accumulated spectra collected at sample potential E_S and reference potential E_R , respectively.



Figure S1. XPS spectra of Fe1.0HTNs and HTNs.



Figure S2. Representative photograph of the used H-type reaction cell for the NRR.



Figure S3. (a) UV-Vis absorption curves of indophenol assays with NH_3 after incubated for 2 h at room temperature. (b) The corresponding calibration curve used for calculation of NH_3 concentrations. All experiments were performed in 0.1 M Na₂SO₄.



Figure S4. (a) UV-Vis absorption curves of various N_2H_4 concentrations after incubated for 20 min at room temperature. (b) The corresponding calibration curve used for estimation of N_2H_4 concentrations. All experiments were performed in 0.1 M Na₂SO₄.



Figure S5. Flow diagram of control experiments to rigorously conduct NRR experiments in this work.



Figure S6. UV-visible absorption spectra of the 0.1 M Na_2SO_4 electrolytes stained with indophenol indicator after charging on the $Fe_{1.0}HTNs$ electrode at open-circuit potential for 2.5 h.



Figure S7. UV-visible absorption spectra of the Ar-bubbled 0.1 M Na_2SO_4 electrolytes stained with indophenol indicator after charging on the $Fe_{1.0}HTNs$ electrode at at -0.7 V vs. RHE for 2.5 h.



Figure S8. (a) UV-Vis absorption curves of Nessler's reagent assays kept with various concentrations of NH_3 . (b) A calibration curve for estimating the concentrations of NH_3 .



Figure S9. (a) ¹H NMR spectra of ¹⁴NH₄⁺ standard samples with different concentrations. (b) The corresponding calibration curve of ¹⁴NH₄⁺ concentration vs. peak area intensity of NMR spectra.



Figure S10. (a) ¹H NMR spectra of ¹⁵NH₄⁺ standard samples with different concentrations. (b) The corresponding calibration curve of $^{15}NH_4^+$ concentration vs. peak area intensity of NMR spectra.



Figure S11. Chronoamperometry curve of $Fe_{1.0}HTNs$ in N₂-bubbled 0.1 M Na₂SO₄ at a fixed overpotential of -0.7 V for 24 h.



Figure S12. XRD patterns of $Fe_{1.0}$ HTNs before and after long-term NRR durability test in 0.1 M Na₂SO₄.



Figure S13. TEM image of Fe_{1.0}HTNs after long-time NRR test in 0.1 M Na₂SO₄.



Figure S14. (a) UV-Vis absorption curves and (b) the corresponding calibration curve of indophenol assays with NH_3 after incubated for 2 h at room temperature in 0.1 M KOH.



Figure S15. (a) UV-Vis absorption curves and (b) the corresponding calibration curve of various N_2H_4 concentration after incubated for 30 min at room temperature in 0.1 M KOH.



Figure S16. (a) UV-Vis absorption curves and (b) the corresponding calibration curve of indophenol assays with NH_3 after incubated for 2 h at room temperature in 0.1 M HCl.



Figure S17. (a) UV-Vis absorption curves and (b) the corresponding calibration curve of various N_2H_4 concentration after incubated for 30 min at room temperature in 0.1 M HCl.



Figure S18. UV-Vis absorption spectra of the electrolytes (0.1 M KOH) estimated by the method of Watt and Chrisp after NRR electrolysis at different potentials.



Figure S19. UV-Vis absorption spectra of the electrolytes (0.1 M HCl) estimated by the method of Watt and Chrisp after NRR electrolysis at different potentials.



Figure S20. Chronoamperometry results of Fe_{1.0}HTNs at each given potential in 0.1 M KOH.



Figure S21. Chronoamperometry results of Fe_{1.0}HTNs at each given potential in 0.1 M HCl.



Figure S22. Chronoamperometry curve of $Fe_{1.0}HTNs$ in N₂-bubbled 0.1 M KOH for 10 h.



Figure S23. Chronoamperometry curve of Fe_{1.0}HTNs in N₂-bubbled 0.1 M HCl for 10 h.



Figure S24. (a) UV-Vis absorption spectra of electrolytes stained with indophenol indicator under different potentials. (b) NH_3 yield rates and FEs for $Fe_{1.0}HTNs$ under different potentials in N_2 -bubbled 0.1 M KOH.



Figure S25. (a) UV-Vis absorption spectra of electrolytes stained with indophenol indicator under different potentials. (b) NH₃ yield rates and FEs for $Fe_{1.0}HTNs$ under different potentials in N₂-bubbled 0.1 M HCl.



Figure S26. LSV curves of samples for HER obtained in Ar-saturated 0.1 M Na₂SO₄ electrolyte.



Figure S27. Nyquist plots of the samples on carbon paper in 0.1 M Na₂SO₄.



Figure S28. Nyquist plots of the samples on carbon paper in 0.1 M Na₂SO₄.



Figure S29. Cyclic voltammograms of (a) $Fe_{1.0}HTNs$, (b) HTNs and (c) SFeTNs coated on carbon paper and measured at different scan rates from 20 to 100 mV s⁻¹. (d) The corresponding plots of current density at -0.14 V versus the scan rate.



Figure S30. The current density of $Fe_{1.0}$ HTNs under different N_2 flow rates: (a) 5 mL/min, (b) 10 mL/min, (c) 20 mL/min, (d) 40 mL/min and (e) 60 mL/min.



Figure S31. The current density of SFeTNs under different N_2 flow rates: (a) 5 mL/min, (b) 10 mL/min, (c) 20 mL/min, (d) 40 mL/min and (e) 60 mL/min.



Figure S32. Schematic illustration of solid Fe-TiO₂@C precursor, Fe_{1.0}HTNs, and crushed Fe_{1.0}HTNs-*t* samples (*t* represents the crushed time). (b-f) The corresponding TEM images of solid Fe-TiO₂@C, Fe_{1.0}HTNs, Fe_{1.0}HTNs-2min, Fe_{1.0}HTNs-5min, and Fe_{1.0}HTNs-20min.

The solid Fe-TiO₂@C sphere precursor with a size of ~100 nm (**Figure S32b**) was carbonized in air to form $Fe_{1.0}$ HTNs hollow nanospheres (**Figure 32c**). Then, $Fe_{1.0}$ HTNs hollow nanospheres were further crushed mechanically by a static pressure of ~32 MPa with the increase over time from 2 min to 20 min (**Figure 32d-f**). The integrity of the spherical shell was gradually destroyed. Herein, the synthesis method used was based on the previous literatures with a minor modification. [*J. Am. Ceram. Soc.*, 2014, 97.2: 407-412; *J. Alloy. Compd.* 2018, 769, 521-531; *J. Mol. Catal. A*: *Chem.* 2005, 226, 93-100; *J. Am. Chem. Soc.* 2007, 129, 8406-8407].



Figure S33. N₂ adsorption-desorption isotherm curves for various samples.



Figure S34. Representative picture of *in situ*-FTIR spectroscopy testing device with Nicolet iS50 spectrometer equipped with VeeMAX III variable angle specular reflectance accessory and an electrochemical reaction cell with a CaF₂ bottom.



Figure S35. The photograph of $Zn-N_2$ battery.

Catalyst	Electrolyte	$\rm NH_3$ yield (µg h ⁻¹ mg ⁻¹ _{cat.})	FE (%)	Ref.
Fe _{1.0} HTNs	0.1 M Na ₂ SO ₄	43.14	16.35	This work.
Fe _{1.0} HTNs	0.1 M KOH	15.2	9.1	This work.
Fe _{1.0} HTNs	0.1 M HCl	12.1	7.9	This work.
Ag ₃ Cu BPNs	0.1 M Na ₂ SO ₄	24.59	13.28	7
PdCuIr-LS	0.1 M Na ₂ SO ₄	13.43	5.29	8
V ₂ O ₃ /C	0.1 M Na ₂ SO ₄	12.3	7.28	9
VO ₂ /CP	0.1 M Na ₂ SO ₄	14.85	3.97	10
TiO ₂ -rGO	0.1 M Na ₂ SO ₄	15.13	3.3	11
Mn ₃ O ₄ nanocubes	0.1 M Na ₂ SO ₄	11.6	3.0	12
Cr ₂ O ₃ microspheres	0.1 M Na ₂ SO ₄	25.3	6.78	13
CoO	0.1 M Na ₂ SO ₄	21.5	8.3	14
VO ₂ hollow microspheres	0.1 M Na ₂ SO ₄	14.85	3.97	15
CeO ₂ nanorod	0.1 M Na ₂ SO ₄	16.4	3.7	16
NiO/G	0.1 M Na ₂ SO ₄	18.6	7.8	17
La ₂ O ₃	0.1 M Na ₂ SO ₄	17.04	4.76	18
MoS ₂ nanoflower	0.1 M Na ₂ SO ₄	29.28	8.34	19
S-CNS	0.1 M Na ₂ SO ₄	19.07	7.47	20
d-FG	0.1 M Na ₂ SO ₄	9.3	4.2	21
BNS	0.1 M Na ₂ SO ₄	13.22	4.04	22
MBN	0.1 M Na ₂ SO ₄	18.2	5.5	23

Table S1. Comparison of NRR activity of some recently reported electrocatalysts in 0.1 M Na₂SO₄.

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