

Electronic Supporting Information

Multivalent cationic dendrofullerenes for gene transfer: synthesis and DNA complexation

Beatriz M. Illescas,^{*[a]} Alfonso Pérez-Sánchez,^[a] Araceli Mallo,^[b] Ángel Martín-Domenech,^[a] Ignacio Rodríguez-Crespo,^{*[c]} and Nazario Martín^{*[a,d]}

^[a] Departamento de Química Orgánica, Facultad de Química, Universidad Complutense de Madrid, Av. Complutense s/n, 28040 Madrid (Spain), Email: beti@ucm.es and nazmar@ucm.es

^[b] Facultad de Ciencias Experimentales, Universidad Francisco de Vitoria, Pozuelo de Alarcón, Madrid 28223, Spain

^[c] Departamento de Bioquímica, Facultad de Química, Universidad Complutense de Madrid, Av. Complutense s/n, 28040 Madrid (Spain), Email: jirodrig@ucm.es

^[d] IMDEA-Nanoscience, C/ Faraday 9, Campus Cantoblanco, 28049 Madrid, Spain.

Table of Contents	S1
General	S2
¹ H NMR (top) and ¹³ C NMR (bottom) spectra of amino compounds	S3
¹ H NMR (top) and FTIR (bottom) spectra of cationic compounds	S13
Determination of the N/P ratios	S17
Cytotoxicity assay	S17
DLS Experiments	S18

General. Reagents and solvents were purchased as reagent grade and used without further purification. 1-(chloromethyl)-3,5-bis(prop-2-yn-1-yloxy)benzene (**1a**),¹ 5-

(chloromethyl)-1,2,3-tris(prop-2-yn-1-yloxy)benzene (**1b**)² and *tert*-butyl (2-(2-(2-azidoethoxy)ethoxy)ethyl)carbamate (**2**)³ were prepared according to previously reported procedures. For column chromatography, silica gel 60 (230-400 mesh, 0.040-0.063 mm) was purchased from Scharlab. Thin Layer Chromatography (TLC) was performed on aluminium sheets coated with silica gel 60 F₂₅₄ purchased from E. Merck, visualization by UV light. IR spectra (cm⁻¹) were measured on a Bruker Tensor 27 instrument equipped with an ATR device FTIR instrument and using KBr for water-soluble compounds or dissolved in the proper solvent when possible. NMR spectra were recorded on a Bruker AC 300, AC 500 or AC 700 with solvent peaks as reference. ¹H and ¹³C NMR spectra were obtained for solutions in CDCl₃ and DMSO-d₆. All the assignments were confirmed by two-dimensional NMR experiments.

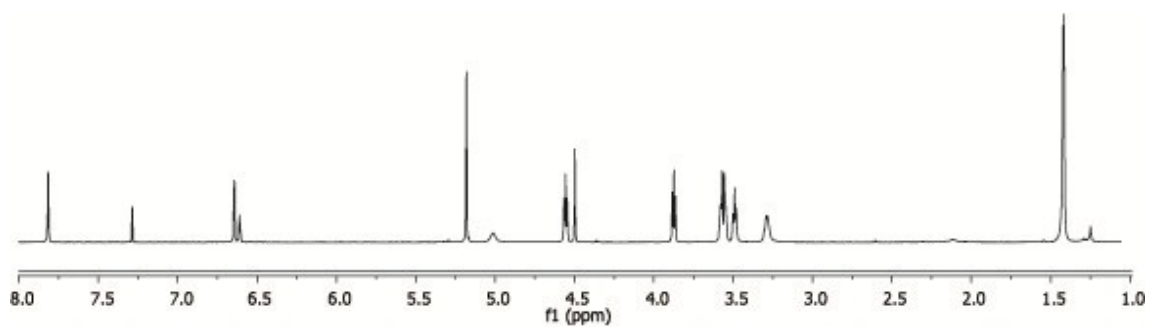
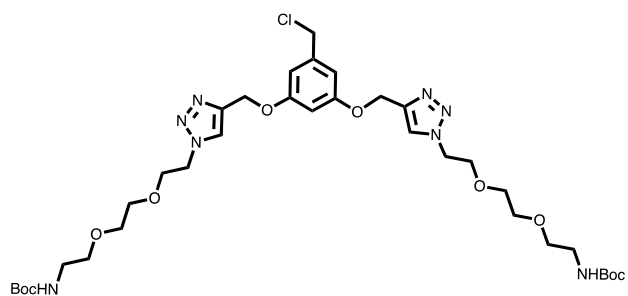
¹H NMR (top) and ¹³C NMR (bottom) spectra of amino compounds.

Compound 3a.

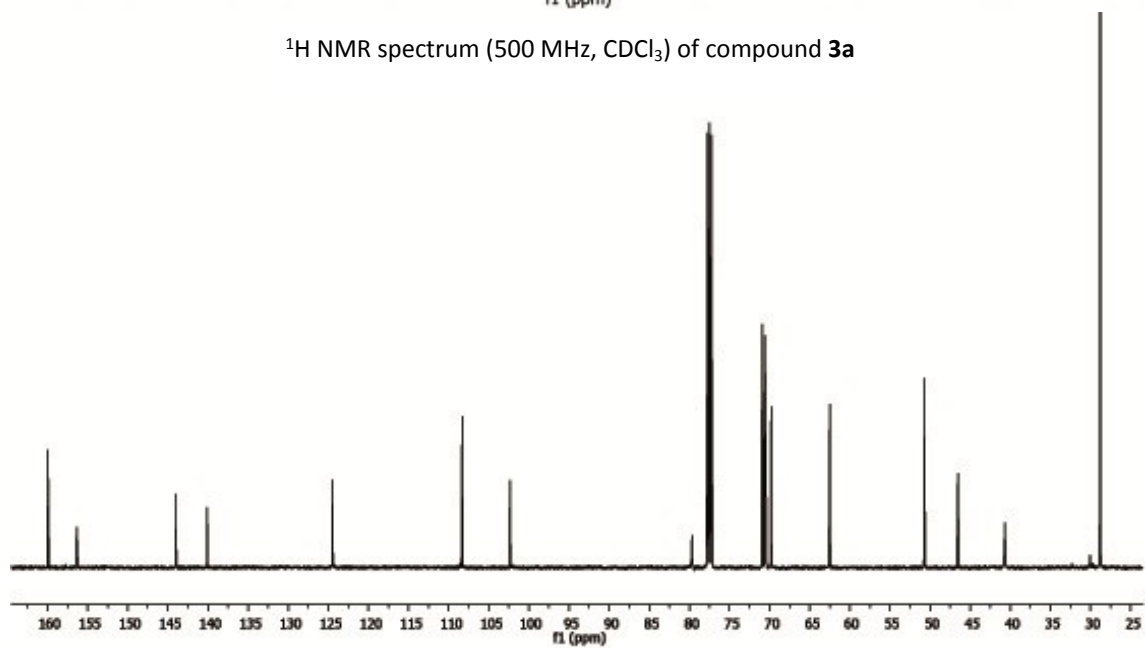
¹ Wu, P.; Feldman, A. K.; Nugent, A. K.; Hawker, C. J.; Scheel, A.; Voit, B.; Pyun, J.; Fréchet, J. M. J.; Sharpless, K. B.; Fokin, V. V. *Angew. Chem. Int. Ed.* **2004**, *43*, 3928-3932.

² Qin, T.; Li, X.; Chen, J.; Zeng, Y.; Yu, T.; Yang, G.; Li, Y. *Chem. Asian J.* **2014**, *9*, 3641-3649.

³ Itoh, Y.; Ishikawa, M.; Kitaguchi, R.; Sato, S.; Naito, M.; Hashimoto, Y. *Bioorg. Med. Chem.* **2011**, *19*, 3229-3241.

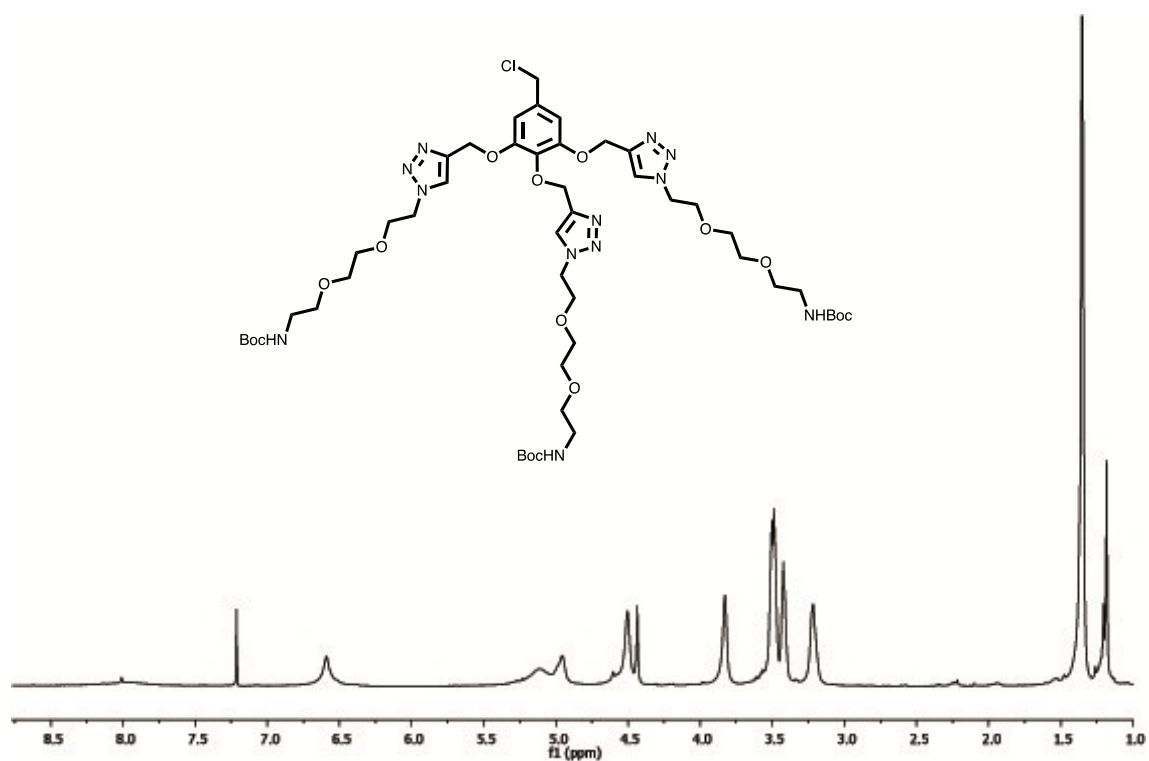


¹H NMR spectrum (500 MHz, CDCl₃) of compound **3a**

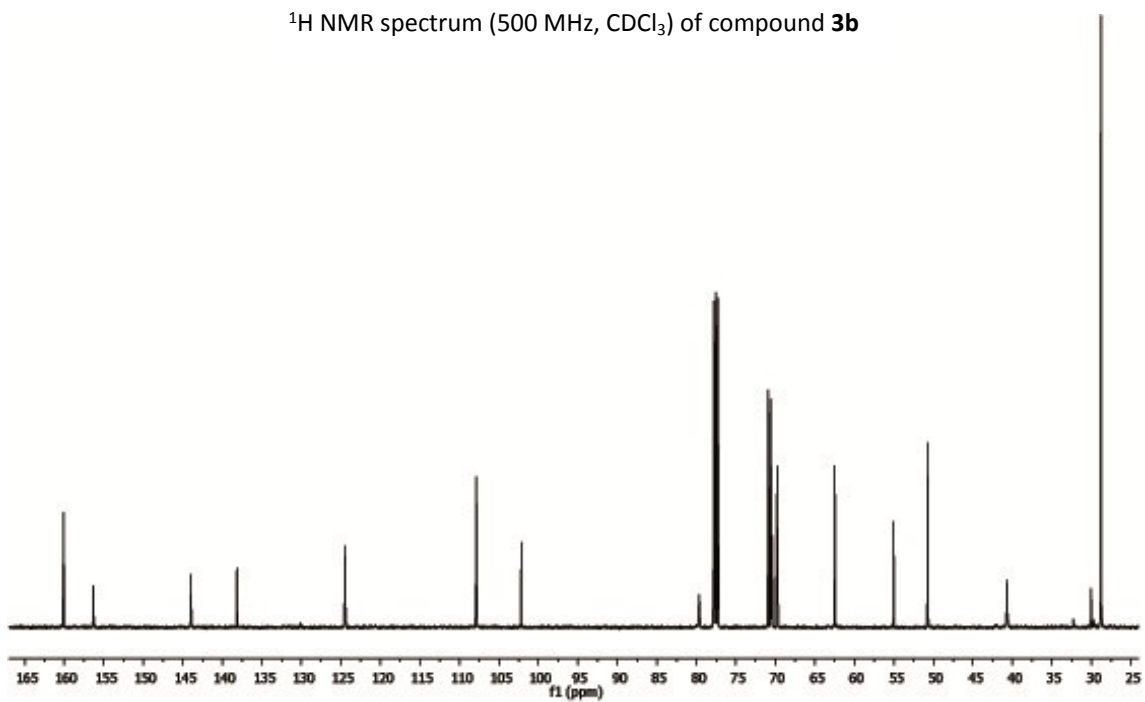


¹³C NMR spectrum (125 MHz, CDCl₃) of compound **3a**

Compound 3b.

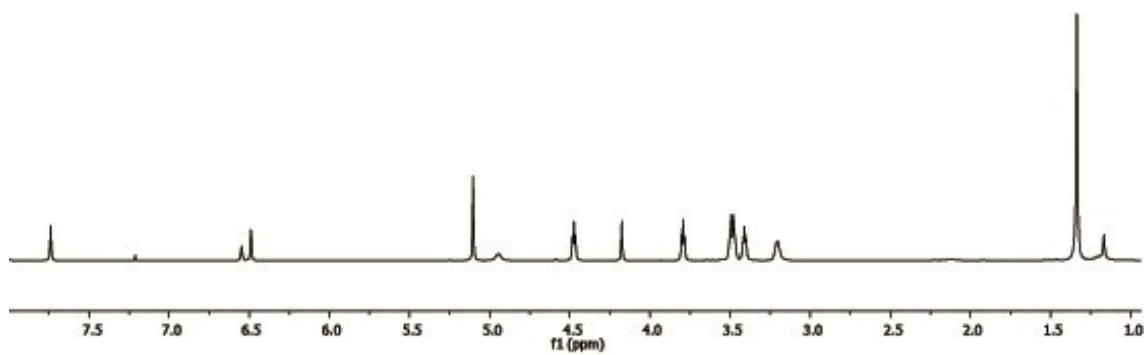
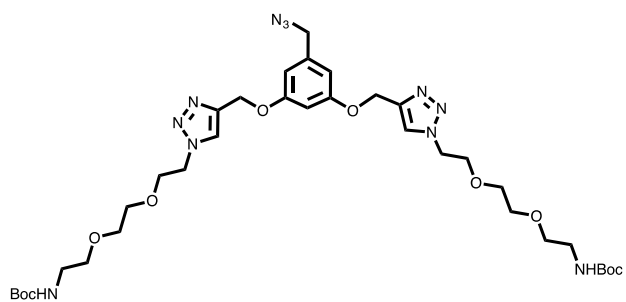


¹H NMR spectrum (500 MHz, CDCl₃) of compound 3b

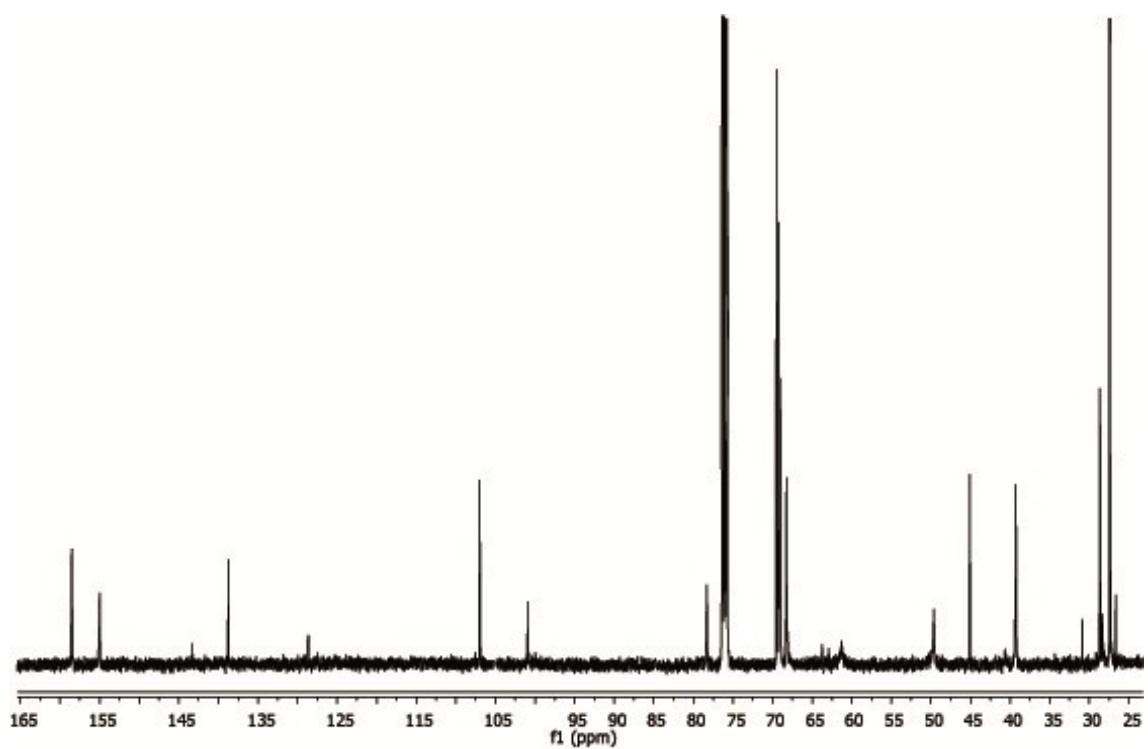


¹³C NMR spectrum (125 MHz, CDCl₃) of compound 3b

Compound 4a.

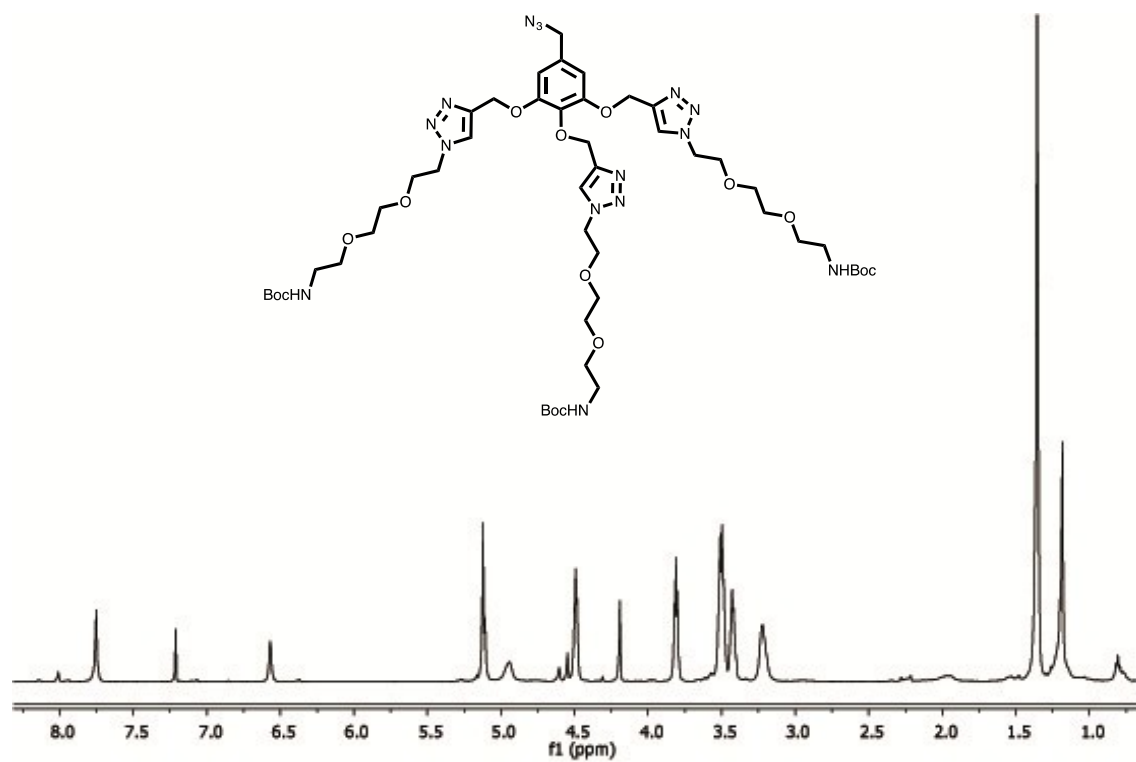


^1H NMR spectrum (500 MHz, CDCl_3) of compound 4a

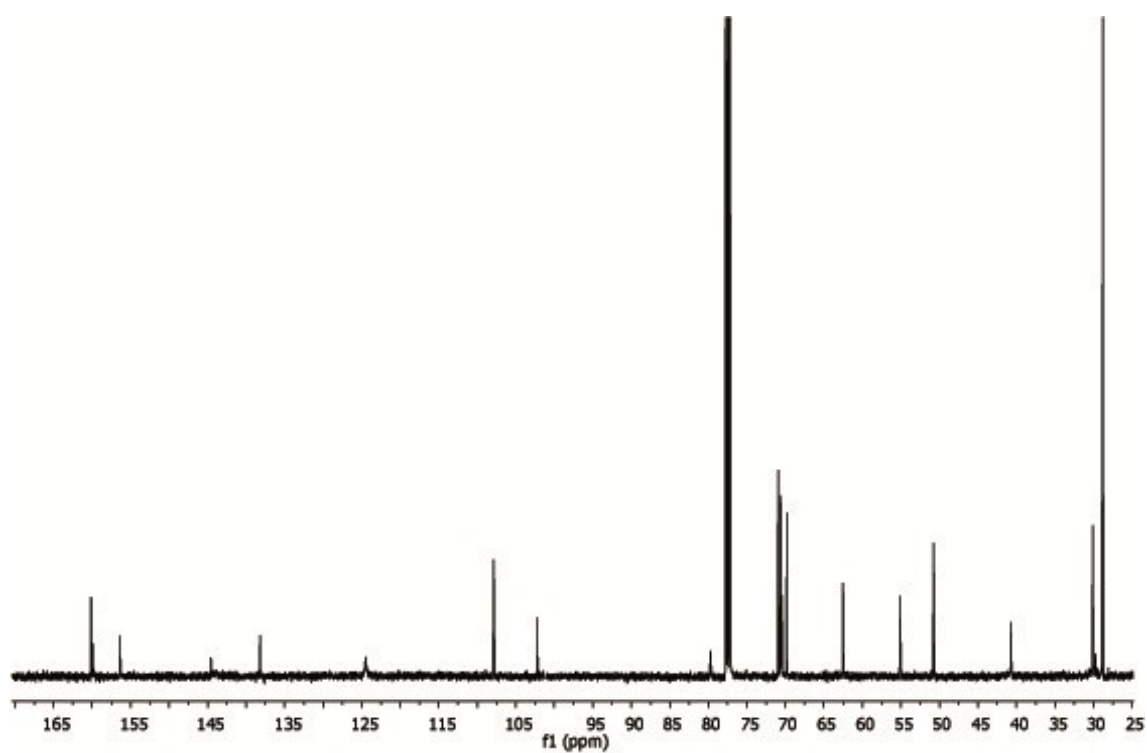


^{13}C NMR spectrum (125 MHz, CDCl_3) of compound 4a

Compound 4b.

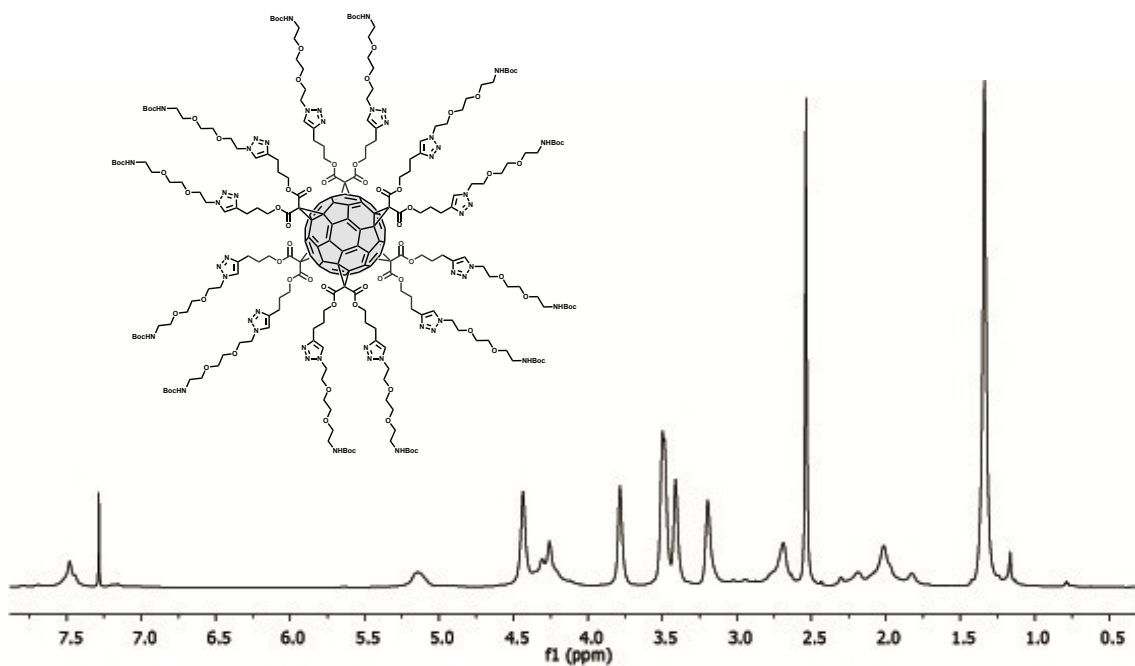


¹H NMR spectrum (500 MHz, CDCl₃) of compound **4b**

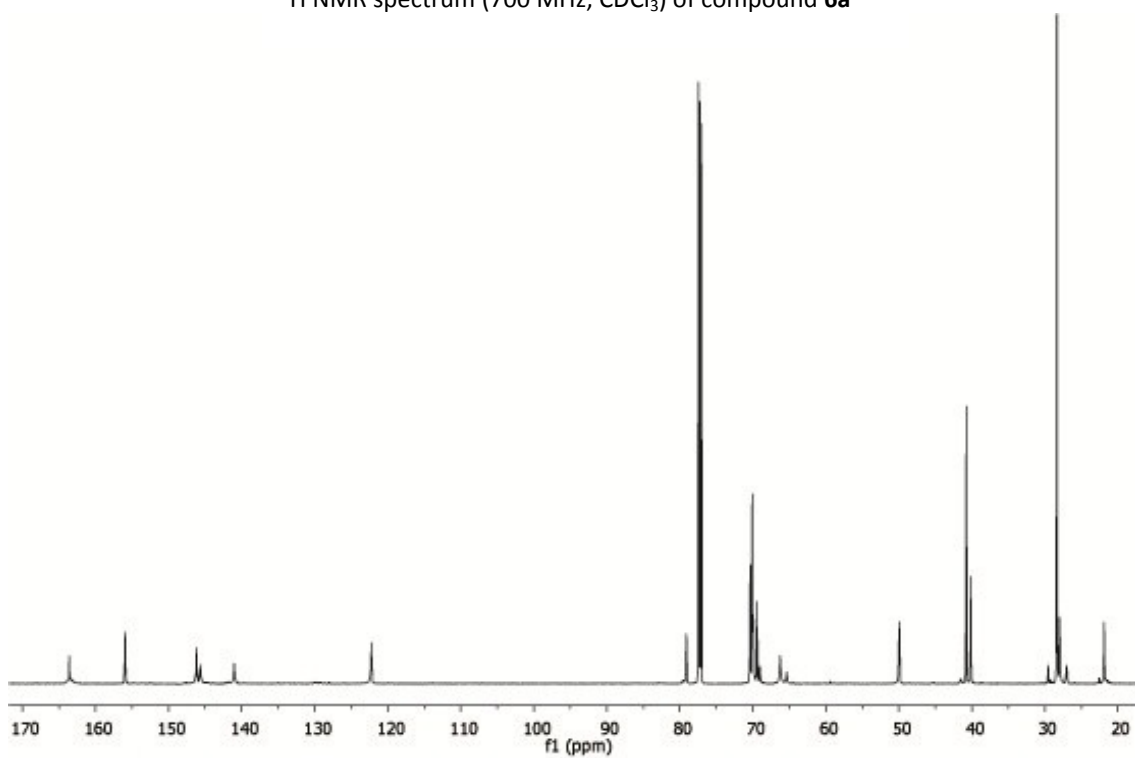


¹³C NMR spectrum (125 MHz, CDCl₃) of compound **4b**

Compound 6a.

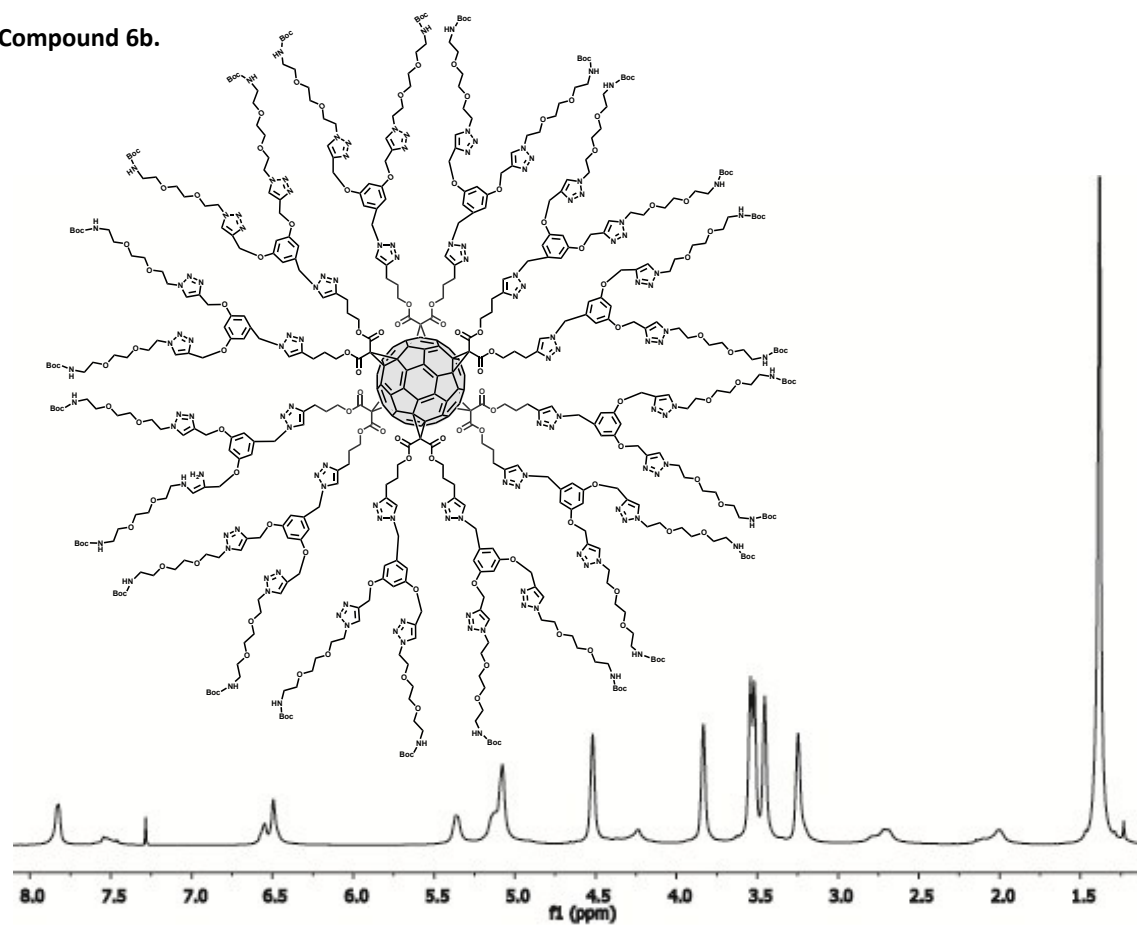


¹H NMR spectrum (700 MHz, CDCl₃) of compound 6a

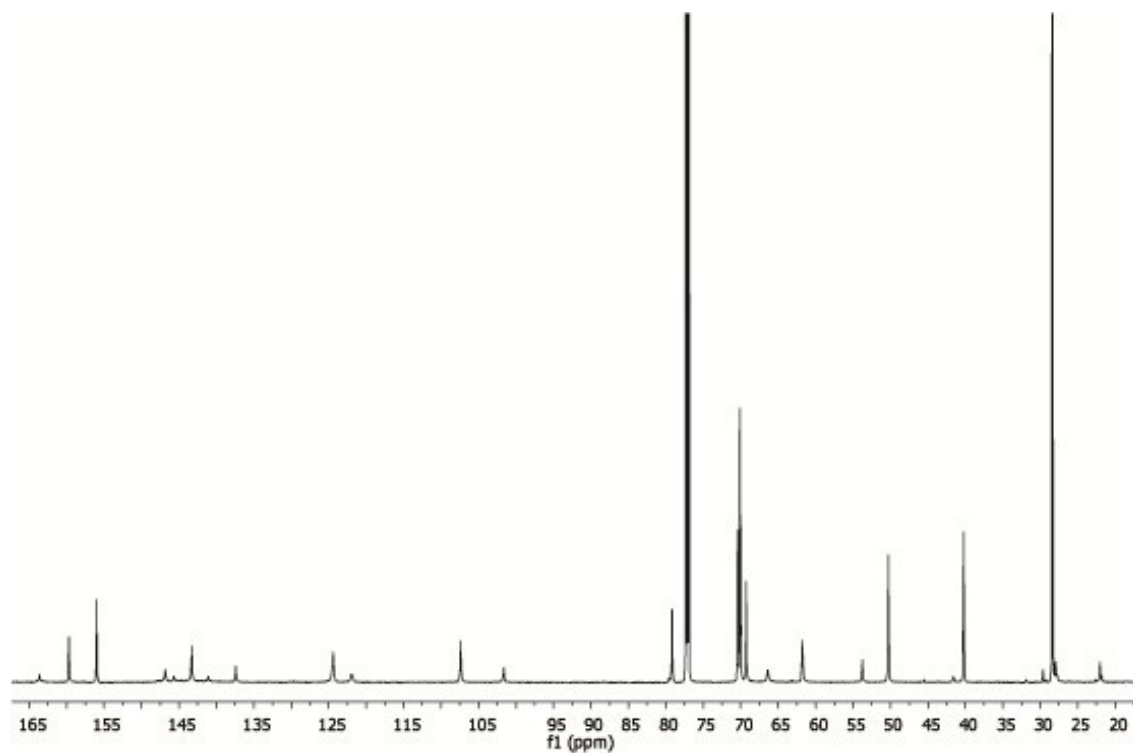


¹³C NMR spectrum (175 MHz, CDCl₃) of compound 6a

Compound 6b.

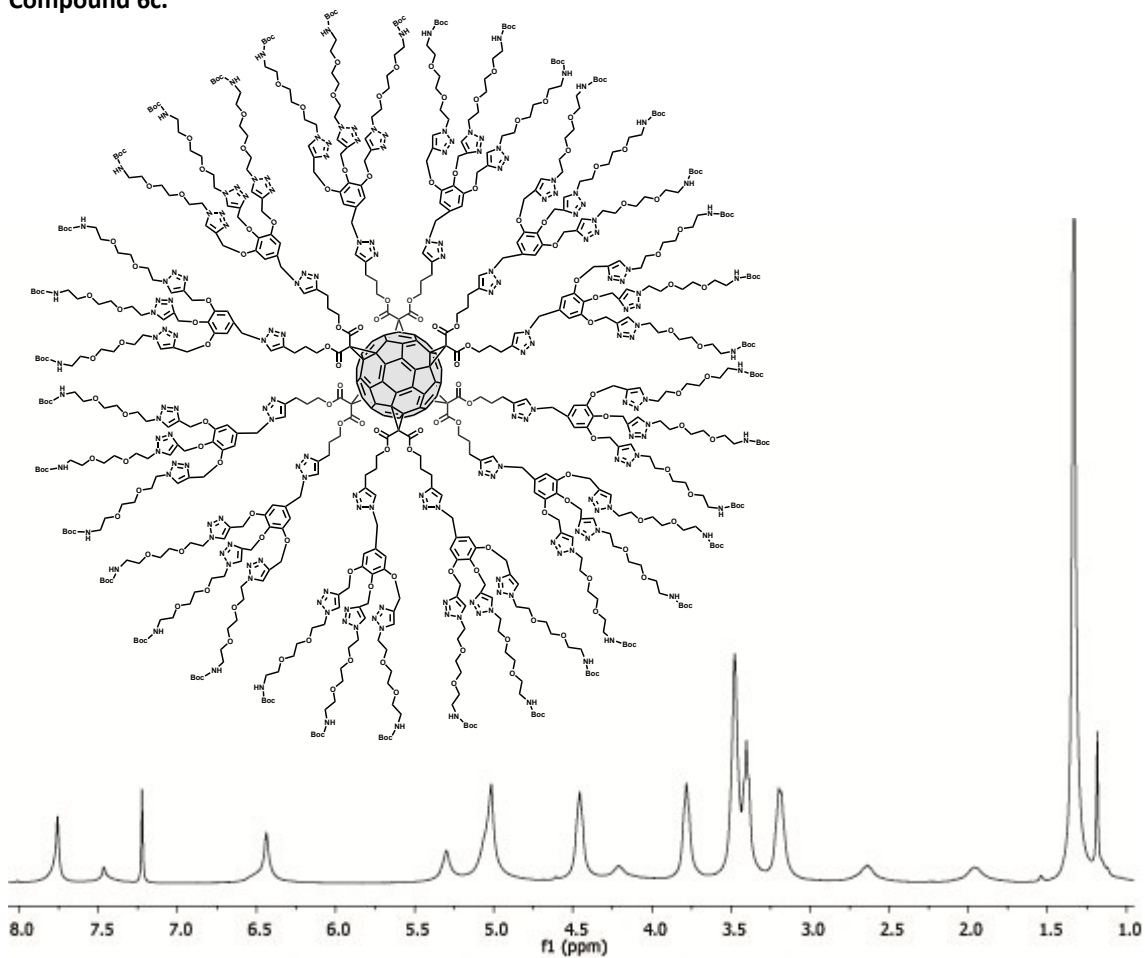


¹H NMR spectrum (700 MHz, CDCl₃) of compound 6b

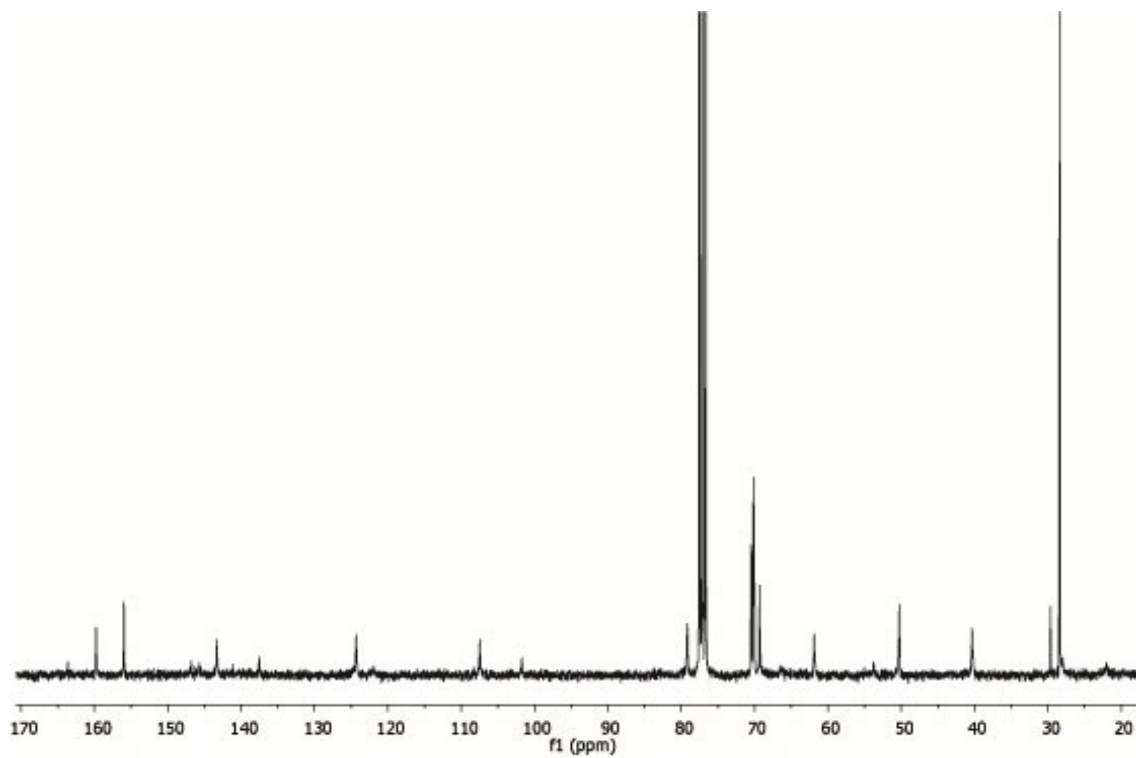


¹³C NMR spectrum (175 MHz, CDCl₃) of compound 6b

Compound 6c.

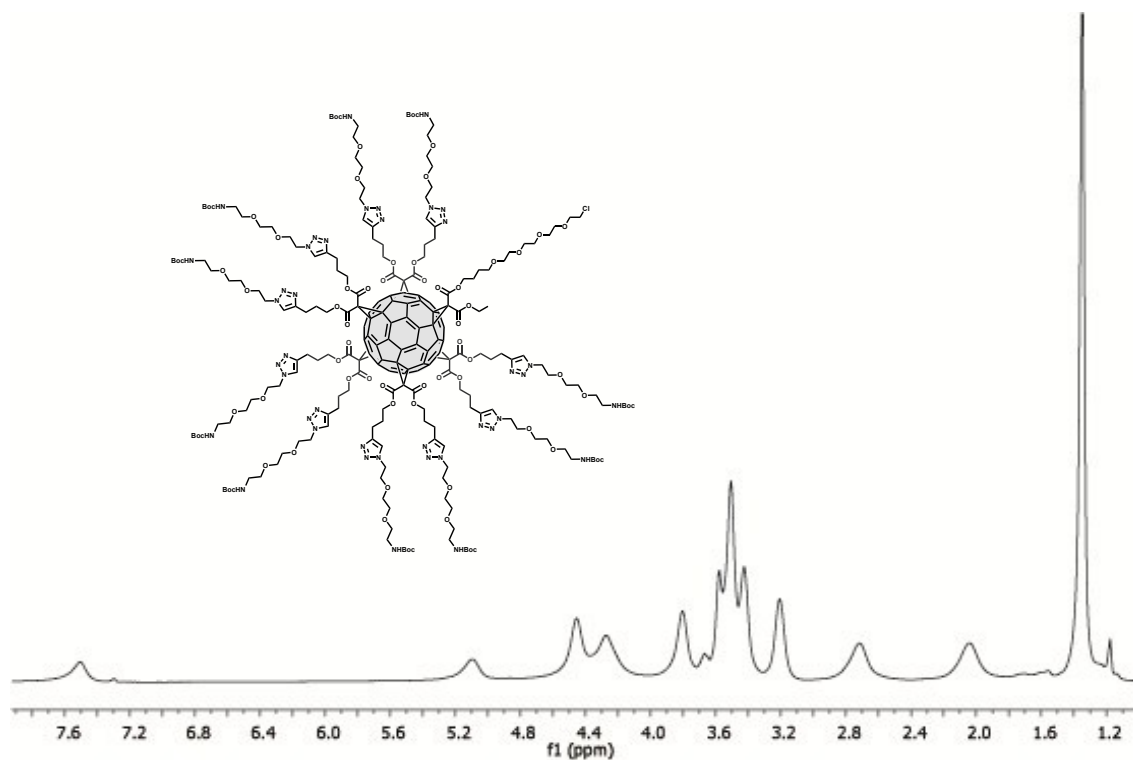


¹H NMR spectrum (700 MHz, CDCl₃) of compound 6c

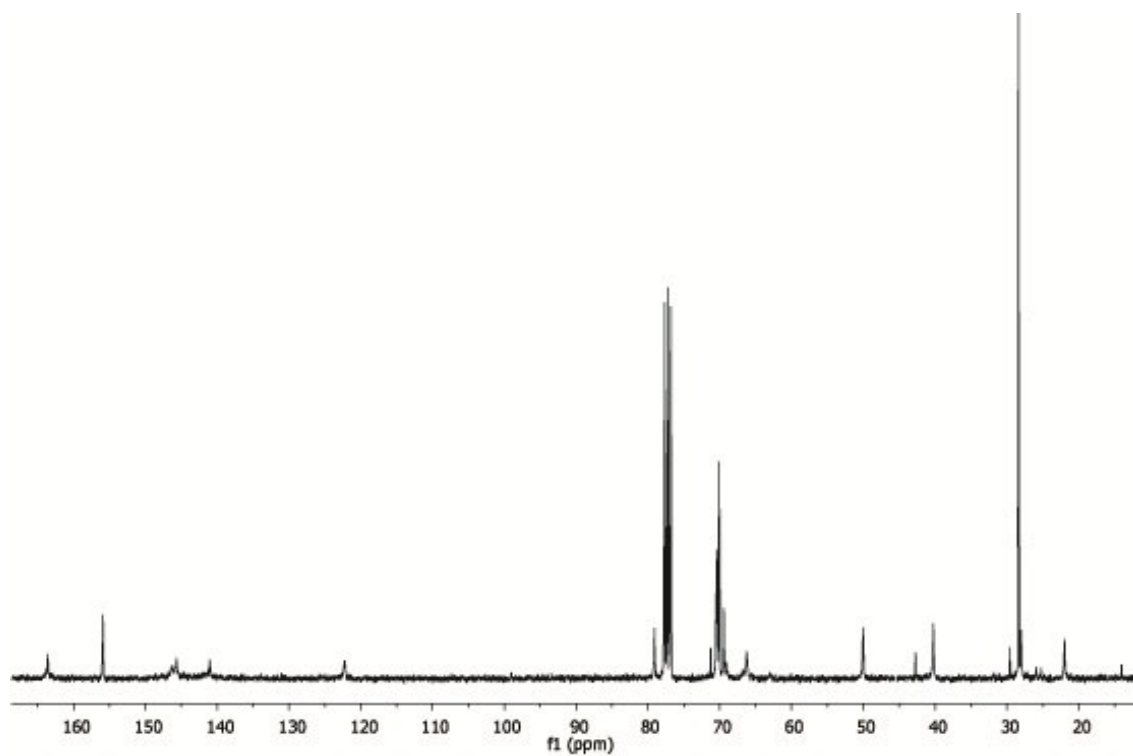


¹³C NMR spectrum (175 MHz, CDCl₃) of compound 6c

Compound 9.

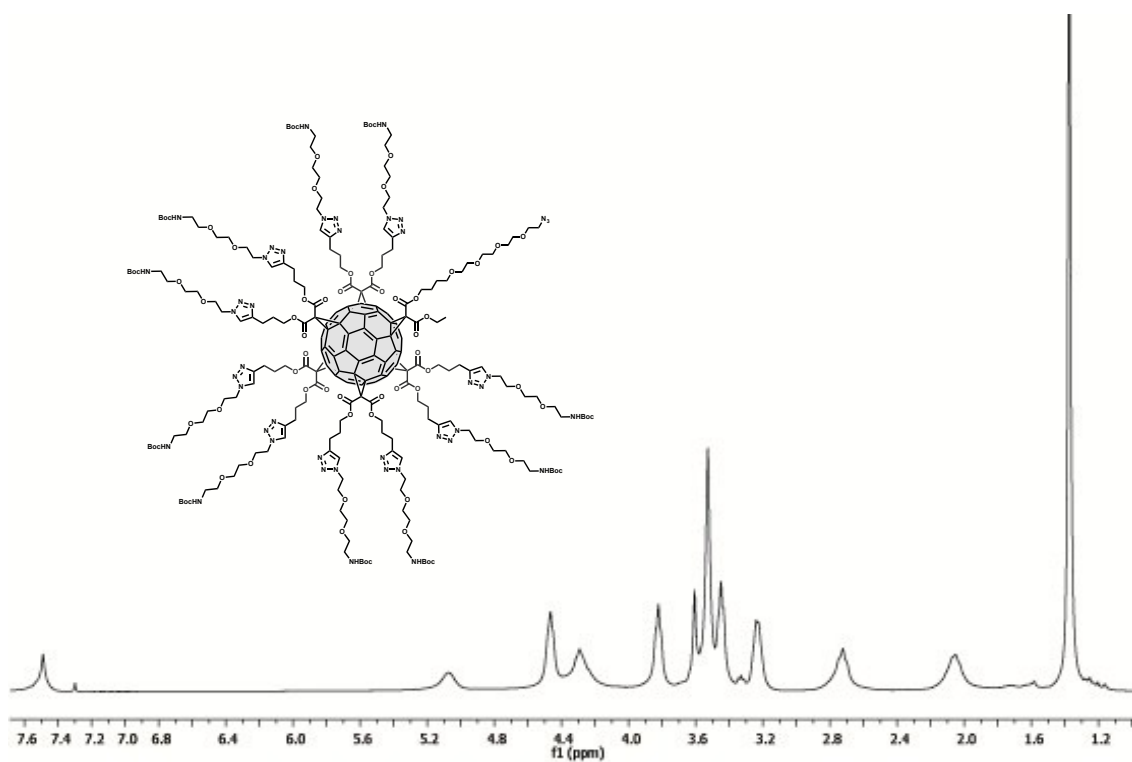


¹H NMR spectrum (700 MHz, CDCl₃) of compound 9

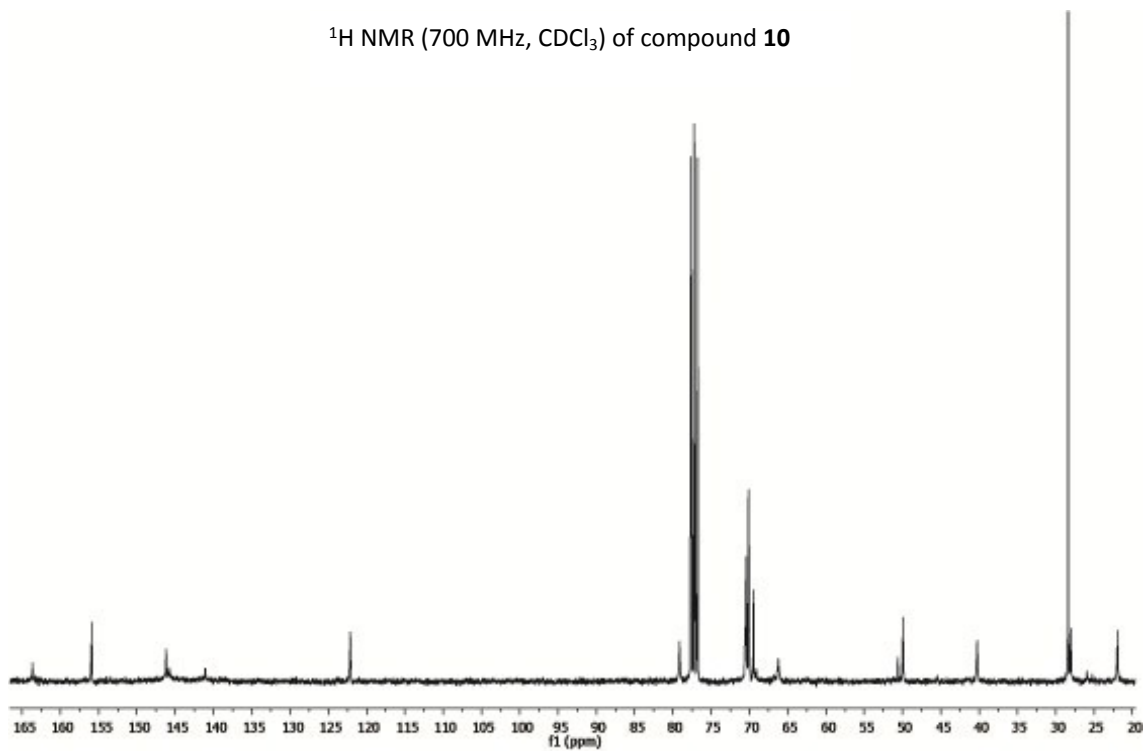


¹³C NMR spectrum (175 MHz, CDCl₃) of compound 9

Compound 10.



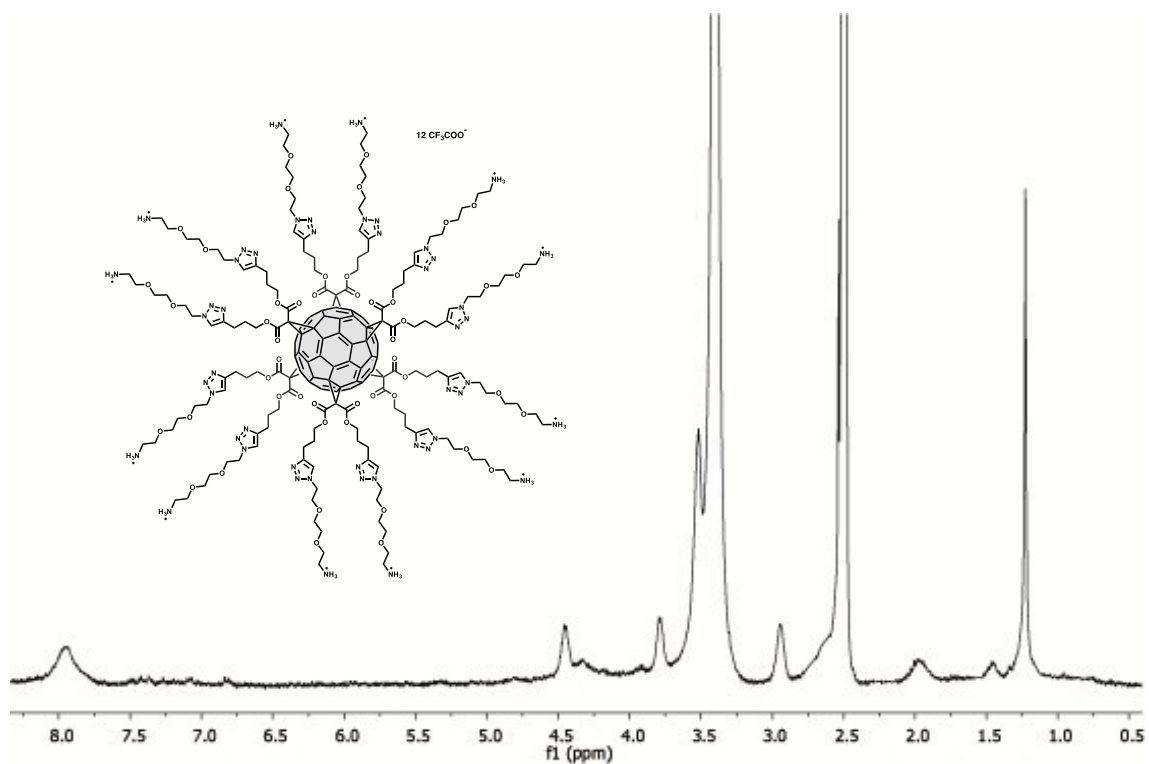
¹H NMR (700 MHz, CDCl₃) of compound 10



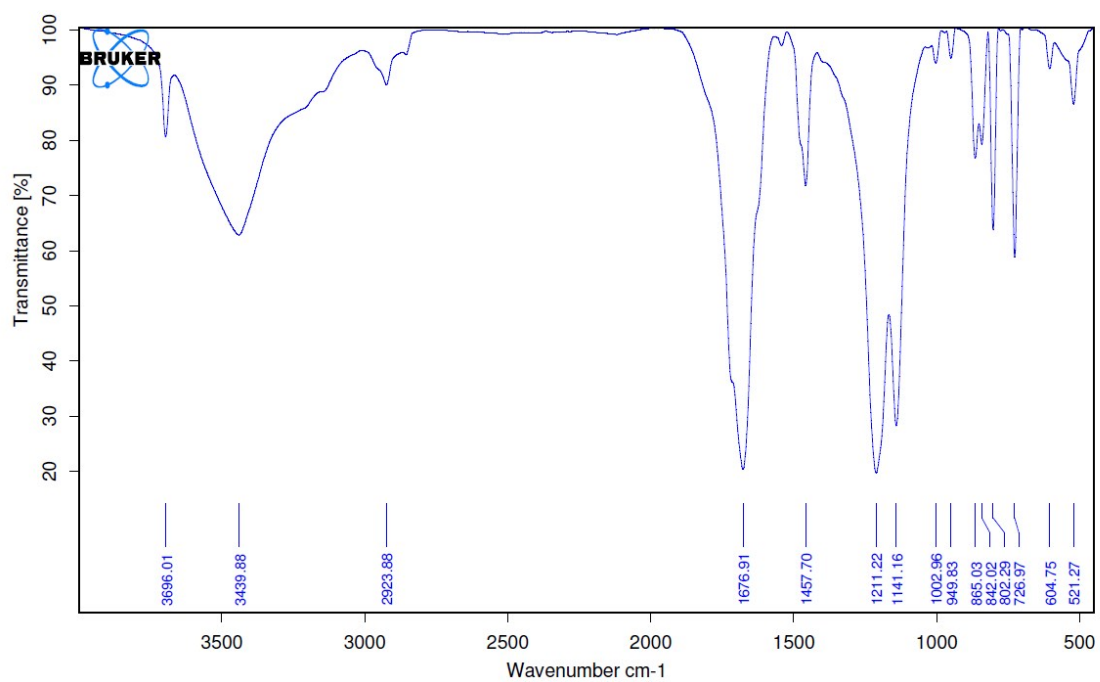
¹³C NMR (175 MHz, CDCl₃) of compound 10

¹H NMR (top) and FTIR (bottom) spectra of compounds 7a-c and 12.

Compound 7a.

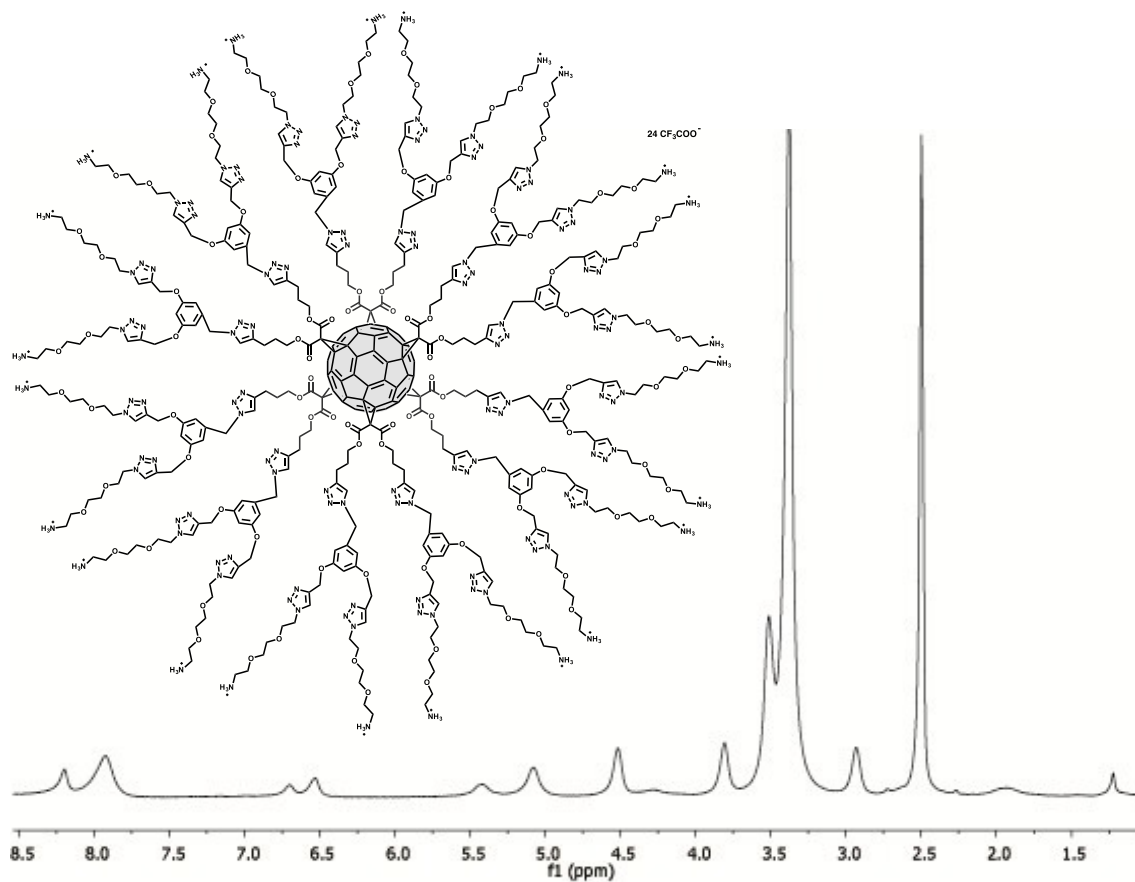


¹H NMR (300 MHz, DMSO-*d*⁶) of compound 7a

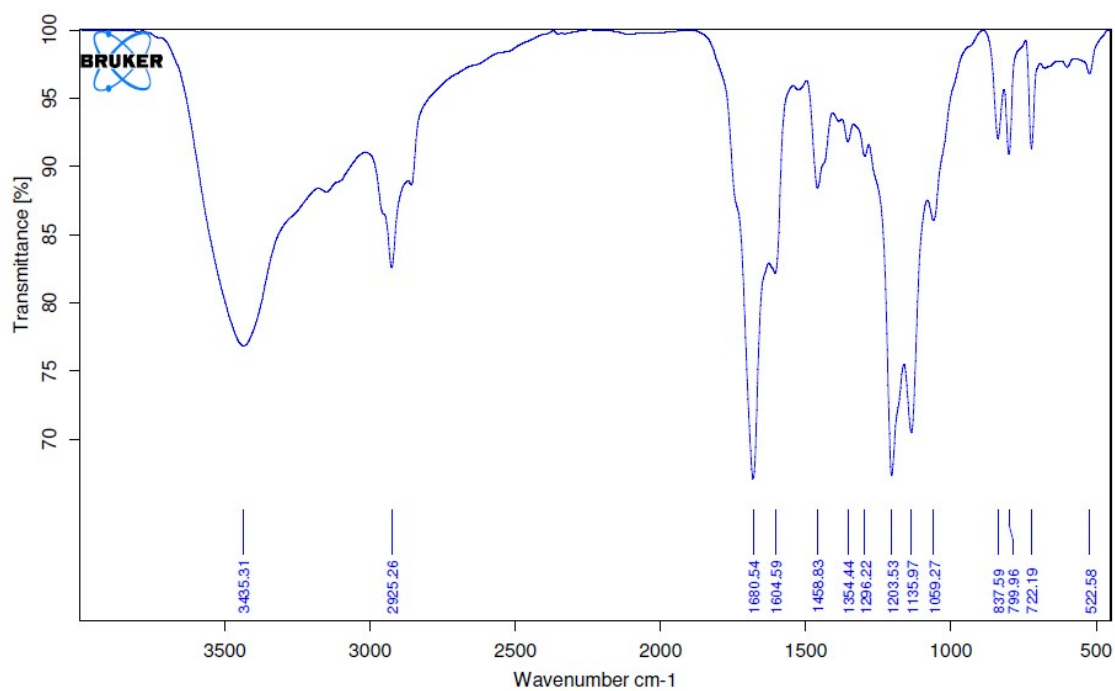


FTIR spectrum (KBr) of compound 7a

Compound 7b.

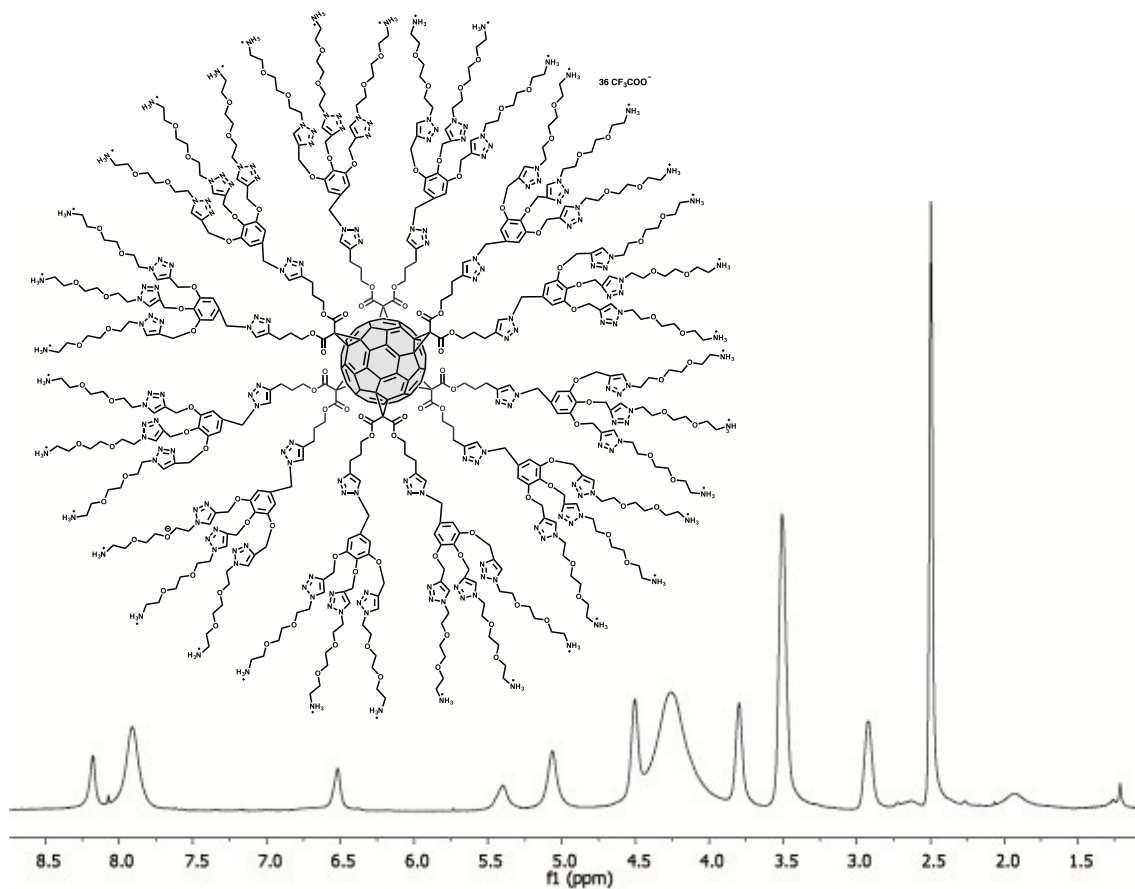


¹H RMN (300 MHz, DMSO-*d*⁶) of compound 7b

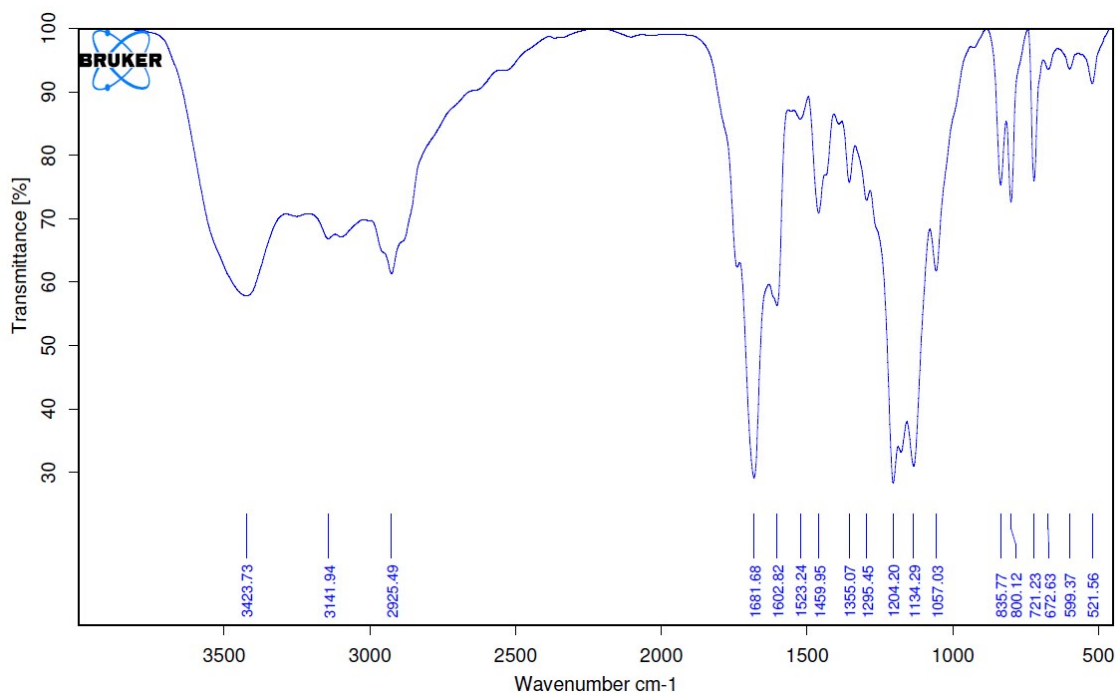


FTIR spectrum (KBr) of compound 7b

Compound 7c.

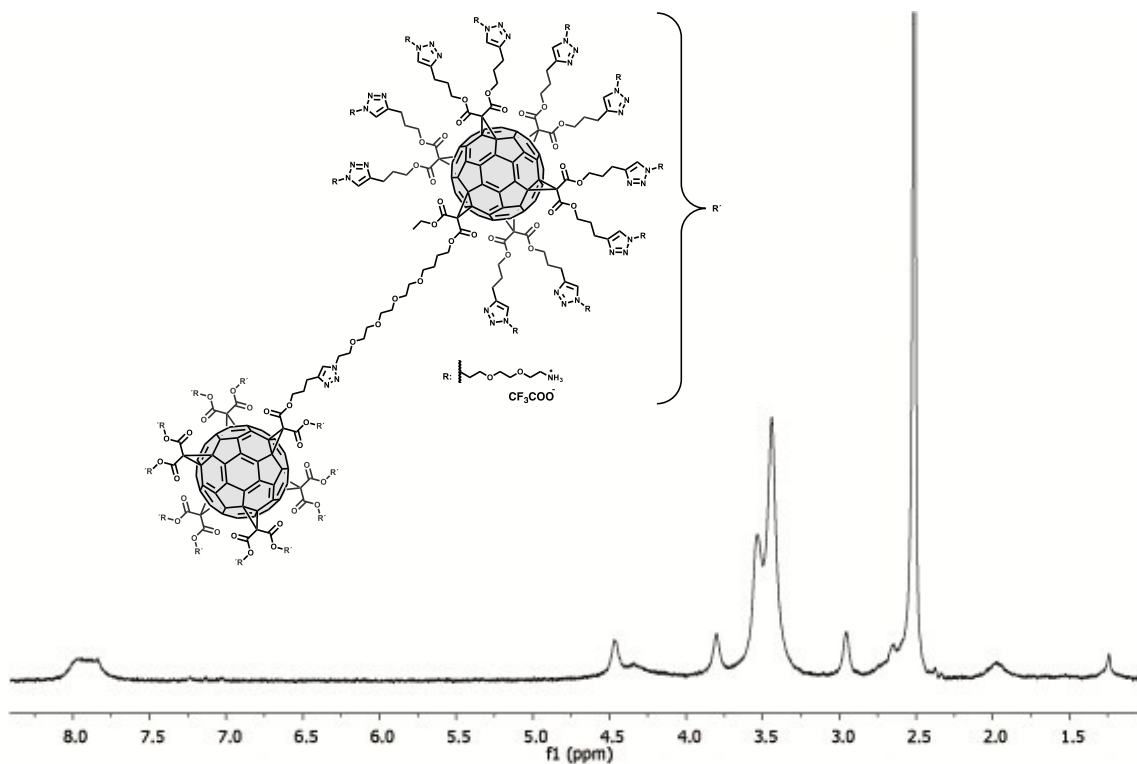


¹H NMR spectrum (300 MHz, DMSO-d₆) of compound 7c

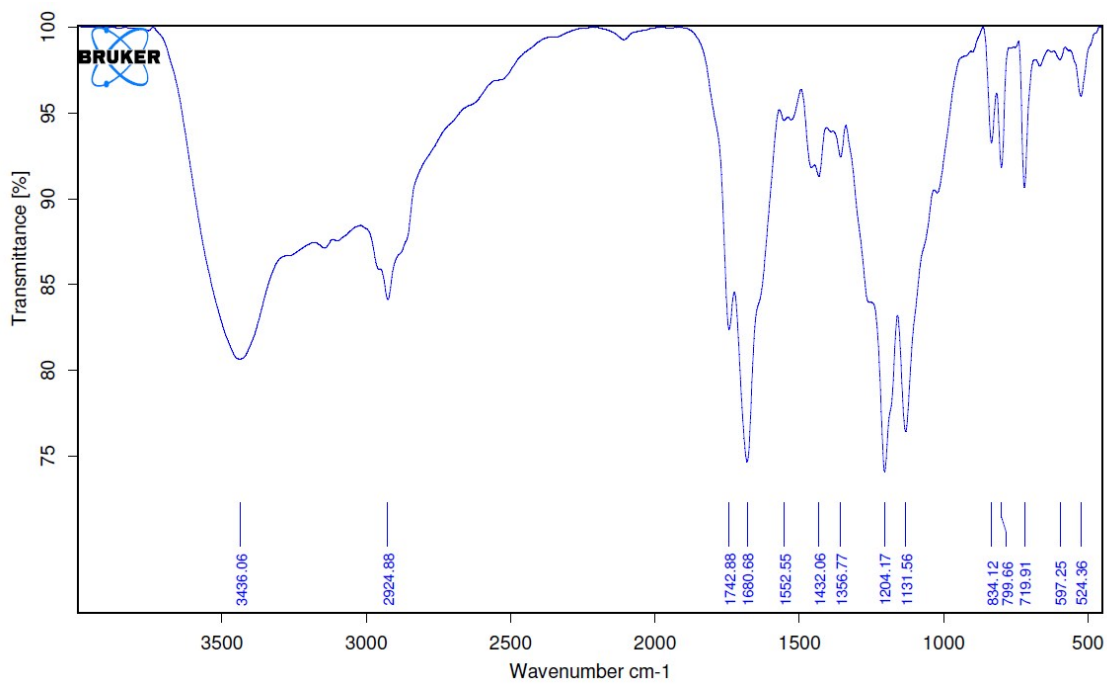


FTIR spectrum (KBr) of compound 7c

Compound 12.



¹H NMR (300 MHz, DMSO-*d*⁶) of compound **12**



FTIR spectrum (KBr) of compound **12**

Determination of the N/P ratios

An average molecular mass of 618 g/mol was considered for each A:T or G:C pair. The EGFP-C1 plasmid has 4700 base pairs and a molecular mass of 2904600 g/mol. Each phosphate group provides one negative charge. That means that each plasmid molecule has 9400 negative charges. In the assays we add 1 μg of plasmid, which is equivalent to $3.44 \cdot 10^{-13}$ mol. Considering the Avogadro number we can calculate $2.074 \cdot 10^{11}$ plasmid units per assay. In summary, for 1 μg of added plasmid we have $1.95 \cdot 10^{15}$ total negative charges. Considering the molecular masses of the fullerene compounds and the amount added, we could obtain the following N/P ratios:

Table S1. N/P Ratios calculated for the concentrations employed in the transfection experiments

	40 μM	8 μM	1.6 μM	0.32 μM	0.064 μM
7a	7.4	1.5	0.3	0.06	0.012
7b	15	3	0.6	0.12	0.025
7c	22.25	4.4	0.9	0.18	0.037
7d	74	15	3	0.6	0.12

Cytotoxicity assay

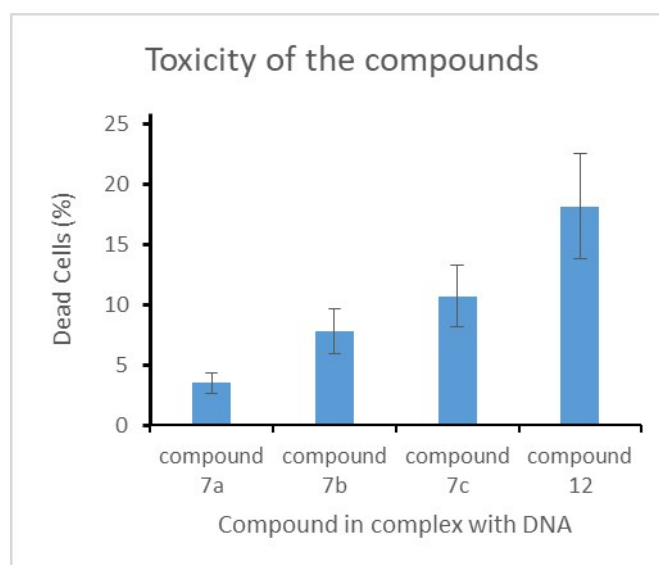


Figure S1. The cytotoxicity exerted by the various fullerene compounds when complexed to plasmidic DNA was determined 12 hours post-addition of the complexes. The amount of dead cells was counted in a field of approximately 1000 cells in total.

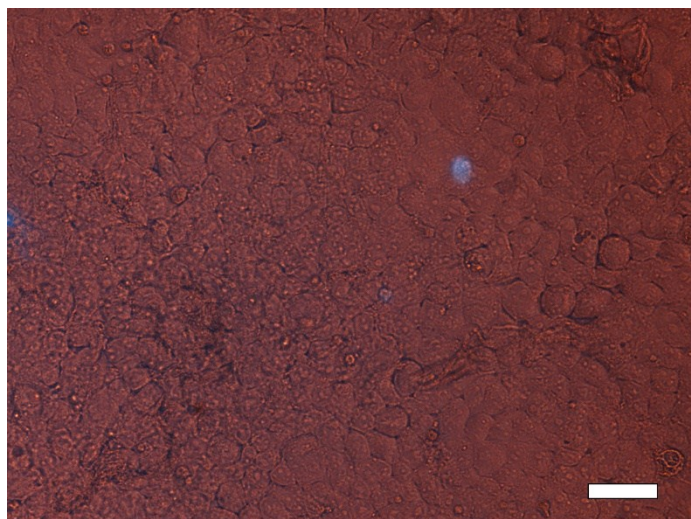


Figure S2. The figure shows an overlay of bright field and 405 nm excitation of confluent HEK293 cells incubated with compound **7a** complexed with plamidic DNA. Emission of blue fluorescence correspond to dead cells with fragmented membranes and the Hoechst dye labelling the cell nuclei. Bar, 50 μ M.

DLS experiments

For compounds **7a-c** and **12**, Dynamic Light Scattering measurements were carried out at 25°C on an ALV GSC08 correlator working in a cross correlation mode with an Ar+ laser operating at $\lambda = 514.5$ nm. The output signals were obtained with backscatter detection at an angle of 90° and processed with a digital correlator that computed intensity-intensity autocorrelation of the scattered light. Measurements were made in a 1-cm path-length round quartz cell maintained at 298 K. Solution samples of 10^{-4} M in PBS buffer were filtered through nylon Acrodisc syringe filters (Pall Life Sciences) with 0.2- μ m pore size.

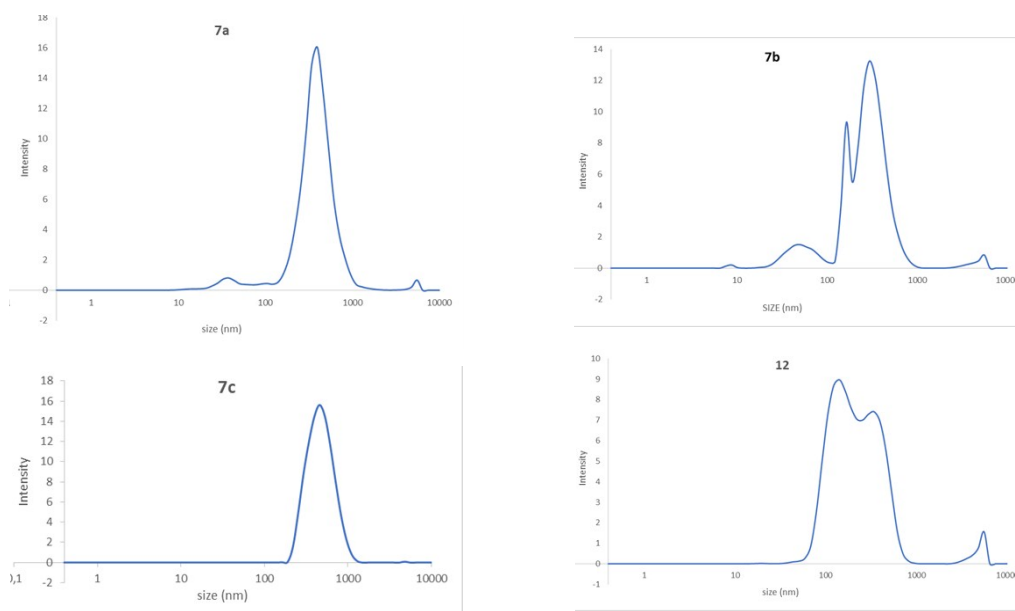


Figure S3. Representative DLS for **7a-c** and **12**, intensity vs. particle size distribution.