Supporting Information

Macrocyclic Polyamine [12]aneN₃ Modified Triphenylamine-Pyrazine Derivatives as Efficient Non-viral Gene Vectors with AIE and Two-photon Imaging Properties

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1. Synthesis



Scheme S1 Schematic route for TDM-A, TDM-B and TDM-C: (i) NaN₃, DMF, 65 °C, 8 h; (ii) NaOH (6 M), Reflux, 24 h; (iii) HCl (2 M); (iv) NMM, EDCI, HOBT, TEA, r.t., 8 h; (v) Propargyl [12]aneN₃, CuBr, CHCl₃, r.t., 30 min; (vi) HCl-EtOAc, DCM, r.t., 2h.

Synthesis of a, b and c. Under argon atmosphere, to the dry DMF (30 mL) solutions of 1- bromhydrin (10 mmol) were slowly added sodium azide (1.30 g, 20 mmol, 2.0 equiv.) at 0 °C. The mixture was slowly warmed up to 65 °C and stirred for overnight. After cooling to room temperature and filtering, saturated brine solution (50 mL) added to the system, and then the mixture solution was extracted with ethyl acetate (20 mL \times 3). The collected organic layers were dried over anhydrous Na₂SO₄. After filtration and vacuum distillation, the crude product was purified by a silica gel column with

PE/EtOAc (v/v = $10:1 \sim 20:1$) as eluent to give corresponding compounds **a**, **b** and **c** as colorless oil in 85 ~ 90% yields.

Compound a: 1.26 g, 8.80 mmol; Yield: 88%. ¹H NMR (600 MHz, CDCl₃) δ 3.55 (t, J = 6.6 Hz, 2H), 3.21 (t, J = 6.9 Hz, 2H), 2.36 (s, 1H), 1.62 – 1.46 (m, 4H), 1.33 (dt, J = 7.0, 3.7 Hz, 4H). ¹³C NMR (101 MHz, CDCl₃): δ 62.57, 51.42, 32.54, 28.84, 26.57, 25.38.

Compound b: 1.54 g, 9 mmol; Yield: 90%. ¹H NMR (600 MHz, CDCl₃) δ 3.48 (t, *J* = 6.7 Hz, 2H), 3.14 (t, *J* = 6.9 Hz, 2H), 2.70 (s, 1H), 1.54 – 1.37 (m, 4H), 1.27 – 1.22 (m, 8H). ¹³C NMR (101 MHz, CDCl₃) δ 62.55, 51.43, 32.67, 29.27, 29.10, 28.79, 26.63, 25.69.δ 62.56, 51.41, 32.53, 28.83 26.56, 25.37.

Compound c: 1.69 g, 8.5 mmol; Yield: 85%.¹H NMR (600 MHz, CDCl₃) δ 3.59 (t, J = 6.7 Hz, 2H), 3.22 (t, J = 7.0 Hz, 2H), 1.80 (s, 1H), 1.61 – 1.48 (m, 4H), 1.36 – 1.22 (m, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 63.00, 51.53, 32.80, 29.53, 29.45, 29.44, 29.18, 28.88, 26.76, 25.79.

Synthesis of compound 2. Compound 1 (1.20 g, 1.56 mmol) were prepared according to literature methods and dissolved in 30 ml of ethanol, and then 60 ml of 6M KOH solution was added. ¹ The mixture was refluxed during 48 h and then cooled to room temperature. The pH was adjusted to 6–7 with 2 M HCl, a yellow color precipitate of colloidal appearance being obtained. The solid was filtered off, washed with ethanol and dried under high vacuum to obtained a pale yellow solid **2** (0.82 g, 1.01 mmol), yield: 65%.¹H NMR (600 MHz, DMSO-*d*₆): δ . 7.55 (dt, *J* = 20.8, 8.0 Hz, 12H), 7.24 (s, 8H), 6.97 (dt, *J* = 21.8, 11.0 Hz, 16H). ¹³C NMR (101 MHz, CDCl₃): δ . 166.41, 151.81,

147.75, 147.44, 142.84, 140.76, 136.05, 133.08, 130.81, 130.13, 128.15, 126.34, 124.83, 123.93, 123.51. HRMS-ESI (m/z): calcd. $[M+H]^+$ for $[C_{54}H_{39}N_4O_4]^+$, 807.2966; found 807.2970.

Synthesis of 3a, 3b and 3c. Under argon atmosphere, compound 2 (2.02 g, 2.5 mmol), HOBT (0.85 g, 6.3 mmol), *N*-methylmorpholine (1.01 g, 10.0 mmol) and EDCI (1.44 g, 7.5 mmol) were dissolved in anhydrous DMF (10 mL). The solution was kept stirring at 0 °C for 30 minutes, a solution of compound **a-c** (7.5 mmol, 3 equiv.) in 5 mL anhydrous DMF was added dropwise, and then allowed to warm up to room temperature and stirred for overnight, 40 mL water was added slowly to quench the reaction and a yellow solid was precipitated. After filtration and vacuum distillation, the crude product was purified by a silica gel column with PE/EtOAc (v/v = 15:1 ~ 20:1) as eluent to give corresponding compounds **3a, 3b, 3c** as yellow solid in 56 ~ 65% yields.

Compound 3a: 1.50 mg, 1.42 mmol; Yield: 56%. ¹H NMR (600 MHz, CDCl₃) δ 7.63 (d, J = 8.4 Hz, 4H), 7.54 (d, J = 8.4 Hz, 4H), 7.48 (d, J = 8.7 Hz, 4H), 7.26 (dd, J = 8.4, 7.5 Hz, 12H), 7.13 – 7.10 (m, 12H), 7.03 (t, J = 7.3 Hz, 4H), 4.44 (t, J = 6.7 Hz, 4H), 3.27 (t, J = 6.9 Hz, 4H), 1.83 (dt, J = 14.3, 7.0 Hz, 4H), 1.63 (dt, J = 14.1, 7.0 Hz, 4H), 1.53 – 1.41 (m, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 164.99, 152.72, 147.88, 147.62, 141.88, 141.67, 135.46, 133.68, 130.45, 129.41, 127.78, 126.59, 124.69, 123.71, 123.25, 66.41, 51.41, 28.84, 28.45, 26.45, 25.57. HRMS-ESI (m/z): calcd. [M+Na]⁺ for [C₆₆H₆₀N₁₀NaO₄]⁺, 1079.4691; found 1079.4703.

Compound 3b: 1.7 g, 1.53 mmol; Yield: 61%. ¹H NMR (600 MHz, CDCl₃) δ 7.63 (d, J = 8.4 Hz, 4H), 7.54 (d, J = 8.5 Hz, 4H), 7.47 (d, J = 8.4 Hz, 4H), 7.27 – 7.24 (d, J =7.4 Hz, 12H), 7.13 – 7.10 (m, 4H), 7.03 (t, J = 7.3 Hz, 4H), 4.42 (t, J = 6.8 Hz, 4H), 3.24 (t, J = 6.9 Hz, 4H), 1.81 (dd, J = 14.5, 7.3 Hz, 4H), 1.61 – 1.56 (m, 4H), 1.48 – 1.42 (m, 4H), 1.40 – 1.34 (m, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 165.00, 152.67, 147.87, 147.62, 141.85, 141.73, 135.48, 133.69, 130.46, 129.41, 127.77, 126.58, 124.68, 123.72, 123.24, 66.63, 51.52, 29.17, 29.12, 28.90, 28.53, 26.71, 25.86. HRMS-ESI (m/z): calcd. [M+Na]⁺ for [C₇₀H₆₈N₁₀NaO₄]⁺, 1135.5317; found 1135.5323.

Compound 3c: 1.90 g, 1.62 mmol; Yield: 65%. ¹H NMR (600 MHz, CDCl₃) δ 7.64 (d, J = 8.4 Hz, 4H), 7.54 (d, J = 8.4 Hz, 4H), 7.48 (d, J = 8.7 Hz, 4H), 7.29 – 7.21 (m, 12H), 7.14 – 7.09 (m, 4H), 7.04 (d, J = 7.4 Hz, 4H), 4.43 (t, J = 6.8 Hz, 4H), 3.24 (t, J = 7.0 Hz, 4H), 1.85 – 1.73 (m, 4H), 1.64 – 1.54 (m, 4H), 1.50 – 1.41 (m, 4H), 1.36 (s), 1.31 (d, J = 1.0 Hz, 16H).¹³C NMR (101 MHz, CDCl₃) δ 164.99, 152.64, 147.86, 147.61, 141.82, 141.74, 135.49, 129.41, 127.77, 124.68, 123.71, 123.25, 66.71, 51.56, 31.53, 29.78, 29.50, 29.48, 29.30, 29.22, 28.93, 28.57, 26.80, 25.94. HRMS-ESI (m/z): calcd. [M+Na]⁺ for [C₇₄H₇₆N₁₀NaO₄]⁺, 1191.5943; found 1191.5948.

Synthesis of target compound TDM-A, TDM-A, TDM-C. Under an argon atmosphere, compounds **3a-3c** (1 mmol) and propargyl [12]aneN₃ (1.02 g, 2.5 mmol)⁴ were dissslved in anhydrous CHCl₃ (10 mL), and catalytic amount CuBr was added. The mixtures were stirred at room temperature for 6 hours, and then adding 30 mL brine, extracted with ethyl acetate (20 mL \times 3), the combined organic layers were dried over anhydrous Na₂SO₄, filtered, and evaporated in vacuo. The obtained crudes were

purified by column chromatography on silica gel with PE/Acetone (v/v = $10:1 \sim 15:1$) as the eluent to give corresponding Boc moieties as yellowish solids. The obtained Boc moieties were subsequently dissolved in a mixture solution with 5 mL anhydrous EtOAc and 1 ml anhydrous EtOH, then 5 mL saturated EtOAc solution of hydrochloric acid was added dropwise to the mixture solution. After stirring for 30 min at room temperature, a large amount of white precipitate obtained. The solid collected by filtering and then washed with aether for several times, and then the solid were dried in vacuo to give the target compounds **TDM-A/B/C** as white solid in 75–85% yields.

TDM-A: 1.10 g, 0.75 mmol; Yield: 75%. FT-IR (KBr, cm-1): 3440 (N–H stretching), 2928 (=C–H stretching), 2736 (CH₂ asymmetrical stretching), 2631 (CH₂ symmetrical stretching), 1597 (C=C and N=N stretching), 1491, 1458 (Ar stretching). ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.56 (d, *J* = 7.7 Hz, 4H), 7.52 (d, *J* = 8.1 Hz, 4H), 7.47 (d, *J* = 7.6 Hz, 4H), 7.22 (t, *J* = 7.2 Hz, 8H), 6.99 (t, *J* = 7.1 Hz, 4H), 6.95 (d, *J* = 7.5 Hz, 8H), 6.91 (d, *J* = 7.7 Hz, 4H), 4.37 (s, 4H), 4.30 (s, 4H), 4.15 (s, 8H), 3.50 – 3.30 (m, 12H), 3.21 (s, 4H), 3.04 (s, 4H), 2.08 (d, *J* = 73.7 Hz, 12H), 1.81 (s, 4H), 1.66 (s, 4H), 1.43 – 1.03 (m, 12H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 164.80, 152.89, 147.79, 147.38, 141.60, 141.00, 135.64, 132.89, 130.85, 130.22, 128.23, 126.43, 124.87, 124.06, 123.44, 66.82, 50.42, 49.61, 47.26, 42.83, 41.79, 30.26, 28.41, 26.90, 26.11, 25.35, 23.10. HRMS-ESI (m/z): calcd. [M+H]⁺ for [C₉₀H₁₀₇N₁₆O₄]⁺, 1475.8656; found 1475.8619.

TDM-B: 1.26 g, 0.82 mmol; Yield: 82%. FT-IR (KBr, cm-1): 3443 (N–H stretching), 2933 (=C–H stretching), 2756 (CH₂ asymmetrical stretching), 2617 (CH2 symmetrical stretching), 1593 (C=C and N=N stretching), 1492, 1462 (Ar stretching). ¹H NMR (600

MHz, DMSO-*d*₆) δ 7.61 (d, *J* = 8.0 Hz, 4H), 7.57 (d, *J* = 8.3 Hz, 4H), 7.51 (d, *J* = 7.8 Hz, 4H), 7.26 (t, *J* = 7.6 Hz, 8H), 7.02 (t, *J* = 7.2 Hz, 4H), 6.98 (d, *J* = 7.8 Hz, 8H), 6.95 (d, *J* = 8.1 Hz, 4H), 4.35 (s, 4H), 4.31 (s, 4H), 4.19 (s, 8H), 3.55 – 3.28 (m, 12H), 3.22 (s, 4H), 3.04 (s, 4H), 2.08 (d, *J* = 62.2 Hz, 12H), 1.78 (s, 4H), 1.66 (s, 4H), 1.40 – 1.10 (m, 20H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 164.81, 152.92, 147.82, 147.42, 141.63, 141.04, 135.70, 132.95, 130.85, 130.20, 128.23, 126.42, 124.88, 124.03, 123.46, 66.77, 50.29, 49.26, 47.04, 43.24, 41.57, 30.25, 29.03, 28.88, 28.56, 28.47, 26.89, 26.36, 25.79. HRMS-ESI (m/z): calcd. [M+H]⁺ for [C₉₄H₁₁₆N₁₆O₄]⁺, 1532.9360; found 1532.9282.

TDM-C: 1.35 g, 0.85 mmol; Yield: 85%. FT-IR (KBr, cm-1): 3443 (N–H stretching), 2928 (=C–H stretching), 2754 (CH₂ asymmetrical stretching), 2623 (CH2 symmetrical stretching), 1591 (C=C and N=N stretching), 1491, 1450 (Ar stretching). ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.62 (d, *J* = 8.0 Hz, 4H), 7.58 (d, *J* = 8.2 Hz, 4H), 7.51 (d, *J* = 7.9 Hz, 4H), 7.27 (t, *J* = 7.6 Hz, 8H), 7.02 (t, *J* = 7.5 Hz, 4H), 6.99 (d, *J* = 7.9 Hz, 8H), 6.95 (d, *J* = 8.2 Hz, 4H), 4.38 – 4.24 (m, 8H), 3.80 (s, 8H), 3.53 – 3.29 (m, 12H), 3.23 – 3.20 (m, 4H), 3.03 (s, 4H), 2.08 (d, *J* = 61.3 Hz, 12H), 1.75 (s, 4H), 1.65 (s, 4H), 1.36 – 1.06 (m, 28H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 164.81, 152.91, 147.82, 147.42, 141.64, 141.04, 135.71, 132.96, 130.84, 130.19, 128.22, 126.41, 124.87, 124.01, 123.46, 66.76, 50.17, 49.13, 47.00, 43.18, 41.45, 30.28, 30.24, 29.42, 29.37, 29.16, 28.95, 28.51, 28.49, 26.39, 25.87. HRMS-ESI (m/z): calcd. [M+H]⁺ for [C₉₈H₁₂₃N₁₆O₄]⁺, 1587.9908; found 1587.9887.



2. Uv-visible spectra of the compounds in different solvents

Fig. S1 UV-vis spectra of TDM-A(A), TDM-B(B) and TDM-C (C) in different solutions. Concentration: $10 \,\mu$ M.

	TDM-A				TDM-B				ТДМ-С			
	λ _{ab} (nm)	λ _{em} (nm)	Stoke's shift	ΦF	λ _{ab} (nm)	λ _{em} (nm)	Stoke's shift	ΦF	λ _{ab} (nm)	λ _{em} (nm)	Stoke's shift	ΦF
			(10^3 cm^{-1})				(10^3 cm^{-1})				$(10^3 \mathrm{cm}^{-1})$	
Toluene	370	547	8.75	0.45	371	542	8.50	0.56	370	543	8.61	0.68
THF	368	610	10.78	0.38	370	607	10.55	0.41	370	607	10.55	0.43
CHCl ₃	380	628	10.39	0.15	377	615	10.27	0.24	380	614	10.03	0.21
CH ₂ Cl ₂	375	655	11.40	0.14	373	657	11.59	0.19	375	649	11.26	0.2
MeCN	365	748	14.03	0.12	365	750	14.06	0.11	365	750	14.06	0.12

Table S1 Summarized photophysical data of TDM-A, TDM-B and TDM-C.

Abbreviation: λ_{ab} = absorption maximum, λ_{em} = emission maximum, ϕ_F = fluorescence quantum yield, concentration: 10 μ M.



Fig. S2 Plot of the emission maximum of TDM-A(A), TDM-B(B), TDM-C(C) in different solvents versus $E_{\rm T}(30)$, $E_{\rm T}(30)$ was the empirical parameter for solvent polarity. Concentrations: 10 μ M, λ ex = 410 nm.

3. Two-photon effect measurement



Fig. S3 TPEF spectra of TDM-A (A), TDM-B (B) and TDM-C (C) at different excitation wavelengths of 720-800 nm. Concentration: $10 \,\mu$ M.



Fig. S4 TPEF spectra of TDM-A (A), TDM-B (B) and TDM-C (C) at different excitation energies at 780 nm. Concentration: 10μ M.





Fig. S5 Fluorescence spectra of TDM-A (A), TDM-B (B), and TDM-C (C) in

water/THF mixtures with THF fractions (f_0) from 0 to 40%. Concentration: 10 μ M;

 $\lambda_{ex} = 410$ nm.



Fig. S6 Fluorescence intensity at 610 nm of **TDM-A** (A), **TDM-B** (B), and **TDM-C** (C) with different THF fractions (f_0). Fluorescence intensity of **TDM-A/B/C** with 99% THF fractions (D). Concentration: 10 μ M; $\lambda_{ex} = 410$ nm.



Fig. S7 SEM images of TDM-A (A), TDM-B (B) and TDM-C (C) in THF/water

mixtures with 99% THF fraction. Scale bars: 1 $\mu m.$



Fig. S8 Particle size distributions of TDM-A (A), TDM-B (B) and TDM-C (C) in

THF/water mixtures with 99% THF fraction. Concentrations = $10 \ \mu$ M.

5. DLS and Zeta potentials



Fig. S9 Particle size distributions of condensed pUC18 DNA (9 μ g/mL) by TDM-A/B/C in Tris-HCl buffer (5 mM, pH 7.4): TDM-A, 10 μ M (A); TDM-B (B), 14 μ M; TDM-C, 16 μ M (C).



Fig. S10 Zeta potentials of pUC18 DNA complexes in 50 mM Trips-HCl (pH 7.4): TDM-A (A), 10 μ M; TDM-B (B), 14 μ M; and TDM-C, 16 μ M (C). Zeta potential histogram of pUC18 DNA complex (D). [DNA] = 9 μ g/mL, 37 °C.

6. Release of the compact DNA



Fig. S11 Gel electrophoresis to measure the reversibility of DNA condensations caused by **TDM-A** (10 μ M), **TDM-B** (14 μ M) and **TDM-C** (16 μ M) at different pH values (A). Gel electrophoresis experiments in the presence of lipase (B). [DNA] = 9 μ g/mL, [lipase] = 10 μ g/mL.

7. Interactions with ctDNAs



Fig. S12 Fluorescence titrations of TDM-A (A), TDM-B (B) and TDM-C (C) with ctDNAs. Plot of emission intensity at 535 nm vs ctDNA concentration (D). Inset: Changes of fluorescent pictures. Concentrations = $10 \ \mu$ M, $\lambda_{ex} = 380$ nm.



Fig. S13 Linearly fitting functions deduced from the plots of emission intensity changes at 535 nm: **TDM-A** (A), **TDM-B** (B), **TDM-C** (C). Concentrations = 10 μ M, λ_{ex} = 380 nm. Plot of I/I_0 - 1 at 535 nm versus the DNA equivalents of **TDM-A/B/C** (D). I₀ = FL intensity in the absence of DNA.

Table S2 Emission enhancement calculated from the fluorescent titration of ctDNA.									
ctDNA (µg/mL)	1	2	3	4	5	6	7	8	
TDM-A	1.69	2.46	3.19	3.93	4.49	5.21	5.9	6.55	
TDM-B	1.63	2.27	2.82	3.27	3.59	3.79	3.98	4.19	
TDM-C	1.61	2.21	2.64	3.18	3.57	3.97	4.21	4.39	

Table S2 Emission enhancement calculated from the fluorescent titration of ctDNA.



Fig. S14 SEM images of condensed ctDNA (9 μ g/mL) by DEDPP-2TPA derivatives in Tris-HCl buffer (5 mM, pH 7.4): TDM-A, 10 *M*m (A); TDM-B, 14 μ M (B); TDM-C, 16 μ M (C). Scale bars: 1 μ m.



Fig. S15 Particle size distributions of DEDPP-2TPA derivatives condensed ctDNA nanoparticles; TDM-A, $10 \,\mu$ M (A); TDM-B, $14 \,\mu$ M (B); TDM-C, $16 \,\mu$ M (C). [pDNA]: $9 \,\mu$ g/mL.

8. EB assay experiment



Fig. S16 Fluorescence quenching curves of EB-bounded ctDNA by TDM-A (A), TDM-B (B) ,TDM-C (C) and plot of $1 - I/I_0$ vs concentrations of TDM-A/B/C (D), , λ_{ex} = 537 nm, [EB] = 20 μ M, [DNA] = 100 μ M, 25 °C, Tris-HCl = 5 mM , NaCl = 50 mM , pH = 7.4.



Fig. S17 Linearly fitting functions deduced from the plots of emission intensity changes at 610 nm by quenching curves of EB-bounded ctDNA: **TDM-A** (A), **TDM-B** (B), **TDM-C** (C).



9. Cellular transfection assays



Fig. S18 Luciferase gene expressions transfected by different concentrations of TDM-A/B/C in three cell lines with Lipo2000 as control. Hela cell lines (A); HepG2 cell lines (B); HEK293T cell lines (C); [pGL-3] = $10 \mu g/mL$.

	TDM-A	TDM-B	TDM-C	TDM-A /DOPE	TDM-B /DOPE	TDM-C /DOPE
Hela	1.1	12	0.3	71	156	85
HepG2	0.6	5.1	< 0.1	64	87	130
HEK293T	<0.1	3.2	< 0.1	50	130	4

Table S3 Optimal luciferase expressions of both the compounds and liposomes indifferent cell lines (% of lipo2000).



Fig. S19 Fluorescence microscope images (10×) of pEGFP-transfected in Hela cells. pEGFP was observed with FITC filter (A); The inherent red fluorescence of **TDM-A/B/C** were observed with the TRITC filter (B); Bright field (C); Merge of A, B, C (D). pEGFP; Lipo2000 and naked pEGFP were used as control. Concentration = 20 μ M; [pEGFP-N1] = 10 μ g/mL, scale bar: 100 μ m.

10. Cellular uptake studies



Fig. S20 Cellular uptake of **TDM-B**/DOPE-DNA complexes incubated at 37 °C (A) and 4 °C (B) in Hela cell lines.



Fig. S21 Cellular uptake of **TDM-B**/DOPE-DNA complexes in the presence of chlorpromazine (A), methyl-β-cyclodextrin (B) and amiloride hydrochloride (C) in Hela cell lines.

11. Elemental analysis of TDM-A, TDM-B and TDM-C

 Table S4 Elemental analysis of the obtained TDM-A, TDM-B and TDM-C for their hydrochloride salts.

 Name
 Nitrogen [%]

 Carbon [%]
 Hydrogen [%]

Name	Nitrogen [%]		Carbo	on [%]	Hydrogen [%]		
	calcd.	found	calcd.	found	calcd.	found	
TDM-A	12.68	12.15	61.16	61.15	6.5	6.46	
TDM-B	12.29	11.88	61.91	62.86	6.74	6.81	
TDM-C	11.92	10.35	62.62	61.56	6.97	6.89	

12. ¹H, ¹³C-NMR Spectra and HR-MS of synthesized compounds



Fig. S23 ¹³C NMR spectrum of a in CDCl₃ (100 MHz, CDCl₃).



Fig. S25 ¹³C NMR spectrum of b in CDCl₃ (100 MHz, CDCl₃).



Fig. S27 ¹³C NMR spectrum of c in CDCl₃ (100 MHz, CDCl₃).

7.582 7.556 7.556 7.556 7.556 7.556 7.556 7.521 7.510 7.508 7.508 7.010 6.974 6.974 6.974 6.974 6.974 6.937





200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm

Fig. S29 ¹³C NMR spectrum of 2 in CDCl₃ (100 MHz, CDCl₃).



Fig. S30 HR-MS spectrum of 2.



Fig. S31 ¹H NMR spectrum of 3a (400 MHz, CDCl₃).



Fig. S33 HR-MS spectrum of 3a.



Fig. S35 ¹³C NMR spectrum of **3b** in CDCl₃ (100 MHz, CDCl₃).



Fig. S36 HR-MS spectrum of 3b.



Fig. S37 ¹H NMR spectrum of 3c (400 MHz, DMSO).



Fig. S38 ¹³C NMR spectrum of 3c in DMSO (100 MHz, DMSO).



Fig. S39 HR-MS spectrum of 3c.



Fig. S41 ¹³C NMR spectrum of TDM-A in DMSO (100 MHz, DMSO).



Fig. S43 ¹H NMR spectrum of TDM-B (400 MHz, DMSO).



Fig. S44 ¹³C NMR spectrum of TDM-B in DMSO (100 MHz, DMSO).



Fig. S45 HR-MS spectrum of TDM-B.



Fig. S47 ¹³C NMR spectrum of TDM-C in CDCl₃ (100 MHz, CDCl₃).



Fig. S48 HR-MS spectrum of TDM-C.

References:

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