

## Supplementary

for

### Circulating Tumour Cells-Targeting Au-Nanocages-Mediated Bimodal Phototherapeutic Properties Enriched by Magnetic Nanocore

Chih-Sheng Chiang,<sup>a</sup> Yu-Che Kao,<sup>b</sup> Thomas J. Webster,<sup>c</sup> Woei-Cherng Shyu,<sup>d</sup> Hung-Wei Cheng,<sup>e</sup> Tse-Ying Liu<sup>f</sup> and San-Yuan Chen<sup>e,g,h,\*</sup>

a. Dr. Chih-Sheng Chiang  
Cell Therapy Center, China Medical University Hospital  
Taichung City 404, Taiwan  
E-mail: [brian78630g@gmail.com](mailto:brian78630g@gmail.com)

b. MS. Yu-Che Kao  
Materials Engineering / School of Materials, The University of Manchester  
Oxford Rd Manchester M13 9PL UK  
E-mail: [kao1031@livemail.tw](mailto:kao1031@livemail.tw)

c. Prof. Thomas J. Webster  
Department of Chemical Engineering, Northeastern University,  
Boston, MA 02115, U.S.A.  
E-mail: [th.webster@neu.edu](mailto:th.webster@neu.edu)

d. Prof. Woei-Cherng Shyu  
Graduate Institute of Biomedical Science, China Medical University  
Taichung, Taiwan 40440  
E-mail: [shyu9423@gmail.com](mailto:shyu9423@gmail.com)

e. MS. Hung-Wei Cheng and Prof. San-Yuan Chen  
Department of Materials Science and Engineering, National Chiao Tung University  
Hsinchu, Taiwan 30010  
E-mail: [sanyuanchen@nctu.edu.tw](mailto:sanyuanchen@nctu.edu.tw)

f. Prof. Tse-Ying Liu  
Department of Biomedical Engineering, National Yang Ming University  
Taipei City 112, Taiwan  
E-mail: [tyliu5@ym.edu.tw](mailto:tyliu5@ym.edu.tw)

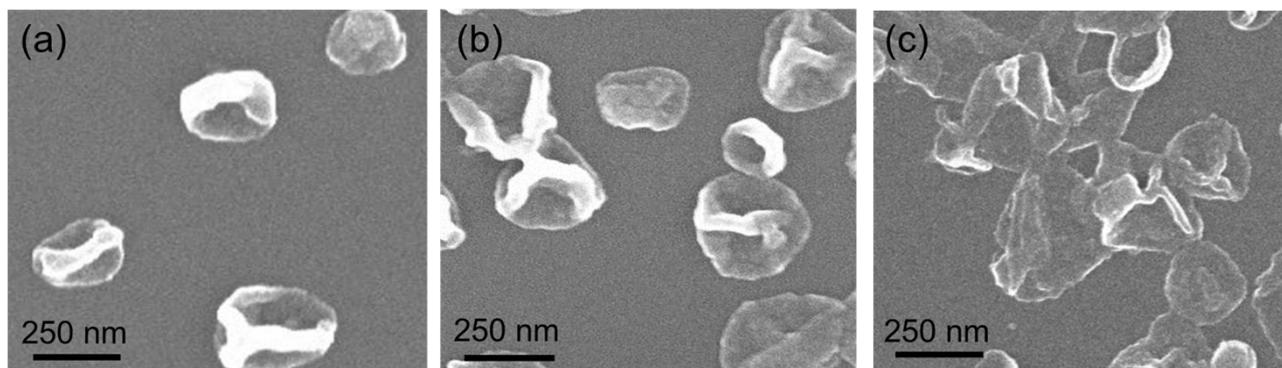
g. Prof. San-Yuan Chen  
Frontier Research Centre on Fundamental and Applied Sciences of Matters, National Tsing Hua  
University,  
Hsinchu, Taiwan  
E-mail: [sanyuanchen@nctu.edu.tw](mailto:sanyuanchen@nctu.edu.tw)

h. Prof. San-Yuan Chen  
School of Dentistry, College of Dental Medicine, Kaohsiung Medical University  
Kaohsiung, Taiwan  
E-mail: [sanyuanchen@nctu.edu.tw](mailto:sanyuanchen@nctu.edu.tw)

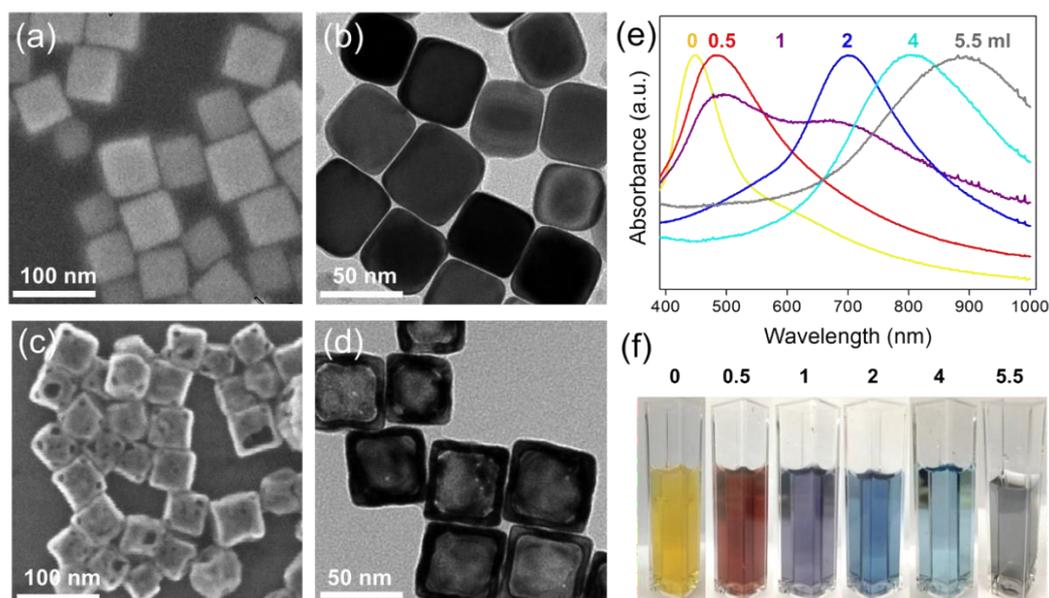
**Table S1.** Characterization of TPVA modified with different TGA volume concentrations. The initial PVA concentration was fixed at 2 wt % for all samples.

Sample	TGA (vol %)	Thiol content (mmol g <sup>-1</sup> )	Size <sup>a</sup> (nm)
TPVA <sub>0.5</sub>	0.5	0.179	223.8 ± 18.1
TPVA <sub>1</sub>	1	0.34	262.8 ± 34.8
TPVA <sub>2.5</sub>	2.5	1.23	358.2 ± 50.5

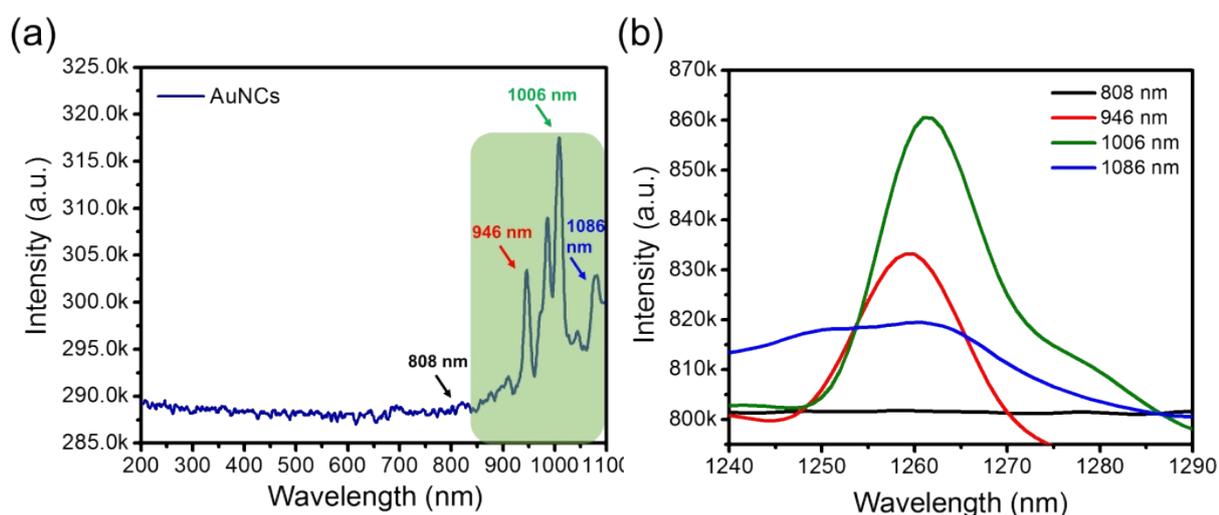
<sup>a</sup> the size of the magnetic core synthesized from TPVA with different thiol content.



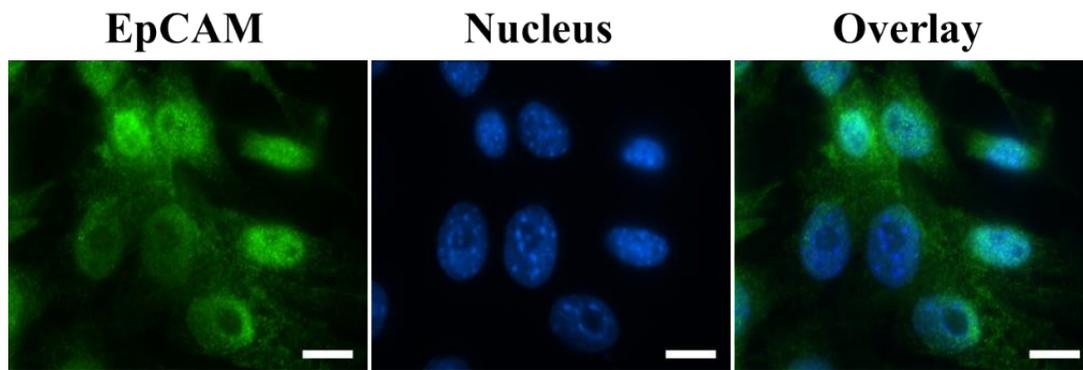
**Fig. S1** The morphology of the magnetic core was related to the thiol content of TPVA. SEM images of MN synthesized from TPVA modified with (a) 0.5, (b) 1, and (c) 2.5 % (v/v) TGA after an esterification reaction, respectively.



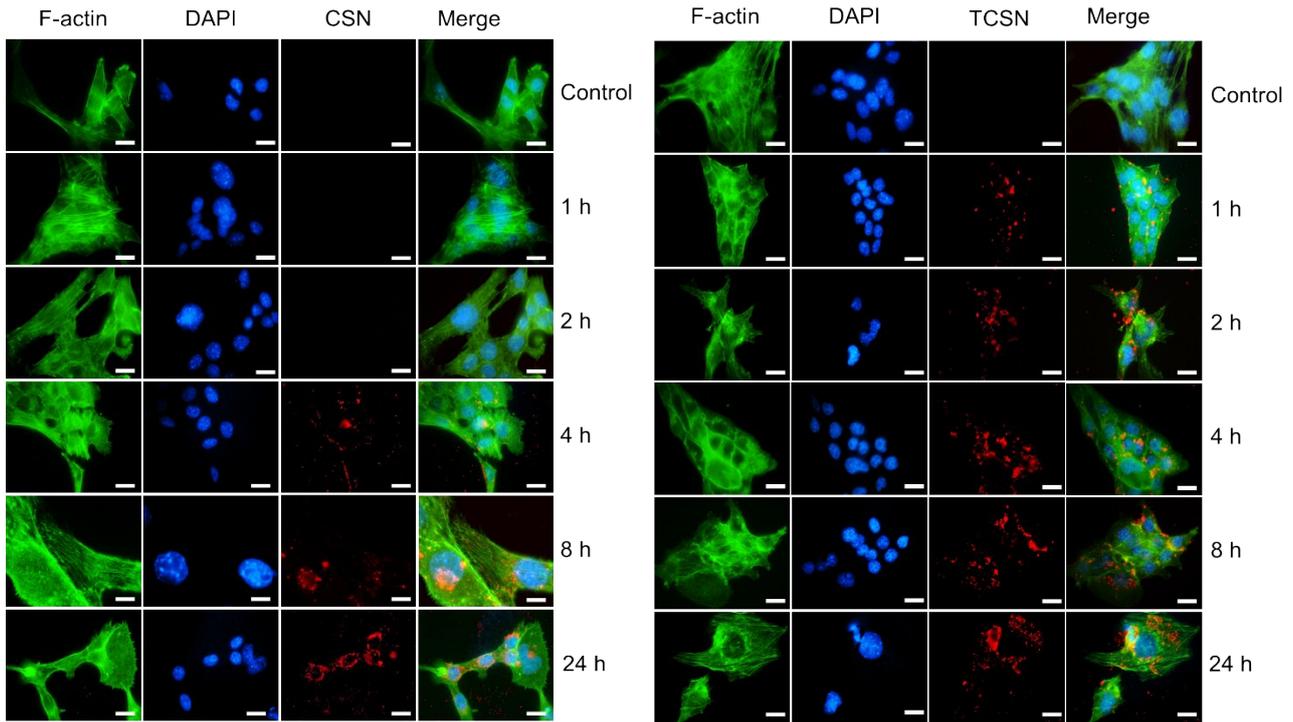
**Fig. S2** Images and optical properties of Ag nanocubes and AuNCs. (a) SEM and (b) TEM image of the Ag nanocubes, which showed approximately 50 nm in edge length. (c) SEM image and (d) TEM image of AuNCs in a hollow and porous structure. (e) UV-visible-NIR absorbance spectra of Ag nanocubes and AuNCs. Different volumes of the  $\text{HAuCl}_4$  solution (0, 0.5, 1, 2, 4, 5.5 ml) were used to titrate the Ag nanocubes suspension. (f) Brilliant color changes during the Ag-Au alloy formation.



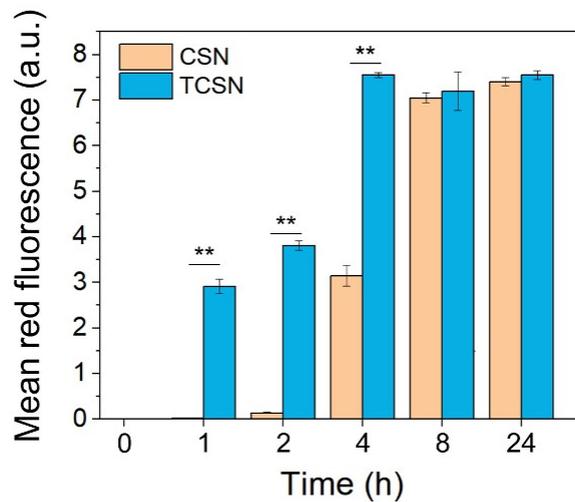
**Fig. S3** Direct detection of singlet oxygen ( $^1\text{O}_2$ ) phosphorescence from the photoexcitation of AuNCs in a  $\text{D}_2\text{O}$  solvent. (a) The excitation spectrum of AuNCs and strong spectral peaks that exist after 900 nm. (b)  $^1\text{O}_2$  phosphorescence emission spectra by sensitization of AuNCs using different NIR wavelengths.



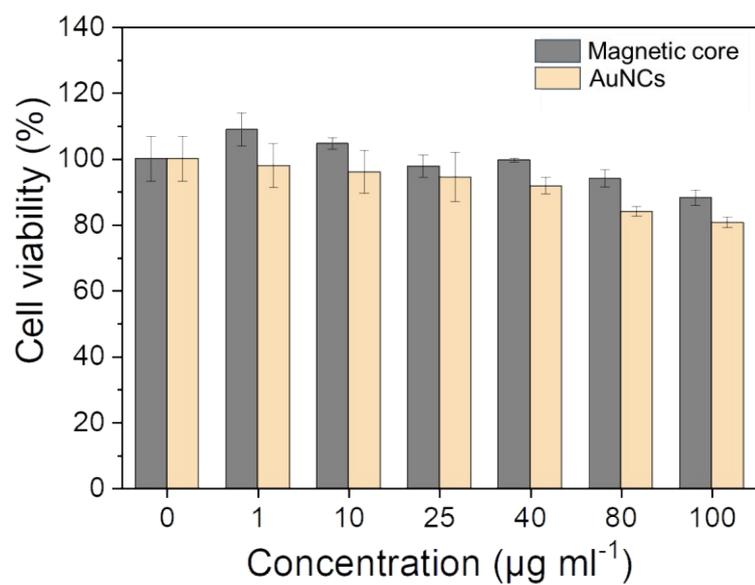
**Fig. S4** EpCAM is a surface antigen used for the targeted delivery of QD-labeled TPMN to 4T1 cells. High expression of EpCAM on 4T1 cells was confirmed using immunofluorescence staining on 4T1 cells with a secondary FITC-labeled anti-rabbit antibody. Scale bars = 10  $\mu\text{m}$ .



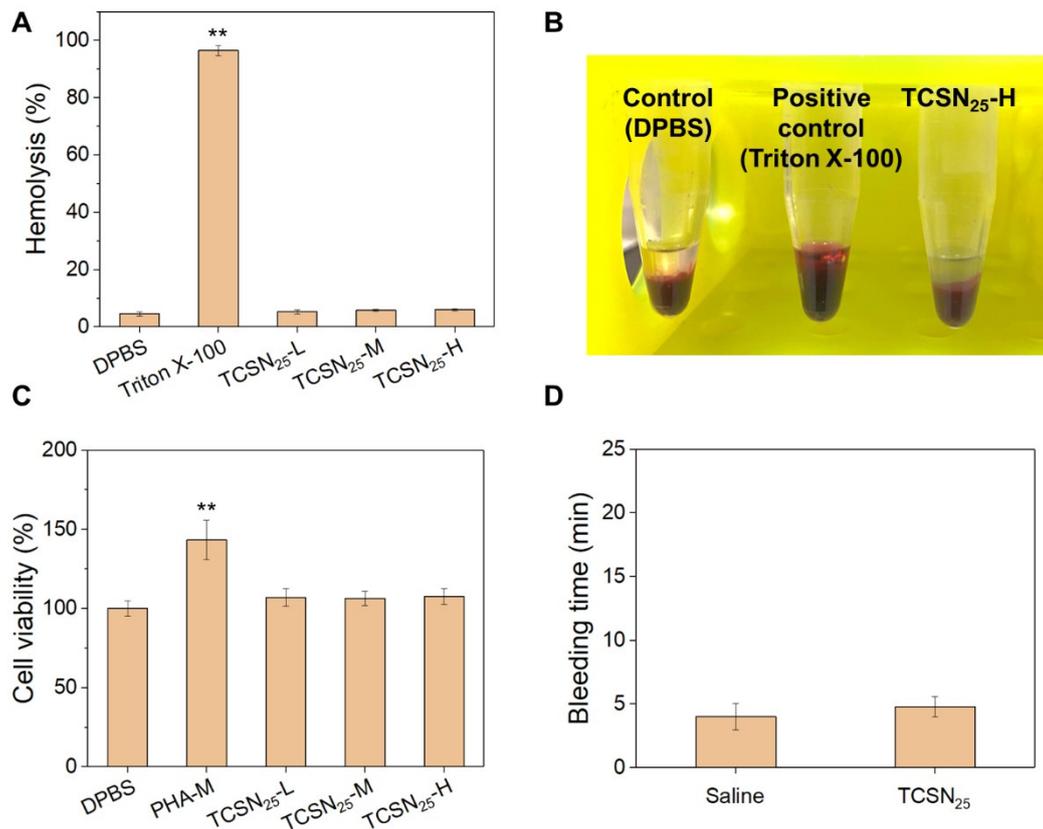
**Fig. S5** Cell association of non-targeted QD-labeled CSN and TCSN (red) incubated for 1, 2, 4, 8, and 24 h with 4T1 cells. F-actin and nuclei of 4T1 cells were stained with Alexa-488 (green) and DAPI (blue). Scale bars = 10  $\mu\text{m}$ .



**Fig. S6** Statistical analysis of cellular association at 0, 1, 2, 4, 8, and 24 incubation with CSN or TCSN. Images from fluorescent microscopy were analyzed using ImageJ. Cell association level between CSN and TCSN showed significant difference at short culture period including 1, 2, and 4h incubation, but no significant difference was found in longer culture time (i.e., 8 and 24h). Data are expressed as mean  $\pm$  SD,  $n = 6$  biologically independent samples. \* $p < 0.05$  and \*\*  $p < 0.01$  between groups using an independent sample t-test.



**Fig. S7** Evaluation of cell viability of 4T1 cells after incubation with various concentrations of the magnetic core and AuNCs in the dark for 24 h. The result is expressed as the mean  $\pm$  SD, n= 5.



**Fig. S8** Blood compatibility of TCSN. (A) Evaluation of hemolytic reaction of DPBS (negative control), Triton X-100 (positive control), low-concentration TCSN<sub>25</sub> (TCSN<sub>25</sub>-L, [Au]= 10  $\mu\text{g mL}^{-1}$ ), medium-concentration TCSN<sub>25</sub> (TCSN<sub>25</sub>-M, [Au]= 20  $\mu\text{g mL}^{-1}$ ), or high-concentration TCSN<sub>25</sub> (TCSN<sub>25</sub>-H, [Au]= 40  $\mu\text{g mL}^{-1}$ ), n= 3. (B) Visual observation of the blood after incubation with DPBS, Triton X-100, and TCSN-H for 3h. (C) Cell viability of peripheral blood mononuclear cell (PBMC) after incubation with DPBS, PHA-M containing RPMI-1640 media, TCSN<sub>25</sub>-L, TCSN<sub>25</sub>-M, and TCSN<sub>25</sub>-H for 24 h, n= 5. (D) Tail bleeding test for monitoring the coagulation time. After the mouse was treated with saline or TCSN<sub>25</sub>, the tail was amputated and the blood were collected and measured to confirm the bleeding time, n= 3. All results are expressed as mean  $\pm$  SD, n  $\geq$  3 biologically independent animals or samples. Comparison between two groups was performed using an unpaired two-tailed t-test. Symbol (\*\*) represents a significant difference (p < 0.01) compared with control.

**Table S2. Semi-quantification of Prussian blue stain**

Group	1 day after treatment			7 day after treatment		
Animal ID	1	2	3	4	5	6
<b>Lung</b>	3	1	1	1	1	1
<b>Liver, red pulp</b>	3	1	2	1	1	1
<b>Spleen, sinusoid</b>	3	1	2	2	1	1
<b>Kidney</b>	0	0	0	0	0	0
<b>Heart</b>	0	0	0	0	0	0

The Perls' Prussian blue stained preparations were graded semi-quantitatively: 0= no iron; 1= minimal or very small amounts; 2= slight and patchy; 3= moderate and diffuse; 4= strong, extensive and diffuse content.

**Table S3. Histopathology incidence table**

Group	1 day after treatment			7 days after treatment		
Animal ID	1	2	3	4	5	6
<b>Lung</b>						
Foreign body inflammation	3	2	3	1	1	1
<b>Liver</b>						
Pigment deposition, sinusoid	3	1	2	1	X	1
<b>Spleen</b>						
Pigment deposition, red pulp	3	1	3	1	1	X

X: No significant lesions; degree of lesions was graded from one to five depending on severity: 1= minimal (< 1%); 2= slight (1-25%); 3= moderate (26-50%); 4= moderate/severe (51-75%); 5= severe/high (76-100%).