

Supplemental Information

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Materials and Methods

Materials: DNA staple strands were purchased from IDT. M13 bacteriophage scaffold DNA (p7560) was produced following a published protocol.¹ 10 nm gold nanoparticles (AuNPs) were purchased from Ted Pella, Inc.

DNA origami hexagon tile design and assembly: DNA origami hexagon tile (DHT) was designed by using caDNAno. In a typical annealing protocol, staples were mixed with single stranded DNA scaffold (p7560, 10 nM) in 5-fold molar excess in aqueous buffer composing of 5 mM Tris base, 1 mM EDTA, and 10 mM MgCl₂, with a total volume of 100 μL. The mixed solution was slowly cooled down to room temperature (24 °C) in a PCR thermal cycler over 18 hrs under the following protocol: 65 °C for 5 mins, 60 to 24 °C, at 30 mins/°C.

Conjugating DNA onto AuNPs: 50 mL of colloidal citrated 10nm AuNP solution was added with 15 mg of Bis(p-sulfonatophenyl) phenylphosphine dihydrate dipotassium salt (BSPP) and shook overnight. Then NaCl was added until a color change of the solution from red to blue was observed. The AuNPs were centrifuged to the bottom at 1,000 rcf for 30 min and the clear supernatant was removed with a pipette. The particles were dissolved in BSPP solution (1 mL, 2.5 mM) followed by addition of 3 mL of methanol. The particles were centrifuged again and re-suspended in 1 mL of BSPP. The concentration of the AuNPs was determined from the optical absorption at a wavelength of 520 nm using a Nanodrop spectrophotometer. For reducing the disulfide bonds of the thiolated ssDNA strands to monothiol, the modified strands were incubated with 100-fold of TCEP (Tris(carboxyethyl) phosphine hydrochloride) for at least 30 mins. AuNPs and thiolated DNA were mixed in 0.5×TBE buffer at a designated ratio (1: 100). Within a period of 24 hrs, NaCl solution was added to the solution to achieve a final concentration of 300 mM. To remove the unbound DNA strands, the mixture was spun for 30 min at 18,000 rcf, and the supernatant was removed carefully without disturbing the pellet. 4 additional washing and spinning steps (30 min, 18,000 rcf) with 100 μL of 0.5 × TBE buffer containing 300 mM NaCl were conducted to fully removed unbound DNA.

Attaching AuNPs onto DNA origami templates: DNA origami structures were assembled first, as described above, which were then added with 5-fold (in terms of binding site, 1.5-fold for 2D lattice designs) of DNA-conjugated AuNPs, incubated for at least 2 hours at room temperature. Cluster structures were then subject to agarose gel electrophoresis and corresponding bands were extracted. Superlattice structures were directly characterized.

Gel electrophoresis: DNA origami with AuNPs forming clusters were run on 1% native agarose gel electrophoresis for 2 hours (gel prepared in 0.5 × TBE buffer supplemented with 10 mM MgCl₂ and 0.005% (v/v) EtBr) in an ice water bath. Then, the target gel bands were excised and placed into a

Freeze 'N Squeeze column (Bio-Rad Laboratories, Inc.). The gel pieces were crushed into fine pieces by a microtube pestle in the column, and the column was then centrifuged at 7000 rcf for 5 minutes. Samples that were extracted through the column were collected for subsequent experiments.

TEM imaging: 10 μ L of samples were deposited onto glow-discharged, carbon-coated copper TEM grids for 5-30 minutes inside a moisture chamber. Residual solution was dried by filter paper. The grids were then stained for 1 minute using a 1% aqueous uranyl formate solution containing 25 mM NaOH. Imaging was performed using a Hitachi-7700 microscope operated at 80 kV.

Stability study: 9-AuNP clusters were prepared as illustrated in the above protocol. 10 μ L of 10 nM nanoparticle clusters were incubated in 1 \times TE/Mg²⁺(10 mM), 1 \times PBS, or RPMI-1640/10% FBS at 37 °C for 0.5, 1, 2, or 4 hours. Immediately after incubation, the samples were subject to 1% native agarose gel electrophoresis running in 0.5 \times TBE buffer supplemented with 10 mM MgCl₂.

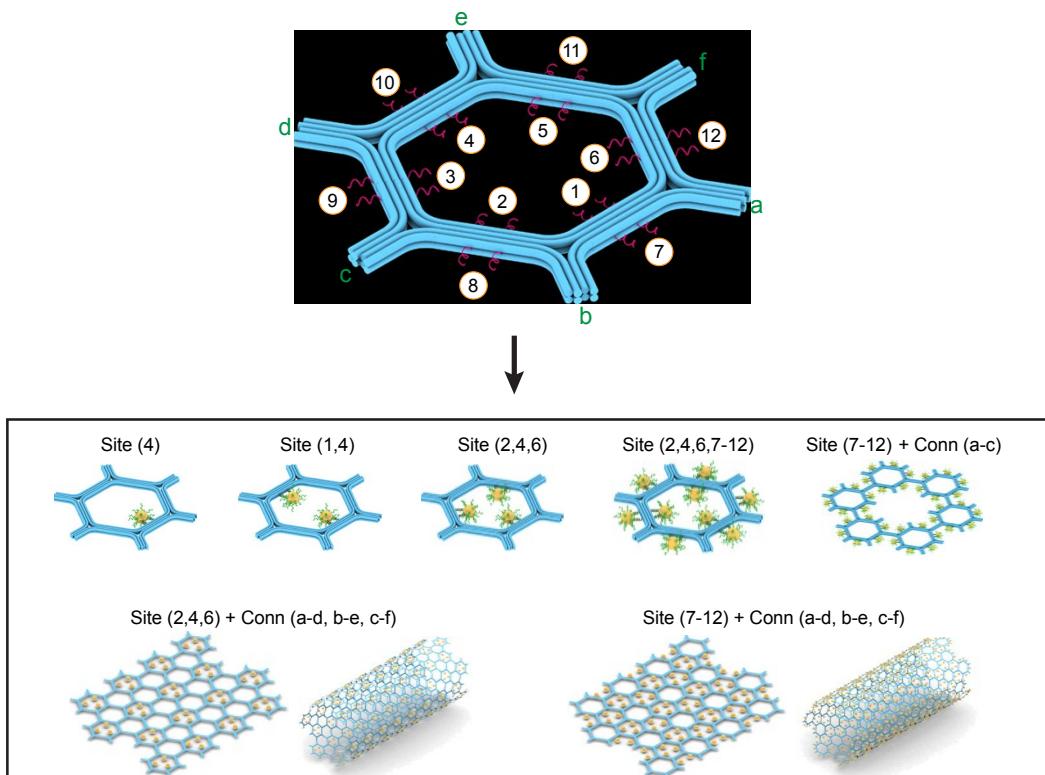


Figure S1. Summary figure. DHT-guided assembly of gold nanoparticle clusters and lattices. The use of docking handle strands and connector strands are indicated for each architecture.

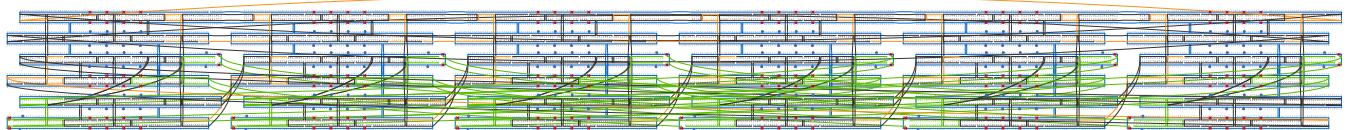


Figure S2. Strand diagram of DHT. Blue: scaffold DNA; black: core staples; orange: capture strands; green: connector strands. Zoom in to see details.

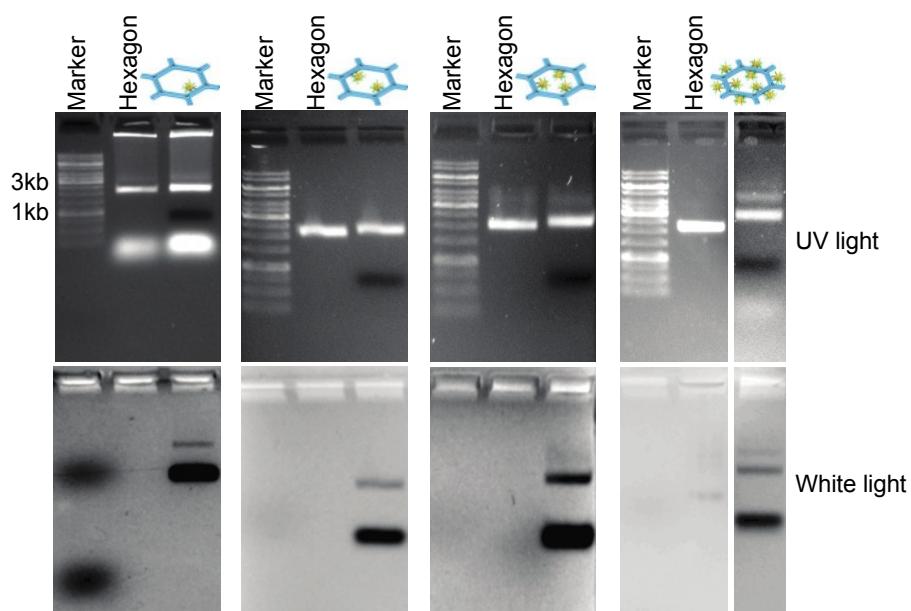


Figure S3. Agarose gel electrophoresis (1%) of nanoparticle clusters assembled on DHT. Top row: UV illumination; bottom row: white light.

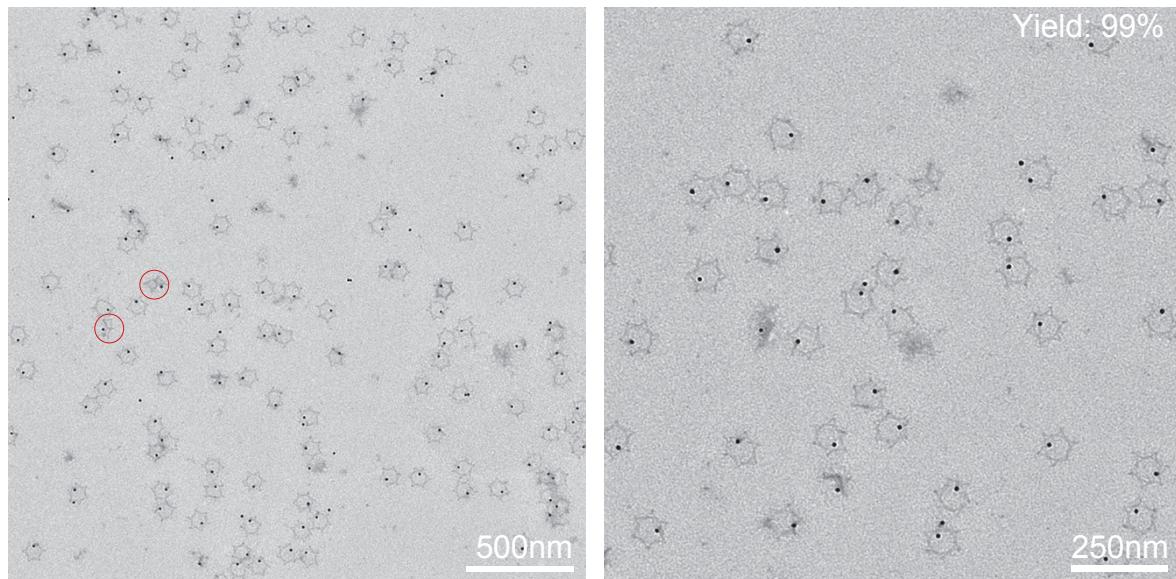


Figure S4. 1-AuNP cluster on DHT. Yield is calculated based on TEM images ($N=116$). Wrongly assembled clusters were marked by a red circle.

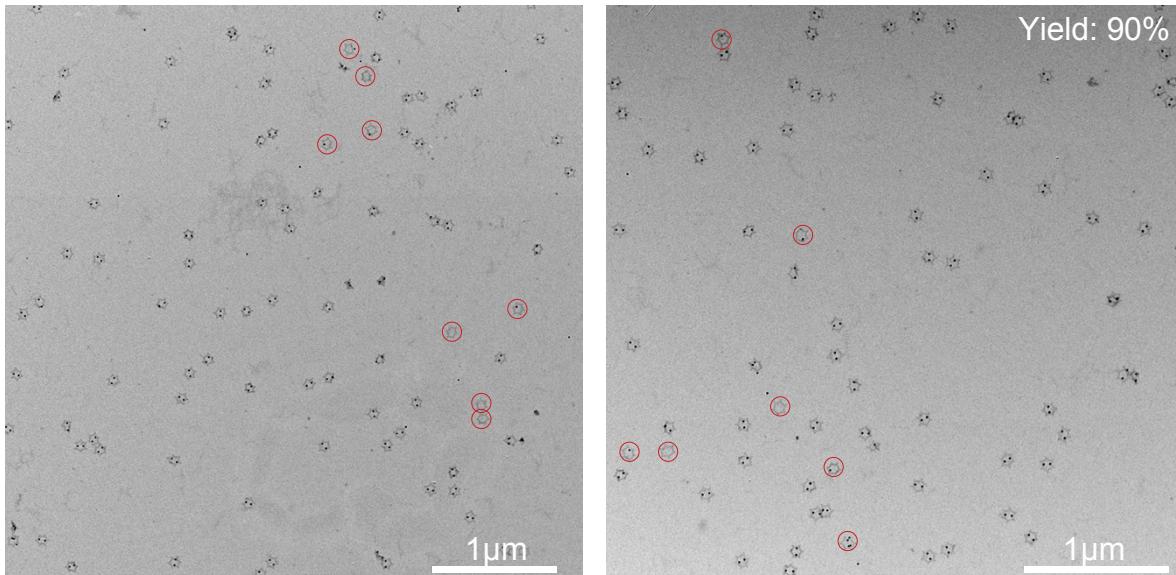


Figure S5. 2-AuNP cluster on DHT. Yield is calculated based on the images ($N=205$). Wrongly assembled clusters were marked by a red circle.

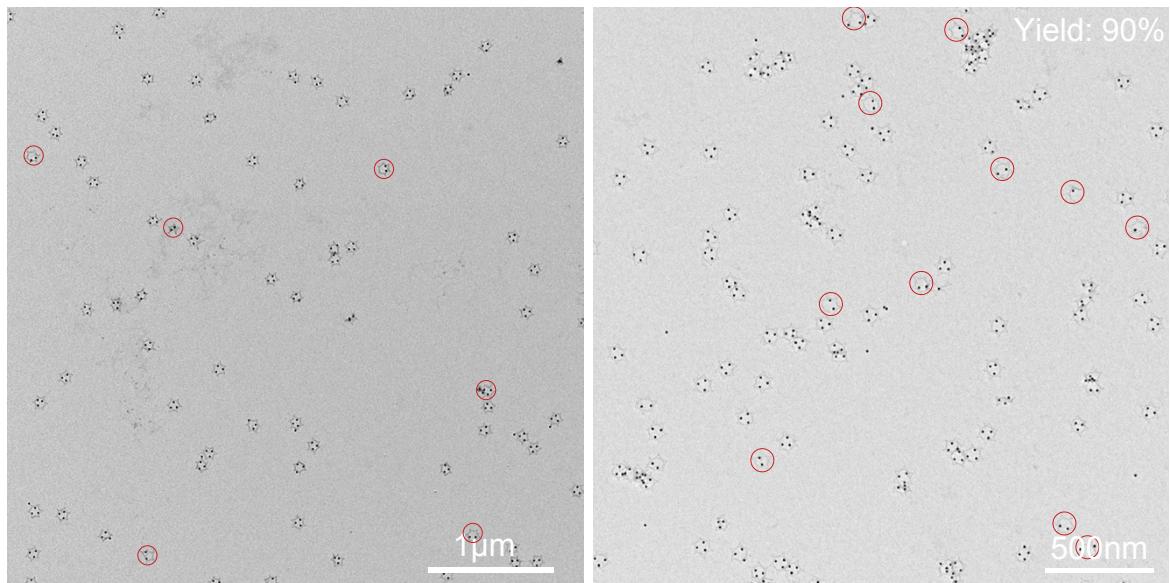


Figure S6. 3-AuNP cluster on DHT. Yield is calculated based on the images ($N=155$). Wrongly assembled clusters were marked by a red circle.

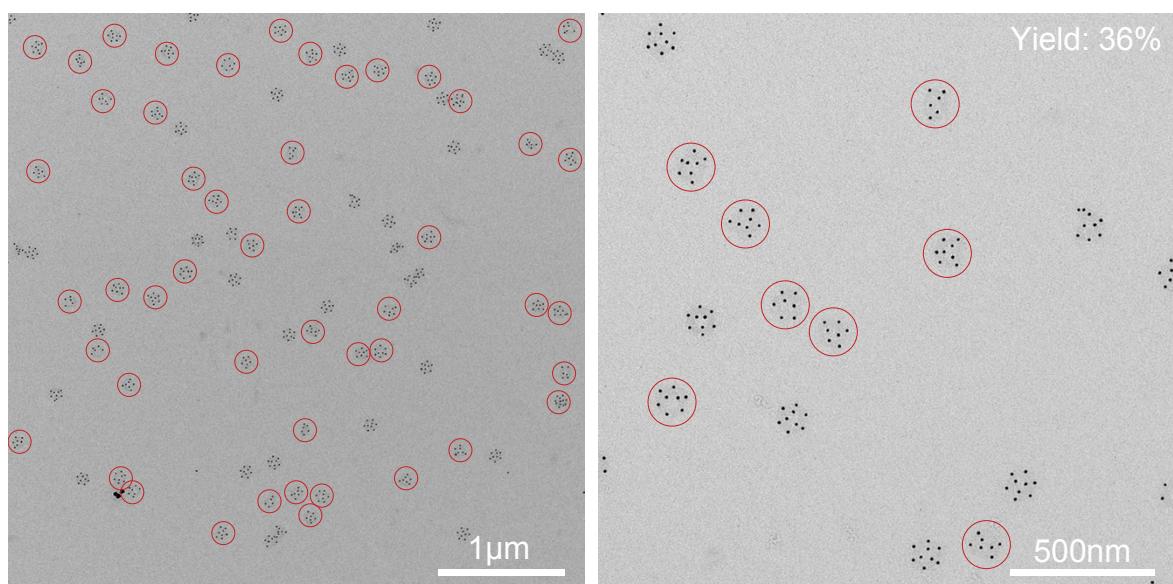


Figure S7. 9-AuNP cluster on DHT. Yield is calculated based on the images ($N=87$). Wrongly assembled clusters were marked by a red circle.

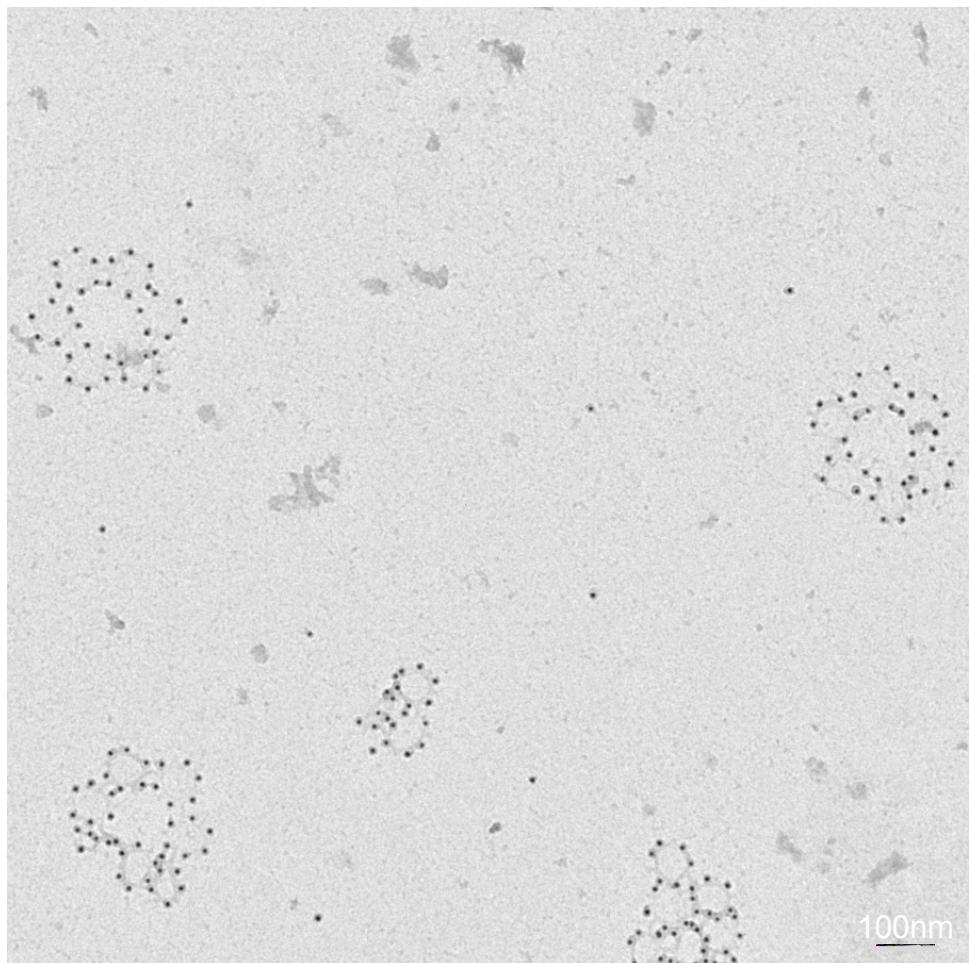


Figure S8. 36-AuNP cluster on DHT hexamer.

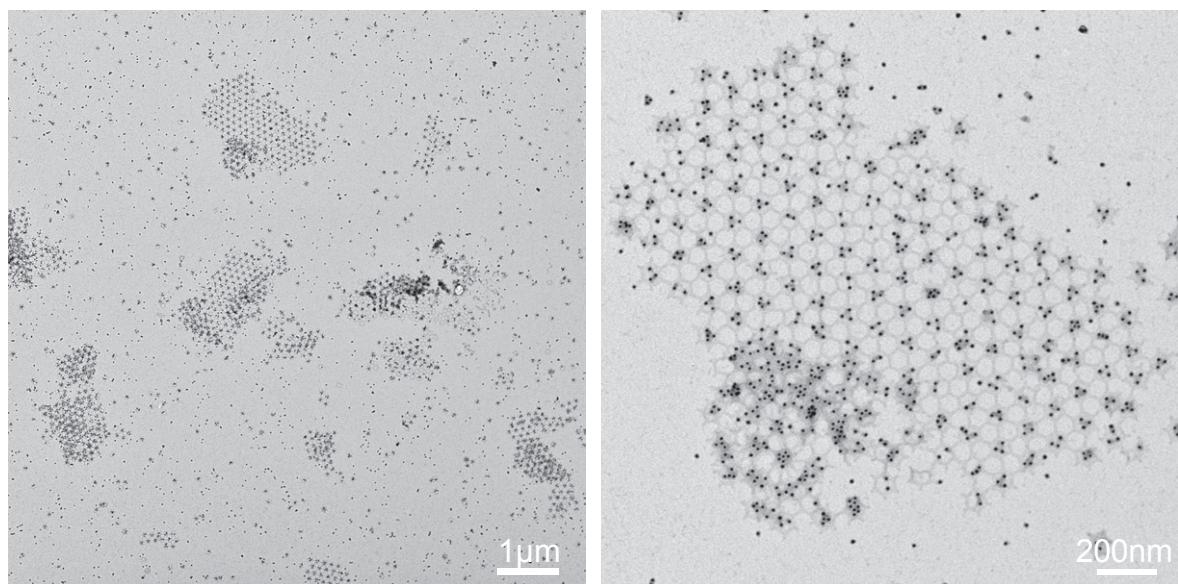


Figure S9. AuNP lattices on honeycomb array with type-1 configuration.

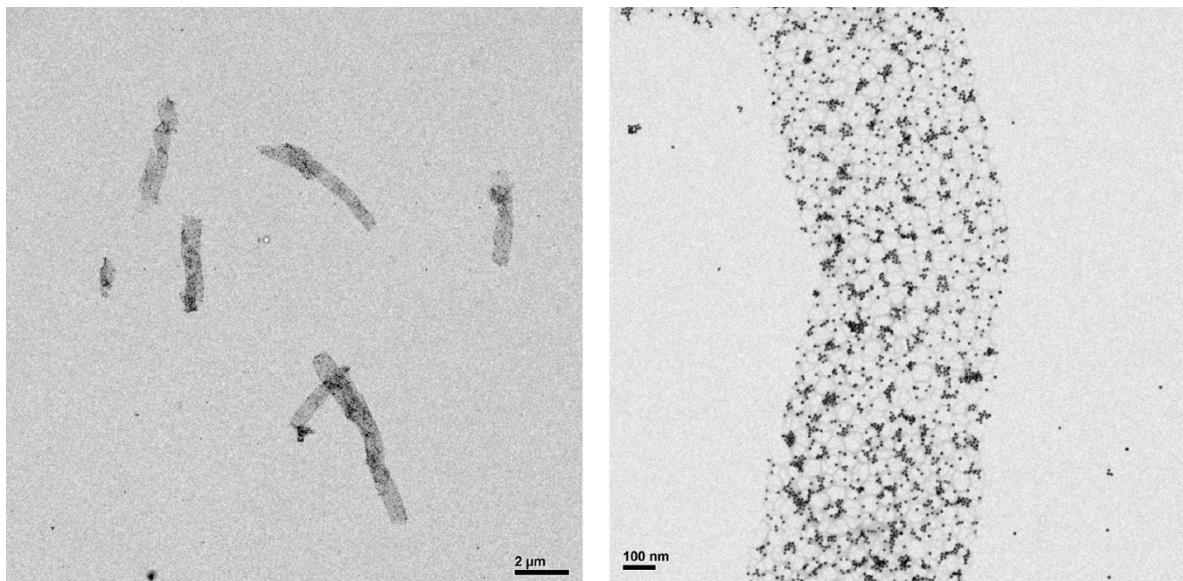


Figure S10. AuNP lattices on honeycomb tube with type-1 configuration.

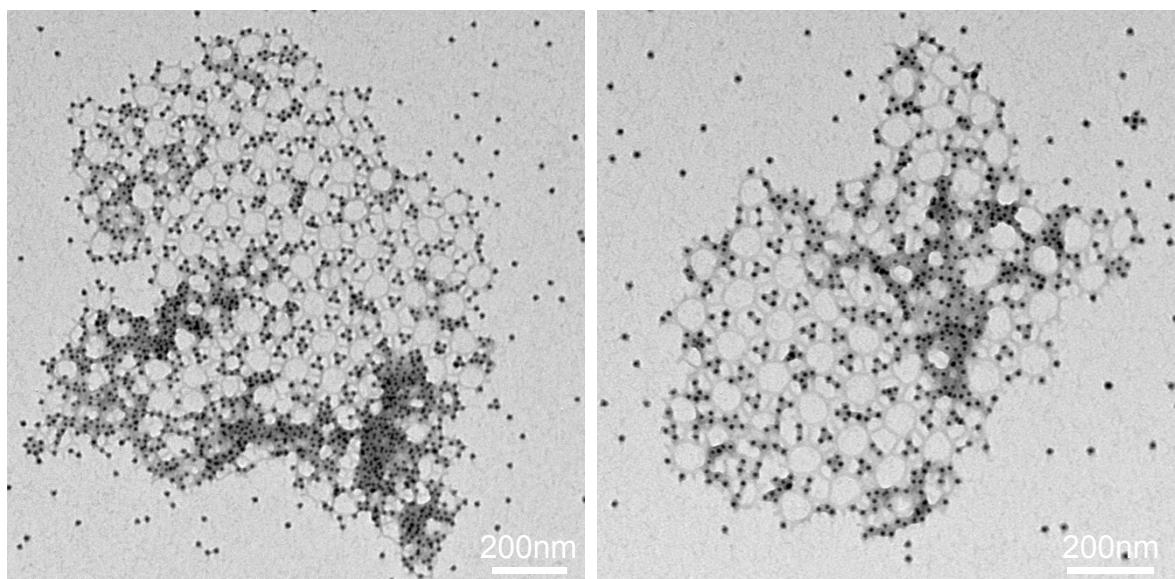


Figure S11. AuNP lattices on honeycomb array with type-2 configuration.

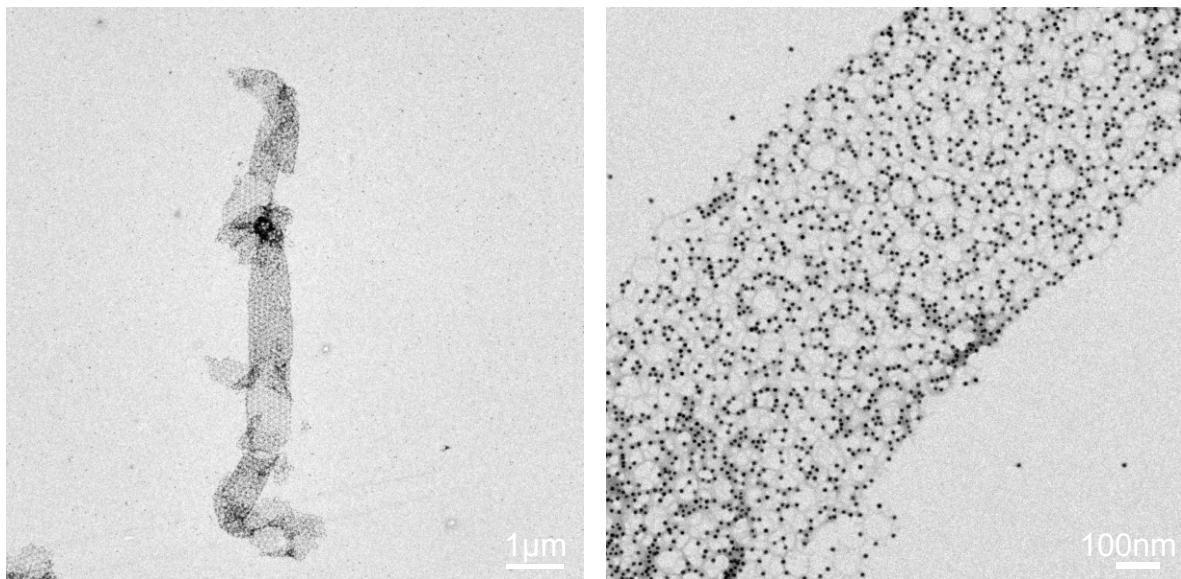


Figure S12. AuNP lattices on honeycomb tube with type-2 configuration.

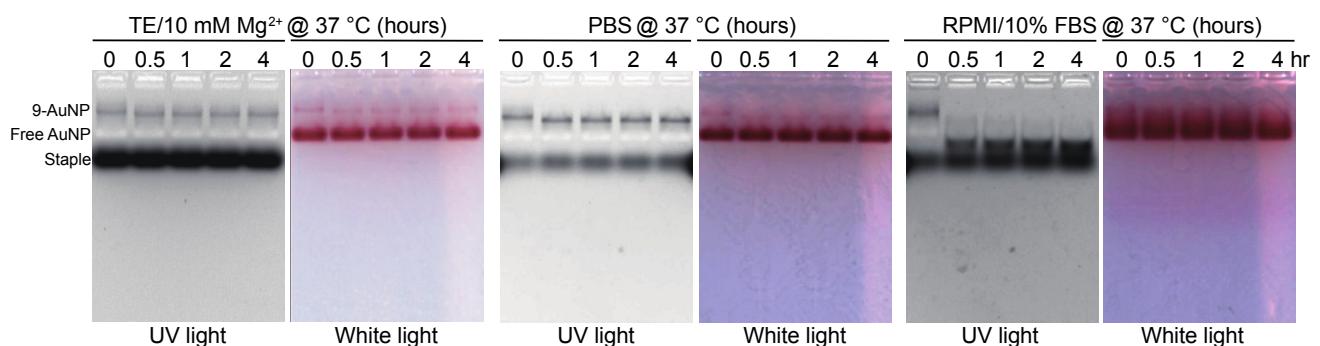


Figure S13. Stability of 9-AuNP clusters in various buffers and cell culture medium at 37 °C.

DNA strands sequences

p7560 scaffold DNA:

Thiolated DNA strands:

5'-ThioMC6-D/TTTTTTTTTTTTTTTTTT-3' for AuNP functionalization.

DNA strands of hexagon tile

Name	Sequence	Note
0[47]	ACAAATTTTGAAATGAGGTGAGGTCGCCATTAGCCAGCA	core
0[79]	CCTGAAAGCGTAAGGTTGGATTAGAGACTACCTTTAACCTCCGGCT	core
0[153]	GAATCCAACGTCAAAGAAGGCCTTCTGACCCCTGTTA	core
0[185]	GAACAAGAGTCCACAAGCAAGCCCAATGAAATAGCAATAGCTATCTTAC	core
0[259]	TCGGCAAAATCCCTTAATACATAAATTAAGACTAAGTTA	core
0[291]	AAAATCCTGTTGAGAGCCACCATAGGTGTATCACCGTACTCAGGAGGT	core
0[365]	CCTTCACCGCCTGGCCTTCCACAGGTAACACTCTTCCA	core
0[397]	ACCAGTGAGACGGGGAGGCTTGAATTCAACTTAAATCATTGTGAATT	core
0[471]	TGAATCGGCCAACCGCGTGCAGATATAGAAAGAACTATCAT	core
0[503]	GGGAAACCTGTCCTTGTGAGTAAAGATTCAAAAGGGTGAGAAAGG	core
0[577]	GGGGTGCCTAATGAGTGGTAATCGCAGGTATCAGAAAAG	core
0[609]	CGAGCCGGAAGCATCCAGGAAACAGGGCTTAAGCTACGTGGTCTTGT	core
0[636]	TATCCGCTCACACATAATCATTCCAGCAGA	core
I[32]	ATAGCCCTAAAGGGTTATATAACGGCACAG	core
I[72]	GATGCAAATAGGTCTGATACTTCTGAATAACGA	core
I[96]	AAGAACCGGAGAAAACCTTTCAAACCGTCTATCATCTGGCCAACAGAATTAAGAC	core
I[138]	TAGTTAATTCCCTTTAAGAAGAACGTG	core
I[178]	CCGAACAAGCAAGAAAGTTTATTTCATGCG	core
I[202]	ACCGAGGAAACGCAATAATAACCGAAGAATAGCCGAGATAGGGTTGAAGAGAGAT	core
I[244]	AAAGAACTGGCCCGCCACCCCTATCCGAAA	core
I[284]	AGAACCGAGCCCCGAACCGAACCGCCCTCCCGTT	core
I[308]	CACCTCATTTCAGGGATAGCAAGTTGCAGCAAGGGTCCACGCTGGTACCGGC	core
I[350]	GGAACCCATGTTGCGATTAAAGTATTG	core
I[390]	ACCAGTCAGATGGTTAGGACTAAAGACTTGAA	core
I[414]	AAATCTACGTTAATAAAACGAACACTGGCGTTGCGTATTGGGCGCCAGGCCCTGAC	core
I[456]	AACATTATTACACAGTCAAATCAGCATTAA	core
I[496]	TTCAACCAATGTGTAGTAAGAGGTATTTGCA	core
I[520]	TAATGCCGGAGAGGGTAGCTATTTCACATTAATTGCGTTGCGCTACAATTITTA	core
I[562]	TCTACAAAGGCATAAAGACGGAAAAGCCT	core
I[602]	CGAGCTCATGAGTAAAGCGCCATTGCCATTGA	core
2[20]	ACGAACCACTCCGAACTCTGATCTGTAAGCAA	core
2[48]	TAGGTTACACGGAATTCAATATA	core
2[87]	TCAAAATCATCCAATCCCTCTGA	core
2[126]	GTTTGAAAGAAGAGTCATAAGTAAATCGTCGC	core
2[154]	CGAACCATCAAACAAGTACCGCACTC	core
2[193]	ATAATAAGAAGTTACCCAGTTG	core
2[232]	CAGTATGTACAAGAATTGAGTAGACGGGAGAA	core
2[260]	TTAGTAATGAGGCATAATCAAATCA	core
2[299]	ATAAGTATCCACCCCTCAGCAGGCG	core
2[338]	CCAGTACAAGTGCCGTCGAGAAAGTATTAAGA	core
2[366]	ACCTTAACCACAGAACGAGGGTAGCA	core
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2[444]	AGATTAGACACCAGAACGAGAACCGGATATT	core
2[472]	CCGGAGAGGCATGATTAGAGAGTACC	core

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2[550]	CTGGAGCACTCATATATTAAAACATTATGA	core
2[578]	TACCTCTATTAAAGGCACCGCTTCTGG	core
2[617]	TTGGTAGAATTCTGTACAACATA	core
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3[374]	ACGGCTACACAAACAGCAACTGGCTCATTAT	core
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10[544]	CCCAATGAAACCATCGATAGCAGCAAAAGGGCGACAGTAGCACCATT	2D connector
11[16]	AAGAAGATGATGAAACAAATTGCGTAGATT	2D connector
11[122]	CAGTTACAAAATAACAGTATTCTAAGAACCC	2D connector
11[228]	GGCTTTGATGATACAGACCAGAACCAACCAG	2D connector
11[334]	GGAACGAGGCGCAGACGCAACCTAAACGAA	2D connector
11[440]	AAAGGTGGCATCAATTCAATGCAACTAAAT	2D connector
11[546]	GACTTAAGTGTCTTAGAGCTGGCAGAACCC	2D connector
4[112]	TTATCATTGCGGAACACAAACAATTGACCC	Tube connector
4[218]	CCTAATTACGAGCATGTCAGCTAATGCAGAG	Tube connector
4[324]	CTTAGCGTCAGACTGTAGCCGAAACGTCAA	Tube connector
4[430]	CGGTCGCTGAGGCTTGCAAAACAGCTTGATAA	Tube connector
4[536]	CGAAAGACTTCAAATATCCAGGTCTTACCCG	Tube connector

4[642]	TAACCGTGCATCTGCCAGTCTCCGTGGGAACA	Tube connector
5[113]	CAAGAGGCAAAAGACCACGCATAACCGATATA	Tube connector
5[219]	AGTACCGGTCTGGAAAAAGATTAAGAGGAAG	Tube connector
5[325]	GGGGATGTGCTGCATGGTAGATGGCGCAT	Tube connector
5[431]	TTTCAGGTTAACGTTAAAAGTTGAGTAA	Tube connector
5[537]	CGCGAGGCAGTACAAGAAAAATAATATCCC	Tube connector
5[643]	CCCAGAGCCGCCAGCGACAGAATCAAGTT	Tube connector
8[39]	AATAAAGAAACATCAACGAATTATTCAATTACCTGAG	Tube connector
8[145]	CTTATCCGGCCATATTAACGAGCGCTTCCAGAGCCTAATT	Tube connector
8[251]	GAGCCGCCAGTGTACAATTACCGTCCAGTAAGCGTCATA	Tube connector
8[357]	GAAGGCACGTCAATCAATCCCGCACCTGCTCCATGTTACTTA	Tube connector
8[463]	TGTTTAATACTAATAGCTATTTCAATTGGGGCGCGAGC	Tube connector
8[569]	ATTACGCCTGCTGAATCTTAAGTGGTTGTGAATTATGCG	Tube connector
10[14]	CCCGATAGTTGCCGACAATGACAACAGTTCAGCGTTGCTTCGA	Tube connector
10[120]	TTGACTATTATAGTCAGAACAAAGTGCAAAGAAGTGAGAACCA	Tube connector
10[226]	CAAACGGCGGATTGACCGTAATGGAATATTTGTTAGAGCGAGTAAC	Tube connector
10[332]	GAACTCGTATTAAATCCTTGCCGCCTCAATCAATAGGATTAGAAG	Tube connector
10[438]	TACGCCTGTTATCAACAATAGACAGTAGGGCTTAACGACGACAAT	Tube connector
10[544]	CCCAATGAAACCATCGATAGCAGCAAAAGGGCGACAGTAGCACCATT	Tube connector
11[15]	AAAGAAGATGATGAAACAATTGCGTAGATT	Tube connector
11[121]	CCAGTTACAAAATAACAGTATTCTAAGAAC	Tube connector
11[227]	TGGCTTGATGATACAGACCAGAACCCACAG	Tube connector
11[333]	CGGAACGAGGCCAGACGCAACCTAAACGAA	Tube connector
11[439]	AAAAGGTGGCATCAATTATGCAACTAAAT	Tube connector
11[545]	CGACTTAAGTGTCTTAGAGCTGGCGAAAGGC	Tube connector

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