## **Supporting Information**

#### Photo-triggered Release of Doxorubicin from Liposomes Formulated by

### Amphiphilic Phthalocyanines for Combination Therapy to Enhance

#### **Antitumor Efficacy**

Ke Zheng, <sup>\*a</sup> Hongyan Liu,<sup>a</sup> Xinxin Liu,<sup>a</sup> Libin Jiang,<sup>b</sup> Linlin Li,<sup>b</sup> Xianggen Wu,<sup>a</sup> Nannan Guo, <sup>b</sup> Caifeng Ding, <sup>\*a</sup> Mingdong Huang<sup>\*b</sup>

<sup>a</sup>Key Laboratory of Optic-electric Sensing and Analytical Chemistry for Life Science, Ministry of Education, Chemical Engineering College, Qingdao University of Science and Technology, Qingdao 266042, China

<sup>b</sup>College of Chemistry, Fuzhou University, Fujian 350116, China

\*Corresponding Author: Ke Zheng, Caifeng Ding, Mingdong Huang

E-mail: zhengke@qust.edu.cn, dingcaifeng@qust.edu.cn, hmd\_lab@fzu.edu.cn

Postal address: 53 Zheng Zhou Road, Qingdao, Shandong 266042, China



Figure S1. Ultraviolet-visible spectra of DOX and ZnPc(PEG)<sub>4</sub> in PBS or DMF.



Figure S2. The PDI variation curve of DOX@LiPOs, ZnPc(PEG)<sub>4</sub>@LiPOs and ZnPc(PEG)<sub>4</sub>:DOX@LiPOs

in PBS during five weeks.



**Figure S3.** The temperature variation curve of  $ZnPc(PEG)_4$  (A) and  $ZnPc(PEG)_4$ :DOX@LiPOs (B) with different concentrations (25µM, 50µM, 100µM, 200µM) in PBS solutions of 200µl under the laser (680 nm) irradiation of 100mW/cm<sup>2</sup> for 20min.

# The determination of singlet oxygen quantum yield and photothermal conversion efficiency for ZnPc(PEG)<sub>4</sub>:DOX@LiPOs

Singlet oxygen quantum yield ( $\Phi_{\Delta}$ ) of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs was determined using 1,3diphenylisobenzofuran as chemical quencher and methylene blue as the standard. Equation (1) was used to measure the  $\Phi_{\Delta}$ .

$$\phi_{\Delta} = \phi_{\Delta}^{Std} \, \frac{R \cdot I_{abs}^{Std}}{R^{Std} \cdot I_{abs}} \tag{1}$$

 $\Phi_{\Delta}^{std}$  was singlet oxygen quantum yield of methylene blue in D<sub>2</sub>O ( $\Phi_{\Delta}^{std}$ =0.52). Where R and R<sup>std</sup> was the photobleaching rate of the quencher in the presence of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs and methylene blue, respectively. I<sub>abs</sub> and I<sub>abs</sub><sup>std</sup> was the rate of light absorption by ZnPc(PEG)<sub>4</sub>:DOX@LiPOs and methylene blue, respectively.

Photothermal conversion efficiency of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs was determined according to the following equation (2).

$$\eta = \frac{hS(T_{MAX} - T_{surr}) - Q_{dis}}{I(1 - 10^{-A680})}$$
(2)

Where  $\eta$  represented the photothermal conversion efficiency, I was the laser power (100 mW/cm<sup>2</sup>), A680 was the absorbance of ZnPc (PEG)<sub>4</sub>:DOX@LiPOs at 680nm, T<sub>surr</sub> was ambient temperature, and T<sub>MAX</sub> was the maximum temperature of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs solution during the illumination, h was heat transfer coefficient, and S was the surface area of the container. The value of hS can be calculated by equation (3) and (4).

$$hS = \frac{mC_{water}}{\tau_s} \tag{3}$$

$$t = -\tau_{S} \ln \theta = -\tau_{S} \ln(\frac{T - T_{surr}}{T_{MAX} - T_{surr}})$$
(4)

Where m was the mass of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs solution,  $C_{water}$  was the heat capacity of solvent (4.2 J/g•°C).  $\tau_s$  was the system time constant, which was defined as the expression in equation (4).  $\tau_s$  was obtained from the linear correlation of cooling time versus -ln $\theta$  (Fig. S4).

Q<sub>dis</sub>, the energy consumption by the container, was measured according to equation (5) using container containing pure water under the same irradiation conditions.



**Figure S4.** A. Photothermal properties of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs in aqueous solution with 680nm light source at 100mW/cm<sup>2</sup>. The irradiation lasted for 900s and then was shut off. B. linear correlation of cooling time versus negative natural logarithm of driving force temperature.



**Figure S5.** The release percentage of DOX from DOX@LiPOs and ZnPc(PEG)<sub>4</sub>:DOX@LiPOs after heating to different temperature (35°C, 40°C, 45°C, 50°C, 55°C) for 30 minutes.



**Figure S6.** (A) Diameter distribution of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs after the irradiation (680 nm) for 20min at light dosage 100mW/cm<sup>2</sup>. (B) There was significant difference in PDI of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs before and after the irradiation. (C) The morphology of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs observed by TEM after the illumination for 20min at light dosage 100mW/cm<sup>2</sup>.



**Figure S7.** The fluorescence intensity profile in the subcellular localization detection of ZnPc(PEG)<sub>4</sub>:DOX@LiPOs (Fig. 5). (A) The distribution of DOX was consistent with that of ZnPc(PEG)<sub>4</sub>, and both of them did not enter the cell nucleus. (B) Through the illumination by LED light source (680nm), some DOX was separated from ZnPc(PEG)<sub>4</sub> and entered the nucleus. In addition, after the incubation with ZnPc(PEG)<sub>4</sub>:DOX@LiPOs for 24 hours, ZnPc(PEG)<sub>4</sub> could distributed in both mitochondria (C) and lysosomes (D) of cells.



**Figure S8.** After the mice were injected with ZnPc(PEG)<sub>4</sub>:DOX@LiPOs, ZnPc(PEG)<sub>4</sub>@LiPOs or DOX@LiPOs at the equal dosage of ZnPc(PEG)<sub>4</sub> (0.2 μmol/kg) or DOX (1.04 μmol/kg), PDT was administered for seven days with a 680nm light source at a light dose of 1 W/cm<sup>2</sup> for 3 min daily. After then, different tissues (heart, liver, spleen, kidney, lung and intestine) were harvested for H&E staining. Representative histopathological images (H&E, 200×) showed no significant pathological changes, demonstrating the safety of these liposomes *in vivo*.