Supporting Information for

Synthesis, molecular docking calculation, fluorescence and bioimaging of mitochondria-targeted ratiometric fluorescent probe for sensing hypochlorite *in vivo*

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1. Materials and methods

All reagents were obtained from commercial suppliers without further purification. All experiments used ultra-pure water. Solvents were purified by standard methods prior. Ultra-pure water is using by ULPURE. The pH measurements were performed with PHS-3E pH meter. UV-vis absorption spectra were obtained on a Shimadzu UV-2700 spectrophotometer, and fluorescence spectra were measured on a HITACHI F4700 fluorescence spectrophotometer. The fluorescence imaging of cells was performed with a Leica TCS SP8 CARS confocal microscope. CCK-8 was purchased from Fluorescence imaging experiments were performed with TransGen Biotechnology. TLC analysis was carried out on silica gel plates, and column chromatography was conducted over silica gel (mesh 200-300); both of them were purchased from the Qingdao Ocean Chemicals. ¹H and ¹³C NMR spectra were measured on a Varian Unity 600 spectrometer. High resolution mass spectrometric (HRMS) analyses were measured on Brooke solanX 70 FT-MS, Agilent 6540T.



Scheme S1. The synthetic route of Mi-OCI-RP.

Compounds 1, 2 and 3 were prepared by the literature procedure.¹ Synthesis of other compounds are described below.

Synthesis of compound 4. Compound **3** (100 mg, 0.16 mmol) was dissolved in C₂H₅OH (20 mL). Then the solution was added slowly into the ammonium hydroxide. The mixture solution was then stirred at 85 °C for 24 h. After cool to room temperature, the residue was purified by silica chromatography using CH₂Cl₂/PE (10:1, V/V) as the eluent, and compound **4** was obtained as dark red solid (48 mg, 50%). ¹H NMR (600 MHz, CDCl₃) δ 10.29 (s, 1H), 7.86 (d, *J* = 12.8 Hz, 1H), 7.27–7.21 (m, 3H), 7.20–7.15 (m, 1H), 6.97 (t, *J* = 7.4 Hz, 1H), 6.75 (d, *J* = 7.9 Hz, 1H), 5.56–5.46 (m, 1H), 5.32 (s, 1H), 3.30–3.19 (m, 4H), 2.63–2.60 (m, 2H), 2.56 (s, 1H), 2.52 (d, *J* = 6.1 Hz, 2H), 1.96–1.87 (m, 1H), 1.83 (d, *J* = 3.7 Hz, 2H), 1.80 (dd, *J* = 12.4, 6.2 Hz, 2H), 1.75 (d, *J* = 7.3 Hz, 2H), 1.69 (s, 5H), 1.33–1.26 (m, 1H), 0.91 (t, *J* = 6.9 Hz, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 190.85 (s), 162.92 (s), 148.70 (s), 144.45 (s), 139.20 (s), 131.15 (s), 128.65 (s), 127.90 (s), 127.64 (s), 127.61 (s), 125.72 (s), 123.34 (s), 121.77 (s), 121.65 (s), 120.89 (s), 119.33 (s), 117.78 (s), 106.78 (s), 105.64 (s), 92.93 (s), 92.50 (s), 53.45 (s), 46.44 (s), 45.67 (s), 44.79 (s), 29.41 (s), 28.32 (s), 26.70 (d), 24.61 (s), 20.95 (d). Anal. Calcd for C₃₂H₃₈IN₃: 591.58. Found: ESI-MS m/z ([C₃₂H₃₈IN₃] + H)⁺: 592.2372; ([C₃₂H₃₈IN₃] + Na)⁺: 614.2021.

Synthesis of the Probe Mi-OCI-RP. Compound **4** (50 mg, 0.084 mmol) was dissolved in DMF (5 mL). Then the solution was added slowly into the phenyl isothiocyanate. The mixture solution was then stirred at 50 °C for 24 h. After cooling to room temperature, the residue was purified by silica chromatography using CH₂Cl₂/MeOH (10:1, V/V) as the eluent, and the probe **Mi-OCI-RP** was obtained as dark red solid (15 mg, 24%). ¹H NMR (600 MHz, CDCl₃) δ 10.66 (d, *J* = 11.9 Hz, 1H), 7.89 (s, 1H), 7.80 (d, *J* = 11.4 Hz, 1H), 7.57 (d, *J* = 5.3 Hz, 3H), 7.47 (d, *J* = 7.5 Hz, 2H), 7.42 (d, *J* = 7.2 Hz, 2H), 7.35 (s, 3H), 7.24 (d, *J* = 7.4 Hz, 1H), 7.16 (s, 1H), 7.12 (s, 1H), 7.03 (dd, *J* = 16.1, 8.8 Hz, 2H), 4.09 (s, 3H), 2.97 (s, 2H), 2.76 (s, 2H), 1.89 (s, 2H), 1.79 (d, *J* = 16.7 Hz, 6H), 1.27 (s, 6H), 0.89 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 167.72 (s), 164.25 (d, *J* = 3.1 Hz), 142.83 (s), 141.04 (s), 140.40 (s), 140.07 (s), 138.79 (s), 136.86 (s), 135.41 (s), 129.57 (s), 129.40 (s), 129.18 (d, *J* = 5.0 Hz), 127.33 (s), 126.50 (s), 126.14 (s), 125.75 (s), 123.76 (s), 123.06 (s), 122.27 (s), 51.96 (s), 35.70 (s), 31.93 (s), 31.66 (s), 31.43 (s), 29.70 (s), 29.37 (s), 28.25 (s), 25.79 (s), 24.75 (s), 22.70 (s), 21.68 (s). Anal. Calcd for C₃₉H₄₃IN₄S: 726.77. Found: ESI-MS m/z ([C₃₉H₄₃IN₄S] + H)⁺: 727.2386.

3. Cell culture and cytotoxicity assays

A549 cells, MCF-7 cells and RAW264.7 were cultured in DMEM or 1640 (Dulbecco's modified Eagle's medium or Roswell Park Memorial Institue medium) supplemented with 10% FBS (fetal bovine serum) in an atmosphere of 5% CO₂ and 95% air at 37 °C. In vitro cytotoxicity was measured using CCK-8 assay on A549 cells, MCF-7 cells and RAW264.7. These Cells were seeded into a 96-well tissue culture plate in the presence of completed Dulbecco's modified Eagle's medium (DMEM) or Roswell Park Memorial Institue medium (1640) supplemented with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin at 37 °C and 5% CO₂ atmosphere overnight followed by incubation for 24 h in the presence of **Mi-OCl-RP** at different concentrations (5, 10, 20, 30, 40 μ M). A commercial cell counting kit-8 (CCK-8) (TransGen Biotechnology, China) was used to detecting the cell viability and the assay was run following the manufactures' instructions. The cell viability was determined by assuming 100% cell viability for cells without **Mi-OCl-RP**.

Reagents	Molecular calculation	Response time	Stokes shift	Ratio imaging	Reference
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	No	-	27 nm	No	2
	No	-	104 nm	No	3
	No	< 3 min	122 nm	No	4
	No	< 10 s	15 nm	No	5
	No	< 5 s	72 nm	No	6
$\overbrace{Et_2N} \xrightarrow{-CO_2H} \xrightarrow{O} \xrightarrow{NEt_2}$	No	< 3 min	200 nm	No	7
	No	1 min	45 nm	No	8
$ \begin{array}{c} & \overset{NC}{\longrightarrow} \overset{CN}{\underset{N}{\longrightarrow}} \\ & \overset{S}{\longrightarrow} \overset{CN}{\longrightarrow} \overset{R}{\underset{R}{\longrightarrow}} \\ \end{array} $	No	$\leq 8 s$	52 nm	No	9
Julia Contraction of the second secon	No	120 s	230 nm	Yes	10
	No	< 40 s	209 nm	Yes	11
	Yes	< 7 s	278 nm	Yes	This work

Table S1 Literature on mitochondria-targeted	OCl ⁻ -sensitive	probes and	our probe.
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**Figure S1** Molecular binding modeling of conformation 2-10. Tertiary structure of ANT, actives site and residues of (2a-10a). The  $\pi$ -cation and  $\pi$ - $\pi$  interactions is indicated by golden block.



**Figure S2** Linear relationship of fluorescence intensity of **Mi-OCI-RP** in different concentration of OCI⁻ (0-10  $\mu$ M) (a:  $\lambda_{ex} = 373$  nm; b:  $\lambda_{ex} = 588$  nm).



**Figure S3** Fluorescence intensity of **Mi-OCI-RP** (10  $\mu$ M) with OCI⁻ and various relevant species (10  $\mu$ M): Hydroxyl radicals; Cys; Hcy; Glutathione; CH₃COOOH; H₂O₂; t-butylhydroperoxide; NO; Ca²⁺; Zn²⁺; Co²⁺; Cu²⁺; Fe²⁺; Mg²⁺; F⁻; Cl⁻; Br⁻; I⁻; HCO₃⁻; SO₃²⁻; HSO₃⁻; and NO₂⁻ in pH 7.4 PBS buffer (a:  $\lambda_{ex} = 373$  nm; b:  $\lambda_{ex} = 588$  nm).



**Figure S4** Fluorescence spectra stability of 10  $\mu$ M **Mi-OCI-RP** in pH 7.4 PBS buffer (a:  $\lambda_{ex} = 373$  nm; b:  $\lambda_{ex} = 588$  nm).



**Figure S5** Fluorescence spectra stability of 10  $\mu$ M **Mi-OCl-RP** in the presence of OCl⁻ (10  $\mu$ M) in pH 7.4 PBS buffer (a:  $\lambda_{ex} = 373$  nm; b:  $\lambda_{ex} = 588$  nm).



**Figure S6** pH dependency of **Mi-OCl-RP** (10  $\mu$ M) in the absence or presence of OCl⁻ (10  $\mu$ M) in PBS buffer (a:  $\lambda_{ex} = 373$  nm; b:  $\lambda_{ex} = 588$  nm).



**Figure S7** Cytotoxicity of A549 cells, MCF-7 cells and RAW264.7 measured CCK-8 assay after treatment with different concentrations of **Mi-OCI-RP**.



**Figure S8** Confocal fluorescence images of **Mi-OCI-RP** responding to OCI⁻ in MCF-7 cells. (1a-1c) MCF-7 cells incubated with **Mi-OCI-RP** (10  $\mu$ M, 20 min); (2a-4c) MCF-7 cells incubated with OCI⁻ (5, 8 and 10  $\mu$ M) for 10 min and then incubated with **Mi-OCI-RP** (10  $\mu$ M, 20 min). First column: ratios images of excitation at 405 and 588 nm; second column: excitation at 588 nm; third column: excitation at 405 nm. (5a-5c) Quantified relative fluorescence intensity of images 1a-4c. Scale bar: 10  $\mu$ m.



Figure S10¹³C NMR spectra of compound 2 in CDCl₃.



Figure S11 HRMS of compound 2.







Figure S13 ¹³C NMR spectra of Mi-OCI-RP in CDCl₃.



Figure S14 HRMS of Mi-OCI-RP.

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