

Supplementary Information

A Rapid Variant-Tolerant Reverse Transcription Loop-Mediated Isothermal Amplification Assay for Point of Care Detection of HIV-1

Yingxue Li,^{a, b, c, ‡} Xin Chen,^{d, ‡} Yongjuan Zhao,^{a, ‡} Zhenzhou Wan,^e Yi Zeng,^a Yingying Ma,^a Lianqun Zhou,^{b, c} Gaolian Xu,^f Julien Reboud,^g Jonathan M. Cooper,^g Chiyu Zhang^{a,*}

Affiliations:

^a Shanghai Clinical Research Center for Infectious Disease (HIV/AIDS), Shanghai Public Health Clinical Center, Fudan University, Shanghai 201508, China

^b CAS key Laboratory of Bio-medical Diagnostics, Suzhou Institute of Biomedical Engineering and Technology, Chinese Academy of Sciences, Suzhou 215163, China

^c School of Biomedical Engineering, University of Science and Technology of China, Hefei 260026, China

^d Department of Pathogenic Biology, School of Basic Medical Sciences, Gannan Medical University, Ganzhou 341000, China

^e Medical Laboratory of Taizhou Fourth People's Hospital, Taizhou 225300 China.

^f Nano Biomedical Research Centre, School of Biomedical Engineering, Shanghai Jiao Tong University, Shanghai 200030, China

^g Division of Biomedical Engineering, University of Glasgow, G12 8LT Glasgow, United Kingdom

‡ These authors contributed equally to this work.

* Corresponding authors:

Chiyu Zhang, Shanghai Public Health Clinical Center, Fudan University, Shanghai 201508, China.

Email: zhangcy1999@shphc.org.cn

Table S1. Primers and probes used in the study.

Aim	Primers	Sequence (5'-3')	Ref.
Mutant construction	F3-Mu-A(F)	CACTTGAAAGGACCAGAAAAGCTCCTCT	This study
	F3-Mu-A(R)	T TGGTCCTTCCAAAGTGGATTCTGCTGT	
	F3-Mu-G(F)	CACTTGAAAGGACCAGGAAAGCTCCTCT	
	F3-Mu-G(R)	C TGGTCCTTCCAAAGTGGATTCTGCTGT	
	F1-Mu-A(F)	GGGGCAGTAGTAATACAAATAATAGTGAC	
	F1-Mu-A(R)	T TTGTATTACTACTGCCCTCACCTTCC	
	F1-Mu-C(F)	GGGGCAGTAGTAATACAAACATAATAGTGAC	
	F1-Mu-C(R)	C TTGTATTACTACTGCCCTCACCTTCC	
	LB-Mu-A(F)	ATGATTGTGGCAAGTAACAGGATGAGG	
	LB-Mu-A(R)	T ACTTGCCACACAATCATCACCTGCCATC	
	LB-Mu-C(F)	ATGATTGTGGCAAGTAACAGGATGAGG	
	LB-Mu-C(R)	G TACTTGCCACACAATCATCACCTGCCATC	
	FIP-Mu-A(F)	AAGCTCCTCTGGAAAGGTAAAGGGCAGTA	
	FIP-Mu-A(R)	CCTTCCAGAGGAGCTTGCTGGCCTTT T	
	FIP-Mu-C(F)	AAGCTCCTCTGGAAAGGTCAAGGGGCAGTA	
	FIP-Mu-C(R)	CCTTCCAGAGGAGCTTGCTGGCCTTT G	
	FIP-Mu-2G(F)	AAAGCTCCTCTGGAAAGGGGAAGGGCAGT	
	FIP-Mu-2G(R)	C CCTTCCAGAGGAGCTTGCTGGCCTTT	
	FIP-Mu-2C(F)	AAAGCTCCTCTGGAAAGGCAGAAGGGCAGT	
	FIP-Mu-2C(R)	G CCTTCCAGAGGAGCTTGCTGGCCTTT	
	FIP-Mu-3A(F)	CAAAGCTCCTCTGGAAAGATGAAGGGCAG	
	FIP-Mu-3A(R)	T CTTCCAGAGGAGCTTGCTGGCCTTT	
	FIP-Mu-3C(F)	CAAAGCTCCTCTGGAAAGCTGAAGGGCAG	
	FIP-Mu-3C(R)	G CCTTCCAGAGGAGCTTGCTGGCCTTT	
	FIP-Mu-5T(F)	AGCAAAGCTCCTCTGGAA T GGTGAAGGGC	(PLoS One. 2015;10(2):e0 117852)
	FIP-Mu-5T(R)	A TTCCAGAGGAGCTTGCTGGCCTTTCCA	
	FIP-Mu-5G(F)	AGCAAAGCTCCTCTGGAA G GGTGAAGGGC	
	FIP-Mu-5G(R)	C TTCCAGAGGAGCTTGCTGGCCTTTCCA	
RT-LAMP assay of HIV-1	AceIN-F3	CCMMTTGGAAAGGACCAGC	
	AceIN-B3b	AACATACATATGRTGYTTACTA	
	AceIN-B3a	TCTTGAAAYATACATATGRTG	
	AceIN-FIPf	CTTGGCACTACYTTATGTCACTAAARCTYCTCT	
		GGAAAGGTG	
	AceIN-FIPE	CTTGGTACTACYTTATGTCACTAAARCTACTCT	
		GGAAAGGTG	
	AceIN-BIP	GGAYTATGGAAAACAGATGGCAGCCATGTTCT	
		AATCYTCATCCTG	
	AceIN-LF	TCTTGTATTACTACTGCCCTT	
	AceIN-LB	GTGMTGATTGTGGCARGTAG	

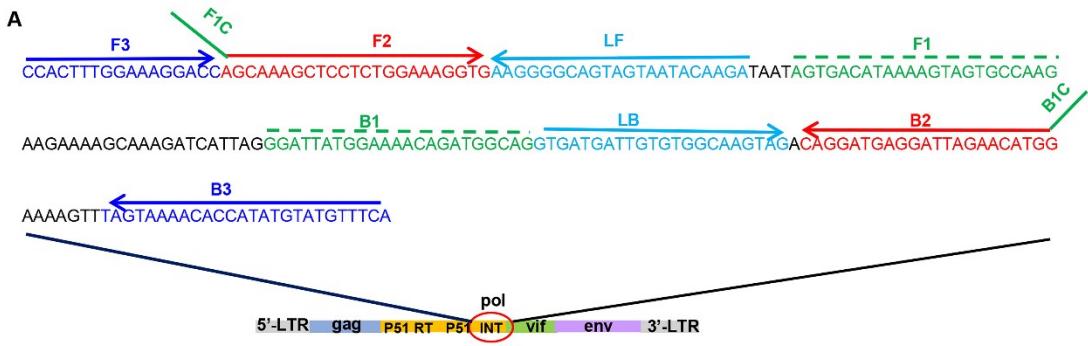
	AceIN-F3	CCMMTTGGAAAGGGACCAGC
RT-qPCR assay	HIV-1 F	CAATTTAAAAGAAAAGGGGGATTG
of HIV-1	HIV-1 R	TAAACCCGAAAATTTGAATT
	HIV-1 P	FAM-ACAGTGCAGGGAAAG-MGB

The artificially introduced mutations are highlighted in red.

Table S2. Detection rates of different HIV-1 serotypes by the conventional and the novel mismatch-tolerant RT-LAMP assays.

HIV-1 subtype	The conventional RT-LAMP			The mismatch-tolerant RT-LAMP			total
	Positive	Negative	Detection rate (%)	Positive	Negative	Detection rate (%)	
B	3	1	75	3	1	75	4
C	1	1	50	1	1	50	2
01_AE	2	4	33.3	4	2	66.7	6
07_BC	0	1	0	0	1	0	1
08_BC	2	1	66.7	3	0	100	3
57_BC	1	1	50	2	0	100	2
87_cpx	0	2	0	0	2	0	2
96_cpx	0	1	0	1	0	100	1
62_BC	0	1	0	0	1	0	1
65_cpx	1	0	100	1	0	100	1
URF	7	14	33.3	14	7	66.7	21
Total	17	27	38.6	29	15	65.9	44

Note: The detection rate was calculated using the formula: (number of positive results/total number) ×100%.



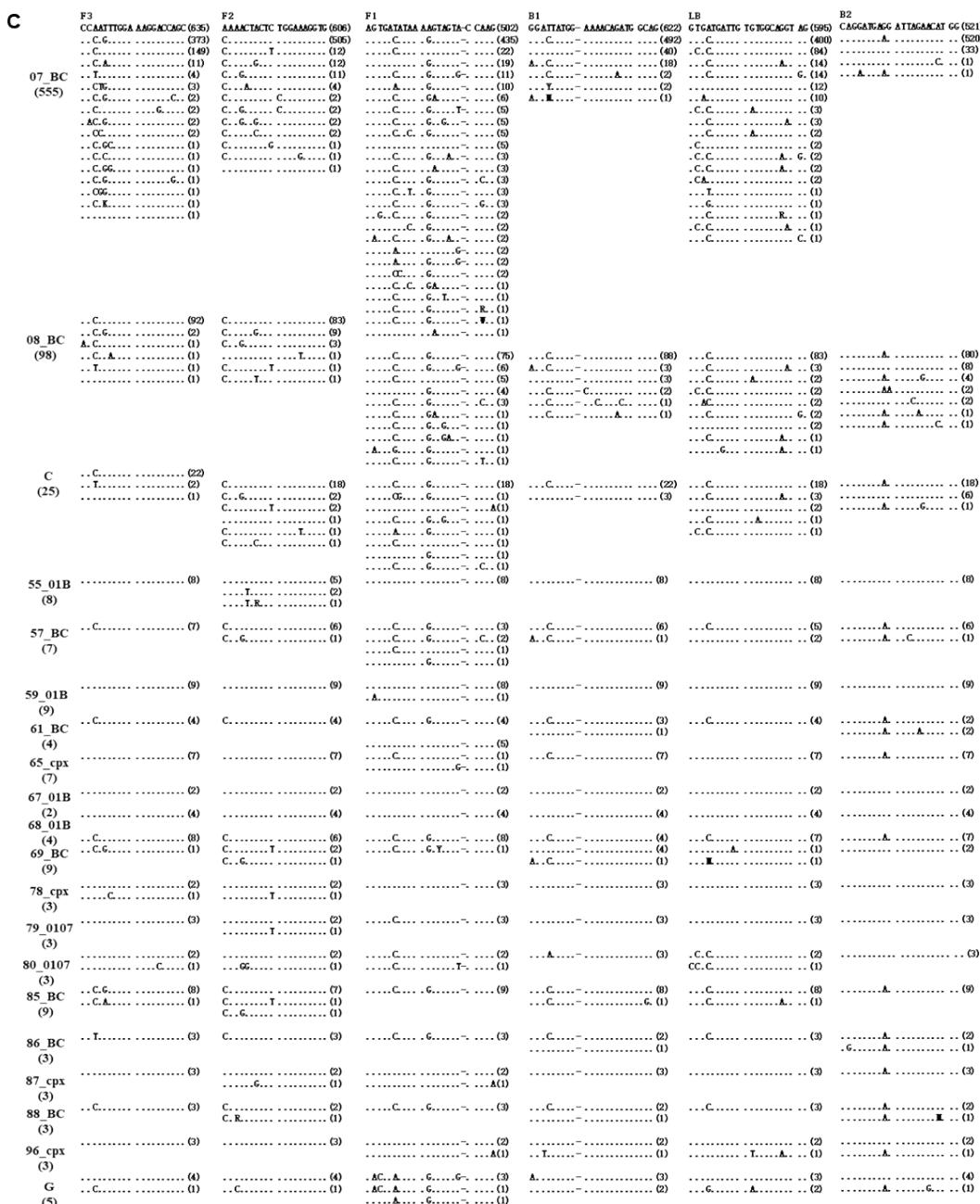


Figure S1. Locations (A) and sequence alignments (B and C) of all available HIV-1 strains. A total of 661_01_AE, 322_B, 555_07_BC, 98_08_BC, 25_C, 8_55_01B, 7_57_BC, 9_59_01B, 4_61_BC, 7_65_cpx, 2_67_01B, 4_68_01B, 9_69_BC, 3_78_cpx, 3_79_0107, 3_80_0107, 9_85_BC, 3_86_BC, 3_87_cpx, 3_88_BC, 3_96_cpx, and 5_G HIV-1 sequences were downloaded from HIV Database (<https://www.hiv.lanl.gov/content/sequence/HIV/mainpage.html>) on April 10, 2019. The number of identical sequences is shown in parenthesis following each unique sequence. Dot, identity with the topmost sequence; Dash, deletion.

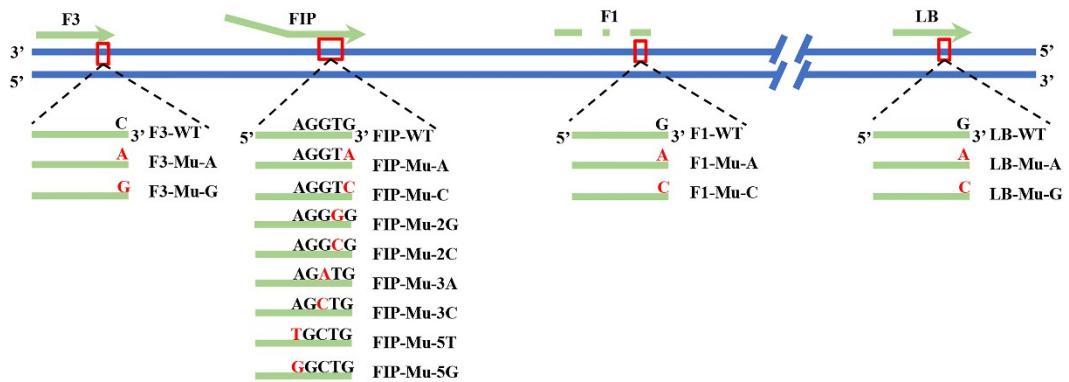


Figure S2. Schematic diagram of different HIV mutation templates. WT, wild-type; Mu, mutant.

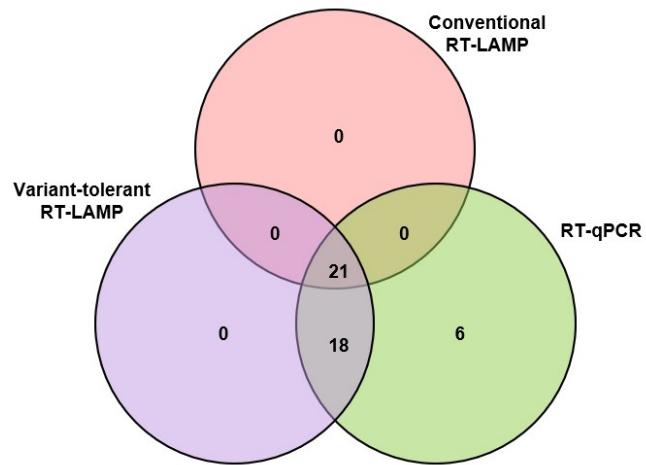


Figure S3. Numbers of HIV-1 positive samples detected by three different assays.

Primer	F3	F2	LF	F1	B1	LB	B2	B3
	CCMMTTGGA AAGGACCAGCAAARCTYCTC	TGGAAAGGAG AAGGGCAGT AGTAATACAA GA	AGTGACATAA ARGTAGTGCC AAG	GGATTATGGA AACAGATGG CAG	GTGAGTATG TGTCGCAGT AG	CAGGATGARG ATTAGAACAT GG	TAGTAAARCA YCATATGTAT RTCTCAAAGA	
4838 (01_AE)	A. AA.....	C. A. A.... G. T.	T. A.	A. G. G. AT. T. G.
4780 (01_AE)	.. AC.....	G. C. T.... G. T.	G.	C. G. G. . T. A. G.
4774 (57_BC)	A. AA.....	C. A. A.... G. T.	C. G. A. AG. T. A. C. G. T.
4799 (96_cpx)	.. TA. C.	C. A. A.... G. T.	C. G. A. AG. T. A. C. G. T.
4663 (B)	A. AA.....	C. A. A.... G. A. T.	A. G. A. T. G.
4635 (08_BC)	.. CA.....	C. A. A.... G. A. T.	C. A. A. A. C. G.
4624 (URF)	.. CA.....	C. A. A.... G. A. T.	C. G. A. A. A. C. A. T.
4651 (URF)	.. AA.....	A. A.... G. T.	A. G. A. T. G.
4662 (URF)	.. AA.....	A. A.... G. T.	A. G. A. T. G.
4665 (URF)	A. AA.....	AT. G.... G. T.	A. G. A. T. G. G.
4808 (URF)	A. AA.....	C. A. A.... G. A.	A. G. A. T. C. C. A.
4825 (URF)	.. CA.....	C. A. A.... G. T.	A. G. G. A. C. G. T.
4867 (URF)	A. AA.....	C. A. A.... G. A. T.	C. A. A. A. C. A. T.

Figure S4. Alignment analysis of primer sequences for clinical samples. A total of 13 samples were analyzed, including seven cases of URF (marked in red), two cases of 01_AE, one case each for 57_BC, 96_cpx, B, and 08_BC, and the black triangle indicates the mutation site. Dot, identity with the topmost sequence; Dash, deletion.

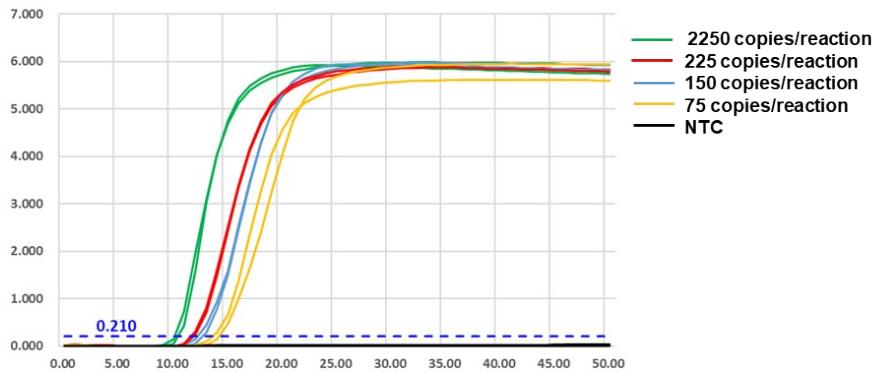


Figure S5. Mismatch-tolerant RT-LAMP detection by rapidly magnetic bead-based RNA extraction. The threshold is determined as 3 standard deviation from the background (0.210, dotted blue line).

NTC: no template control.

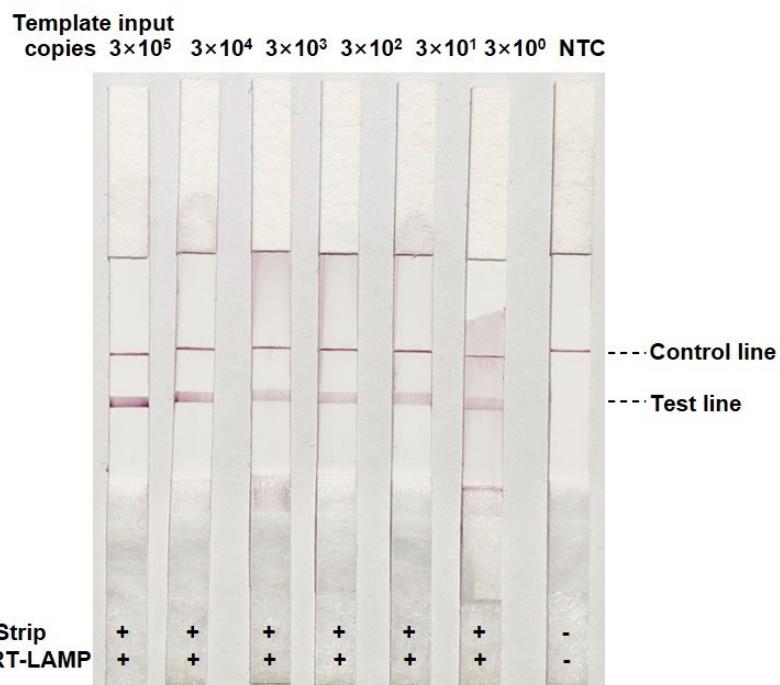


Figure S6. Lateral flow strip test of the sensitivity of RT-LAMP products. A single red line (control line) indicates a negative result, whilst two single red lines (control and test lines) indicate a positive result. ‘+’: positive results; ‘-’: negative results; Pos: positive control; NTC: no template control; Control line: represents the control result; Test line: represents the test result.

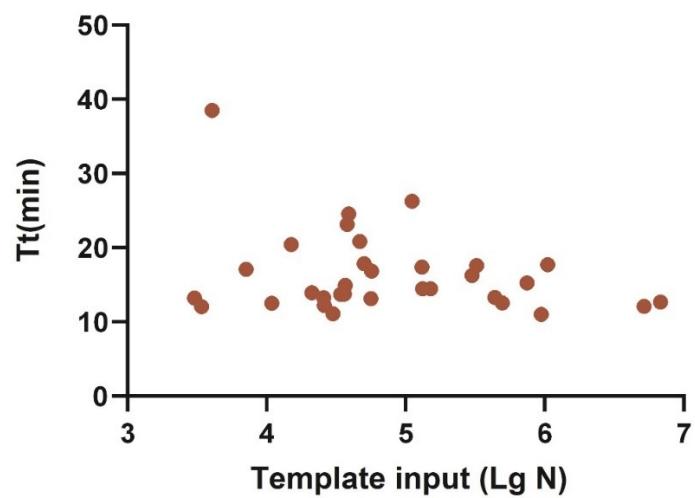


Figure S7. Correlation between the number of copies measured in 32 positive clinical samples and the novel RT-LAMP detection time. Tt: time threshold.

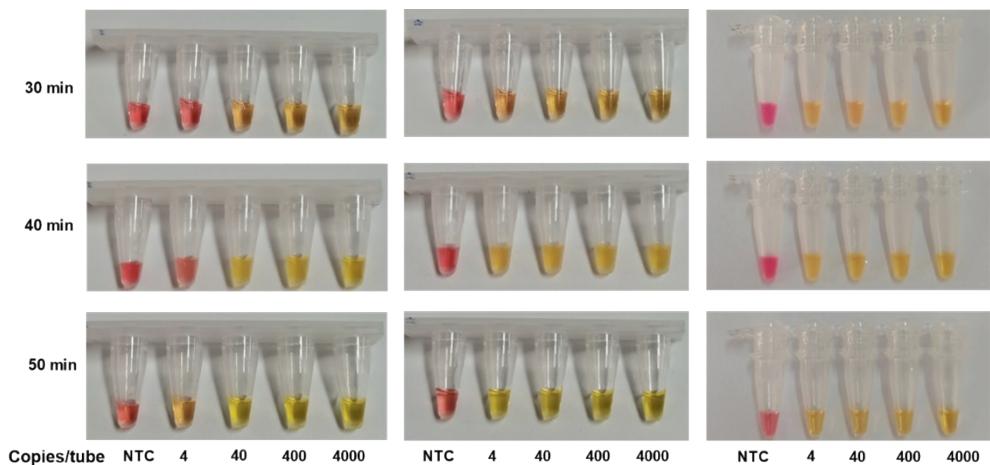


Figure S8. Visual detection of HIV-1 using the novel mismatch-tolerant RT-LAMP assays with

cresol red. The color change from burgundy to orange or yellow is considered as positive. NTC: no template control.