

SUPPORTING INFORMATION

An artificial enzyme cascade amplification strategy for highly sensitive and specific detection of breast cancer-derived exosomes

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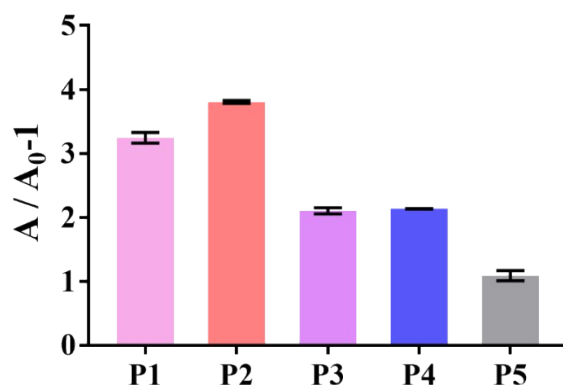


Fig. S1 Optimization of the G-quadruplex mimicking DNAzyme sequence by comparing the relative signal generated from five common G-quadruplex mimicking DNAzyme sequence. A is the absorbance at 420 nm in the presence of exosomes and A₀ is the absorbance at 420 nm in the absence of exosomes. Error bars show the standard deviation of three experiments.

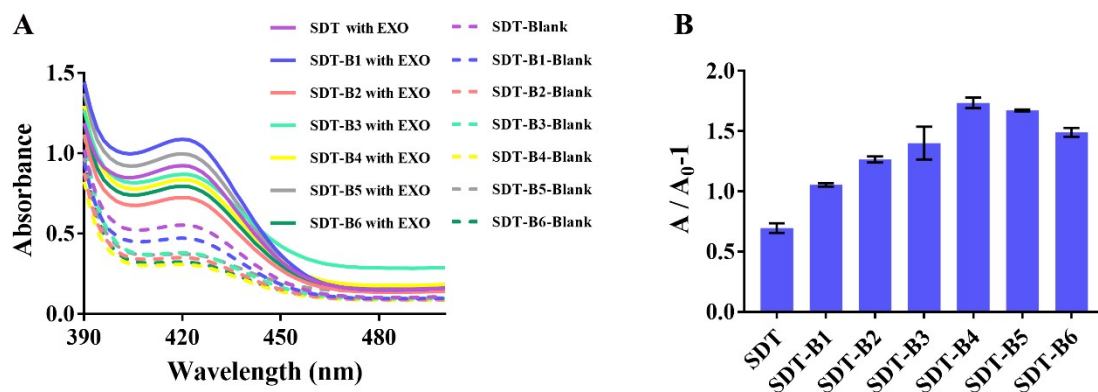


Fig. S2 Optimization of the switch strands. (A) Absorption spectra of the SDT sensing scaffold coupled with different switch strands (B1, B2, B3, B4, B5, and B6) in the presence and absence of exosomes. (B) The relative signal of the SDT sensing scaffold coupled with different switch strands.

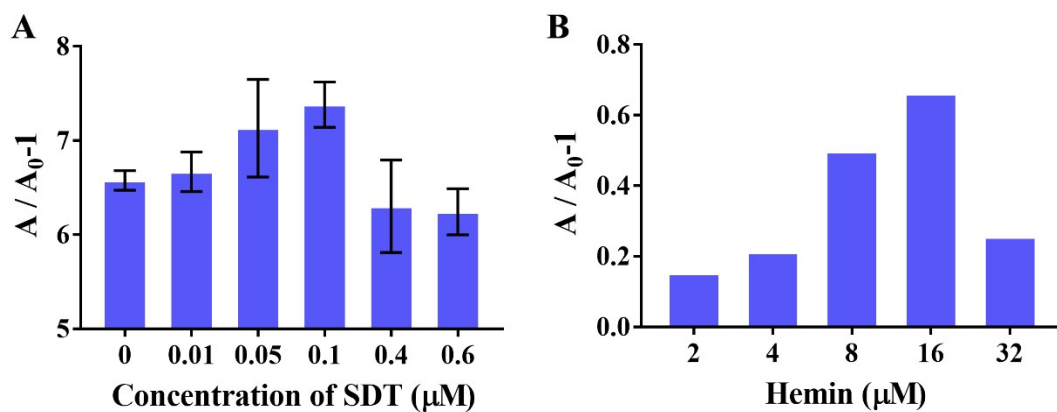


Fig. S3 (A) The relative signal of different concentration of SDT for detection of MCF-7 exosomes. (B) The relative signal of different concentration of hemin for detection of MCF-7 exosomes.

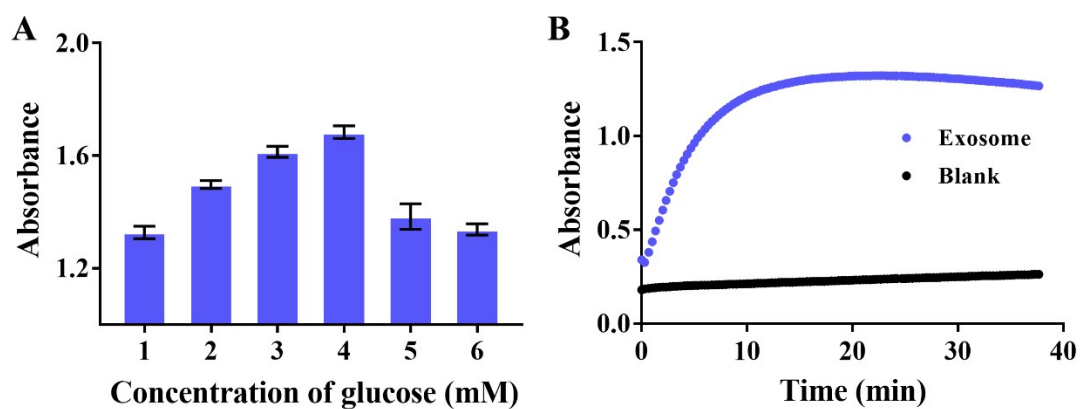


Fig. S4 (A) The relative signal of the SDT sensing scaffold at 420 nm after incubated with different concentration of glucose. (B) Time-course absorbance monitoring of the SDT sensing scaffold after adding all the reagents in the presence and absence exosomes.

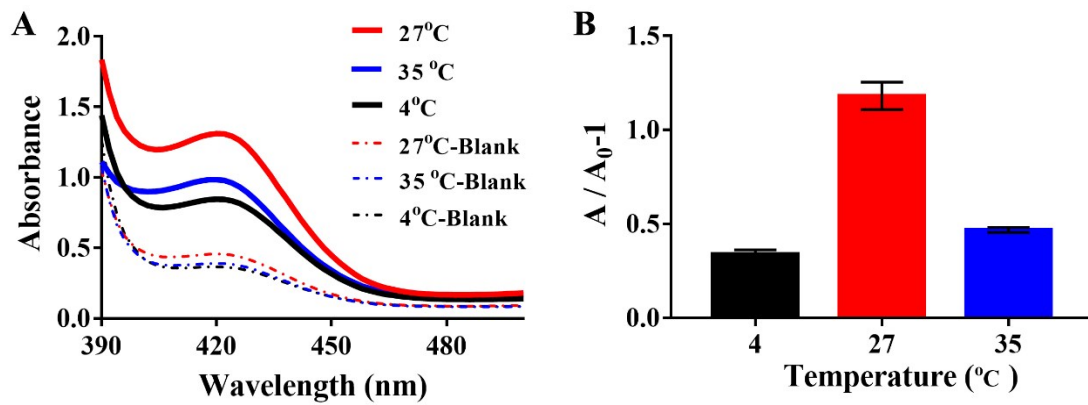


Fig. S5 Influence of temperature (4, 27, and 35°C) on the formation of G-quadruplex. (C) Histogram of absorbance change ratio (A/A_0-1) for the different reaction temperature.

2 Supporting Tables

Table S1. Sequence of oligonucleotides designed in this study^a

DNA	Sequence (from 5' to 3')
P1	GGG TTG GGC GGG ATG GGG GTA TCCA
P2	TTT GGG TAG GGC GGG TTG GGG GTA TCC A
P3	GGG TAG GGC GGG TTG GGG GTA TCC A
P4	CTG GGA GGG AGG GAG GGA GGT ATC CA
P5	ACC TGG GGG AGT ATT GCG GTA TCC A
A76	TCAACTTTGGGTAGGGCGGGTTGGGGGTATCCATGATAAAACGACACTACGT GGGAA TCTACTATGGCGGCTCTTC
B76	TTCAGTTTGGGTAGGGCGGGTTGGGGGTATCCAAATGTGCTTCCACGTAGT GTCGTTTGTATTGGACCCTCGCAT
C59	CTGAA TTTTTT ATTACAGCTTGCTACACTTTTTGAAGAGCCGCCATAGTATTTTTA CATT
D59	GTTGA TTTTTT GTGTAGCAAGCTGTAATTTTTATGCGAGGGTCCAATACTTTTTT ATCA
B1	<u>TTGATCCTTTGGATACC</u> CCC AAC
B2	<u>TTGATCCTTTGGATACC</u> CCC AAC CCG
B3	<u>TTGATCCTTTGGATACC</u> CCC AAC CCG CCC
B4	<u>TTGATCCTTTGGATACC</u> CCC AAC CCG CCC TAC
B5	<u>TTGATCCTTTGGATACC</u> CCC AAC CCG CCC TAC CCA
B6	<u>TTGATCCTTTGGATACC</u> CCC AAC CCG CCC TAC CCA AA

^a Bold bases are the G-quadruplex sequence, and underlined bases are MUC1 aptamer.

Table S2. Comparison of our method and reported methods for exosomes detection.

Platform	Linear range (particles/mL)	LOD (particles/mL)	Method	Assay time	Reference
B-Chol anchor assay with enzyme-linked HCR	$2.3 \times 10^6 - 2.3 \times 10^8$	2.2×10^6	Colorimetric	~16.5 h	1
large-AR AuNBP@MnO ₂ NSs	$8.5 \times 10^5 - 8.5 \times 10^7$	1.35×10^5	Colorimetric	~2.5 h	2
Aptamer-based DNA nanodevices	$10^6 - 10^{11}$	10^6	Fluorescence	-	3
“on-off”-type aptasensor	$1.0 \times 10^8 - 1.6 \times 10^9$	4.2×10^7	Fluorescence	32 min	4
Aptasensor based on g-C ₃ N ₄ nanosheets	$1.9 \times 10^9 - 3.38 \times 10^{10}$	1.35×10^9	Colorimetric	~30 min	5
HRP-pSC ₄ -AuNPs@COFs	$5 \times 10^5 - 1.5 \times 10^{10}$	1.6×10^5	electrochemical	~3 h	6
Switchable DNA tetrahedral scaffolds	$3.8 \times 10^6 - 1.2 \times 10^8$	1.51×10^5	Colorimetric	~2 h	This work

REFERENCES

- 1 F. He, H. Liu, X. Guo, B.-C. Yin and B.-C. Ye, *Anal. Chem.*, 2017, **89**, 12968-12975.
- 2 Y. Zhang, J. Jiao, Y. Wei, D. Wang, C. Yang and Z. Xu, *Anal. Chem.*, 2020, **92**, 15244-15252.
- 3 D. He, S.-L. Ho, H.-N. Chan, H. Wang, L. Hai, X. He, K. Wang and H.-W. Li, *Anal. Chem.*, 2019, **91**, 2768-2775.
- 4 J. Zhang, J. Shi, W. Liu, K. Zhang, H. Zhao, H. Zhang and Z. Zhang, *Sens. Actuators B Chem.*, 2018, **276**, 552-559.
- 5 Y.-M. Wang, J.-W. Liu, G. B. Adkins, W. Shen, M. P. Trinh, L.-Y. Duan, J.-H. Jiang and W. Zhong, *Anal. Chem.*, 2017, **89**, 12327-12333.
- 6 M. Wang, Y. Pan, S. Wu, Z. Sun, L. Wang, J. Yang, Y. Yin and G. Li, *Biosens. Bioelectron.*, 2020, **169**, 112638.