Localized Surface Plasmon Resonance Aptasensor for Selective Detection of SARS-CoV-2

S1 Protein

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Fig. S1. SEM images of LSPR chip at various magnifications. The chip surface contains biotin film on gold nanoparticles.



Fig. S2. Absorbance spectrum of LSPR Biotin-Au chip.



Fig. S3. Sensorgram showing fabrication process involving streptavidin protein loading to the biotin-gold chip followed by immobilization of S1 and S1-T aptamers on two different channels ([streptavidin] = 0.5μ M; [S1 aptamer] = 50μ g/mL; [S1 aptamer-T] = 50μ g/mL; flow rate = 20μ L/min).



Fig. S4. Signal response after immobilization of N, S1- and S1-T aptamer on individual channels with minimum target values provided for each ([Aptamer] = $50 \mu g/mL$).



Fig. S5. Signal response after immobilization of S1 aptamer with 3 subsequent aptamer injections ([Aptamer] = $16 \mu g/mL$).



Fig. S6. Representative sensorgram of the S1 protein binding (1) followed by the regeneration optimization with subsequent injections of 0.5% SDS buffer (2-3) ([S1 Protein] = 33 nM (2.5 μ g/mL), regeneration flow rate = 150 μ L/min).



Fig. S7. (A) Representative sensorgram of fitted data (black lines) for varying concentrations of S1 protein (colored lines) using the S1 aptasensor ([S1 protein] = 32 pM to 131 nM). (B) LSPR signal change as a function of S1 protein concentration determined from sensorgram (A) ([S1 protein] = 0 to 131 nM).



Fig. S8. Comparison of LSPR signal responses for S1 protein using the S1 Aptamer-T or the S1 Aptamer ([Aptamer] = $50 \mu g/mL$, data represent average and error bars of triplicate measurements).

Protein	k _a (1/M*s)	<i>k</i> _d (1/s)	K _D (nM)
SARS-CoV-2 S1	9.26 x $10^4 \pm 1.05$ x 10^4	$3.72 \ge 10^{-5} \pm 1.66 \ge 10^{-5}$	0.41 ± 0.23
SARS-CoV-2 S2	$2.37 \ge 10^5 \pm 3.82 \ge 10^4$	$1.51 \ge 10^{-3} \pm 4.03 \ge 10^{-4}$	6.29 ± 0.69
SARS-CoV-2 RBD	4.10 x $10^4 \pm 1.13$ x 10^3	$6.98 \ge 10^{-4} \pm 5.40 \ge 10^{-4}$	17.28 ± 13.75

 Table S1. Experimental fitting parameters for various SARS-CoV-2 proteins using S1 aptamer

Protein	k _a (1/M*s)	k _d (1/s)	K _D (nM)
SARS-CoV-2 S1	6.65 x $10^4 \pm 1.22$ x 10^4	1.01 x $10^{-4} \pm 5.19$ x 10^{-5}	1.48 ± 0.7
SARS-CoV-2 S2	1.86 x $10^5 \pm 9.39$ x 10^4	$1.84 \ge 10^{-2} \pm 2.91 \ge 10^{-2}$	104.13 ± 160.2
SARS-CoV-2 RBD	8.25 x $10^4 \pm 3.36$ x 10^3	5.02 x $10^{-4} \pm 8.21$ x 10^{-4}	5.94 ± 9.7

 Table S2. Experimental fitting parameters for various SARS-CoV-2 proteins using S1-T aptamer

Protein	k _a (1/M*s)	k _d (1/s)	K _D (nM)
SARS-CoV-2 S1	2.22 x $10^5 \pm 9.05$ x 10^4	8.44 x $10^{-4} \pm 3.62$ x 10^{-4}	3.78 ± 0.07
SARS-CoV-2 S2	$3.92 \text{ x } 10^4 \pm 2.55 \text{ x } 10^4$	4.55 x $10^{-2} \pm 1.08$ x 10^{-2}	2389.67 ± 2867.2
SARS-CoV-2 RBD	$1.52 \ge 10^5 \pm 1.83 \ge 10^5$	$7.87 \ge 10^{-3} \pm 8.78 \ge 10^{-3}$	152.69 ± 167.1

 Table S3. Experimental fitting parameters for various SARS-CoV-2 proteins using N-T aptamer



Fig. S9. Representative sensorgrams of fitted data (black lines) for varying concentrations of A) S1 protein (colored lines), B) S2 protein, C) RBD protein using the S1 aptasensor ([protein] = $0 - 2.5 \mu g/mL$).



Fig. S10. Representative sensorgrams of fitted data (black lines) for varying concentrations of A) S1 protein (colored lines), B) S2 protein, C) RBD protein using the S1-T aptasensor ([protein] = $0 - 2.5 \ \mu g/mL$).



Fig. S11. Representative sensorgrams of fitted data (black lines) for varying concentrations of A) S1 protein (colored lines), B) S2 protein, C) RBD protein using the N-T aptasensor ([protein] = $0 - 2.5 \,\mu g/mL$).



Fig. S12. A) Representative sensorgram and B) plot of LSPR signals at t=610 s, of SARS-CoV-2 S1, and various mixtures; S1+N, S1+N+RBD, S1+N+RBD+S2 ([each protein] = $2.5 \ \mu g/mL$).



Fig. S13. Molecular docking modelling of SARS-CoV S1 protein (PDB ID: 6CRZ) and S1 aptamer using PatchDock WebServer and visualized with Discovery Studio Visualizer Software.



Fig. S14. Representative sensorgram of fitted data (black lines) for various concentrations of SARS-CoV S1 protein (colored lines) using the S1 aptasensor ([SARS CoV S1] = 4.2, 8.4, 16.8 and 33.6 nM (0.3 to 2.5 μ g/mL)).

Table S4. Curve fitting data for SARS-CoV S1 protein with the S1 aptasensor. The 1:1 binding model was used in the TraceDrawer Software to obtain the k_a , k_d , and K_D values. Values represent triplicate measurements.

	$k_a \left(1/\mathrm{M*s}\right)$	k_d (1/s)	K_D (nM)
SARS-CoV	$3.99 \ge 10^5 \pm 2.65 \ge 10^4$	$1.17 \ge 10^{-3} \pm 2.10 \ge 10^{-4}$	2.9 ± 0.4



Fig. S15. Sensorgram showing sensor fabrication and performance: (1) streptavidin protein loading to the biotin-gold chip (flow rate = $20 \ \mu L/min$); followed by (2) immobilization of S1 aptamer (flow rate = $20 \ \mu L/min$); then (3) injection of 1000x diluted serum albumin spiked with S1 protein sample (flow rate = $10 \ \mu L/min$); (4) surface regeneration with 0.5% SDS (flow rate = $150 \ \mu L/min$); (5) injection of S1 protein in buffer (flow rate = $20 \ \mu L/min$); (6) surface regeneration with 0.5% SDS (flow rate = $150 \ \mu L/min$) ([streptavidin] = $0.5 \ \mu M$; [S1 aptamer] = $50 \ \mu g/mL$; [S1 protein] = $0.25 \ \mu g/mL$).



Fig. S16. Plot of LSPR signal change for SARS-CoV-2 S1 protein in spiked buffer solution compared to the 1000x diluted spiked serum albumin solution with or without S1 protein ([S1 Protein] = $2.5 \mu \text{g/mL}$; signals represent average of triplicate measurements with standard deviations shown as error bars).



Fig. S17. Representative sensorgrams showing repeatability of the SARS-CoV-2 S1 protein injections using offline prepared S1 aptamer sensor after specific storage time: (A) at t = 0 days; (B) t = 10 days; (C) t = 24 days (all S1 protein injections were followed by regeneration using 0.5% SDS buffer; [S1 Protein] = 1 µg/mL).



Fig. S18. (A) LSPR signals for 9 repeat injections of S1 protein using the online sensor or offline-prepared sensor with the shelf-life = 0, 10 or 24 days (data represent average of duplicate measurements with error bars showing standard deviation); (B) Average LSPR signals for the online or offline-prepared sensors used for 9 repeat injections of S1 protein on 0, 10 or 24 days; (C) Average LSPR signals for the offline-prepared sensor used for triplicate injections of blank buffer, BSA and S1 Protein ([S1 protein] = [BSA] = 1 μ g/mL; regeneration conditions = 0.5% SDS).