Manganese Doped Iron Coordination Polymer Nanoparticles with Enhanced Peroxidase-like Activity for Colorimetric Detection of Antioxidants

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1. Materials and reagents

Ferrous(II) chloride (FeCl₂), manganese(II) chloride (MnCl₂), o-phenylenediamine (OPD), 3,3',5,5'tetramethylbenzidine (TMB), cysteine (Cys), L-proline (Pro), L-serine (Ser), L-phenylalaine (Phe), Glycine (Gly), L-arginine (Arg), L-histidine (His), and ascorbic acid (AA) were purchased from Sigma-Aldrich (USA). Glutathione (GSH), 2,5-dihydroxyterephthalic acid (H₄DHTA), 2,2'-azinobis(3ethylbenzothiazoline-6-sulfonic acid ammonium salt) (ABTS) were supplied by Aladdin Industrial Corporation (Shanghai, China). N, N-Dimethylformamide (DMF), ethanol (EtOH), CaCl₂·2H₂O, Zn(NO₃)₂·6H₂O, KCl, glucose, maltose, fructose, lactose, sucrose, H₂O₂, acetic acid and sodium acetate were obtained from Sinopharm Chemical Reagent Co., Ltd.

2.2 Material Characterization

The Fourier transform infrared (FTIR) absorption spectra were collected in the range of 400–4000 cm⁻¹ on a Nicolet Impact 410 FTIR spectrometer with KBr pellet. Powder X-ray diffraction (PXRD) patterns were recorded on a Rigaku D-Max 2550 diffractometer using Cu-K α radiation ($\lambda = 0.15418$ nm). The ratio of metal ions was determined by an iCAP 7600 Inductively Coupled Plasma Optical Emission Spectrometer (ICP-OES) analyzer. UV-vis absorption spectra were carried out on a UV-8000 spectrophotometer (Metash, China). Transmission electron microscopy (TEM), high-resolution transmission electron microscopy (HRTEM) images and scanning transmission electron microscope (STEM-EDS) elemental mapping were conducted on a JEOL JEM-2100F at an accelerating voltage of 200 kV. X-ray photoelectron spectroscopy (XPS) recorded on a Thermo ESCALAB 250XI photoelectron spectrometer equipped with a monochromatic Al-K α radiation.



Fig. S1 TEM images of (A) Fe-DHTP, (B) Fe₂Mn-DHTP, (C) Fe₈Mn-DHTP and (D) Mn-DHTP.



Fig. S2 PXRD patterns of Fe₂Mn-DHTP, Fe₅Mn-DHTP, Fe₈Mn-DHTP, Fe-DHTP and Mn-DHTP.



Fig. S3 Relative activity of Fe_5Mn -DHTP as a function of (A) buffer pH and (B) temperature.



Fig. S4 The storage stability of Fe₅Mn-DHTP.



Fig. S5 High resolution XPS spectra of Mn 2p of Fe₅Mn-DHTP.



Fig. S6 The EPR spectra of (a) Fe_5Mn -DHTP methanol suspension and (b) Fe_5Mn -DHTP aqueous suspension.



Fig. S7 UV-vis spectra of H_2O_2 + TMB solutions in presence of (a) fresh Fe₅Mn-DHTP and (b) recycled Fe₅Mn-DHTP after treated with Cys.



Fig. S8 Specificity of this antioxidant assay toward different interference (a. blank, b. glucose, c. lactose, d. sucrose, e. fructose, f. maltose, g. His, h. Arg, i. Phe, g. Ser, k. Pro, l. Gly, m. Zn^{2+} , n. Ca^{2+} , o. Mg^{2+} , p. K⁺ and q. Na⁺, r. AA, s. Cys, t. GSH). The concentration for interference and AA was 140 μ M, for Cys and GSH was 60 μ M.

Catalyst	Substrate	$K_{\rm m}({\rm mM})$	$V_{\rm max}$ (M s ⁻¹)	Reference
	TMB	0.434	1×10^{-7}	
НКР	H_2O_2	3.7	8.71×10^{-8}	1
ZnFe ₂ O ₄ MNPs	TMB	0.85	1.33×10^{-7}	
	H_2O_2	1.66	7.74×10^{-8}	2
Fe ₃ O ₄ MNPs	TMB	0.098	3.44×10^{-8}	
	H_2O_2	154	9.78×10^{-8}	1
Fe-N-C	TMB	3.6	1.16 × 10 ⁻⁸	
	H_2O_2	12.2	3.61 × 10 ⁻⁸	3
	TMB	0.82	1.07×10^{-5}	
re _{0.5} -1N-C	H_2O_2	20.53	6.17 × 10 ⁻⁶	4
Ea MSN	TMB	0.407	7.6×10^{-8}	
re-misin	H2O2	0.01	4.33 × 10 ⁻⁸	5
	TMB	0.064	1.94×10^{-8}	
rerror-1	H_2O_2	13.33	2.12×10^{-8}	6
E- MOE	TMB	2.6	5.6×10^{-8}	
I'C-MOI	H_2O_2	1.3	2.5×10^{-8}	2.5×10^{-8} 7
	TMB	0.076	8.59 × 10 ⁻⁸	
1°eO _x @31138-0.03	H_2O_2	0.154	8.31 × 10 ⁻⁸	8
Mn-DHTP	TMB	0.082	8.92×10^{-9}	
	H_2O_2	1.09	2.19 × 10 ⁻⁹	
Fe-DHTP	TMB	0.051	2.88×10^{-8}	This work
	H_2O_2	0.88	2.43×10^{-8}	This work
Fe ₅ Mn-DHTP	TMB	0.022	6.42×10^{-8}	
	H ₂ O ₂	0.71	4.39 × 10 ⁻⁸	

 Table S1. Comparison of the kinetic constants of different nanozymes.

Materials	Antioxidant	Linear range (μM)	LOD (µM)	Reference
PANi-MnO ₂ -Pd NWs	AA	0.5–5.0	0.034	9
MIL-68	AA	30-485	6	10
IrO ₂ /GO	AA	5-70	0.324	11
Co-POP	AA	20–400	1.6	12
AuNCs	AA	0.5-200	0.22	13
Fe ₅ Mn-DHTP	AA	5-120	1.47	This work
N-MoS ₂ NFS	Cys	0-4000	0.68	14
MoS ₂ -Au@Pt	Cys	0.8–54.4	0.5	15
NiMo ₆ @Co ₃ O ₄	Cys	1–20	0.018	16
MoS ₂ @CoFe ₂ O ₄	Cys	0.5-15	0.1	17
FeOC1	Cys	3–33	2.76	18
Fe ₅ Mn-DHTP	Cys	1–26	0.33	This work
Co-POP	GSH	5–90 90–300	0.71	12
ZIF-67	GSH	0.1–25	0.07	19
MnO ₂ /CDs	GSH	0.1–10	0.095	20
Ir/NC	GSH	0.05-15	0.5	21
MnO ₂ nanosheets	GSH	1.0–20 20–80		22
Fe ₅ Mn-DHTP	GSH	1-40	0.51	This work

 Table S2. Comparison of different colorimetric assays for antioxidant.

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