

## Electric Supplementary Information

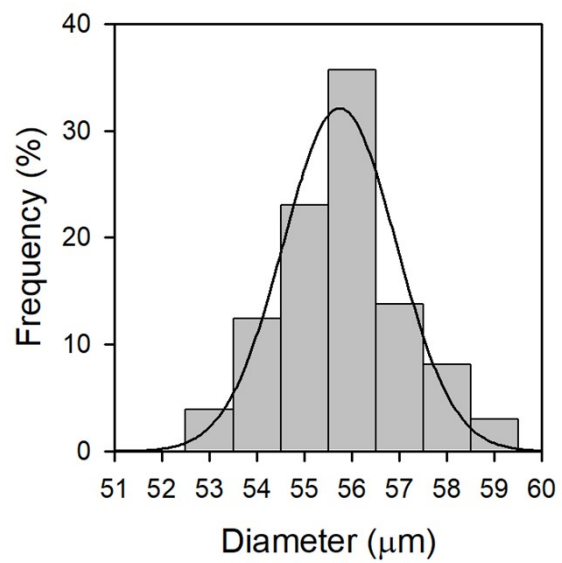
### Stretch-Responsive Adhesive Microcapsules for Strain-Regulated Antibiotic Release from Fabric Wound Dressings

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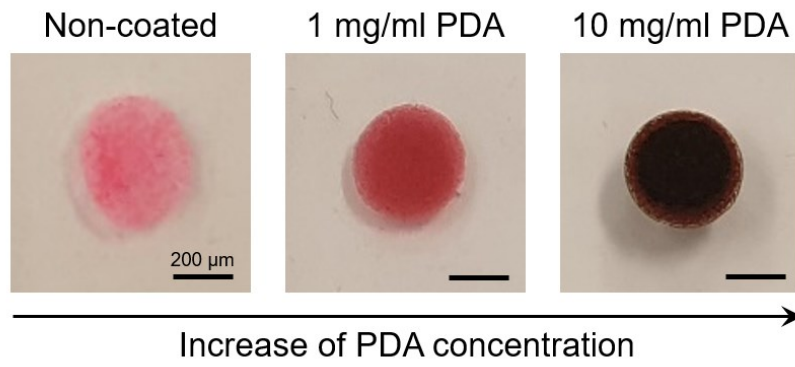
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**Table S1.** Adhesion strength of PDA-MAMCs at different concentration of PDA coating to porcine skin and a plastic surface (n ≥ 4).

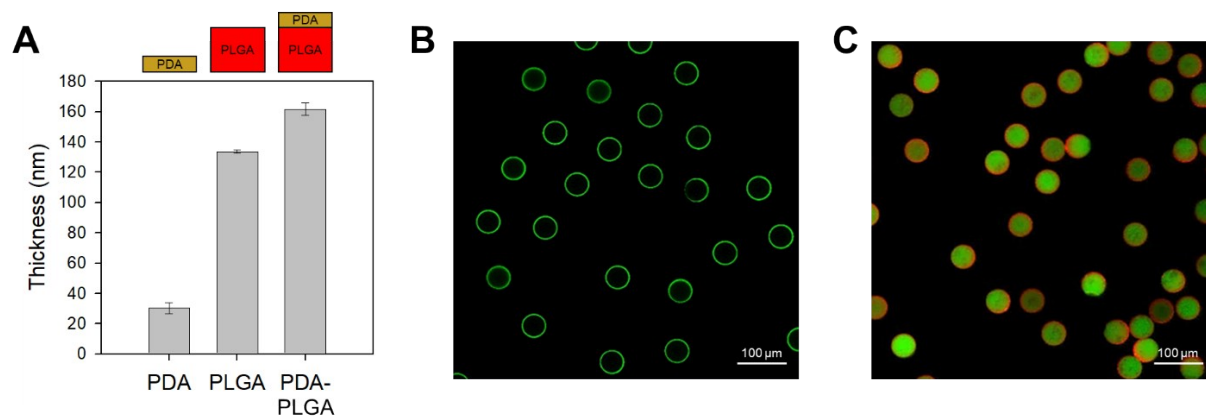
<b>Substrate</b>	<b>1 mg/ml PDA</b>	<b>10 mg/ml PDA</b>
Porcine skin	633 ± 106 g	689 ± 93 g
Plastic	510 ± 77 g	608 ± 109g



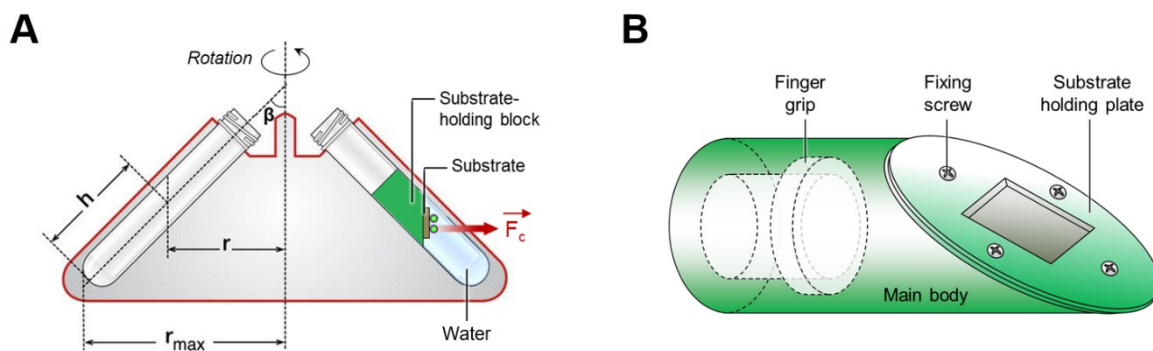
**Fig. S1** Size distribution of MAMCs.



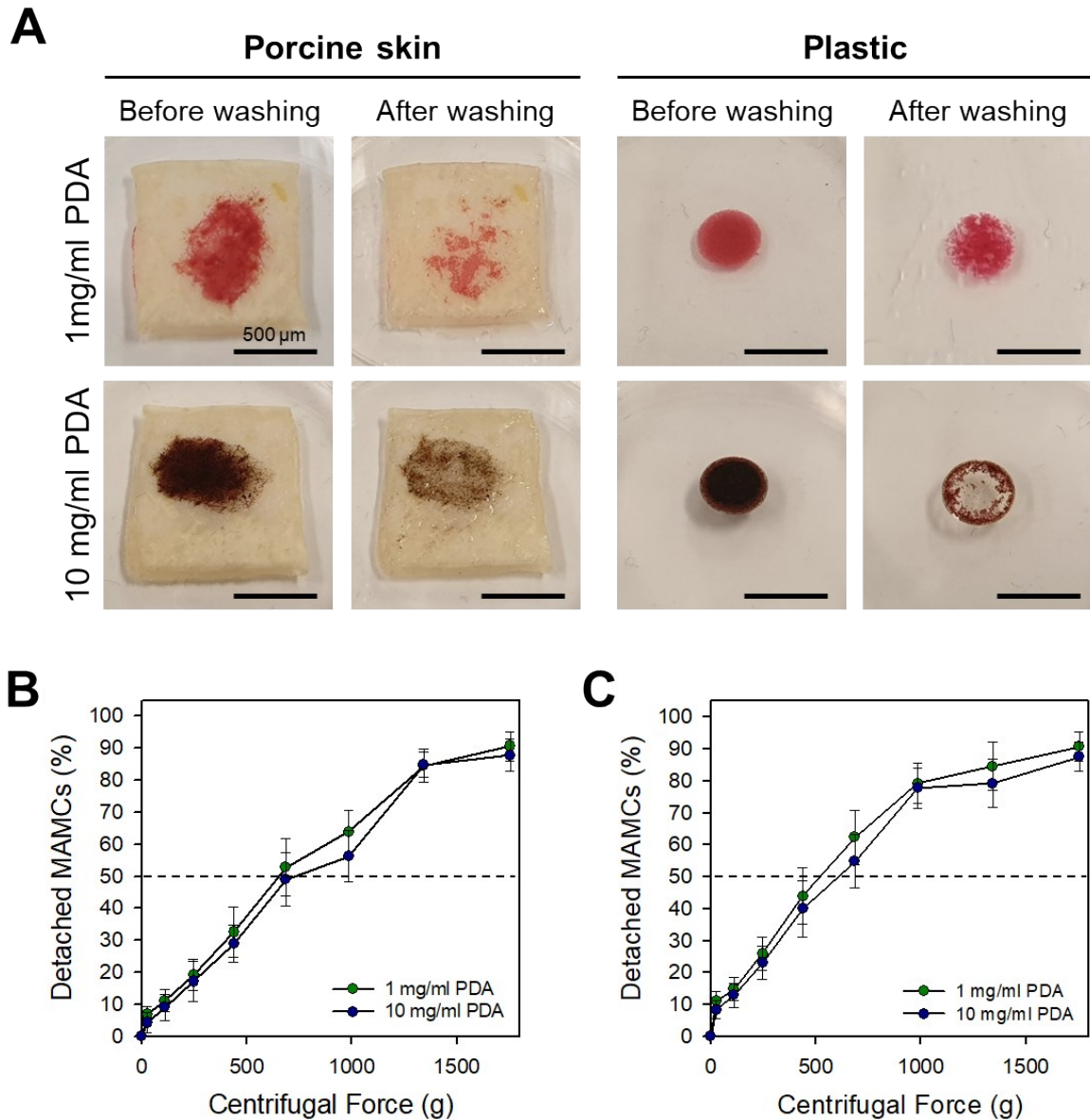
**Fig. S2** Optimal images of MAMCs before and after PDA coating. The brown color becomes darker with the increase of the concentration of PDA.



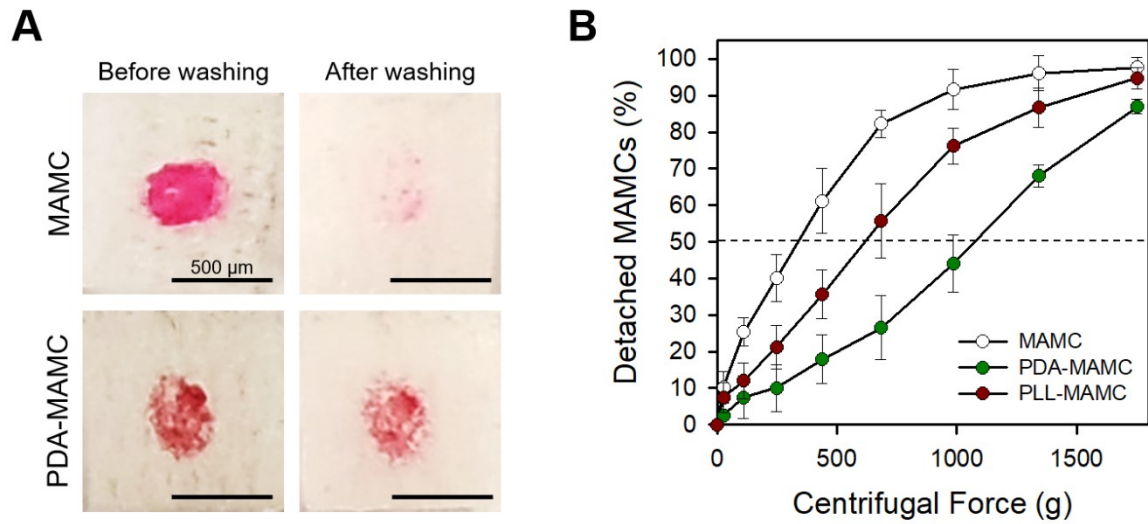
**Fig. S3** (A) Thickness of PDA coating layer, PLGA film and PDA-coated PLGA film measured by ellipsometry. (B) Confocal microscopy image of the PDA-MAMCs with labeled PDA coating layer (green). To clearly visualize the PDA layer, non-labeled BSA is used as the aqueous core. (C) Confocal microscopy image of the PDA-MAMCs with labeled PLGA shells (red) used for mechano-activation analyses (% Full > 95). To accurately quantify the percentage of full microcapsules, fluorescent BSA (green) is encapsulated into the PDA-MAMC with non-labeled PDA coating layer.



**Fig. S4** Measurement of adhesion strength of PDA-MAMCs. Schematic illustration of (A) the centrifuge method for measurement of adhesion strength and (B) the custom-designed substrate-holding block.  $\beta$ , the angle of the centrifuge tube;  $h$ , the height of sample in the centrifuge tube;  $r$ , the radial diameter of the samples;  $r_{max}$ , the maximum radial diameter of the centrifuge tube.

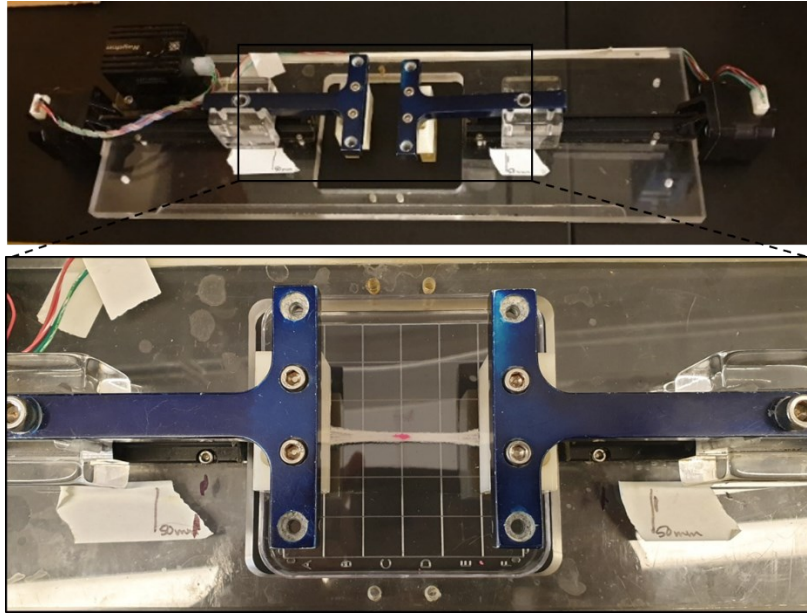


**Fig. S5** Adhesiveness of PDA-MAMCs at different concentration of PDA coating. (A) Images showing adhesion of microcapsules to porcine skin and a plastic surface. (B, C) Microcapsule detachment profiles from (B) porcine skin and (C) a plastic surface as a function of centrifugal force ( $n \geq 4$  specimens,  $***p < 0.005$ ,  $**p < 0.01$ ,  $*p < 0.05$ ; Kolmogorov-Smirnov test).

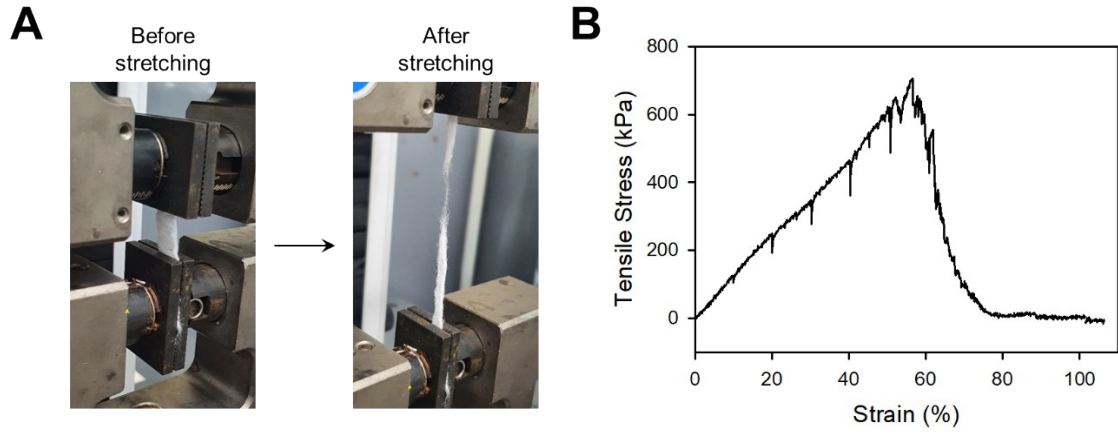


**Fig. S6** Adhesion of PDA-MAMCs on the fabric dressing. (A) Images showing adhesion of MAMCs and PDA-MAMCs on a gauze. (B) Microcapsule detachment profiles from the gauze at different centrifugal forces ( $n \geq 4$  specimens; \*\*\* $p < 0.005$ , \*\* $p < 0.01$ , \* $p < 0.05$ ; Kolmogorov-Smirnov test).

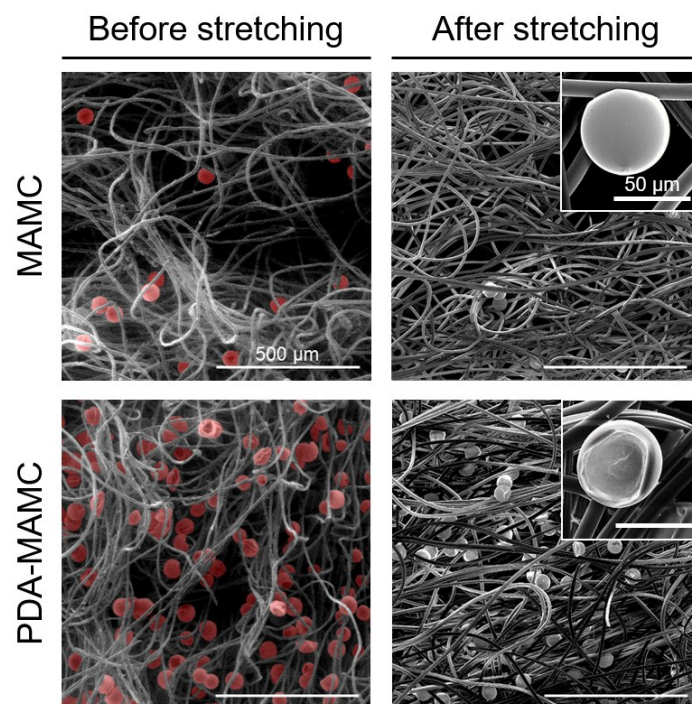




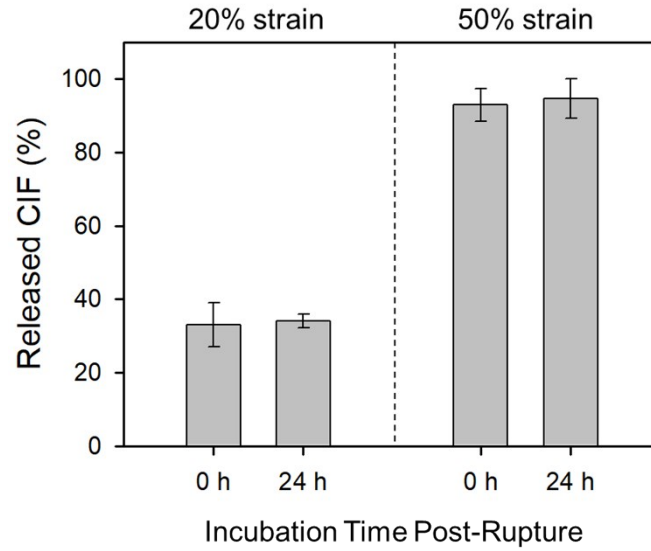
**Fig. S7** Experimental setup of a custom-built micromechanical tester.



**Fig. S8** (A) Photographs of a gauze before and after application of tensile strain. (B) Strain–stress curve of the gauze upon uniaxial stretching with 10% stepwise increments at a strain rate of 1%/s.



**Fig. S9** Stretch-induced mechano-activation of PDA-MAMCs in the fibrous matrix of gauze. SEM images of the gauze loaded with pseudo-red-colored microcapsules before and after stretching.



**Fig. S10** Released CIF from the CIF@PDA-MAMCs-laden gauze immediately (0 h) after or 24 h after rupture induced by stretching with tensile strains of 20% and 50% ( $n \geq 500$  microcapsules/loading regimen/type, 4 specimens/loading regimen/type).