

## Supplementary Information

### Surface engineering of oncolytic adenovirus for combinations of immune checkpoint blockade and virotherapy

Peng Lv<sup>1\*</sup>, Xiaomei Chen<sup>1</sup>, Shiyong Fu<sup>1</sup>, En Ren<sup>1</sup>, Chao Liu<sup>1</sup>, Xuan Liu<sup>1</sup>, Lai Jiang<sup>1</sup>, Yun Zeng<sup>2</sup>, Xiaoyong Wang<sup>1,3</sup>, and Gang Liu<sup>1\*</sup>

<sup>1</sup> State Key Laboratory of Molecular Vaccinology and Molecular Diagnostics and Center for Molecular Imaging and Translational Medicine, School of Public Health, Xiamen University, Xiamen 361102, China;

<sup>2</sup> Department of Pharmacy, Xiamen Medical College, Xiamen 361023, China;

<sup>3</sup> Amoy Hopeful Biotechnology Co., Ltd., Xiamen 361027, China.

\*Email: gangliu.cmitm@xmu.edu.cn; lv.peng@xmu.edu.cn

### Experimental Section

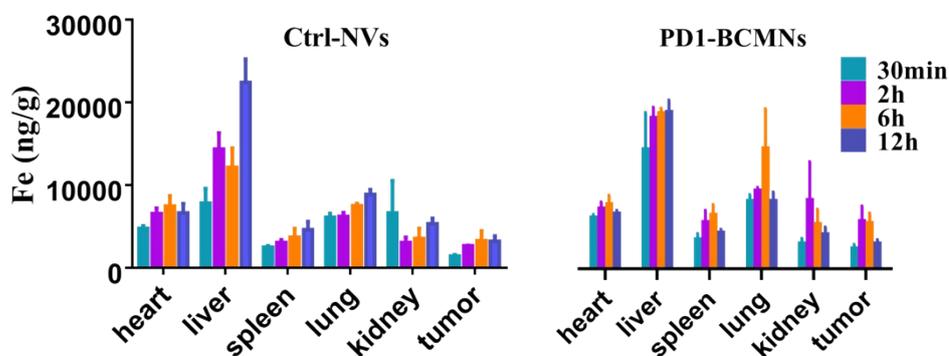
**Plasmid Construction.** The PD-1 (mouse) gene with GFP gene sequence were subcloned to a PLV vector and expressed in L929 cell line, generating PD-1 gene stable expression cell line L929-PD1.

**Localization of PD-1.** L929-PD1 were seeded onto confocal glass-bottom dishes in DMEM medium supplemented with 10% FBS. After 24h, L929-PD1 cells were stained with DAPI for 15 min. Live cells were monitored with confocal laser scanning microscopes (Olympus). For the evaluation of PD-1 on the cell membrane surface, PD-1 antibodies were loaded on the surface of BeaverBeads™ Streptavidin

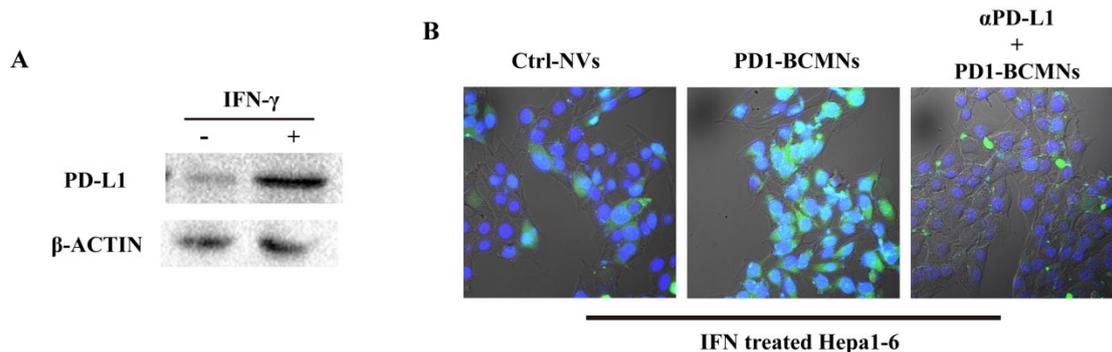
(BEAVER). PD1-BCMNs-GFP were mixed with BeaverBeads™ Streptavidin at room temperature for 30 min, blank EVs were used as control.

**Real-time PCR analysis.** After cells, tissues or plasma were collected, total RNA were isolated by Trizol (Sangon, SK1321). Reverse transcription was performed with RevertAid Premium Reverse Transcriptase kit (Thermo Scientific™ EP0733). Expression levels of PD-L1 mRNA was measured by SG Fast qPCR Master Mix (High Rox) (2X) (BBI) (ABI) (B639273). Primers for PD-L1 gene (5'ACTTGCTACGGGCGTTTACT3' and 5'ACTAACGCAAGCAGGTCCAG3') was used.

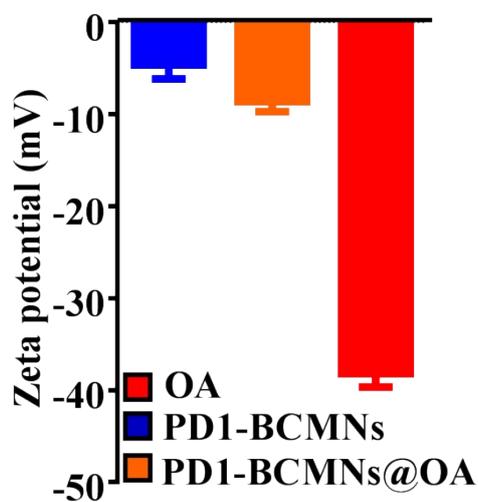
## Results



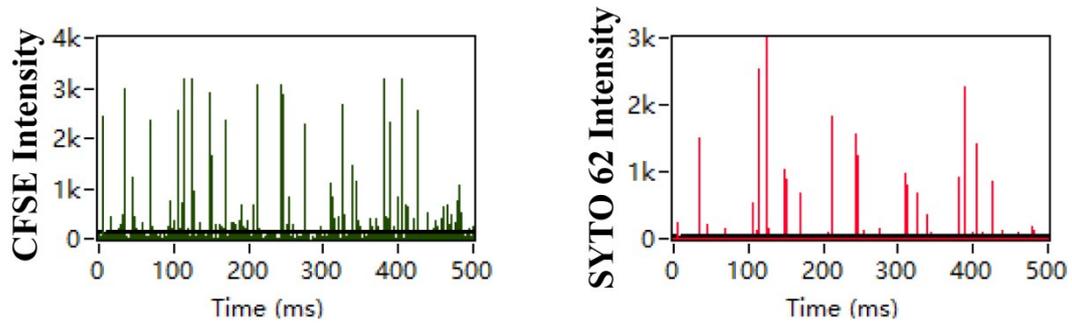
**Figure S1.** Distribution of Ctrl-NVs and PD1-BCMNs in tumor and major organs. Iron intensity per gram of tissue in tumor and major organs (n = 5). Error bar, mean ± s.d.



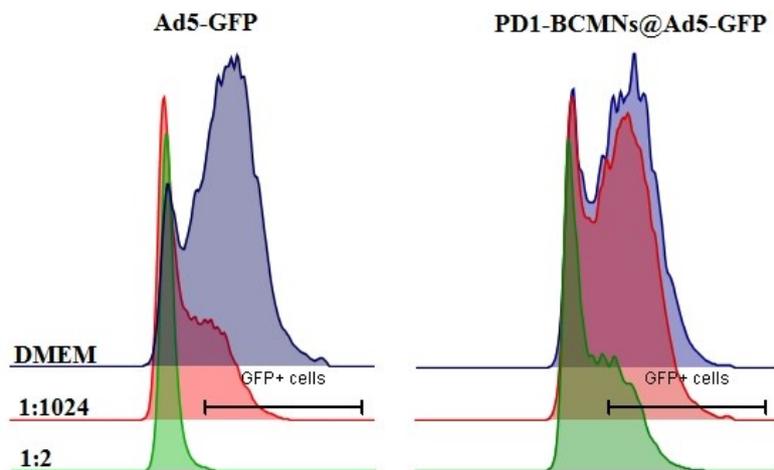
**Figure S2.** Characterization of PD1-BCMNs on Hepa1-6 cells with high levels of endogenous PD-L1. (A) Western blot analysis of PD-L1 expression of Hepa1-6 cells after treated with IFN- $\gamma$  (200 U/mL). (B) (F) CLSM image for the binding capacity of PD1-BCMNs detection. PD1-BCMNs and Ctrl-NVs were stained with CFSE (green).



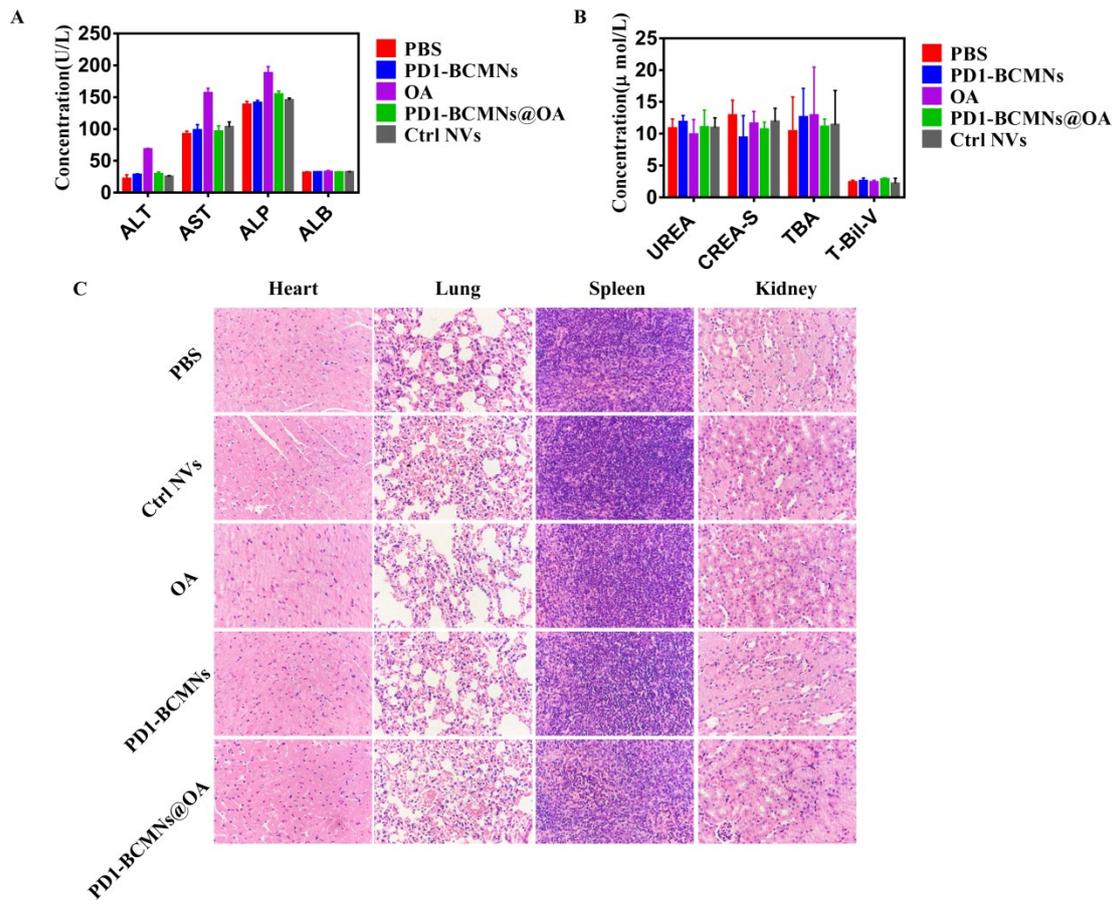
**Figure S3.** Zeta-potential of OA, PD1-BCMNs and PD1-BCMNs@OA measured by dynamic light scattering (DLS).



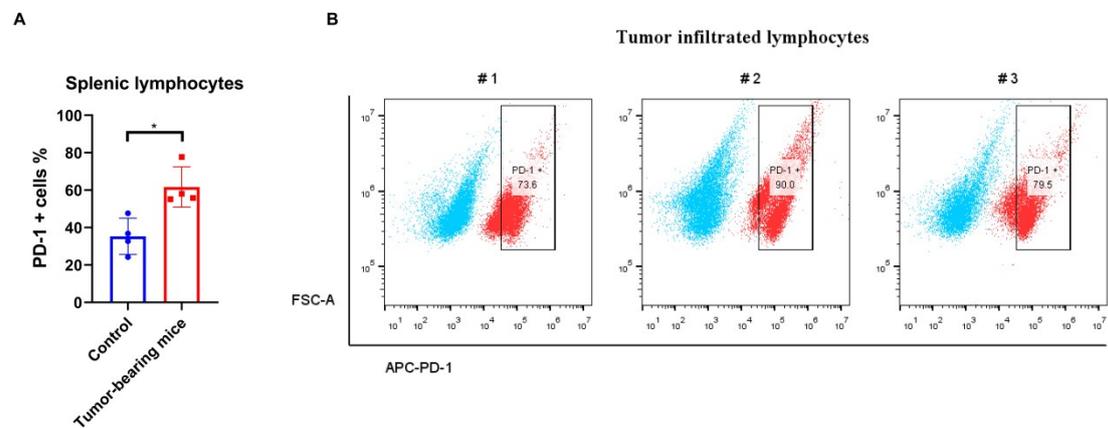
**Figure S4.** The encapsulation efficiency of PD1-BCMNs@OA was further evaluated by using a High Sensitivity Flow Cytometry (HSFCM). FITC-CFSE intensity, PC5.5-SYTO62 intensity represented PD1-BCMNs and OA, respectively.



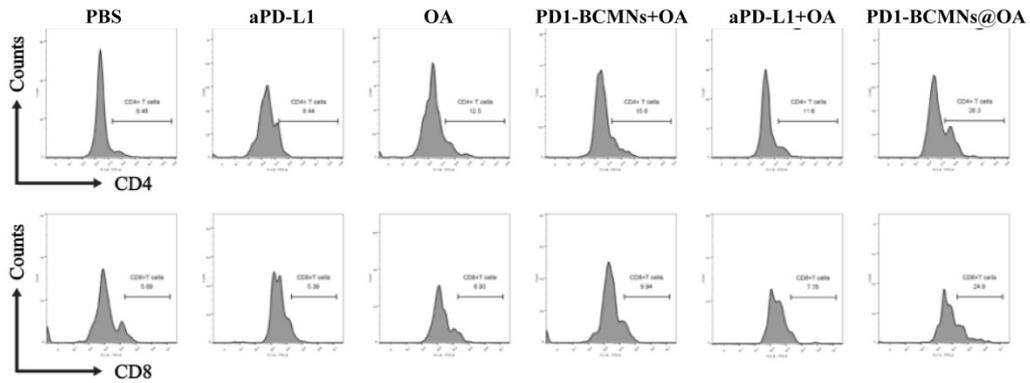
**Figure S5.** Flow cytometry assay of Ad5-GFP@BCMNs and Ad5-GFP in HEK 293 cells with anti-Ad5 serum at indicated dilutions.



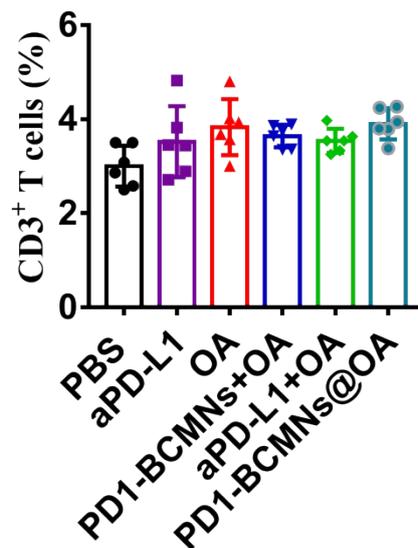
**Figure S6.** Biological safety evaluation of OA, PD1-BCMNs, Ctrl NVs, and PD1-BCMNs@OA after intravenous injection.



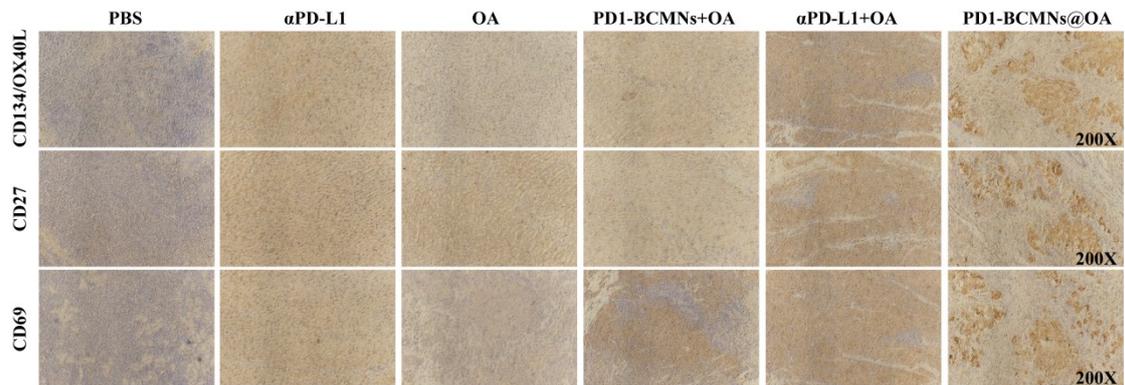
**Figure S7.** Quantitative analysis of PD1 expressing levels in splenic lymphocytes (A) and tumor infiltrated lymphocytes (B) by the flow cytometry. \* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\* $p < 0.001$ .



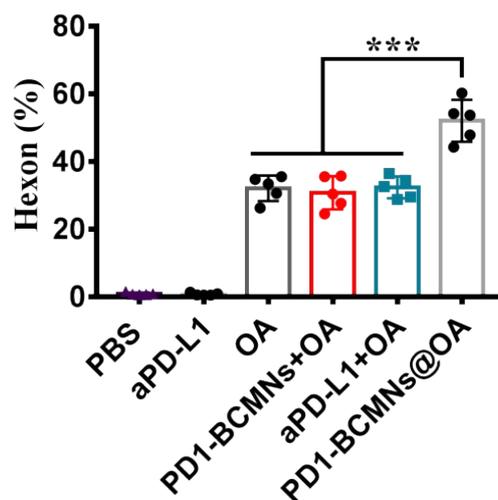
**Figure S8.** Representative plots of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in different treatment groups analyzed by the flow cytometry.



**Figure S9.** Quantitative analysis of infiltrating CD3<sup>+</sup> T cells in different treatment groups analyzed by the flow cytometry.



**Figure S10.** Representative images of T cell activation markers (CD134/OX40L, CD27, and CD69 proteins) immunohistochemistry staining for tumor from Hepa1-6 subcutaneous tumor models at day 23. Error bar, mean  $\pm$  s.d.



**Figure S11.** Quantitative analysis of Hexon rate in histology sections with anti-Hexon antibody staining, respectively. \* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\* $p < 0.001$ .