

Supporting Information

Ultrasound-Activable Piezoelectric Membranes for Accelerating Wound Healing

Xingxing Shi,^{ab} Yingxin Chen,^{ac} Yi Zhao,^{ad} Mingzhou Ye,^{ad} Shuidong Zhang,^b

Shaoqin Gong^{*adef}

^aWisconsin Institute for Discovery, University of Wisconsin–Madison, Madison,
Wisconsin 53706, United States

^bSchool of Mechanical and Automotive Engineering, South China University of
Technology, Guangzhou, Guangdong 510640, China

^cKey Laboratory of Novel Materials for Sensor of Zhejiang Province, College of
Materials and Environmental Engineering, Hangzhou Dianzi University, Hangzhou,
Zhejiang 310018, China

^dDepartment of Biomedical Engineering, University of Wisconsin–Madison, Madison,
Wisconsin 53706, United States

^eDepartment of Chemistry, University of Wisconsin–Madison, Madison, Wisconsin
53706, United States

^fDepartment of Materials Science and Engineering, University of Wisconsin–Madison,
Madison, Wisconsin 53706, United States

*Corresponding author.

E-mail addresses: shaoqingong@wisc.edu (S. Gong).

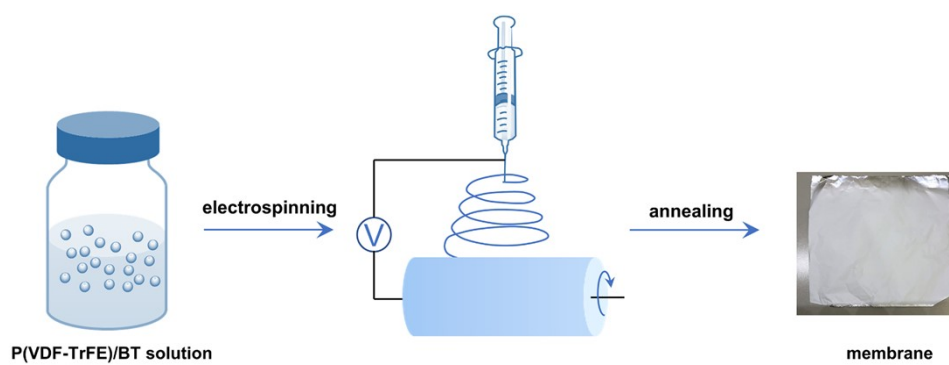


Fig. S1 Schematic illustration of the fabrication of P(VDF-TrFE)/BT membranes.

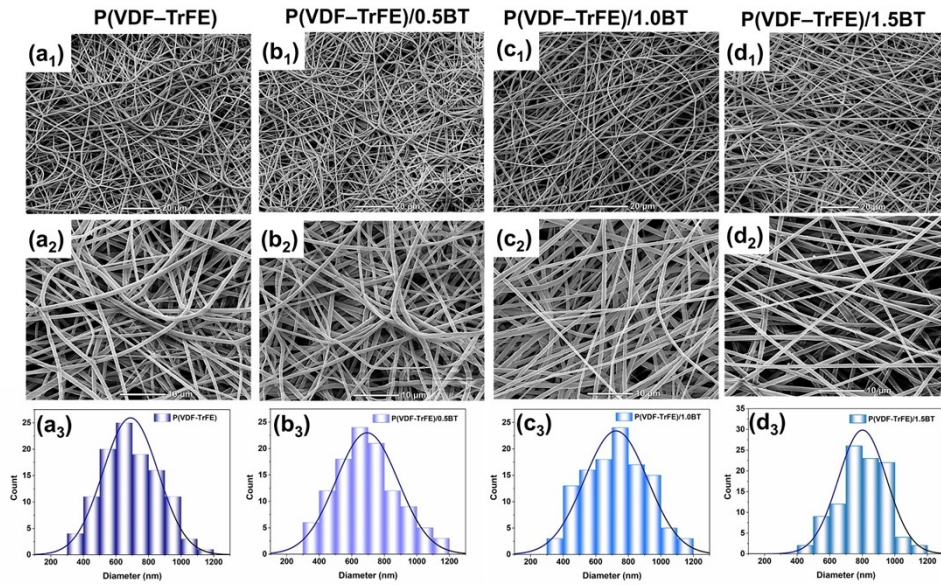


Fig. S2 SEM images showing the nanofibrous microstructure and diameter distributions of (a₁, a₂, a₃) P(VDF-TrFE), (b₁, b₂, b₃) P(VDF-TrFE)/0.5BT, (c₁, c₂, c₃) P(VDF-TrFE)/1.0BT, and (d₁, d₂, d₃) P(VDF-TrFE)/1.5BT membranes.

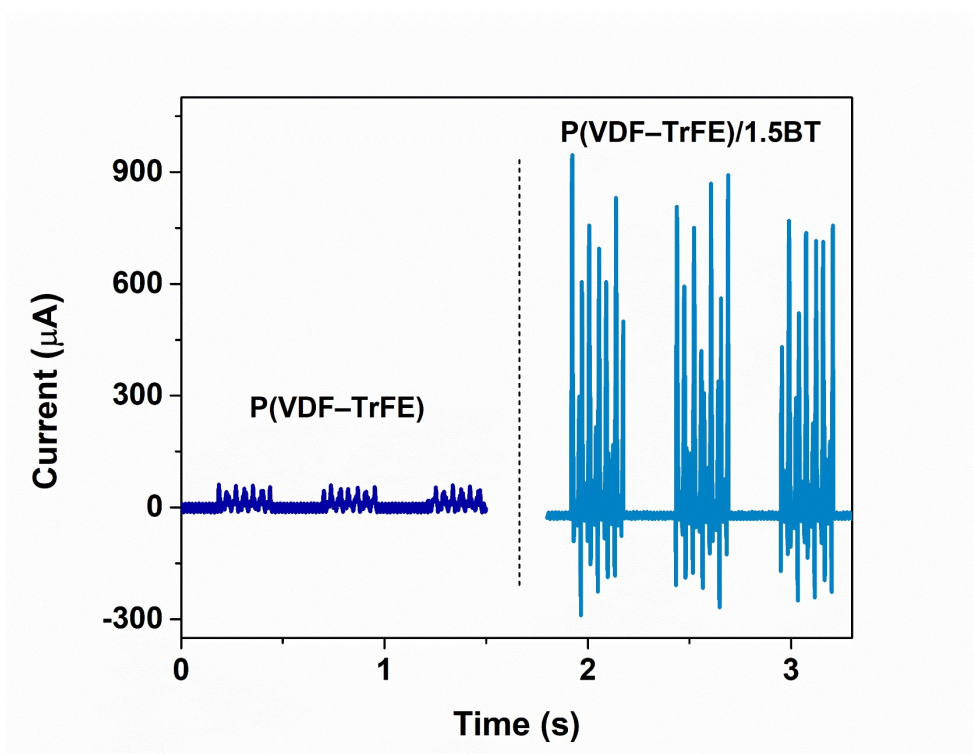


Fig. S3 The output currents generated from the P(VDF-TrFE) and P(VDF-TrFE)/1.5BT membranes under ultrasonic stimulation.

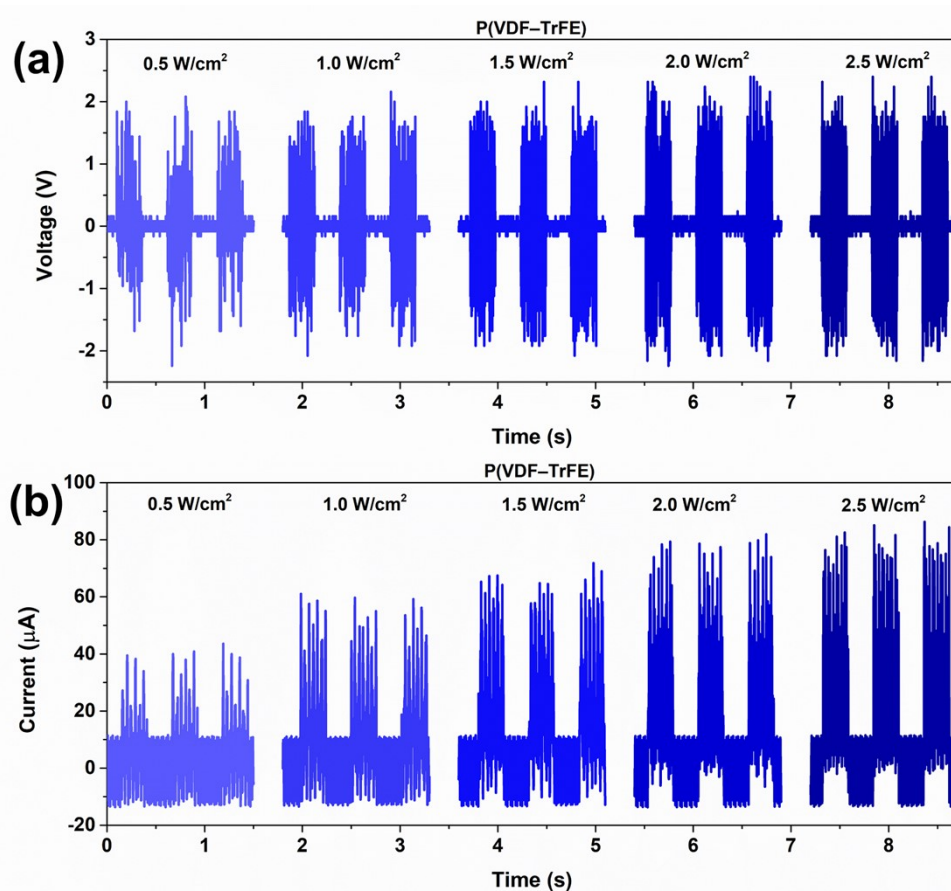


Fig. S4 The output (a) voltages and (b) currents of the P(VDF-TrFE) membranes under ultrasonic stimulation with different power intensities (i.e., 0.5 W/cm², 1.0 W/cm², 1.5 W/cm², 2.0 W/cm², and 2.5 W/cm²).

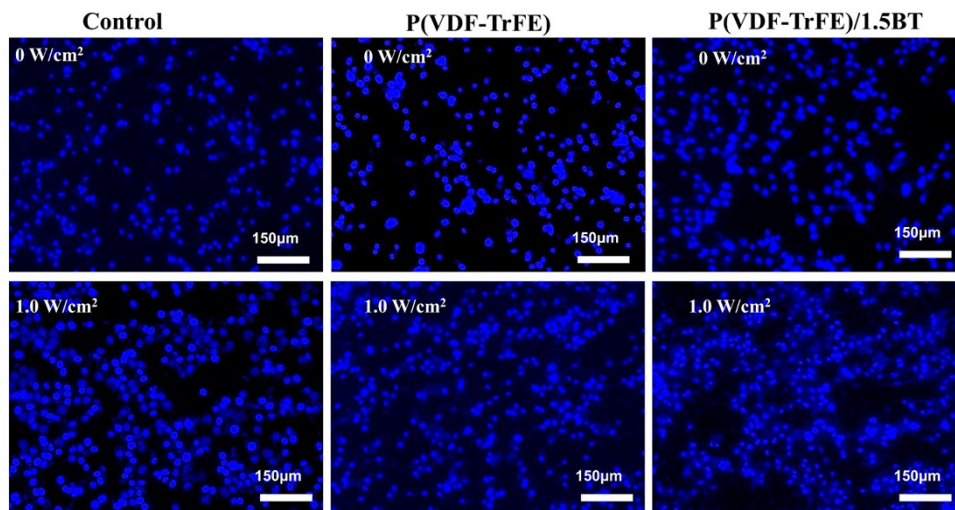


Fig. S5 The fluorescence images of DAPI-stained NIH-3T3 fibroblasts with/without ultrasound treatment for three days (scale bar = 150 μm).

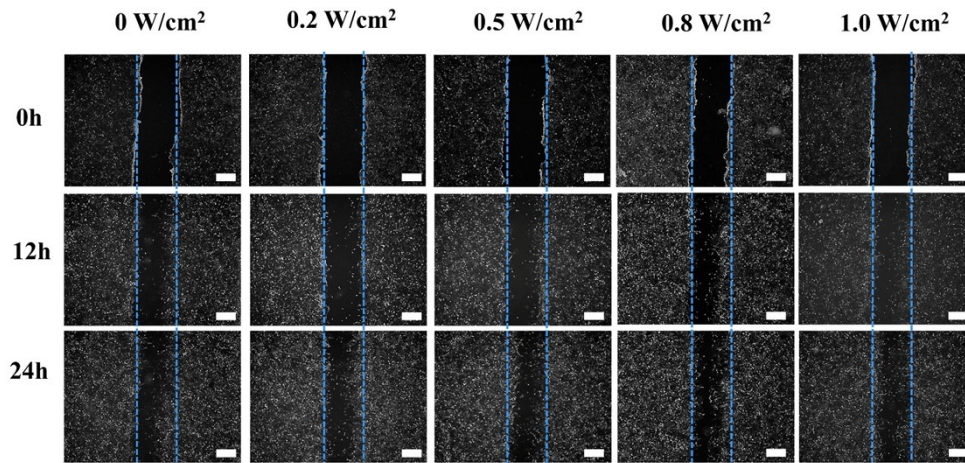


Fig. S6 The cell migration images of NIH-3T3 fibroblasts on the culture plates (control group) stimulated by ultrasound with different power intensities (scale bar = 500 μ m).

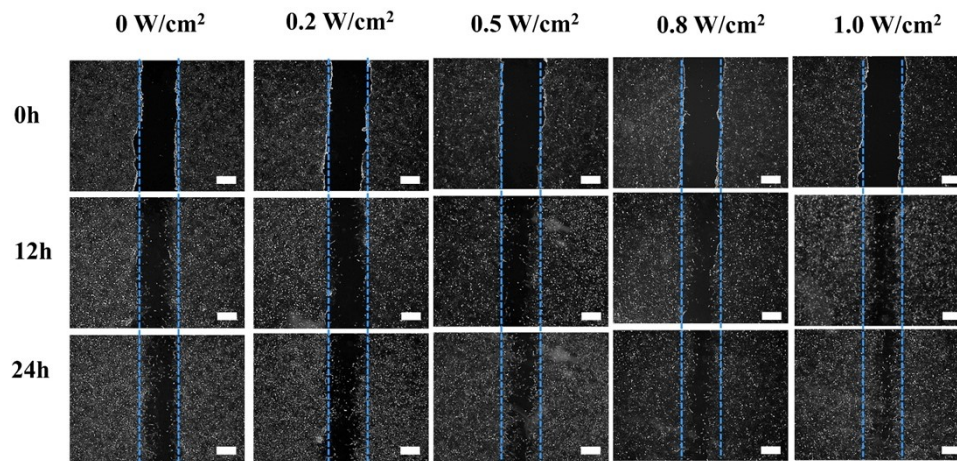


Fig. S7 The cell migration images of NIH-3T3 fibroblasts on the P(VDF–TrFE) membranes stimulated by ultrasound with different power intensities (scale bar = 500 μm).

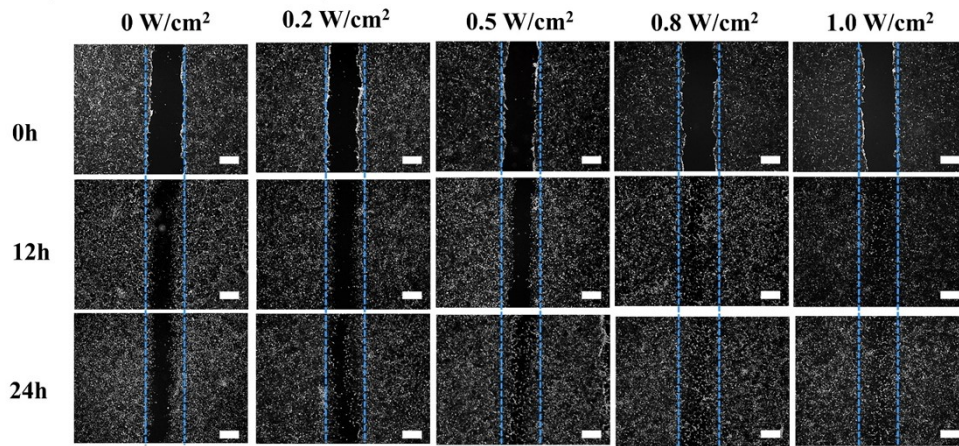


Fig. S8 The cell migration images of NIH-3T3 fibroblasts on the P(VDF–TrFE)/1.5BT membranes stimulated by ultrasound with different power intensities (scale bar = 500 μm).

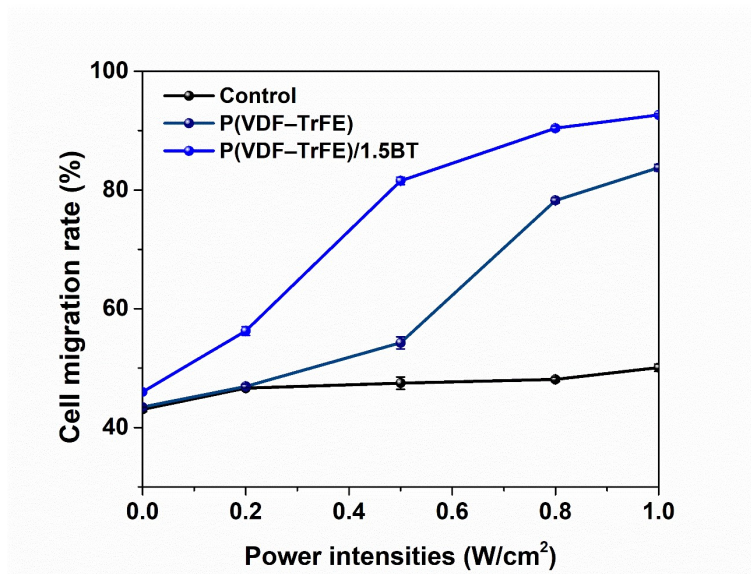


Fig. S9 The quantitative analysis of the cell migration rate of NIH-3T3 fibroblasts on the culture plates (control group) and P(VDF-TrFE)/BT membranes from the scratch tests

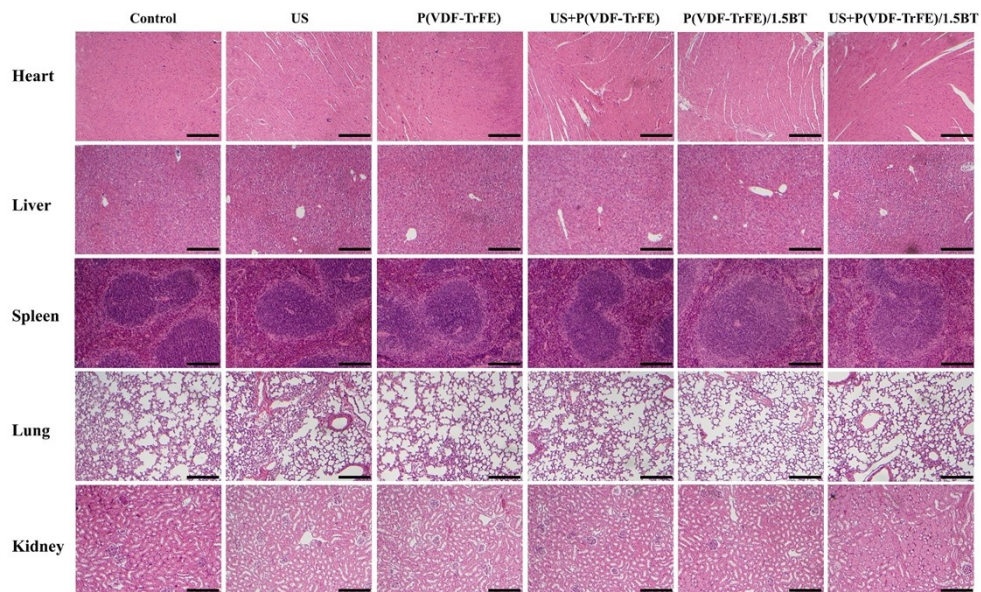


Fig. S10. H&E staining of the main tissues of the mice in different groups at the end of the experiments (scale bar = 200 μm).

Video lists

Video S1. The output voltages generated from the ultrasound-activable P(VDF–TrFE)/1.5BT membrane on the mouse skin.

Video S2. The output voltages of P(VDF–TrFE) membrane implanted at 1.5 cm.

Video S3. The output voltages of P(VDF–TrFE)/1.5BT membrane implanted at 1.5 cm.

Video S4. A flashed LED charged by the P(VDF-TrFE)/1.5BT membrane implanted at 1.5 cm under 1.4 ms pulse duration.