Optimizing CSP1 Analogs for Modulating Quorum Sensing in *Streptococcus pneumoniae* with Bulky, Hydrophobic Nonproteogenic Amino Acid Substitutions

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HPLC Traces for CSP1 Analogs



CSP1-L4Cha

CSP1-L4HLeu







CSP1-F7HLeu



CSP1-F8HLeu

CSP1-F11HLeu

CSP1-I12Cha

CSP1-I12HLeu

CSP1-F7Cha/F8HLeu

CSP1-F7HLeu/F8Cha

CSP1-F7HLeu/F8HLeu

CSP1-F7Cha/I12HLeu

CSP1-F7HLeu/I12Cha

CSP1-F7HLeu/I12HLeu

CSP1-F8Cha/I12HLeu

CSP1-F8HLeu/I12HLeu

CSP1-E1A/F7HLeu

CSP1-E1A/F8HLeu

CSP1-E1A/I12HLeu

CSP1-E1A/F7Cha/I12HLeu

MS and HPLC Data for CSP1 Analogs

| Compound | Calc. EM | Obs. EM | Purity | |
|---------------------|------------|----------------|--------|--|
| Name | $MH2^{2+}$ | $MH2^{2+}$ | (%) | |
| CSP1-L4Cha | 1142.1528 | 1142.1529 | ≥98 | |
| CSP1-L4HLeu | 1129.1449 | 1129.1451 | ≥99 | |
| CSP1-F7Cha | 1125.1606 | 1125.1561 | ≥99 | |
| CSP1-F7HLeu | 1112.1528 | 1112.1537 | ≥99 | |
| CSP1-F8Cha | 1125.1606 | 1125.1616 | ≥99 | |
| CSP1-F8HLeu | 1112.1528 | 1112.1535 | ≥99 | |
| CSP1-F11Cha | 1125.1606 | 1125.1614 | ≥99 | |
| CSP1-F11HLeu | 1112.1528 | 1112.1532 | ≥99 | |
| CSP1-I12Cha | 761.7709* | 761.7678^{*} | ≥98 | |
| CSP1-I12HLeu | 1129.1449 | 1129.1437 | ≥98 | |
| CSP1-F7Cha/F8Cha | 1128.1841 | 1128.1838 | ≥99 | |
| CSP1-F7Cha/F8HLeu | 1115.1762 | 1115.1743 | ≥99 | |
| CSP1-F7HLeu/F8Cha | 1115.1762 | 1115.1754 | ≥99 | |
| CSP1-F7HLeu/F8HLeu | 1102.1684 | 1102.1718 | ≥99 | |
| CSP1-F7Cha/I12Cha | 1145.1762 | 1145.1771 | ≥98 | |
| CSP1-F7Cha/I12HLeu | 1132.1684 | 1132.1691 | ≥99 | |
| CSP1-F7HLeu/I12Cha | 1132.1684 | 1132.1725 | ≥98 | |
| CSP1-F7HLeu/I12HLeu | 1119.1606 | 1119.1620 | ≥99 | |
| CSP1-F8Cha/I12Cha | 1145.1762 | 1145.1775 | ≥98 | |
| CSP1-F8Cha/I12HLeu | 1132.1684 | 1132.1720 | ≥99 | |
| CSP1-F8HLeu/I12Cha | 1132.1684 | 1132.1711 | ≥98 | |
| CSP1-F8HLeu/I12HLeu | 1119.1606 | 1119.1622 | ≥99 | |

Table S1. MS and HPLC data for CSP1 single and double mutant analogs.

EM = Exact Mass. See methods above, * MH_3^{3+} .

Table S2. MS and HPLC data for CSP1-E1A mutant analogs.

| Compound Name | Calc. EM MH2 ²⁺ | Obs. EM MH2 ²⁺ | Purity (%) | |
|------------------------|-------------------------------|------------------------------|---------------|--|
| CSP1-E1A/F7Cha | 1096.1578 | 1096.1562 | ≥99 | |
| CSP1-E1A/F7HLeu | 1083.1500 | 1083.1493 | ≥99 | |
| CSP1-E1A/F8Cha | 1096.1578 | 1096.1598 | ≥99 | |
| CSP1-E1A/F8HLeu | 1083.1500 | 1083.1488 | ≥99 | |
| CSP1-E1A/I12Cha | 1113.1500 | 1113.1506 | ≥99 | |
| CSP1-E1A/I12HLeu | 1100.1422 | 1100.1461 | ≥99 | |
| CSP1-E1A/F7Cha/I12Cha | 1116.1735 | 1116.1759 | ≥99 | |
| CSP1-E1A/F7Cha/I12HLeu | 1103.1657 | 1103.1672 | ≥97 | |

EM = Exact Mass. See methods above.

Primary Reporter Gene Assay Data

S. pneumoniae D39pcomX::lacZ (ComD1)

Agonism assays were performed at 10 μ M concentration of synthetic CSP1 analogs. CSP1 was used as the positive control (100%) while DMSO as the negative control (0%). Percent (%) ComD1 activation was measured by normalizing the Miller units obtained for each peptide to that of the native CSP1. All peptides were screened in triplicate over three separate trials. Error bars indicate standard error of the mean of nine values.

Figure S-1. Primary agonism screening assay data for the CSP1 single mutant analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} values.

Figure S-2. Primary agonism screening assay data for the CSP1 double mutant analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} values.

Figure S-3. Primary agonism screening assay data for the CSP1-E1A mutant analogs. None of the peptides exhibited activation of the ComD1 receptor and peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

Antagonism assays were performed at 10 μ M concentration of peptides against 50 nM concentration of CSP1. CSP1 (50 nM) was used as the positive control (100%) while DMSO as the negative control (0%). Percent (%) *comX* activation was measured by normalizing the Miller units obtained for each peptide to that of CSP1. All peptides were screened in triplicate over three separate trials. Error bars indicate standard error of the mean of nine values.

Figure S-4. Primary antagonism screening assay data for the CSP1-E1A mutant analogs. Peptides that exhibited less than 50% activation were further evaluated to determine their IC_{50} values.

S. pneumoniae TIGR4 pcomX::lacZ (ComD2)

Agonism assays were performed at 10 μ M concentration. CSP2 was used as the positive control (100%) while DMSO as the negative control (0%). Percent (%) ComD2 activation was measured by normalizing the Miller units obtained for each peptide to that of the native CSP2. All peptides were screened in triplicate over three separate trials. Error bars indicate standard error of the mean of nine values.

Figure S-5. Primary agonism screening assay data for the CSP1 single mutant analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} values.

Figure S-6. Primary agonism screening assay data for the CSP1 double mutant analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} values.

Figure S-7. Primary agonism screening assay data for the CSP1-E1A mutant analogs. None of the peptides exhibited activation of the ComD2 receptor and peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

Antagonism assays were performed at 10 μ M concentration of peptides against 250 nM concentration of CSP2. CSP2 (250 nM) was used as the positive control (100%) while DMSO as the negative control (0%). Percent (%) *comX* activation was measured by normalizing the Miller units obtained for each peptide to that of CSP2. All peptides were screened in triplicate over three separate trials. Error bars indicate standard error of the mean of nine values.

Figure S-8. Primary antagonism screening assay data for the CSP1-E1A mutant analogs. None of the peptides exhibited inhibition of the ComD2 receptor.

Agonism and Antagonism Dose Response Curves

CSP1 analogs were tested to determine their EC_{50} or IC_{50} values over varying concentrations in the two indicated *S. pneumoniae* β -galactosidase reporter strains. Each dose response curve is representative of three independent experiments performed on three separate occasions (i.e., experiments (Exp.) #1-3; shown for each peptide below). Error bars represent standard error of the mean of triplicate values. In each plot, the peptide, as well as its EC_{50} or IC_{50} value (in nM) and 95% confidence interval (95% CI) values (in nM), are indicated at top left or top right.

S. pneumoniae D39 pcomX::lacZ (ComD1)

Activation dose response curves

Inhibition dose response curves

S. pneumoniae TIGR4 pcomX::lacZ (ComD2)

Activation dose response curves

Metabolic Stability Analysis of Lead CSP1 Analogs

Figure S-9. Degradation pattern analysis of CSP1 in the presence of trypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 0.5, 1, 2, 3, 4, 5, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed trypsin cleavage sites annotated on the CSP1 sequence.

Figure S-10. Degradation pattern analysis of CSP1-F7ChaI12Cha in the presence of trypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 0.5, 1, 2, 3, 4, 5, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed trypsin cleavage sites annotated on the CSP1-F7ChaI12Cha sequence.

Figure S-11. Degradation pattern analysis of CSP1-E1AF7Cha in the presence of trypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 0.5, 1, 2, 3, 4, 5, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed trypsin cleavage sites annotated on the CSP1-E1AF7Cha sequence.

Figure S-12. Degradation pattern analysis of CSP1 in the presence of chymotrypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 0.5, 1, 2, 3, 4, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed chymotrypsin cleavage sites annotated on the CSP1 sequence.

Figure S-13. Degradation pattern analysis of CSP1-F7ChaI12Cha in the presence of chymotrypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 0.5, 1, 2, 3, 4, 5, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed chymotrypsin cleavage sites annotated on the CSP1-F7ChaI12Cha sequence.

Figure S-14. Degradation pattern analysis of CSP1-E1AF7Cha in the presence of chymotrypsin. A) Comparison of analytical HPLC chromatograms (220 nm) across the time points taken at 0, 2, 4, 6 and 24 h. B) Zoomed view of panel A with all the time points (0, 1, 2, 3, 4, 5, 6 and 24 h). All peaks were collected and analyzed via mass spectrometry (MALDI-TOF MS and ESI-TOF LC-MS). The degradation products corresponding to each peak are listed on the left. C) A summary of observed chymotrypsin cleavage sites annotated on the CSP1-E1AF7Cha sequence.