# Computational design and experimental characterisation of a stable human heparanase variant 

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## Supplementary Material



Supplementary Figure 1. Wild type human heparanase expression in E. coli. The 50 and 8 kDa subunits and chaperones ${ }^{28}$ in total (annotated as T ) and in soluble fraction (annotated as S ) are shown with LMW protein marker (GE healthcare). 8 kDa subunit with N -terminal 6 xHis tag and 50 kDa subunit were amplified by PCR using gBlocks (IDT, Supplementary Table 1) and inserted into multiple cloning sites of pETDuet-1 coexpression vector (Novagen). The plasmid DNA was transformed into Shuffle T7 Express competent cells (NEB) together with chaperones in PACYC vector. Overnight seed culture from single colony was inoculated by $1 \%$ into LB media supplemented with ampicillin and chloramphenicol antibiotics. The culture was incubated at $37^{\circ} \mathrm{C}$ for about 3-5 hours until the OD was about 0.8. 0.4 mM IPTG was added and further incubated for about 3 hours at $37^{\circ} \mathrm{C}$. The cell pellet was resuspended in 50 mM acetate buffer at pH 5 and lysed using BugBuster protein extraction reagent (Merck) for the solubility measurement.


Supplementary Figure 2. Design of stable heparanase mutants from PROSS (A) the resulting mutations are shown for each design. (B) The crystal structure of heparanase (PDB ID: 5E9C) was used as template whereby 50 kDa subunit (cyan cartoon) was targeted with restriction on the residues contacting the ligand (black sticks) and the 8 kDa subunit (blue cartoon).


Supplementary Figure 3. Pross variant expression trials in E. coli SHuffle cells. P1-P7 plasmid DNA was transformed into Shuffle T7 Express competent cells (NEB) together with varying chaperones in pACYC vector. Skp+TF was tested in 1, 4, 7, 10 and 13 . GroEL/ES+TF was tested in $2,5,8,11,14,17$, and 20 . Skp + GroEL/ES was tetesd in $3,6,9,12,15,18$ and 21 . LMW protein marker (GE healthcare) is shown on the left. Overnight seed culture from single colony was inoculated by $1 \%$ into LB media supplemented with ampicillin and chloramphenicol antibiotics. The culture was incubated at $37^{\circ} \mathrm{C}$ for about $3-5$ hours until the OD was about 0.8. 0.4 mM IPTG was added and further incubated for about 3 hours. The cell pellet was resuspended in 50 mM acetate buffer at pH 5 and lysed using BugBuster protein extraction reagent (Merck) for the solubility measurement.


Supplementary Figure 4: Crystallographic B-Factors of WT HSPE and HSPE P6. Comparison between the B factors from the two structures (with different Wilson B-factors and resolution) show an overall decrease in magnitude of the $B$ factors in the Mutant P6, yet the overall trend in terms of regions of high/low B-factors is very similar.


Supplementary Figure 5: The disulfide bond between Cys542 and Cys437 that is stabilised by A540P mutation, which is in a favourable conformation is the kinked loop preceding Cys542.


Supplementary Figure 6: Principal component analysis (PCA) sampling quality for WT and P6 simulations. Sampling shows that the 10 simulations sample a large range of conformational space for both WT and mutant replica simulations, and using all replica simulations is viable in the analysis.

Supplementary Table 1. Nucleotide sequences of heparanase variants used for cloning

| Name | Sequences |
| :---: | :---: |
| $8 \quad k D a$  <br> subunit of <br> wild type <br> (gBlock)  | CAGGATGTGGTCGATCTTGATTTCTTCACGCAGGAGCCTTTGCACCTTGTATCCCC CTCATTTTTAAGCGTTACCATAGACGCAAACCTTGCCACTGACCCGCGCTTCTTAA TCTTGCTTGGTAGTCCAAAGTTAAGAACGCTGGCGCGGGGGCTTAGTCCAGCATA TCTGCGCTTTGGCGGAACGAAAACAGACTTCCTGATTTTTGATCCTAAAAAGGAA |
| 50 kDa subunit of wild type (gBlock) | AAAAAATTTAAGAACTCCACTTATAGCCGCAGTTCAGTCGACGTGCTGTACACCTT CGCGAACTGTTCGGGATTGGATTTGATATTTGGATTAAATGCATTGTTGCGCACGG CGGATCTGCAGTGGAACTCTAGTAACGCGCAATTATTGTTAGATTATTGTAGTTCG AAGGGCTACAATATATCGTGGGAATTGGGTAATGAGCCGAACAGCTTTTTGAAGAA AGCCGACATCTTTATTAATGGGTCTCAGCTGGGCGAAGATTTTATACAACTTCACA AGCTGTTACGCAAATCAACATTTAAGAACGCGAAGTTATATGGACCAGATGTTGGG CAGCCACGTAGAAAGACCGCCAAGATGCTGAAAAGCTTCCTTAAAGCAGGAGGTG AAGTGATTGACTCGGTGACCTGGCATCACTACTACTTAAACGGAAGAACAGCAACT CGTGAGGATTTCTTGAACCCGGATGTCCTTGATATATTTATTTCATCTGTACAAAAA GTCTTCCAAGTTGTAGAATCCACCAGACCTGGCAAAAAAGTGTGGTTAGGAGAGA CTTCAAGCGCTTACGGCGGTGGTGCACCTCTTTTGTCCGACACCTTCGCGGCAGG CTTCATGTGGCTGGACAAATTGGGCTTAAGCGCGCGTATGGGGATCGAAGTGGTG ATGCGGCAGGTATTCTTTGGCGCCGGGAACTATCACCTGGTCGATGAAAATTTTGA TCCTTTACCTGATTATTGGTTGTCATTACTGTTTAAAAAGTTGGTCGGGACAAAGGT CCTTATGGCTTCTGTCCAGGGGAGTAAAAGAAGAAAATTGAGAGTTTACTTGCATT GCACCAATACGGACAACCCGAGATATAAGGAAGGAGACTTGACCTTATACGCTATC AATTTGCACAATGTTACGAAATATTTGCGTTTACCTTACCCATTCTCCAACAAACAA GTTGACAAATACTTGCTGCGCCCTCTGGGTCCGCATGGCTTATTATCCAAATCGGT TCAGTTGAATGGCTTAACTCTGAAAATGGTAGATGATCAGACATTGCCACCATTGA tGGAGAAA |
| 50 kDa subunit of design (gBlock) | ccccatcttagtatattagttaagtataagaaggagatatacatATGAAAAAATTCAAAAACTCGACGTAT AGCCGGTCTTCTGTGGATGTGCTCTATACTTTTGCGAAGTGTTCGGGCCTGGACTT AATCTTCGGCTTAAATGCACTGCTTCGGACTTCAGATGGGCAGTGGAATTCTAGCA ATGCTCAGCTCCTGCTCGATTACTGTGCCTCTAAAGGGTATAACATCGACTGGGAG TTGGGCAACGAGCCAAATAGCTTCCGTAAAAAGGCTGGGATCTTCATCAACGGGT CGCAATTAGGCAAGGACTTCATTCACCTTCACAAACTGCTCCGGAAATCGACATTT AAGAATGCGAAACTGTATGGCCCTGATGTAGGTCAACCGCGCGGGAAAACGGCCA AAATGCTTAAATCGTTCCTGAAGGCGGGCGGCGAAGTCATTGATGCAGTAACATG GCACCATTACTATTTGAATGGTCGCACCGCCACCTTAGAAGATTTCCTGAATCCGG ACGTATTGGACACGTTTATTTCTCAGGTTCAAAAGGTCTTGCAAGTTGTCGAATCG |



## Supplementary Table 2. Amino acid sequences of heparanase variants

| $8 \quad \mathrm{kDa}$ <br> subunit | MGSSHHHHHHSQDPNSSSQDVVDLDFFTQEPLHLVSPSFLSVTIDANLATDPRFLIL LGSPKLRTLARGLSPAYLRFGGTKTDFLIFDPKKE* |
| :---: | :---: |
| 50 kDa subunit of wild type | MKKFKNSTYSRSSVDVLYTFANCSGLDLIFGLNALLRTADLQWNSSNAQLLLDYCSS KGYNISWELGNEPNSFLKKADIFINGSQLGEDFIQLHKLLRKSTFKNAKLYGPDVGQP RRKTAKMLKSFLKAGGEVIDSVTWHHYYLNGRTATREDFLNPDVLDIFISSVQKVFQ VVESTRPGKKVWLGETSSAYGGGAPLLSDTFAAGFMWLDKLGLSARMGIEVVMRQ VFFGAGNYHLVDENFDPLPDYWLSLLFKKLVGTKVLMASVQGSKRRKLRVYLHCTN TDNPRYKEGDLTLYAINLHNVTKYLRLPYPFFSNKQVDKYLLRPLGPHGLLSKSVQLN GLTLKMVDDQTLPPLMEKPLRPGSSLGLPAFSYSFFVIRNAKVAACI* |
| 50 kDa subunit of design | MKKFKNSTYSRSSVDVLYTFAKCSGLDLIFGLNALLRTSDGQWNSSNAQLLLDYCA SKGYNIDWELGNEPNSFRKKAGIFINGSQLGKDFIHLHKLLRKSTFKNAKLYGPDVG QPRGKTAKMLKSFLKAGGEVIDAVTWHHYYLNGRTATLEDFLNPDVLDTFISQVQKV LQVVESTRPGKKVWLGETSSAYGGGAPGLSDTFAAGFMWLDKLGLSARMGIEVVM RQVFFGAGNYHLVDENFDPLPDYWLSLLFKKLVGTKVLMASVQGQDRRKLRVYLH CTNTDNPRYKEGDLTLYAINLHNVTKYLRLPYPFSNKQVDQYLLRPHGPDGLLSKSV QLNGQTLKMVDDQTLPPLKPKPLRPGSSLGLPAFSYAFFVIRNAKVPACI* |

## Supplementary Table 3. Data collection and refinement statistics

| PDB ID | 7RG8 |
| :---: | :---: |
| Data collection |  |
| Space group | P $2_{1} 2_{1} 2_{1}$ |
| Cell dimensions |  |
| $a, b, c$ ( A$)$ | 59.7876 .09124 .43 |
| $\alpha, \beta, \gamma\left({ }^{\circ}\right)$ | 90.0090 .0090 .00 |
| Resolution ( A ) | 47.01-1.30 (1.346-1.3) |
| $R_{\text {merge }}$ | 0.07 (1.11) |
| $1 / \mathrm{\sigma l}$ | 11.82 (0.33) |
| $\mathrm{CC}_{1 / 2}$ | 0.998 (0.882) |
| Completeness (\%) | 99.78\% (99.0\%) |
| Redundancy | 13.2 (11.3) |
| Refinement |  |
| Resolution (Å) | 47.01-1.30 (1.346-1.3) |
| No. reflections | 139516 (13656) |
| $R_{\text {work }} / R_{\text {free }}$ | 0.142/0.164 |
| No. atoms |  |
| Protein | 3660 |
| Ligand/ion | 11 |
| Water | 363 |
| $B$-factors |  |
| Protein | 24.89 |
| Ligand/ion | 21.94 |
| Water | 41.23 |
| R.m.s. deviations |  |
| Bond lengths (Å) | 0.007 |
| Bond angles ( ${ }^{\circ}$ ) | 1.01 |
| Ramachandran plot |  |
| Preferred (\%) | 98.68 |
| Allowed (\%) | 1.32 |
| Outliers (\%) | 0.00 |

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[^0]:    *X-ray data were collected from single crystals. *Values in parentheses are for highest-resolution shell.

