Computational design and experimental characterisation of a stable human heparanase variant

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Supplementary Material



Supplementary Figure 1. Wild type human heparanase expression in *E. coli*. The 50 and 8 kDa subunits and chaperones²⁸ in total (annotated as T) and in soluble fraction (annotated as S) are shown with LMW protein marker (GE healthcare). 8 kDa subunit with N-terminal 6xHistag and 50 kDa subunit were amplified by PCR using gBlocks (IDT, **Supplementary Table 1**) and inserted into multiple cloning sites of pETDuet-1 coexpression vector (Novagen). The plasmid DNA was transformed into Shuffle T7 Express competent cells (NEB) together with chaperones in pACYC vector. Overnight seed culture from single colony was inoculated by 1% into LB media supplemented with ampicillin and chloramphenicol antibiotics. The culture was incubated at 37 °C for about 3-5 hours until the OD was about 0.8. 0.4 mM IPTG was added and further incubated for about 3 hours at 37 °C. The cell pellet was resuspended in 50 mM acetate buffer at pH 5 and lysed using BugBuster protein extraction reagent (Merck) for the solubility measurement.



Supplementary Figure 2. Design of stable heparanase mutants from PROSS (**A**) the resulting mutations are shown for each design. (**B**) The crystal structure of heparanase (PDB ID: 5E9C) was used as template whereby 50 kDa subunit (cyan cartoon) was targeted with restriction on the residues contacting the ligand (black sticks) and the 8 kDa subunit (blue cartoon).



Supplementary Figure 3. Pross variant expression trials in *E. coli* SHuffle cells. P1-P7 plasmid DNA was transformed into Shuffle T7 Express competent cells (NEB) together with varying chaperones in pACYC vector. Skp+TF was tested in 1, 4, 7, 10 and 13. GroEL/ES+TF was tested in 2, 5, 8, 11, 14, 17, and 20. Skp + GroEL/ES was tetesd in 3, 6, 9, 12, 15, 18 and 21. LMW protein marker (GE healthcare) is shown on the left. Overnight seed culture from single colony was inoculated by 1% into LB media supplemented with ampicillin and chloramphenicol antibiotics. The culture was incubated at 37 °C for about 3-5 hours until the OD was about 0.8. 0.4 mM IPTG was added and further incubated for about 3 hours. The cell pellet was resuspended in 50 mM acetate buffer at pH 5 and lysed using BugBuster protein extraction reagent (Merck) for the solubility measurement.



Supplementary Figure 4: Crystallographic B-Factors of WT HSPE and HSPE P6. Comparison between the B factors from the two structures (with different Wilson B-factors and resolution) show an overall decrease in magnitude of the B factors in the Mutant P6, yet the overall trend in terms of regions of high/low B-factors is very similar.



Supplementary Figure 5: The disulfide bond between Cys542 and Cys437 that is stabilised by A540P mutation, which is in a favourable conformation is the kinked loop preceding Cys542.



Supplementary Figure 6: Principal component analysis (PCA) sampling quality for WT and P6 simulations. Sampling shows that the 10 simulations sample a large range of conformational space for both WT and mutant replica simulations, and using all replica simulations is viable in the analysis.

Name	Sequences	
8 kDa	CAGGATGTGGTCGATCTTGATTTCTTCACGCAGGAGCCTTTGCACCTTGTATCCCC	
subunit of	CTCATTTTTAAGCGTTACCATAGACGCAAACCTTGCCACTGACCCGCGCTTCTTAA	
wild type	TCTTGCTTGGTAGTCCAAAGTTAAGAACGCTGGCGCGGGGGCTTAGTCCAGCATA	
(gBlock)	TCTGCGCTTTGGCGGAACGAAAACAGACTTCCTGATTTTTGATCCTAAAAAGGAA	
50 kDo		
SU KDa		
suburiit or		
(aBlock)		
(дыоск)		
	AGCIGITACGCAAATCAACATTAAGAACGCGAAGTTATATGGACCAGATGTTGGG	
	CGIGAGGAIIICIIGAACCCGGAIGICCIIGAIAIAIIIAII	
	GTCTTCCAAGTTGTAGAATCCACCAGACCTGGCAAAAAGTGTGGTTAGGAGAGA	
	CTTCAAGCGCTTACGGCGGTGGTGCACCTCTTTTGTCCGACACCTTCGCGGCAGG	
	CTTCATGTGGCTGGACAAATTGGGCTTAAGCGCGCGTATGGGGATCGAAGTGGTG	
	ATGCGGCAGGTATTCTTTGGCGCCGGGAACTATCACCTGGTCGATGAAAATTTTGA	
	TCCTTTACCTGATTATTGGTTGTCATTACTGTTTAAAAAGTTGGTCGGGACAAAGGT	
	CCTTATGGCTTCTGTCCAGGGGGGGGAGTAAAAGAAGAAAATTGAGAGTTTACTTGCATT	
	GCACCAATACGGACAACCCGAGATATAAGGAAGGAGACTTGACCTTATACGCTATC	
	AATTTGCACAATGTTACGAAATATTTGCGTTTACCTTACCCATTCTCCAACAAACA	
	GTTGACAAATACTTGCTGCGCCCTCTGGGTCCGCATGGCTTATTATCCAAATCGGT	
	TCAGTTGAATGGCTTAACTCTGAAAATGGTAGATGATCAGACATTGCCACCATTGA	
	TGGAGAAA	
50 kDa	ccccatcttagtatattagttaagtataagaaggagatatacatATGAAAAAATTCAAAAACTCGACGTAT	
subunit of	AGCCGGTCTTCTGTGGATGTGCTCTATACTTTTGCGAAGTGTTCGGGCCTGGACTT	
design	AATCTTCGGCTTAAATGCACTGCTTCGGACTTCAGATGGGCAGTGGAATTCTAGCA	
(gBlock)	ATGCTCAGCTCCTGCTCGATTACTGTGCCTCTAAAGGGTATAACATCGACTGGGAG	
	TTGGGCAACGAGCCAAATAGCTTCCGTAAAAAGGCTGGGATCTTCATCAACGGGT	
	CGCAATTAGGCAAGGACTTCATTCACCTTCACAAACTGCTCCGGAAATCGACATTT	
	AAGAATGCGAAACTGTATGGCCCTGATGTAGGTCAACCGCGCGGGAAAACGGCCA	
	AAATGCTTAAATCGTTCCTGAAGGCGGGCGGCGAAGTCATTGATGCAGTAACATG	
	GCACCATTACTATTTGAATGGTCGCACCGCCACCTTAGAAGATTTCCTGAATCCGG	
	ACGTATTGGACACGTTTATTTCTCAGGTTCAAAAGGTCTTGCAAGTTGTCGAATCG	

Supplementary Table 1. Nucleotide sequences of heparanase variants used for cloning

ACCCGGCCTGGGAAGAAGTTTGGCTCGGGGAGACAAGTTCCGCCTATGGCGGT GGCGCTCCTGGCCTCTCAGATACCTTTGCTGCTGGTTTCATGTGGCTTGATAAACT GGGCCTCTCCGCTCGCATGGGGATCGAAGTCGTGATGCGCCAAGTATTTTTTGGC GCTGGCAACTACCACCTCGTCGACGAAAACTTCGATCCATTGCCTGACTACTGGCT GAGCCTCCTTTTCAAAAAGTTAGTTGGTACAAAGGTGTTGATGGCAAGTGTTCAGG GTCAGGATCGCCGCAAACTTCGCGTTTATCTCCATTGCACAAATACGGATAATCCT CGCTACAAAGAAGGCGACCTGACGCTCTATGCTATCAACCTCCATAACGTCACCA AGTATCTCCGCCTGCCATATCCTTTTAGTAATAAACAGGTGGATCAATATCTCTTGC

GCCCTCATGGCCCTGATGGTTTACTGTCCAAGAGCGTGCAGTTGAATGGCCAGAC CCTCAAGATGGTTGATGACCAGACTTTGCCTCCTTTGAAGCCAAAACCACTGCGTC CGGGGAGCAGTCTTGGCCTGCCTGCCTTCTCCTACGCATTTTTTGTAATTCGTAAC GCAAAGGTCCCAGCCTGCATCTGATAACTCGAGTCTGGTAAAGAAACCG

Supplementary Table 2. Amino acid sequences of heparanase variants

8 kDa subunit	MGSSHHHHHHSQDPNSSSQDVVDLDFFTQEPLHLVSPSFLSVTIDANLATDPRFLIL LGSPKLRTLARGLSPAYLRFGGTKTDFLIFDPKKE*
50 kDa subunit of wild type	MKKFKNSTYSRSSVDVLYTFANCSGLDLIFGLNALLRTADLQWNSSNAQLLLDYCSS KGYNISWELGNEPNSFLKKADIFINGSQLGEDFIQLHKLLRKSTFKNAKLYGPDVGQP RRKTAKMLKSFLKAGGEVIDSVTWHHYYLNGRTATREDFLNPDVLDIFISSVQKVFQ VVESTRPGKKVWLGETSSAYGGGAPLLSDTFAAGFMWLDKLGLSARMGIEVVMRQ VFFGAGNYHLVDENFDPLPDYWLSLLFKKLVGTKVLMASVQGSKRRKLRVYLHCTN TDNPRYKEGDLTLYAINLHNVTKYLRLPYPFSNKQVDKYLLRPLGPHGLLSKSVQLN GLTLKMVDDQTLPPLMEKPLRPGSSLGLPAFSYSFFVIRNAKVAACI*
50 kDa subunit of design	MKKFKNSTYSRSSVDVLYTFAKCSGLDLIFGLNALLRTSDGQWNSSNAQLLLDYCA SKGYNIDWELGNEPNSFRKKAGIFINGSQLGKDFIHLHKLLRKSTFKNAKLYGPDVG QPRGKTAKMLKSFLKAGGEVIDAVTWHHYYLNGRTATLEDFLNPDVLDTFISQVQKV LQVVESTRPGKKVWLGETSSAYGGGAPGLSDTFAAGFMWLDKLGLSARMGIEVVM RQVFFGAGNYHLVDENFDPLPDYWLSLLFKKLVGTKVLMASVQGQDRRKLRVYLH CTNTDNPRYKEGDLTLYAINLHNVTKYLRLPYPFSNKQVDQYLLRPHGPDGLLSKSV QLNGQTLKMVDDQTLPPLKPKPLRPGSSLGLPAFSYAFFVIRNAKVPACI*

PDB ID	7RG8
Data collection	
Space group	P 2 ₁ 2 ₁ 2 ₁
Cell dimensions	
a, b, c (Å)	59.78 76.09 124.43
α, β, γ (°)	90.00 90.00 90.00
Resolution (Å)	47.01-1.30 (1.346-1.3)
R _{merge}	0.07 (1.11)
Ι / σΙ	11.82 (0.33)
CC _{1/2}	0.998 (0.882)
Completeness (%)	99.78% (99.0%)
Redundancy	13.2 (11.3)
Refinement	
Resolution (Å)	47.01-1.30 (1.346-1.3)
No. reflections	139516 (13656)
R _{work} / R _{free}	0.142/ 0.164
No. atoms	
Protein	3660
Ligand/ion	11
Water	363
B-factors	
Protein	24.89
Ligand/ion	21.94
Water	41.23
R.m.s. deviations	
Bond lengths (Å)	0.007
Bond angles (°)	1.01
Ramachandran plot	
Preferred (%)	98.68
Allowed (%)	1.32
Outliers (%)	0.00

Supplementary Table 3. Data collection and refinement statistics

*X-ray data were collected from single crystals. *Values in parentheses are for highest-resolution shell.