

Electronic Supplementary Information

**Construction of optically controllable CRISPR-Cas9 system using a DNA origami nanostructure**

Katsuhiko Abe,<sup>a</sup> Hiroshi Sugiyama,\*<sup>a,b</sup> and Masayuki Endo\*<sup>a,b,c</sup>

<sup>a</sup>Department of Chemistry, Graduate School of Science, Kyoto University, Kitashirakawa-oiwakecho, Sakyo-ku, Kyoto 606-8502, Japan.

<sup>b</sup>Institute for Integrated Cell-Material Sciences, Kyoto University, Yoshida-ushinomiya-cho, Sakyo-ku, Kyoto 606-8501, Japan.

<sup>c</sup>Organization for Research and Development of Innovative Science and Technology, Kansai University, 3-3-35 Yamate, Suita, Osaka 564-8680, Japan.

## **Experimental Procedure**

**Materials.** All the staple DNAs for the DNA nanoring were purchased from Eurofins Genomics (Tokyo, Japan). Single-stranded p8064 DNA was purchased from Tilibit nanosystems (Garching, Germany). DNA strand containing a cleavable linker was purchased from Japan Bio Services (Saitama, Japan). Cas9 and Streptavidin Magnetic Beads for purification were purchased from New England Biolabs (Ipswich, MA). The gel-filtration column and the Sephacryl S-300 and S-400 were purchased from BioRad Laboratories (Hercules, CA) and GE healthcare (Buckinghamshire, UK), respectively.

**Preparation of DNA nanoring.** A solution was prepared with 40 nM p8046 ssDNA (template strand), 4 equivalents of staple strands, 10 mM MgCl<sub>2</sub>, 1 mM EDTA, and pH was adjusted with 20 mM Tris-HCl (pH 7.6) as a buffer. This solution was incubated at 65 °C for 15 min, then cooled stepwise by 1 °C every 60 min from 60 °C to 40 °C to assemble nanoring. Sequences of staple DNAs used here are listed in Table 1.

**Preparation of sgRNA.** Single-guide RNA (sgRNA) was prepared by in vitro transcription from a template dsDNA containing T7 promoter sequence. The template dsDNA was prepared by performing overlap extension PCR with an 84 nt and 56 nt template DNA strands (Table S1). The PCR amplification was performed using Go Taq polymerase (Promega) with a forward and reverse primer (Table S1) by following the manufacturer's protocol. The amplified product was purified using a QIAquick PCR purification Kit (Qiagen).

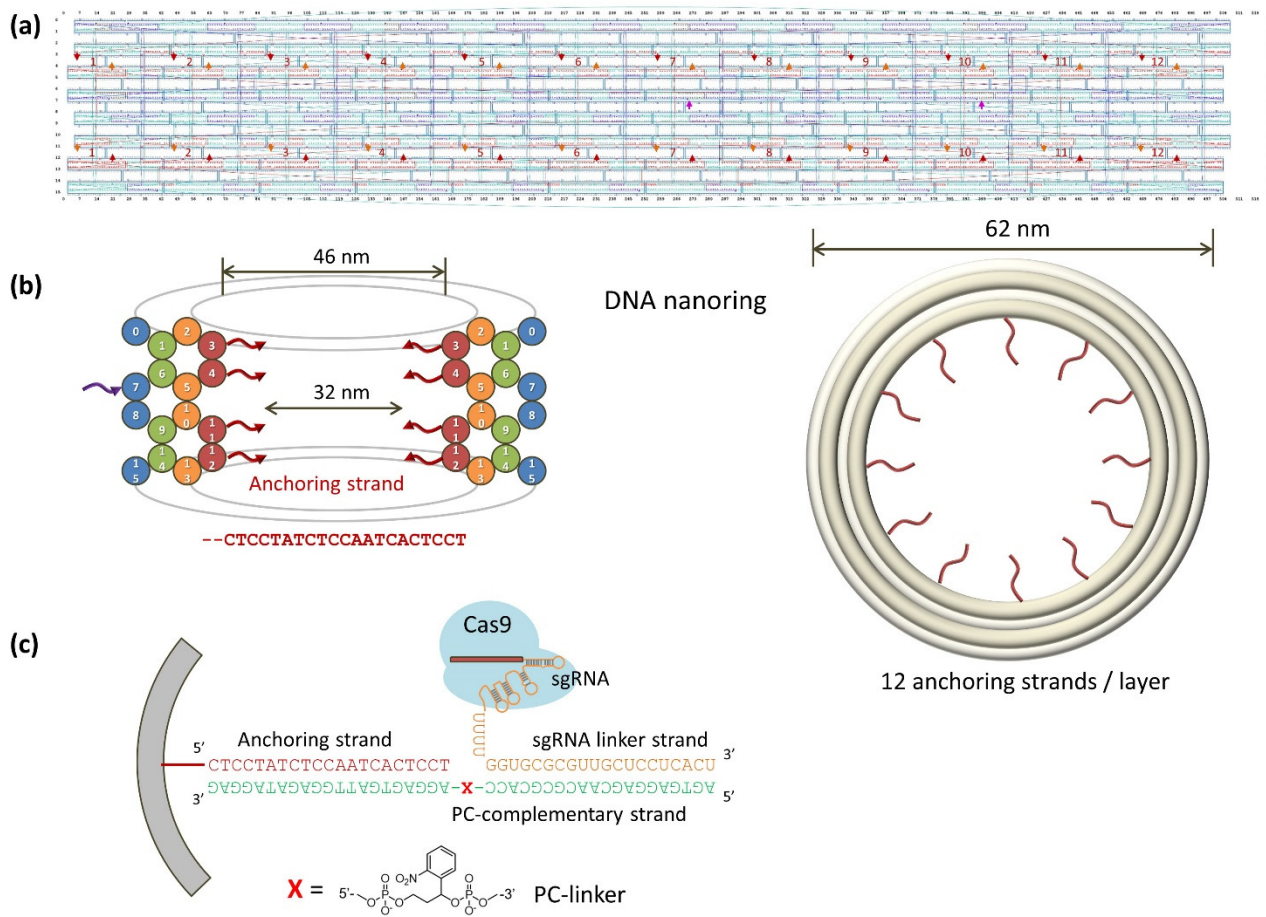
The in vitro transcription reaction was performed using 30 nM dsDNA template, 40 mM Tris-HCl (pH 8.0), 5 mM DTT, 8 mM MgCl<sub>2</sub>, 2 mM spermidine, 2 mM NTP and 0.25 μM T7 RNA polymerase (Takara) at 37 °C for 2 h. The resulting sgRNA was purified by RNA purification kit (Qiagen) and the quality was confirmed by running the sgRNA on an 8% denaturing polyacrylamide gel. Sequence of template dsDNA and sgRNA are listed in Table S2.

**Introduction of protein.** For incorporation of Cas9/sgRNA into the nanoring (Figure S1), 1 μM Cas9 and 500 nM sgRNA were incubated in a buffer containing 20 mM HEPES, 100 mM NaCl, 5 mM

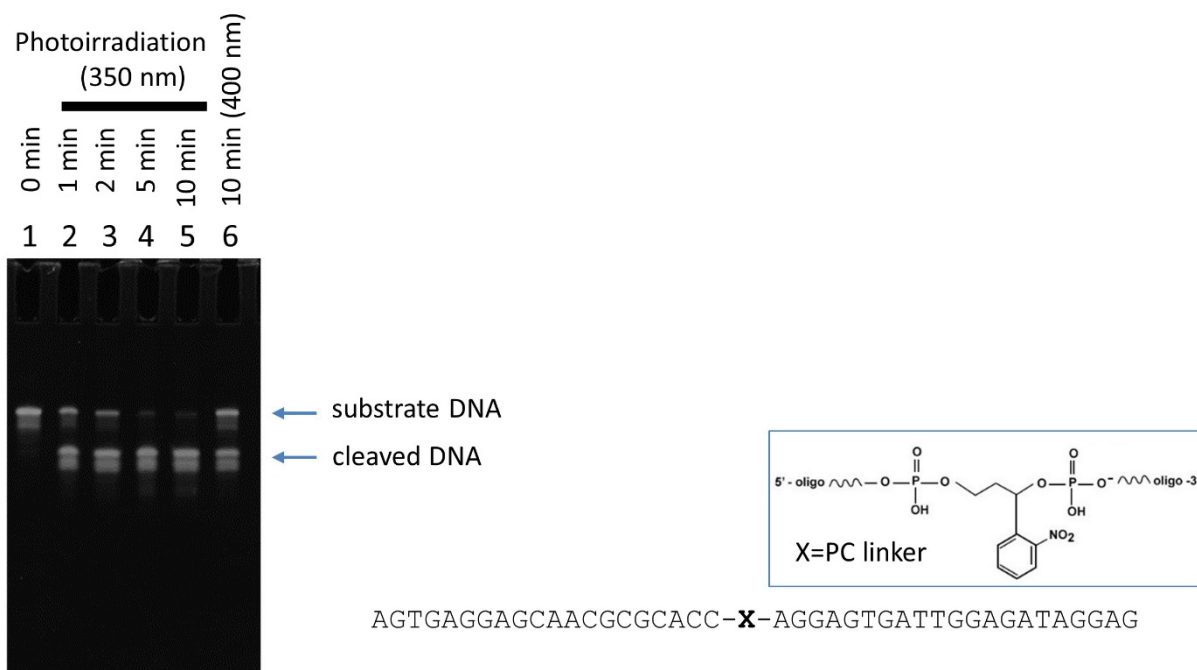
MgCl<sub>2</sub> and 0.1 mM EDTA at 25 °C for 10 min. A solution of 250 nM Cas9/sgRNA complex was added to a solution of 10 nM DNA nanorings with PC-linker-containing complementary strand in a buffer (10 mM HEPES, 50 mM NaCl, 5 mM MgCl<sub>2</sub>, 0.01 mM EDTA), and then incubated at 37 °C for 1 h. Excess staple DNAs were removed by gel filtration using Sephacryl-S400.

**Purification of nanoring.** For purification of a Cas9-attached nanoring, a single-stranded DNA (ssDNA, 25 nt) was introduced to the nanoring as a capture strand (Figure S2). A 50 μM of biotinylated complementary strand (16 nt) to hybridize with above 25-nt capture ssDNA was added to a mixture of streptavidin-attached magnetic beads (4 mg/mL), and the mixture was kept at rt for 5 min, then the magnetic beads was washed to remove excess biotinylated strand. The Cas9-attached nanoring with 25-nt capture ssDNA was added to the magnetic beads mixture. The mixture was kept at 25 °C for 30 min, then the beads were trapped using a magnet stand, and the remaining supernatant was removed to remove excess Cas9. After three times washing, the Cas9-attached nanoring was separated from the beads using strand displacement with 50 μM complementary ssDNA (25 nt) by incubating at 25 °C for 30 min, and the purified Cas9-attached nanoring was recovered. The sequences of DNA strands used for purification are listed in Table S3.

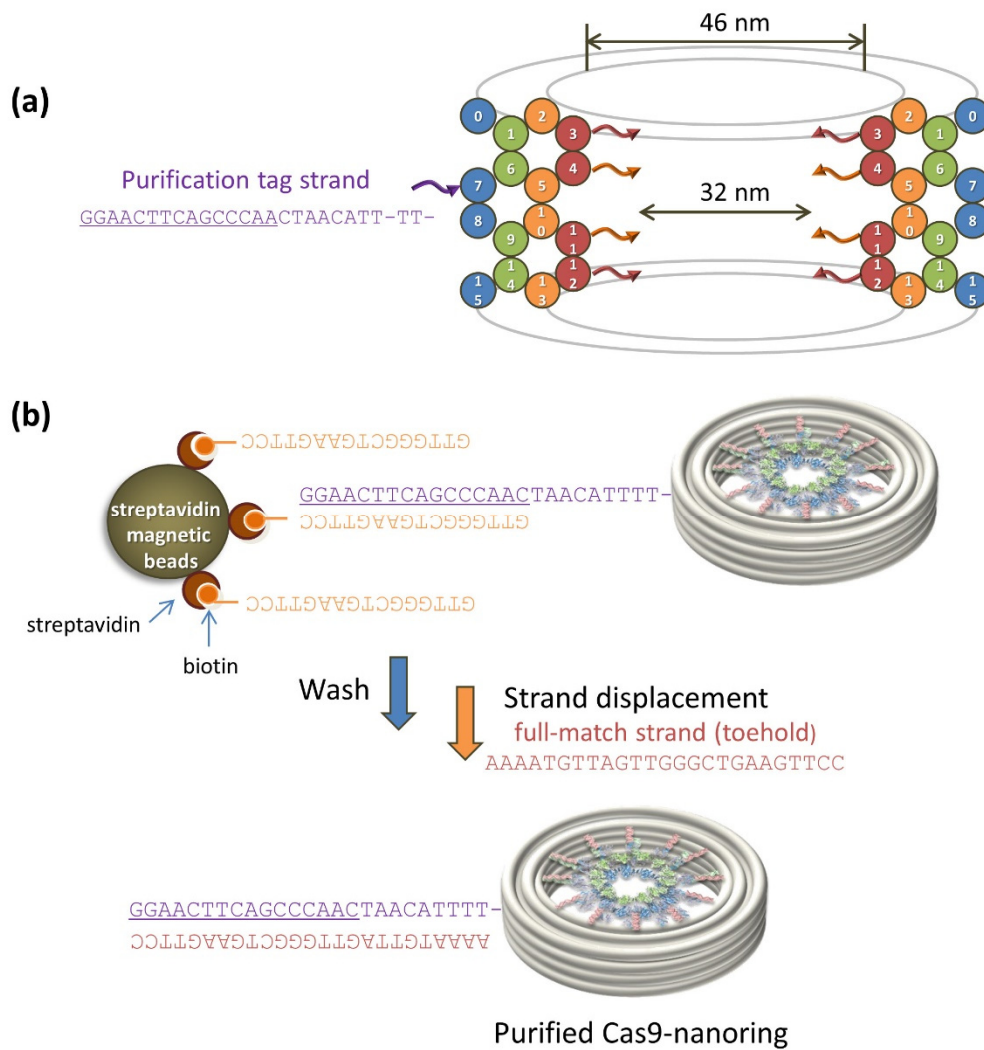
**Confirmation of the Cas-9 activity.** For photoirradiation, the purified Cas9-attached nanoring (7 nM) was irradiated with UV light (350 nm) for 10 min using Xenon Light Source (ASAHI SPECTRA MAX-303, 300W). After irradiation, PCR-amplified EGFP-coding dsDNA (850 bp, 10 nM) was added to the solution of photoirradiated Cas9-nanoring, and the mixture was incubated at 37 °C for 60 min. The reaction solution was then heated at 65 °C for 10 min to dissociate Cas9 from the substrate dsDNA, and then the decomposed stapled DNAs were removed by Sephacryl-S300 gel filtration and the reaction substrate was recovered. The cleavage reaction was observed by agarose gel electrophoresis. Sequence of a target dsDNA (EGFP gene) is presented in Table S4



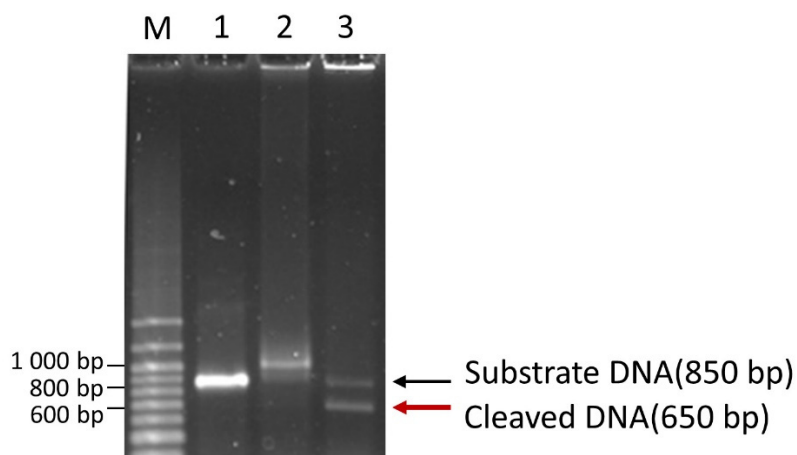
**Figure S1.** Schematic illustration of introduction of Cas9 into the nanoring. (a) caDNAno design of DNA origami nanoring. (b) Illustration of the nanoring with anchoring strands inside the cavity. Twelve anchoring strands were incorporated for one layer, and total 48 strands were incorporated. (c) Incorporation of Cas9 inside the nanoring. Linker RNA strand was added to 3'-end of sgRNA. Cas9/sgRNA with linker strand was bound to the anchoring DNA strand inside the nanoring with the PC-linker-containing complementary strand.



**Figure S2.** Image of denatured PAGE gel for investigating the efficiency of photo-cleavage reaction. Substrate DNA containing PC-linker without irradiation (lane 1), with 350 nm UV for 1 min (lane 2), 2 min (lane 3), 5 min (lane 4), 10 min (lane 5), and with 400 nm UV for 10 min (lane 6). Electrophoresis conditions: 16% acrylamide, 8 M Urea, TBE buffer, 150 V, 60 min, rt.



**Figure S3.** Schematic illustration of the purification. (a) Purification tag strands were incorporated to the outer side of the nanoring. (b) Biotinylated DNA strand was immobilized on the streptavidin magnetic beads. Purification-tag attached Cas9-nanoring was attached onto the magnetic beads to wash out excess Cas9, and then separated to recover the purified Cas9-nanoring using strand displacement.



**Figure S4.** Image of agarose gel for investigating cleavage reaction efficiency of free Cas9. Lane 1: substrate DNA only, lane 2: Substrate DNA incubated with Cas9 at 37 °C for 1 h (without denaturation to dissociate Cas9 from the substrate DNA), lane 2: Substrate DNA incubated with Cas9 at 37 °C for 1 h followed by denaturation at 60 °C for 30 min. Electrophoresis conditions: 1% agarose, TBE buffer, 100 V, 60 min, r.t. In this experiment, Cas9: DNA = 400: 24 (DNA 10 nM) was used.

After the reaction, 74% of the substrate DNA strand were cleaved. Therefore, it is considered that ideally the DNA strand should be cleaved almost completely at the ratio of Cas9: DNA = 400: 1 (DNA 10 nM) as shown in Figure 3c, if Cas9 is used as a free form without being introduced into the nanoring.

**Table S1. Staple DNA sequences for ring-shape DNA origami**

Staple	sequence
ring 001 h0-1A	TTCACTACTTACCCTCTACC-GGGCGATGTTGGGGTCGAGGTAACGCTCATGTACCGCCAGCCTTGCT
ring 002 h0-2A	TTCACTACTTACCCTCTACC-TAAAGAACCCTTCTGACCTGCCTCATATTTCAAACCACCAGTAACAT
ring 003 h0-3A	TTCACTACTTACCCTCTACC-CGAGATAGCAGAAATAAAGAACATATATTTTCAAAGAACAAATGCTG
ring 004 h0-4A	TTCACTACTTACCCTCTACC-GGTGGTCTTTTAGTATCATATTTTTAAATCATCCCGACTCTAAGAAC
ring 005 h0-5A	TTCACTACTTACCCTCTACC-AGCGGTCTTTATCCCAATCCGAGATTGAGGCCAAAGACAATAGAAA
ring 006 h0-6A	TTCACTACTTACCCTCTACC-CAACAGCTCATTAGCAAGGCCACTTCGGAACTATAAACAGGGTCAGT
ring 007 h0-7A	TTCACTACTTACCCTCTACC-TGCGTATTATATAAGTATAGCGGTATATTCGCAACAACCTTGATACC
ring 008 h0-8A	TTCACTACTTACCCTCTACC-TGCCAGCAAAGACTTTTTTACAATTATACCACCTTATGTTAATTTT
ring 009 h0-9A	TTCACTACTTACCCTCTACC-TAATTGCGAATGCAGATACATCAACTAAAGTTGTAGCTCCTTAGAGC
ring 010 h0-10A	TTCACTACTTACCCTCTACC-CATAAAGTTATATTTTCATTTCCAGATTGTAATAATCAGAGCATGTC
ring 011 h0-11A	TTCACTACTTACCCTCTACC-GAAATGTCTGGCCTTCCTGGCCTTTCTCCCCAGTCCCACGGCGA
ring 012 h0-12A	TTCACTACTTACCCTCTACC-GGTACCAGAAAATCCCGTAAAGCGTGCCTCGATCCAGCAGCGGTGC
ring 013	CGTGAACCATCCTCACAGTTGAGGATCCCCG
ring 014	CCAACGTCAAAGGGCGATTCTGGACACACGACCAGTAATGCCG
ring 015	TGTTGTTCCAGTTTGGTCAAAATGTTGGAAGGGTTAGAAAAAG
ring 016	GCAAAATCCCTTATACATAATTTATAAATAAGAATAAAATTG
ring 017	TTTGCCCGCAGCGGTACAAAACTTTCCAGAGCCTAAGCGT
ring 018	CTTACCAGCCTGGCCAAATCACGACCATTTGGGAATTTAAAT
ring 019	GGGTGGTTTTTCTTTATAAGTGCAGGGTTTTGCTCAGTGGA
ring 020	TGAATCGGCAACCGCGGCTACAAAAGCATCGGAACGAGCCGG
ring 021	CACTGCCCGCTTCCATTTAGGATAGTAGAAAGATTTCATTGAG
ring 022	TGGGGTGCCTAATGAAATGGTCTCTTGACCATTAGATAAACG
ring 023	TCACAATTCACACAAAACGCCATTGAGCTCATTTTTTAAGGGG
ring 024	TTCGTAATCATGGTCACGGTTGTTGCTCATTTGCCGCCATAGC
ring 025	TAAAGCATTACATTGGCAGGTCTGATACCTATTGCTT
ring 026	CGTAAGAGTTTGGATTATACAATATATTATCATGACGC
ring 027	CGTAGATCGACCGTGTGATTAATGGAATTTCCGACAA
ring 028	TATACAATCTTACCAACGCACAATTTAGTTGAAGACG
ring 029	AAGAAACATTATCACCGTCAATTCATGAAGGTACTTGA
ring 030	ACGTCACCAAGAGAAGGATAGAGGCTATTCTCACCAC
ring 031	AATAGGTCGTCACCCCTCAGGCCGCTCTGAGGTTAACG
ring 032	GAAGTTTCTAACGGAACAACCGTTAAAGGACGGATTGT
ring 033	CCAAAAGTCTGCGAACGAAACAGTTGTCTGTTGTCA
ring 034	CGCGAGCTCGCATTAATTAATATTTCAAATAGACGGG
ring 035	CAGCTTTAGCGGATCAAACCTAAGAGATGAAGGCCGGA
ring 036	GCCGACTACCGGGGTTTCCGTTGTGCTGCGGTCT
ring 037	AGGGAGCCCCATCTAAATCGGAACCCTCGTGCCTGTTCTTCAAAA
ring 038	TATTTTTGAATTATACGTGGCACAGCCAACAGAAAATACCG
ring 039	GATGAATATACACTTTCAGGTTTAACTATTTGCAGCTTTGAA
ring 040	GCCAACGCTCAAATTTTACCAGTAAGTAAATTTAGGC
ring 041	CAAAAATGAAAAGATTTTTTGTTTATAACAGCGGGAGAAT
ring 042	GCAGCACCGTACTCAATGAAACCATCCAGTAGCATTTTCATCG
ring 043	AGGTTTAGTAATGTATCACCGTACTCCGTCGAGACCCTCAT
ring 044	TACGTAATGCCACCATTAAACGGGTGAGGCTTTACTCATCT
ring 045	AAGAGCAACACAAGAATTACGAGGCAATACCACAGCTTTTGC
ring 046	TTCTACTAATATGAAAAGGTGGCAAATAACCTCAAAATTA



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ring 047	AGCGAGTAAACACCATCAACATTAACAAAAATAGGTCACGT
ring 048	GAAGGGTAAAGCAAGGCGCCTTTAGGTACATCGCGTCGCTG
ring 049 h3-1C	TTCGATTTAGAGCTTGATGCGGCGCTGCCAGCACGAA-CTCCTATCTCCAATCACTCCT
ring 050 h3-2C	TGGCTATTAGTCTTTAAGCTCAATCATTCACCAGTCAA-CTCCTATCTCCAATCACTCCT
ring 051 h3-3C	ATAGTAACAGTACCTTTAATTCATCTTCTGAATAACA-CTCCTATCTCCAATCACTCCT
ring 052 h3-4C	TGAACAGTAGGGCTTAACCTAAATTAATAAGGCGTAA-CTCCTATCTCCAATCACTCCT
ring 053 h3-5C	GATAGCAGCCTTTACAGACCCAGCTTAACGAGCGTCGT-CTCCTATCTCCAATCACTCCT
ring 054 h3-6C	TCATCAGTAGCGACAGACGGAAATTCGACTTGAGTA-CTCCTATCTCCAATCACTCCT
ring 055 h3-7C	GGCCGCCACCCTCAGAAAAGTATTATAGGATTAGCGGG-CTCCTATCTCCAATCACTCCT
ring 056 h3-8C	AACTACGAAGGCACCAAAGTTAAAGCAGCGAAAGACAA-CTCCTATCTCCAATCACTCCT
ring 057 h3-9C	CCTATCATAACCCTCGTAAATCTAATTATTACAGGT-CTCCTATCTCCAATCACTCCT
ring 058 h3-10C	AAAGTAGTAGCATTAACTTCCATATGTAGATTTAGTAA-CTCCTATCTCCAATCACTCCT
ring 059 h3-11C	AACCCGTCGGATTCTCCTAAACGTTTTTGTAAATCTG-CTCCTATCTCCAATCACTCCT
ring 060 h3-12C	GTTTAAACGATGCTGATCAGGAAATAAATTTCTGAT-CTCCTATCTCCAATCACTCCT
ring 061	CGAAATGCGCGCCTGTCCGTGAGCCTCACCCAAATCGAAGAAAG
ring 062	AACGACAACCTGCGGGAAGTGAATGACAGTATTACAAATCAA
ring 063	TACCAGAGCTACATCGGGAGAAATTAATTTACCTTGAGTGAA
ring 064	AGAGGTTGATTGAGAATCGCGTTCAGACAAAAGTCTTAATT
ring 065	TAACTTTGCAGAGAATAACATAAGAGCTAATATCCGAACAAA
ring 066	GCATTGAGAATCAAGTTTGCCCTCATTCATAATACCAGAGC
ring 067	TTTACCGACCGCCACCCTCCAGACAACCTGAGTTTCTGTATG
ring 068	TTGACCAGGCTAAAACGAACCTGCTACGGAGATAAAGAGGA
ring 069	AAAAGAAGATTACCAGACGAATCCCCACTGGATAGACTATTA
ring 070	AGCAAAACAATCCAATAAATCAACGCAAAACATTAGATTCAA
ring 071	TGGTGAATGGTGGGAACAAACAGCTTTGAGGGGATGGGAAGG
ring 072	GCAGCAGCTTGCCGTTCCGGGCAAGAGGACTTGTATCAGCG
ring 073	AACAATATGAGAGAAAGGAAGGAGTTTTGCCCCTA
ring 074	AACAAAGCTCATCGCCATTAAGATAGAACGTGGACT
ring 075	TCGCAAGAAGATTTCGCTGATTTCGTAAGGTTGAG
ring 076	AGCGAACCGCGCAACATGTAAAGCCTGCGAAATCG
ring 077	ACCAGCGGAAGCGCATTAGACCATATTACACGCTGG
ring 078	AGTGCCCGCTACTGTAGCGCTCCATTACGATTGCC
ring 079 tag1	GGAACTTCAGCCCAACTAACATTTT-GACAATGAGTCTCAGAGCCACCAGGGTTGGGGCGCCA
ring 080	GTGAATTAGATACTAAACGAGGACTTGCAATTAA
ring 081	TATAATGCACAAATAGCGAGAGTTCAACTTTGCGCT
ring 082 tag2	GGAACTTCAGCCCAACTAACATTTT-CCCGGTTGTAGCAAAGAATTAGGTTTAGCGTAAAGCC
ring 083	ATGTTTAGTCGTAATGGGATAATTGCGGTTATCCGC
ring 084	CATCAGACTGGTTTTTTTCTGTCTACATAAAGCTCGAA
ring 085 h15-1B	GGTAATATCCACACCCCGCGCATCAGTGAGCCAGAATCCTGAGA-TTCCTCTACCACCTACATCAC
ring 086 h15-2B	TATCATTCTGCAACAGTGCCCGATTAGAGTATCTTTAGGAGCACT-TTCCTCTACCACCTACATCAC
ring 087 h15-3B	ATGCAATAGAAGATGATGAACTAGCGATAGATTTCCCTTAGAATC-TTCCTCTACCACCTACATCAC
ring 088 h15-4B	GCGAGGGGATTTCTGTCCAGACTCATTAAACCTCTTCCCTTATCATT-TTCCTCTACCACCTACATCAC
ring 089 h15-5B	ATTCATATAACCACAAGAACAATTAAGACAACGGAATACCCAAAA-TTCCTCTACCACCTACATCAC
ring 090 h15-6B	GCCTTGAGACCGGAACAGAGGAAATAAATAGACGATTGGCCTTGA-TTCCTCTACCACCTACATCAC
ring 091 h15-7B	GATAGTTGACGTACAACTACGCGGAATTGCGAGTGAGAATAGAAA-TTCCTCTACCACCTACATCAC
ring 092 h15-8B	AACTTTAATCGCCTGATAAAACGACAAGAATGGCTGACCTTCATCA-TTCCTCTACCACCTACATCAC
ring 093 h15-9B	TTAATTGCATACTGCGGAATCTTATCGGTTTTAAGAGGAAGCCCGAA-TTCCTCTACCACCTACATCAC
ring 094 h15-10B	AATCATATTGTAATACTTTTGGATGATAAATATCAATATGATATTC-TTCCTCTACCACCTACATCAC
ring 095 h15-11B	AACGTACAGTATCGGCCTCAACGCGATTAAGCTGGCGAAAGGGGA-TTCCTCTACCACCTACATCAC

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ring 096 h15-12B CGGTGCCCCAGCGTGGTGTGGCACTGTTGCGGGCGCGGTTGCGGTA-TTCCTCTACCACCTACATCAC

ring 097 ACTATCGGCCATTGCAAACCACCAGCAGAAGATAACGG

ring 098 AAAGTTTGAGAAGGAGCGAGTTACAAAATCGCGCAGAAA

ring 099 CTATATGTGCGAGAAAACCATTTTCGAGCCAGTAATATT

ring 100 CGGTATTTGCGGGAGGTGAACACCCTGAACAAAGTTTT

ring 101 TCACAATCAAAGGGCGATTTCGGTCATAGCCCCCTAAC

ring 102 TTTTAAACGGTTAATGCCCGGGATAGCAAGCCCAATAGCC

ring 103 AAACAGCATCGCCACGCCCCAGCGATTATACCAAATT

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ring 105 GCGGATGGAACATGTTTTTAAAGCCTCAGAGCATAAAAC

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ring 108 CTGCAGCCGCGAGTGTACGGAGCGGGCGCTAGGGCGCAG

ring 109 AATGCGCTGTAGCAATACTCACGCAACCGAG

ring 110 ACATCACCATTTTGGGAAAAAAGGGCAAAAACCGTCTATCA

ring 111 TGAGAGCTTTACAAACAATTTTAGAACAATAG

ring 112 AATCCTTGATGATGATTATCATAACCATAAAACAAGAGTCCACTAT

ring 113 ACATCAAATCATAGGTCTATAGTGGATTAA

ring 114 AACCTCCATCTTCTTTTCAAACCGGAATAATCAAAGAATAGCC

ring 115 ACAATAAATCGTAGGAATCAGCCGTACCGCA

ring 116 GCAAGCACTATTTTTGAAGCCTGCCAGTAAAATCCTGTTTGAT

ring 117 GTTAAGCATAAAGGTGGCAAGTAGAATATTAC

ring 118 CAAAGACAAATATTTTCAAACCGCCAGCATGAGAGAGTTGCAGCA

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ring 123 GTCGAAATCAGTGAATAAGGAACAAAATATTC

ring 124 AACACCATTGGGAACTGGCTCGTTGAGAGTCGGGAAACCTGTCC

ring 125 TAAATATACAGGTCAGGATCCGGAATTCGAG

ring 126 TTGCTCCGAAGTTTATATGCATTTTCGAGTGAGCTAACTCACAT

ring 127 AGAAGCCTATCAGGTCAATAGAGATGCCGGA

ring 128 CAAACAATTTAAATAACAGGAAATAGGACATACGAGCCGGAAG

ring 129 GATCGCACGACGCCAGTGCCACGACGGTAAC

ring 130 GGAGCCGATAGCTAGATAGAAGTTGGGTAGCTGTTTCTGTGT

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ring 132 TCGTTAACGGCCAGAACGGGGAAGCCGGGCGCGTGCAAA

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ring 134 h4-2C TCTAAAGCATCCCAGCAGCATAGCCCTAAAAGAATCCTGA-CTCCTATCTCCAATCACTCCT

ring 135 h4-3C CATTTAACAAATAAGAAAACAAAACAATAACGGTTTTTGAA-CTCCTATCTCCAATCACTCCT

ring 136 h4-4C TGCAGAACGCTCACAACATCATATTTAACAAATTTATCC-CTCCTATCTCCAATCACTCCT

ring 137 h4-5C AAACAATGAAACCCAATAATAAAAACAGGGAGGTAAAGGT-CTCCTATCTCCAATCACTCCT

ring 138 h4-6C CGCCACCCTCACCACCGCTCTTTAGCGTCAGATTGAGAC-CTCCTATCTCCAATCACTCCT

ring 139 h4-7C TCATAGTTAGCTCATTTCCAAGAACCGCCACCCGTTTGGC-CTCCTATCTCCAATCACTCCT

ring 140 h4-8C GTTACTTAGCCTTCCGCGAAGAGGCAAAAAGATCTAAAACG-CTCCTATCTCCAATCACTCCT

ring 141 h4-9C AATGCTTTAAACATCATTTGACGATAAAAACCAGGTGATTC-CTCCTATCTCCAATCACTCCT

ring 142 h4-10C ATAAAAATTTGCTTTATTTTCATACAGGCAAGAGTTGTTA-CTCCTATCTCCAATCACTCCT

ring 143 h4-11C GCACCGCTTCTACTCCAGCCGGCGGATTGACGGCGCAGAA-CTCCTATCTCCAATCACTCCT

ring 144 h4-12C CAACGGCAGCAAAGCAACCCAACCGGGTCTGTTTCAG-CTCCTATCTCCAATCACTCCT

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ring 145 h11-1C	CCTTGACGAGCACGTATAAAGGTTTTGGTGTGTTCAAC-CTCCTATCTCCAATCACTCCT
ring 146 h11-2C	AACCTTGCTGAACCTCACTGTCCATTCCTTTGATTAGAA-CTCCTATCTCCAATCACTCCT
ring 147 h11-3C	TCTTCATTTGAATTACCTTGAGGATCGACAACTCGTA-CTCCTATCTCCAATCACTCCT
ring 148 h11-4C	TTGCCTGTTTATCAACAAAGAGTCAGAGAGACTACCAA-CTCCTATCTCCAATCACTCCT
ring 149 h11-5C	ATAGCAATAGCTATCTTAAACAAGCAATTACCGCGCCAG-CTCCTATCTCCAATCACTCCT
ring 150 h11-6C	TAGAACC GCCACCCTCATAGCAAACCATATAAAAGGC-CTCCTATCTCCAATCACTCCT
ring 151 h11-7C	GGCGTAACGATCTAAAGGAAAGCGGTCATACATGGCC-CTCCTATCTCCAATCACTCCT
ring 152 h11-8C	AGGAACGAGGCGCAGACTTGAAAATTTTAAATTGTATAT-CTCCTATCTCCAATCACTCCT
ring 153 h11-9C	TCCAGTTCAGAAAACGAATCAACGCTTTGCCCTGACA-CTCCTATCTCCAATCACTCCT
ring 154 h11-10C	AGTTAGAACCCTCATATGAACCAGATAGAGAGTACCGG-CTCCTATCTCCAATCACTCCT
ring 155 h11-11C	AGGTGCCGGAACAGGTATTTTTGGCCTGAGAGTCCG-CTCCTATCTCCAATCACTCCT
ring 156 h11-12C	TACCGTCGGTGGTGCCATTCCCAGTCAAGCTTTCAGC-CTCCTATCTCCAATCACTCCT
ring 157	TAAAAGATTTTATACTGAGTAGAAGAGTAATAATA
ring 158	ATAATACACTAATACGAACGTTATATCATTATTA
16HB ring 159	GACGCTGAAAACATTAGGTTGGGTTACGCTTTTTT
16HB ring 160	CTCATCGAACGGGTAGATATAGAAGGCGCACATA
16HB ring 161	GCAGTATTGGCATGCGGAATAAGTTTTTGAAAACG
16HB ring 162	GCCAGAACACAAACAGTGTACTGGTAACGCCTTTT
16HB ring 163	TTTTTCAAATAAATTTTCGAGGTGAACACCCGGTT
16HB ring 164	ATTACCCTAATCTTGAGTAGTAAATTAGCTCGAGA
16HB ring 165	CTTCAAATTCAAATGATAAGAGGTCACAAATTTAA
16HB ring 166	GAGGGTATTCTAGCATCGATGAACGGTTTCTGGAG
16HB ring 167	GCCAGGGCTGCAAGGGGAACGGATAACCATGAGGT
16HB ring 168	GGCCGAGCCGGGTCATCAGATGCCGGCACGCTGCGCAAACAGGA
16HB ring 169 h12-1C	GTAATGGGTAACGTGCTTTCCTCGTTAGAATCCCAATTAA-CTCCTATCTCCAATCACTCCT
16HB ring 170 h12-2C	CAGTTGTGTAATATCAAACCCTCAATCAATATGTGTATTAG-CTCCTATCTCCAATCACTCCT
16HB ring 171 h12-3C	TAACCCAAATTTTTTAAATGAAACAGTACATATAAATTTA-CTCCTATCTCCAATCACTCCT
16HB ring 172 h12-4C	TACGATGAGATAGATAAGTCTGAACAAGAAAGTTTTTAT-CTCCTATCTCCAATCACTCCT
16HB ring 173 h12-5C	GTTACAGAGACCGAAGCCCTTTTTAAGAAAAGCTAATACAT-CTCCTATCTCCAATCACTCCT
16HB ring 174 h12-6C	CGCCGACGTGAGCCACCACCCTCAGAGCCGCTCTGAATT-CTCCTATCTCCAATCACTCCT
16HB ring 175 h12-7C	GGATTTTTGTTTTGTGCTCTTCCAGACGTTAATAAAAAA-CTCCTATCTCCAATCACTCCT
16HB ring 176 h12-8C	CAGATACCGGGTCAATCATAAGGGAACCGAACGGGCTGCTC-CTCCTATCTCCAATCACTCCT
16HB ring 177 h12-9C	TAGTCAGAAGAATGACCATAAATCAAAAATCAAAGCAAAC-CTCCTATCTCCAATCACTCCT
16HB ring 178 h12-10C	AAGGGACGATTTTAAATGCAATGCGCTGAGTAATCTACAA-CTCCTATCTCCAATCACTCCT
16HB ring 179 h12-11C	GCGATCGGCCAAAGCGCCATTGCGCATTGAGGTGGTTGTAA-CTCCTATCTCCAATCACTCCT
16HB ring 180 h12-12C	GGGTCTGTTTCCCACGCAACCAGCTTACGGCTTGTGCTCA-CTCCTATCTCCAATCACTCCT
16HB ring 181	GGGAGCTGTAACCACCTGGCAAGTGTAGCGGTGTTAC
16HB ring 182	CAGTTGGACACCGCTTGAACAGAGGTGAGGCGACTCAA
16HB ring 183	ATATATGGAGCAAACCGGCGAATTATTCATTATTTTA
16HB ring 184	TATCCCATAAAGTATTAGAGAATATAAAGTACTATAA
16HB ring 185	AGATAGCAGAGAGATGGCAGAGGGTAATTGAGCTTATC
16HB ring 186	AACCACCCAAAATCTATTAGCGTTTGCCATCATTTTG
16HB ring 187	TGAATTTTCGTCACCGGAACCCATGTACCGTAATAAG
16HB ring 188	AACTTTGTGTATCATCGCGCAAACAAAGTATTTCTT
16HB ring 189	TTACCCTGCGTCCATGAATAGTAAATGTTGGGCTT
16HB ring 190	TAGGTAATGACCCGTGCTAAATCGGTTGTACTTTTT
16HB ring 191	CAACTGTCGACGACAGCCCGTGCATCTGCCAGTAATCG
16HB ring 192	TGTCCAGAGAACGCTCTCATACGGAACGTCCTCAC
16HB ring 193	AGTGAAAGGAATTGAGGAAGGTTATCTAAAACCTGGT

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16HB ring 194	AATTGCTTCTGTAATCGTCGCTATTAATTACTAATCA
16HB ring 195	CTGCATGTAGAAACCAATCAATAATCGGCTGAAAATAA
16HB ring 196	CCACAGAAGGAAACCGAGGAAACGCAATAATTCTAAGC
16HB ring 197	GACCAGCATTGACAGGAGGTTGAGGCAGGTCCCACCAG
16HB ring 198	TATTGCTAAACAACCTTCAACAGTTTCAGCGGAGTAAA
16HB ring 199	GGAGAACGGTGTACAGACCAGGCGCATAGGCCCTGACC
16HB ring 200	AGAGAAGCAAAGCGGATTGCATCAAAAAGATTTGGTCT
16HB ring 201	AGTGAGAAAGGCCGGAGACAGTCAAATCACCTAATGTG
16HB ring 202	AACCGGTGCGGGCCTCTTCGCTATTACGCCAGTCTGCG
16HB ring 203	TGATTGCAGGCGCTTTCGCACTCAATCCGCCCGGAGG
16HB ring 204	TGATTAAGGGATTTTAGACAGGAACGGTACGGAGAGC
complementary strand A	GGTAGAGTGGTAAGTAGTGAA
complementary strand B	GTGATGTAGGTGGTAGAGGAA
complementary strand C	AGGAGTGATTGGAGATAGGAG

**Table S2.** Sequences of sgRNA and temple dsDNA.

	sequence
sgRNA + connector	GGGUUUUAGAGCUAGAAUAGCAAGUUAAAAUAAGGCUAGUCCGUUAUCAACUUGAAAAAGUGGCACCGA GUCGGUGCUUUUGGUGCGCGUUGCUCUCACU
T7 promoter +	TTCTAATACGACTCACTATA-
sgRNA template + connector top 84 nt	GGGCACGGGCAGCUUGCCGGGTTTTAGAGCTAGAAATAGCAAGTTAAAATAAGGCTAGTCCGTT
T7 promoter +	AGTGAGGAGCAACGCGCACC-AAAA-
sgRNA template + connector bottom 80 nt	GCACCGACTCGGTGCCACTTTTTCAAGTTGATAACGGACTAGCCTTATTTTAACTT
Forward primer for template	TTCTAATACGACTCACTATAGG
Reverse Primer for template	AGTGAGGAGCAACGCGCACCAA

**Table S3.** DNA sequences of biotinylated strand and fully complementary strand for purification.

Staple	sequence
biotinylated strand	GTTGGGCTGAAGTTCC [BIOTEG]
complementary strand	AAAATGTTAGTTGGGCTGAAGTTCC

**Table S4.** Substrate sequence (EGFP complementary strand, 714 bp)

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GTACAGCTCGTCCATGCCGAGAGTGATCCCGGCGGGCGGTCACGAACTCCAGCAGGACCATGTGATCGCGCTTCTCGTTGGGGTCTTTG  
CTCAGGGCGGACTGGGTGCTCAGGTAGTGGTTGTCGGGCAGCAGCACGGGGCCGTCGCCGATGGGGGTGTTCTGCTGGTAGTGGTCGG  
CGAGCTGCACGCTGCCGTCCTCGATGTTGTGGCGGATCTTGAAGTTCACCTTGATGCCGTTCTTCTGCTTGTCGGCCATGATATAGAC  
GTTGTGGCTGTTGTAGTTGTACTIONCCAGCTTGTGCCCCAGGATGTTGCCGTCCTCCTTGAAGTCGATGCCCTTCAGCTCGATGCGGTTTC  
ACCAGGGTGTGCCCTCGAACTTCACCTCGGGCGGGTCTTGTAGTTGCCGTCGTCCTTGAAGAAGATGGTGGCGCTCCTGGACGTAGC  
CTTCGGGCATGGCGGACTTGAAGAAGTCGTGCTGCTTCATGTGGTCGGGGTAGCGGCTGAAGCACTGCACGCCGTAGGTCAGGGTGGT  
CACGAGGGTGGGCCAGGGCACGGGCAGCTTGCCGGTGGTGCAGATGAACTTCAGGGTCAGCTTGCCGTAGGTGGCATCGCCCTCGCCC  
TCGCCGGACACGCTGAACTTGTGGCCGTTTACGTGCGCGTCCAGCTCGACCAGGATGGGCACCACCCCGGTGAACAGCTCCTCGCCCT  
TGCTCACCAT

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The underline presents the target sequence.

## Reference

1. P. Ketterer, A. N. Ananth, D. S. Laman Trip, A. Mishra, E. Bertolin, M. Ganji, J. van der Torre, P. Onck, H. Dietz and C. Dekker, *Nature Communications*, **2018**, *9*, 902.