# **Electronic Supplementary Information (ESI)**

# Mimicking reactive high-valent diiron- $\mu_2$ -oxo intermediates of nonheme enzymes by an iron tetracarbene complex

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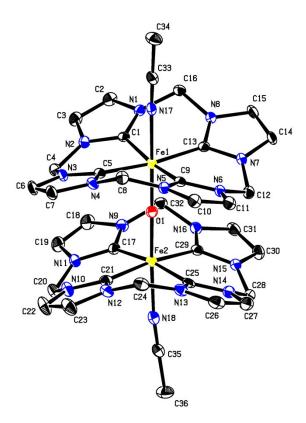
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#### 1. General Considerations

**Experimental Remarks.** Unless otherwise stated, all manipulations were carried out under an argon atmosphere using standard Schlenk and glovebox techniques. Dry, degassed acetonitrile was obtained from an Mbraun solvent purification system. Acetonitrile- $d_3$  was dried over phosphorous pentoxide, distilled, and stored over molecular sieves prior to use. All solvents were degassed by using at least three freeze/pump/thaw cycles. The diiron(III)-oxo complex **2** and thianthrenyl hexafluorophosphate were synthesized according to known literature procedures.<sup>1, 2</sup> All other reagents were purchased from commercial suppliers and used without further purification. NMR spectra were recorded on a Bruker Advance DPX 400 (<sup>1</sup>H NMR, 400.13 MHz; <sup>13</sup>C NMR, 100.53 MHz). Chemical shifts are reported in ppm and referenced to the residual signal of the deuterated solvent (acetonitrile- $d_3$ ; 1.94 ppm). Investigations concerning the magnetic susceptibility in solution were performed via Evans Method.<sup>3, 4</sup>

UV/Vis-spectra were recorded from liquid samples using a JASCO V-500 spectrometer. Elemental analyses (C/H/N) were obtained by the microanalytical laboratory at the Technische Universität München. Electrospray (ESI) mass spectrometry (MS) data were carried out on an LCQ-Fleet by Thermo Scientific. CV measurements were performed at -40 °C on a GRAMY reference 600 potentiostat and a DAQ electrochemical reaction vessel (3 mL) was used as electrochemical cell. Platinum electrodes were used as working/ counter electrodes, Ag/AgCl (3.4 M in KCl) was used as reference electrode (all from eDAQ) and ferrocene was used as internal standard. The potential was scanned with scan rates of 50 mV/s to 400 mV/s.

#### 2. Single crystal X-ray diffraction:



**Fig. S1**: ORTEP-style drawing of the cationic fragment of complex **3**. Hydrogen atoms, cocrystallized solvent molecules and five  $PF_{6}$  anions are omitted for clarity and thermal ellipsoids are shown at a 50% probability level.

Selected bond lengths (Å) and angles (°): Fe1-C1: 1.929(2), Fe1-C5: 1.924(3), Fe1-C9: 1.931(2), Fe1-C13: 1.939(3), Fe1-O1: 1.7272(17), Fe1-N17: 1.984(2), N17-C33: 1.138(3), Fe2-C17: 1.929(3), Fe2-C21: 1.931(3), Fe2-C25: 1.931(3), Fe2-C29: 1.929(3), Fe2-O1: 1.7298(17), Fe2-N18: 1.987(2), N18-C35: 1.141(3), C1-Fe1-C5: 91.72(9) C5-Fe1-C9: 90.29(10), C9-Fe1-C13: 89.52(10), C1-Fe1-C13: 89.63(10), O1-Fe1-N17: 178.09(8), C17-Fe2-C21: 89.96(11) C21-Fe2-C25: 89.01(11), C25-Fe2-C29: 90.39(11), C17-Fe2-C29: 89.29(11), O1-Fe2-N18: 177.18(9), Fe1-O1-Fe2: 176.34(11)

Single crystals of **3** suitable for X-ray diffraction were obtained by slow diffusion of diethyl ether into an acetonitrile solution at -40 °C.

Substance identification	Complex	
CCDC	2077633	
Empirical formula	C <sub>42</sub> H <sub>47</sub> F <sub>30</sub> Fe <sub>2</sub> N <sub>21</sub> OP <sub>5</sub>	
Fw [g mol <sup>-1</sup> ]	1689.55	
T [K]	123(2)	
Crystal system	triclinic	
Space group	<i>P</i> -1	
a [Å]	12.1708(3)	
b [Å]	12.6323(3)	
c [Å]	23.6169(5)	
α [deg]	29.9980(10)	
β [deg]	93.5180(10)	
γ [deg]	115,2720(10)	
V [ų]	3264.86(13)	
Ζ	2	
Density (calcd) [g cm <sup>-3</sup> ]	1.728	
μ [mm <sup>-1</sup> ]	0.706	
F (000)	1702	
Crystal size (mm <sup>3</sup> )	0.16 × 0.27 × 0.33	
heta range for data collection	1.98 to 25.41	
[deg]		
Reflections collected	84113	
Independent reflections	11929	
	[R(int)= 0.0405]	
Data/restrains/parameters	11929 / 81 / 983	
GOF on F <sup>2</sup>	1.024	
Final R1	R1 = 0.0558,	
wR2 [I > $2\sigma(I)$ ]	wR2 = 0.0868	
Largest diff. peak and	0.631 and	
hole [eÅ-3]	-0.348	

 Tab. S1: Crystallographic data of complex 3.

## 3. Experimental Data

#### Bis-µ<sub>2</sub>-oxo[calix[4]imidazoyl]iron(III)/(IV) hexafluorophosphate (3)

Bis- $\mu_2$ -oxo[calix[4]imidazoyl]iron(III) hexafluorophosphate **2** (0.100 g, 0.074 mmol, 1 eq.)<sup>2</sup> was dissolved in 2 mL acetonitrile and cooled to -40 °C. An equimolar amount of thianthrenyl hexafluorophosphate (0.027 g, 0.074 mmol, 1 eq.) in acetonitrile was added. The burgundy solution was stirred for 15 minutes at -40 °C and precipitated with diethylether. After drying *in vacuo*, the product was obtained as burgundy powder (0.099 g, 0.063 mmol, 85%). <sup>1</sup>**H-NMR** (400 MHz, CD<sub>3</sub>CN, 296 K): no signals observable.

Anal. Calcd. for  $C_{36}H_{38}F_{30}Fe_2N_{18}OP_5$ : C 27.45; H 2.43; N 16.00. Found: C 27.41; H 2.54; N 15.76.

# 4. UV/Vis-Spectra

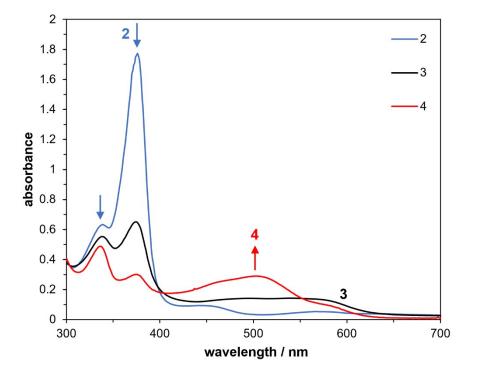
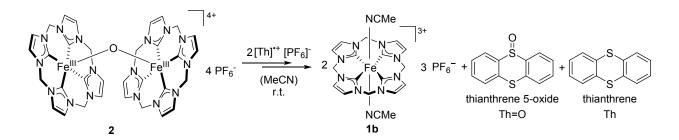


Fig. S2: UV/Vis spectrum of 2, 3, and 4. T = -40 °C, c =  $3.5 \cdot 10^{-4}$  M in dry and degassed MeCN.

#### 5. <sup>1</sup>H-NMR-spectra in the paramagnetic region

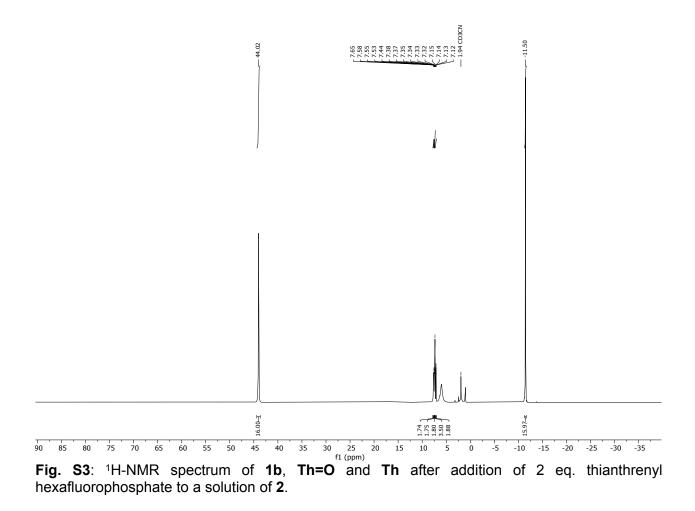


15 mg of diiron(III)-oxo complex **2** (0.011 mmol, 1 eq.) were dissolved in ~0.5 mL dry and degassed acetonitrile- $d_3$  in a dried *J*-Young NMR tube. After addition of thianthrenyl hexafluorophosphate (8 mg, 0.022 mmol, 2 eq.), the mixture was vigorously shaken resulting in an instantaneous color change from dark green (**2**) to reddish-purple indicating the formation of **1b**.

In the <sup>1</sup>H-NMR spectrum (Fig. S3) the signals of **1b** are located at 44.02 ppm and -11.50 ppm. The first is representing the  $CH_2$ -bridge and the latter the CH-backbone of the imidazole units of the tetracarbene ligand. They are both in a 1:1 ratio. The peaks fit well with a blank spectrum of **1b** taken at 293 K (Fig. S5). Fig. S6 furthermore shows the signals of **1b** at various temperatures from -40 °C to 50 °C.

Fig. S4 depicts the area between 8.00 ppm and 6.90 ppm of Fig. S3; the signals of both Th=O and Th are present. The signals of Th can be assigned to the peaks at 7.14 and 7.35 ppm, where the latter is overlapping with one signal of Th=O. The remaining Th=O signals are at 7.65, 7.57 and 7.44 ppm. The integrals of Th=O are very close to each other in a 1:1:1:1 ratio. The chemical shifts of both Th and Th=O match well with those reported in the literature.<sup>5, 6</sup>

<sup>1</sup>**H-NMR** (400 MHz, CD<sub>3</sub>CN, 296 K): δ 44.02 (s, 16H, **1b**, C*H*<sub>2</sub>), 7.65 (m, 2H, Th=O, C*H*), 7.57 (m, 2H, Th=O, C*H*), 7.44 (m, 2H, Th=O, C*H*), 7.35 (m, 6H, Th=O (2H), Th (4H), C*H*), 7.14 (m, 4H, Th, C*H*), -11.50 (s, 16H, **1b**, C*H*).





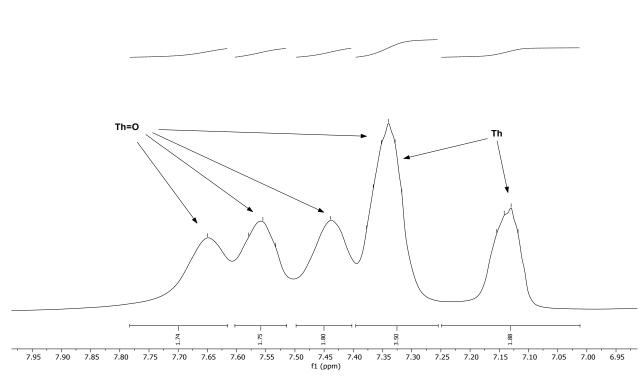
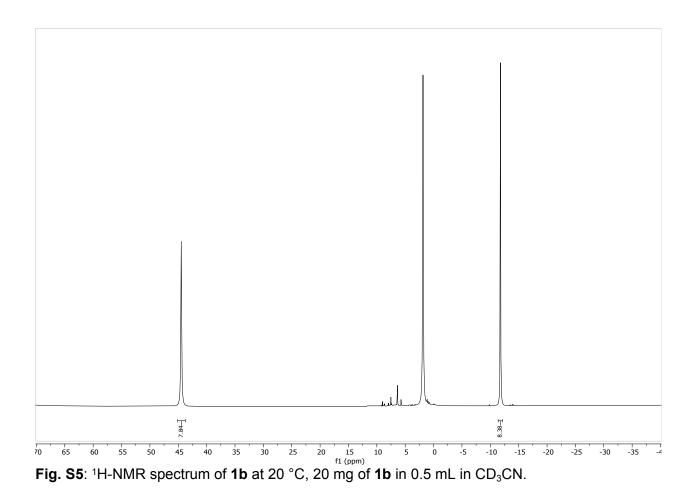


Fig. S4: Section from 8.00 to 6.90 ppm of Fig. S3.



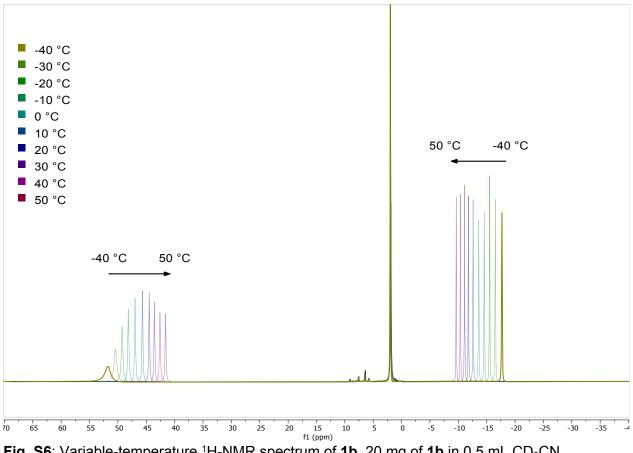


Fig. S6: Variable-temperature <sup>1</sup>H-NMR spectrum of 1b. 20 mg of 1b in 0.5 mL CD<sub>3</sub>CN.

#### 6. Evans' Method

The following formula were used to calculate the magnetic susceptibility  $X_m$  (neglecting the solvent correction term) and the magnetic moment  $\mu$ , with  $\Delta f$  being the shift in frequency [Hz],  $f_0$  the operating frequency of the NMR spectrometer [Hz],  $S_f$  the shape factor of the magnet ( $S_f = (4\pi)/3$  due to cylindrical sample in superconducting magnet) and c with the concentration of the sample [mol/cm<sup>3</sup>].<sup>3, 4, 7, 8</sup>

$$X_m = \frac{\Delta f}{f_0 S_f c}$$

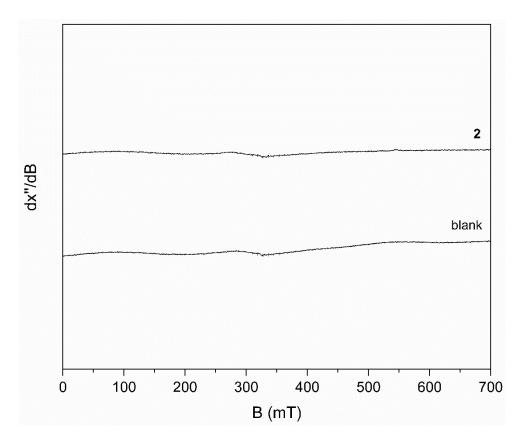
 $\mu = \sqrt{8(X_mT)}$ 

Tab. S2: Data of Evans-Method for 2 and 3.

	2	3
f <sub>0</sub> [MHz]	400	400
m [mg]	10.4	14.7
M [g/mol]	1348.27	1575.34
m <sub>solvent</sub> [g]	0.4	0.526
<u></u> δ [g/mL]	0.844	0.844
V [mL]	0.474	0.623
c [mol/mL]	1.628 · 10 <sup>-5</sup>	1.498 · 10 <sup>-5</sup>
Т [К]	233.15	233.15
original solvent peak [ppm]	2.3	1.99
shifted solvent peak [ppm]	2.3	1.89
∆f [ppm]	0	0.1
∆f [Hz]	0	40
X <sub>m</sub> [cm <sup>3</sup> /mol]	0	159.388 · 10 <sup>-5</sup>
μ [μB]	0	1.724

#### 7. EPR Spectroscopy

X-band EPR spectra (approx. 9.27 GHz) were recorded at 173 K in a frozen solution of MeCN using a JEOL JES-FA 200 EPR spectrometer equipped with a variable temperature control unit ES-DVT4 and cooling with liquid nitrogen. The microwave frequency was measured via an Advantest R5372 microwave frequency counter. Measurements were performed with a modulation frequency of 0.4 mT, a receiver gain of 6·100 and a microwave power of 5 mW. The values for g and A were determined using a  $Mn^{2+}$  (I = 5/2) standard embedded in MgO. The fourth low field line is taken as reference signal set to g = 1.981. The measured EPR spectra were simulated using EasySpin 5.2.28<sup>9</sup> and MatLab R2019b<sup>10</sup> for determining the presented g and A values.



**Fig. S7**: EPR X-band spectra of complex **2** (top, 173 K, 4.6·10<sup>-4</sup> mol/L) and of the blank EPR tube (bottom, no solvent, 273 K). No signal observable.

## Supplementary References:

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