## **Supporting Information**

# Mono-mercapto-functionalized pillar[5]arene: a host-guest

## complexation accelerated reversible redox dimerization

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#### 1. General Materials and Methods

All reagents were purchased from commercial suppliers: Innochem, Aladdin, Adamas, Greagent and used without further purification. All solvents used in synthesis were purchased from commercial suppliers: KESHI. Anhydrous solvents were purchased from commercial suppliers: Energy Chemical and used without further purification from a drying agent under a nitrogen atmosphere.

<sup>1</sup>H NMR, <sup>13</sup>C NMR, <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>1</sup>H NOESY, <sup>1</sup>H-<sup>13</sup>C HSQC, <sup>1</sup>H-<sup>13</sup>C HMBC spectra were recorded on 400 MHz Agilent ProPulse (<sup>1</sup>H 400 MHz, <sup>13</sup>C 100 MHz) instruments. Chemical shifts ( $\delta$ ) are reported in parts per million (ppm). Coupling constants are reported in Hertz (Hz), and signal multiplicity is denoted as singlet (s), doublet (d), doublet of doublets (dd), triplet (t), dt (doublet of triplets), td (triplet of doublets), dtd (doublet of triplets of doublets), q (quartet), h (heptet), multiplet (m) and broad (br). All spectra were acquired at 20 °C (293 K) and were referenced to the residual solvent peaks. The common solvent impurities in <sup>1</sup>H and <sup>13</sup>C NMR in small amounts were water, dichloromethane.

The high-resolution mass spectra (HR-MS) data was acquired on a Bruker MaXis HD ESI-TOF mass spectrometer for high mass accuracy, coupled to a Thermo Scientific Dionex Ultra-High Performance Liquid Chromatography (UPLC) unit.

UPLC-MS data was collected by Ultimate 3000 Ultra-High

Performance Liquid Chromatography-ISQEM Mass Spectrometer. Acetonitrile (CH<sub>3</sub>CN) for UPLC-MS was purchased from Adamas. Mass spectra (positive mode) were acquired in ultra-scan mode. Analytical separations were achieved by injecting 5  $\mu$ L (for 1 mM DCL, scaled accordingly for dynamic combinatorial libraries (DCLs) at different concentrations) of DCL solution on to a Symmetry C18 reverse phase column (ThermoFisher Scientific, Accucore RP-MS 100 × 2.1 mm, 2.6  $\mu$ m particle size) at 35 °C and a flow rate of 0.30 mL/min.

2. Syntheses and Characterizations of Mono-SH-pillar[5]arene J1 and Di(mono-SH-pillar[5]arene)mer J2



## Synthetic route of J1

Scheme S1 The synthesis of Mono-SH-pillar[5]arene J1

## Synthetic route of J2



Scheme S2 The synthesis of dimer of Mono-SH-pillar[5]arene

#### **Synthetic Procedures**

Compound **S1** and compound **1** were synthesized according to literature procedures with necessary modification.<sup>1,2</sup>

**Compound S1: Dimethoxypillar**[5]arene.<sup>1</sup> To the mixture of 1,4dimethoxybenzene (5.80 g, 42.0 mmol), paraformaldehyde (3.60 g, 120.0 mmol, 2.86 eq) and anhydrous iron (III) chloride (1.00 g, 6.17 mmol, 0.15 eq) was added dichloromethane (A. R., analytical reagent) (CH<sub>2</sub>Cl<sub>2</sub>) (320 mL) in the water bath (30 °C) under nitrogen atmosphere. After stirring at 30 °C for 1 h, the reaction was quenched with distilled water (125 mL). Then, the mixture was filtrated under reduced pressure and the filtrate was extracted with  $CH_2Cl_2$  (150 mL  $\times$  2). The combined organic extracts were dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was further purified by column chromatography over silica gel (petroleum ether: ethyl acetate = 3:1) to afford S1 as whitish vellow solid powder in 58% yield (3.65 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) 6.79 (s, 10H, phenyl), 3.78 (s, 10H, -CH<sub>2</sub>-), 3.67 (s, 30H, -OCH<sub>3</sub>). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>): δ (ppm) 150.85 (phenyl), 128.32 (phenyl), 114.10 (phenyl), 55.87 (-OCH<sub>3</sub>), 29.71 (-CH<sub>2</sub>-). HR-MS (ESI-TOF): m/z Calcd for  $[M + NH_4]^+ C_{45}H_{54}NO_{10}$ : 768.3748; found: 768.3725, error: -3.0 ppm.

**Compound 1: Mono-OH-pillar[5]arene.** <sup>2</sup> To a solution of **S1** (2.00 g, 2.66 mmol) in chloroform (A. R., analytical reagent, 140 mL) was added boron tribromide (2.67 g, 10.7 mmol, 1.00 mL, 4.00 eq) in the ethanol bath (-10 °C). After stirring at -10 °C for 80 min, the reaction was quenched with

distilled water (100 mL). Then, the mixture was extracted with  $CH_2Cl_2$ (100 mL  $\times$  2). The combined organic extracts were dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was further purified by column chromatography over silica gel (eluent: first with (petroleum ether: ethyl acetate = 9:1); next with (petroleum ether: ethyl acetate = 4:1); last with (petroleum ether: ethyl acetate = 3:1)) to afford 1 as pink solid powder in 50% yield (0.98 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 6.89 (s, 1H, -OH), 6.76-6.58 (m, 10H); -CH<sub>2</sub>-: 3.84-3.76 (m, 10H); -OCH<sub>3</sub>: 3.75-3.50 (m, 27H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 151.95, 151.17, 151.07, 151.03, 151.01, 150.99, 150.90, 148.69, 147.65, 130.09, 129.41, 128.78, 128.47, 128.41, 128.36, 128.22, 127.80, 126.92, 125.13, 119.01, 114.65, 114.57, 114.37, 114.15, 113.95, 113.06, 113.00; -OCH<sub>3</sub>: 56.47, 56.25, 56.17, 56.09, 56.00, 55.97, 55.95, 55.94; -CH<sub>2</sub>-: 31.06, 30.17, 30.04, 29.71, 28.95. HR-MS (ESI-TOF): m/z Calcd for  $[M + NH_4]^+ C_{44}H_{52}NO_{10}$ : 754.3591; found: 754.3563, error: -3.7 ppm.

#### Compound 2: Mono-[(*N*,*N*'-dimethyl)thiocarbamoyl]-

pillar[5]arene.<sup>3,4</sup> To a solution of 1 (0.98 g, 1.33 mmol) and N,N'dimethylthiocarbamoyl chloride (DMTCC, 0.85 g, 6.65 mmol, 5.00 eq) in anhydrous THF (50 mL) was added sodium hydride (NaH, 60% wt suspension in mineral oil, 0.53 g, 13.3 mmol, 10.0 eq) at 0 °C. The mixture was stirred at 60 °C in the oil bath for 29 h. After cooling to room temperature, the reaction was quenched by adding 20 mL solution of 2.0 M HCl in the ice-water bath.  $CH_2Cl_2$  (150 mL) and distilled  $H_2O$  (100 mL) was added to the mixture. The mixture was extracted with  $CH_2Cl_2$  (100 mL  $\times$  2). The obtained yellow solution was dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was purified first by recrystallization with methanol and dichloromethane. Then, the obtained solid was further purified by column chromatography over silica gel (eluent: first with (petroleum ether: ethyl acetate = 4:1); then with (petroleum ether: ethyl acetate = 3:1); last with (petroleum ether: ethyl acetate = 2:1)) to afford **2** as white solid powder in 87% yield (0.96) g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 6.80-6.74 (m, 8H), 6.70 (s, 1H), 6.56 (s, 1H); -CH<sub>2</sub>-: 3.79-3.76 (m, 8H), 3.68 (s, 2H); -OCH<sub>3</sub>: 3.66-3.58 (m, 27H); -N(CH<sub>3</sub>)<sub>2</sub>: 3.48 (s, 3H), 3.29 (s, 3H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 188.43, 154.91, 151.12, 151.10, 151.07, 150.91, 150.88, 150.87, 150.85, 150.84, 144.92, 132.58, 128.83, 128.74, 128.67, 128.60, 128.51, 128.46, 128.08, 128.04, 127.74, 124.96, 114.67, 114.40, 114.28, 114.22, 114.03, 113.92, 112.61; -OCH<sub>3</sub>: 56.20, 56.06, 56.03, 56.00, 55.98, 55.94, 55.89, 55.24; -N(CH<sub>3</sub>)<sub>2</sub>: 43.46, 38.52; -CH<sub>2</sub>-: 30.20, 30.17, 29.95, 29.91, 29.80. HR-MS (ESI-TOF): *m*/*z* Calcd for  $[M + H]^+ C_{47}H_{54}NO_{10}S$ : 824.3468; found: 824.3455, error: -1.6 ppm.

Compound 3: Mono-[(N,N'-dimethylcarbamoyl)thio]pillar[5]arene.<sup>4,5</sup> The mixture of 2 (2.51 g, 3.05 mmol) and diphenyl ether (32.1 g, 188.6 mmol) was heated at 260-280 °C in the heatingmantle for 140 min under N<sub>2</sub> to undergo the Newman-Kwart rearrangement reaction. Then, the mixture was cooled to the room temperature and further purified by column chromatography over silica gel (eluent: first with petroleum ether; then with (petroleum ether: ethyl acetate =4:1); next with (petroleum ether: ethyl acetate = 2:1); last with (petroleum ether: ethyl acetate = 7:5)) to afford **3** as white solid powder in 48% yield (1.22 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 7.32 (s, 1H), 6.86 (s, 1H), 6.80-6.70 (m, 8H); -CH<sub>2</sub>-: 3.98 (s, 2H), 3.80-3.72 (m, 8H); -OCH<sub>3</sub>: 3.67-3.59 (m, 27H); -N(CH<sub>3</sub>)<sub>2</sub>: 3.20-2.92 (br, 6H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 167.90, 158.68, 151.26, 151.07, 151.02, 150.90, 150.88, 150.85, 150.74, 150.67, 145.94, 139.69, 128.59, 128.54, 128.44, 128.41, 128.38, 128.32, 128.17, 127.90, 117.08, 114.56, 114.40, 114.27, 114.22, 114.19, 114.11, 113.94, 112.97; -OCH<sub>3</sub>: 56.13, 56.00, 55.95, 55.92, 54.91; -N(CH<sub>3</sub>)<sub>2</sub>: 37.06; -CH<sub>2</sub>-: 33.60, 30.22, 30.07, 29.85, 29.74. HR-MS (ESI-TOF): m/z Calcd for  $[M + H]^+ C_{47}H_{54}NO_{10}S$ : 824.3468; found: 824.3468, error: 0.0 ppm.

**Compound J1: Mono-SH-pillar[5]arene.** <sup>5</sup> To the mixture of **3** (1.22 g, 1.48 mmol) and lithium aluminum hydride (LiAlH<sub>4</sub>) (0.84 g, 22.2 mmol,

15 eq) was added anhydrous THF (75 mL) in the ice-water bath under  $N_2$ atmosphere. After that, the mixture was stirred at 40 °C in the oil bath for 4.0 h under N<sub>2</sub> and quenched by slowly adding 50 mL of HCl (2.0 M, deoxygenated) solution at 0 °C under nitrogen protection. The obtained solution was extracted with  $CH_2Cl_2$  (100 mL  $\times$  2) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was further purified by column chromatography over silica gel (eluent: first with (petroleum ether: ethyl acetate = 6:1); then with (petroleum ether: ethyl acetate = 4:1)) to afford **J1** as white solid powder in 90% yield (1.00 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 7.20 (s, 1H), 6.84-6.76 (m, 7H), 6.75 (s, 1H), 6.73 (s, 1H); -CH<sub>2</sub>-: 3.95 (s, 2H), 3.80-3.77 (br, 6H), 3.72 (s, 2H); -OCH<sub>3</sub>: 3.67-3.62 (m, 27H); -SH: 3.18 (s, 1H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 156.36, 151.07, 151.03, 150.96, 150.90, 150.86, 150.85, 150.82, 150.67, 140.82, 135.31, 128.80, 128.71, 128.51, 128.49, 128.45, 128.35, 128.17, 127.86, 127.62, 117.97, 114.35, 114.24, 114.21, 114.19, 114.14, 113.90, 113.37; -OCH<sub>3</sub>: 56.04, 55.97, 55.96, 55.93, 55.80, 55.15; -CH<sub>2</sub>-:34.22, 29.95, 29.80, 29.75, 29.53. HR-MS (ESI-TOF): *m*/*z* Calcd for [M + NH<sub>4</sub>] <sup>+</sup> C<sub>44</sub>H<sub>52</sub>NO<sub>9</sub>S: 770.3363; found: 770.3359, error: -0.52 ppm.

Compound J2: Di(mono-SH-pillar[5]arene)mer. <sup>6</sup> To a solution of J1 (0.55 g, 0.73 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (A. R., analytical reagent) (130 mL)

containing triethylamine (1.48 g, 14.6 mmol, 2.00 mL, 20 eq), I<sub>2</sub> (0.56 g, 2.20 mmol, 3.00 eq) was added at room temperature (20 °C). After that, the solution was stirred at room temperature for 100 min and quenched by adding 30 mL of sodium thiosulfate solution (2.0 M) at room temperature. CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and distilled H<sub>2</sub>O (100 mL) was added to the mixture. The obtained brown solution was extracted with  $CH_2Cl_2$  (100 mL  $\times$  2) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was further purified by column chromatography over silica gel (eluent: first with (petroleum ether: ethyl acetate = 6:1); then with (petroleum ether: ethyl acetate = 4:1); last with (petroleum ether: ethyl acetate = 2:1)) to afford J2 as whitish yellow solid powder in 67% yield (0.37 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$ (ppm) phenyl: 7.40 (s, 2H), 6.85-6.72 (m, 18H); -CH<sub>2</sub>-: 3.89 (s, 4H), 3.77 (s, 12H), 3.72 (s, 4H); -OCH<sub>3</sub>: 3.68-3.63 (br, 48H), 3.57 (s, 6H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 158.11, 151.20, 151.03, 150.89, 150.82, 150.67, 150.51, 144.01, 137.76, 128.50, 128.38, 128.25, 128.19, 127.69, 126.11, 114.27, 114.12, 114.05, 113.80, 112.57; -OCH<sub>3</sub>: 55.99, 55.95, 55.92, 55.89, 55.86, 55.79, 54.98, -CH<sub>2</sub>-: 33.16, 29.89, 29.64. HR-MS (ESI-TOF): m/z Calcd for  $[M + NH_4]^+ C_{88}H_{98}NO_{18}S_2$ : 1520.6225; found: 1520.6176, error: -3.2 ppm.

Compound J2: Di(mono-SH-pillar[5]arene)mer. <sup>6</sup> To a solution of J1

(0.12 g, 0.16 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (A. R., analytical reagent) (30 mL) containing triethylamine (0.73 g, 7.20 mmol, 1.00 mL, 45 eq) and distilled H<sub>2</sub>O (15 mL), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) (30 wt%, 1.10 g, 9.70 mmol, 1.00 mL, 61.0 eq) was added at room temperature (20 °C). After that, the mixture was stirred at room temperature for 42.5 h and quenched by adding 10 mL of sodium thiosulfate solution (2.0 M) at room temperature.  $CH_2Cl_2$  (30 mL) and distilled  $H_2O$  (30 mL) was added to the mixture. The obtained solution was extracted with  $CH_2Cl_2$  (50 mL  $\times$  2) and dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of the solvents under reduced pressure, the residue was further purified by column chromatography over silica gel (eluent: first with (petroleum ether: ethyl acetate = 6:1); then with (petroleum ether: ethyl acetate = 4:1); last with (petroleum ether: ethyl acetate = 3:1)) to afford **J2** as white solid powder in 58% yield (0.070 g). <sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$  (ppm) phenyl: 7.40 (s, 2H), 6.85-6.72 (m, 18H); -CH<sub>2</sub>-: 3.89 (s, 4H), 3.77 (s, 12H), 3.72 (s, 4H); -OCH<sub>3</sub>: 3.68-3.63 (br, 48H), 3.57 (s, 6H). <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>):  $\delta$ (ppm) phenyl: 158.11, 151.20, 151.03, 150.89, 150.82, 150.67, 150.51, 144.01, 137.76, 128.50, 128.38, 128.25, 128.19, 127.69, 126.11, 114.27, 114.12, 114.05, 113.80, 112.57; -OCH<sub>3</sub>: 55.99, 55.95, 55.92, 55.89, 55.86, 55.79, 54.98, -CH<sub>2</sub>-: 33.16, 29.89, 29.64. HR-MS (ESI-TOF): *m/z* Calcd for  $[M + NH_4]^+ C_{88}H_{98}NO_{18}S_2$ : 1520.6225; found: 1520.6176, error: -3.2 ppm.

#### **1D NMR Spectra**



Fig. S2. <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of S1.



Fig. S3. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>, 293 K) of 1.



Fig. S4. <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of 1.



**Fig. S5.** <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>, 293 K) of **2**.



Fig. S6. <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of 2.



**Fig. S7.** DEPT ( $\theta$  = 135 °) spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of **2**.



**Fig. S8.** <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>, 293 K) of **3**.







Fig. S11. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>, 293 K) of J1.



Fig. S12. <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of J1.



**Fig. S13.** DEPT ( $\theta$  = 135 °) spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of **J1**.



**Fig. S14.** <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>, 293 K) of **J2**.



Fig. S15. <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of J2.



**Fig. S16.** DEPT ( $\theta$  = 135 °) spectrum (100 MHz, CDCl<sub>3</sub>, 293 K) of **J2**.

# 2D NMR Spectra





Fig. S17. Partial <sup>1</sup>H-<sup>1</sup>H COSY spectra of 2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S18. Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of 2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S19. Partial <sup>1</sup>H-<sup>13</sup>C HSQC spectra of 2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S20. Partial <sup>1</sup>H-<sup>13</sup>C HMBC spectra of 2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).





**Fig. S21.** Partial <sup>1</sup>H-<sup>1</sup>H COSY spectra of **3** (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S22. Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of 3 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S23. Partial <sup>1</sup>H-<sup>13</sup>C HSQC spectra of 3 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S24. Partial <sup>1</sup>H-<sup>13</sup>C HMBC spectra of 3 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).



J1



Fig. S25. Partial <sup>1</sup>H-<sup>1</sup>H COSY spectra of J1 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S26. Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of J1 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S27. Partial <sup>1</sup>H-<sup>13</sup>C HSQC spectra of J1 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S28. Partial <sup>1</sup>H-<sup>13</sup>C HMBC spectra of J1 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).








Fig. S31. Partial <sup>1</sup>H-<sup>13</sup>C HSQC spectra of J2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).



Fig. S32. Partial <sup>1</sup>H-<sup>13</sup>C HMBC spectra of J2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, 293 K, CDCl<sub>3</sub>)).

# HR-MS (ESI-TOF) Spectra





















#### 3. Reversible Redox Conversion between J1 and J2



**Fig. S45.** <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion of **J1** and **J2** using I<sub>2</sub> as oxidant and DTT as reductant. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) **J1**, (b) a reaction mixture of **J1** and I<sub>2</sub> after reacting for 100 min, (b) a reaction mixture from (c) adding DTT (dithiothreitol) for 5 h 40 min, (d) **J2**. The chemical shifts assigned to phenyl of **J1** (**Fig. S45-c**) are not completely consistent with pure **J1** (**Fig. S45-a**) probably due to the existence of host-guest

complexation among DTT, the oxidized product of DTT, **J1**. Peaks are highlighted in red and blue for **J1** ( $H_m$ ) and **J2** ( $H_d$ ), respectively.



Fig. S46. The reversible redox conversion in the presence of oxidant  $O_2$  and reductant DTT (dithiothreitol) between J1 and J2.

## The oxidative self-dimerization of J1 in the presence of O2 and K2CO3





**Fig. S47.** The partial <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 293 K) spectra changes of **J1** in the presence of  $O_2$  and K<sub>2</sub>CO<sub>3</sub>(1: **J1**; 2: 0 d; 3: 1 d; 4: 2 d; 5: 5 d; 6: 16 d; 7: 30 d; 8: 47 d; 9: **J2**). Asterisk represents dichloromethane.

# The reduction of J2 in the presence of DTT and K<sub>2</sub>CO<sub>3</sub>





**Fig. S48.** The full <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) changes of **J2** in the presence of **DTT** and  $K_2CO_3(1: J2; 2: 10 min; 3: 40 min; 4: 163 min; 5: 590 min; 6: 23 h 43 min; 7: 34 h 10 min; 8: 47 h; 9:$ **J1**). Asterisk represents dichloromethane.



**Fig. S49.** The partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) changes of **J2** in the presence of **DTT** and  $K_2CO_3(1: J2; 2: 10 min; 3: 40 min; 4: 163 min; 5: 590 min; 6: 23 h 43min; 7: 34 h 10 min; 8: 47 h; 9:$ **J1**). Asterisk represents dichloromethane.



Fig. S50. <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) DTT, (b) J1 in the presence of DTT, (c) J1.



Fig. S51. The UPLC chromatogram changes of J2 in the presence of DTT and  $K_2CO_3$  (blue: J2; red: J1) (a: J2; b: 76 min; c: 5 h; d: 24 h).



**Fig. S52.** The UPLC chromatogram (left) and the corresponding mass spectra of **J2** (right). The split peak is ascribed to the nonoptimal separation conditions including stationary phase, mobile phase, separation column temperature, solvent effect, etc.

#### Determination of oxidative self-dimerization rate constant of J1 by linear fitting



Fig. S53. The oxidative self-dimerization of J1.

The oxidative self-dimerization rate constant of **J1** was determined by linear fitting. A linear fitting method was applied to calculate the oxidative self-dimerization rate constant for **J1**, which was based on the second order reaction equation as follows:

$$\frac{1}{c} - \frac{1}{c_0} = k_2 t \qquad (\text{eq-1})$$

Where  $c_0$  is the fixed initial concentration of J1 ( $c_0 = 42 \text{ mM}$ ), c obtained by calculation from relative proportion of H<sub>SH</sub> and H<sub>d</sub> in <sup>1</sup>H NMR spectra is the concentration of J1 when reaction time is t, t is reaction time of the oxidative self-dimerization of J1.



Fig. S54. The linear fitting plot of oxidative self-dimerization rate constant of J1.

#### The changes of oxidative self-dimerization conversion ratio of J1 by nonlinear curve-fitting

The oxidative self-dimerization is a kind of second order reaction according to the result of Fig. S63. For oxidative self-dimerization conversion ratio of J1  $\alpha$ , its expression is obtained as follows:

$$\alpha = \frac{n_0 - n}{n_0}$$

$$n = cV_0, n_0 = c_0V_0$$

$$\frac{1}{c} - \frac{1}{c_0} = k_2t$$

$$c = \frac{c_0}{c_0k_2t + 1}$$

$$\alpha = \frac{c_0k_2t}{c_0k_2t + 1} = \frac{At}{At + 1} = \frac{1}{1 + \frac{1}{At}}$$
(eq-2)

Where  $c_0$  is the fixed initial concentration of J1 ( $c_0 = 42$  mM);

*c*: the concentration of J1 obtained by calculation from relative proportion of  $H_{SH}$  and  $H_d$  in <sup>1</sup>H NMR spectra when reaction time is *t*;

*t*: reaction time of the oxidative self-dimerization of **J1**;

 $\alpha$ : oxidative self-dimerization conversion ratio of J1;

*n*: the amount of substance of J1 obtained by calculation from relative proportion of  $H_{SH}$  and  $H_d$  in <sup>1</sup>H NMR spectra when reaction time is *t*;

- $n_0$ : the fixed initial amount of substance of J1 ( $n_0 = 0.021 \text{ mmol}$ );
- $k_2$ : the oxidative self-dimerization rate constant of J1 ( $k_2 = (0.902 \pm 0.016)$  L• mol<sup>-1</sup>•d<sup>-1</sup>);

$$A = c_0 k_2.$$



Fig. S55. The changes of oxidative self-dimerization conversion ratio of J1  $\alpha$  by nonlinear curve-fitting.

# 4. Host-guest Complexation

NMR spectra of HDPB (1-hexadecylpyridinium bromide) in the absence and presence of J1



**Fig. S56.** <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) **J1**, (b) **HDPB** in the presence of **J1**, (c) **HDPB**. **[J1]** = **[HDPB]** = 44.8 mM. Asterisk represents dichloromethane.



**Fig. S57.** Full <sup>1</sup>H-<sup>1</sup>H NOESY spectra of **HDPB** in the presence of **J1** (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). [**J1**] = [**HDPB**] = 44.8 mM. Asterisk represents dichloromethane.



**Fig. S58.** Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of **HDPB** in the presence of **J1** (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). [**J1**] = [**HDPB**] = 44.8 mM.



**Fig. S59.** Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of **HDPB** in the presence of **J1** (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). [**J1**] = [**HDPB**] = 44.8 mM.

NMR spectra of HDPB (1-hexadecylpyridinium bromide) in the absence and presence of J2







**Fig. S61.** Full <sup>1</sup>H-<sup>1</sup>H NOESY spectra of **HDPB** in the presence of **J2** (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). [**HDPB**] = 2[J2] = 40.0 mM. Asterisk represents dichloromethane.



Fig. S62. Partial <sup>1</sup>H-<sup>1</sup>H NOESY spectra of HDPB in the presence of J2 (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). [HDPB] = 2[J2] = 40.0 mM.

# NMR spectra of titration





**Fig. S63.** The <sup>1</sup>H NMR spectra of titration of **G0 (HDPB)** and **H0 (J1)**, **G0** in the presence of **H0** (0.1-3.0 eq) (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). Asterisk represents dichloromethane.





**Fig. S64.** The <sup>1</sup>H NMR spectra of titration of **G0** (**HDPB**) and **H1** (**J2**), **G0** in the presence of **H1** (0.1-2.6 eq) (<sup>1</sup>H NMR (400 MHz, 293 K, CDCl<sub>3</sub>)). Asterisk represents dichloromethane.

## Molar ratio plot for the complexation by <sup>1</sup>H NMR titration



Fig. S65. Molar ratio plot for the complexation between G0 (HDPB) and H0 (J1), indicating a 1:1 stoichiometry.



Fig. S66. Molar ratio plot for the complexation between G0 (HDPB) and H1 (J2), indicating a 2:1 stoichiometry.

## Determination of association constants by <sup>1</sup>H NMR titration

The association constant between the host H0 (J1) and the guest G0 (HDPB) was determined by <sup>1</sup>H NMR titration. A nonlinear curve-fitting method (Fitter: NMR 1:1; Method: Nelder-Mead; Flavour: None (Full))<sup>7</sup> was applied to calculate the association constant for complexation between H0 and G0, which was based on the online tools for supramolecular chemistry research and analysis (<u>http://supramolecular.org</u>).



Fig. S67. The chemical shift changes and the nonlinear-fitting curves by BindFit v0.5 of  $H_1$  (Proton 1),  $H_4$  (Proton 2) and  $H_5$  (Proton 3) of G0 (42 mM) upon the addition of H0 (0–3.00 eq).

The association constant between the host H1 (J2) and the guest G0 (HDPB) was determined by <sup>1</sup>H NMR titration. A nonlinear curve-fitting method (Fitter: NMR 2:1; Method: Nelder-Mead; Flavour: None (Full))<sup>8</sup> was applied to calculate the association constant for complexation between H1 and G0, which was based on the online tools for supramolecular chemistry research and analysis (<u>http://supramolecular.org</u>).


Fig. S68. The chemical shift changes and the nonlinear-fitting curves by BindFit v0.5 of  $H_1$  (Proton 1),  $H_5$  (Proton 2) and  $H_6$  (Proton 3) of G0 (42 mM) upon the addition of H1 (0–2.60 eq).

5. The Reversible Redox Conversion of the Host-guest Complexes between  $G0 \subset J1$  and  $(G0)_2 \subset J2$ .





Fig. S69. <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion of the host-guest complexes between  $G0 \subset J1$  and  $(G0)_2 \subset J2$ . Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) the mono-SH-pillar[5]arene J1, (b) the host-guest complex  $G0 \subset J1$ , (c) a reaction mixture containing  $(G0)_2 \subset J2$  after performing oxidation by O<sub>2</sub> in the air for 4 h, (d) a reaction mixture containing  $G0 \subset J1$  after performing reduction by adding DTT (dithiothreitol) for 68 min, and (e) the di(mono-SH-pillar[5]arene)mer J2. Peaks are highlighted in red and blue for J1 (H<sub>m</sub>) and J2 (H<sub>d</sub>), respectively; peaks which chemical shifts changed are highlighted in red, blue, for H<sub>1</sub>, H<sub>2</sub>, H<sub>3</sub> and H<sub>4</sub>, H<sub>5</sub>, H<sub>6</sub> of G0, respectively.



**Fig. S70.** <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion between J1 and J2 in the presence of catalytic amount of G0 (9 mol%). Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) **mono-SH-pillar[5]arene J1**, (b) a reaction mixture containing J2 and catalytic amount of G0 after performing oxidation by  $O_2$  in the air for 5 d, (c) a reaction mixture containing J1 and catalytic amount of G0 after performing reduction by adding DTT (dithiothreitol) for 130 min, and (d) di(mono-SH-pillar[5]arene)mer J2. Peaks are highlighted in red and blue for J1 (H<sub>m</sub>) and J2 (H<sub>d</sub>), respectively.

Host	$K_a$ / $M^{-1}$	Reaction	$t_c^{[a]}$	tc' <sup>[b]</sup>
J1	270	Dimerization	53 d <sup>[c]</sup>	4 h
J2	1200 110	Reduction	56.5 h <sup>[d]</sup>	68 min

Table S1 Association constants of host (J1, J2) with guest (G0), the time of redox conversion between J1 and J2 in the absence or presence of G0.

<sup>[a]</sup>The time of redox conversion between J1 and J2 in the absence of G0

<sup>[b]</sup>The time of redox conversion between **J1** and **J2** in the presence of **G0** 

<sup>[c]</sup>Conversion ratio of **J1** (71%) by <sup>1</sup>H NMR

<sup>[d]</sup>Conversion ratio of **J2** (76%) by <sup>1</sup>H NMR

## 6. Control Experiments for Further Elucidating the Mechanism of Accelerated Redox Conversion Process.

In order to elucidate the mechanism of accelerated redox conversion process between J1 and J2 in the presence of G0 (pyridinium salt), several control experiments have been performed. Since we realized the redox reactions are heterogeneous systems owing to the low solubility of base ( $K_2CO_3$ ) in CHCl<sub>3</sub>, we first replaced G0 with TBAB (tetrabutylammonium bromide) and found out that TBAB can accelerate the reaction (Fig S75, S76, Table S2). The result of TBAB confirms that phase transfer catalyst can accelerate the reaction, but not as good as G0 because TBAB is too bulky to get into the cavity of J1 and J2. So, we think the host-guest interaction is also important. To further confirm this hypothesis, we mixed G0 with 4-methoxybenzenethiol (MBT) and found that the dimerization of MBT was accelerated (Fig S72, S73, Table S2), which confirms that G0 also functions as a phase transfer catalyst.<sup>9</sup> The acceleration effect is not as good as J1 again because there is no host-guest interaction between G0 and MBT.



Fig. S71. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) J1, (b) J1 in the presence of G0 (HDPB). The  $H_{SH}$  assigned to G0 $\subset$ J1 experienced significant downfield shift compared with J1 owing to deshielding effect derived from complexation of electron-deficient G0 and J1.



Fig. S72. <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion between 4-methoxybenzenethiol and 1,2-bis(4-methoxybenyl)disulfane in the absence of G0. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) 4-methoxybenzenethiol, (b) a reaction mixture containing 4-methoxybenzenethiol and K<sub>2</sub>CO<sub>3</sub> after performing oxidation by O<sub>2</sub> in the air for 72 h (conversion ratio: 7%), (c) a reaction mixture containing 4-methoxybenzenethiol (conversion ratio: 50%) and K<sub>2</sub>CO<sub>3</sub> after performing reduction by adding DTT (dithiothreitol) into 1,2-bis(4-methoxybenyl)disulfane for 59 h 52 min, and (d) 1,2-bis(4-methoxybenyl)disulfane. Peaks are highlighted in red for 4-methoxybenzenethiol.



Fig. S73. <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion between 4-methoxybenzenethiol and 1,2-bis(4-methoxybenyl)disulfane in the presence of G0. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) 4-methoxybenzenethiol, (b) a reaction mixture containing 4-methoxybenzenethiol, G0 and K<sub>2</sub>CO<sub>3</sub> after performing oxidation by O<sub>2</sub> in the air for 16 h 10 min, (c) a reaction mixture containing 1,2-bis(4-methoxybenyl)disulfane, G0 and K<sub>2</sub>CO<sub>3</sub> after performing reduction by adding DTT (dithiothreitol) for 165 min, and (d) 1,2-bis(4-methoxybenyl)disulfane. Peaks are highlighted in red for 4-methoxybenzenethiol.



**Fig. S74.** <sup>1</sup>H NMR spectroscopic analysis of host-guest complexation between **J1** and **octylbenzene**. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) **mono-SH-pillar[5]arene J1**, (b) a reaction mixture containing **J1** and **octylbenzene**, and (c) **octylbenzene**. No host-guest complexation between **J1** and **octylbenzene**.



**Fig. S75.** <sup>1</sup>H NMR spectroscopic analysis of host-guest complexation between J1 and TBAB (tetrabutylammonium bromide). Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) mono-SH-pillar[5]arene J1, (b) a reaction mixture containing J1 and TBAB, and (c) TBAB. No host-guest complexation between J1 and TBAB.



**Fig. S76.** <sup>1</sup>H NMR spectroscopic analysis of reversible redox conversion between **J1** and **J2** in the presence of **TBAB**. Partial <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 293 K) of (a) a reaction mixture containing **J1** and **TBAB**, (b) a reaction mixture containing **J2** (conversion ratio: 50%) and K<sub>2</sub>CO<sub>3</sub> after performing oxidation by O<sub>2</sub> in the air for 25 d, (c) a reaction mixture containing **J2** (conversion ratio: 59%), **TBAB** and K<sub>2</sub>CO<sub>3</sub> after performing oxidation by O<sub>2</sub> in the air for 49 h 50 min, (d) a reaction mixture containing **J1**, **TBAB** and K<sub>2</sub>CO<sub>3</sub> after performing reduction by adding DTT (dithiothreitol) for 5 h 49 min, and (e) **di(mono-SH-pillar[5]arene)mer J2**. Peaks are highlighted in red and blue for **J1** (H<sub>m</sub>) and **J2** (H<sub>d</sub>), respectively.

System	Reaction	t <sub>ox</sub>	t <sub>red</sub>
J1	Dimerization	25 d <sup>[a]</sup> , 53 d <sup>[b]</sup>	—
J2	Reduction	_	56.5 h <sup>[c]</sup>
J1+G0	Dimerization	4 h	—
J2+G0+DTT	Reduction	—	68 min
J1+TBAB	Dimerization	49 h 50 min <sup>[d]</sup>	—
J2+TBAB+DTT	Reduction	—	5 h 49 min
MBT	Dimerization	72 h <sup>[e]</sup>	—
BMPD	Reduction	—	59 h 52 min <sup>[f]</sup>
MBT+G0	Dimerization	16 h 10 min	—
BMPD +G0+DTT	Reduction	—	165 min

Table S2 The time of redox conversions between monomers and dimers in the absence or presence of G0, TBAB.

<sup>[a]</sup>Conversion ratio of J1 (50%) by <sup>1</sup>H NMR
<sup>[b]</sup>Conversion ratio of J1 (71%) by <sup>1</sup>H NMR
<sup>[c]</sup>Conversion ratio of J2 (76%) by <sup>1</sup>H NMR
<sup>[d]</sup>Conversion ratio of J1 (59%) by <sup>1</sup>H NMR
<sup>[e]</sup>Conversion ratio of MBT (7%) by <sup>1</sup>H NMR, MBT: 4-methoxybenzenethiol
<sup>[f]</sup>Conversion ratio of BMPD (50%) by <sup>1</sup>H NMR, BMPD: 1,2-bis(4-methoxyphenyl)disulfane (dimer of 4-methoxybenzenethiol)

The Proposed Mechanism of a Faster Reversible Redox Conversion in the Presence of Guest



G0 (quaternary ammonium salt, phase transfer catalyst (PTC) & Guest)

Fig. S77. The proposed mechanism of the faster oxidative dimerization of J1.<sup>10</sup>



Fig. S78. The proposed mechanism of the faster reduction of J2.<sup>11</sup>

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