Supplementary Information

for

Deoxygenative hydroboration of primary and secondary amides: A catalyst-free and solvent-free approach

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General experimental information

All reactions were performed under argon atmosphere using Schlenk techniques or inside a MBraun glove box. Pinacolborane (HBpin), amides were purchased from Sigma–Aldrich, TCI and used without further purification. C_6D_6 , DMSO-d₆, CDCl₃ were purchased from Sigma-Aldrich, were degassed by three freeze-pump-thaw cycles and stored over molecular sieves. ¹H and ¹³C{¹H} spectra were recorded on Bruker AV-200 MHz, AV-400 MHz and AV-500 MHz and referenced to the resonances of the solvent used.

		-	-	-	-	
Entry	Catalyst	Equiv. of	Solvent	Temperature	Time	NMR Yield
		HBpin		(°C)	(h)	(%)
1.	-	4.5	Neat	60	20	55
2.	-	4.5	Neat	70	20	60
3.	-	5.0	Neat	80	12	56
4.	-	5.0	Neat	80	16	73
5.	-	5.0	Neat	90	12	72
6.	-	5.0	Neat	90	16	85
7.	-	5.0	Neat	95	12	91
8.	-	5.0	Neat	100	13	98

Table S1. Optimization table of catalyst and solvent free for the hydroboration of benzamide

🚔 General procedure for the hydroboration of primary amides



Scheme S1. General procedure for the hydroboration of primary amides.

Primary amide (0.127 mmol), pinacolborane (5.0 equiv, 0.635 mmol) were mixed together in a Schlenk tube or in a screw cap NMR tube inside the glove box. The reaction mixture was allowed

to heat at 100 °C for 13 h without adding any solvent. Upon completion of the reaction, mesitylene (0.127 mmol) as an internal standard, was added while making the NMR in an appropriate deuterated solvent. The progress of the reaction was monitored by ¹H NMR, which indicates the completion of the reaction by the appearance of a new $R-CH_2-N(Bpin)_2$ resonance (Scheme S1). Upon completion, the reaction mixture was stirred with 1.0 mL 2.0 (M) NaOH solutions and 1.0 mL Et_2O for 1 h to complete the hydrolysis. Next, the reaction mixture was worked up with Et_2O : H_2O mixture (1:1) and the Et_2O part were concentrated in vacuum to get corresponding reduced amines. Consequently, 1.0 mL 1.0 (M) methanolic HCl solution was added to the concentrated amines followed by addition of 1.0 mL Et_2O and the corresponding amine hydrochloride salt was purified by washing with Et_2O . Isolated amine hydrochlorides were characterized through NMR spectroscopy in DMSO-d₆.

Hydroboration product of primary amides, **1a**, **1e** were isolated and in both the cases little discrepancy was observed with the NMR yield.

Entry	Amide Precursor	Temperature	Time	NMR	Product
		(°C)	(h)	yield (%)	
1.	NH ₂	100	13	97	1a
2.	MeO NH ₂	100	13	98	1b
3.	Me NH ₂	100	13	98	1c

Table S2. Substrates scope for the primary amide hydroboration.

4.	Ph NH ₂	100	13	95	1d
5.	MeO OMe OMe	100	13	95	1e
6.	F NH2	100	13	96	1f
7.	F O NH ₂	100	13	22	1g
8.	F F F F	100	13	28	1h
9.	O NH2	100	13	>99	1i
10.	NH ₂	100	13	70	1j
11.	NH ₂	100	13	90	1k

12.	NH ₂	100	13	66	11
13.		100	13	99	1m
14.	Et NH ₂	100	13	87	1n
15.	NH ₂	100	13	85(reduction of both C=C and C=O bond)	10
16.		100	13	69	1p
17.	NH ₂	100	13	94	1q
18.	N NH2	100	13	76	1r
19.		100	13	70	1s
20.	NH ₂	100	13	n. r.	1t
21.	Br NH ₂	100	13	n. r.	1u

Analytical data (¹H and ¹³C NMR) of the borylamines of corresponding primary amides: For every reaction, along with the *N*-borylated amine Bpin-O-Bpin formed as second product which show a singlet at δ 1.17-1.18 ppm in ¹H NMR and peak at δ 24.1-24.4 and 82.4-82.9 ppm in ¹³C NMR.

1. 1a: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.36-7.32 (m, 2H, *Ph*), 7.22-7.20 (t, 2H, *Ph*), 7.17 N(Bpin)₂
 7.14 (t, 1H, *Ph*), 4.26(s, 2H, PhCH₂N(Bpin)₂), 1.29 (s, 24H,N(Bpin)₂) ppm;
 ¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 142.9 (*Ph*), 137.5(*Ph*), 127.6
 (*Ph*), 127.4 (*Ph*), 126.8 (*Ph*), 125.9(*Ph*),82.95(PhCH₂N(Bpin)₂), 82.19(PhCH₂N(Bpin)₂),
 81.78(PhCH₂N(Bpin)₂),47.1(PhCH₂N(Bpin)₂), 24.4 (PhCH₂N(Bpin)₂) ppm.

The spectroscopic data is consistent with literature data.¹

Phenylmethanamine hydrochloride (1a').

The general procedure was followed to isolate phenylmethanamine hydrochloride salt, **1a'** (15.15 mg, 85.2% yield) as a colorless solid.



¹H NMR (200 MHz, DMSO-d₆): δ 8.47(br, 3H, NH₂·HCl), 7.52-7.49 (d, ³J_{H-H} = 6.50 Hz, 2H, *Ph*), 7.39-7.41 (m, 3H, *Ph*), 4.00 (br,2H, *CH*₂) ppm. ¹³C{¹H} (100.6 MHz, DMSO-d₆): δ 134.18 (*Ph*),129.0 (*Ph*), 128.6 (*Ph*), 128.6 (*Ph*), 42.2 (*C*H₂) ppm.

The spectroscopic data is consistent with literature data.²



2. 1b: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.27-7.23 (m, 2H, *Ph*),
6.77 (m, 2H, Ph),4.17(s, 2H, PhC*H*₂N(Bpin)₂), 3.78 (s, 3H, OC*H*₃),
1.21 (s, 24H, N(B*pin*)₂) ppm; ¹³C{¹H} NMR (CDCl₃, 100.6 MHz,
298 K): δ 157.9 (*ArC*-OMe), 137.4 (*Ph*), 135.3 (*Ph*), 133.8 (*Ph*),

128.6 (*Ph*),127.7 (*Ph*), 126.7 (*Ph*),82.8 (PhCH₂N(B*pin*)₂), 54.9 (OM*e*), 46.4 (PhCH₂N(B*pin*)₂), 24.39(PhCH₂N(B*pin*)₂) ppm.

The spectroscopic data is consistent with literature data.¹

 $\begin{array}{c} \text{Me} \\ & \text{N(Bpin)}_2 \\ \text{Me} \\ & \text{N(Bpin)}_2 \end{array} \begin{array}{c} \text{3. 1c: } ^1\text{H NMR (CDCl_3, 200 MHz, 298 K): } \delta \ 7.16\text{-}7.00 \ (\text{m}, 5\text{H}, Ph), \\ & 4.23(\text{s}, 2\text{H}, \text{Ph}CH_2\text{N(Bpin)}_2), \ 2.32 \ (\text{s}, 3\text{H}, CH_3), \ 1.22 \ (\text{s}, 24\text{H}, \\ & \text{N(Bpin)}_2) \ \text{ppm;} \ ^{13}\text{C} \{^1\text{H}\} \ \text{NMR (CDCl_3, 100.6 MHz, 298 K): } \delta \ 142.8 \\ & (ArC\text{-Me}), \ 137.5 \ (Ph), \ 128.1 \ (Ph), \ 127.3 \ (Ph), \ 126.7 \ (Ph), \ 124.3 \ (Ph), \ 123.6 \ (Ph), \ 82.9 \\ & (\text{PhCH}_2\text{N(Bpin)}_2), \ 82.1 \ (\text{PhCH}_2\text{N(Bpin)}_2), 46.9 \ (\text{PhCH}_2\text{N(Bpin)}_2), \ 24.4(\text{PhCH}_2\text{N(Bpin)}_2), \ 21.2 \\ & (\text{Ar-CH}_3)\text{ppm.} \end{array}$

The spectroscopic data is consistent with literature data.³

4. **1d:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.63-7.61 (m, 2H, *Ph*), N(Bpin)₂ 7.54-7.52 (d, 2H, *Ph*), 7.46-7.41 (m, 4H, *Ph*), 7.35-7.33 (m, 1H, *Ph*), 4.32(s, 2H, PhCH₂N(Bpin)₂), 1.24 (s, 24H, N(B*pin*)₂) ppm; ¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 142.1 (CH₂Ar*C*-Ph), 141.1 (CH₂Ar-CAr), 138.8(*Ph*), 137.5 (*Ph*), 128.5 (*Ph*), 127.8 (*Ph*),126.7 (*Ph*), 126.4 (*Ph*), 82.2 (PhPhCH₂N(B*pin*)₂), 46.8 (PhPhCH₂N(B*pin*)₂), 24.4(PhCH₂N(B*pin*)₂) ppm.

5. **1e:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 6.52-6.51 (m, 2H, *Ph*), 6.29 (s, 1H, *Ph*), 4.18 (s, 2H, MeO $N(Bpin)_2$ $PhCH_2N(Bpin)_2$), 3.76 (s, 6H, OCH₃), 1.22 (s, 24H, N(Bpin)_2) pm;¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 160.6(ArC-OMe), 160.2(ArC-OMe), 145.3(*Ph*), 137.4 (*Ph*), 126.7 (*Ph*), 104.3(*Ph*), 98.4 (*Ph*), 82.8 (PhCH₂N(Bpin)₂), 55.0 (-OMe), 45.1

(PhCH₂N(Bpin)₂), 24.3 (PhCH₂N(Bpin)₂) ppm.

(3,5-dimethoxyphenyl)methanamine hydrochloride (1e').

The general procedure was followed for the isolation of (3,5-dimethoxyphenyl)methanamine hydrochloride salt, **1e'** (22.58 mg, 87.6% yield) as a colorless solid.



¹H NMR (200MHz, DMSO-d₆): δ 8.59 (br, 3H, NH₂·HCl), 6.74-6.73 (d, 2H, Ph), 6.46 (t, 1H, Ph),
3.90 (q, 2H, CH₂), 3.73 (s, 6H, OMe) ppm.
¹³C{¹H} (100.6 MHz, DMSO-d₆): δ 160.4 (Ph), 136.2 (Ph), 106.8 (Ph), 99.9 (Ph), 55.3 (OMe),42.1 (CH₂) ppm.

The spectroscopic data is consistent with literature data.²

 $6. 1f: {}^{1}H NMR (CDCl_{3}, 200 MHz, 298 K): \delta 7.33-7.30 (m, 2H, Ph), 6.98-6.89 (t, 2H, Ph), 4.20(s, 2H, PhCH_2N(Bpin)_2), 1.21 (s, 24H, N(Bpin)_2) ppm; {}^{13}C {}^{1}H NMR (CDCl_{3}, 100.5 MHz, 298 K): \delta 162.5-160.2 (ArC-F), 138.6-137.4 (Ph), 129.8 (Ph), 128.7 (Ph), 114.7-114.1 (Ph), 82.8 (PhCH_2N(Bpin)_2), 46.3 (PhCH_2N(Bpin)_2), 24.3 (PhCH_2N(Bpin)_2) ppm; {}^{19}F {}^{1}H NMR (CDCl_{3}, 128 MHz, 298 K): -117.45 ppm.$

The spectroscopic data is consistent with literature data.¹

7. **1g:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.05-7.00 (m, 1H, *Ph*), 6.75-6.71 (t, 2H, *Ph*), 4.33(s, 2H, PhCH₂N(Bpin)₂), 1.27 (s, 24H, N(B*pin*)₂) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 163.0 (Ar*C*-F), 160.6-160.5(Ar*C*-F), 128.0-127.9 (*Ph*), 112.0-111.7 (*Ph*), 110.5-110.2 (*Ph*), 81.5 (PhCH₂N(B*pin*)₂),35.7 (PhCH₂N(B*pin*)₂), 24.1 (PhCH₂N(B*pin*)₂) ppm; ¹⁹F{¹H} NMR (CDCl₃, 128 MHz, 298 K): -103.58 (*product*), -113.38 (*starting material*) ppm.



 O_2N

8. 1h: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 4.35(s, 2H, PhCH₂N(Bpin)₂), 1.28 (s, 24H, N(Bpin)₂) ppm; ¹³C {¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 137.6-126.6 (Ph), 83.0 (PhCH₂N(Bpin)₂), 36.0 (PhCH₂N(Bpin)₂), 24.4 (PhCH₂N(Bpin)₂) ppm; ¹⁹F {¹H} NMR (CDCl₃, 128 MHz, 298 K): δ - 131.34 (*product*), -140.57 (product), -141.87

(starting material), -152.89(product), -157.90 (starting material), -162.37 (starting material), -164.33 (starting material) ppm.

9. 1i: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 8.15-8.11 (m, 2H, Ph), 7.47-7.43 (m, 2H, Ph), 4.31

 $(s, 2H, PhCH₂N(Bpin)₂), 1.20 (s, 24H, N(Bpin)₂) ppm; {}^{13}C{}^{1}H} NMR$ $(CDCl₃, 100.5 MHz, 298 K): <math>\delta$ 150.6 (*ArC*-NO₂), 146.4 (*Ph*), 137.5 (*Ph*), 127.9 (*Ph*), 126.7 (*Ph*), 123.4 (*Ph*), 82.9 (NO₂PhCH₂N(B*pin*)₂), 46.8 (NO₂PhCH₂N(B*pin*)₂), 24.4 (NO₂PhCH₂N(B*pin*)₂) ppm.

The spectroscopic data is consistent with literature data.⁴

10. **1j**: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 8.42-8.40 (d,1H, *Ar*), 7.86-N(Bpin)₂ 7.84 (d,1H, *Ar*), 7.73-7.71 (d,1H, *Ar*), 7.53-7.48 (m, 2H, *Ar*), 7.40-7.36 (m, 2H, *Ar*), 3.46-3.42 (t, 2H, ArCH₂CH₂N(Bpin)₂), 3.24-3.20 (t, 2H,ArCH₂CH₂N(Bpin)₂), 1.25 (s, 24H, N(B*pin*)₂) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 137.5 (*Ar*), 136.4(*Ar*), 133.7(*Ar*), 132.4(*Ar*), 128.3 (*Ar*), 126.8 (*Ar*), 126.4 (*Ar*), 125.3 (*Ar*), 125.1 (*Ar*), 124.6 (*Ar*),82.9 (ArCH₂CH₂N(B*pin*)₂), 44.8 (ArCH₂CH₂N(B*pin*)₂), 37.1 (ArCH₂CH₂N(B*pin*)₂), 24.4 (ArCH₂CH₂N(B*pin*)₂) ppm.

11. **1k:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.29-7.24 (m, 5H, *Ph*), **N(Bpin)**₂ 3.36-3.33 (t, 2H, PhCH₂CH₂N(Bpin)₂), 2.77-2.74 (t, 2H, PhCH₂CH₂N(Bpin)₂), 1.22 (s, 24H, N(B*pin*)₂) ppm;¹³C{¹H} NMR (CDCl₃, 100.53 MHz, 298 K): δ 139.7(ArC-R), 137.5 (*Ph*), 128.8 (*Ph*), 126.7(*Ph*), 82.9 (PhCH₂CH₂N(B*pin*)₂), 42.2 (PhCH₂CH₂N(B*pin*)₂), 39.7 (PhCH₂CH₂N(B*pin*)₂), 24.4 (PhCH₂CH₂N(B*pin*)₂) ppm.

The spectroscopic data is consistent with literature data.³

12. **1I:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 2.90-2.87 (d, 2H, Cy*CH*₂N(Bpin)₂), 1.71-1.66 (m, N(Bpin)₂ 4H, *Cy*), 1.59 (br, 1H, Cy), 1.22 (s, 24H, N(B*pin*)₂), 0.88-0.83 (m, 2H, *Cy*) pm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.8 (CyCH₂N(B*pin*)₂), 49.5 (Cy*C*H₂N(B*pin*)₂), 40.4 (*Cy*CH₂N(B*pin*)₂), 30.4 (*Cy*CH₂N(B*pin*)₂), 26.6 (*Cy*CH₂N(B*pin*)₂), 26.0 (*Cy*CH₂N(B*pin*)₂), 24.77 (*Cy*CH₂N(B*pin*)₂), 24.4 (CyCH₂N(B*pin*)₂) ppm.

The spectroscopic data is consistent with literature data.³

13. **1m:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 3.08-3.03 (q, 2H, CH₂), 1.22 (s, H₃C N(Bpin)₂ 24H, N(Bpin)₂),1.04-1.00 (t, 3H, CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.8 (CH₃CH₂N(Bpin)₂), 38.4 (CH₃CH₂N(Bpin)₂), 24.4 (CH₃CH₂N(Bpin)₂), 18.5 (CH₃CH₂N(Bpin)₂) ppm. The spectroscopic data is consistent with literature data.³

14. **1n**: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 3.03-2.96 (t, 2H, CH₂), 1.47-1.39 Et N(Bpin)₂ (m, 2H, CH₂), 1.22 (s, 24H, N(B*pin*)₂), 0.87-0.80 (t, 3H, CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.8 (CH₃CH₂CH₂N(B*pin*)₂), 45.2 (CH₃CH₂CH₂N(B*pin*)₂), 26.5 (CH₃CH₂CH₂N(B*pin*)₂), 24.4 (CH₃CH₂CH₂N(B*pin*)₂), 10.9 (CH₃CH₂CH₂N(B*pin*)₂) ppm.

The spectroscopic data is consistent with literature data.⁶

15. 10: NMR data is same as entry 14 1n.

N(Bpin)₂

16. **1p:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 2.88-2.84 (d, 2H, CH₂), 1.67-1.60 (m, 1H, CH), N(Bpin)₂ 1.22 (s, 24H, N(Bpin)₂), 0.85-0.82 (d, 6H, CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.8 ((CH₃)₂CHCH₂N(Bpin)₂), 50.8 ((CH₃)₂CHCH₂N(Bpin)₂),30.6 ((CH₃)₂CHCH₂N(Bpin)₂), 24.7 ((CH₃)₂CHCH₂N(Bpin)₂), 19.7 ((CH₃)₂CHCH₂N(Bpin)₂) ppm.

The spectroscopic data is consistent with literature data.³

17. **1q:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 3.06-3.00(t, 2H, CH₂), 1.40- $N(Bpin)_2$ 1.38 (t, 3H, CH), 1.22 (s, 24H, N(Bpin)₂), 0.92-0.85 (m, 4H, CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.5 (CH₃CH₂CH₂CH₂N(Bpin)₂), 46.7 (CH₃CH₂CH₂CH₂N(Bpin)₂), 30.9 (CH₃CH₂CH₂CH₂CH₂N(Bpin)₂), 24.5(CH₃CH₂CH₂CH₂N(Bpin)₂), 21.0 (CH₃CH₂CH₂CH₂N(Bpin)₂) ppm.

18. **1r**: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 3.15-3.11 (t,2H, CH₂), 2.41-NC N(Bpin)₂ 2.38 (t, 2H, CH₂), 1.19 (s, 24H, N(Bpin)₂) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 81.7 (N(Bpin)₂), 79.2 (-NC), 37.2 (CH₂N(Bpin)₂), 20.96 (CH2CH₂N(Bpin)₂ppm.

19. 1s: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 2.66 (s, 3H, CH₃), 1.22 (s, 24H, N(Bpin)₂); ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 82.95 (CH₃N(Bpin)₂), 30.5 (CH₃N(Bpin)₂), 24.4(CH₃CH₂N(Bpin)₂) ppm.

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Note: In case of **1t** (nicotinamide) and **1u** (5-bromo nicotinamide), no reduction product was observed even after using 6 equivalents of HBpin.

Control experiments for the mechanistic investigation

To understand the mechanism associated with hydroboration of primary amide, we have carried out a series of experimental studies as mentioned below.

> Detection of molecular hydrogen in hydroboration of benzamide.

An oven dried screw-cap NMR tube was charged with benzamide (15.37 mg, 0.127 mmol, 1.0 qeuiv), HBpin (32.48 mg, 0.254 mmol, 2.0 equiv), and benzene-d₆ (500 μ L) and immediate evolution of molecular hydrogen was observed which was characterized through ¹H NMR spectroscopy. A sharp resonance at δ 4.48 ppm in ¹H NMR spectrum indicates the formation of molecular hydrogen (Scheme S2 and Figure S1).

PhCONH₂ + 2.0 equiv HBpin
$$\longrightarrow$$
 Ph H_2^{\uparrow}
0.127 mmol 0.254 mmol 0.254 mmol $\delta = 4.47$ ppm

Scheme S2. Detection of molecular hydrogen in hydroboration of benzamide.



Figure S1. ¹H NMR spectrum of molecular hydrogen recorded in C₆D₆ for hydroboration of benzamide.

> Characterization of in situ generated imine intermediate.

A Screw cap tube was charged with benzamide (15.37 mg, 0.127 mmol), HBpin (56.88 mg, 0.444 mmol) in an argon filled glovebox and after the reaction time of 14 h, at room temperature, ¹H NMR of the reaction mixture was recorded in CDCl₃. A sharp resonance at δ 10.24 ppm for benzamide was observed in ¹H NMR spectrum, indicating an imine intermediate formation (Scheme S3 and Figure S3).

PhCONH₂ + 3.5 equiv HBpin
$$\xrightarrow{14 \text{ h, RT}}$$
 - Bpin-O-Bpin $\xrightarrow{\text{Ph}}$ Ph $\xrightarrow{\text{Ph}}$ N $\xrightarrow{\text{Bpin}}$
0.127 mmol 0.444 mmol 1 H NMR:
 δ 10.24 ppm

Scheme S3. Synthetic scheme for the formation of intermediate imine.



Figure S2. ¹H NMR spectrum of in situ generated N-borylated imine recorded in CDCl₃.

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Reaction in presence of TMEDA.

PhCONH2 + 4.5 equiv HBpin
0.127 mmol
$$\frac{TMEDA (10 \text{ mol}\%)}{\text{Neat, 100 °C, 13 h}}$$

- Bpin-O-Bpin
 $- 2 \text{ H}_2^{\uparrow}$ Ph $\widehat{N(Bpin)_2}$
78 %

Scheme S4. Synthetic scheme for the hydroboration of benzamide in presence of TMEDA.

To identify whether the BH₃ is behaving as hidden catalyst in the hydroboration of benzamide or not, we performed the reaction in presence of N, N, N', N'-Tetramethylethylenediamine (TMEDA) as a BH₃ scavenger. A screw cap NMR tube was charged with HBpin (81.22 mg, 0.635 mmol, 5.0 equiv), and TEMDA (0.15 mg, 10 mol%) inside the argon-filled glovebox. Subsequently benzamide (15.37 mg, 0.127 mmol, 1.0 equiv), was added to the reaction mixture and heated at 100 °C for 13h in an oil bath. After completion of the reaction 0.127 mmol mesitylene was added as an internal standard prior to the NMR measurement in CDCl₃ solvent. For the mentioned reaction, 78% NMR yield (98% for without TMEDA) for the corresponding N-borylated product was observed (Scheme S4) suggesting for the non-involvement or very less influence of BH_3 as an active catalyst for the hydroboration of benzamide.



Figure S3. ¹H NMR for the hydroboration of benzamide in presence of TMEDA.

Effect of heating on HBpin

In another experiment, to identify whether HBpin is decomposing into BH_3 , HBpin was heated at 100 °C for 13 h in benzene-d₆. ¹¹B NMR shows a doublet at 27.78-29.18 ppm similar to HBpin which clearly indicates that only HBpin is doing the reduction not the BH_3 (figure S4).



Figure S4. ¹¹B NMR spectrum of heating of HBpin at 100 °C for 13 h (C₆D₆, 200 MHz, 298 K).

Competitive experiment for chemo selective hydroboration in case of ester, aldehyde,

alkene and alkyne with amide functionality

Selective hydroboration of benzamide over methyl benzoate, benzaldehyde, styrene and phenyl acetylene in presence of 5 equiv. of HBpin has been performed and only negligible conversion was found in case of ester which is confirmed by ¹H NMR spectra and in other cases significant conversion was observed (Scheme S5).

Methyl benzoate (17.28 mg, 0.127 mmol), benzaldehyde (13.46 mg, 0.127 mmol), styrene (13.26 mg, 0.127 mmol) and phenyl acetylene (12.96 mg, 0.127 mmol) were charged for separate reactions with benzamide (15.37 mg, 0.127 mmol), pinacolborane (82 mg, 0.635 mmol) in a Schlenk tube/ screw cap nmr tube inside the glove box. The reaction mixture was heated at 100 °C for 13 h. The progress of the reaction was monitored by ¹H NMR after addition of mesitylene (0.127 mmol) as an internal standard in CDCl₃. A sharp resonance at δ = 4.26 ppm indicates for the formation of hydroboration product from benzamide and resonance at δ = 4.95 ppm (figure S5)

indicates for the formation of hydroboration product from ester. Similarly, hydroborated product of benzaldehyde shows singlet resonance at δ = 4.96 ppm (figure S6), while hydroborated product of styrene shows triplet resonance at δ = 2.77-2.81 ppm (figure S7) and hydroborated product of phenyl acetylene shows doublet resonance at δ = 6.19-6.23 ppm (figure S8).



eme S5. Chemo selective experiments for hydroboration of benzamide vs methyl benzoate, benzaldehyde, styrene and phenyl acetylene, respectively in presence of 5 equiv. of HBpin.



gure S5. ¹H NMR for competitive experiments for hydroboration for ester and amide.



Figure S6. ¹H NMR for competitive experiments for hydroboration of aldehyde and amide.



gure S7. ¹H NMR for competitive experiments for hydroboration of alkene and amide.



Figure S8. ¹H NMR for competitive experiments for hydroboration of alkyne and amide.

General procedure for the hydroboration of secondary amides



Scheme S6. General procedure for the hydroboration of secondary amides.

Secondary amide (0.127 mmol), pinacolborane (3.0 equiv., 0.381mmol) were mixed together in a Schlenk tube or in a screw cap NMR tube inside the glove box. The reaction mixture was allowed to heat at 100 °C for 18 h without adding any solvent. Volatiles of the mixture were removed under reduced pressure and mesitylene (0.127 mmol) as an internal standard, was added while making the NMR in appropriate deuterated solvent. The progress of the reaction was monitored by ¹H NMR, which indicated the completion of the reaction by the appearance of a new R-C H_2 -NR'(Bpin) resonance (Scheme S5).

Entry	Substrate	Temperature	Time	NMR	Product
		(°C)	(h)	yield (%)	
1.	H N O Me	100	18	73	2a
2.	Br H Me	100	18	72	2b
3.	Me N O Me	100	18	81	2c
4.	NH NH	100	18	70	2d

Table S4. Substrate scope for the secondary amide hydroboration:

5.	HO Ne O	100	18	95	2e
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Analytical data (¹H and ¹³C NMR) of the borylamines of the corresponding secondary amides:

1. **2a:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.28-7.27 (m, 4H, *Ph*), 6.97-6.90 (m, 1H, *Ph*), 3.59- **Bpin** 3.54 (q, 2H, CH₂), 1.31 (s, 12H, PhN(B*pin*)CH₂CH₃), 1.19-1.15 (t,3H, ^N Me PhN(B*pin*)CH₂CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 148.3 (*Ph*), 129.0 (*Ph*), 119.8(*Ph*), 117.1(*Ph*), 112.7 (*Ph*), 83.0 (PhN(B*pin*)CH₂Me), 38.3 (PhN(B*pin*)CH₂Me), 24.4 (PhN(B*pin*)CH₂Ph), 14.7 (PhN(B*pin*)CH₂Me)ppm.

The spectroscopic data is consistent with literature data.¹¹

2. **2b:** ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.34-7.32 (d, 2H, *Ph*), 7.15-7.13 (d, 2H, *Ph*), 3.54- **Bpin** 3.48 (q, 2H, CH₂), 1.28 (s, 12H, PhN(B*pin*)CH₂CH₃), 1.14-1.11(t, 3H, Me PhN(B*pin*)CH₂CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 147.1 (*Ar*), 131.6 (*Ar*), 121.1 (*Ar*), 114.5 (*Ar*), 114.2 (*Ar*), 82.9 (ArN(B*pin*)CH₂Me), 38.3 (ArN(B*pin*)CH₂Me), 24.6 (ArN(B*pin*)CH₂Ph), 14.4 (ArN(B*pin*)CH₂Me)ppm.

The spectroscopic data is consistent with literature data.⁷

3. 2c: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.15-7.08 (m, 3H, Ph), 6.80 (s, 1H, Ph), 3.58-3.53
Bpin (q, 2H, CH₂), 2.35 (s, 3H, Me), 1.31 (s, 12H, PhN(Bpin)CH₂CH₃), 1.18Me Me 1.15 (t, 3H, PhN(Bpin)CH₂CH₃) ppm; ¹³C{¹H} NMR (CDCl₃, 100.5 MHz, 298 K): δ 148.3 (Ar), 138.6 (Ar), 128.9 (Ar), 117.8 (Ar), 113.3 (Ar), 109.7 (Ar), 82.9 (PhN(Bpin)CH₂Me), 38.2 (ArN(Bpin)CH₂Me), 24.3 (ArN(Bpin)CH₂Ph), 21.0 (Ar-Me), 14.7 (ArN(Bpin)CH₂Me)ppm.



4. 2d: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 3.02-2.99 (t, 4H, RCH₂N(Bpin)CH₂R), 1.60 (br, 4H, RCH₂CH₂N(Bpin)CH₂CH₂R), 1.49 (br, 4H, -CH₂CH₂CH₂N(Bpin)CH₂CH₂CH₂), 1.19 (s, 12H, $CH_2CH_2CH_2N(Bpin)CH_2CH_2CH_2-)$ ppm; ¹³C{¹H} NMR (CDCl₃, 100.6 MHz, 298 K): δ 82.0 (PhCH₂N(Me)Bpin), 47.3 (-CH₂CH₂CH₂N(Bpin)CH₂CH₂CH₂-), 28.7 (-24.5(-CH₂CH₂CH₂N(Bpin)CH₂CH₂CH₂-), 26.6 $(-CH_2CH_2CH_2N(Bpin)CH_2CH_2CH_2-),$ CH₂CH₂CH₂N(Bpin)CH₂CH₂CH₂-) ppm.

The spectroscopic data is consistent with literature data.⁸

5. 2e: ¹H NMR (CDCl₃, 200 MHz, 298 K): δ 7.14-7.12 (d, 2H, Ph), 7.00-6.98 (d, 2H, Ph), 3.51-3.46 (q, 2H, CH₂), 1.33 (s, 12H, PhN(Bpin)CH₂CH₃), 1.28 (s, 12H, Bpin *pin*BO-PhN(Bpin)CH₂CH₃), 1.13-1.09(t, 3H, PhN(Bpin)CH₂CH₃) Ν. Me ppm; ${}^{13}C{}^{1}H$ NMR (CDCl₃, 100.6 MHz, 298 K): δ 148.0 (Ar), н н **BpinO** 141.0 (Ar), 128.8 (Ar), 122.2 (Ar), 119.2 (Ar), 82.9 (ArN(Bpin)CH₂Me), 42.2 $(ArN(Bpin)CH_2Me),$ 24.4 (ArN(Bpin)CH₂Ph), 14.4 (ArN(Bpin)CH₂Me)ppm.

Representative NMR Spectra:





gure S12. ¹³C NMR spectrum of **1a'** (DMSO-d₆,100.6 MHz, 298 K).



gure S14. ¹³C NMR spectrum of **1b** (CDCl₃, 100.5 MHz, 298 K).



gure S16. 13 C NMR Spectrum of **1c** (CDCl₃, 200 MHz, 298 K).



gure S18. ¹³C NMR spectrum of 1d (CDCl₃, 100.6 MHz, 298 K).



gure S20. ¹³C NMR spectrum of **1e** (CDCl₃, 100.6 MHz, 298 K).



gure S22. 13 C NMR spectrum of **1e'** (DMSO-d₆, 100.6 MHz, 298 K).





Figure S25. ¹⁹F NMR Spectrum of **1f** (CDCl₃, 400 MHz, 298 K).



gure S26. ¹H NMR Spectrum of 1g (CDCl₃, 200 MHz, 298 K).



Figure S28. ¹⁹F NMR Spectrum of **1g** (CDCl₃, 400 MHz, 298 K).



gure S30. ¹³C NMR spectrum of **1h** (CDCl₃, 100.6 MHz, 298 K).



Figure S32. ¹H NMR spectrum of 1i (CDCl₃, 200 MHz, 298 K).



gure S34. ¹H NMR Spectrum of 1j (CDCl₃, 200 MHz, 298 K).



gure S36. ¹H NMR Spectrum of 1k (CDCl₃, 200 MHz, 298 K).



gure S38. ¹H NMR Spectrum of **11** (CDCl₃, 200 MHz, 298 K).



gure S40. ¹H NMR Spectrum of 1m (CDCl₃, 200 MHz, 298 K).



gure S42. ¹H NMR Spectrum of 1n (CDCl₃, 200 MHz, 298 K).



Note: ¹H NMR spectra of **10** is same as **1n**.



gure S45. ¹³C NMR spectrum of **1p** (CDCl₃, 100.5 MHz, 298 K).



gure S47. 13 C NMR spectrum of 1q (CDCl₃, 100.5 MHz, 298 K).



gure S49. ¹³C NMR spectrum of **1r** (CDCl₃, 100.6 MHz, 298 K).



Figure S51. ¹³C NMR spectrum of **1s** (CDCl₃, 100.6 MHz, 298 K).



gure S53. ¹³C NMR spectrum of **2a** (CDCl₃, 100.6 MHz, 298 K).



gure S55. ¹³C NMR spectrum of **2b** (CDCl₃, 100.6 MHz, 298 K).



gure S57. ¹³C NMR spectrum of **2c** (CDCl₃, 100.5 MHz, 298 K).



gure S59. ¹³C NMR spectrum of 2d (CDCl₃, 100.6 MHz, 298 K).



Figure S61. ¹³C NMR spectrum of **2e** (CDCl₃, 100.6 MHz, 298 K).

🚢 Kinetic study

Kinetics for hydroboration of amides was studied by performing NMR scale reactions. Benzamide is used as substrate for the kinetic study because it is the simplest compound of aromatic amides. The reaction was monitored by ¹H NMR analysis at time interval of 10 min and the reaction was performed at 100 °C. The kinetic data was obtained by the proton integral of $-CH_2$ peak relative to CH₃ protons of mesitylene (internal standard).

General procedure for the rate determination

In argon atmosphere, required amount of HBpin and benzamide were taken along with mesitylene. NMR was recorded at room temperature considering it at T=0 min (no product was observed). Then NMR tube was kept in oil bath, 100 °C temperature and spectra was recorded at the interval of 10 min up to 1 hour.

> HBpin rate order assessment

Varying concentration of HBpin while keeping benzamide concentration constant.

HBpin [M]	Benzamide [M]	Rate (M/min)	R ²
0.635	0.127	$8.06 \times 10^{-4} \pm 2.41 \times 10^{-5}$	0.99555
0.508	0.127	$7.29 \times 10^{-4} \pm 2.61 \times 10^{-5}$	0.99360
0.381	0.127	$5.17 \times 10^{-4} \pm 1.70 \times 10^{-5}$	0.99463



Figure S62. Plot of conc. of product vs. time at different HBpin concentrations.



Figure S63. Plot of log (rate) vs. log (conc. of product). This shows first order dependence over HBpin.

Benzamide rate order assessment

Varying concentration of HBpin while keeping benzamide concentration constant.

HBpin [M]	Benzamide [M]	Rate (M/min)	R ²
0.635	0.381	$1.85 \times 10^{-4} \pm 1.73 \times 10^{-5}$	0.95804
0.635	0.254	$5.60 \times 10^{-4} \pm 2.50 \times 10^{-5}$	0.99006
0.635	0.063	$4.68 \times 10^{-4} \pm 3.08 \times 10^{-5}$	0.97824



Figure S64. Plot of conc. of product vs. time at different benzamide concentrations. The reaction follows first order dependence over benzamide.

Details of DFT calculations:

The quantum chemical calculations have been performed using density functional theory (DFT), as a tool with the aid of the Turbomole 7.0 suite of programs.⁹ The PBE functional,¹⁰ and the TZVP¹¹ basis set has been employed. The resolution of identity (RI),¹² along with the multipole accelerated resolution of identity (marij)¹³ approximations have been used for an accurate and efficient treatment of the electronic Coulomb term in the DFT calculations. The option "disp" provided in the Turbomole package (DFT-D3) was employed for dispersion-corrected DFT calculations.¹⁴ The values reported are ΔG values, with zero point energy corrections, internal energy and entropic contributions included through frequency calculations on the optimized minima with the temperature taken to be 298.15 K. Harmonic frequency calculations were performed for all stationary points to confirm them as a local minima or transition state structures. Furthermore, intrinsic reaction coordinate (IRC) calculations were carried out with all transition states to further confirm that they were the correct transition state, yielding the correct reactant and product structures.¹⁵

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PBE/TZVP optimized geometries for all the compounds and transition states

HBpin

В	-0.002940	-0.022707	1.379727
0	-1.256412	-0.097901	1.943236
0	-0.001661	0.004803	0.003578
С	-1.379910	-0.283326	-0.424518
С	-2.222148	0.103121	0.851007
С	-1.684023	0.542131	-1.667273
С	-1.431267	-1.778098	-0.750980
Η	-1.208299	-2.387917	0.135810
Η	-0.675978	-2.001305	-1.516525
Η	-2.417030	-2.070548	-1.138994
Η	-1.041854	0.211961	-2.495326
Н	-1.500296	1.609691	-1.498585
Η	-2.732217	0.408439	-1.973182
С	-3.436256	-0.778575	1.108553
Η	-3.947403	-0.445026	2.022164
Η	-3.153306	-1.829682	1.241207
Η	-4.148906	-0.707921	0.273440
Н	-3.392159	1.807092	0.138702
Н	-1.754037	2.232777	0.706883
С	-2.620143	1.580999	0.887438
Η	-3.021576	1.816207	1.882447
Η	0.999625	0.015650	2.030356
Ber	zamide		

С	0.356516	-0.715766	0.220395
С	0.268498	-0.364556	1.575127
С	1.345287	0.247878	2.215627
С	2.516203	0.532016	1.504578
С	2.604010	0.204573	0.148382
С	1.530769	-0.420291	-0.490719
Н	-0.660776	-0.578258	2.105859
Н	1.270789	0.510327	3.272270
Н	3.356410	1.017981	2.004985
Н	3.507848	0.442702	-0.415342
Н	1.599648	-0.637610	-1.558988
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Ν	0.789539	-4.449663	2.127348
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Η	-0.207575	0.929279	-1.827308
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С	-4.586246	3.879345	5.395316	Н	2.922665	4.209006	4.488064
С	-3.416724	3.861026	6.157801	Н	0.990778	2.905224	2.643172
С	-1.846852	2.729524	7.723397	Н	0.898664	1.240367	3.256653
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62

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