A triple-pore tessellated square array by a metal-hexagonal ligand with reinforced tetra-

connectors

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1. General Procedures

All starting materials were purchased from Aldrich and Alfa Aesar and used without further purification. Compounds 5-Methoxyisophthalaldehyde, 3, 5-diformylbromobenzene and 4'-(4-boronophenyl)-terpyridine were synthesized according to the reported methods.^{S1-S3} Column chromatography was conducted using basic Al₂O₃ (Sinopharm Chemical Reagents Co. Ltd, 200-300 mesh) or SiO₂ (Qingdao Haiyang Chemical Co., Ltd, 200-300 mesh) and the separated products were confirmed by ¹H NMR and ¹³C NMR spectra using a Bruker Avance 400-MHz and 500-MHz NMR spectrometers in CDCl₃, CD₃OD, and CD₃CN with a TMS standard. Transmission electron microscopy (TEM) was conducted on JEOL 2010. Electro-spray ionization (ESI) mass spectra were recorded with a Bruker microOTOF-QII or a Waters Synapt HDMS G2-Si instrument, using solutions of 0.01 mg/mL in CHCl₃/MeCN (1:3, v/v) for ligands and 0.2 mg/mL in MeCN or MeCN/MeOH (3:1, v/v) for complexes. TWIM MS. ESI mass spectrometry and traveling wave ion mobility (TWIM) experiments were conducted on a Waters Synapt HDMS G2-Si instrument with a LockSpray ESI source, using the following parameters: ESI capillary voltage, 1.3-3.0 kV; sample cone voltage, 20-25 V; extraction cone voltage, 1.1-3 V; desolvation gas flow, 800 L/h (N₂); trap collision energy (CE), 4 V; transfer CE, 0 V; trap gas flow, 2.0 mL/min (Ar); source temperature, 30 °C; and desolvation temperature, 30 °C. All samples were dissolved in CH₃CN or CH₃CN/CH₃NO₂ (1:1, v/v) and then infused into the -100, KD Scientific). For TWIM experiments, the helium cell gas flow was held at 180.0 mL/min and the ion mobility cell gas flow was held at 90.0 mL/min (N₂). The TWIM DC traveling wave velocity and height were set as 683 m/s and 26.3 V, respectively. Data were collected and analyzed by using MassLynx 4.2 and DriftScope 2.4 (Waters).

TEM. The samples were dissolved in MeCN at a concentration of $\sim 10^{-7}$ M. The solutions were dropped cast on to a carbon-coated Cu grid (300-400 mesh) and the extra solution was blotted by filter paper to avoid aggregation. The TEM images of the samples were taken with a JEOL 2010 Transmission Electron Microscope.

2. Synthesis of Hexagon-Containing Metallo-organic Ligands L1 and L2





Scheme S1: The synthetic route of metallo-organic ligand LA and parallelogram $Fe_2(LA)_2$. Reagents and conditions: (i) 2-acetylpyridine, KOH, EtOH, NH₃•H₂O, reflux; (ii) RuCl₃·3H₂O, EtOH, reflux, 1 d; (iii) N-ethyl morpholine, MeOH:CHCl₃ (v/v, 1:1), reflux, 1 d; (iv) RuCl₃·3H₂O, EtOH, reflux, 1 d; (v) Pd(PPh₃)₄, THF, NaOH, reflux, 1 d; (vi) Br₂, CHCl₃, reflux; (vii) N-ethyl morpholine, MeOH:CHCl₃ (v/v, 1:1), reflux, 2 d; (viii) Tpy-B(OH)₂, Pd(PPh₃)₄, CH₃CN/CH₃OH(V:V, 2:1), K₂CO₃, reflux.



Compound S2: To a solution of 5-Methoxyisophthalaldehyde (3.2 g, 2 mmol) in EtOH (200 mL), 2-acetylpyridine (5.1 g, 4.8 mmol) and NaOH powder (240.0 mg, 6 mmol) was added. After stirring at room temperature for 24 h, aqueous NH₃•H₂O (28%, 150 mL) was added, the resulting mixture was refluxed for 20 h. After cooling to room temperature, the solid was collected by suction filtration and was washed with CH₃OH to give the product as a white solid: 8.1 g, 72.6%). ¹H NMR (500 MHz, CDCl₃) δ 8.83 (s, 4H, tpy- $H^{3',5'}$), 8.77-8.76 (d, 4H, *J* = 2Hz, tpy- $H^{3,3''}$), 8.72-8.70 (d, 4H, *J* = 4Hz, tpy- $H^{6,6''}$), 7.99 (s, 1H, Ph- H^a), 7.93-7.90 (t, 4H, tpy- $H^{4,4''}$), 7.53 (s, 2H, PH- H^b), 7.40-7.38 (t, 4H, tpy- $H^{5,5''}$), 4.04 (s, 3H, H^{-OMe}). ¹³C NMR (101 MHz, CDCl₃) δ 160.67, 156.24, 156.02, 150.21, 149.15, 140.97, 136.89, 123.85, 121.45, 119.33, 119.06, 113.68, 55.87.



Compound S3: To a solution of **S2** (200.0 mg, 0.4 mmol) and RuCl₃•3H₂O (105.0 mg, 0.4 mmol) in EtOH (50 mL). The mixture was stirred at 75 °C for 24 h. After cooling to ambient temperature, the precipitates were filtered and washed with MeOH to afford **S3**: 250.0 mg, 90%.



Compound S4: To a flask containing a mixture of **S3** (300 mg, 0.30 mmol)) and **S2** (521 mg, 0.91 mmol), McOH (150 mL), CHCl₃ (150 mL), and 10 drops of N-ethylmorpholine were added. The mixture was stirred at 75 °C for 48 h. After cooling to ambient temperature, the solvent was evaporated in vacuo and the residue was purified by column chromatography (Al₂O₃), eluting with a mixture of MeOH and CH₂Cl₂. The complex was obtained, as a red precipitate: 420 mg, 67.1%.¹H NMR (400 MHz, CD₃OD) δ 9.67 (s, 4H, A-tpy-H^{3',5}), 9.48 (s, 4H, B-tpy-H^{3',5'}), 9.22 (d, J = 8.0 Hz, 4H, A-tpy-H^{3,3''}), 9.07 (m, 8H, C-tpy-H^{3',5'}, B-tpy-H^{3,3''}), 8.99 (d, J = 8.0 Hz, 4H, C-tpy-H^{3,3''}), 8.94 (s, 1H,H^b), 8.89 (d, J = 8.0 Hz, 4H, C-tpy-H^{6,6''}), 8.53(s, 2H, H^c), 8.34 (t, J = 8.0 Hz, 4H, C-tpy-H^{4,4''}), 8.13 (s, 2H, H^a), 8.07 (m, 10H, A-tpy-H^{4,4''}, B-tpy-H^{4,4''}, H^d), 7.84 (s, 2H, H^c), 7.78 (t, J = 8.0Hz, 4H, C-tpy-H^{4,4''}), 8.13 (s, 2H, H^a), 8.07 (m, 10H, A-tpy-H^{4,4''}, B-tpy-H^{4,4''}, H^d), 7.84 (s, 2H, H^c), 7.78 (t, J = 8.0Hz, 4H, C-tpy-H^{5,5''}), 7.68 (d, J = 8.0Hz, 4H, B-tpy-H^{6,6''}), 7.64 (d, J = 8.0 Hz, 4H, A-tpy-H^{6,6''}), 7.34 (m, 8H, A-tpy-H^{5,5''}, B-tpy-H^{5,5''}), 4.30 (s, 3H, H^{-OCH3}), 4.19 (s, 6H, H^{-OCH3}). ESI-MS (2055.91 calcd. For C₁₁₁H₇₈Cl₄N₁₈O₃Ru₂): m/z 748.6031 [M - 4Cl⁻]⁴⁺ (calcd m/z: 748.5245), 649.8176 [M - 3Cl⁻]³⁺ (calcd m/z: 649.8503), m/z 992.7487 [M - 2Cl⁻]²⁺ (calcd m/z: 992.5020).



Compound M1: To a solution of **S4** (200.0 mg, 0.4 mmol) and RuCl₃•3H₂O (105.0 mg, 0.4 mmol) in EtOH (50 mL). The mixture was stirred at 75 °C for 24 h. After cooling to ambient temperature, the precipitates were filtered and washed with MeOH to afford **M1**: 250.0 mg, 90%.



Compound S6: To a solution of NaOH powder (240.0 mg, 6 mmol) in EtOH (200 mL), 3, 5-diformylbromobenzene **S5** (3.54 g, 2 mmol) and 2-acetylpyridine (5.14 g, 4.8 mmol) was added. After stirring at room temperature for 24 h, aqueous NH₃•H₂O (28%, 150 mL) was added, the resulting mixture was refluxed for 20 h. After cooling to room temperature, the solid was collected by suction filtration and was washed with CH₃OH to give the product as a white solid: 9.97 g, 85.0%). ¹H NMR (400 MHz, CDCl₃) δ 8.78 (s, 4H, tpy- $H^{3',5'}$), 8.75-8.74 (d, *J* = 4 Hz, 4H, tpy- $H^{6,6''}$), 8.71- 8.69 (d, *J* = 8 Hz, 4H, tpy- $H^{3,3''}$), 8.29 (s, 1H, Ph- H^a), 8.12(s, 2H, Ph- H^b), 7.92-7.88 (t, *J* = 16 Hz, 4H, tpy- $H^{4,4''}$), 7.39-7.36 (t, *J* = 12 Hz, 4H, tpy- $H^{5,5''}$). ¹³C NMR (101 MHz, CDCl₃) δ 156.21, 155.99, 149.18, 148.75, 141.50, 136.96, 130.78, 125.13, 124.01, 121.47, 119.10, 77.33, 77.22, 77.01, 76.70. ESI-MS (621.54 calcd. For C₃₆H₂₅BrN₆): m\z 622.47[M+H⁺]⁺ (calcd m/z: 622.55).



Compound S8: 1, 3-dimethoxybromobenzene **S7** (0.78 g, 3.6 mmol) and **S6** (2.00 g, 3.4 mmol) was added to a 500 mL flask, then THF (250 mL) and NaOH (0.41 g, 10.3 mmol) in 10 mL of water was added. The system was degassed for 10 min, and Pd(PPh₃)₄ (0.46 g, 0.4 mmol) as the catalyst was added. The mixture was stirred at 85 °C

under nitrogen for 12 h, after cooled to ambient temperature, then concentrated in vacuo followed by column chromatography (Al₂O₃), eluting with the mixture of petroleum ether and CH₂Cl₂ to pure the product, as white solid: 1.50 g, 65%. ¹H NMR (500 MHz, CDCl₃) δ 8.87 (s, 4H, tpy- $H^{3,5'}$), 8.75-8.74 (d, J = 4 Hz, 4H, tpy- $H^{6,6''}$), 8.72-8.70 (d, J = 8 Hz, 4H, tpy- $H^{3,3''}$), 8.38 (s, 1H, H^d), 7.98 (s, 2H, H^c), 7.92-7.89 (t, J = 12 Hz, 4H, tpy- $H^{4,4''}$), 7.38-7.35 (m, 5H, tpy- $H^{5,5''}$, Ph- H^a) 6.73-6.72 (d, J = 4 Hz, 2H, H^b), 3.79 (s, 6H, H^{-OCH3}). ¹³C NMR (101 MHz, CDCl₃) δ 161.86, 158.19, 157.70, 156.34, 155.68, 150.45, 150.29, 149.10, 139.41, 138.90, 136.90, 131.04, 130.85, 124.66, 123.76, 121.45, 119.49, 118.61, 118.47, 105.22, 104.14, 55.93. ESI-MS (676.26 calcd. For C₄₆H₃₂N₆O₂): m/z 699.28[M+Na⁺]⁺ (calcd m/z: 699.24).



Compound S9: To a solution of **S8** (500.0 mg, 0.74 mmol) in CHCl₃ (50 mL), a solution of Br₂ (5 mL, 97.5 mmol) in CHCl₃ (5 mL) was added dropwise. After refluxing for 24 h, the mixture was washed by saturated NaHSO₃ until colorless. The organic layer was dried (anhydrous Na₂SO₄), and then concentrated in vacuo to give **S9**, as white solid: 494.0 mg, 80%. ¹H NMR (500 MHz, CDCl₃) δ 8.87 (s, 4H, tpy- $H^{3',5'}$), 8.76-8.75 (d, *J* = 4 Hz, 4H, tpy- $H^{6,6''}$), 8.73-8.71 (d, *J* = 8 Hz, 4H, tpy- $H^{3,3''}$), 8.49 (s, 1H, H^c), 8.12 (s, 2H, H^b), 7.94-7.91 (t, *J* = 12 Hz, 4H, tpy- $H^{4,4''}$), 7.86 (s, 1H, H^a), 7.41-7.38 (d, J = 12 Hz, 4H, tpy- $H^{5,5''}$), 3.51 (s, 6H, H^{-OCH3}). ¹³C NMR (101 MHz, CDCl₃) δ 156.12,

155.06, 149.75, 149.11, 139.72, 137.01, 135.53, 134.44, 131.48, 129.75, 125.94, 123.95, 121.52, 119.31, 113.31, 60.85.¹³C NMR (101 MHz, CDCl₃) δ 156.29, 156.12, 149.90, 149.21, 136.88, 136.45, 133.73, 133.66, 128.52, 127.38, 125.80, 123.86, 121.39, 119.08, 77.33, 77.01, 76.69. ESI-MS (834.08 calcd. For C₄₆H₃₀N₆O₂Br₂): m\z 857.11[M+Na⁺]⁺ (calcd m/z: 857.07).



Compound S10: To a solution of **S9** (200.0 mg, 0.4 mmol) and RuCl₃•3H₂O (105.0 mg, 0.4 mmol) in EtOH (50 mL). The mixture was stirred at 75 °C for 24 h. After cooling to ambient temperature, the precipitates were filtered and washed with MeOH to afford **S10**: 250.0 mg, 90%.



Compound M2: To a flask containing a mixture of S10 (330 mg, 0.26 mmol) and S2 (393 mg, 0.63 mmol), MeOH

(150 mL), CHCl₃ (150 mL), and 10 drops of N-ethylmorpholine were added. The mixture was stirred at 75 °C for 48 h. After cooling to ambient temperature, the solvent was evaporated in vacuo and the residue was purified by column chromatography (Al₂O₃), eluting with a mixture of MeOH and CH₂Cl₂. The complex was obtained, as a red precipitate: 150 mg, 24%.¹H NMR (500 MHz, MeOD) δ 9.74 (s, 4H, A-tpy-H^{3',5'}), 9.56 (s, 1H, H^b), 9.47 (s, 4H, B-tpy-H^{3',5'}), 9.23 (d, *J* =10.0 Hz, 4H, A-tpy-H^{3,3''}), 9.03 (d, *J* = 10.0 Hz, 4H, B-tpy-H^{3,3''}), 8.96 (s, 4H, C-tpy-H^{3',5'}), 8.86 (s, 2H, H^c), 8.78 (m, 8H, C-tpy-H^{3,3''}, C-tpy-H^{6,6''}), 8.69 (m, 4H, H^e, H^a), 8.46 (m, 12H, A-tpy-H^{4,4''}, B-tpy-H^{4,4''}, C-tpy-H^{4,4''}), 7.70 (d, *J* = 5.0 Hz, 4H, A-tpy-H^{6,6''}), 7.63 (d, *J* = 5.0 Hz, 4H, B-tpy-H^{6,6''}), 7.57(t, *J* = 5.0 Hz, 4H, C-tpy-H^{5,5''}), 7.34(m, 8H, A-tpy-H^{5,5''}), 3.75 (s, 6H, H^{-OCH3}). ESI-MS (2417.56 calcd. For C₁₁₆H₇₆Br₄Cl₄N₁₈O₂Ru₂): m/z 568.9478 [M - 4Cl⁻]⁴⁺ (calcd m/z: 568.9370), 770.6383 [M - 3Cl⁻]³⁺ (calcd m/z: 770.4003), m/z 1173.5205 [M - 2Cl⁻]²⁺ (calcd m/z: 1173.3270).



Compound H1: The M1 (62 mg, 0.025 mmol) and M2 (58 mg, 0.025 mmol) was added to a 250 mL flask, then

100 mL CH₃OH and 100 mL CHCl₃ was added as solvent. After adding 10 drops N-ethyl morpholine, the suspension was refluxed for 48 h. The solvent was evaporated in vacuo to give a residue that was purified with column chromatography (SiO₂) using a H₂O/MeCN/sat.KNO_{3(aq)} (1:12:1; v/v/v) mixture as eluent; the complex was obtained, as a red precipitate: 40 mg, 42%. ¹H NMR (500 MHz, CD₃OD) δ 9.31 (m, 24H, A-tpy-H^{3,5}, B-tpy-H^{3,5}, C-tpy-H^{3,5}, D-tpy-H^{3,5}, F-tpy-H^{3,5}), 9.20 (s, 1H, H^h), 9.18 (s, 2H, Hⁱ), 9.12 (s, 2H, H^c), 9.01 (s, 1H, H^b), 8.84 (m, 24H, A-tpy-H^{3,3}, B-tpy-H^{3,3}, C-tpy-H^{3,3}, D-tpy-H^{3,3}, F-tpy-H^{3,3}, F-tpy-H^{3,3}), 8.64 (s, 2H, Hⁱ), 8.60 (m, 2H, H^s), 8.56 (m, 2H, Hⁱ), 8.05 (m, 31H, A-tpy-H^{4,4}, B-tpy-H^{4,4}, C-tpy-H^{4,4}, D-tpy-H^{4,4}, F-tpy-H^{4,4}, F-tpy-H^{4,4}, H^m, H^a, H^d, H^c), 7.57 (m, 24H, A-tpy-H^{6,6*}, B-tpy-H^{6,6*}, C-tpy-H^{6,6*}, E-tpy-H^{6,6*}, F-tpy-H^{6,6*}), 7.29 (m, 24H, A-tpy-H^{5,5*}, B-tpy-H^{5,5*}, F-tpy-H^{5,5*}, F-tpy-H^{6,5*}, C-tpy-H^{6,5*}, F-tpy-H^{6,6*}), 3.78 (s, 6H, H^{-OCH3}). ESI-MS (6033.84 calcd. For C₂₂₉H₁₆₀Br₂F₇₂N₃₆O₇P₁₂Ru₆): m/z 525.3549 [M – 9PF₆⁻]⁹⁺ (calcd m/z: 525.4667), 609.4097 [M – 8PF₆⁻]⁸⁺ (calcd m/z: 609.2700), 717.1891[M – 7PF₆⁻]⁷⁺ (calcd m/z: 717.0171), 860.7324 [M – 6PF₆⁻]⁶⁺ (calcd m/z: 860.6800), 1061.9016 [M – 5PF₆⁻]⁵⁺ (calcd m/z: 1061.808), m/z 1363.6615 [M – 4PF₆⁻]⁴⁺ (calcd m/z: 1363.5).



Hexagon-Containing Metallo-organic Ligand L1: To a mixture of H1 (60.0 mg, 0.01 mmol) and 4'-(4boronophenyl)-terpyridine (42.0 mg, 0.12 mmol) in CH₃CN (30 mL), aqueous K₂CO₃ (0.24 mL, 1 M) was added. The system was pumped and backfilled with nitrogen; then Pd(PPh₃)₄ (7.1 mg, 6 unol) was added. After refluxing for 48 h under argon, the mixture was cooled to 25 °C and added access NH₄PF₆ solution to form the precipitate. The resultant precipitate was washed with 100 mL CH₃OH and purified by flash column chromatography (Al₂O₃), eluting with CH₂Cl₂/CH₃OH to give L1, as a red solid, 32.8 mg: 53% yield. ¹H NMR (400 MHz, CD₃CN) δ 9.42 – 9.20 (m, 24H, A-tpy-H^{3,5}, B-tpy-H^{3,5}, C-tpy-H^{3,5}, D-tpy-H^{3,5}, F-tpy-H^{3,5}, F-tpy-H^{3,5}), 9.12 (s, 2H, Hⁱ), 8.97 (s, 1H, H^h), 8.81 (m, 39H, A-tpy-H^{3,3}", B-tpy-H^{3,3}", C-tpy-H^{3,3}", D-tpy-H^{3,3}", F-tpy-H^{3,3}", G-tpy-H^{3,3}", G-tpy-H^{6,6}" , G-tpy-H^{3,5}, H^e, H^b), 8.61 (s, 2H, Hⁱ), 8.53 (s, 4H, H^s, H^f), 8.12-8.02 (m, 38H, A-tpy-H^{4,4}", B-tpy-H^{4,4}", C-tpy-H^{4,4}", Dtpy-H^{4,4}", E-tpy-H^{4,4}", F-tpy-H^{4,4}", G-tpy-H^{4,4}", H^s, H^c, H^d, H^y, H^s), 7.85 (s, 1H, H^m), 7.51 (m, 28H, A-tpy-H^{6,6}", B-tpy-

 $H^{6,6"}, C-tpy-H^{6,6"}, D-tpy-H^{6,6"}, E-tpy-H^{6,6"}, F-tpy-H^{6,6"}, G-tpy-H^{5,5"}), 7.26 \text{ (m, } 24\text{H, } A-tpy-H^{5,5"}, B-tpy-H^{5,5"}, C-tpy-H^{5,5"}, D-tpy-H^{5,5"}, E-tpy-H^{5,5"}, F-tpy-H^{5,5"}), 4.28 \text{ (s, } 15\text{H, } H^{-OCH3}), 3.54 \text{ (s, } 6\text{H, } H^{-OCH3}). ESI-MS (6490.76 calcd. For <math>C_{271}H_{188}F_{72}N_{42}O_7P_{12}Ru_6)$: m/z 576.3352 [M – 9PF $_6$]⁹⁺ (calcd m/z: 576.2355), 666.4828 [M – 8PF $_6$]⁸⁺ (calcd m/z: 666.3850), 782.4011[M – 7PF $_6$]⁷⁺ (calcd m/z: 782.2914), 936.9478 [M – 6PF $_6$]⁶⁺ (calcd m/z: 1477.7300).



Compound H2: The **M1** (100 mg, 0.04 mmol) and **M3** (98 mg, 0.04 mmol) was added to a 250 mL flask, then 100 mL CH₃OH and 100 mL CHCl₃ was added as solvent. After adding 10 drops N-ethyl morpholine, the suspension was refluxed for 48 h. The solvent was evaporated in vacuo to give a residue that was purified with column chromatography (SiO₂) using a H₂O/MeCN/sat.KNO_{3(aq)} (1:12:1; v/v/v) mixture as eluent; the complex was obtained, as a red precipitate: 75 mg, 49%. 1H NMR (400 MHz, MeOD) δ 9.56 (s, 8H, A-tpy-H^{3',5'}, B-tpy-H^{3',5'}), 9.52 (m, 8H, C-tpy-H^{3',5'}, D-tpy-H^{3',5'}), 9.50 (s, 8H, E-tpy-H^{3',5'}, F-tpy-H^{3',5'}), 9.46 (s, 1H, H^h), 9.32 (s, 2H, H^j), 9.16

(s, 2H, H^e), 9.01 (m, 24H, A-tpy-H^{3,3"}, B-tpy-H^{3,3"}, C-tpy-H^{3,3"}, D-tpy-H^{3,3"}, E-tpy-H^{3,3"}, F-tpy-H^{3,3"}, F-tpy-H^{3,3"}), 8.76 (m, 4H, Hⁱ, H^a), 8.74 (s, 2H, Hⁱ), 8.65 (s, 1H, H^b), 8.11 (m, 6H, H^m, H^a, H^d), 8.08 (s, 2H, H^e), 8.04 (m, 24H, A-tpy-H^{4,4"}, B-tpy-H^{4,4"}, C-tpy-H^{4,4"}, D-tpy-H^{4,4"}, E-tpy-H^{4,4"}, F-tpy-H^{4,4"}), 7.68 (m, 24H, A-tpy-H^{6,6"}, B-tpy-H^{6,6"}, C-tpy-H^{6,6"}, D-tpy-H^{6,6"}, E-tpy-H^{6,6"}, F-tpy-H^{6,6"}), 7.32 (m, 24H, A-tpy-H^{5,5"}, B-tpy-H^{5,5"}, C-tpy-H^{5,5"}, D-tpy-H^{5,5"}, E-tpy-H^{5,5"}, F-tpy-H^{5,5"}, F-tpy-H^{5,5"}, F-tpy-H^{5,5"}, F-tpy-H^{5,5"}, F-tpy-H^{5,5"}, F-tpy-H^{5,5"}, C-tpy-H^{5,5"}, D-tpy-H^{5,5"}, E-tpy-H^{5,5"}, F-tpy-H^{5,5"}), 4.29 (s, 9H, H^{-OCH3}), 3.75 (s, 6H, H^{-OCH3}). ESI-MS (5136.06 calcd. For C₂₂₇H₁₅₄Br₄N₄₈O₄₁Ru₆): m/z 508.8199 [M – 9NO₃⁻]⁹⁺ (calcd m/z: 508.6683), 580.0477 [M – 8NO₃⁻]⁸⁺ (calcd m/z: 580.0025), 671.7687 [M – 7NO₃⁻]⁷⁺ (calcd m/z: 671.7178), 794.0674 [M – 6NO₃⁻]⁶⁺ (calcd m/z: 794.005), 965.2892 [M – 5NO₃⁻]⁵⁺ (calcd m/z: 965.207), m/z 1222.1252 [M – 4NO₃⁻]⁴⁺ (calcd m/z: 1222.01).



Hexagon-Containing Metallo-organic Ligand L2: To a mixture of H2 (51 mg, 0.01 mmol) and 4'-(4boronophenyl)-terpyridine (42 mg, 0.12 mmol) in CH₃CN (30 mL), aqueous K_2CO_3 (0.24 mL, 1 M) was added. The system was pumped and backfilled with nitrogen; then Pd(PPh₃)₄ (7.1 mg, 6 umol) was added. After refluxing

for 48 h under argon, the mixture was cooled to 25 °C and added access NH₄PF₆ solution to form the precipitate. The resultant precipitate was washed with 100 mL CH₃OH and purified by flash column chromatography (Al₂O₃), eluting with CH₂Cl₂/CH₃OH to give L2, as a red solid, 30.5 mg: 47% yield. ¹H NMR (400 MHz, CD₃CN) δ 9.64-9.60 (m, 12H, D-tpy-H^{3',5'}, E-tpy-H^{3',5'}, F-tpy-H^{3',5'}), 9.46 (m, 12H, A-tpy-H^{3',5'}, B-tpy-H^{3',5'}, C-tpy-H^{3',5'}), 9.31 (s, 1H, H^h), 9.09-8.74 (m, 53H, G-tpy-H^{3',5'}, H-tpy-H^{3',5'}, A-tpy-H^{3,3"}, B-tpy-H^{3,3"}, C-tpy-H^{3,3"}, D-tpy-H^{3,3"}, E-tpy-H^{3,3"}, F-tpy-H^{3,3"} $, G-tpy-H^{3,3"}, H-tpy-H^{3,3"}, G-tpy-H^{6,6"}, H-tpy-H^{6,6"}, H^{j}, H^{e}, H^{b}), 8.65 \ (s, 2H, H^{a}), 8.44 \ (m, 8H, G-Ph-H^{x}, H-Ph-H^{x}), 8.32 \ (m, 8H, G-Ph-H^{x}), 8.32$ (m, 8H, G-Ph-H^y, H-Ph-H^y), 8.16–7.93 (m, 37H, A-tpy-H^{4,4"}, B-tpy-H^{4,4"}, C-tpy-H^{4,4"}, D-tpy-H^{4,4"}, E-tpy-H^{4,4"}, F-tpy-H^{4,4}", G-tpy-H^{4,4}", H-tpy-H^{4,4}", H^g, H^f, H^d, Hⁱ, H^c), 7.87 (s, 2H, H^m), 7.63 – 7.51 (m, 38H, A-tpy-H^{6,6}", B-tpy-H^{6,6}", C-tpy-H^{6,6}", D-tpy-H^{6,6}", E-tpy-H^{6,6}", F-tpy-H^{6,6}", G-tpy-H^{5,5}", H-tpy-H^{5,5}"), 7.29 (m, 24H, A-tpy-H^{5,5}", B-tpy-H^{5,5}", C-tpy-H^{5,5}", D-tpy-H^{5,5}", D-tpy-H^{5,5}", C-tpy-H^{5,5}", C-tpy-H^{5,5}", D-tpy-H^{5,5}", C-tpy-H^{5,5}", C-tpy-H^{5,5}", D-tpy-H^{5,5}", C-tpy-H^{5,5}", C-tpy-H^{5,5} tpy-H^{5,5}", E-tpy-H^{5,5}", F-tpy-H^{5,5}"), 4.29 (s, 9H, H^{-OCH3}), 3.53 (s, 6H, H^{-OCH3}). ESI-MS (7045.42 calcd. For $C_{311}H_{210}F_{72}N_{48}O_5P_{12}Ru_6): m/z \ 638.0178 \ [M-9PF_6^-]^{9+} \ (calcd \ m/z: \ 637.8644), \ 735.7637[M-8PF_6^-]^{8+} \ (calcd \ m/z: \ m/z) \ (calcd \ m/z: \ m/z) \ (calcd \$ 735.7175), 861.8739 $[M - 7PF_6]^{7+}$ (calcd m/z: 861.5285), 1029.1887 $[M - 6PF_6]^{6+}$ (calcd m/z: 1029.2766), $1264.2231 \ [M - 5PF_6^-]^{5+} \ (calcd \ m/z: \ 1264.124), \ m/z \ 1616.5441 \ [M - 4PF_6^-]^{4+} \ (calcd \ m/z: \ 1616.395).$

3. Experimental test of first designed dual-pore tessellated square array



Scheme S2: Experimental test of first designed dual-pore tessellated square array: MOL ligand L1 (4.0 mg, 0.61 μ mol), and Zn(NO₃)₂·6H₂O (0.19 mg, 0.61 μ mol) or (0.38 mg, 1.22 μ mol) was added in a 50 mL flask, then a solvent mixture of CH₃CN/MeOH (20 ml, V:V, 2/1) was added. The mixture was refluxed for 24 h, after cooled to ambient temperature, excess NH₄PF₆ in MeOH was added to get a red precipitate, which was filtered and washed with H₂O and MeOH, to generate a red solid.



Figure S1: (A) Comparison of the ¹H NMR spectra (500 MHz) of hexagon-containing metallo-organic ligand **L1** and the resultant assembly at different stoichiometric ratio (1:1 and 1:2) in CD₃CN; (B) ESI-MS of hexagoncontaining metallo-organic ligand **L1** and the resultant assembly at different stoichiometric ratio (1:1 and 1:2). The tpy $H^{6',6''}$ protons of free terpyridine has no expected change in chemical shift, and ESI-MS showed that the resultant assembly is mainly the peak of ligand, suggesting that such a construction has been unsuccessful, we suspected this could be put down to the molecular hexagon framework has relatively large molecular volume and mass, which breaking the coordination connection of singly bridged tpy-Zn(II)-tpy.



Figure S2: (A) Comparison of the DOSY spectra (500 MHz) of hexagon-containing metallo-organic ligand L1 and the resultant assembly at a stoichiometric ratio of 1:2 in CD_3CN , both showed narrow bands at log D = -9.71 m²s⁻¹, also confirmed that the metallo-organic ligand L1 is the major component.

4. Self-Assembly of supramolecules of tri-pore tessellated square array



Scheme S3: Self-Assembly of supramolecules of tri-pore tessellated square array.



Triple-pore tessellated square array Zn₈L2₄: MOL ligand L2 (4.0 mg, 0.56 µmol), and Zn(NO₃)₂·6H₂O (0.4 mg, 1.2 µmol) was added in a 50 ml flask, then a solvent mixture of CH₃CN/MeOH (20 ml, V:V, 2/1) was added. The mixture refluxed ambient temperature, for 12 h, after cooled NH_4PF_6 was to excess or bistrifluoromethanesulfonimide lithium salt (LiNTf₂) in MeOH was added to get a red precipitate, which was filtered and washed with H₂O and MeOH, to generate a red solid (~93%). ¹H NMR (500 MHz, CD₃CN) & 9.48-9.43(m, 12H, D-tpy-H^{3',5'}, E-tpy-H^{3',5'}, F-tpy-H^{3',5'}), 9.33-9.30(m, 12H, A-tpy-H^{3',5'}, B-tpy-H^{3',5'}, C-tpy-H^{3',5'}), 9.20(m, 8H, G-tpy-H^{3',5'}, H-tpy-H^{3',5'}), 9.09-8.94(4H, H^h, H^j, H^e, H^b), 8.92-8.82(m, 32H, A-tpy-H^{3,3"}, B-tpy-H^{3,3"}, C-tpy-H^{3,3"}, D-tpy-H^{3,3"}, E-tpy-H^{3,3"}, F-tpy-H^{3,3"}, G-tpy-H^{3,3"}, H-tpy-H^{3,3"}), 8.65-8.62 (m, 6H, H^a, H^d, H^c), 8.43-8.41(m, 4H, G-Ph-H^y, H-Ph-H^y), 8.29-8.22(m, 10H, Hⁱ, H^g, H^f, G-ph-H^{x'}, H-ph-H^{x'}), 8.16-8.07 (m, 32H, A-tpy-H^{4,4"}, B-tpy-H^{4,4"}, C-tpy-H^{4,4"}, D-tpy-H^{4,4"}, E-tpy-H^{4,4"}, F-tpy-H^{4,4"}, G-tpy-H^{4,4"}, H-tpy-H^{4,4"}), 7.96-7.63(m, 9H, G-tpy-H^{6,6"}, H-tpy-H^{6,6"}, H^m), 7.70-7.52(m, 32H, A-tpy-H^{6,6"}, B-tpy-H^{6,6"}, C-tpy-H^{6,6"}, D-tpy-H^{6,6"}, E-tpy-H^{6,6"}, F-tpy-H^{6,6"}, G-tpy-H^{5,5"}, H-tpy-H^{5,5"}), 7.48-7.29 (m, 24H, A-

tpy-H^{5,5}", B-tpy-H^{5,5}", C-tpy-H^{5,5}", D-tpy-H^{5,5}", E-tpy-H^{5,5}", F-tpy-H^{5,5}"), 4.33-4.31 (s, 9H, H^{-OCH3}), 3.68 (s, 6H, H^{-OCH3}). ESI-MS (31024.16 with 64 PF6⁻ calcd. For $C_{1244}H_{840}F_{384}N_{192}O_{20}P_{64}Ru_{24}Zn_8$): m/z 1048.3783 [M - 26PF6⁻]²⁶⁺ (calcd m/z: 1048.2769), 1096.0315 $[M - 25PF_6]^{25+}$ (calcd m/z: 1096.0064), 1147.6342 $[M - 24PF_6]^{24+}$ (calcd m/z: 1147.7133), 1203.9335 $[M - 23PF_6^{-1}]^{23+}$ (calcd m/z: 1203.9165), 1265.2479 $[M - 22PF_6^{-1}]^{22+}$ (calcd m/z: 1265.2290), m/z 1332.4351 $[M - 21PF_6]^{21+}$ (calcd m/z: 1332.3809), 1406.3335 $[M - 20PF_6]^{20+}$ (calcd m/z: 1406.248), 1488.1493 $[M - 19PF_6]^{19+}$ (calcd m/z: 1487.8905), 1578.9857 $[M - 18PF_6]^{18+}$ (calcd m/z: 1578.6044), 1679.99732 $[M - 17PF_6]^{17+}$ (calcd m/z: 1679.9905); ESI-MS (39675.11 with 64 NTf₂⁻ calcd. For $C_{1372}H_{840}F_{384}N_{256}O_{276}Ru_{24}S_{128}Zn_8$): m/z 3026.7109 [M - 12NTf₂]¹²⁺ (calcd m/z: 3026.1131), 2771.2622 [M - 12NTf₂]¹²⁺ $13NTf_2^{-13+}$ (calcd m/z: 2771.7855), 2554.0914 [M - 14NTf_2^{-14+} (calcd m/z: 2553.7904), 2365.1586 [M - $15NTf_2^{-}]^{15+}$ (calcd m/z: 2364.8613), 2200.0632 [M - $16NTf_2^{-}]^{16+}$ (calcd m/z: 2199.5483), 2054.3015 [M - 16NTf_2^{-}]^{16+} $17NTf_2^{-17+}$ (calcd m/z: 2053.684), 1924.6033 [M - $18NTf_2^{-18+}$ (calcd m/z: 1924.0267), 1808.4157 [M - 18NTf_2^{-18+} $19NTf_2^{-}$ ¹⁹⁺ (calcd m/z: 1808.0176), 1703.9834 [M - 20NTf_2^{-}]²⁰⁺ (calcd m/z: 1703.6095), 1609.6814 [M - 20NTf_2^{-}]²⁰⁺ (calcd m/z) [M - 20NTf_2^{-}]²⁰⁺ (calcd m/z $21NTf_2^{-1}^{21+}$ (calcd m/z: 1609.1449), 1523.3942 [M - $22NTf_2^{-1}^{22+}$ (calcd m/z: 1523.2680), 1445.1146 [M - $23NTf_2^{-}^{23+}$ (calcd m/z: 1444.8587), 1373.2389 [M - 24NTf_2^{-}]^{24+} (calcd m/z: 1372.9835), 1307.0752 [M - $25NTf_2^{-}$ ²⁵⁺ (calcd m/z: 1306.8584).



Figure S3: ESI–MS of Zn_8L2_4 with $64PF_6^-$. By further ion exchange, a series of exclusive peaks with successive charge states at m/z 1048.38, 1096.03, 1147.63, 1204.11, 1266.45, 1332.44, 1406.33, 1488.15, 1578.99 and 1679.97 were also observed due to the loss of sequential numbers of PF_6^- anions based on the molecular formula of $(Zn_8L_{24})^{64+}(PF_6^-)^{64-}$ with a molecular weight of 31 024 Da.



Figure S4: ESI-MS of Zn₈L2₄with 64(NTf₂⁻).



Figure S5: Theoretical (top) and experimental (bottom) isotope patterns for the different charge states observed from $Zn_8L2_4-64(NTf_2^{-})$.



Figure S6: gMS^2 of Zn_8L2_4 ·64(NTf_2^-) at m/z 1609.68 with different collision energies.



Figure S7: TEM imaging of discrete tessellated square array Zn₈L2₄.



Figure S8: TEM imaging of tubular-like nanostructures assembled by Zn₈L2₄.^{S4}

5. NMR spectra data of ligands and complexes



Figure S9:¹H NMR spectrum (400 MHz) of S2 in CDCl₃



Figure S10: 2D COSY spectrum (400 MHz) of S2 in CDCl₃.



Figure S11: ¹³C NMR spectrum (400 MHz) of S2 in CDCl₃.



Figure S13: ¹³C NMR spectrum (100 MHz) of S6 in CDCl₃.





Figure S14: ¹H NMR spectrum (400 MHz) of S8 in CDCl₃



Figure S15: 2D COSY spectrum (400 MHz) of S8 in CDCl₃.









Figure S17: ¹H NMR spectrum (400 MHz) of S9 in CDCl₃.



Figure S18: 2D COSY spectrum (400 MHz) of S9 in CDCl₃.



Figure S19: ¹³C NMR spectrum (100 MHz) of S9 in CDCl₃.



Figure S20: ¹H NMR spectrum (400 MHz) of S4 in CD₃OD.



Figure S21: 2D COSY spectrum (400 MHz) of S4 in CD₃OD.



Figure S22: 2D NOESY spectrum (400 MHz) of S4 in CD₃OD.



Figure S23: ¹H NMR spectrum (400 MHz) of M2 in CD₃OD.



Figure S24: 2D COSY spectrum (400 MHz) of M2 in CD₃OD.



Figure S25: 2D NOESY spectrum (400 MHz) of M2 in CD₃OD.



Figure S26: ¹H NMR spectrum (400 MHz) of M3 in CD₃OD.



Figure S27: 2D COSY spectrum (400 MHz) of M3 in CD₃OD.



Figure S28: 2D NOESY spectrum (400 MHz) of M3 in CD₃OD.



Figure S29: ¹H NMR spectrum (400 MHz) of H1 in CD₃CN.



Figure S30: 2D COSY spectrum (400 MHz) of H1 in CD₃CN.



Figure S31: 2D NOESY spectrum (400 MHz) of H1 in CD₃CN.



Figure S32: ¹H NMR spectrum (400 MHz) of H2 in CD₃CN.



Figure S33: 2D COSY spectrum (400 MHz) of H2 in CD₃CN.



Figure S34: 2D NOESY spectrum (400 MHz) of H3 in CD₃CN.



Figure S35: ¹H NMR spectrum (400 MHz) of L1 in CD₃CN.



Figure S36: 2D COSY spectrum (400 MHz) of L1 in CD₃CN.



Figure S37: 2D NOESY spectrum (400 MHz) of L1 in CD₃CN.



Figure S38: ¹H NMR spectrum (400 MHz) of L2 in CD₃CN.



Figure S39: 2D COSY spectrum (400 MHz) of L2 in CD₃CN.



Figure S40: 2D NOESY spectrum (400 MHz) of L2 in CD₃CN.



Figure S41: ¹H NMR spectrum (400 MHz) of Zn_8L2_4 in CD₃CN.



Figure S42: 2D COSY spectrum (400 MHz) of Zn₈L2₄ in CD₃CN.



Figure S43: 2D NOESY spectrum (400 MHz) of Zn₈L2₄ in CD₃CN.



Figure S44: Variable temperature ¹H NMR spectra (500 MHz) of Zn₈L2₄ in CD₃CN (from 298 K to 338 K).

6. ESI-MS spectra data of ligands and complexes



Figure S45: ESI-MS spectrum of S8.



Figure S46: ESI-MS spectrum of S9.



Figure S47: ESI-MS spectrum of S4.



Figure S48: ESI-MS spectrum of M2.



Figure S49: ESI-MS spectrum of M3.



Figure S50: Isotope patterns and ESI-MS spectrum of H1.



Figure S51: Isotope patterns and ESI-MS spectrum of L1.



Figure S52: Isotope patterns and ESI-MS spectrum of H2.



Figure S53: Isotope patterns and ESI-MS spectrum of L2.

7. Reference

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