# Pyridinium-conjugated polynorbornenes for nanomolar ATP

## Sensing using Indicator Displacement Assay and PET strategy

Ling-Xi Huang<sup>*a*,‡</sup>, Yue-Bo He<sup>*a*,‡</sup>, Jaewon Kim<sup>b</sup>, Amit Sharma<sup>c</sup>, Qian-Yong Cao<sup>*a*,\*</sup> Jong Seung Kim<sup>*b*,\*</sup>

<sup>a</sup> Department of Chemistry, Nanchang University, Nanchang 330031, P. R. China.

<sup>b</sup> Department of Chemistry, Korea University, Seoul 02841, Korea.

<sup>c</sup> CSIR-CSIO, Sector 30C, Chandigarh 160030, India.

<sup>‡</sup> These two authors contributed equally to this work.

\*Corresponding author. E-mail: <u>cqyong@ncu.edu.cn</u> (Q.-Y. Cao), jongskim@korea.ac.kr (JSK)

#### **Experimental Section:**

Materials and Instruments:

All chemicals and reagents for the reaction are analytical pure and used without further purification. The intermediates **2**, **PY2** and **PY3** were prepared by literature method [1, 2]. A Hitachi F-4500 spectrofluorophotometer was used for fluorescence measurements. The absorption spectra were measured on a Hitachi U-3900/3900H spectrophotometer. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded by a Varian instrument (400 MHz). The particle size of the nanoaggregations was recorded by a Zetasizer Nano-ZS90 instrument.



Scheme S1 Synthesis route of PNPY-n.

#### General procedure for Synthesis of NPY-n:

Equal amount (1.0 mmol) of compound 4 (300 mg, 1.16 mmol) and **PY-n** (n =1, 2, 3) were dissolved in acetonitrile (40 mL) under a nitrogen atmosphere. The stirred solution was heated and refluxed overnight. The solvent was evaporated in vacuo and the crude residue was purified by column chromatography (CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 95: 5, v/v) to obtain **NPY-n** as a white solid.

**NPY-1**: Yield 70%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 10.88 (s, 1H), 9.42 (s, 1H), 8.76 (d, J = 5.7 Hz, 1H), 8.33 (d, J = 8.9 Hz, 1H), 8.06 (d, J = 2.8 Hz, 1H), 6.11 (s, 1H), 5.82 (s, 1H), 4.73 – 4.61 (m, 2H), 4.09 (d, J = 5.8 Hz, 1H), 4.05 – 3.93 (m, 2H), 2.99 (s, 1H), 2.80 (s, 1H), 2.37 (d, J = 8.3 Hz, 2H), 2.21 (dq, J = 12.7, 6.4 Hz, 3H), 1.81 – 1.71 (m, 1H), 1.56 (s, 2H), 1.33 – 1.10 (m, 24H), 0.80 (d, J = 5.7 Hz, 3H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 146.09, 145.40, 138.10, 132.81, 128.51, 61.35, 58.97, 49.48, 45.45, 42.88, 42.33, 30.03, 29.11. TOF-HRMS (*m/z*): Calcd. for (C<sub>30</sub>H<sub>47</sub>BrN<sub>2</sub>O<sub>3</sub>): 483.3581 (M – Br<sup>–</sup>), found 483.3570 (M – Br<sup>–</sup>).

**NPY-2:** Yield 87%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 11.15 (s, 1H), 9.41 (s, 1H), 8.80 (d, J = 6.1 Hz, 1H), 8.43 (d, J = 8.6 Hz, 1H), 8.11 – 8.04 (m, 1H), 6.12 (s, 1H), 5.83 (s, 1H), 4.67 (t, J = 7.0 Hz, 2H), 4.00 (t, J = 6.2 Hz, 2H), 3.01 (s, 1H), 2.80 (s, 1H), 2.19 (t, J = 6.6 Hz, 2H), 2.17 – 2.11 (m, 3H), 1.77 (t, J = 10.3 Hz, 1H), 1.28 – 1.21 (m, 2H), 1.18 – 1.11 (m, 1H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 170.24,

139.78, 139.57, 138.09, 134.93, 134.19, 132.81, 128.67, 61.20, 59.42, 49.51, 45.47, 42.35, 30.19, 29.12. TOF-HRMS (*m/z*): Calcd. for (C<sub>18</sub>H<sub>23</sub>BrN<sub>2</sub>O<sub>3</sub>): 314.1703 (M – Br<sup>–</sup> – H<sup>+</sup>), found 314.1712.

**NPY-3:** Yield 88%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 9.08 (d, J = 6.0 Hz, 2H), 8.60 (t, J = 7.8 Hz, 1H), 8.16 (t, J = 6.9 Hz, 2H), 6.12 (dd, J = 5.7, 3.0 Hz, 1H), 5.83 (dd, J = 5.7, 2.8 Hz, 1H), 4.66 (t, J = 7.0 Hz, 2H), 4.01 (t, J = 6.0 Hz, 2H), 2.99 (s, 1H), 2.81 (s, 1H), 2.25 (d, J = 6.5 Hz, 1H), 1.77 (ddd, J = 12.4, 9.5, 3.6 Hz, 1H), 1.27 – 1.20 (m, 2H), 1.15 (dt, J = 11.8, 3.3 Hz, 1H). <sup>13</sup>C NMR (400 MHz, DMSO- *d*<sub>6</sub>)  $\delta$  ppm: 146.09, 145.40, 138.10, 132.81, 128.51, 61.35, 58.97, 49.48, 45.45, 42.88, 42.33, 30.0, 29.11. TOF-HRMS (*m/z*): Calcd. for (C<sub>16</sub>H<sub>20</sub>BrNO<sub>2</sub>): 257.1489 (M – Br<sup>–</sup> – H<sup>+</sup>), found 257.1487.

#### General procedure for Synthesis of PNPY-n:

Monomer NPY-n (0.5 mmol) and the Grubbs third-generation catalyst (1% of the amount of monomer substance) were dissolved in 5 mL of dry dichloromethane. Under nitrogen protection, the mixture was stirred at room temperature for 10 minutes, and then 1 mL of ethyl-vinyl ether was added to terminate the polymerization. It was then precipitated by crystallization in ether and concentrated to get the off-white solid **PNPY-n**.

**PNPY-1**: Yield 72%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 11.01 (s, 1H), 9.47 (s, 1H), 8.81 (s, 1H), 8.34 (s, 1H), 8.09 (s, 1H), 4.67 (s, 2H), 3.93 (s, 1H), 3.31 (d, J = 2.8 Hz, 6H), 3.08 (s, 1H), 2.76 (s, 2H), 2.19 (s, 2H), 1.82 (s, 2H), 1.53 (s, 3H), 1.16 (s, 14H), 0.78 (s, 2H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 173.05, 139.55, 128.77, 36.50, 31.74, 29.51, 29.19, 29.00, 25.05, 22.54, 14.37. GPC (THF, polystyrene standards): M<sub>n</sub> = 33240, M<sub>w</sub> = 34902, and PDI = 1.05.

PNPY-2: Yield 70%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 11.38 (s, 1H), 9.44 (s, 1H), 8.85 (s, 1H), 8.49 (s, 1H), 8.09 (s, 1H), 4.69 (s, 2H), 4.00 (d, J = 40.9 Hz, 2H), 3.08 (s, 1H), 2.78 (s, 1H), 2.17 (d, J = 21.2 Hz, 3H), 1.87 (s, 1H), 1.50 (s, 1H), 1.20 (s, 1H), 1.20

1H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ ppm: 170.25, 139.63, 134.72, 134.11, 129.85, 128.66, 30.22, 24.27. GPC (THF, polystyrene standards): Mn = 33199, Mw = 33858, and PDI = 1.05.

**PNPY-3**: Yield 70%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 9.13 (s, 2H), 8.62 (s, 1H), 8.17 (s, 2H), 4.69 (s, 2H), 4.06 (s, 1H), 3.93 (s, 1H), 3.11 (s, 1H), 2.78 (s, 2H), 2.24 (s, 2H), 1.84 (s, 2H), 1.49 (s, 1H), 1.15 (s, 1H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  ppm: 146.14, 145.27, 128.46, 61.07, 58.67, 30.08. GPC (THF, polystyrene standards): M<sub>n</sub> = 33068, M<sub>w</sub> = 34721, and PDI = 1.05.



Fig. S1 (a) Fluorescence intensity of UD at 515 nm versus the number of equivalents of PNPY-1.(b) The binding constant of PNPY-1/UD using the Benesi-Hildebrand method.



**Fig. S2** (a) Fluorescence spectra and (b) relative fluorescence quenching ratios (%) of UD (6  $\mu$ M,  $\lambda_{ex} = 490$  nm) at 515 nm upon titration 2 equivalents of **PNPY-1** storage different time in HEPES (10 mM, pH = 7.4) buffer solution.



Fig. S3 The size distribution of (a) PNPY-2; (b) PNPY-2/UD; (c) PNPY-3; (d) PNPY-3/UD.



Fig. S4 (a) UV-vis spectra and (b) size distribution of PNPY-1/UD (12  $\mu$ M/6  $\mu$ M) with storage different time in a HEPES (10 mM, pH = 7.4) buffered solution.



Fig. S5 (a) UV-vis spectra of PNPY-1/UD (12  $\mu$ M/6  $\mu$ M,  $\lambda_{ex} = 490$  nm) upon addition of various amounts of ATP in a HEPES (10 mM, pH = 7.4) buffered solution. (b) UV titration curve of PNPY-1/UD at 490 nm versus the equiv of ATP.



**Fig. S6** The time course of fluorescence spectra of (a) **PNPY-1/UD** (6  $\mu$ M,  $\lambda_{ex} = 490$  nm) and the fluorescence intensity of **UD** at 515 nm (b) **PNPY-1/UD** (6  $\mu$ M,  $\lambda_{ex} = 490$  nm) upon addition of 20 equiv of ATP in HEPES (10 mM, pH = 7.4) buffer solution.



Fig. S7 The fluorescence (a) and UV-vis (c) spectra of PNPY-1/UD (6  $\mu$ M,  $\lambda_{ex} = 490$  nm) upon addition of various amounts of ADP in HEPES (10 mM, pH = 7.4) buffer solution. The binding constant of PNPY-1/ADP using the Benesi-Hildebrand method (b) and UV absorbance of PNPY-1/UD at 490 nm (d) versus the equiv of ADP.



Fig. S8 The fluorescence (a) and UV-vis (c) spectra of PNPY-1/UD (6  $\mu$ M,  $\lambda_{ex} = 490$  nm) upon addition of various amounts of PPi in HEPES (10 mM, pH = 7.4) buffer solution. The binding constant of PNPY-1/PPi using the Benesi-Hildebrand method (b) and UV absorbance of PNPY-1/UD at 490 nm (d) versus the equiv of PPi.



Fig. S9 Fluorescence intensity of (a) PNPY-2/UD (36  $\mu$ M/6  $\mu$ M,  $\lambda_{ex} = 490$  nm) and (b) PYNP-3/UD (120  $\mu$ M/6  $\mu$ M,  $\lambda_{ex} = 490$  nm) at 515 nm versus the equiv of anions.



Fig. S10 Fluorescence spectra of UD (6  $\mu$ M,  $\lambda_{ex}$  = 490 nm) upon the addition of various amounts of NPY-1 (a), NPY-2 (b) and NPY-3 (c) in a HEPES (10 mM, pH = 7.4) buffered solution. The inset shows the fluorescence intensity of UD at 515 nm versus the equiv of NPY-1, NPY-2 or NPY-3.

![](_page_7_Figure_4.jpeg)

**Fig. S11** (a) Fluorescence spectra of **NPY-1/UD** (66  $\mu$ M/6  $\mu$ M,  $\lambda_{ex} = 490$  nm) upon the addition of various amounts of ATP in a HEPES (10 mM, pH = 7.4) buffered solution. (b) Fluorescence intensity of **NPY-1/UD** at 515 nm versus the number of equiv of ATP.

![](_page_8_Figure_0.jpeg)

Fig. S12 (a) Competitive fluorescence experiment of probe PNPY-1/UD toward ATP in the presence of various anions in HEPES (10 mM, pH = 7.4) buffer solutions. 1-ADP, 2-PPi, 3-AMP, 4-H<sub>2</sub>PO<sup>4-</sup>, 5-AcO<sup>-</sup>, 6-F<sup>-</sup>, 7-Cl<sup>-</sup>, 8-Br<sup>-</sup>, 9-I<sup>-</sup>, 10-CO<sub>3</sub><sup>2-</sup>, 11-SO<sub>4</sub><sup>2-</sup>. (b) The relative absorption loss (%) of UD (6  $\mu$ M, record at 490 nm) and PNPY-1/UD (12  $\mu$ M/6  $\mu$ M, record at 517 nm) with exposure time at xenon lamp (300 W) irritation.

![](_page_8_Figure_2.jpeg)

Fig. S13 Fluorescence intensity at 515 nm of UD (6  $\mu$ M), PNPY-1/UD (12  $\mu$ M/6  $\mu$ M) and PNPY-1/UD + ATP (10 equivalents) at different pH values.

![](_page_8_Figure_4.jpeg)

Fig. S14 The relative fluorescence loss of UD (12  $\mu$ M/6  $\mu$ M) and PNPY-1/UD (12  $\mu$ M/6  $\mu$ M) with increasing the exposure time of xenon lamp (300 W) at 517 nm.

![](_page_9_Figure_0.jpeg)

**Fig. S15** Cell viability after incubation of Hepa1-6 murine liver cancer cells with various concentrations of **PNPY-1/UD** in aqueous solutions.

![](_page_9_Picture_2.jpeg)

Fig. S16 Laser scanning confocal images of Hepa1-6 cells co-stained with PNPY-1/UD (12  $\mu$ M/6  $\mu$ M) and CytoFixRed (10  $\mu$ M) for 30 min. (a) Ex. 488 nm, Em. 500–570 nm, (b) Ex. 488 nm, Em. 650–710 nm. (c) Merged image of (a) and (b). Scale bar, 20  $\mu$ m. (d) Correlation plot of PNPY-1/UD and CytoFixRed intensities (Pearson Correlation Coefficient: 0.82).

![](_page_10_Figure_0.jpeg)

**Fig. S18**<sup>13</sup>C NMR spectrum of **NPY-1** in a DMSO-*d*<sub>6</sub> solution.

![](_page_11_Figure_0.jpeg)

![](_page_11_Figure_1.jpeg)

![](_page_12_Figure_0.jpeg)

Fig. S22 <sup>13</sup>C NMR spectrum of NPY-3 in a DMSO-*d*<sub>6</sub> solution.

![](_page_13_Figure_0.jpeg)

![](_page_13_Figure_1.jpeg)

![](_page_13_Figure_2.jpeg)

Fig. S24 The HR-MS spectrum of NPY-2

![](_page_14_Figure_0.jpeg)

Fig. S25 The HR-MS spectrum of NPY-3

0

![](_page_14_Figure_2.jpeg)

Fig. S26 <sup>1</sup>H NMR spectrum of PNPY-1 in a DMSO-*d*<sub>6</sub> solution.

![](_page_15_Figure_0.jpeg)

![](_page_15_Figure_1.jpeg)

![](_page_15_Figure_2.jpeg)

![](_page_15_Figure_3.jpeg)

![](_page_16_Figure_0.jpeg)

Fig. S29 The GPC curve of (a) PNPY-1; (b) PNPY-2; (c) PNPY-3 in THF, polystyrene standard.

### References

- [1] S. Brahmachari, S. Debnath, S. Dutta, P. K. Das, *Beilstein J. Org. Chem.*, 2010, 6, 859–868.
- [2] S. K. Yang, M. Weck, *Macromolecules*, 2008, 41, 346–351.