Preparation of enantioenriched helical- and axial-chiral molecules

by dynamic asymmetric induction

Yuuya Kawasaki,^a Ryota Kamikubo,^b Yuta Kumegawa,^b Kouhei Ogawa,^b

Takeru Kashiwagi,^a Yusuke Ano,^a Kazunobu Igawa^{a,b} and Katsuhiko Tomooka^{*,a,b}

^{*a}Institute for Materials Chemistry and Engineering, and IRCCS, Kyushu University, 6-1 Kasuga-koen, Kasuga, Fukuoka 816-8580, Japan*</sup>

^bDepartment of Molecular and Material Sciences, Kyushu University, 6-1 Kasuga-koen, Kasuga, Fukuoka 816-8580, Japan

E-mail: ktomooka@cm.kyushu-u.ac.jp

Table of Contents:

1.	General Experimental	S 3
2.	Detailed Reaction Procedures and Analytical Data	S4–S19
3.	Determination of the Absolute Stereochemistry	S20
4.	¹ H NMR Analysis	
	for Tautomerization of Lactam $3d$ and Lactim $4d$ in CDCl ₃ at 25 °C	S21
5.	Kinetic Measurements of the Racemization of Lactam 3 and Lactim 4	S22
6.	General Procedure of DYASIN	S23
7.	HPLC Chromatograms for Determination of Enantiomeric Ratio	
	Measurement for DYASIN of Heterohelicenes	S24–S27
	Measurement for Conversion of Heterohelicenes to Binaphthyl Compounds	S28
8.	Computational Results	S29–S31
9.	¹ H, ¹³ C NMR Spectra	S32–S46
10.	References and Notes	S47

1. General Experimental:

All reactions were carried out in heat-gun-dried glassware under an argon atmosphere unless otherwise noted. Dry tetrahydrofuran (THF), 1,4-dioxane, CH₂Cl₂, CH₃CN, MeOH, EtOH, N,Ndimethylformamide (DMF), N.N-dimethylacetamide (DMA), and AcOEt were purchased from Kanto Chemical Co., Inc. and used without further purification. OCS1,¹ OCS2,² OCS3,³ OCS4,⁴ and OCS5⁵ were provided by Daicel corporation. ¹H NMR spectra were recorded on Varian Mercury spectrometer (300 MHz) or JEOL JNM-ECA600 (600 MHz) and ¹³C NMR spectra were recorded on Varian Mercury spectrometer (75 MHz) or JEOL JNM-ECA600 (151 MHz) at ambient temperature using $CDCl_3$ as a solvent: $CHCl_3$ (¹H, δ 7.26), $CDCl_3$ (¹³C, δ 77.1) were used as internal references. The peak multiplicities were given as followed: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. Specific rotations were measured on a digital polarimeter (JASCO DIP-370). Circular dichroism (CD) spectra were measured on a spectrophotometer (JASCO J-720W). Infrared spectra (IR) were recorded on a Fourier transfer infrared spectrophotometer (Perkin Elmer SpectrumOne) as neat liquid on NaCl plates and as crystals used a diffuse reflector. High performance liquid chromatography (HPLC) were performed on circular dichroism chiral detector (JASCO CD-2095 or CD-4095), photodiode array detector (JASCO MD-2018 or MD-4010) and column oven (JASCO CO-2067 or CO-4060) equipped with a JASCO PU-2089 using Daicel CHIRALCEL OD-3 column (0.46 cm × 5.0 cm), CHIRALPAK AD-3 column (0.46 cm × 5.0 cm), CHIRALPAK AS-H column (0.46 cm × 25 cm), CHIRALPAK AS-3 column (0.46 cm × 5.0 cm), CHIRALPAK IB column (0.46 cm × 25 cm), and CHIRALPAK IE $(0.46 \text{ cm} \times 25 \text{ cm})$. Analytical thin-layer chromatography (TLC) was carried out on silica gel 60 F₂₅₄ (Merck 5715) plates and developed plates were visualized by UV (254 nm) and by heating on a hot plate after staining with a 4% solution of phosphomolybdic acid in ethanol or a 2.5% solution of panisaldehyde in ethanol. Column chromatography were performed using Kanto spherical silica gel 60N (neutral, 63-210 μm) or Fuji Silysia silica gel FL100D (neutral, 100 μm). X-ray crystallographic data were collected using a Rigaku AFC HyPix-6000 using MoK α radiation ($\lambda = 0.71073$ Å) to a maximum 2θ range for data collection 4.812° to 60.972°. HRMS analyses were recorded on a JEOL JMS-700 at the Analytical Center in IMCE, Kyushu University.

Chemicals Unless otherwise noted, all reagents were purchased and used as delivered.

2. Detailed Reaction Procedures and Analytical Data:

lactone 2a



The compound **2a** was prepared according to the literature procedure with slight modification.⁶

To a solution of **s1** (377 mg, 1.00 mmol) in DMA (20 mL) was added PPh₃ (262 mg, 1.00 mmol), NaOAc (328 mg, 4.00 mmol) and Pd(OAc)₂ (89.8 mg, 0.400 mmol) at ambient temperature, then the mixture was degassed (3 times of freeze-pump-thaw). After stirred at 120 °C for 24 h, the mixture was cooled to ambient temperature and filtered through a pad of silica gel using AcOEt as eluent. The filtrate was concentrated under reduced pressure and the residue was purified by silica gel chromatography (n-C₆H₁₄/AcOEt = 4:1) to afford a mixture of **2a** and its regioisomer. The mixture was suspended to n-C₆H₁₄/*i*-PrOH = 1:1 and the filtration afforded 160 mg (54%) of **2a** as colorless crystals. The spectroscopic data were in good agreement with the reported values.⁶

Analytical HPLC [column: CHIRALPAK AS-H (0.46 cm × 25 cm), eluent: n-C₆H₁₄/i-PrOH = 90:10, flow rate: 1.0 mL/min, detection: UV 254 nm, temperature: 10 °C]: $t_1 = 10.5$ min for (*M*)-isomer, $t_2 = 13.3$ min for (*P*)-isomer.

Analytical HPLC [column: CHIRALPAK AS-3 (0.46 cm × 5.0 cm), eluent: n-C₆H₁₄/EtOH = 90:10, flow rate: 0.5 mL/min, detection: UV 254 nm, temperature: 10 °C]: $t_1 = 5.1$ min for (*M*)-isomer, $t_2 = 7.4$ min for (*P*)-isomer.

naphthoic acid s4

To a solution of 3-hydroxy-2-naphthoic acid (10.0 g, 53.1 mmol) in MeOH (250 mL) was added conc. H₂SO₄ (25 mL) at 0 °C. After stirred under reflux for 24 h, the mixture was cooled to ambient temperature. The mixture was poured into 28-30 % aq. NH₃ at 0 °C, and then the aqueous phase was extracted with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure to afford crude s2. To a solution of the crude s2 in DMF (25 mL) was added K₂CO₃ (10.1 g, 73.1 mmol) and BnBr (9.30 ml, 77.8 mmol) at ambient temperature. After stirred at 80 °C for 20 h, the mixture was cooled to ambient temperature. H₂O was added to the mixture, and then the aqueous phase was extracted with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure to afford crude product s_3 . To a solution of the crude s_3 in MeOH (200 mL) / H₂O (80 mL) was added NaOH (4.24 g, 106 mmol) at ambient temperature. The mixture was stirred at 40 °C for 12 h, and then cooled to ambient temperature. The mixture was washed with Et₂O, and the aqueous phase was acidified with 35-37% aq. HCl, and extracted with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure to afford 14.8 g (up to 99% in 3 steps) of s4 as a yellow solid. The analytical data of the naphthoic acid s4 were collected after purification by recrystallization from CHCl₃/n-C₆H₁₄ providing colorless crystals.

¹H NMR (300 MHz, CDCl₃): δ 11.0 (br, 1H), 8.82 (s, 1H), 7.92 (d, *J* = 8.4 Hz, 1H), 7.77 (d, *J* = 8.1 Hz, 1H), 7.60 (ddd, *J* = 8.4, 7.5, 1.2 Hz, 1H), 7.52-7.41 (m, 7H), 5.39 (s, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 165.3, 153.6, 136.6, 136.5, 134.4, 129.6, 129.4, 129.3, 128.6, 128.2,

126.7, 125.7, 108.7, 72.4 (two aromatic carbon's peaks are overlapping).

IR (reflection, cm⁻¹): 3228, 3058, 1953, 1727, 1632, 1502, 980, 710.

HRMS (EI, positive): Exact mass calcd. for $C_{18}H_{14}O_3$ [M]⁺ requires *m/z*: 278.0943, found *m/z*: 278.0946.

Mp: 134.7-135.0 °C

ester s5



To a solution of s4 (1.29 g, 4.64 mmol) in CH₂Cl₂ (20 mL) was added oxalyl chloride (519 μ L, 6.05 mmol) and 1-methyl-2-pyrrolidone (NMP) (1 mL) at 0 °C. After stirred at ambient temperature for 1 h, Et₃N (2.70 mL, 19.4 mmol) and 1-bromo-2-naphthol (864 mg, 3.87 mmol) were added to the mixture. After stirred at that temperature for 14 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/CH₂Cl₂ = 3:2) to afford 1.71 g (76%) of s5 as colorless crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.78 (s, 1H), 8.32 (d, *J* = 8.4 Hz, 1H), 7.93 (d, *J* = 8.1 Hz, 1H), 7.90-7.87 (m, 2H), 7.76 (d, *J* = 8.1 Hz, 1H), 7.67-7.51 (m, 5H), 7.47-7.29 (m, 6H), 5.35 (s, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 163.5, 155.2, 146.8, 136.7, 134.3, 132.9, 132.6, 129.1, 129.03, 128.95, 128.7, 128.5, 128.3, 127.9, 127.8, 127.1, 126.7, 126.4, 124.8, 122.4, 120.7, 115.5, 108.8, 70.7. (two aromatic carbon's peaks are overlapping)

IR (reflection, cm⁻¹): 3057, 2889, 2349, 1741, 1627, 1127, 1047, 822.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{19}BrO_3$ [M]⁺ requires m/z: 482.0518, found m/z: 482.0519.

Mp: 139.7-140.1 °C

lactone s6



To a solution of **s5** (483 mg, 1.00 mmol) in 1,4-dioxane (20 mL) was added PPh₃ (118 mg, 0.450 mmol), dried K₂CO₃ (415 mg, 3.00 mmol), pivalic acid (30.6 mg, 0.300 mmol) and Pd(OAc)₂ (33.7 mg, 0.150 mmol) at ambient temperature, then the mixture was degassed (3 times of freeze-pump-thaw). After stirred at 100 °C for 20 h, the mixture was cooled to ambient temperature and filtered through a pad of silica gel using AcOEt as eluent. The filtrate was concentrated under reduced pressure and the residue was purified by silica gel chromatography (n-C₆H₁₄/AcOEt = 15:1) to afford 281 mg (70%) of **s6** as yellow crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.01 (d, *J* = 9.3 Hz, 1H), 7.91 (dd, *J* = 8.4, 8.4 Hz, 2H), 7.79 (dd, *J* = 9.6, 9.6 Hz, 2H), 7.70 (d, *J* = 7.5 Hz, 2H), 7.59-7.55 (m, 2H), 7.50-7.41 (m, 4H), 7.36-7.31 (m, 2H), 7.23-7.17 (m, 1H), 5.50 (d, *J* = 13.8 Hz, 1H), 5.44 (d, *J* = 13.8 Hz, 1H).

¹³C NMR (75 MHz, CDCl₃): δ 158.2, 155.5, 150.6, 138.6, 137.6, 136.7, 132.1, 131.0, 130.2, 129.8, 129.3, 128.8, 128.5, 127.9, 127.10, 127.07, 126.9, 126.0, 125.3, 124.2, 123.4, 117.0, 114.3, 112.7, 109.7, 71.2.

IR (reflection, cm⁻¹): 3459, 3070, 2941, 1740, 1552, 1291, 1018, 821.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{18}O_3$ [M]⁺ requires *m/z*: 402.1256, found *m/z*: 402.1257.

Mp: 208.7-209.2 °C

lactone 2ba



To a solution of **s6** (140 mg, 0.348 mmol) in AcOEt (20 mL) was added 10% palladium on active charcoal (370 mg, 0.348 mmol) at ambient temperature. The mixture was stirred for 14 h under atmospheric pressure of hydrogen at that temperature. The mixture was filtered through a pad of Celite using AcOEt as eluent, and the filtrate was concentrated under reduced pressure to afford **s7**. To a solution of crude **s7** in CH₂Cl₂ (5 mL) was added Hünig base (566 μ L, 3.28 mmol) and 2-methoxyethoxymethyl chloride (MEMCl) (374 μ L, 3.28 mmol) at ambient temperature. After stirred at that temperature for 20 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (toluene/AcOEt = 15:1) to afford 131 mg (94% in 2 steps) of **2ba** as yellow crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.00 (d, J = 9.0 Hz, 1H), 7.93-7.85 (m, 3H), 7.77-7.74 (m, 2H), 7.62-7.57 (m, 1H), 7.56 (d, J = 9.0 Hz, 1H), 7.50-7.44 (m, 1H), 7.35-7.30 (m, 1H), 7.26-7.20 (m, 1H), 5.63 (d, J = 6.9 Hz, 1H), 5.60 (d, J = 6.9 Hz, 1H), 4.05-4.01 (m, 2H), 3.65-3.62 (m, 2H), 3.40 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 158.2, 154.0, 150.3, 138.3, 137.5, 132.1, 131.0, 130.1, 129.3, 129.1, 128.5, 127.4, 127.0, 126.0, 125.3, 124.7, 123.8, 116.9, 114.4, 113.0, 112.7, 94.8, 71.6, 68.3, 59.1. IR (reflection, cm⁻¹): 2923, 1736, 1621, 1331, 1209, 1009, 752.

HRMS (EI, positive): Exact mass calcd. for $C_{25}H_{20}O_5$ [M]⁺ requires m/z: 400.1311, found m/z: 400.1310.

Mp: 97.7-98.3 °C

Analytical HPLC [column: CHIRALPAK AD-3 (0.46 cm × 5.0 cm), eluent: n-C₆H₁₄/*i*-PrOH = 80:20, flow rate: 1.0 mL/min, detection: UV 240 nm, temperature: 10 °C]: t_1 = 3.0 min for (*P*)-isomer, t_2 = 5.8 min for (*M*)-isomer.

lactone 2bb



To a solution of lactone **s6** (50.0 mg, 0.124 mmol) in AcOEt (6 mL) was added 10% palladium on active charcoal (132 mg, 0.124 mmol) at ambient temperature. The mixture was stirred for 1 h under atmospheric pressure of hydrogen at that temperature. The mixture was filtered through a pad of Celite using AcOEt as eluent, and the filtrate was concentrated under reduced pressure to afford **s7**. To a solution of crude **s7** in CH₂Cl₂ (10 mL) was added Hünig base (210 μ L, 1.21 mmol) and 2-(trimethylsilyl)ethoxymethyl chloride (SEMCl) (110 μ L, 0.622 mmol) at ambient temperature. After stirred at that temperature for 5 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/AcOEt = 9:1) to afford 43.7 mg (80% in 2 steps) of lactone **2bb** as a yellow amorphous.

¹H NMR (300 MHz, CDCl₃): δ 8.00 (d, *J* = 9.0 Hz, 1H), 7.93-7.84 (m, 3H), 7.78-7.72 (m, 2H), 7.62-7.55 (m, 2H), 7.47 (ddd, *J* = 6.9, 6.9, 1.2 Hz, 1H), 7.32 (ddd, *J* = 8.7, 8.7, 1.5 Hz, 1H), 7.22 (ddd, *J* = 8.4, 8.4, 1.5 Hz, 1H), 5.59 (d, *J* = 6.9 Hz, 1H), 5.56 (d, *J* = 6.9 Hz, 1H), 4.03-3.88 (m, 2H), 1.07-1.01 (m, 2H), 0.021 (s, 9H).

¹³C NMR (75 MHz, CDCl₃): δ 158.2, 154.3, 150.4, 138.3, 137.6, 132.0, 131.0, 130.2, 129.7, 129.2, 128.5, 127.4, 127.0, 126.0, 125.3, 124.6, 123.7, 116.9, 114.4, 112.8, 94.4, 67.1, 18.3, -1.3. (one aromatic carbon's peak is overlapping).

IR (reflection, cm⁻¹): 2923, 1736, 1620, 1554, 1332, 1009, 751.

HRMS (EI, positive): Exact mass calcd. for $C_{27}H_{26}O_4Si$ [M]⁺ requires m/z: 442.1600, found m/z: 442.1600.

Analytical HPLC [column: CHIRALCEL OD-3 (0.46 cm × 5.0 cm), eluent: n-C₆H₁₄/i-PrOH = 50:50, flow rate: 0.5 mL/min, detection: UV 240 nm, temperature: 15 °C]: $t_1 = 4.1$ min for (*M*)-isomer, $t_2 = 21.8$ min for (*P*)-isomer.

naphthol s8



To a solution of 2,3-dihydroxynaphthalene (4.81 g, 30.0 mmol) in DMF (30 mL) was added K₂CO₃ (4.15 g, 30.0 mmol) at ambient temperature. The mixture was stirred at 50 °C for 30 min then BnCl (3.45 ml, 30.0 mmol) was added to the mixture. After stirred at 80 °C for 12 h, the reaction was quenched with H₂O and extracted with Et₂O. The combined organic phase was poured into 10 M aq. NaOH. The desired product **s8** and 2,3-dihydroxynaphthalene were extracted with 10 M aq. NaOH as a sodium salt. The combined aq. NaOH phase was acidified with 35-37% aq. HCl, and then the aqueous phase was extract with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/AcOEt = 15:1) to afford 4.40 g (59%) of **s8** as colorless crystals.

¹H NMR (300 MHz, CDCl₃): δ 7.69-7.63 (m, 2H), 7.54-7.30 (m, 8H), 7.22 (s, 1H), 5.98 (s, 1H), 5.24 (s, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 146.6, 145.8, 136.0, 129.8, 129.0, 128.9, 128.6, 128.0, 126.6, 126.4, 124.5, 124.0, 109.7, 107.1, 71.1.

IR (reflection, cm⁻¹): 3533, 3061, 2883, 1961, 1693, 1638, 1590, 1454, 1162, 942.

HRMS (EI, positive): Exact mass calcd. for $C_{17}H_{14}O_2$ [M]⁺ requires *m/z*: 250.0994, found *m/z*: 250.0994.

Mp: 91.3-92.2 °C

ester s9



To a solution of 1-bromo-2-naphthoic acid (500 mg, 1.99 mmol) in CH_2Cl_2 (10 mL) was added oxalyl chloride (267 µL, 3.11 mmol) and 1-methyl-2-pyrrolidone (1 mL) at 0 °C. The mixture was stirred at ambient temperature for 1 h then Et_3N (1.39 mL, 9.97 mmol) and **s8** (498 mg, 1.99 mmol) were added to the mixture. After stirred at that temperature for 14 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/CH₂Cl₂ = 3:2) to afford 894 mg (93%) of **s9** as colorless crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.50 (d, *J* = 8.1 Hz, 1H), 7.88-7.76 (m, 6H), 7.71-7.61 (m, 2H), 7.49-7.31 (m, 8H), 5.27 (s, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 165.6, 149.4, 140.6, 136.4, 135.5, 132.8, 132.4, 130.6, 128.8, 128.6, 128.4, 128.3, 128.22 128.15, 127.83, 127.75, 127.5, 126.7, 126.4, 126.3, 124.6, 123.4, 120.7, 108.8, 70.8. (one aromatic carbon's peak is overlapping)

IR (reflection, cm⁻¹): 3065, 2942, 1943, 1755, 1361, 1228, 991, 752.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{19}BrO_3$ [M]⁺ requires m/z: 482.0518, found m/z: 482.0523.

Mp: 127.7-128.4 °C

lactone s10



To a solution of **s9** (483 mg, 1.00 mmol) in DMA (20 mL) was added PPh₃ (262 mg, 1.00 mmol), dried NaOAc (328 mg, 4.00 mmol) and Pd(OAc)₂ (89.8 mg, 0.400 mmol) at ambient temperature, then the mixture was degassed (3 times of freeze-pump-thaw). After stirred at 120 °C for 24 h, the mixture was cooled to ambient temperature and filtered through a pad of silica gel using AcOEt as eluent. The filtrate was concentrated under reduced pressure and the residue was purified by silica gel chromatography (n-C₆H₁₄/toluene = 1:4) to afford 257 mg (64%) of **s10** as yellow crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.39 (d, *J* = 8.7 Hz, 1H), 8.12 (d, *J* = 8.7 Hz, 1H), 8.06 (d, *J* = 8.4 Hz, 1H), 8.01 (d, *J* = 8.4 Hz, 1H), 7.81 (d, *J* = 8.7 Hz, 1H), 7.78 (d, *J* = 9.0 Hz, 1H), 7.68 (ddd, *J* = 6.9, 6.9, 0.9 Hz, 1H), 7.59 (d, *J* = 7.5 Hz, 2H), 7.47-7.35 (m, 6H), 7.22 (ddd, *J* = 8.1, 6.6, 1.2 Hz, 1H), 5.42 (s, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 161.4, 146.3, 142.9, 136.7, 136.4, 134.9, 131.0, 129.7, 129.3, 129.1, 128.8, 128.6, 128.4, 128.2, 127.4, 127.3, 127.1, 125.8, 125.5, 124.9, 124.2, 123.5, 121.7, 114.3, 111.2, 71.1.

IR (reflection, cm⁻¹): 3059, 1729, 1592, 1470, 1065, 802, 759, 694.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{18}O_3$ [M]⁺ requires *m/z*: 402.1256, found *m/z*: 402.1255.

Mp: 84.0-84.6 °C

lactone 2ca



To a solution of **s10** (136 mg, 0.338 mmol) in AcOEt (20 mL) was added 10% palladium on active charcoal (353 mg, 0.332 mmol) at ambient temperature. The mixture was stirred for 3 h under atmospheric pressure of hydrogen at that temperature. The mixture was filtered through a pad of Celite using AcOEt as eluent, and the filtrate was concentrated under reduced pressure to afford **s11**. To a solution of crude **s11** in CH₂Cl₂ (5 mL) was added Hünig base (290 μ L, 1.66 mmol) and MEMCl (190 μ L, 1.66 mmol) at ambient temperature. After stirred at that temperature for 4 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (toluene/AcOEt = 15:1) to afford 114 mg (84% in 2 steps) of **2ca** as yellow crystals.

¹H NMR (300 MHz, CDCl₃): δ 8.37 (d, *J* = 8.4 Hz, 1H), 8.11 (d, *J* = 8.7 Hz, 1H), 8.06 (d, *J* = 8.4 Hz, 1H), 8.00 (d, *J* = 8.1 Hz, 1H), 7.86-7.80 (m, 3H), 7.68 (ddd, *J* = 7.8, 7.8, 0.9 Hz, 1H), 7.49-7.38 (m, 2H), 7.27-7.22 (m, 1H), 5.61 (d, *J* = 7.2 Hz, 1H), 5.57 (d, *J* = 7.2 Hz, 1H), 4.03-4.00 (m, 2H), 3.65-3.62 (m, 2H), 3.40 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 161.4, 144.8, 143.0, 136.8, 135.0, 131.1, 129.7, 129.3, 129.2, 128.7 128.4, 127.7, 127.0, 125.9, 125.6, 125.5, 124.2, 124.0, 121.6, 114.4, 114.2, 94.7, 71.6, 68.3, 59.1.

IR (reflection, cm⁻¹): 3449, 2922, 1730, 1592, 1404, 1338, 1055, 920, 793, 759.

HRMS (EI, positive): Exact mass calcd. for $C_{25}H_{20}O_5$ [M]⁺ requires m/z: 400.1311, found m/z: 400.1312.

Mp: 50.3-50.9 °C

Analytical HPLC [column: CHIRALPAK AD-3 (0.46 cm × 5.0 cm), eluent: n-C₆H₁₄/i-PrOH = 80:20, flow rate: 1.0 mL/min, detection: UV 240 nm, temperature: 15 °C]: t_1 = 2.9 min for (*P*)-isomer, t_2 = 6.4 min for (*M*)-isomer.

lactone 2cb



To a solution of **s10** (50.0 mg, 0.124 mmol) in AcOEt (6 mL) was added 10% palladium on active charcoal (132 mg, 0.124 mmol) at ambient temperature. The mixture was stirred for 5 h under atmospheric pressure of hydrogen at that temperature. The mixture was filtered through a pad of Celite using AcOEt as eluent, and the filtrate was concentrated under reduced pressure to afford **s11**. To a solution of crude **s11** in CH₂Cl₂ (5 mL) was added Hünig base (128 μ L, 0.735 mmol) and SEMCl (66.0 μ L, 0.373 mmol) at ambient temperature. After stirred at that temperature for 17 h, the solvent was removed under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/AcOEt = 9:1) to afford 50.5 mg (92% in 2 steps) of **2cb** as a yellow amorphous.

¹H NMR (300 MHz, CDCl₃): δ 8.38 (d, *J* = 8.7 Hz, 1H), 8.13 (d, *J* = 8.7 Hz, 1H), 8.06 (d, *J* = 8.4 Hz, 1H), 8.01 (d, *J* = 8.4 Hz, 1H), 7.85 (d, *J* = 8.4 Hz, 1H), 7.82 (d, *J* = 8.7 Hz, 1H), 7.79 (d, *J* = 9.6 Hz, 1H), 7.68 (ddd, *J* = 7.2, 7.2, 1.1 Hz, 1H), 7.49-7.30 (m, 2H), 7.27-7.20 (m, 1H), 5.57 (d, *J* = 7.2 Hz, 1H), 5.53 (d, *J* = 7.2 Hz, 1H), 3.96-3.87 (m, 2H), 1.07-1.01 (m, 2H), 0.02 (s, 9H).

¹³C NMR (75 MHz, CDCl₃): δ 161.3, 144.9, 142.8, 136.6, 134.8, 131.0, 129.5, 129.2, 129.1, 128.5, 128.3, 127.5, 126.9, 125.7, 125.4, 125.2, 124.0, 123.7, 121.5, 114.1, 113.7, 94.4, 66.9, 18.2, -1.4. IR (reflection, cm⁻¹): 2953, 1735, 1592, 1248, 1056, 1001, 834.

HRMS (EI, positive): Exact mass calcd. for $C_{27}H_{26}O_4Si$ [M]⁺ requires m/z: 442.1600, found m/z: 442.1600.

Analytical HPLC [column: CHIRALCEL OD-3 (0.46 cm × 5.0 cm), eluent: n-C₆H₁₄/EtOH = 90:10, flow rate: 0.5 mL/min, detection: UV 254 nm, temperature: 10 °C]: t_1 = 4.5 min for (*M*)-isomer, t_2 = 30.2 min for (*P*)-isomer.

lactam 3d



The compound **3a** was prepared according to the literature procedure.⁷

To a solution of **3a** (148 mg, 0.500 mmol) in THF (5 mL) was added MeMgBr (1.07 M in THF, 514 μ L, 0.550 mmol) at 0 °C. The mixture was stirred at that temperature for 30 min, and benzoyl chloride (63.9 μ L, 0.550 mmol) was added to the mixture. After stirred at that temperature for 48 h, the reaction was quenched with sat. aq. NH₄Cl and extracted with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/CHCl₃/Et₂O = 8:20:1) to afford 206 mg (79%) of **3d** as a yellow crystal.

¹H NMR (600 MHz, CDCl₃): δ 8.47 (d, *J* = 7.8 Hz, 1H), 8.16 (d, *J* = 8.7 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 8.03 (d, *J* = 7.8 Hz, 1H), 7.99 (d, *J* = 7.8 Hz, 2H), 7.97 (d, *J* = 7.2 Hz, 1H), 7.91 (d, *J* = 9.0 Hz, 1H), 7.89 (d, *J* = 8.4 Hz, 1H), 7.69 (dd, *J* = 8.4, 8.4 Hz, 1H), 7.65 (dd, *J* = 7.2, 7.2 Hz, 1H), 7.48 (t, *J* = 8.4 Hz, 1H), 7.48 (dd, *J* = 8.4, 8.4 Hz, 2H), 7.38 (ddd, *J* = 8.4, 8.4, 1.2 Hz, 1H), 7.33 (dd, *J* = 8.4, 8.4 Hz, 1H), 7.31 (d, *J* = 9.0 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃): δ 173.0, 161.6, 136.0, 135.2, 134.7, 133.9, 132.58, 131.0, 130.8, 130.6, 130.4, 129.41, 129.38, 129.3, 129.1, 128.6, 128.4, 128.2, 127.8, 125.9, 125.5, 125.4, 125.3, 123.1, 114.9, 113.2.

IR (reflection, cm⁻¹): 3062, 1734, 1647, 1251, 1176, 986, 808, 759.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{17}NO_2$ [M]⁺ requires m/z: 399.1259, found m/z: 399.1260.

Mp: Due to the thermal tautomerization of **3** and **4**, it was difficult to measure the correct melting point of them. Actually, we attempted to measure the melting points of lactim **4d**, however, the resulting melted sample contained lactam **3d** and **4d**.

Analytical HPLC [column: CHIRALPAK IE (0.46 cm × 25 cm), eluent: n-C₆H₁₄/CH₂Cl₂ = 10:90, flow rate: 1.0 mL/min, detection: UV 254 nm, temperature: 10 °C]: $t_1 = 12.8 \text{ min}$, $t_2 = 14.5 \text{ min}$.

lactim 4d



To a solution of **3a** (395 mg, 1.00 mmol) in THF (5 mL) was added *n*-BuLi (1.31 M in *n*-C₆H₁₄, 840 μ L, 1.10 mmol) at 0 °C. The mixture was stirred at ambient temperature for 30 min, and benzoyl chloride (174 μ L, 1.50 mmol) was added to the mixture. After stirred at that temperature for 6 h, the reaction was quenched with sat. aq. NH₄Cl and extracted with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The mixture was suspended to Et₂O and the filtration afforded 301 mg (75%) of **4d** as colorless crystals.

¹H NMR (600 MHz, CDCl₃): δ 8.65 (d, *J* = 8.4 Hz, 1H), 8.52 (d, *J* = 8.4 Hz, 1H), 8.41 (dd, *J* = 8.4, 1.2 Hz, 2H), 8.12 (d, *J* = 8.4 Hz, 1H), 8.09 (d, *J* = 8.4 Hz, 1H), 8.03-7.99 (m, 4H), 7.79 (tt, *J* = 7.8, 1.2 Hz, 1H), 7.66 (ddd, *J* = 6.6, 6.6, 1.2 Hz, 1H), 7.62-7.57 (m, 3H), 7.39 (ddd, *J* = 6.6, 6.6, 1.2 Hz, 1H), 7.35 (ddd, *J* = 6.6, 6.6, 1.2 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃): δ 165.1, 154.9, 144.0, 135.1, 134.5, 134.2, 132.6, 130.7, 130.6, 130.0, 129.28, 129.25, 129.0, 128.84, 128.81, 128.6, 128.2, 128.1, 127.1, 126.9, 125.4, 124.9, 121.0, 120.1, 120.0. (one aromatic carbon is overlapping)

IR (reflection, cm⁻¹): 3074, 1742, 1600, 1577, 1404, 1234, 1061, 817.

HRMS (EI, positive): Exact mass calcd. for $C_{28}H_{17}NO_2$ [M]⁺ requires *m*/*z*: 399.1259, found *m*/*z*: 399.1258.

Analytical HPLC [column: CHIRALPAK IE (0.46 cm × 25 cm), eluent: n-C₆H₁₄/CH₂Cl₂ = 10:90, flow rate: 2.0 mL/min, detection: UV 300 nm, temperature: 10 °C]: $t_1 = 6.4$ min for (*M*)-isomer, $t_2 = 12.3$ min for (*P*)-isomer.

Specific rotation value:

 $[\alpha]_{D}^{-10} = -1,073 \ (c \ 6.0 \times 10^{-3}, n-C_{6}H_{14}/CH_{2}Cl_{2} = 10:90) \ \text{for } (M)\text{-isomer } (96-86\% \ \text{ee}).$ $[\alpha]_{D}^{-10} = +937 \ (c \ 3.0 \times 10^{-2}, n-C_{6}H_{14}/CH_{2}Cl_{2} = 10:90) \ \text{for } (P)\text{-isomer } (92-76\% \ \text{ee}).$ X-ray crystal structure analysis:



ORTEP Drawing of **4d** (50% probability ellipsoids) selected crystal data: crystal system; monoclinic space group: $P2_1/c$, a = 14.4902(13) Å, b = 5.7693(5) Å, c = 25.421(2) Å, $\beta = 102.486(9)^\circ$, V = 2074.9(3) Å³, Z = 4, $R_1 = 0.0627$, $wR_2 = 0.1539$. CCDC Number: 2025507. lactim 4e



To a solution of **3a** (200 mg, 0.677 mmol) in THF (10 mL) was added *n*-BuLi (1.29 M in *n*-C₆H₁₄, 579 μ L, 0.747 mmol) at 0 °C. The mixture was stirred at ambient temperature for 10 min and 2-naphthoyl chloride (194 mg, 1.02 mmol) was added to the mixture. After stirred at that temperature for 1 h, the reaction was quenched with sat. aq. NH₄Cl and extracted with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The mixture was suspended to Et₂O and the filtration afforded 223 mg (73%) of **4e** as colorless crystals.

¹H NMR (600 MHz, CDCl₃): δ 9.02 (s, 1H), 8.67 (d, *J* = 7.8 Hz, 1H), 8.54 (d, *J* = 9.0 Hz, 1H), 8.36 (dd, *J* = 8.4, 1.2 Hz, 1H), 8.13 (d, *J* = 9.0 Hz, 1H), 8.10 (d, *J* = 9.0 Hz, 1H), 8.07-8.05 (m, 2H), 8.04-8.00 (m, 4H), 7.97 (d, *J* = 7.8 Hz, 1H), 7.69 (dd, *J* = 6.6, 6.6 Hz, 1H), 7.67 (dd, *J* = 7.8, 7.8 Hz, 1H), 7.62 (dd, *J* = 7.8, 7.8 Hz, 1H), 7.60 (dd, *J* = 8.4, 8.4 Hz, 1H), 7.42 (ddd, *J* = 8.4, 8.4, 1.2 Hz, 1H), 7.37 (ddd, *J* = 7.8, 7.8, 1.2 Hz, 1H).

¹³C NMR (151 MHz, CDCl₃): δ 165.3, 155.1, 144.0, 136.2, 135.1, 134.6, 132.8, 132.7, 132.6, 130.7, 130.1, 129.8, 129.33, 129.31, 129.1, 128.8, 128.72, 128.68, 128.24, 128.16, 128.2, 127.2, 127.1, 127.0, 126.2, 125.8, 125.38, 125.0, 121.1, 120.2, 120.1. (one aromatic carbon is overlapping) IR (reflection, cm⁻¹): 3055, 1738, 1578, 1275, 1189, 1078, 813, 760.

HRMS (EI, positive): Exact mass calcd. for $C_{32}H_{19}NO_2$ [M]⁺ requires m/z: 449.1416, found m/z: 449.1415.

Analytical HPLC [column: CHIRALPAK IE (0.46 cm × 25 cm), eluent: n-C₆H₁₄/CH₂Cl₂ = 10:90, flow rate: 2.0 mL/min, detection: UV 300 nm, temperature: 10 °C]: $t_1 = 7.5$ min for (*M*)-isomer, $t_2 = 15.4$ min for (*P*)-isomer.

binaphthyl (S)-5cb



To a solution of (*P*)-**2cb** (20.0 mg, 0.0452 mmol, 96% ep) in EtOH (84 mL) was added LiOH (21.6 mg, 0.902 mmol) at -78 °C. After stirred at -30 °C for 15 min, the solvent was concentrated under reduced pressure. The mixture was purified by silica gel chromatography (*n*-C₆H₁₄/AcOEt = 3:1) to afford 19.1 mg (86%, 96% ep) of (*S*)-**5cb** as a yellow syrup.

¹H NMR (300 MHz, CDCl₃): δ 8.15 (d, *J* = 8.4 Hz, 1H), 8.03 (d, *J* = 8.7 Hz, 1H), 7.96 (d, *J* = 8.1 Hz, 1H), 7.77 (d, *J* = 7.8 Hz, 1H), 7.61 (s, 1H), 7.54 (ddd, *J* = 6.9, 6.9, 1.5 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 1H), 7.35-7.26 (m, 2H), 7.12 (ddd, *J* = 7.2, 7.2, 1.2 Hz, 1H), 6.94 (d, *J* = 8.1 Hz, 1H), 5.95 (s, 1H), 5.49 (s, 2H), 3.94-3.85 (m, 4H), 1.08-1.02 (m, 2H), 0.72 (dd, *J* = 6.9, 6.9 Hz, 3H), 0.04 (s, 9H).

¹³C NMR (75 MHz, CDCl₃): δ 167.8, 145.0, 143.0, 135.3, 135.2, 132.7, 130.1, 129.7, 128.7, 128.4, 128.2, 127.8, 127.5, 127.1, 126.9, 126.3, 124.8, 124.7, 123.9, 119.2, 109.9, 94.4, 67.2, 60.6, 18.3, 13.4, -1.3.

IR (neat, cm⁻¹): 2954, 1705, 1471, 1328, 1270, 1057, 983.

HRMS (EI, positive): Exact mass calcd. for $C_{29}H_{32}O_5Si$ [M]⁺ requires m/z: 488.2019, found m/z: 488.2020.

Analytical HPLC [column: CHIRALPAK AS-H (0.46 cm × 25 cm), eluent: n-C₆H₁₄/i-PrOH = 95:5, flow rate: 0.5 mL/min, detection: UV 254 nm, temperature: 25 °C]: $t_1 = 14.9$ min for (*S*)-isomer, $t_2 = 21.4$ min for (*R*)-isomer.

Specific rotation value: $[\alpha]_D^{26} = -15.8 (c \ 1.00, CHCl_3)$ for (S)-isomer (>99% ep).

binaphthyl (R)-6b



To a solution of (*M*)-**4e** (20.0 mg, 0.0445 mmol, 94% ep) in THF (1 mL) was added MeMgBr (1.06 M in THF, 1.68 mL, 1.78 mmol) at -20 °C. The mixture was stirred at -20 °C for 25 min and Boc₂O (292 µL, 1.34 mmol) was added to the mixture. After stirred at that temperature for 15 min, EtOH (2 mL) was added to the mixture and stirred at that temperature for 5 min then sat. aq. NH₄Cl was added to the mixture and extracted with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. Trifluoroacetic acid (3 mL) was added to the mixture and extracted with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The sat. aq. NaHCO₃ was added to the mixture and extracted with CH₂Cl₂. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and the filtrate was concentrated under reduced pressure. The residue was purified by silica gel chromatography (*n*-C₆H₁₄/AcOEt = 90:10) to afford 12.5 mg (85%, 94% ep) of (*R*)-**6b** as a pale yellow amorphous.

¹H NMR (300 MHz, CDCl₃): δ 8.10 (d, *J* = 9.0 Hz, 1H), 8.03 (d, *J* = 9.0 Hz, 1H), 7.97 (d, *J* = 8.4 Hz, 1H), 7.80 (d, *J* = 9.0 Hz, 1H), 7.78 (dd, *J* = 9.0, 9.0 Hz, 1H), 7.57 (dd, *J* = 8.4, 8.4 Hz, 1H), 7.44 (d, *J* = 8.4 Hz, 1H), 7.34 (dd, *J* = 8.1, 8.1 Hz, 1H), 7.20 (dd, *J* = 7.8, 7.8 Hz, 1H), 7.15 (dd, *J* = 8.1, 1.2 Hz, 1H), 7.10 (d, *J* = 9.0 Hz, 1H), 6.85 (d, *J* = 8.4 Hz, 1H), 3.87-3.94 (m, 2H), 0.66 (dd, *J* = 6.9, 6.9 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 168.1, 141.6, 136.0, 135.5, 134.3, 132.5, 130.7, 129.1, 128.5, 128.3, 128.03, 127.96, 127.86, 127.3, 127.2, 126.5, 126.4, 124.1, 122.2, 118.0, 116.4, 60.8, 13.4.

IR (reflection, cm⁻¹): 3376, 3056, 1705, 1621, 1239, 1137, 815, 769.

HRMS (EI, positive): Exact mass calcd. for $C_{23}H_{19}NO_2$ [M]⁺ requires m/z: 341.1416, found m/z: 341.1416.

Analytical HPLC [column: CHIRALPAK IB (0.46 cm × 25.0 cm), eluent: n-C₆H₁₄/i-PrOH = 90:10, flow rate: 1.0 mL/min, detection: UV 300 nm, temperature: 25 °C]: t_1 = 8.1 min for (S)-isomer, t_2 = 10.2 min for (R)-isomer.

Specific rotation value: $[\alpha]_D^{25} = +28.9 (c \ 1.15, CHCl_3)$ for (*R*)-isomer (>99% ep).

3. Determination of the Absolute Stereochemistry:

The stereochemical stability of 2 and 4 is low at ambient temperature; thus, its absolute stereochemistry was determined after transformation to stereochemically more stable 5 and 6.

Determination of the Absolute Stereochemistry of Lactone 2 and Binaphthyl 5

Bringmann and colleagues determined the absolute stereochemistry of (M)-2a and (R)-5a [Nu = (–)menthoxy, Z, Z' = H] by X-ray crystallography and reported correlation with CD spectra.⁶ By comparison with these data, the absolute stereochemistry of 2a, 2ba, 2bb, 2ca, 2cb, and 5cb resulting from DYASIN using OCS5 was determined as (P), (P), (P), (P), (P), and (S)-configuration, respectively.



Figure S1. CD spectra of (*P*)-2cb and (*S*)-5cb.

Determination of the Absolute Stereochemistry of Lactim 4 and Binaphthyl 6

Furuta, Kan, Kawabata, and colleagues reported the absolute stereochemistry and specific rotation values of (*S*)-**6a**.⁷ We synthesized **6a** via **6b** from enantioenriched **4d** and **4e** prepared by DYASIN with **OCS1** and compared its specific rotation with the literature values of **6a**.

Reported specific rotation value of (S)-**6a**: $[\alpha]_D^{20} = -133$ (c 0.55, CHCl₃, >99% ep).

Observed specific rotation value of **6a**: $[\alpha]_D^{25} = +144$ (*c* 0.055, CHCl₃, >99% ep).

By comparison with these data, the absolute stereochemistry of 4d, 4e, and 6b resulting from DYASIN using **OCS1** was determined as (*M*), (*M*), and (*R*)-configuration, respectively.

4. ¹H NMR Analysis for Tautomerization of Lactam 3d and Lactim 4d in CDCl₃ at 25 °C:

A solution of lactam **3d** or lactim **4d** in CDCl₃ was left at 25 °C and its tautomerization was measured using ¹H NMR (300 MHz) analysis.



5. Kinetic Measurements of the Racemization of Lactam 3 and Lactim 4:

HPLC analysis using a chiral stationary column equipped with a CD and a UV detector was carried out at 10 °C after proper time intervals in n-C₆H₁₄/CH₂Cl₂ = 10:90. Plotting ln a ($a = ee / ee_0$) against time, furnished a straight line, afforded the rate constants k and the half-lives of optical activity ($t_{1/2}$) of **3** and **4**.

"ee" is the observed enantiomeric excess value and " ee_0 " is initial value.



y = -1.98×10^{-2} x + 0.031600 (R² = 0.9985), k = 1.98×10^{-2} , $t_{1/2}$ = 35.1 h at 10 °C



time [h]	ee [%]	а	In a
0.00	80.0	1.00	0.00
0.230	62.0	0.775	-0.255
0.470	46.0	0.575	-0.553
0.850	30.0	0.375	-0.981
1.10	22.0	0.275	-1.29

y = -1.17 x + 0.005370 (R² = 0.9997), k = 1.17, t_{1/2} = 0.592 h at 10 °C





time [h]	ee [%]	а	In a
0.00	74.0	1.00	0.00
0.300	50.0	0.676	-0.392
0.580	36.0	0.486	-0.721
0.930	24.0	0.324	-1.13
1.23	18.0	0.243	-1.41
1.55	10.0	0.135	-2.00

y = -1.24 x + 0.004719 (R² = 0.9925), k = 1.24, t_{1/2} = 0.559 h at 10 °C

6. General Procedure of DYASIN:

Typical procedure: To a solution of the *rac*-2cb [20.0 mg in a mixed solution of *n*-hexane / *i*-Pr₂O = 10 : 1 (3.5 mL)] in a sample bottle was added OCS5 (4.0 g). After the mixture was shaken well by hand for a few seconds, the sample bottle was left at 25 °C for 24 h. The mixture was poured into the glass funnel attached PTFE filter (pore size: 0.48 μ m) and washed with ice-cooled Et₂O (80 mL). The filtrate was concentrated under reduced pressure at -10 to 0°C to afford 96% ep of (*P*)-2cb quantitatively.

More large scale of DYASIN can be performed with a similar procedure.

7. HPLC Chromatograms for Determination of Enantiomeric Ratio:

Measurement for DYASIN of Heterohelicenes: Lactone 2

rac-2a



Analysis condition Column: CHIRALPAK AS-H (0.46 cm × 25 cm) Eluent: $n-C_6H_{14}/i-PrOH = 90:10$ Flow rate: 1.0 mL/min Detection: UV 254 nm Temp.: 10 °C $t_1 = 10.5 \text{ min for } (M)\text{-isomer}, t_2 = 13.3 \text{ min for } (P)\text{-isomer}.$





Column: CHIRALPAK AS-3 (0.46 cm × 5.0 cm) Eluent: $n-C_6H_{14}/EtOH = 90:10$ Flow rate: 0.5 mL/min Detection: UV 254 nm Temp.: 10 °C $t_1 = 5.1 \text{ min for } (M)\text{-isomer}, t_2 = 7.4 \text{ min for } (P)\text{-isomer}.$



Solv.: n-C₆H₁₄ Temp.: 25 °C Time: 24 h % ep: 50 (rac)

Peak	tR [min]	Area [µV·sec]	Area %
1	10.1	6887719	50.0
2	13.0	6887774	50.0



Solv.: *n*-C₆H₁₄

001114				
Temp.: 25 °C	Peak	tR [min]	Area [µV·sec]	Area %
Time: 24 h	1	5.4	4076168	47.1
% ep: 53 (P)	2	7.6	4571967	52.9

Entry 3: 58% ep of (P)-2a



OCS: OCS3

Solv.: <i>n</i> -C ₆ H ₁₄				
Temp.: 25 °C	Peak	tR [min]	Area [µV·sec]	Area %
Time: 24 h	1	5.3	3306407	41.8
% ep: 58 (P)	2	7.5	4607209	58.2
1 \ /				

same as above





OCS: **OCS5** Solv.: *c*-C₅H₁₀ Temp.: 25 °C

Time: 24 h

% ep: 75 (P)

Peak

tR [min] Area [µV⋅sec] Area %

1113094

3381735

24.8

75.2

5.3

7.6



Analysis condition Column: CHIRALPAK AD-3 (0.46 cm × 5.0 cm) Eluent: n-C₆H₁₄/*i*-PrOH = 80:20 Flow rate: 1.0 mL/min Detection: UV 240 nm Temp.: 10 °C t_1 = 3.0 min for (*P*)-isomer, t_2 = 5.8 min for (*M*)-isomer.

rac-2bb



 $t_1 = 4.1 \text{ min for } (M)\text{-isomer}, t_2 = 21.8 \text{ min for } (P)\text{-isomer}.$





Analysis condition

Column: CHIRALPAK AD-3 (0.46 cm × 5.0 cm) Eluent: n-C₆H₁₄/*i*-PrOH = 80:20 Flow rate: 1.0 mL/min Detection: UV 240 nm Temp.: 15 °C $t_1 = 2.9$ min for (*P*)-isomer, $t_2 = 6.4$ min for (*M*)-isomer. Entry 7: 89% ep of (P)-2ba



OCS: **OCS5** Solv.: *c*-C₅H₁₀ Temp.: 25 °C Time: 24 h

% ep: 89 (P)

Peak	tR [min]	Area [µV·sec]	Area %
1	3.1	2910814	89.2
2	5.5	353288	10.8





Solv.: *c*-C₅H₁₀ Temp.: 25 °C Time: 24 h

% ep: 93 (P)

Peak	tR [min]	Area [µV·sec]	Area %
1	4.1	448807	7.4
2	22.3	5640678	92.6

Entry 9: 93% ep of (P)-2ca



OCS: **OCS5** Solv.: *i*-C₆H₁₄

Temp.: 25 °C	Peak	tR [min]	Area [µV·sec]	Area %
Time: 24 h	1	2.9	3365872	92.9
% ep: 93 (P)	2	5.6	256547	7.1





Measurement for DYASIN of Heterohelicenes: Lactim 4





Analysis condition Column: CHIRALPAK IE (0.46 cm × 25 cm) Eluent: n-C₆H₁₄/CH₂Cl₂ = 10:90 Flow rate: 2.0 mL/min Detection: UV 300 nm Temp.: 10 °C $t_1 = 6.4$ min for (*M*)-isomer, $t_2 = 12.3$ min for (*P*)-isomer.



Column: CHIRALPAK IE (0.46 cm × 25 cm) Eluent: n-C₆H₁₄/CH₂Cl₂ = 10:90 Flow rate: 2.0 mL/min Detection: UV 300 nm Temp.: 10 °C $t_1 = 7.5$ min for (*M*)-isomer, $t_2 = 15.4$ min for (*P*)-isomer. 92% ep of (*M*)-4d



Solv.: n-C₆H₁₄/i-PrOH

5017					
Temp.: 25 °C	Peak	tR [min]	Area [µV·sec]	Area %	
Time: 1 h	1	6.1	3902826	91.6	
% ep: 92 (<i>M</i>)	2	11.9	357226	8.4	



OCS: OCS1

Solv.: n-C ₆ H ₁₄ /i-PrOH						
Temp.: 25 °C	Peak	tR [min]	Area [µV·sec]	Area %		
Time: 1 h	1	7.1	8757420	94.0		
% ep: 94 (<i>M</i>)	2	14.5	557326	6.0		



Conversion of Heterohelicenes to Binaphthyl Compounds

*rac-*5**cb**





Analysis condition Column: CHIRALPAK AS-H (0.46 cm × 25 cm) Eluent: n-C₆H₁₄/*i*-PrOH = 95:5 Flow rate: 0.5 mL/min Detection: UV 254 nm Temp.: 25 °C t_1 = 14.9 min for (*S*)-isomer, t_2 = 21.4 min for (*R*)-isomer.





Analysis condition Column: CHIRALPAK IB (0.46 cm × 25 cm) Eluent: n-C₆H₁₄/*i*-PrOH = 90:10 Flow rate: 1.0 mL/min Detection: UV 300 nm Temp.: 25 °C $t_1 = 8.1$ min for (S)-isomer, $t_2 = 10.2$ min for (R)-isomer.



Peak	tR [min]	Area [µV·sec]	Area %
1	14.6	10441900	96.3
2	21.6	401421	3.7





Peak	tR [min]	Area [µV·sec]	Area %
1	8.2	235385	6.2
2	10.2	3541851	93.8

8. Computational Results:

All the DFT and TD-DFT (NStates = 30) calculations were performed at B3LYP/6-311G(d,p) level of theory by use of Gaussian 16 program at the computer facilities at Research Institute for Information Technology, Kyushu University.⁸ The initial structures were prepared by PM3 calculations using spartan 08 program.⁹

Standard state Gibbs free energies (G°) were obtained with thermal correction by frequency calculations of optimized geometries at the same level of theories.





G°: −1282	2.387843 a.u.		
Н	-0.99577	2.96028	-3.98538
С	-0.90923	2.33555	-3.08911
С	-0.82717	0.85814	-0.6836
С	0.15575	2.46246	-2.24613
C	-1.89614	1.34101	-2.84142
С	-1.8405	0.56913	-1.66054
Č	0.22499	1.68186	-1.06282
Ĥ	0.96798	3.16751	-2.46341
С	-0.79388	0.2824	0.65826
Č	-0.57027	-1.21247	3.04049
Č	0.44852	0.0566	1.25227
Č	-1.96892	0.00128	1.43575
Č	-1.85409	-0.77714	2.60639
Č	0 55357	-0.76852	2 41159
н	1 54068	-1.05126	2 80013
н	_0.50015	_1 87141	3 91396
C	-2 90632	1.07141	-3 80614
н	-2.96147	1 72391	-4 69524
C	_3 78178	0.04688	-3 63449
н	_4 55938	_0 15712	_4 37788
C	-3 67445	_0 78548	_2 49945
н	_4 35146	_1 6401	_2 39726
C	_2 73242	_0 53065	_1 53825
н	-2 64699	_1 18728	-0.6626
C	_3 2388	0.56036	1 1 2 9 6 8
н	-3.32903	1 230/6	0.26/03
C	_4 33503	0.28829	1 90597
н	-5 30639	0.73239	1.56383
C	_4 22635	_0 56032	3 02784
н	-5 1189	_0 78445	3 62084
C	-3.0071	-1.07835	3 37588
н	-2 90142	_1 71747	4 26
C	1 47316	1.6958	_0 27949
N	1 62331	0.63992	0.68585
C	2 929	0.53054	1 40662
õ	2.525	2 45135	-0.43736
õ	3 07592	1 12671	2 44498
Č	3 95216	_0 32422	0 74371
C	5 88503	-1 91077	-0.46464
Č	3 86675	-1 71299	0.83836
Č	5.00075	0 27435	0.03030
Č	5 96572	-0 52527	-0.55809
Č	4 83669	-2 50356	0.23277
н	3 04144	-2.18039	1.38841
н	5 06014	1 36735	_0 02692
н	6 78974	-0.0596	-1 10898
н	4 77318	-3 59444	0 30429
Н	6.64732	-2.53655	-0.94066

Transition Sate of Lactam 3d



	•		-
G°: −1282	2.350000 a.u.		
Imaginary	Frequency:	-104.111	5
н	-1.26324	2.83732	-4.35962
С	-0.83314	2.24989	-3.54095
С	0.1055	0.93258	-1.19514
C	-1.3308	1.02964	-3.2027
C	0.33402	2.72376	-2.87552
Ċ	0.90263	1.9734	-1.81817
Č	-0.86425	0.37314	-2.03007
H	-2.131	0.56073	-3.78887
C	0.03241	0.58403	0.22836
Č	-1.15589	0.73313	2.81586
Č	-1.00271	-0.27754	0.62692
č	0 72893	1 23404	1 32345
C	0.04453	1.25101	2 54233
C C	-1 58762	_0.20129	1 02008
ч	2 / 3060	-0.20129	2 10244
п	-2.43909	-0.84201	2.19244
С	-1.07078	0.09932	2 26474
U U	0.9639	3.00373	-3.304/4
н	0.47282	4.30439	-4.10849
C	2.24881	4.20132	-2.9365
Н	2.75564	5.10309	-3.29513
C	2.92072	3.31631	-2.07304
H	3.97602	3.49163	-1.83883
C	2.26882	2.22588	-1.55143
Н	2.85489	1.48449	-0.99961
С	2.10108	1.57725	1.32611
Н	2.76335	1.15231	0.56635
С	2.65905	2.33462	2.32752
Н	3.7217	2.59805	2.29148
С	1.88016	2.7575	3.41953
Н	2.31392	3.41792	4.17691
С	0.60068	2.27913	3.54674
Н	0.00009	2.5135	4.43319
С	-1.52705	-0.89222	-1.68576
Ν	-1.55253	-1.21216	-0.29458
0	-2.02015	-1.69763	-2.46233
С	-2.44008	-2.31721	0.17851
0	-3.62931	-2.1187	0.2309
С	-1.75308	-3.59404	0.51217
С	-0.49014	-5.99332	1.11698
С	-1.93139	-4.69889	-0.32068
С	-0.94767	-3.68627	1.64722
Ċ	-0.31872	-4.88961	1.94733
Č	-1.29475	-5.89765	-0.01394
Ĥ	-2.5642	-4.61968	-1.21319
н	-0.81255	-2.81288	2,2978
H	0.31413	-4.96657	2.83801
H	-1.42872	-6.76651	-0.66725
н	0 00964	-6.93874	1.35445
		N/ N/ N/ N/ / T	1/./TT.

Minimum Structure of Lactim **4d**

<i>G</i> °:−1	282.390740 a.u	1.	
Н	-0.62348	2.55788	-3.97801
С	-0.60115	2.01142	-3.02813
С	-0.70071	0.78689	-0.47346
С	0.48468	2.06212	-2.21552
С	-1.71619	1.18368	-2.68336
С	-1.74841	0.52801	-1.43616
С	0.46913	1.39948	-0.94958
Н	1.37782	2.62746	-2.50741
C	-0.73752	0.39069	0.91267
C	-0.57322	-0.80215	3.48361
Č	0.4916	0.20653	1.56766
Č	-1.93232	0.23877	1.71132
Č	-1.84645	-0.40145	2.96457
Č	0.56812	-0.45485	2.83786
й	1 5541	_0 67586	3 2639
н	_0 5417	-1 34631	4 43468
C	_2 74881	0.96435	-3 62544
н	-2.74001	1 521	-1 56958
C	3 74096	0.05101	3 3673
с ц	-5.74090	0.05101	-5.5075
C	2 71022	-0.12339	2 16054
с ц	-3.71932	-0.08000	1 00525
С	-4.46323	-1.4517	1 22607
с ц	-2.74700	-0.45425	-1.22097
С	-2.72070	-1.04364	-0.50215
с ц	-3.17127	1 40520	0.41940
С	-3.23231	0.67001	0.41049
U U	-4.20423	1.12675	1.04011
п	-3.23402	1.12073	2.22405
U U	-4.21292	-0.00138	2.0409
н	-5.1148/	-0.19509	3.9408
U U	-3.01022	-0.5/98/	3./483
H	-2.93302	-1.12107	4.09842
N	1.08/39	0.00904	1.02585
C	1.65062	1.31034	-0.13388
0	2./9481	1.96538	-0.53145
C	4.03158	1.35702	-0.46941
0	4.86625	2.22471	-0.5816
C	4.24009	-0.1094	-0.37932
С	4.71232	-2.84331	-0.24402
С	3.84093	-0.93481	-1.43102
C	4.88051	-0.64659	0.73675
С	5.11093	-2.01702	0.80125
С	4.08017	-2.30227	-1.35937
Н	3.34506	-0.50904	-2.31057
Н	5.19608	0.00646	1.55837
Н	5.60908	-2.44409	1.6782
Н	3.76984	-2.95337	-2.18332
Н	4.89684	-3.92146	-0.19026

Transition Sate of Lactim 4d



<i>G</i> °: −12	82. <u>3</u> 57924 a.ı	1.	
Imagina	ry Frequency	: -106.58	308
Н	-1.34534	1.83783	-3.73981
С	-0.65774	1.38857	-3.0146
С	0.9615	0.37236	-0.88538
С	-0.72155	0.07453	-2.69444
С	0.39831	2.18623	-2.46893
С	1.32026	1.6148	-1.56116
С	0.06577	-0.43707	-1.61453
Н	-1.42568	-0.59706	-3.20048
С	1.24808	-0.02638	0.47787
С	0.6583	-0.47247	3.24881
С	0.59323	-1.18611	0.94915
С	1.95954	0.71151	1.51544
С	1.51032	0.60988	2.85346
С	0.32575	-1.4199	2.34173
Н	-0.21125	-2.3305	2.63604
Н	0.35219	-0.55134	4.29833
C	0.59155	3,49607	-2.962
н	-0.18942	3 95214	-3 58146
C	1 75877	4 17197	-2 69604
н	1 91099	5 19443	_3.05582
C	2 7883	3 50651	_2 01326
н	3 77014	3 9828	_1 91939
C	2 57583	2 2526	_1 48527
н	3 /368/	1 70735	_1.40527
C	3 14762	1 44652	1 32781
н	3 73625	1 30/11	0.418
C	3 66707	2 25622	2 31206
с ц	4 58130	2.23022	2.51290
C	2.04850	2.02900	2.12500
U U	2 40757	2.55525	1 22655
С	2 00700	1.49	2 9 4 9 5 3
U U	2.00799	1.40	1 95297
п	1.57046	2 12665	4.03207
N C	0.047	-2.15005	1 15204
C O	-0.19227	-1.//112	-1.13294
C	-0.05564	-2.13262	-2.03402
C O	-1./0834	-3.34070	-1./2203
C	-1.08800	-4.46505	-2.4//4
C	-2.73115	-3.20248	-0./0109
C	-4.70775	-2.01/39	1.10057
C	-2.95854	-4.0/418	0.36282
C	-3.49247	-2.04081	-0.83949
C	-4.48177	-1.75295	0.09338
C	-3.94696	-5.77424	1.29511
Н	-2.36123	-4.98774	0.46402
H	-3.30985	-1.35916	-1.6/944
H	-5.08298	-0.84374	-0.01267
H	-4.12663	-4.45409	2.13481
H	-5.4869	-2.38664	1.89479

9. ¹H, ¹³C NMR Spectra:

¹H NMR (300 MHz) Chart of s4 in CDCl₃



¹³C NMR (75 MHz) Chart of s4 in CDCl₃



 ^1H NMR (300 MHz) Chart of s5 in CDCl_3



 ^{13}C NMR (75 MHz) Chart of s5 in CDCl_3



¹H NMR (300 MHz) Chart of s6 in CDCl₃



¹³C NMR (75 MHz) Chart of s6 in CDCl₃



¹H NMR (300 MHz) Chart of **2ba** in CDCl₃



 ^{13}C NMR (75 MHz) Chart of 2ba in CDCl_3



¹H NMR (300 MHz) Chart of **2bb** in CDCl₃



¹³C NMR (75 MHz) Chart of **2bb** in CDCl₃



¹H NMR (300 MHz) Chart of s8 in CDCl₃



¹³C NMR (75 MHz) Chart of s8 in CDCl₃



¹H NMR (300 MHz) Chart of **s9** in CDCl₃



¹³C NMR (75 MHz) Chart of **s9** in CDCl₃



¹H NMR (300 MHz) Chart of **s10** in CDCl₃



¹³C NMR (75 MHz) Chart of **s10** in CDCl₃



¹H NMR (300 MHz) Chart of **2ca** in CDCl₃



¹³C NMR (75 MHz) Chart of **2ca** in CDCl₃



¹H NMR (300 MHz) Chart of **2cb** in CDCl₃



¹³C NMR (75 MHz) Chart of **2cb** in CDCl₃



¹H NMR (300 MHz) Chart of **5cb** in CDCl₃



¹³C NMR (75 MHz) Chart of **5cb** in CDCl₃



¹H NMR (600 MHz) Chart of **3d** in CDCl₃



¹³C NMR (151 MHz) Chart of **3d** in CDCl₃



¹H NMR (600 MHz) Chart of 4d in CDCl₃



¹³C NMR (151 MHz) Chart of 4d in CDCl₃



¹H NMR (600 MHz) Chart of **4e** in CDCl₃



¹³C NMR (151 MHz) Chart of 4e in CDCl₃



¹H NMR (300 MHz) Chart of **6b** in CDCl₃



 ^{13}C NMR (75 MHz) Chart of 6b in CDCl_3



10. References and Notes:

(1) T. Zhang, D. Nguyen, P. Franco, Y. Isobe, T. Michishita and T. Murakami, *J. Pharm. Biomed. Anal.*, 2008, **46**, 882.

- (2) Y. Okamoto, R. Aburatani, T. Fukumoto and K. Hatada, Chem. Lett., 1987, 16, 1857.
- (3) Y. Kaida and Y. Okamoto, J. Chromatogr. A, 1993, 641, 267.
- (4) Y. Okamoto, M. Kawashima and K. Hatada, J. Chromatogr. A, 1987, 389, 95.
- (5) Y. Okamoto, M. Kawashima, and K. Hatada, J. Chromatogr. A, 1986, 363, 173.
- (6) G. Bringmann, B. Schöner, O. Schupp, K. Peters, E.-M. Peters and H. G. von Schnering, *Liebigs Ann. Chem.* 1994, 91.
- (7) T. Furuta, J. Yamamoto, Y. Kitamura, A. Hashimoto, H. Masu, I. Azumaya, T. Kan and T. Kawabata, *J. Org. Chem.* 2010, **75**, 7010.
- (8) Gaussian 16, Revision C.01, M. J. Frisch, et al, Gaussian, Inc., Wallingford CT, 2016.
- (9) MacSpartan 08 version 1.0.1, B. J. Deppmeier, et al, Wavefunction, Inc., Irvine CA, 2009.