# Ultrasound-assisted dopamine polymerization: rapid and oxidizing agent-free polydopamine coatings on membrane surfaces

Aydın Cihanoğlu,<sup>a</sup> Jessica D. Schiffman<sup>b</sup> and Sacide Alsoy Altınkaya\*<sup>a</sup>

<sup>a</sup> İzmir Institute of Technology, Faculty of Engineering, Department of Chemical Engineering, 35430, Urla, İzmir, TURKEY
 <sup>b</sup> University of Massachusetts-Amherst, Department of Chemical Engineering, Massachusetts, 01003, USA

\*Corresponding author (E-mail: sacidealsoy@iyte.edu.tr)

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#### Materials

Dopamine hydrochloride, Tris-HCl buffer, L-ascorbic acid, Cysteine, nitrotetrazolium blue chloride (NBT) were purchased from Sigma-Aldrich. Polysulfone (PSF) (Mw= 35 kDa) purchased from Sigma-Aldrich, and Sulfonated polyethersulfone (SPES) (Mw= 80 kDa, Sulfonation degree (SD)<30%) kindly donated by Konishi Chemicals, Japan, were used to prepare flat sheet membranes. 1-methyl-2-pyrrolidone (NMP, 99.5%) and N, N-Dimethylacetamide (DMAc, 99%) purchased from Fluka and Sigma-Aldrich, were used to dissolve the polymers. NaOH, HCl, polyethylene glycol (PEG: 35 kDa) and polyethylene oxide (PEO: 100 kDa) were purchased from Sigma-Aldrich. The water/paraffin emulsion used to determine the fouling resistance of the membranes was kindly supplied by Işıksan Kimya Corporation and Polyester non-woven fabric (05TH-100) (thickness: 161  $\mu$ m and base weight: 100 g/m<sup>2</sup>) was purchased from Hirose Paper Mfg. Co. Ltd, Japan, and used as a support layer for manufacturing the membranes. All chemicals were used without further purification and solutions were prepared using deionized (18.2 M $\Omega$  cm) water.

### **Preparation of Flat Sheet Membranes**

PSF and PSF-SPES blend membranes were prepared by the non-solvent induced phase inversion technique. The polymers were dried in a vacuum oven at 80 °C for 24 h to remove moisture. Dried PSF and the PSF:SPES blend (blending ratio is 3:1) were dissolved in NMP and DMAc:NMP mixture (DMAc:NMP ratio of 2:1), respectively by stirring at 100 rpm for 24 h. In order to eliminate air bubbles, solutions rested for 24 h without stirring and then were cast on a clean glass plate and non-woven with the help of an automated film applicator (Sheen Instrument Ltd., model number: 1133N). The initial thickness of the cast membranes was adjusted using a four-sided applicator with a gap size of 200  $\mu$ m. Following casting, the glass plate was immediately immersed into the coagulation bath including only deionized (18.2 M $\Omega$  cm) water at 25 °C. The polymer concentration in both casting solutions was adjusted to 20 wt.%.

#### **Conventional and Ultrasound-Assisted Polymerization of Dopamine**

Dopamine hydrochloride (2 mg/mL) was dissolved in Tris-HCl buffer solution (10 mM, pH 8.5, 25 °C). For conventional polymerization (Scheme 1a), the reaction solution (100 mL) was gently shaken at 70 rpm and room (25 °C) temperature. For ultrasound-assisted polymerization, an ultrasonic horn (Scheme 1b) and an ultrasonic reactor (Scheme 1c) operated at 30 W (frequency 20 kHz) and 25 W were used. In both experimental setup, the dopamine solution (100 mL) was not stirred, and the temperature was controlled by a chiller. At specific time intervals, the sample was removed to measure the formed polydopamine intensity using UV-vis spectroscopy. For coating the membranes, the sample coupons were immersed in the dopamine solution for 1 h under the same conditions used for bulk polymerization.



**Scheme 1 a)** Conventional polymerization set-up, **b)** Ultrasonic horn polymerization set-up **c)** Ultrasonic reactor polymerization set-up

#### **Characterization of Membranes**

The chemical structure of the bare and modified membranes was determined by Attenuated Total Reflectance Fourier Transformed Infrared Spectrometer (ATR-FTIR), (Perkin Elmer). Spectra were collected at ambient temperature over a scanning range of 4000-650 cm<sup>-1</sup> with a resolution of 4.0 cm<sup>-1</sup>. The water contact angle of the membranes was measured (Attension Optical tensiometer) with a 5  $\mu$ L water droplet (*n*=5). The surface free energy (SFE) calculations based on the OWRK method (Owens, Wendt, Rabel and Kaelble) were carried out using the contact angle measurements of water and diiodomethane. The X-ray photoelectron spectra (XPS, Thermo Scientific) analysis at the emission angle of 0° was used to determine the elemental composition of the membranes (n=3). The surface morphology of the membranes was characterized using a scanning electron microscope (SEM) (FEI Quanta 250 FEG). Before taking the images, the membrane surfaces were coated with gold nanoparticles with a Magnetron Sputter Coating Instrument. The surface roughness of the membranes (arithmetic mean (R<sub>a</sub>) and root-meansquare  $(R_{\alpha})$  was determined using an atomic force microscope (AFM) (MMSPM Nanoscope 8, Bruker). 5 × 5  $\mu$ m<sup>2</sup> sample area was scanned at a rate of 1 Hz using tapping mode in the air at room temperature by the TAP150 model tip (Bruker) (n=3). Prior to analysis, all the membranes were dried in a vacuum oven (Memmert) at 25 °C. (n is the number of repeated experiments). The surface free energy (SFE) values were determined by the OWRK method.<sup>1</sup> Equation 1 describes the surface tension of the solid ( $\sigma_s$ ) with respect to the interfacial tension between solid and liquid  $(\sigma_{sl})$  and the surface tension of the liquid  $(\sigma_l)$  by the Young equation:

$$\sigma_s = \sigma_{sl} + \sigma_l . \cos\theta \tag{1}$$

The OWRK method requires contact angle measurements with at least two liquids, one polar (water) and the other nonpolar (diiodomethane) to calculate  $\sigma_s$  and the  $\sigma_l$  from Equation 2.

$$\sigma_l = \sigma_l^d + \sigma_l^p, \qquad \sigma_s = \sigma_s^d + \sigma_s^p \tag{2}$$

where  $\sigma_l^d / \sigma_s^d$  and  $\sigma_l^p / \sigma_s^p$  are the disperse and polar components of the liquid and solid, respectively. The OWRK model uses geometric mean to combine the solid and liquid contributions as follows:

$$\sigma_{sl} = \sigma_s + \sigma_l - 2\left(\sqrt{\sigma_s^d \cdot \sigma_l^d} + \sqrt{\sigma_s^p \cdot \sigma_l^p}\right)$$
(3)

Substituting this term for  $\sigma_{st}$  in the Young equation (1) and solving the unknowns results in a linear expression.

$$y = mx + c \tag{4}$$

where

$$y = \frac{1 + \cos\theta}{2} \frac{\sigma_l}{\sqrt{\sigma_l^d}}, \qquad x = \sqrt{\frac{\sigma_l^p}{\sigma_l^d}}, \qquad m = \sqrt{\sigma_s^p}, \qquad c = \sqrt{\sigma_s^d}$$
(5)

Thus, plotting y versus x enables the calculation of  $\sigma_s^{\mathbb{P}}$  from the slope, and  $\sigma_s^{\mathbb{Z}}$  from the intersection with the vertical axis.

# **Membrane Filtration Performance**

The filtration performance of membranes was determined by a 50 mL dead-end stirred cell (Millipore, Amicon Stirred Cell 8050) with an effective area of 13.4 cm<sup>2</sup>. Before filtration, membrane coupons were compacted until a constant flux is reached. Next, pure water was filtered at 1 bar and collected permeate volume was recorded for specific time intervals. The volumetric flux was calculated from the slope of the permeate volume vs. time graph and converted to hydraulic pure water permeability (PWP) using following equation:

$$PWP = \frac{\Delta V}{A\Delta t \Delta P} \tag{6}$$

where  $\Delta V$  is the volume of permeated water (L), A (m<sup>2</sup>) is the membrane area,  $\Delta t$  (h) is the permeation time and  $\Delta P$  (bar) is the transmembrane pressure difference applied through the membrane. To determine the rejection characteristics of the membranes, 1 g/L aqueous solutions of 35 kDa PEG and 100 kDa PEO were filtered at 1 bar. The concentrations of the permeate, retentate and feed solutions were measured by Rudolph-J357 Automatic Refractometer. The solute rejection (%) was calculated using the equation:

$$R(\%) = \left(1 - \frac{C_p}{\frac{C_f + C_r}{2}}\right) x 100 \tag{7}$$

where  $C_{p}$ ,  $C_r$  and  $C_f$  are the concentrations of permeate, retentate and feed solution, respectively. To eliminate concentration polarization, the solution was stirred at 300 rpm. The fouling behaviour of the membranes was evaluated by filtering of water/paraffin emulsion at 1 bar. Following filtration, the membrane coupons were rinsed with pure water for 30 min and water flux was re-measured to calculate the flux recovery ratio (FRR).

$$FRR(\%) = \left(\frac{J_R}{J_W}\right) x 100 \tag{8}$$

where  $J_W$  and  $J_R$  are the pure water fluxes of the clean and the washed membranes. The experiments were carried out at room (25 °C) temperature. (*n*=3 where *n* is the number of repeated experiments).

The hydraulic resistance of the porous bare and PDA coated membranes is defined as follows:<sup>2</sup>

$$R_i = \frac{\Delta p}{\mu J_i} \tag{9}$$

where  $\Delta p$  is the transmembrane pressure,  $\mu$  is the viscosity of the feed solution,  $J_i$  is the steadystate water flux and  $R_i$  is the hydraulic resistance of the membrane.

Equation 9 can be employed for coated membranes to quantify the effect of PDA coating on membrane flux. The coating layer adds resistance to the membrane's overall hydraulic resistance, and it can be expressed as:

$$J_{PDA} = \frac{\Delta p}{\mu (R_0 + R_{PDA})} \tag{10}$$

where  $J_{PDA}$  is the pure water flux of PDA coated membrane,  $R_0$  is the hydraulic resistance of bare membrane,  $R_{PDA}$  is the hydraulic resistance of the PDA coated membrane. By combining equations 9 and 10,  $R_{PDA}$  can be calculated as follows:

$$R_{PDA} = \frac{\Delta p}{\mu} \left( \frac{1}{J_{PDA}} - \frac{1}{J_0} \right) \tag{11}$$

where  $J_0$  is the steady-state pure water flux of bare membrane.

# **Stability of Coating Layer**

The chemical stability of the PDA-coating layer on membranes was evaluated in a strong acidic and alkaline environment. To this end, coated membranes with 1.5 cm x 1.5 cm sizes were immersed in 5 mL 0.1 M NaOH and 0.1 M HCl for 24 h. Next, the leached PDA in solution was quantified by measuring the absorbance of the solution at 420 nm with UV-vis spectroscopy.<sup>3</sup>

	Peak Area				
Membranes	<b>v(N-H)</b> and <b>v(O-H)</b> peaks at 3300 cm <sup>-1</sup>				
	Bare	СР	USP		
PSF	41.1	400.5	515.2		
PSF-SPES	345.7	2030.1	3918.4		

Table S1. Peak area of the bare and PDA coated membranes.

Mombranos	Contact Angle (°)			The change in Contact Angl		
Weitibranes	Bare	СР	USP	$(\theta_{B}-\theta_{CP})/\theta_{B}$	$(\theta_{B}-\theta_{USP})/\theta_{B}$	
PSF	97.4±0.2*	76.9±0.6*	69.9±0.6*	21.0	28.2	
PSF-SPES	72.9±0.8 <sup>**</sup>	58.9±0.8 <sup>**</sup>	46.8±0.1**	19.1	35.9	

Table S2 Contact angles of the bare and PDA coated membranes.

 $\theta_{B}$ ,  $\theta_{CP}$ ,  $\theta_{USP}$  are the contact angle of bare, CP, and UPS membranes, respectively.

\*represents statistically significant difference (p<0.05) in contact angle of the bare and coated PSF membranes.

\*\*represents statistically significant difference (p<0.05) in contact angle of the bare and coated PSF-SPES membranes.

Coating		Cont	Contact Angle (°)		SFE components (mN/m)			
	technique	Water	Diiodomethane	$\sigma_{s}$	$\sigma_{s}{}^{d}$	$\sigma_s{}^p$	(0 <sub>5</sub> <sup>r</sup> -0 <sub>50</sub> <sup>r</sup> )/0 <sub>50</sub> <sup>r</sup>	$(O_s - O_{so})/O_{so}$
DCE	СР	76.9±0.6	32.2±0.4	47.1	43.3	3.8	250.9	0.080
P3F	USP	69.9±0.6	31.7±0.4	49.9	43.5	6.4	427.5	0.146
	СР	58.9±0.8	29.6±0.3	55.7	44.4	11.3	1.2	0.130
POR-OPEO	USP	46.8±0.1	31.3±0.5	61.9	43.7	18.2	2.6	0.255

**Table S3.** Surface free energy and its components of the PDA coated membranes.

 $\sigma_{so}$ : The total surface free energies of the modified membranes.

 $\sigma_{so}^{d}$  and  $\sigma_{so}^{p}$ : Dispersive and polar components of the surface free energies of the modified membranes.

 $(\sigma_s - \sigma_{so})/\sigma_{so}$  describes the change in  $\sigma_s$  of the PDA coated membranes in comparison to that of the bare one.

A high value of  $(\sigma_s - \sigma_{so})/\sigma_{so}$  means that the total surface free energy of the membranes after the PDA coating increases.

Mombranas	Conta	Contact Angle (°)		SFE components (mN/m)		
	Water	Diiodomethane	$\sigma_{so}$	$\sigma_{so}{}^d$	$\sigma_{so}{}^{p}$	
PSF	97.4±0.2	31.6±0.5	43.6	43.5	0.015	
PSF-SPES	72.9±0.8	29.8±0.4	49.3	44.3	5.048	

Table S4. Surface free energy and its components of the bare membranes.

 $\sigma_{so}$ : The total surface free energies of the bare membranes.

 $\sigma_{so}{}^{d}$  and  $\sigma_{so}{}^{p}$ : Dispersive and polar components of the surface free energies of the bare membranes.

Membranes	Coating technique	S%	С%	0%	N%	N/S	C/O
	*	3.86	82.74	13.4	-	-	6.17
PSF	СР	3.00	76.82	16.14	4.04	1.35	4.76
	USP	1.82	74.83	19.11	4.24	2.33	3.92
	*	7.86	71.12	21.02	_	_	3.38
PSF-SPES	СР	4.91	73.61	19.53	1.95	0.40	3.77
	USP	3.73	69.03	21.88	5.36	1.44	3.15

**Table S5.** XPS analysis of the bare and PDA coated membranes.

\* Bare membrane.

**Table S6.** Surface roughness of the bare and PDA coated membranes.

Membranes	R <sub>a</sub> (nm)	R <sub>q</sub> (nm)
PSF	2.53±0.16	3.15±0.19
PSF_CP	5.41±1.45	7.22±2.01
PSF_USP	3.70±0.76	4.67±0.98
PSF-SPES	3.34±0.01	4.26±0.24
PSF-SPES_CP	8.52±3.18	12.01±4.89
PSF-SPES_USP	4.03±0.45	5.23±0.70

**Table S7.** Influence of PDA deposition on membrane hydraulic resistance.

Mombranes	Hydraulic Resist	ance x 10 <sup>-10</sup> (m <sup>-1</sup> )
	PSF	PSF-SPES
Bare	415.7	39.4
PDA coated membranes with CP	1547.8	10.7
PDA coated membranes with USP	3549.9	69.1



**Fig. S1** The rate of dopamine polymerization **a)** Effect of ultrasound frequency on the absorbance of dopamine solution **b)** Absorbance of blue NBT formazan at 560 nm as function of time.



Fig. S2 The effect of scavengers on the absorbance of dopamine solution.



Fig. S3 ATR-FTIR spectra of the bare and modified a) PSF and b) PSF-SPES membranes.



Fig. S4 The general survey of the bare and modified a) PSF and b) PSF-SPES membranes.





PSF





PSF\_CP

PSF-SPES\_CP













PSF-SPES





PSF\_CP













**Fig. S7** UV-visible spectra of the eluent of PDA coated membranes immersed in strongly acidic and alkaline solutions: **a)** 0.1 M HCl, **b)** 0.1 M NaOH solutions. The immersion time is 24 h.



**Fig. S8** Size distribution and zeta potential of water/paraffin emulsion used as synthetic foulant in the experiments.

# Notes and references

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