# **Supporting Information**

# Enantioselective synthesis of *cis*-hexahydro-γcarboline derivatives via Ir-catalysed asymmetric hydrogenation

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#### I. General Information

Unless otherwise mentioned, all experiments were carried out under an atmosphere of argon in a glovebox or using standard Schlenk techniques. Solvents were dried with standard procedures and degassed with argon gas. Flash column chromatography was performed using Tsingdao silica gel (particle size 300-400 mesh). NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz for <sup>1</sup>H NMR, 101 MHz for <sup>13</sup>C NMR, 376 MHz for <sup>19</sup>F NMR or a Bruker DPX 600 spectrometer at 600 MHz for <sup>1</sup>H NMR, 151 MHz for <sup>13</sup>C NMR, 565 MHz for <sup>19</sup>F NMR in CDCl<sub>3</sub> or *d*<sup>6</sup>-DMSO with tetramethyl silane (TMS) as internal standard. Chemical shifts are reported in ppm and coupling constants are given in Hz. Chemical shifts were reported relative to TMS (0.00 ppm), *d*<sup>6</sup>-DMSO (2.50 ppm) or CDCl<sub>3</sub> (7.26 ppm) for <sup>1</sup> H NMR and relative to CDCl<sub>3</sub> (77.16 ppm) or *d*<sup>6</sup>-DMSO (39.52 ppm) for <sup>13</sup>C NMR. LC-MS analysis was carried out on Agilent 1200 Series instrument using a reversed phase column (Athena C18, 120 Å, 4.6 \*150 mm, 5 µm). The starting materials aryl hydrazine and 1-carbethoxy-4-piperidone were purchased from commercial suppliers and used without further purification. High resolution mass spectra (HRMS) were obtained on Thermo Scientific Q Exactive hybrid quadrupole-Orbitrap mass spectrometer. PE refers to petroleum ether, DCE refers to 1,2-dichloroethane, DCM refers to dichloromethane.

#### II. General Procedure for Preparation of Substrates 5a-5i, 5k, 5n-5p



Following a literature procedure<sup>1</sup>, aryl hydrazine hydrochloride (3.6 mmol) and 1-carbethoxy-4piperidone (3 mmol) were suspended in EtOH (5 mL) and heated to reflux for overnight, then cooled down to room temperature. The resulting mixture was concentrated, filtered and washed with 50% aqueous EtOH to afford the desired product. The residue was purified by flash column.



**Ethyl 1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**5a**)<sup>2</sup>: white solid, 659 mg, 90% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.96 (s, 1H), 7.46 (d, *J* = 7.8 Hz, 1H), 7.31 (d, *J* = 7.8 Hz, 1H), 7.16 (t, *J* = 7.5 Hz, 1H), 7.11 (t, *J* = 7.4 Hz, 1H), 4.70 (s, 2H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.88 (s, 2H), 2.84 (s, 2H), 1.31 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  156.3, 134.7, 133.0, 126.8, 124.1, 120.9, 116.8, 108.5, 104.4, 61.8, 41.3, 23.5, 14.9. The resulting data is according to the reported literature.



**Ethyl 8-methyl-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**5b**)<sup>3</sup> : white solid, 666 mg, 86% yield. <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) δ 10.76 (s, 1H), 7.20 – 7.18 (m, 2H), 6.89 – 6.84 (m, 1H), 4.57 (s, 2H), 4.11 (q, *J* = 7.1 Hz, 2H), 3.75 (t, *J* = 5.5 Hz, 2H), 2.78 (t, *J* = 5.5 Hz, 2H), 2.36 (s, 3H), 1.23 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) δ 155.6, 134.7, 133.0, 127.4, 125.8, 122.6, 117.4, 111.1, 105.4, 61.3, 41.4, 23.4, 21.6, 15.1. <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) δ 155.2, 134.2, 132.5, 127.0, 125.4, 122.1, 116.9, 110.6, 105.0, 60.8, 41.00, 22.9, 21.2, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup>: 259.1441, found: 259.1441. The resulting data is according to the reported literature.



Ethyl 8-(tert-butyl)-1,3,4,5-tetrahydro-2H-pyrido[4,3-b]indole-2-carboxylate (5c): white solid,

630 mg, 70% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.79 (s, 1H), 7.44 (s, 1H), 7.25 (s, 2H), 4.70 (s, 2H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.86 (s, 2H), 2.83 (s, 2H), 1.39 (s, 9H), 1.32 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO) δ 155.2, 140.8, 134.0, 132.4, 124.9, 118.7, 112.9, 110.4, 105.4, 60.8, 41.1, 34.2, 31.9, 23.0, 14.7. HRMS (ESI/ion trap) *m/z*:  $[M + H]^+$  calcd for C<sub>18</sub>H<sub>25</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup>:301.1911, found:301.1912.



**Ethyl 8-methoxy-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b***]<b>indole-2-carboxylate** (**5d**)<sup>4</sup> : white solid, 477 mg, 58% yield. <sup>1</sup>H NMR (600 MHz,  $d^6$ -DMSO)  $\delta$  10.72 (s, 1H), 7.17 (d, J = 8.6 Hz, 1H), 6.93 (s, 1H), 6.67 (d, J = 8.6 Hz, 1H), 4.55 (s, 2H), 4.10 (q, J = 7.0 Hz, 2H), 3.80 – 3.70 (m, 5H), 2.77 (s, 2H), 1.22 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (151 MHz,  $d^6$ -DMSO)  $\delta$  155.2, 153.1, 133.2, 130.8, 125.4, 111.5, 110.3, 105.3, 99.5, 60.8, 55.3, 41.1, 23.3, 23.0, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 275.1390, found: 275.1391. The resulting data is according to the reported literature.



**Ethyl 8-(benzyloxy)-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b***]<b>indole-2-carboxylate** (**5e**)<sup>5</sup> : white solid, 714 mg, 68% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.10 – 8.07 (brs, 1H), 7.78 (d, *J* = 7.4 Hz, 2H), 7.69 (t, *J* = 7.5 Hz, 2H), 7.62 (t, *J* = 7.2 Hz, 1H), 7.50 (d, *J* = 8.3 Hz, 1H), 7.30 (d, *J* = 1.5 Hz, 1H), 7.19 (d, *J* = 8.6 Hz, 1H), 5.40 (s, 2H), 4.95 (s, 2H), 4.51 (q, *J* = 7.1 Hz, 2H), 4.17 (s, 2H), 3.12 (s, 2H), 1.61 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.2, 152.2, 137.9, 133.3, 131.1, 128.3, 127.6, 127.6, 125.5, 111.6, 111.0, 105.5, 101.1, 69.8, 60.9, 41.1, 23.3, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup>: 351.1703, found: 351.1705.



**Ethyl 8-fluoro-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate**(**5f**)<sup>6</sup> : white solid, 652 mg, 83% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (s, 1H), 7.20 (s, 1H), 7.09 (d, *J* = 8.7 Hz, 1H), 6.89 (t, *J* = 8.2 Hz, 1H), 4.64 (s, 2H), 4.21 (q, *J* = 7.1 Hz, 2H), 3.86 (s, 2H), 2.83 (s, 2H), 1.31 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  156.8 (d, *J* = 231.0 Hz), 155.1, 134.8, 125.3 (d, *J* = 10.1 Hz), 111.7 (d, *J* = 9.8 Hz), 108.4 (d, *J* = 25.9 Hz), 106.0, 102.3 (d, *J* = 23.4 Hz), 60.9, 40.9, 23.0, 14.7. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -124.4. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub><sup>-</sup>: 261.1039, found: 261.1044.



Ethyl 8-chloro-1,3,4,5-tetrahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (5g)<sup>5</sup> : white solid, 678 mg, 81% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.97 (s, 1H), 7.41 (s, 1H), 7.21 (d, *J* = 8.4 Hz, 1H), 7.14 – 7.06 (m, 1H), 4.64 (s, 2H), 4.20 (q, *J* = 7.1 Hz, 2H), 3.87 (s, 2H), 2.84 (s, 2H), 1.30 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  155.1, 134.5, 134.3, 126.2, 123.2, 120.5, 116.7, 112.3, 105.6, 60.9, 40.8, 22.9, 14.6. HRMS (ESI/ion trap) *m/z*: [M - H]<sup>-</sup> calcd for C<sub>14</sub>H<sub>14</sub>ClN<sub>2</sub>O<sub>2</sub><sup>-</sup> : 277.0744, found: 277.0747.



**Ethyl 8-bromo-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**5h**)<sup>5</sup> : white solid, 844 mg, 87% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.96 (s, 1H), 7.57 (s, 1H), 7.23 (dd, *J* = 8.5, 1.6 Hz, 1H), 7.17 (d, *J* = 8.4 Hz, 1H), 4.64 (s, 2H), 4.20 (q, *J* = 7.1 Hz, 2H), 3.86 (s, 2H), 2.84 (s, 2H), 1.30 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  155.1, 134.5, 126.9, 123.0, 119.7, 112.8, 111.1, 105.6, 60.9, 40.8, 22.9, 14.6. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>16</sub>BrN<sub>2</sub>O<sub>2</sub><sup>-</sup> : 323.0390, found: 323.0392.



Ethyl 8-(trifluoromethyl)-1,3,4,5-tetrahydro-2H-pyrido[4,3-b]indole-2-carboxylate (**5i**): white solid, 805 mg, 86% yield. <sup>1</sup>H NMR (600 MHz,  $d^6$ -DMSO) δ 11.42 (d, J = 6.7 Hz, 1H), 7.85 (d, J = 7.1 Hz, 1H), 7.59 – 7.39 (m, 1H), 7.34 (t, J = 7.6 Hz, 1H), 4.64 (s, 2H), 4.22 – 4.04 (m, 2H), 3.90 – 3.68 (m, 2H), 2.83 (s, 2H), 1.36 – 1.05 (m, 3H). <sup>13</sup>C NMR (151 MHz,  $d^6$ -DMSO) δ 155.6, 137.9, 135.6, 126.2 (q, J = 271.1 Hz), 124.9, 119.9 (q, J = 31.3 Hz), 117.6, 115.5, 111.9, 107.2 (m), 61.4, 41.24, 23.3, 15.1. <sup>19</sup>F NMR (565 MHz,  $d^6$ -DMSO) δ -58.3. HRMS (ESI/ion trap) m/z: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>16</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup>: 313.1158, found: 313.1154.



Ethyl 7-methoxy-1,3,4,5-tetrahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (5k): following the general procedure, the crude product was purified by flash column (EtOAc : hexane, 2:1), afforded

the desired product (30% yield, 247 mg). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.85 (s, 1H), 7.32 (d, J = 8.5 Hz, 1H), 6.82 (s, 1H), 6.77 (dd, J = 8.5, 2.0 Hz, 1H), 4.65 (s, 2H), 4.20 (q, J = 7.1 Hz, 2H), 3.84 (d, J = 10.7 Hz, 5H), 2.80 (s, 2H), 1.30 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz,  $d^6$ -DMSO)  $\delta$  155.1, 153.1, 137.1, 130.5, 121.5, 115.4, 105.1, 104.5, 99.0, 60.8, 55.1, 42.4, 40.8, 22.8, 14.7. HRMS (ESI/ion trap) m/z: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 275.1390, found: 275.1391.



**Ethyl 9-methoxy-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**5n**) : following the general procedure, the crude product was purified by flash column (EtOAc : hexane, 2:1), afforded the desired product (16% yield,132 mg). <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) δ 10.86 (s, 1H), 6.92 (t, *J* = 7.8 Hz, 1H), 6.88 (d, *J* = 8.0 Hz, 1H), 6.44 (d, *J* = 7.6 Hz, 1H), 4.70 (s, 2H), 4.10 (d, *J* = 6.6 Hz, 2H), 3.83 (s, 3H), 3.71 (t, *J* = 5.6 Hz, 2H), 2.74 (d, *J* = 5.0 Hz, 2H), 1.21 (t, *J* = 7.0 Hz, 3H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) δ 155.1, 153.1, 137.1, 130.5, 121.5, 115.4, 105.3, 104.5, 99.0, 60.8, 55.1, 42.3, 23.1, 22.8, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 275.1390, found: 275.1391.



**Ethyl 7,9-dimethyl-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**50**) <sup>7</sup> : white solid, 685 mg, 84% yield. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.73 (s, 1H), 6.91 (s, 1H), 6.68 (s, 1H), 4.90 (s, 2H), 4.21 (q, *J* = 7.1 Hz, 2H), 3.83 (s, 2H), 2.80 (s, 2H), 2.58 (s, 3H), 2.39 (s, 3H), 1.31 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  155.1, 136.3, 130.9, 129.6, 127.9, 122.6, 121.3, 108.7, 105.8, 60.8, 42.5, 40.8, 40.2, 23.2, 21.2, 19.4, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>16</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup>: 273.1598, found: 273.1599.



**Ethyl 6-fluoro-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**5p**) : white solid, 597 mg, 76% yield. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.20 (s, 1H), 7.21 (d, *J* = 7.8 Hz, 1H), 7.01 (td, *J* = 7.9, 4.8 Hz, 1H), 6.87 (dd, *J* = 11.1, 8.0 Hz, 1H), 4.69 (s, 2H), 4.22 (q, *J* = 7.1 Hz, 2H), 3.87 (s, 2H), 2.86 (s, 2H), 1.31 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  155.1, 148.9 (d, *J* = 241.9 Hz), 134.9, 129.0 (d, *J* = 3.4 Hz), 123.4 (d, *J* = 13.0 Hz), 119.0 (d, *J* = 6.3 Hz), 113.5 (d, *J* = 2.5 Hz),

106.6, 105.8 (d, J = 16.3 Hz), 60.9, 40.9, 22.9, 14.6. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.1. HRMS (ESI/ion trap) m/z: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>16</sub>FN<sub>2</sub>O<sub>2</sub><sup>+</sup>: 263.1190, found: 263.1191.

#### **III. Procedure for Preparation of Substrate 5j**



Following the general procedure, <sup>8</sup> *N*-Benzyl-4-piperidone (2.9 g, 17 mmol), 2-fluoro-5methylaniline (1.88 g, 15 mmol) and catalytic *p*-TsOH (50 mg) were combined in toluene (80 mL) and heated at reflux with removal of  $H_2O$  using a Dean–Stark trap. After 15 h the reaction was cooled to r.t. washed with sat. NaHCO<sub>3</sub> and concentrated in vacuo. The crude reaction mixture was then purified under vacuum under heating to remove unreacted starting materials, to give the imine as a bright yellow oil (1.69 g, 90%). This material was sufficiently pure to be used without further purification.



**2-Benzyl-7-methyl-2,3,4,5-tetrahydro-***1H***-pyrido[4,3-***b***]<b>indole** (**S5j**) : The imine (1.39 g, 5 mmol) was dissolved in anhydrous THF (15 mL) under Ar in an oven dried flask equipped with a reflux condenser. The solution was cooled to -78 °C and a fresh prepared LDA (0.69 M, 12.5 mmol) in THF (18 mL) was added via syringe. The reaction was stirred at -78 °C for 15 min, then warmed to r.t. and heated to reflux under an atmosphere of argon gas. TLC analysis showed a quantitative conversion to a new product after 5 h at reflux and the reaction was cooled to rt, quenched with H<sub>2</sub>O (2 mL) and filtrated through celite, washed with CH<sub>2</sub>Cl<sub>2</sub> (20 mL). The mixture was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated in vacuo. The residue was purified by flash chromatography (EtOAc : hexane, 1:1) to give **S5j** as a yellow solid (1.27 g, 92% yield). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.74 (s, 1H), 7.41 (d, *J* = 7.6 Hz, 2H), 7.33 (t, *J* = 7.5 Hz, 2H), 7.27 (t, *J* = 7.1 Hz, 1H), 7.23 (d, *J* = 7.8 Hz, 1H), 6.94 (s, 1H), 6.87 (d, *J* = 8.0 Hz, 1H), 3.78 (s, 2H), 3.70 (s, 2H), 2.85 (t, *J* = 5.7 Hz, 2H), 2.71 (t, *J* = 5.5 Hz, 2H), 2.40 (s, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  138.6, 136.5, 131.3, 130.7, 129.2, 128.3, 127.1, 124.0, 120.8, 117.1, 110.7, 108.5, 62.4, 50.2, 49.8, 23.7, 21.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>20</sub>N<sub>2</sub><sup>-</sup>: 277.1699, found: 277.1689.



Ethyl 7-methyl-1,3,4,5-tetrahydro-2H-pyrido[4,3-b]indole-2-carboxylate (5j) : Following the literature procedure, S5j (1.0 g, 3.6 mmol) and 10 mol% Pd/C (5 wt%) (766 mg) were suspended in a 70% EtOH/ H<sub>2</sub>O solution (20 mL) and placed under hydrogen at atmospheric pressure. The reaction was heated to 70 °C and stirred for 24 h, after which the reaction was filtered through filter paper before cooling to remove Pd/C. The filter cake was washed with 70% EtOH/H<sub>2</sub>O ( $3 \times 50$  mL) and the combined washes and filtrate were concentrated in vacuo. The crude product was recrystallized from a 70% EtOH/H2O solution, and the precipitate was isolated by filtration, washed with cold MeOH, and dried in vacuo. The resulted product was used directly in next step. The crude product was dissolved in DCM (20 mL), and saturated NaHCO<sub>3</sub> (10 mL) was added, then the resulted mixture was cold at ice-water bath. Ethyl chloroformate (1.3 eq.) was added dropwise to the mixture. After completion, the organic phase was separated and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, concentrated under vacuum. The resulted residue was purified by flash column (EtOAc : hexane, 2:1), afford product 5j (817 mg, 88% yield). <sup>1</sup>H NMR (400 MHz, d<sup>6</sup>-DMSO) δ 10.73 (s, 1H), 7.26 (d, J = 7.9 Hz, 1H), 7.08 (s, 1H), 6.79 (d, J = 8.0 Hz, 1H), 4.54 (s, 2H), 4.09 (q, J = 7.1 Hz, 2H), 3.74 (t, J = 5.6 Hz, 2H), 2.76 (t, J = 5.1 Hz, 2H), 2.37 (s, 3H), 1.21 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, d<sup>6</sup>-DMSO) δ 155.1, 136.3, 131.7, 129.6, 123.0, 120.1, 116.9, 110.9, 105.2, 60.8, 41.0, 40.1, 22.9, 21.4, 14.7. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub><sup>-</sup>: 259.1441, found: 259.1440.

#### IV. Procedure for Preparation of Substrate 51 and 5m



To a suspension of 2-chloro-3-fluoroaniline (5.0 g, 33.3 mmol) in water (8.4 mL) and concentrated HCl (8.4 mL) at 0 °C, a solution of sodium nitrite (3.68 g, 53.3 mmol) in water (8 mL) was added dropwise. After the mixture was stirred at the same temperature for 0.5 h, a solution of tin(II) chloride hydrate (13.6 g, 60 mmol) in concentrated hydrochloric acid (15 mL) was added dropwise at 0 °C, and the mixture was stirred at ambient temperature overnight. The reaction mixture was filtered, and the filtrate was extracted with EA, the organic phase was discarded and the water phase was neutralized by NaOH (12 M) solution, extracted with DCM, and dried under reduced pressure, the residue was formed as hydrochloride salt in 1,4-dioxane to give the (2-chloro-3-fluorophenyl) hydrazine hydrochloride as a white powder (2.7 g, yield 42%). <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  10.52 (brs, 3H), 8.41 (s, 1H), 7.35 (dd, *J* = 14.5, 8.3 Hz, 1H), 6.98 (t, *J* = 7.3 Hz, 2H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  157.8 (d, *J* = 244.8 Hz), 143.4, 128.5 (d, *J* = 9.5 Hz), 110.1, 108.8 (d, *J* = 21.3 Hz), 106.5 (d, *J* = 20.9 Hz). <sup>19</sup>F NMR (565 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  -115.6.



**Ethyl 6-chloro-7-fluoro-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**S5I**): (2-chloro-3-fluorophenyl)hydrazine hydrochloride (1.96 g, 10 mmol) and 1-carbethoxy-4-piperidone (1.54 g, 9 mmol) were suspended in EtOH (15 mL) and heated to reflux for overnight, then cooled down to room temperature. The resulting mixture was concentrated, filtered and washed with 50% aqueous EtOH to afford the desired product **S5I** (1.5 g, 56% yield). <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) δ 11.43 (s, 1H), 7.39 (dd, *J* = 8.0, 4.5 Hz, 1H), 7.04 – 6.94 (m, 1H), 4.56 (s, 2H), 4.09 (q, *J* = 7.1 Hz, 2H), 3.74 (t, *J* = 5.7 Hz, 2H), 2.80 (t, *J* = 5.5 Hz, 2H), 1.21 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) δ 155.1, 153.5 (d, *J* = 236.5 Hz), 134.7, 133.2, 123.1, 116.7 (d, *J* = 9.1 Hz), 107.7 (d, *J* = 23.5 Hz), 106.9, 101.9 (d, *J* = 21.9 Hz), 60.9, 40.7, 40.1, 23.2, 14.6. <sup>19</sup>F NMR (565 MHz, *d*<sup>6</sup>-DMSO) δ -128.8. HRMS (ESI/ion trap) *m/z*: [M – H]<sup>-</sup> calcd for C<sub>14</sub>H<sub>13</sub>ClFN<sub>2</sub>O<sub>2</sub><sup>-</sup>: 295.0655, found: 295.0652.



**Ethyl 7-fluoro-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b***]<b>indole-2-carboxylate** (**5l**): A mixture of S1I (1.2 g, 4.0 mmol), 5 mol% Pd/C (824 mg, 10 mol%) and Et<sub>3</sub>N (1.44 g, 12.1 mmol) in MeOH (10 mL) and THF (3 mL) was stirred under hydrogen atmosphere (10 bar) at room temperature overnight. The mixture was filtered and the filtrate was concentrated to afford the crude product. After removing the ammonium salt through extraction, product 5I (0.99 g, 95%yield) was afforded. <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) δ 11.01 (s, 1H), 7.39 (s, 1H), 7.08 (d, *J* = 10.1 Hz, 1H), 6.81 (t, *J* = 9.2 Hz, 1H), 4.57 (s, 2H), 4.10 (q, *J* = 7.0 Hz, 2H), 3.74 (t, *J* = 5.4 Hz, 2H), 2.77 (s, 2H), 1.21 (t, *J* = 7.0 Hz, 3H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) δ 158.6 (d, *J* = 233.3 Hz), 155.1, 135.8 (d, *J* = 12.1 Hz), 133.2, 121.2, 118.0 (d, *J* = 10.1 Hz), 106.7 (d, *J* = 24.1 Hz), 105.8, 97.2 (d, *J* = 25.5 Hz), 60.9, 40.9, 40.1, 23.1, 14.6. <sup>19</sup>F NMR (565 MHz, *d*<sup>6</sup>-DMSO) δ -122.5. HRMS (ESI/ion trap) *m/z*: [M – H]<sup>-</sup> calcd for C<sub>14</sub>H<sub>14</sub>FN<sub>2</sub>O<sub>2</sub><sup>-</sup>: 261.1045, found: 261.1042.

Following the procedure for preparation of 5l, substrate 5m was prepared as follows:



Followed by the procedure of synthesis of **5**l, using 2-bromo-5-methylaniline (5 g, 25 mmol) as starting material, (2-bromo-5-methyl)hydrazine hydrochloride (2.8 g, 48% yield) was afford.



**Ethyl 6-bromo-9-methyl-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-***b*]**indole-2-carboxylate** (**S5m**) : prepared using (2-bromo-5-methyl)hydrazine hydrochloride (2.0 g, 8.4 mmol) and 1-carbethoxy-4-piperidone (1.28 g, 7.5 mmol), afford 1.82 g, 72% yield. <sup>1</sup>H NMR (600 MHz,  $d^6$ -DMSO) δ 11.04 (s, 1H), 7.10 (d, J = 7.7 Hz, 1H), 6.66 (d, J = 7.8 Hz, 1H), 4.81 (s, 2H), 4.11 (q, J = 7.1 Hz, 2H), 3.72 (t, J = 5.7 Hz, 2H), 2.81 (t, J = 5.5 Hz, 2H), 2.50 (s, 3H), 1.22 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (151 MHz,  $d^6$ -DMSO) δ 155.1, 133.9, 133.4, 128.2, 126.2, 123.0, 121.0, 107.5, 101.2, 60.9, 42.2, 40.6, 23.3, 19.1, 14.6. HRMS (ESI/ion trap) m/z: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>18</sub>BrN<sub>2</sub>O<sub>2</sub><sup>+</sup>: 337.0546, found: 337.0544.



**Ethyl 9-methyl-1,3,4,5-tetrahydro-***2H***-pyrido**[**4,3-b**]**indole-2-carboxylate** (**5m**) : prepared by using **S5m** (1.35 g, 4.0 mmol), afford **5m** with 1.01 g, 98% yield. <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  10.84 (s, 1H), 7.09 (d, *J* = 8.0 Hz, 1H), 6.92 – 6.85 (m, 1H), 6.69 (d, *J* = 7.1 Hz, 1H), 4.82 (s, 2H), 4.10 (q, *J* = 7.1 Hz, 2H), 3.72 (t, *J* = 5.8 Hz, 2H), 2.77 (t, *J* = 5.7 Hz, 2H), 2.52 (s, 3H), 1.22 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO)  $\delta$  155.1, 135.8, 131.8, 128.3, 124.6, 120.7, 119.5, 108.7, 106.0, 60.8, 54.9(CH<sub>2</sub>Cl<sub>2</sub>), 42.5, 40.7, 23.2, 19.6, 14.6. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub><sup>-</sup>: 259.1441, found: 259.1441.

#### V. Procedure for Preparation of Substrate 5q



Following the procedure of literature<sup>9</sup>, sodium acetate (1.98 g, 24.2 mmol) and hydroxylamine hydrochloride (1.68 g, 24.2 mmol) were added to commercially available 1,2,3,9-tetrahydro-*4H*-carbazol-4-one (2.98 g, 16.1 mmol), in 40 mL of EtOH/water 2/1 v/v, and the mixture was refluxed 24 h under nitrogen atmosphere. After cooling, the solvent was removed and the residue was suspended in 150 mL of water and triturated, until precipitation of the desired oxime product as brown solid (**S1q**), which was collected by filtration and recrystallized from EtOH and water (2.74 g, yield 85%). Analytical and spectral data were in agreement with those of literature. <sup>1</sup>H NMR (600 MHz,  $d^6$ -DMSO)  $\delta$  11.41 (s, 1H), 8.17 (d, *J* = 7.9 Hz, 1H), 7.38 (t, *J* = 5.0 Hz, 1H), 7.28 (d, *J* = 8.0 Hz, 1H), 7.09 – 7.03 (m, 1H), 7.04 – 6.96 (m, 1H), 3.33 (s, 1H), 3.20 (dd, *J* = 9.6, 5.0 Hz, 2H), 3.10 (t, *J* = 6.7 Hz, 2H), 1.99 (dt, *J* = 11.0, 7.4 Hz, 2H).

Compound **S1q** (2.6 g, 13 mmol) of was added portionwise to 97 g of preheated (110 °C) PPA under vigorous stirring, and the mixture was stirred at this temperature for 30 min. Then, 200 g of ice were carefully poured into the mixture and triturated until complete dissolution of PPA and formation of a grey precipitate, which was collected by filtration under reduced pressure, washed with 100 mL of water, 10 mL of 5% diluted ammonia and further 100 mL of water. The solid was then suspended into 50 mL of MeOH and, after addition of 1.0 g of vegetal carbon, refluxed for 1 h. After cooling, the suspension was filtered on a Celite pad and the solvent removed to furnish **S2q** as pale brown solid (1.1 g) without further purification.



#### Ethyl 3,4,5,6-tetrahydroazepino[4,3-b]indole-2(1H)-carboxylate (5q) :

LiAlH<sub>4</sub> (1.25 g, 33 mmol) was added to a solution of **S2q** (0.6 g, 3 mmol) in 100 mL of dry 1,4dioxane, and the mixture was refluxed until disappearance of the starting material (TLC). After cooling, the reaction was quenched by adding 10 mL of a saturated solution of sodium sulfate (Na<sub>2</sub>SO<sub>4</sub>) and stirred at rt for 30 min. The mixture was then filtered and the precipitate was washed with 20 mL of 1,4-dioxane. The collected filtrates were concentrated and the residue, suspended in 80 mL of distilled water, was extracted with  $3 \times 50$  mL of CH<sub>2</sub>Cl<sub>2</sub>. The organic fractions were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated under reduced pressure, afforded an oil residue (0.5 g) without further purification. Then the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and 10 mL saturated NaHCO<sub>3</sub> at an ice bath, ethyl chloroformate (1.3 eq.) was added dropwise, then warmed the reaction to room temperature, after 0.5 h traced by TLC until disappearance of the starting material. Separation of the organic phase and the water phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL x3), combined the organic phase and washed with saturated brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, concentrated under evaporator, afforded the crude residue. Then crude product was purified through flash column (EtOAc/PE, 1:2), afforded compound **5q** as white solid. (558 mg, 72% yield). <sup>1</sup>H NMR (600 MHz, CD<sub>2</sub>Cl<sub>2</sub>)  $\delta$  8.54 – 8.07 (m, 1H), 7.52 (d, *J* = 7.2 Hz, 1H), 7.41 – 7.17 (m, 1H), 7.18 – 6.96 (m, 2H), 4.87 – 4.62 (m, 2H), 4.24 – 3.95 (m, 2H), 3.96 – 3.67 (m, 2H), 2.93 (s, 2H), 2.03 – 1.80 (m, 2H), 1.32 – 0.95 (m, 3H). <sup>13</sup>C NMR (151 MHz, CD<sub>2</sub>Cl<sub>2</sub>)  $\delta$  156.3, 137.1, 134.8, 128.1, 121.3, 119.7, 117.8, 111.6, 110.9, 61.6, 49.7, 43.2, 27.8, 27.2, 14.9. **rotamer** <sup>13</sup>C NMR (151 MHz, CD<sub>2</sub>Cl<sub>2</sub>)  $\delta$  156.3, 137.7, 134.8, 128.0, 121.1, 119.7, 117.6, 111.8, 110.8, 61.6, 50.4, 45.0, 44.2, 42.9, 27.8, 26.8, 14.8. HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 259.1441, found: 259.1440.

Ethyl 1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indole-2-carboxylate (**5r**)<sup>10</sup>: Following the reported literature, to a solution of 1,2,3,4-Tetrahydro- $\beta$ -carboline (344 mg, 2 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) was added slowly ethyl carbonochloridate (245 mg, 2.3 mmol) and Et<sub>3</sub>N (607 mg, 6 mmol) at 0 °C. After the addition was completed, the reaction mixture was allowed to reach room temperature and stirred for 3 h. And then the solvent was evaporated to get the crude product, which was crystallized from water to give the desired product (450 mg, 92%) as a white solid.

<sup>1</sup>H NMR (400 MHz, DMSO)  $\delta$  10.83 (s, 1H), 7.39 (d, J = 7.7 Hz, 1H), 7.31 (d, J = 8.0 Hz, 1H), 7.10 – 7.02 (m, 1H), 7.00 – 6.91 (m, 1H), 4.61 (s, 2H), 4.10 (q, J = 7.1 Hz, 2H), 3.72 (t, J = 5.7 Hz, 2H), 2.70 (t, J = 5.6 Hz, 2H), 1.22 (t, J = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO)  $\delta$  155.1, 135.9, 131.1, 126.5, 120.8, 118.5, 117.5, 111.0, 106.7, 61.0, 41.8, 21.0, 14.6.

HRMS (ESI/ion trap) m/z:  $[M + H]^+$  calcd for  $C_{14}H_{17}N_2O_2^+$ : 245.1285, found: 245.1286.

#### VI. General Procedure for the Asymmetric Hydrogenation Reaction

In the argon-filled glovebox, a solution of  $[Ir(COD)Cl]_2$  (6.72 mg, 0.01 mmol) and ZhaoPhos (18.2 mg, 0.021 mmol) in 2.0 mL anhydrous solvent was stirred at room temperature for 20 min. A specified volume of the resulting solution (100 µL, 1 mol% Ir-Zhaophos catalyst) was transferred by syringe to a score-break ampule charged with substrate (0.1 mmol in 1.0 mL dichloromethane) and TsOH (1.2 eq.) as acid additive. The ampule was placed into an autoclave, which was then charged with desired pressure of hydrogen gas. The autoclave was stirred at desired temperature for the indicated period of time. After release of H<sub>2</sub>, saturated sodium bicarbonate solution and dichloromethane was added and the mixture was stirred for 10 min. The organic layer was dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of solvent, the crude product was analyzed by <sup>1</sup>H NMR to determine the conversion. Purification was performed by silica gel column chromatography, eluted with Petrol ether / EtOAc, to give the desired product. The enantiomeric excess was determined by HPLC analysis.



**Ethyl (4aS,9b***R*)- 1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6a): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 24.2 mg, 98% yield, 93% ee,  $[\alpha]_D^{23}$  + 75.5 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.13 (d, *J* = 7.2 Hz, 1H), 7.05 (td, *J* = 7.6, 1.2 Hz, 1H), 6.73 (td, *J* = 7.4, 0.8 Hz, 1H), 6.66 (d, *J* = 7.8 Hz, 1H), 4.20 – 4.06 (m, 2H), 3.97 (dt, *J* = 6.8, 4.9 Hz, 1H), 3.96 – 3.64 (m, 2H), 3.63 – 3.53 (m, 1H), 3.49 – 3.11 (m, 3H), 1.90 (ddt, *J* = 14.1, 9.3, 4.6 Hz, 1H), 1.76 (ddd, *J* = 14.4, 9.3, 5.3 Hz, 1H), 1.26 (t, *J* = 6.6 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.7, 150.9, 130.1, 128.1, 124.3, 119.1, 110.0, 61.3, 57.5, 43.9, 41.0, 39.8, 28.1, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 11.140 min (*R*, *S*) (minor), 12.028 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>19</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 247.1441, found: 247.1442.



Ethyl (4a*S*,9b*R*)-8-methyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6b): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.2 mg, 97% yield, 94% ee, [ $\alpha$ ]<sub>D</sub><sup>25</sup> + 53.9 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.95 (s, 1H), 6.86 (d, *J* = 7.8 Hz, 1H), 6.58 (d, *J* = 7.8 Hz, 1H), 4.13 (dtt, *J* = 10.6, 7.1, 3.6 Hz, 2H), 3.97 – 3.92 (m, 1H), 3.91 – 3.50 (m, 3H), 3.49 – 3.04 (m, 3H), 2.25 (s, 3H), 1.89 (ddt, *J* = 14.1, 9.4, 4.7 Hz, 1H), 1.76 (td, *J* = 9.2, 4.7 Hz, 1H), 1.27 (t, *J* = 6.3 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  155.8, 148.5, 130.9, 130.5, 128.4, 125.1, 110.1, 61.3, 57.8, 44.0, 41.2, 39.9, 28.2, 20.9, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 7.090 min (*R*, *S*) (minor), 9.004 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 261.1598, found: 261.1599.



**Ethyl (4aS,9b***R*)-8-(*tert*-butyl)-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6c): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 29.5 mg, 97% yield, 91% ee,  $[\alpha]_D^{25}$  + 35.0 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.20 (s, 1H), 7.11 (dd, *J* = 8.1, 2.0 Hz, 1H), 6.64 (d, *J* = 8.1 Hz, 1H), 4.16 (q, *J* = 7.1 Hz, 2H), 4.03 – 3.95 (m, 2H), 3.93 – 2.93 (m, 5H), 1.93 (ddt, *J* = 14.2, 9.4, 4.6 Hz, 1H), 1.80 (dd, *J* = 9.6, 4.7 Hz, 1H), 1.31 (s, 12H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 155.8, 148.4, 142.4, 130.1, 124.8, 121.2, 109.6, 61.4, 57.8, 44.2, 41.2, 39.9, 34.3, 31.8, 28.2, 14.9. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak AD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 5.955 min (*S*, *R*) (major), 6.906 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>27</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 303.2067, found: 303.2070.



**Ethyl** (4a*S*,9b*R*)-8-methoxy-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6d): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.7 mg, 93% yield, 92% ee,  $[\alpha]_D^{25}$  + 42.7 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 6.76 (s, 1H), 6.62 (dt, *J* = 14.0, 5.3 Hz, 2H), 4.20 – 4.07 (m, 2H), 3.99 – 3.94 (m, 1H), 3.93 – 3.76 (m, 1H), 3.74 (s, 3H), 3.66 – 3.09 (m, 5H), 1.88 (ddd, *J* = 18.6, 9.3, 4.5 Hz, 1H), 1.75 (brs, 1H), 1.26 (brs, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.8, 153.8, 144.6, 131.8, 113.0, 111.2, 110.8, 61.4, 58.0, 56.1, 44.0, 41.4, 40.0, 28.2, 14.9. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 8.012 min (*R*, *S*) (minor), 21.165 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 277.1547, found: 277.1548.



Ethyl (4a*S*,9b*R*)-8-(benzyloxy)-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2carboxylate (6e): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 34.8 mg, 99% yield, 94% ee,  $[\alpha]_D^{25} + 28.5$  (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.42 (d, J = 7.3 Hz, 2H), 7.37 (t, J = 7.5 Hz, 2H), 7.31 (t, J = 7.3 Hz, 1H), 6.84 (d, J = 2.3 Hz, 1H), 6.70 (dd, J = 8.4, 2.5 Hz, 1H), 6.60 (d, J = 8.4 Hz, 1H), 4.98 (s, 2H), 4.18 – 4.09 (m, 2H), 3.98 – 3.94 (m, 1H), 3.84 (d, J = 94.5 Hz, 1H), 3.67 – 3.10 (m, 5H), 1.88 (ddd, J = 13.8, 9.3, 4.6 Hz, 1H), 1.76 (dd, J = 9.2, 4.2 Hz, 1H), 1.31 – 1.25 (m, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.7, 152.9, 144.9, 137.6, 131.9, 128.6, 127.9, 127.6, 114.2, 112.3, 110.7, 71.1, 61.4, 58.0, 44.0, 41.4, 39.9, 28.1, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>/</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda = 210$  nm, t<sub>R</sub>: 8.050 min (*R*, *S*) (minor), 19.245 min (*S*, *R*) (major). HRMS (ESI/ion trap) m/z: [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>25</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 353.1860, found: 353.1860.



**Ethyl** (4a*S*,9b*R*)-8-fluoro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6f): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.8 mg, 97% yield, 90% ee,  $[\alpha]_D^{25}$  + 61.4 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.85 (dd, *J* = 8.2, 2.5 Hz, 1H), 6.74 (td, *J* = 8.9, 2.6 Hz, 1H), 6.56 (dd, *J* = 8.4, 4.3 Hz, 1H), 4.18 – 4.09 (m, 2H), 3.98 (dt, *J* = 10.1, 5.0 Hz, 1H), 3.80 (brd, *J* = 63.1 Hz, 1H), 3.69 – 3.13 (m, 5H), 1.88 (ddd, *J* = 13.8, 6.8, 4.8 Hz, 1H), 1.80 – 1.66 (m, 1H), 1.27 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 157.0 (d, *J* = 235.7 Hz), 155.6, 146.8, 131.7 (d, *J* = 34.0 Hz), 114.0 (d, *J* = 23.2 Hz), 111.6 (d, *J* = 23.7 Hz), 110.2 (d, *J* = 7.5 Hz), 61.3, 58.0, 43.6, 41.2, 39.8, 27.9, 14.7. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -125.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>i</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 6.487 min (*R*, *S*) (minor), 9.634 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub><sup>+</sup> : 265.1347, found: 265.1349.



Ethyl (4a*S*,9b*R*)-8-chloro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6g): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 27.4 mg, 98% yield, 86% ee,  $[\alpha]_D^{25}$  + 30.4 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.07 (s, 1H), 6.99 (dd, *J* = 8.2, 2.1 Hz, 1H), 6.55 (d, *J* = 8.3 Hz, 1H), 4.20 – 4.06 (m, 2H), 3.98 (dt, *J* = 10.1, 5.0 Hz, 1H), 3.94 – 3.65 (m, 2H), 3.62 – 3.07 (m, 4H), 1.94 – 1.83 (m, 1H), 1.72 (brs, 1H), 1.26 (t, *J* = 6.3 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.7, 149.5, 131.8, 127.9, 124.6, 123.6, 110.8, 61.5, 57.9, 43.6, 41.2, 39.8, 28.0, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 6.739 min (*R*, *S*) (minor), 12.626 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>ClN<sub>2</sub>O<sub>2</sub><sup>+</sup> : 281.1051, found: 281.1053.



Ethyl (4a*S*,9b*R*)-8-bromo-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6h): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 31.4 mg, 98% yield, 84% ee,  $[\alpha]_D^{25}$  + 19.8 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.20 (d, *J* = 1.8 Hz, 1H), 7.13 (dd, *J* = 8.2, 2.0 Hz, 1H), 6.52 (d, *J* = 8.2 Hz, 1H), 4.19 – 4.07 (m, 2H), 3.97 (dt, *J* = 10.1, 5.0 Hz, 1H), 3.92 – 3.65 (m, 2H), 3.62 – 3.13 (m, 4H), 1.88 (td, *J* = 13.9, 4.8 Hz, 1H), 1.70 (s, 1H), 1.27 (brs, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.7, 149.9, 132.3, 130.7, 127.3, 111.3, 110.5, 61.5, 57.8, 43.6, 41.1, 39.8, 28.0, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 6.989 min (*R*, *S*) (minor), 13.105 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>1</sub>4H<sub>18</sub>BrN<sub>2</sub>O<sub>2</sub><sup>+</sup> : 325.0546, found: 325.0548.



**Ethyl (4a***S*,9b*R*)-8-trifluoromethyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6i): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 30.4 mg, 97% yield, 66% ee,  $[\alpha]_D^{25}$  + 18.5 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 7.33 (s, 1H), 7.30 (d, *J* = 8.2 Hz, 1H), 6.64 (d, *J* = 8.1 Hz, 1H), 4.16 – 4.08 (m, 2H), 4.08 – 4.00 (m, 2H), 3.98 – 3.11 (m, 5H), 1.92 (ddt, *J* = 13.9, 9.2, 4.7 Hz, 1H), 1.74 (s, 1H), 1.31 – 1.19 (m, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.6, 153.7, 129.9, 126.0 (q, *J* = 3.4 Hz), 123.1 (q, *J* = 270.6 Hz), 121.4, 120.65 (d, *J* = 32.2 Hz), 108.8, 61.4, 57.6, 43.2, 40.7, 39.7, 27.8, 14.6. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -60.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>1</sup>PrOH = 85:15, flow rate = 0.8 mL/min,  $\lambda$  = 254 nm, t<sub>*R*</sub>: 8.52 min (*R*, *S*) (minor), 10.46 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m*/*z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>18</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 315.1315, found: 315.1311.



Ethyl (4a*S*,9b*R*)-7-methyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6j): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.6 mg, 98% yield, 92% ee,  $[\alpha]_D^{23}$  + 56.9 (*c* 1.0, CHCl<sub>3</sub>).<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.00 (d, *J* = 7.4 Hz, 1H), 6.55 (d, *J* = 7.4 Hz, 1H), 6.50 (s, 1H), 4.13 (p, *J* = 7.2 Hz, 2H), 4.07 – 3.63 (m, 3H), 3.57 (dt, *J* = 11.4, 5.1 Hz, 1H), 3.42 (d, *J* = 12.7 Hz, 1H), 3.25 (t, *J* = 33.4 Hz, 2H), 2.26 (s, 3H), 1.89 (ddt, *J* = 14.4, 9.6, 4.6 Hz, 1H), 1.76 (dt, *J* = 14.4, 5.2 Hz, 1H), 1.31 – 1.23 (m, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$ 

155.8, 151.1, 138.1, 127.4, 124.0, 119.9, 111.0, 61.3, 57.7, 44.1, 39.9, 28.1, 21.7, 14.9. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 7.836 min (*S*, *R*) (major), 9.540 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 261.1598, found: 261.1596.



Ethyl (4a*S*,9b*R*)-7-methoxy-1,3,4,4a,5,9b-hexahydro-*2H*-pyrido[4,3-*b*]indole-2-carboxylate (6k): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 26.2 mg, 95% yield, 90% ee,  $[\alpha]_D^{25}$  + 79.4 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.01 (d, *J* = 7.7 Hz, 1H), 6.34 – 6.17 (m, 2H), 4.12 (dtd, *J* = 10.3, 7.0, 3.6 Hz, 2H), 3.98 (dd, *J* = 11.5, 4.9 Hz, 1H), 3.93 – 3.65 (m, 2H), 3.65 – 3.51 (m, 1H), 3.47 – 3.35 (m, 1H), 3.31 – 3.10 (m, 2H), 1.94 – 1.84 (m, 1H), 1.75 (ddd, *J* = 14.3, 9.4, 5.2 Hz, 1H), 1.26 (t, *J* = 6.3 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  160.4, 155.7, 152.2, 124.5, 122.7, 103.7, 96.9, 61.3, 57.9, 55.4, 44.1, 40.2, 39.8, 28.1, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane//PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 8.288 min (*S*, *R*) (major), 19.862 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 277.1547, found: 277.1548.



Ethyl (4a*S*,9b*R*)-7-fluoro-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6l): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.9 mg, 98% yield, 82% ee,  $[\alpha]_D^{25}$  + 3.1 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 7.03 – 6.97 (m, 1H), 6.40 – 6.36 (m, 1H), 6.33 (dd, *J* = 9.7, 2.2 Hz, 1H), 4.16 – 4.08 (m, 2H), 4.01 (dd, *J* = 11.9, 4.9 Hz, 1H), 3.89 – 3.68 (m, 2H), 3.58 – 3.52 (m, 1H), 3.44 – 3.18 (m, 3H), 1.89 (ddt, *J* = 13.8, 9.1, 4.3 Hz, 1H), 1.74 (ddd, *J* = 14.4, 9.3, 5.3 Hz, 1H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 163.6 (d, *J* = 241.9 Hz), 155.8, 152.4 (d, *J* = 11.6 Hz), 125.7 (d, *J* = 34.2 Hz), 124.8, 105.1 (d, *J* = 16.3 Hz), 97.7 (d, *J* = 26.4 Hz), 61.4, 58.1, 43.9, 40.2, 39.7, 28.0, 14.8. <sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) δ -114.9. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>/</sup>PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 7.574 min (*S*, *R*) (major), 9.122 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub><sup>+</sup> : 265.1347, found: 265.1345.



Ethyl (4aS,9b*R*)-9-methyl-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6m): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.5 mg, 98% yield, 91% ee,  $[\alpha]_D^{22}$  + 54.9 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.96 (t, *J* = 7.7 Hz, 1H), 6.55 (d, *J* = 7.6 Hz, 1H), 6.52 (d, *J* = 7.7 Hz, 1H), 4.16 (q, *J* = 7.1 Hz, 2H), 4.09 – 3.62 (m, 4H), 3.26 (d, *J* = 13.9 Hz, 1H), 2.73 (brs, 1H), 2.28 (s, 3H), 1.96 (s, 1H), 1.91 – 1.80 (m, 1H), 1.28 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  155.7, 150.9, 134.4, 129.7, 128.0, 120.6, 107.6, 61.3, 57.7, 43.7, 39.6, 27.9, 18.4, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 9.997 min (*S*, *R*) (major), 11.021 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 261.1598, found: 261.1596.



Ethyl (4a*S*,9b*R*)-9-methoxy-1,3,4,4a,5,9b-hexahydro-2*H*-pyrido[4,3-*b*]indole-2-carboxylate (6n): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 27.3 mg, 99% yield, 95% ee,  $[\alpha]_D^{25}$  + 29.1 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.01 (t, *J* = 8.0 Hz, 1H), 6.30 (t, *J* = 8.0 Hz, 2H), 4.27 – 3.96 (m, 4H), 3.79 (s, 3H), 3.78 – 3.66 (m, 2H), 3.40 (s, 1H), 3.37 – 3.27 (m, 1H), 3.18 – 2.76 (m, 1H), 1.95 (dd, *J* = 14.7, 9.6 Hz, 1H), 1.88 – 1.80 (m, 1H), 1.26 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 157.0, 155.9, 152.6, 129.4, 117.3, 103.4, 101.9, 61.2, 57.4, 55.3, 43.1, 39.6, 38.8, 27.8, 14.9. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>i</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$ = 210 nm, t<sub>*R*</sub>: 10.693 min (*R*, *S*) (minor), 12.109 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub><sup>+</sup> : 277.1547, found: 277.1548.



**Ethyl (4aS,9bR)-7,9-dimethyl-1,3,4,4a,5,9b-hexahydro-***2H***-pyrido[4,3-***b***]<b>indole-2-carboxylate** (**6o**): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 26.1 mg, 95% yield, 89% ee,  $[\alpha]_D^{25}$  + 73.9 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.39 (s, 1H), 6.36 (s, 1H), 4.15 (q, *J* = 7.1 Hz, 2H), 4.10 (brs, 1H), 3.96 (dt, *J* = 7.0, 3.6 Hz, 1H), 3.93 – 3.60 (m, 2H), 3.24 (dd, *J* = 25.8, 11.8 Hz, 2H), 2.72 (brs, 1H), 2.23 (d, *J* = 1.4 Hz, 6H), 2.02 – 1.89 (m, 1H), 1.89 – 1.78 (m, 1H), 1.27 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.6, 151.1, 137.9, 134.0, 126.9, 121.3, 108.3, 61.2, 57.8, 43.8, 39.5, 39.0, 27.9, 21.5, 18.2, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/<sup>/</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 5.654 min (*S*, *R*) (major), 6.425 min (*R*, *S*) (minor). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>16</sub>H<sub>23</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 275.1754, found: 275.1755.



Ethyl (4a*S*,9b*R*)-6-fluoro-1,3,4,4a,5,9b-hexahydro-*2H*-pyrido[4,3-*b*]indole-2-carboxylate (6p): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 25.6 mg, 97% yield, 77% ee,  $[\alpha]_D^{25}$  + 53.9 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 6.90 (d, *J* = 7.3 Hz, 1H), 6.82 (dd, *J* = 9.7, 8.4 Hz, 1H), 6.65 (ddd, *J* = 8.1, 7.5, 4.6 Hz, 1H), 4.11 (ddq, *J* = 14.3, 7.2, 3.5 Hz, 2H), 4.01 (dt, *J* = 6.8, 4.9 Hz, 1H), 3.97 – 3.66 (m, 2H), 3.60 – 3.52 (m, 1H), 3.48 – 3.38 (m, 1H), 3.36 – 3.13 (m, 2H), 1.90 (ddt, *J* = 14.1, 9.3, 4.6 Hz, 1H), 1.77 (dt, *J* = 14.2, 4.6 Hz, 1H), 1.24 (t, *J* = 6.7 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 155.6, 149.4 (d, *J* = 240.4 Hz), 137.7 (d, *J* = 12.9 Hz), 133.9 (d, *J* = 7.5 Hz), 119.6 (d, *J* = 23.1 Hz), 114.7 (d, *J* = 17.5 Hz), 61.3, 58.2, 43.7, 41.3, 39.7, 27.9, 14.8. <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>) δ -135.5. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak AD-3 column (0.46 x 25 cm), Hexane/<sup>†</sup>PrOH = 85:15, flow rate = 1.0 mL/min, *λ* = 210 nm, t<sub>*R*</sub>: 6.800 min (*R*, *S*) (minor), 7.666 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m/z*: [M + H]<sup>+</sup> calcd for C<sub>14</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>2</sub><sup>+</sup> : 265.1347, found: 265.1349.



**Ethyl (5aS,10b***R***)-3,4,5,5a,6,10b-hexahydroazepino**[**4**,3-*b*]indole-2(*1H*)-carboxylate (**6**q): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 16.9 mg, 65% yield, 98% ee,  $[\alpha]_D^{25}$  + 81.2 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.15 – 6.97 (m, 2H), 6.70 (t, *J* = 7.4 Hz, 1H), 6.57 (d, *J* = 7.9 Hz, 1H), 4.18 (dt, *J* = 22.1, 11.0 Hz, 3H), 4.02 (tt, *J* = 9.8, 4.8 Hz, 2H), 3.88 – 3.51 (m, 2H), 3.16 – 2.98 (m, 1H), 2.96 – 2.80 (m, 1H), 2.02 – 1.90 (m, 2H), 1.87 – 1.78 (m, 1H), 1.70 – 1.58 (m, 1H), 1.33 – 1.26 (m, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.9, 150.7, 129.4, 128.1, 124.3, 118.4, 108.9, 61.3, 49.1, 48.1, 45.5, 31.7, 25.4, 14.8. **rotamer :** <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 155.9, 150.7, 129.6, 127.9, 124.6, 118.5, 108.8, 62.5, 49.0, 48.4, 44.7, 31.9, 25.5, 14.8. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak AD-3 column (0.46 x 25 cm), Hexane/<sup>*i*</sup>PrOH = 80:20, flow rate = 1.0 mL/min,  $\lambda$  = 210 nm, t<sub>*R*</sub>: 6.599 min (*R*, *S*) (minor), 12.318 min (*S*, *R*) (major). HRMS (ESI/ion trap) *m*/*z*: [M + H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub><sup>+</sup> : 261.1598, found: 261.1597.

Ethyl (4aS,9aR)-1,3,4,4a,9,9a-hexahydro-2H-pyrido[3,4-b]indole-2-carboxylate (6r): purified by flash chromatography (silica, petroleum ether/ethyl acetate = 2/1, v/v), oily solid, 23.6 mg, 96% yield, 75% ee,  $[\alpha]_D^{25}$ -60.4 (*c* 1.0, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.04 (dd, *J* = 14.8, 7.4 Hz, 2H), 6.73 (t, *J* = 7.3 Hz, 1H), 6.63 (d, *J* = 7.7 Hz, 1H), 4.12 (dd, *J* = 13.8, 6.8 Hz, 2H), 4.00 – 3.81 (m, 1H), 3.66 – 3.47 (m, 1H), 2.00 (td, *J* = 12.3, 5.9 Hz, 1H), 1.85 (s, 1H), 1.32 – 1.13 (m, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.9, 150.3, 130.9, 127.6, 123.5, 118.7, 109.6, 61.1, 57.2, 44.0, 40.8, 39.1, 26.0, 14.5. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/iPrOH = 80 : 20, flow rate = 1.0 mL/min,  $\lambda$  = 254 nm, t<sub>R</sub>: 10.68 min (*R*, *S*) (minor), 11.92 min (*S*, *R*) (major).

HRMS (ESI/ion trap) m/z:  $[M + H]^+$  calcd for  $C_{14}H_{19}N_2O_2^+$ : 247.1411, found: 247.1412.

#### **VII. Gram Scale Reaction and High Turnover Reaction**



In the argon-filled glovebox, a solution of  $[Ir(COD)Cl]_2$  (6.7 mg, 0.01 mmol) and ZhaoPhos (18.2 mg, 0.021 mmol) in 2.0 mL anhydrous solvent was stirred at room temperature for 20 min. A specified volume of the resulting solution (100 µL, 1 mol% Ir-Zhaophos catalyst) was transferred by syringe to a score-break ampule charged with substrate (5 mmol, 1.22 g) in 40 mL dichloromethane and TsOH (904 mg, 5.25 mmol, 1.05 eq.) as acid additive. The ampule was placed into an autoclave, which was then charged with hydrogen gas (65 atm). The autoclave was stirred at room temperature for 48 h. After release of H<sub>2</sub>, saturated sodium bicarbonate solution and dichloromethane was added and the mixture was stirred for 10 min. The organic layer was dried with anhydrous Na<sub>2</sub>SO<sub>4</sub>. After removal of solvent, the crude product was analysed by <sup>1</sup>H NMR to determine the conversion. Purification was performed by silica gel column chromatography, eluted with Petrol ether / EtOAc, to give the desired product **6a** (90% yield, 91% ee).

#### VIII. Synthetic Application from 6a to Lumateperone Intermediate



Synthesis of compound 8: In argon atmosphere, to a solution of 2a (739 mg, 3 mmol) in acetone (15 mL) were added 2-chloro-N-methylacetamide (387 mg, 3.6 mmol), KI (250 mg, 1.5 mmol), and K<sub>2</sub>CO<sub>3</sub> (620 mg, 4.5 mmol). The reaction mixture was heated at 70 °C for 24 h, cooled to room temperature, and then the solid was removed by filtration through celite, and the filtrate was concentrated under vacuum. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>, washed by saturated NaCl, and then was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the filtrate was concentrated under vacuum to give crude product 7. Then to a solution the crude product 7 in CH<sub>2</sub>Cl<sub>2</sub> (15 mL), monopyridin-1-ium tribromide (2.11 g, 6.6 mmol) was added in portions, and the reaction was completed after 1h traced by TLC. Then the reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL), quenched by aq. Na<sub>2</sub>CO<sub>3</sub>, separated the organic phase, washed with aq. Na<sub>2</sub>SO<sub>3</sub>, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, then the filtrate was concentrated under vacuum to give crude residue. The residue was purified by silica gel flash column chromatography eluting with a gradient of 30-50% ethyl acetate in petrol ether to give the title compound 8 (1.21 g, 85% yield after two steps, 93% ee). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 7.38 (s, 1H), 7.16 (s, 1H), 6.81 (brs, 1H), 4.32 – 4.18 (m, 1H), 4.14 – 3.96 (m, 2H), 3.89 – 3.12 (m, 7H), 2.87 (d, J = 4.9 Hz, 3H), 1.96 – 1.85 (m, 1H), 1.83 – 1.72 (m, 1H), 1.21 (t, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) & 171.1, 155.7, 147.8, 136.1, 135.4, 126.7, 113.0, 105.1, 65.6, 61.7, 54.6, 43.5, 41.1, 39.9, 26.2, 25.0, 14.7. The enantiomeric excess was determined by HPLC analysis on Daicel Chiralpak OD-3 column (0.46 x 25 cm), Hexane/PrOH = 85:15, flow rate = 1.0 mL/min,  $\lambda$ = 210 nm,  $t_R$ : 7.88 min (*R*, *S*) (minor), 8.49 min (*S*, *R*) (major). HRMS (ESI/ion trap) m/z:  $[M + H]^+$ calcd for C<sub>17</sub>H<sub>22</sub>Br<sub>2</sub>N<sub>3</sub>O<sub>3</sub><sup>+</sup> : 474.0022, found: 474.0020.

## IX. NMR Spectra of Compounds 5a-5r, 6a-6r, 8





## <sup>1</sup>H NMR (600 MHz, $d^6$ -DMSO) of compound **5b**



<sup>1</sup>H NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **5b** 



## $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) of compound 5c



)0 110 100 f1 (ppm) ć 

## <sup>1</sup>H NMR (600 MHz, $d^6$ -DMSO) of compound **5d**



<sup>1</sup>H NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **5d** 



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **5**e



## <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of compound **5**e





#### $^1\text{H}$ NMR (600 MHz, CDCl\_3) of compound $\mathbf{5f}$



## <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO) of compound **5**f









## $^{13}\text{C}$ NMR (101 MHz, $d^{6}\text{-}\text{DMSO})$ of compound $\mathbf{5g}$



## <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **5h**



## $^{13}$ C NMR (101 MHz, $d^6$ -DMSO) of compound **5h**



<sup>19</sup>F NMR (565 MHz, *d*<sup>6</sup>-DMSO) of compound **5**i



<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **S5j** 



<sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of compound **S5j** 



## <sup>1</sup>H NMR (400 MHz, *d*<sup>6</sup>-DMSO) of compound **5**j



 $^{13}$ C NMR (151 MHz,  $d^6$ -DMSO) of compound **5**j


#### $^1\text{H}$ NMR (600 MHz, CDCl<sub>3</sub>) of compound 5k



<sup>13</sup>C NMR (101 MHz,  $d^6$ -DMSO) of compound **5**k



## <sup>1</sup>H NMR (600 MHz, d<sup>6</sup>-DMSO) of compound S5I



<sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **S5**I





#### <sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **5**l



 $^{19}\mathrm{F}$  NMR (565 MHz,  $d^6\text{-}\mathrm{DMSO})$  of compound **51** 



-74 -76 -78 -80 -82 -84 -86 -88 -90 -92 -94 -96 -98 -102 -106 -110 -114 -118 -122 -126 -130 f1 (ppm)

#### <sup>1</sup>H NMR (600 MHz, $d^6$ -DMSO) of compound **S5m**



<sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **S5m** 



#### <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) of compound **5m**



<sup>13</sup>C NMR (151 MHz, *d*<sup>6</sup>-DMSO) of compound **5m** 



#### <sup>1</sup>H NMR (600 MHz, *d*<sup>6</sup>-DMSO) of compound **5n**



<sup>13</sup>C NMR (151 MHz,  $d^6$ -DMSO) of compound **5n** 



#### <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **50**



#### <sup>13</sup>C NMR (101 MHz, *d*<sup>6</sup>-DMSO) of compound **50**



#### $^1\text{H}$ NMR (600 MHz, CDCl<sub>3</sub>) of compound 5p









 $^{13}\text{C}$  NMR (151 MHz, CD<sub>2</sub>Cl<sub>2</sub>) of compound 5q



# <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of compound **5r**



#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound 6a



#### $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) of compound 6b



#### <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of compound **6b**





## $^1\text{H}$ NMR (400 MHz, CDCl<sub>3</sub>) of compound 6c



#### <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of compound **6c**



 $^1\text{H}$  NMR (600 MHz, CDCl<sub>3</sub>) of compound 6d



#### <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of compound **6d**



#### $^1\mathrm{H}$ NMR (600 MHz, CDCl<sub>3</sub>) of compound $\mathbf{6e}$



<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of compound **6e** 



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound 6f



## $^{13}\mathrm{C}$ NMR (101 MHz, CDCl<sub>3</sub>) of compound $\mathbf{6f}$





OEt

-70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -22 f1 (ppm)



-60

-30 -40 -50

20

10 0 -10 -20

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound **6g**



## $^{13}\text{C}$ NMR (151 MHz, CDCl<sub>3</sub>) of compound $\mathbf{6g}$



#### $^1\mathrm{H}$ NMR (400 MHz, CDCl<sub>3</sub>) of compound $\mathbf{6h}$



#### <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **6i**





<sup>19</sup>F NMR (565 MHz, CDCl<sub>3</sub>) of compound 6i



----60.82







<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound **6k** 



<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of compound **6k** 



## $^{13}\mathrm{C}$ NMR (151 MHz, CDCl\_3) of compound **61**

10

0 -10 -20 -30







-60

-40 -50

-70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1(ppm)

#### <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound 6m



## <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of compound 6m



#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound **6n**



<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) of compound **6n** 





#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound **60**



#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) of compound **6p**





 $^1\text{H}$  NMR (400 MHz, CDCl<sub>3</sub>) of compound 6q



## $^{13}\text{C}$ NMR (151 MHz, CDCl<sub>3</sub>) of compound 6q



## $^{13}\mathrm{C}$ NMR (151 MHz, CDCl<sub>3</sub>) of compound 6r



#### <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of compound **8**





X. HPLC Spectra of *cis*-(±)-6a-6r, (±)-8 and Chiral Compounds 6a-6r, 8



Joi ceu by	•	Dignar	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

#### Signal 1: DAD1 A, Sig=210,4 Ref=360,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	7.090	MM	0.1749	126.10619	12.01711	3.2044
2	9.004	BB	0.2091	3809.24194	275.90793	96.7956






Peak	RetTime	Туре	Width	Area	Height	Area	
#	[min]		[min]	[mAU*s]	[mAU]	%	
1	8.012	BB	0.2029	153.82138	11.15668	3.8424	
2	21.165	BB	0.4908	3849.46509	119.82839	96.1576	



Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	a Dilution	Factor with	ISTDs

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	8.050	FM	0.2072	292.89197	23.56196	3.0967
2	19.245	BB	0.4986	9165.36328	280.94107	96.9033









Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.989	мм	0.1448	244.62207	28.15826	7.9510
2	13.105	BB	0.2801	2832.00342	155.73909	92.0490



Use Multiplier & Dilution Factor with ISTDs

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	8.484	BB	0.1844	560.01794	46.54556	17.0085
2	10.385	BB	0.2450	2732.56836	170.49403	82.9915







Area Percent Report

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	7.528	BB	0.1470	2886.17676	298.04651	51.0999
2	9.093	BB	0.1890	2761.92651	222.24333	48.9001





Area Percent Report

Area refeete kepore

Sorted By		:	Sig	nal	
Multiplier		:	1.00	900	
Dilution		:	1.00	900	
Use Multiplier	&	Dilution	Factor	with	ISTDs

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	10.090	BB	0.2194	1379.77441	96.15178	49.7651
2	11.088	BB	0.2316	1392.80249	91.51515	50.2349













Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	6.599	BB	0.1877	2880.95923	215.87976	50.1722
2	12.318	BB	0.5760	2861.18213	68.96182	49.8278





Area Percent Report


Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

Signal 1: DAD1 B, Sig=254,4 Ref=360,100

Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	%
1	10.684	BB	0.2427	745.30450	47.60540	49.9376
2	11.925	BB	0.2435	747.16589	47.50566	50.0624



Area Percent Report

-----

Sorted By	:	Signal	
Multiplier	:	1.0000	
Dilution	:	1.0000	
Use Multiplier &	Dilution	Factor with	ISTDs

Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.626	BB	0.2370	157.53870	10.27008	12.3351
2	11.818	BB	0.2404	1119.62085	72.39935	87.6649



# XI. X-Ray Data of Compound 6a (CCDC 2108430)

checkCIF/PLATON report Structure factors have been supplied for datablock(s) 1 THIS REPORT IS FOR GUIDANCE ONLY. IF USED AS PART OF A REVIEW PROCEDURE FOR PUBLICATION, IT SHOULD NOT REPLACE THE EXPERTISE OF AN EXPERIENCED CRYSTALLOGRAPHIC REFEREE. No syntax errors found. CIF dictionary Interpreting this report Datablock: 1 Bond precision: C-C = 0.0028 A Wavelength=1.54178 Cell: a=8.5260(7) b=7.7633(6) c=9.5887(8) alpha=90 beta=98.910(3) gamma=90 Temperature: 100 K Calculated Reported Volume 627.02(9) 627.02(9) Space group P 21 P 1 21 1 Hall group P 2yb P 2yb Moiety formula C14 H18 N2 O2 C14 H18 N2 O2 Sum formula C14 H18 N2 O2 C14 H18 N2 O2 Mr 246.30 246.30 Dx,g cm-3 1.305 1.305 Z 2 2 Mu (mm-1) 0.710 0.710 F000 264.0 264.0 F000' 264.78 h,k,lmax 10,9,11 10,9,11 Nref 2506[1350] 2463 Tmin, Tmax 0.912, 0.931 0.469, 0.754 Tmin' 0.912 Correction method= # Reported T Limits: Tmin=0.469 Tmax=0.754 AbsCorr = MULTI-SCAN Data completeness= 1.82/0.98 Theta(max)= 72.625 R(reflections)= 0.0331(2445) wR2(reflections)= 0.0859(2463) S = 1.069 Npar= 168 The following ALERTS were generated. Each ALERT has the format test-name ALERT alert-type alert-level. Click on the hyperlinks for more details of the test. Alert level G PLAT791 ALERT 4 G Model has Chirality at C7 (Sohnke SpGr) R Verify PLAT791 ALERT 4 G Model has Chirality at C8 (Sohnke SpGr) S Verify PLAT912 ALERT 4 G Missing # of FCF Reflections Above STh/L= 0.600 11 Note PLAT961 ALERT 5 G Dataset Contains no Negative Intensities ...... Please Check

PLAT978\_ALERT\_2\_G Number C-C Bonds with Positive Residual Density. 6 Info 0 ALERT level A = Most likely a serious problem - resolve or explain 0 ALERT level B = A potentially serious problem, consider carefully
0 ALERT level C = Check. Ensure it is not caused by an omission or oversight 5 ALERT level G = General information/check it is not something unexpected 0 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
1 ALERT type 2 Indicator that the structure model may be wrong or deficient
0 ALERT type 3 Indicator that the structure quality may be low
3 ALERT type 4 Improvement, methodology, query or suggestion
1 ALERT type 5 Informative message, check

## PLATON version of 13/07/2021; check.def file version of 13/07/2021

Datablock 1 - ellipsoid plot



# **XII. References**

- <sup>1</sup> J. B. Hester, A. D. Rudzik, P. F. VonVoigtlander, J. Med. Chem. 1980, 23, 643-647.
- <sup>2</sup> Bridoux, A.; Goossens, L.; Houssin, R.; Héanichart, J.-P. J. Hetero. Chem., 2006, 43, 571-578
- <sup>3</sup> C. A. Harbert, J. J. Plattner, W. M. Welch, A. Weissman, B. K. Koe, *J. Med. Chem.* **1980**, *23*, 635–643.
- <sup>4</sup> N. Khorana, A. Purohit, K. Herrick-Davis, M. Teitler, R. A. Glennon, *Bioorg. Med. Chem.* 2003, *11*, 717–722.
- <sup>5</sup> L. E. J. Kennis, J. C. Mertens Tetrahydro-γ-carbolines with serotonin receptor affinity, WO 9912926 A1
- <sup>6</sup> C. Sheng, X. Che, W. Wang, S. Wang, Y. Cao, J. Yao, Z. Miao, W. Zhang, *Eur. J. Med. Chem.* **2011**, *46*, 1706–1712.
- <sup>7</sup> S. Svorad, P. Frantisek, B. Viktor *et al.* Preparation of pyridoindole derivatives with antioxidant properties, Slovakia (2010), SK 287506 B6 Dec 07, 2010.
- <sup>8</sup> F. Kudzma, Synthesis **2003**, 11, 1661 1666
- <sup>9</sup> M. de Candia, G. Zaetta, N. Denora, D. Tricarico, M. Majellaro, S. Cellamare, C. D. Altomare, *Eur. J. Med. Chem.* 2017, 125, 288–298.
- <sup>10.</sup> J. C. Tomesch *et al.* Process for preparation of heterocycle fused gamma-carbolines, PCT Int. Appl. (2008), WO 2008112280 A1 20080918
- <sup>10</sup> J. Ye, J. Wu, T. Lv, G. Wu, Y. Gao, H. Chen, *Angew. Chem. Int. Ed.* **2017**, *56*, 14968–14972.