Electronic Supporting Information

Solvent-free dynamic nuclear polarization enhancements in organically modified mesoporous silica

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Table S1 – Weights	of Fmoc-Gly-OH	and 3-Carboxy-PROXYL pr	recursors used in the
synthesis. The overall	mass of the final	materials (yield mass) is also g	jiven.
Sample	Weigh	Weigh	Vield

Sample	Weigh	Weigh	Yield
	Fmoc-Gly-OH	3-Carboxy-PROXYL	[mg]
	[mg]	[mg]	-
99.5Fmoc/0.5Prx	79	0.3	54
95Fmoc/5Prx	75	2.5	59
90Fmoc/10Prx	71	5	58
75Fmoc/25Prx	59	12	50
50Fmoc/50Prx	40	25	64

Table S2: Median pore diameters calculated by the BJH (left) and NLDFT (right) method of the prepared SBA-15 with APTES linker and functionalized mesoporous silica materials as obtained from BET analysis.

Sample	Pore diameter	Pore diameter
	by BJH	by NLDFT
	/ nm	/ nm
SBA-15 with APTES	5.5	7.9
99.5Fmoc/0.5Prx	5.3	6.6
95Fmoc/5Prx	5.3	7.0
90Fmoc/10Prx	5.4	6.6
75Fmoc/25Prx	5.3	6.6
50Fmoc/50Prx	5.2	6.6

Table S3. Pore volume (Gurvich at $p/p^0 0.95$), specific surface area (BET), nitrogen loading and carbon/nitrogen ratio of the support material determined by EA.

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Pore	Surface Area	Nitrogen loading		
Volume	(BET)	(EA)	C/N ratio	C/N ratio
(BET)	/ m ² ·g ⁻¹	/ mmol·g ⁻¹	(Theo.)	(Exp.)
/ cm ³ ·g ⁻¹	, iii g	/ minor g		(LAP.)
0.72	537	1.78 ± 0.3	3.00	3.13 ± 0.5
	Pore Volume (BET) / cm ³ ·g ⁻¹ 0.72	Pore VolumeSurface Area(BET) $/ cm^3 \cdot g^{-1}$ $/ m^2 \cdot g^{-1}$ 0.72537	Pore VolumeSurface Area (BET)Nitrogen loading (EA)(BET) $/ cm^3 \cdot g^{-1}$ $/ m^2 \cdot g^{-1}$ $/ mmol \cdot g^{-1}$ 0.72537 1.78 ± 0.3	Pore VolumeSurface Area (BET)Nitrogen loading (EA)C/N ratio (Theo.) (BET) $/ cm^3.g^{-1}$ $/ m^2.g^{-1}$ $/ mmol \cdot g^{-1}$ C/N ratio (Theo.) 0.72 537 1.78 ± 0.3 3.00

Table S4 - Elemental Analysis	Two measurements were	performed for eac	h sample
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Sample	C/N ratio	Carbon (%)	Hydrogen (%)	Nitrogen (%)
Amine functionalized SBA-15	2.835	8.018	3.11	2.829
Amine functionalized SBA-15	2.842	7.728	3.399	2.719
X 0.5	3.611	17.24	3.013	4.775
X 0.5	3.65	17.1	2.859	4.685
X 5	3.408	17.05	3.271	5.004
X 5	3.29	16.71	3.682	5.079
X 10	3.68	16.59	3.101	4.509
X 10	3.511	16.39	3.108	4.667
X 25	3.092	15.48	3.291	5.006
X 25	3.112	15.43	3.349	4.958
X 50	3.266	15.51	3.024	4.747
X 50	3.268	14.96	3.182	4.578



Figure S1 – ¹H spin-latice relaxation times (T_1 , μw off) and DNP build-up times (T_B , μw on) as a function of composition parameter *x* for the investigated samples. Values were obtained from unconstrained least-square fittings of saturation-recovery curves by using stretched exponential functions.



Figure S2 – Experimental (black curve, bottom) and simulated (red curve, top) cw EPR spectra for the sample with x = 10 mol%. The experimental spectrum was measured at 100 K. The simulation was performed using the EasySpin¹ package for the Matlab[®] software. The simulation uses the "pepper" function from EasySpin,¹ which considers a solid state powder. The system consists of an electron spin with rhombic g and ¹⁴N-hyperfine tensors (¹⁵N was also considered in natural abundance). The parameters were optimized by comparing experimental and simulated spectra, using a combination of least-square fitting and genetic algorithms and taking the parameters reported by Dhanasekaran at al.² as initial guess. The final parameters for the simulation are $g_{xx} = 2.0088$, $g_{yy} = 2.0057$, $g_{zz} = 2.0021$, $A_{xx} = 12 \pm 1$ MHz and $A_{yy} = 14 \pm 1$ MHz and $A_{zz} = 102 \pm 1$ MHz. The simulation also considers a Gauss-Lorentz (0.6-0.4) isotropic line broadening of 28 MHz (peak-to-peak).



Figure S3 – ²⁹Si MAS NMR spectra for the investigated samples (x = 0.5 to 50) and for a sample without carboxy-PROXYL radical (x = 0). The spectral line width increases for increasing radical concentration due to electron-nucleus hyperfine interaction.



Figure S4 – Top: ¹H–¹H MAS FSLG spectrum of a) mesoporous SBA-15 functionalized with APTES and Fmoc-Glycine (diamagnetic sample) and b) mesoporous SBA-15 functionalized with APTES, Fmoc-Glycine and Carboxy-PROXYL radicals (paramagnetic sample, x = 50). Skyline projections are also displayed. Bottom: comparison of the projections taken from the sum over the indirectly detected (δ_1) dimension for both spectra shown on top. 2D spectra were measured with a zfiltered ¹H-¹H correlation pulse sequence with Frequency Switched Lee–Goldberg decoupling during t_1 evolution.³ The experiments were performed using ¹H carrier frequency of 400 MHz, spinning rate of 10 kHz and z-filter delay of 1 μ s. In this experiment a MAS spectrum is detected in the direct domain (δ_2) while FSLG decoupled signal is achieved in the indirect domain (δ_1). Due to the FSLG decoupling the ¹H-¹H dipolar coupling is averaged out and the line broadening caused by ¹H spin-spin relaxation mechanism is suppressed. The result is a resolution enhancement in the $\delta 1$ projection and the detection of the δ^2 projections similar to the 1D spectrum. On the other hand, the electron-nucleus hyperfine interaction is not averaged out by FSLG decoupling leading to fast relaxation of areas containing the radicals. As a result, only components outside of the effective paramagnetic shell will survive seen as relative enhancement of the peak at 1.2 ppm in the projections, accompanied by a higher resolved δ_1 projections. This peak belongs most probably to mobile and isolated water molecules on the surface,⁴ far away from the radicals.



Figure S5 – ¹H magnetization as a function of the spin-lock time for samples x = 10 and x = 50, measured using a 90°-spinlock pulse sequence, nuclei are excited by a 90° pulse and then spin-locked by a continuous wave pulse (with a phase shift of $\pi/2$ relative to the excitation pulse) for a time τ .⁵ The experimental points are normalized to the extrapolated equilibrium magnetization. The evolution of the magnetization curves gives information about the ¹H spin lattice relaxation in the rotating frame (T₁^ρ). Solid curves are exponential fit to the data, using a stretched exponential function of the form $I = I_0 \exp\left(-\left(t/T_1^{\rho}\right)^{\beta}\right)$ (see parameter values in Table S5). Due to the distribution of distances between H species and the unpaired electrons in the sample, a continuous distribution of relaxation times is expected. Clearly, for the sample with x = 50 the effective ¹H spin relaxation is much faster than for the sample with x = 10. For this reason, ¹H-¹³C CP transfer could not be achieved for sample x = 50 (see main text).

Table S5: Fitting parameters to a stretched exponential decay function, $I = I_0 \exp\left(-\left(\frac{t}{T_1^{\rho}}\right)^{\beta}\right)$, adjusted on the data shown in Figure S5.

Sample	T_1^{ρ} (ms)	β
x = 10	2.1	0.5
x = 50	2×10 ⁻³	0.1

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