SUPPLEMENTAL INFORMATION

Destruction and reconstruction of UO₂²⁺ using gas-phase reactions

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Table S1. Product ion m/z values and compositions to accompany Figures 1 and 2 in the main text.

Table S2. Comparison of accurate m/z measurements, theoretical monoisotopic values, and molecule formulae for relevant ions in MSⁿ CID experiments initiated with [UO₂(O₂C-C₆F₂H₃)(CH₃CH₂OH)₂]⁺ (Figure 1 of the main text).

Figure S1. Product in spectra generated by isolation of $[UF_2(C_2H)]^+$ for reaction with H₂O: (a) 1 ms isolation time, (b) 10 ms isolation time and (c) 100 ms isolation time. Figure 1d shows the product ion spectrum generated by dissociation of the ion-molecule reaction product at m/z 293.

Figure S2. Product ion spectra generated by isolation of $[UF_2(OCH_3)]^+$ for reaction with CH₃OH: (a) 1 ms, (b) 10 ms and (c) 100 ms.

Figure S3. Product ion spectra generated by CID of $[UF(OCH_3)_2(CH_3OH)]^+$ at m/z 351: (a) CID (MS⁸ stage) of $[UF(OCH_3)_2(CH_3OH)]^+$, (b) CID (MS⁹ stage) of $[U(OCH_3)_3]^+$ at m/z 331 and (c) CID (MS¹⁰) of $[UO(OCH_3)_2]^+$ at m/z 316.

Figure S4. Product ion spectra generated by CID of species created by reaction of $[UF_2(C_2H)]^+$ with a mix of CH₃OH and CD₃OH: (a) isolation (MS⁸ stage) of $[UF_2(C_2H)]^+$ for 100 ms for reaction with CH₃OH/CD₃OH, (b) CID (MS⁹ stage) of $[UF(OCH_3)_2]^+$ at *m*/*z* 319, (c) CID (MS⁹ stage) of $[UF(OCH_3)(OCD_3)]^+$ at *m*/*z* 322 and (d) CID (MS⁹ stage) of $[UF(OCD_3)_2]^+$ at *m*/*z* 325. The spectrum shown in (e) shows the product ions generated by CID (MS¹⁰ stage) of $[UOF(CD_3)]^+$ at *m*/*z* 307.

Figure S5. Product ion spectra generated by CID of species created by reaction of $[UF_2(C_2H)]^+$ with CD₃OH: (a) CID (MS⁸ stage) of $[UF(OCD_3)_2(CD_3OH)]^+$ at m/z 360, (b) CID (MS⁹ stage) of $[U(OCD_3)_3]^+$ at m/z 340 and (c) CID (MS¹⁰ stage) of $[UOF(OCD_3)_2]^+$ at m/z 322.

Figure S6. Comparison of product ion spectra generated by isolation of species for reaction with H₂O. Figures S6(a)-S6(c) were generated by isolation of $[UO_2(F)]^+$ created by electrospray ionization of a solution of $[UO_2(F)_2]$. Figures S6(d)-S6(f) were created by isolation of the species at *m*/*z* 289 created by MSⁿ CID of $[UF(OCH_3)_2]^+$.

Figure S7. Comparison of product ion spectra generated by CID of species at m/z 301: (a) dissociation of $[UO_2(OCH_3)]^+$ created by solution of $UO_2(NO_3)_2$ in CH₃OH and (b) dissociation of ion at m/z 301 created by MSⁿ CID of $[UF(OCH_3)_2(CH_3OH)]^+$ that was generated by ion-molecule reactions of $[UF_2(C_2H)]^+$.

Scheme S1. Reaction pathways (dissociation and ion-molecule reaction) for species created by isolation and exposure of $[UF_2(C_2H)]^+$ to ca. 1 x 10⁻⁶ torr of CD₃OH directly admitted ion trap mass spectrometer.

Table S1.

(Figure 1)

	MS ⁿ CID of [UO ₂ (O ₂ C-C ₆ F ₂ H ₃)(CH ₃ CH ₂ OH) ₂] ⁺ (Figure					
m/z	Assigned composition					
519	$[UO_2(O_2C\text{-}C_6F_2H_3)(CH_3CH_2OH)_2]^+$					
491	$[UO_2(O_2C-C_6F_2H_3)(CH_3CH_2OH)(H_2O)]^+$					
473	$[UO_2(O_2C-C_6F_2H_3)(CH_3CH_2OH)]^+$					
445	$[UO_2(O_2C-C_6F_2H_3)(H_2O)]^+$					
427	$[UO_2(O_2C-C_6F_2H_3)(H_2O)]^+$					
401	$[UO_2(C_6F_2H_3)(H_2O)]^+$					
383	[UO ₂ (C ₆ F ₂ H ₃)] ⁺					
381	[UO ₂ (C ₆ FO ₄)]+					
355	$[UO(C_5F_2H_3)]^+$					
329	$[UO(C_3F_2H_2)]^+$					
327	$[U(C_4F_2H_3)]^+$					
301	$[UF_2(C_2H)]^+$					
293	[UF ₂ (OH)]*					
289	[UO ₂ (F)] ⁺					
287	[UO ₂ (OH)] ⁺					
276	[UF ₂]+					
273	[OUF]+					

Reaction of [UF₂(C₂H)]⁺ with CH₃OH (Figure 2)

m/z	Assigned composition
305	[UF(OH)(OCH ₃)] ⁺
307	$[UF_2(OCH_3)]^+$
319	[UF(OCH ₃) ₂] ⁺
325	$[U(C_2H)(OCH_3)_2]^+$
331	[U(OCH ₃) ₃]+
337	[UF(OH)(OCH ₃)(CH ₃ OH)] ⁺
339	[UF ₂ (OCH ₃)(CH ₃ OH)] ⁺
351	[UF(OCH ₃) ₂ (CH ₃ OH)] ⁺
363	[U(OCH ₃) ₃ (CH ₃ OH)] ⁺
369	[UF(OH)(OCH ₃)(CH ₃ OH) ₂] ⁺
371	$[UF_2(OCH_3)(CH_3OH)_2]^+$

CID of Ion-molecule Reaction Products (Figure 3)

304	[UOF(OCH ₃)] ⁺
292	[UOF ₂]+

Table S2.

Measured m/z	Monoisotopic m/z	Error (ppm calc)	Formula	Composition Assignment
519.1344	519.1345	0.1926	$C_{11}H_{15}F_2O_6U$	$[UO_2(O_2C-C_6F_2H_3)(CH_3CH_2OH)_2]^+$
491.1022	491.1031	1.8326	C ₉ H ₁₁ F ₂ O ₆ U	$[UO_2(O_2C-C_6F_2H_3)(CH_3CH_2OH)(H_2O)]^+$
473.0919	473.0925	1.2683	C ₉ H ₉ F ₂ O ₅ U	$[\mathrm{UO}_2(\mathrm{O}_2\mathrm{C}\text{-}\mathrm{C}_6\mathrm{F}_2\mathrm{H}_3)(\mathrm{CH}_3\mathrm{CH}_2\mathrm{OH})]^{+}$
463.0710	463.0718	1.7276	C7H7F2O6U	$[UO_2(O_2C-C_6F_2H_3)(H_2O)_2]^+$
445.0605	445.0613	1.7750	$C_7H_5F_2O_5U$	$[UO_2(O_2C-C_6F_2H_3)(H_2O)]^+$
427.0500	427.0507	1.6392	$C_7H_3F_2O_4U$	$[UO_2(O_2C-C_6F_2H_3)]^+$
401.0708	401.0715	1.6456	$C_6H_5F_2O_3U$	$[UO_2(C_6F_2H_3)(H_2O)]^+$
399.0752	399.0757	1.2529	C ₆ H ₆ FO ₄ U	$\left[\mathrm{UO}_{2}(\mathrm{C_{6}FOH_{4}})(\mathrm{H_{2}O})\right]^{+}$
383.0602	383.0608	1.5663	$C_6H_3F_2O_2U$	$[UO_2(C_6F_2H_3)]^+$
381.0647	381.0652	1.3121	C ₆ H ₄ FO ₃ U	$[UO_2(C_6FOH_4)]^+$
355.0655	355.0659	1.1266	C ₅ H ₃ F ₂ OU	$[UO(C_5H_3F_2)]^+$
353.0699	353.0703	1.1329	C ₅ H ₄ FO ₂ U	$[UO(C_5H_4OF)]^+$
329.0498	329.0503	1.4891	C ₃ HF ₂ OU	$[UO(C_3H_2F_2)]^*$
327.0706	327.0711	1.4370	$C_4H_3F_2U$	$[U(C_4H_3F_2)]^+$
301.0550	301.0554	1.4947	C_2HF_2U	$\left[\mathrm{UF}_{2}(\mathrm{C}_{2}\mathrm{H})\right]^{+}$
293.0499	293.0503	1.3650	HF ₂ OU	$[UF_2(OH)]^+$
292.0420	292.0425	1.7121	F ₂ OU	$[UF_2(O)]^+$
289.0385	289.0390	1.7299	FO ₂ U	$[UO_2(F)]^+$
287.0430	287.0434	1.2890	HO ₃ U	$[UO_2(OH)]^+$
276.0472	276.0476	1.4490	F_2U	$[UF_2]^+$
273.0437	273.0441	1.4650	FOU	[OUF] ⁺
270.0402	270.0406	1.3331	O ₂ U	$[UO_2]^+$



Figure S1.



Figure S2.



Figure S3.



Figure S4.



Figure S5.



Figure S6.



Figure S7.

Scheme S1.



EXPERIMENTAL METHODS

Sample Preparation

Methanol (CH₃OH), labeled methanol (CD₃OH), ethanol (CH₃CH₂OH) and 2,6difluorobenzoic acid were purchased from Millipore-Sigma (St. Louis, MO) and used as received. The sample used to prepare gas-phase $[UO_2(O_2C-C_6F_2H_3)]^+$ was created by combining 2-3 mg of U^{VI}O₃ (Strem Chemicals, Newburyport MA), corresponding to approximately 7 x 10⁻⁶ to 1 x 10⁻⁵ moles, with a 2-fold mole excess of propiolic acid (Sigma Aldrich, St. Louis MO) and 400 @L of deionized/distilled H₂O in a glass scintillation vial. The solutions were incubated on a hot plate at 70°C for 12 hours. *Caution: uranium oxide is radioactive (\alpha- and \gamma-emitter), and proper shielding, waste disposal and personal protective gear should be used when handling the material*. When cooled, 20 µL of the resulting solution was diluted with 800 µL of 90:10 (by volume) H₂O:CH₃CH₂OH and used without further work up as the spray solution for electrospray ionization (ESI).

ESI and CID experiments were performed on a ThermoScientific (San Jose, CA) LTQ-XL linear ion trap (LIT) mass spectrometer. Solutions for ESI were infused into the instrument using the incorporated syringe pump at a flow rate of 5 μ L/min. In the positive ion mode, the auto-tune routine within the LTQ Tune program was used to optimize the atmospheric pressure ionization stack settings for the instrument (lens voltages, quadrupole and octopole voltage offsets, etc.) to achieve maximum transmission of singly charged ions such as [UO₂(O₂C-C₆F₂H₃)(CH₃CH₂OH)₂]⁺ to the LIT. Helium was used as the bath/buffer gas to improve trapping efficiency and as the collision gas for CID experiments.

For multiple-stage (MSⁿ) CID experiments to probe fragmentation and ion-molecule reaction (IMR) pathways, precursor ions were isolated using a width of 1.0 to 1.5 mass to charge (m/z) units. The exact value was determined empirically to provide maximum ion intensity while ensuring isolation of a single isotopic peak. To probe CID behavior, the (mass) normalized collision energy (NCE, as defined by ThermoScientific) was set between 5 and 18%, which corresponds to 0.075 - 0.27 V applied for CID with the current instrument calibration. The activation Q, which defines the frequency of the applied radio frequency potential, was set at 0.30 and a 30 ms activation time was used.

We have installed a gas manifold on the buffer gas inlet of our instrument to allow the mixing of reagents with the helium buffer gas before introduction into the ion trap. Liquid reagents are introduced into the manifold from a metered syringe pump, where they evaporate. The partial pressure of these reagents may be controlled through the syringe pump rate, the helium flow rate, and by heating or cooling the manifold, which is done by wrapping the manifold tubing with a temperature-controlled water coil. All gas ports are controlled by manually actuated precision needle valves, which provide control of the flow rates. The helium buffer gas flow rate is monitored with an electronic mass flow meter and flow is controlled by the needle valve to the exhaust line. When operating the mass spectrometer without additional

reagents, three-way valves route the buffer gas through a clean section of tubing. The reagent pathway may be purged by flushing with clean gas (e.g., nitrogen) into a vacuum pump. The vacuum pump is preceded by a cold trap to aid in the removal of volatile reagents. The entire manifold is constructed from stainless steel tubing and components using stainless steel compression fittings, so that it may be periodically removed and baked in an oven for cleaning.

To probe gas-phase reactions of selected precursor ions with background neutrals, ions were isolated using widths of 1-2 m/z units. Here too, the specific width used was chosen empirically to ensure maximum ion isolation efficiency. The ions were then stored in the LIT for periods ranging from 1 ms to 10 s. When examining ion-molecule reactions (IMRs), our intent was not to measure or report rates or rate constants, but to identify the *pathways* by which ions react with neutrals such as H₂O or CH₃OH in the LIT. Because the experiments were performed using the multi-dimensional tandem mass spectrometry capabilities of the linear ion trap, care had to be taken to ensure that enough neutral reagent was present in the ion trap for ion-molecule reaction studies, without hampering the "synthesis" of the [OUCH]+ precursor ion by repeated CID steps. For both CID and IMR experiments, the mass spectra displayed were created by accumulating and averaging at least 30 isolation, dissociation, and ejection/detection steps.

High-resolution/high-accuracy measurements were performed on a ThermoScientific (San Jose, CA) LTQ-Orbitrap Elite mass spectrometer using experimental conditions and settings like those outlined above for the LTQ-XL instrument. With the Orbitrap instrument, the experiments were performed with the 120,000 resolution setting, NCE values of 8 - 16%, activation Q setting of 0.25, and a 10 ms activation time. Mass spectra were collected for 1 minute at each MSⁿ stage.