## **Electronic Supplementary Information**

## Electrochemical Surface-Enhanced Raman Spectroscopy (EC-SERS): A Tool for the Identification of Polyphenolic Components in Natural Lake Pigments

M. M. Eisnor<sup>a</sup>, K.E.R. McLeod<sup>a</sup>, S. Bindesri<sup>a</sup>, S. A. Svoboda<sup>b</sup>, K. L. Wustholz<sup>c</sup>, C. L. Brosseau<sup>a\*</sup>

\* To whom correspondence should be addressed: Christa L. Brosseau (christa.brosseau@smu.ca) Phone (902) 496-8175 Fax (902) 496-8104 Department of Chemistry, Saint Mary's University, Halifax, Nova Scotia, Canada, B3H 3C3.

<sup>*a*</sup> Department of Chemistry, Saint Mary's University, Halifax, Nova Scotia, Canada.

<sup>b</sup> Department of Conservation, The Colonial Williamsburg Foundation, Williamsburg, Virginia, USA.

<sup>c</sup> Department of Chemistry, The College of William and Mary, Williamsburg, Virginia, USA.





**Figure S-1:** Cathodic EC-SERS spectra for 2000 ppm caffeic acid and 2000 ppm chlorogenic acid on the surface of the AgNP coated screen-printed electrode in 0.1 M NaF supporting electrolyte. Applied voltage was stepped in the cathodic direction in 100 mV increments. Laser excitation was 780 nm. Power at the sample was 80 mW, and acquisition time was 30 s.

Dye	Raman Shift (cm <sup>-1</sup> )			Dof	
	In Air	OCP	-1.0V	KCI	
Apigenin	1178 w	1168 w	1166 w, 831 m	29	-
Luteolin	1504 m, 510 w, 468 w	1504 m, 510 w, 468 w	512 w	29	-
Quercetin	-	1598 s, 1516 m, 1463 m, 1408 m, 1334 m, 1248 m, 1214 m, 1203 m, 1103 w, 1087 m, 1007 w, 962 w, 845 w, 680 w, 634 w, 586 w, 567 sh, 481 s, 417 m	1595 w, 1461 w, 1207 w,1101 w, 844 w, 427 w	30	42
Rhamnetin	1101 w,1001 w	1598 m , 1516 w, 1465 w, 1406 w, 1346 w, 1318 w, 1252 w, 1207 w, 1163 w, 1135 w, 1103 w, 1088 w, 963 w, 676 w, 633 w, 590 w, 568 w, 553 w, 477 m, 463 sh, 437 w, 402 w, 321 w	1465 w, 1409 w, 1310 w, 844 w, 643 w, 604 w	-	-
Kaempferol	-	1169 w, 967 w, 480 m	1583 w	42	
Emodin	1321 w, 1278 w, 1262 w, 641 w, 471 w	1323 w, 1281 w, 1263 w, 942 w, 91 w, 642 w, 472 w	1557 w, 1364 w	42	43
Caffeic acid	1613 w, 1287 w	1598 m, 1492 m, 1448 m, 1432 m, 1343 w, 1163 s, 1109 s, 980 w, 614 w, 573 w, 491 s, 441 s	-	44	-
Chlorogenic acid	1596 w, 1108 m, 487 w, 436 w	1595 w, 1220 m, 1185 m, 754 w, 564 w, 487 s, 436 m, 358 w	1184 w, 485 w, 437 w	45	-

Table S-1: Discriminant peaks of apigenin, luteolin, quercetin, rhamnetin, kaempferol, emodin, caffeic acid and chlorogenic acid

Abbreviations used: s, strong; m, medium; w, weak; sh, shoulder



signal of caffeic acid.



**Figure S-3:** Comparison of EC-SERS spectra of in air and at the voltage that gives the best SERS signal of rhamnetin.



**Figure S-4:** Comparison of EC-SERS spectra of in air and at the voltage that gives the best SERS signal of chlorogenic acid.



signal of kaempferol.



**Figure S-6:** Comparison of EC-SERS spectra of *Reseda Lake* extract at OCP and -0.1V cathodic, luteolin at OCP anodic and quercetin at -0.5V anodic. Laser excitation was 780 nm. Power at the sample was 80 mW, and acquisition time was 30 s. Orange dashed line indicates *Reseda Lake* OCP cathodic being compared to quercetin and purple dashed line indicates *Reseda Lake* -0.1V being compared to luteolin.



**Raman Shift / cm<sup>-1</sup> Figure S-7:** Anodic progression of EC-SERS spectra for 2.0 mg *Stil de Grain* extract. Laser excitation was 780 nm. Power at the sample was 80 mW, and acquisition time was 30 s.



**Figure S-8:** Anodic progression of EC-SERS spectra for simulated *Stil de Grain* art sample extract. Laser excitation was 780 nm. Power at the sample was 120 mW, and acquisition time was 60 s.