Supporting information

The catalytic reaction mechanism of tyrosylprotein sulfotransferase-1

Pavel Šmak¹, Igor Tvaroška^{1,2,*}, Jaroslav Koča^{1,3,†}

¹ National Center for Biomolecular Research (NCBR), Faculty of Science, Masaryk University,

Brno, Czech Republic

² Institute of Chemistry, Slovak Academy of Sciences, Bratislava, Slovak Republic

³ Central European Institute of Technology (CEITEC), Masaryk University, Brno, Czech Republic

* To whom correspondence should be addressed. E-mail: <u>chemitsa@savba.sk</u>

[†] deceased



Fig. S1 Development of the RMSD of protein non-hydrogen atoms towards the initial X-ray structure for the two MD simulations. Two different force fields were used for the description of the enzyme.



Fig. S2 Mean values of selected geometrical parameters from MD simulation with FF14SB force field.

Table S1 Distances (in Angstroms) between selected residues obtained from the metadynamicssimulations and potential energy scan. PAP denotes the phosphate part of PAP, the smallestdistances between the pairs of residues are provided.

Distance / Å	CPMD Metadynamics			Optimized geometries		
	MC	TS	Р	MC	TS	Р
d _{OT-S} / Å	3.6	2.5	1.9	3.5	2.2	1.9
d _{OP-S} / Å	1.7	2.4	2.8	1.7	2.4	2.9
d _{T-H} / Å	1.0	1.4	1.6	1.3	1.5	1.7
d _{G-H} / Å	1.7	1.2	1.0	1.1	1.0	1.0
PAP – ARG 79	2.56	2.49	1.88	2.42	2.24	1.42
PAP – SER 286	3.07	3.49	4.09	1.72	1.50	1.59
PAP – THR 82	2.86	1.83	1.99	2.07	1.81	1.85
PAP – THR 83	1.71	1.46	1.53	2.12	1.80	1.86
PAP-GLY 81	2.20	2.48	2.19	2.50	2.65	2.58
PAP – SER 80	2.18	3.04	2.89	2.36	2.60	2.49
Sulfate – SER 286	1.93	4.33	4.91	3.49	3.74	4.19
Sulfate – THR 83	2.77	2.85	3.39	3.01	3.28	3.58
Sulfate – THR 82	2.94	3.52	3.66	2.99	3.25	3.41
Sulfate – LYS 159	2.49	2.60	2.69	4.57	4.44	4.60
Sulfate – ARG 79	2.25	2.36	2.70	1.61	1.83	2.66
Sulfate – ARG 285	2.11	2.28	1.95	3.48	2.70	2.61



Fig. S3 Dependence of distances d_{OP-S} and d_{OT-S} , and the shape of the transferred sulfate (torsion of 0° indicates planarity of the group), on the collective variable CV_1 . The data from the metadynamics simulation were smoothed using a moving average filter.



Fig. S4 Probability of the different structures with respect to the collective variables CV_1 and CV_2 within the CPMD simulation. The local minima of free energy are implied by the maxima of probability in the graph. A snapshot was recorded for every 10 MD steps.



Fig. S5 Development of the value of CV_1 within the metadynamics (MTD) simulation (blue line) and positions of added metadynamics potentials (orange dots). After adding the last shown metadynamics potential, the walker crossed the reaction barrier to the side of the MC.



Fig. S6 The development of the value of CV2 within the potential energy scan along CV1.



Fig. S7 The development of the distance between the hydroxyl oxygen of TYR 343 (acceptor) and the proximal oxygen of the carboxyl group of GLU 100 (base) within the potential energy scan along CV1.



Fig. S8 Distance between ARG 79 and transferred sulfate residue within the potential energy scan.

The schemes were created with the use of Marvin.⁶⁸ Trajectories were processed with the use of cpptraj (Amber Tools),⁶⁹ and the visualization created with VMD.⁵³