

Supporting Information for the manuscript:

**Stereo-selective synthesis of non-canonical γ -hydroxy- α -amino acids
by enzymatic carbon-carbon bond formation**

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Supplementary Figure S1-S6

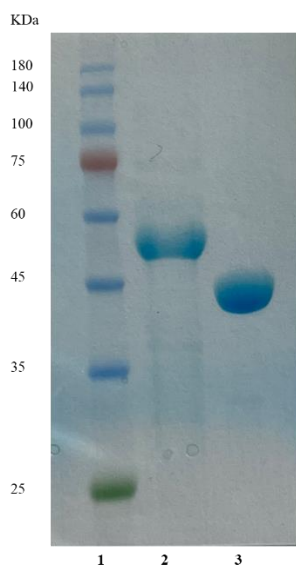


Figure S1. SDS PAGE of ApUstD and PfK. Lane 1: protein markers, lane 2: purified ApUstD, lane 3: purified PfK.

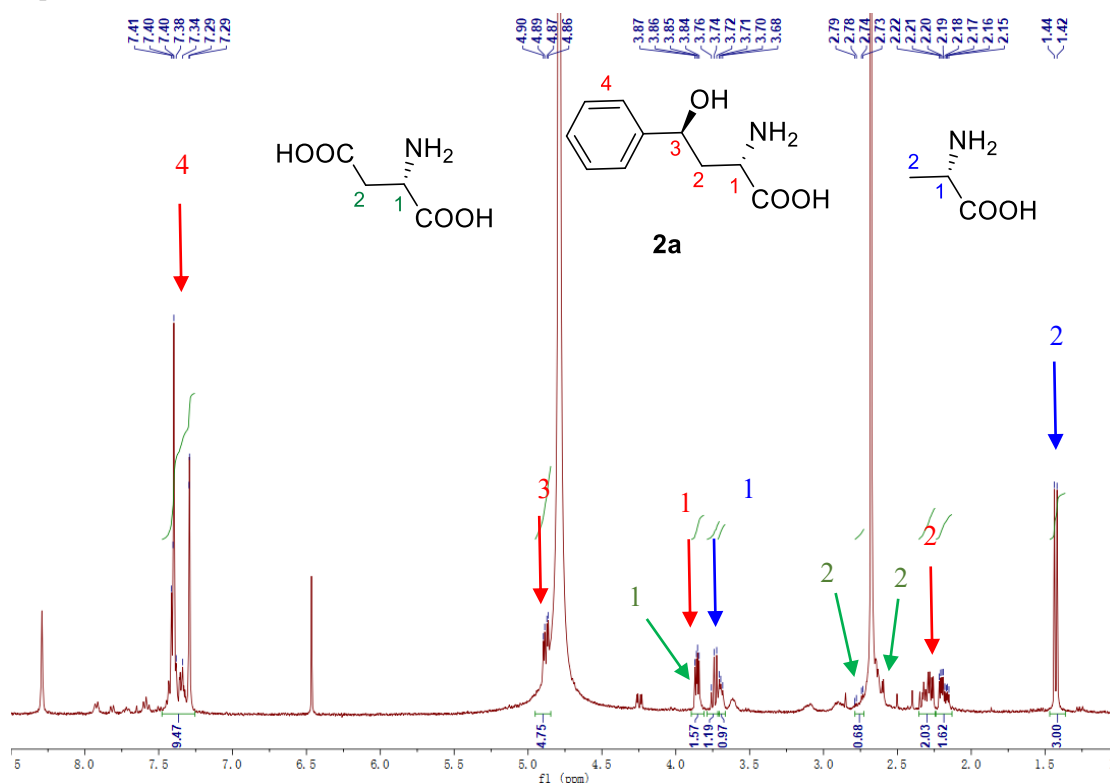


Figure S2. ApUstD reaction with L-Asp and benzaldehyde. 1 ml reaction containing 4 mM L-aspartic acid, 5 mM benzaldehyde (**1a**), 1mM dithiothreitol, 100 μ M PLP and 0.3 mg/ml ApUstD in 50 mM phosphate buffer, at pH 7.4 were run at 25 °C for 24 hours. Reaction mixture was lyophilized, and then was dissolved in 600 μ l D₂O. ¹H NMR was recorded thereafter. Resonances of different compound were assigned.

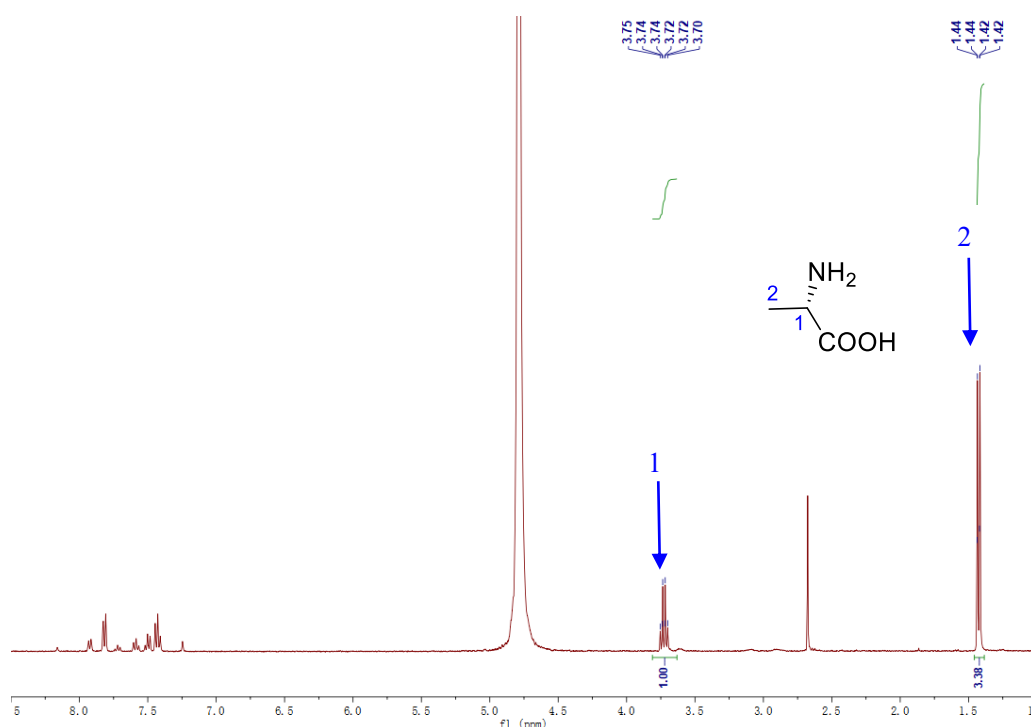


Figure S3. ApUstD reaction with L -Alanine and benzaldehyde. 1 ml reaction containing 4 mM L -alanine, 5 mM benzaldehyde (**1a**), 1mM dithiothreitol, 100 μ M PLP and 0.3 mg/ml ApUstD in 50 mM phosphate buffer, at pH 7.4 were run at 25 $^{\circ}$ C for 24 hours. Reaction mixture was lyophilized, and then was dissolved in 600 μ l D_2O . 1H NMR was recorded thereafter. No product was observed.

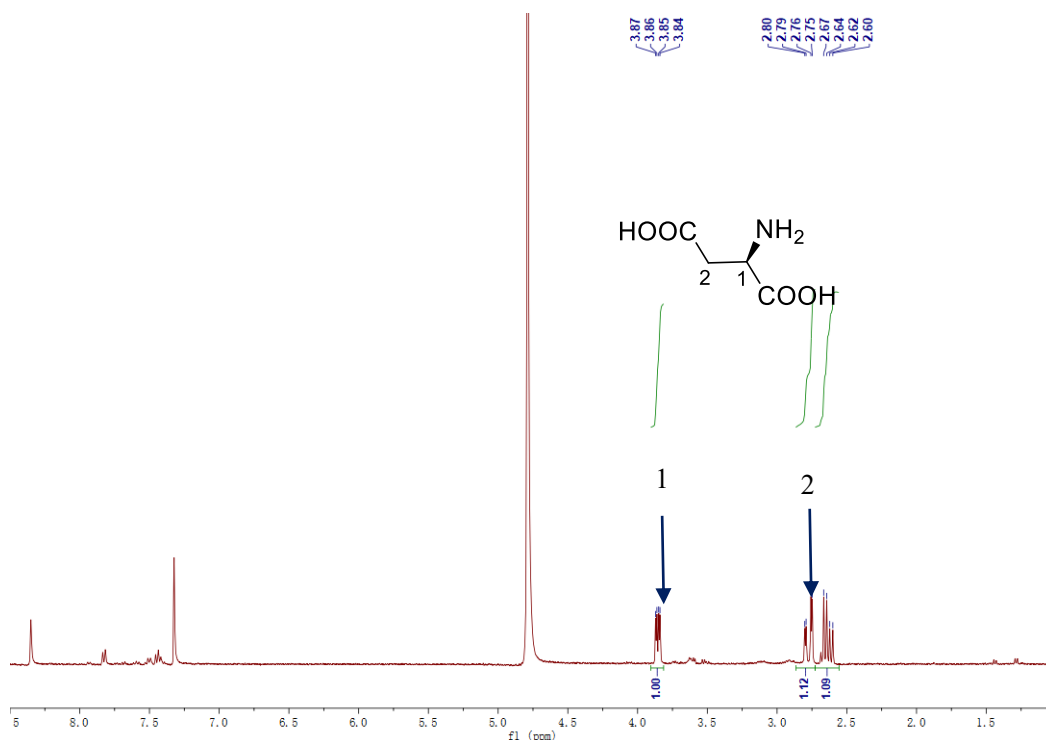


Figure S4. ApUstD reaction with D -Asp and benzaldehyde. 1 ml reaction containing 4 mM D -aspartic acid, 5 mM benzaldehyde (**1a**), 1mM dithiothreitol, 100 μ M PLP and 0.3 mg/ml ApUstD in 50 mM phosphate buffer, at pH 7.4 were run at 25 $^{\circ}$ C for 24 hours. Reaction mixture was lyophilized, and then was dissolved in 600 μ l D_2O . 1H NMR was recorded thereafter. No product

was observed.

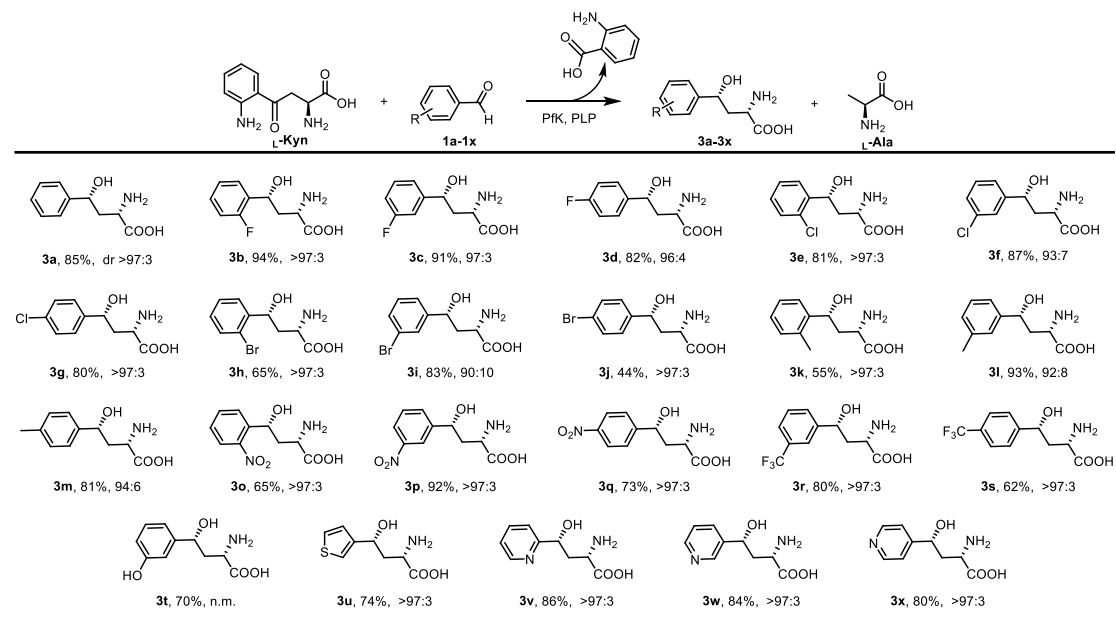


Figure S5. Conversion and diastereoisomeric ratio (d.r.) of **3a-3x** from PfK reaction. Reaction details were indicated in the method section.

X-ray crystallographic analyses of **2c**

Colorless needle crystals of **2c** were obtained by recrystallization from methanol. The crystal data was collected on a Bruker D8 VENTURE diffractometer by CuK α radiation ($\lambda = 1.54178$). Integration and scaling of intensity data was performed using the SAINT program. Data were corrected for the effects of absorption using SADABS. The structures were solved by direct method and refined with full-matrix least-squares technique using SHELX-2014 software. Non-hydrogen atoms were refined with anisotropic displacement parameters, and hydrogen atoms were placed in calculated positions and refined with a riding model. The crystallographic data have been deposited at the Cambridge Crystallographic Data Center (deposition no. CCDC 2084340), which can be obtained free of charge from the CCDC via https://www.ccdc.cam.ac.uk/data_request/cif.

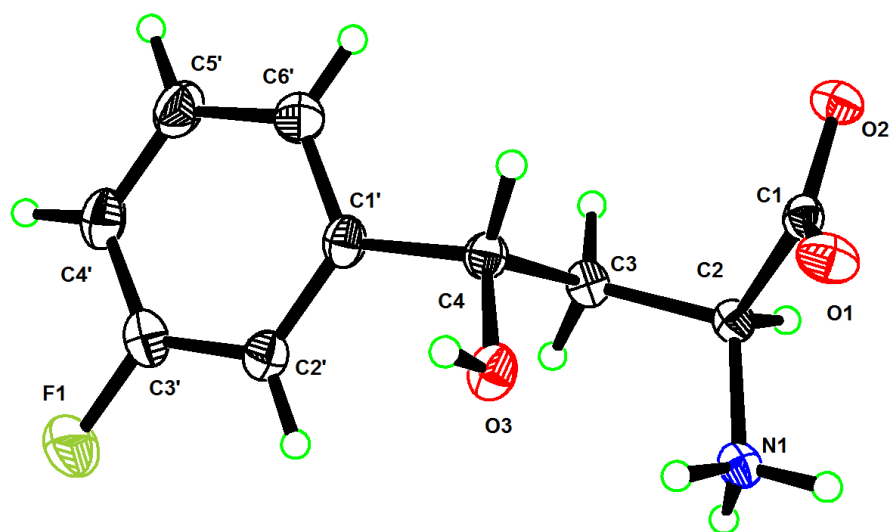


Figure S6. Perspective ORTEP drawing for **2c**

Supplementary Tables

Table S1. ApUstD reaction with L-Asp and benzaldehyde.

	Enzyme	L-Asp	1a	time	temp	2a	L-Ala	L-Asp
1	0.3 mg/ml	4 mM	5 mM	24 h	25 °C	49.1 %	30.3 %	20.6 %
2	0.3 mg/ml	4 mM	10 mM	24 h	25 °C	75.9 %	24.1 %	<0.1 %
3	0.3 mg/ml	4 mM	20 mM	24 h	25 °C	84.4 %	12.2 %	3.4 %
4	0.3 mg/ml	4 mM	40 mM	24 h	25 °C	84.9 %	6.1 %	9.0 %
5	0.3 mg/ml	8 mM	40 mM	24 h	25 °C	76.9 %	6.7 %	16.4 %

1 ml reaction containing both substrates, 1mM dithiothreitol, 100 μ M PLP and 0.4 mg/ml ApUstD in 50 mM phosphate buffer, at pH 7.4 were run at 25 °C. Substrate concentration was indicated in different entries. Relative content of different compounds were determined based on the integrals from ^1H NMR.

Table S2. X-ray crystallographic data for **2c**

Identification code	cu_2021524_0m
Empirical formula	$\text{C}_{10}\text{H}_{12}\text{FNO}_3$
Formula weight	213.21
Temperature/K	170.0
Crystal system	monoclinic
Space group	$P2_1$
a/Å	5.93840(10)
b/Å	5.30530(10)
c/Å	15.9518(4)
$\alpha/^\circ$	90
$\beta/^\circ$	93.2890(10)
$\gamma/^\circ$	90
Volume/Å ³	501.734(18)
Z	2
$\rho_{\text{calc}}/\text{g/cm}^3$	1.411
μ/mm^{-1}	0.993
F(000)	224.0
Crystal size/mm ³	0.15 × 0.08 × 0.05
Radiation	$\text{CuK}\alpha$ ($\lambda = 1.54178$)
2 θ range for data collection/ $^\circ$	5.55 to 127.37
Index ranges	$-6 \leq h \leq 6, -6 \leq k \leq 6, -18 \leq l \leq 18$
Reflections collected	5180
Independent reflections	1596 [$R_{\text{int}} = 0.0275, R_{\text{sigma}} = 0.0272$]
Data/restraints/parameters	1596/1/149
Goodness-of-fit on F^2	1.095
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0260, wR_2 = 0.0656$
Final R indexes [all data]	$R_1 = 0.0268, wR_2 = 0.0676$
Largest diff. peak/hole / e Å ⁻³	0.15/-0.13
Flack parameter	0.13(8)

Table S3. Kinetic parameters of ApUstD with different substrates.

Substrate	k_{cat} (min^{-1})	K_{M} (μM)	$k_{\text{cat}}/K_{\text{M}}$ ($\text{M}^{-1}\text{min}^{-1}$)
3-F (1c)	5.0 ± 0.1	3400 ± 200	1400
2-Br (1h)	3.5 ± 0.1	1700 ± 200	2000
4-NO ₂ (1q)	6.9 ± 0.3	400 ± 40	17000

200 μl reaction containing 12 mM L-aspartic acid, 100 μM PLP, 0.3 mg/ml ApUstD and varied aldehyde concentration in 50 mM phosphate buffer at pH 7.4 were run at 25 °C. 40 μl of reaction mixture were quenched by 20 μl of 1M HCl after 1, 2, 3, 4, 5 minutes respectively. The product formation was monitored by HPLC-UV.

Supplementary Methods

General

All enzyme-encoding pET-28a(+) based expression plasmids were purchased from GenScript and expression strain *E. coli* BL21 (DE3) were purchased from Sangon Biotech. Proteins were analyzed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) under denaturing conditions. The gels were stained with Coomassie brilliant blue.

The crystal data was collected on a Bruker D8 VENTURE diffractometer. Semi-Preparative HPLC was performed on a Waters 1525 instrument with a Waters 2489 UV detector using a DAICEL CHIRALPAK IF column (S-5 μ m, 250 \times 10 mm) and a Water SunFire column (S-5 μ m, 100 \times 19 mm). HRESIMS spectra were recorded on an Angilent G6520 Q-ToF mass detector. NMR spectra were recorded using the Bruker AVANCE III 400, 500 and 600 MHz instrument. Standard abbreviations indicating multiplicity were used as follows: s (singlet), d (doublet), t (triplet), and m (multiplet). Chemical shifts were reported in ppm (δ), coupling constants (J) in hertz. Econosep C18 60A (50 μ m, DIKMA, China) was used for column chromatography (CC).

Antibiotics, isopropyl β -D-thiogalactoside (IPTG) and dithiothreitol (DTT) were purchased from Sangon Biotech. L-Aspartic acid was purchased from RHAWN. Aldehyde substrates were purchased from J & K SCIENTIFIC LTD. and bidepharmarmatech. PLP and L-kynurenine were from J & K. Deuterated solvents were purchased from J & K SCIENTIFIC LTD. High pressure cell disrupter and Ni-NTA unionrose 6FF agarose were purchased from Union Biotech.

Protein expression and purification

E. coli BL21 (DE3) cells were transformed with pET-28a(+) expression plasmids by heat shock. Transformed cells were first cultivated on solid medium (LB-AGAR plates with 50 μ g/ml kanamycin) and then in liquid medium (LB with 50 μ g/ml kanamycin). After incubation at 37°C overnight, 15 ml of pre-culture was used to inoculate fresh Terrific Broth (TB) medium (1L) with kanamycin (50 μ g/ml). The cells were grown in 3L shaking flask at 37 °C (170 rpm) until OD600 reached 1.0. The culture was cooled to 18 °C and supplemented with 100 μ M isopropyl β -D-thiogalactoside (IPTG). This culture was incubated at 18 °C for 20 hours. Cells were harvested by centrifugation at 13000 g for 15 minutes and stored at -20 °C.

For purification of ApUstD, cell pellets (10 g) were suspended in 50 ml lysis buffer (50 mM sodium phosphate, 300 mM NaCl, pH 8.0). Cells were disrupted by high pressure

cell disrupter for 3 times. Lysates were centrifuged at 42000 g for 15 min at 4 °C. The cleared lysate was loaded onto a Ni-NTA unionrose 6FF agarose column. The agarose beads were washed with 150 ml lysis buffer containing 100 mM, 150 mM and 200 mM imidazole respectively. The protein was eluted in a lysis buffer solution containing 500 mM imidazole. Protein containing fractions were collected and dialyzed against dialysis buffer (50 mM sodium phosphate, 50 mM NaCl, 1 mM dithiothreitol, 100 µM PLP, pH 7.4). Protein was used directly after dialysis (yield, 2.3 mg/L).

For purification of PfK, cell pellets (10 g) were suspended in 50 ml lysis buffer (50 mM sodium phosphate, 300 mM NaCl, pH 8.0). Cells were disrupted by high pressure cell disrupter for 3 times. Lysates were centrifuged at 42000 g for 20 min at 4 °C. The cleared lysate was loaded onto a Ni NTA unionrose 6FF agarose column. The agarose beads were washed with 100 ml lysis buffer containing 10 mM and 20 mM imidazole respectively. The protein was eluted in a lysis buffer solution containing 250 mM imidazole. Protein containing fractions were collected and dialyzed against dialysis buffer (50 mM sodium phosphate, 50 mM NaCl, 100 µM PLP, pH 7.4), aliquoted and stored at -80 °C.

Protein sequence:

ApUstD

MGSSHHHHHHSSGLVPGSHMKSVANSSSLHDVDKDSVPLSSGTNGTTQAETPLDNVID
VESVRSHPVLEGETAAFNNASGTVVLKEAIESTSKFMFSFPFPPGVDAKSMEAITA
YTGNKGKVATFINALPDEITFGQSTTCLFRLGLSLKPMLNSDCEIVCSTLCHEAAA
SAWIHLSDRLGITIKWWSPTTTPNSPDDPVLTTDSLKPLLSPKTRLVTCNHVSNVVG
TIHPIREIADVHAIPGCMLIVDGVACVPHRPVDVKELDVDFYCFSWYKLFGPHMGT
LYASRKAQDRYMTSINHFLSSSSLDGKLALGMPSFELQLMCSPIVSYLQDIVGWRD
IVRQETVLVKILLQYLLSKPGVYRVFGRNSDPSQRVSIIVTFEVVGRSSGDVAMRVN
TRNRFRTSGICLAPRPTWDVLKPVSAAGLIRVSFVHYNTVEEVREFCNELDEIVTQ
DT

PfK

MGSSHHHHHHSSGLVPGSHMGTTTRNDCLALDAQDSLAPLRQQFALPEGVIYLDGNSL
GARPVAALARAQAVIAEEWGNGLIRSWNSAGWRDLSERLGNRLATLIGARDGEVVVT
DTTSINLFKVLSAALRVQATRSPERRVIVTETS NFPTDLYIAEGLADMLQQGYTLRL
VDSPEELPQAIDQDTAVVMLTHVNYKTGYMHDMQALTALSHECGALAIWDLAHSAGA
VPVDLHQAGADYAIGCTYKYLNNGPGSQAFVWVSPQLCDLVPQPLSGWFGHSRQFAM
EPRYEPSNGIARYLCGTQPITSLAMVECGLDVFAQTDMASLRRKSLALTDLFIELVE
QRCAAHETLTVTPREHAKRGSHVSFEHPEGYAVIQALIDRGVIGDYREPRIMRFGFT
PLYTTTFTEVWDAVQILGEILDRKTWAQAQFQVRHSVT

Time dependent Progress of ApUstD reaction

Reaction containing 12 mM L-aspartic acid, 80 mM benzaldehyde (**1a**), 100 μ M PLP and 0.3 mg/ml ApUstD in 50 mM phosphate buffer, at pH 7.4 were run at 25 °C. 1 ml reaction mixture was lyophilized at different time point (12h, 24h, 48h, 72h, 96h). After lyophilization, the mixture was dissolved in 600 μ l D₂O and submitted to ¹H NMR measurement. Relative content of different compounds was calculated based on integrals of key ¹H NMR resonances and shown in Figure 2b.

Determination of conversion and diastereoisomeric ratio (d.r.)

Products of **2a-2f**, **2h**, **2i**, **2r**, **2s**, **2u-2aa** and **2ad**: 1 ml reactions containing 0.4 mg ApUstD, 12 mM L-Aspartic acid, 80 mM aldehyde, 1 mM DTT, 100 μ M PLP and 50 mM sodium phosphate, 50 mM NaCl, at pH 7.4 were run at 25 °C. After 72 hours, the reaction mixtures were centrifugated and lyophilized. After lyophilization, the mixtures were dissolved in 600 μ l of D₂O. ¹H NMR were recorded. Conversion efficiency (% of consumed amino acid) and ratio of different products were determined by integration of characteristic ¹H NMR resonances from different compounds.

Products of **2g**, **2j**, **2o-2q** and **2t**: 1 ml reactions containing 0.4 mg ApUstD, 12 mM L-Aspartic acid, 80 mM aldehyde, 4% DMSO (v/v), 1 mM DTT, 100 μ M PLP and 50 mM sodium phosphate, 50 mM NaCl, at pH 7.4 were run at 25 °C. After 72 hours, the reaction mixtures were centrifugated and lyophilized. After lyophilization, the mixtures were dissolved in 600 μ l of D₂O. ¹H NMR were recorded. Conversion efficiency (% of consumed amino acid) and ratio of different products were determined by integration of characteristic ¹H NMR resonances from different compounds.

Products of **2k-2n**, **2ab** and **2ac**: 1 ml reactions containing 0.4 mg ApUstD, 12 mM L-Aspartic acid, 16 mM aldehyde, 2% DMSO (v/v), 1 mM DTT, 100 μ M PLP and 50 mM sodium phosphate, 50 mM NaCl, at pH 7.4 were run at 25 °C. After 72 hours, the reaction mixtures were centrifugated and lyophilized. After lyophilization, the mixtures were dissolved in 600 μ l of D₂O. ¹H NMR were recorded. Conversion efficiency (% of consumed amino acid) and ratio of different products were determined by integration of characteristic ¹H NMR resonances from different compounds.

Products of **3a-3x**: 1 ml reactions containing 0.5 mg PfK, 24 mM L-kynurenine, 180 mM aldehyde, 100 μ M PLP and 50 mM sodium phosphate, 50 mM NaCl, at pH 7.4 were run at 25 °C. After 12 hours, the reaction mixtures were centrifugated and lyophilized. After lyophilization, the mixtures were dissolved in 600 μ l of D₂O. ¹H NMR were recorded. Conversion efficiency (% of consumed L-kynurenine) and ratio of different products were determined by integration of characteristic ¹H NMR resonances from different compounds.

The diastereoisomeric ratio of ApUstD and PfK reactions were determined by

integration of ^1H NMR resonances of $\gamma\text{-H}$ of the products and their diastereo-isomers (Figure S58-S85).

Determination of Kinetic Parameters

200 μl reaction containing 12 mM L-aspartic acid, 100 μM PLP, 0.3 mg/ml ApUstD and varied aldehyde concentration in 50 mM phosphate buffer at pH 7.4 were run at 25 $^\circ\text{C}$. 40 μl of reaction mixture were quenched by 20 μl of 1M HCl after 1, 2, 3, 4, 5 minutes respectively. The product formation was monitored by HPLC-UV. Product concentration were calculated based on the respective calibration curve. Velocities were calculated and plotted against substrate concentrations. The velocity was fitted to equation of $v = v_{\text{max}} * [\text{s}] / (K_{\text{M}} + [\text{s}])$ and values of k_{cat} , K_{M} and $k_{\text{cat}}/K_{\text{M}}$ were obtained thereafter.

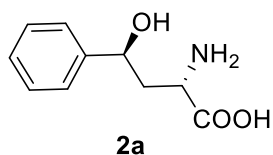
Semi-preparative reaction and characterization of Compounds

For the structural characterization of products **2a-2x** and **2ab**, 15 ml reactions were incubated for 72 h, lyophilized. Products of **2a-2x** were loaded on C18 chromatography eluted with methanol in water in a stepwise manner (0%, 20%, 40%, 60%, 95%). The purification was monitored with HPLC. Fractions containing products were collected and the solvent was removed by rotavapor to yield products.

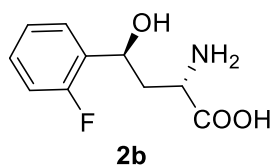
Product of **2ab** was purified by semi-preparative HPLC using a DAICEL CHIRALPAK IF HPLC column (methanol: 20 mM ammonium bicarbonate in water = 1:99, 4 ml/min). The purified compounds were dissolved in methanol to detect the HRESIMS, and dissolved in NaOD/D₂O analyzed by ^1H NMR and ^{13}C NMR.

For the structural characterization of products **3a-3x**, 5 ml reactions were incubated for 12 h, lyophilized. Products of **3a-3g**, **3o-3q** and **3u** were purified by semi-preparative HPLC using a Water sunfire column eluted with MeOH/H₂O (0-30 min, 5-95%, 10 ml/min). Products of **3h-3m** were purified by semi-preparative HPLC using a Water sunfire column eluted with MeOH/H₂O (0-30 min, 25-95%, 10 ml/min). Products of **3r** and **3s** were purified by semi-preparative HPLC using a Water sunfire column eluted with MeOH/H₂O (0-30 min, 40-95%, 10 ml/min). Products of **3t** and **3v-3x** were purified by semi-preparative HPLC using a Water sunfire column eluted with MeOH/H₂O (0-30 min, 2-50%, 10 ml/min). The solvent was removed by rotavapor to yield products.

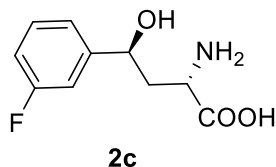
All reported chemical shifts was adjusted with reference of D₂O (4.79). Raw NMR Data of all compounds has been deposited on Zendo.org (DOI: <https://doi.org/10.5281/zenodo.5205646>)



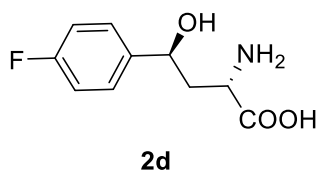
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.44 – 7.33 (m, 5H), 4.89 (dd, J = 9.5, 3.7 Hz, 1H), 3.86 (dd, J = 7.0, 4.2 Hz, 1H), 2.30 (ddd, J = 14.8, 9.5, 4.2 Hz, 1H), 2.19 (ddd, J = 14.8, 7.0, 3.7 Hz, 1H) ppm. ^{13}C NMR (125 MHz, pD = 7.0) δ 174.3, 143.0, 128.9, 128.9, 128.1, 125.8, 125.8, 70.9, 52.9, 37.9 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{14}\text{NO}_3$ calculated 196.0968, observed 196.0968.



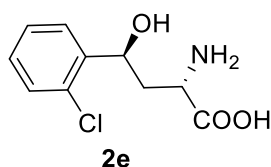
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.46 (t, J = 7.6 Hz, 1H), 7.33 (q, J = 6.8 Hz, 1H), 7.22 (m, 1H), 7.11 (m, 1H), 5.12 (dd, J = 9.1, 4.6 Hz, 1H), 3.34 (dd, J = 8.6, 4.8 Hz, 1H), 2.16 (ddd, J = 14.0, 9.1, 4.8 Hz, 1H), 1.87 (ddd, J = 14.0, 8.6, 4.6 Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 182.8, 159.7 (d, $^1J_{\text{C-F}}$ = 244.3 Hz), 130.2 (d, $^2J_{\text{C-F}}$ = 13.2 Hz), 127.5 (d, $^3J_{\text{C-F}}$ = 8.5 Hz), 127.5 (d, $^3J_{\text{C-F}}$ = 4.4 Hz), 124.6 (d, $^4J_{\text{C-F}}$ = 3.3 Hz), 115.6 (d, $^2J_{\text{C-F}}$ = 21.7 Hz), 65.4 (d, $^3J_{\text{C-F}}$ = 2.6 Hz), 53.5, 41.7 ppm. ^{19}F NMR (471 MHz, D_2O , pD = 8.0) δ -119.7 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{FNO}_3$ calculated 214.0874, observed 214.0874.



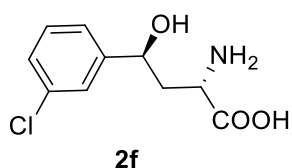
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.37 (td, J = 8.0, 6.0 Hz, 1H), 7.15 (m, 2H), 7.03 (m, 1H), 4.81 (m, 1H, partially overlapped with solvent), 3.31 (m, 1H, partially overlapped with solvent), 2.08 (ddd, J = 14.0, 9.1, 5.0 Hz, 1H), 1.80 (ddd, J = 14.0, 8.5, 4.6 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 182.8, 162.6 (d, $^1J_{\text{C-F}}$ = 243.4 Hz), 146.4 (d, $^3J_{\text{C-F}}$ = 6.8 Hz), 130.3 (d, $^3J_{\text{C-F}}$ = 8.2 Hz), 121.7 (d, $^4J_{\text{C-F}}$ = 2.8 Hz), 114.3 (d, $^2J_{\text{C-F}}$ = 21.1 Hz), 112.7 (d, $^2J_{\text{C-F}}$ = 21.9 Hz), 70.7, 53.4, 43.0 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 8.0) δ -113.7 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{FNO}_3$ calculated 214.0874, observed 214.0870. $[\alpha]_{\text{D}}^{20}$ = -30.0 (c = 0.10, Methanol).



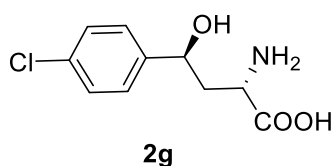
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.38 (m, 2H), 7.11 (t, J = 8.0 Hz, 2H), 4.81 (m, 1H, partially overlapped with solvent), 3.38 (m, 1H), 2.14 (ddd, J = 14.4, 8.7, 5.0 Hz, 1H), 1.89 (m, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 180.4, 162.0 (d, $^1J_{\text{C-F}}$ = 243.0 Hz), 139.2 (d, $^4J_{\text{C-F}}$ = 3.1 Hz), 127.8 (d, $^3J_{\text{C-F}}$ = 8.3 Hz), 127.8 (d, $^3J_{\text{C-F}}$ = 8.3 Hz), 115.3 (d, $^2J_{\text{C-F}}$ = 21.4 Hz), 115.3 (d, $^1J_{\text{C-F}}$ = 21.4 Hz), 70.5, 53.3, 41.6 ppm. ^{19}F NMR (471 MHz, D_2O , pD = 8.0) δ -115.5 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{FNO}_3$ calculated 214.0874, observed 214.0873.



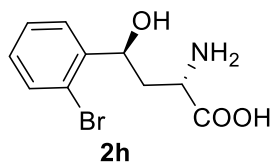
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.55 (d, J = 7.7 Hz, 1H), 7.41 (d, J = 7.7 Hz, 1H), 7.37 (t, J = 7.7 Hz, 1H), 7.28 (t, J = 7.7 Hz, 1H), 5.28 (dd, J = 9.6, 3.6 Hz, 1H), 3.41 (dd, J = 9.1, 4.2 Hz, 1H), 2.04 (ddd, J = 14.0, 9.6, 4.2 Hz, 1H), 1.86 (ddd, J = 14.0, 9.1, 3.6 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 182.8, 140.8, 131.5, 129.5, 128.9, 127.4, 126.9, 67.5, 53.5, 41.5 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{ClNO}_3$ calculated 230.0578, observed 230.0574.



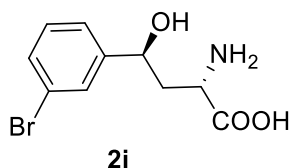
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.41 (s, 1H), 7.33 (m, 3H), 4.79 (m, 1H, partially overlapped with solvent), 3.32 (m, 1H), 2.07 (m, 1H), 1.81 (m, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 182.8, 145.9, 133.7, 130.1, 127.6, 125.9, 124.3, 70.6, 53.3, 43.0 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{ClNO}_3$ calculated 230.0578, observed 230.0580.



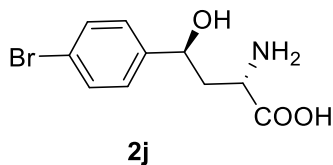
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.39 (d, J = 8.4 Hz, 2H), 7.34 (d, J = 8.4 Hz, 2H), 4.81 (m, 1H, partially overlapped with solvent), 3.36 (dd, J = 8.1, 5.0 Hz, 1H), 2.11 (ddd, J = 14.2, 9.1, 5.0 Hz, 1H), 1.84 (ddd, J = 14.2, 8.1, 4.5 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 181.6, 142.2, 132.7, 128.5, 128.5, 127.5, 127.5, 70.5, 53.3, 42.3 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{ClNO}_3$ calculated 230.0578, observed 230.0575.



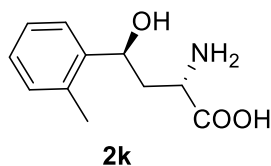
^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.60 (d, $J = 8.0$ Hz, 1H), 7.54 (d, $J = 8.0$ Hz, 1H), 7.41 (t, $J = 8.0$ Hz, 1H), 7.21 (t, $J = 8.0$ Hz, 1H), 5.23 (dd, $J = 10.3, 2.4$ Hz, 1H), 3.43 (dd, $J = 9.2, 4.2$ Hz, 1H), 2.00 (m, 1H), 1.87 (m, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $\text{pD} = 8.0$) δ 182.8, 142.4, 132.7, 129.2, 128.0, 127.1, 121.5, 69.8, 53.5, 41.6 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{BrNO}_3$ calculated 274.0073, observed 274.0066.



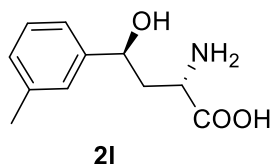
^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.56 (s, 1H), 7.46 (d, $J = 7.6$ Hz, 1H), 7.33 (d, $J = 7.6$ Hz, 1H), 7.28 (t, $J = 7.6$ Hz, 1H), 4.76 (m, 1H, partially overlapped with solvent), 3.32 (m, 1H, partially overlapped with solvent), 2.08 (ddd, $J = 14.0, 9.2, 4.9$ Hz, 1H), 1.78 (ddd, $J = 14.0, 8.5, 4.5$ Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $\text{pD} = 8.0$) δ 182.6, 146.2, 130.6, 130.4, 128.8, 124.8, 122.0, 70.6, 53.3, 43.0 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{BrNO}_3$ calculated 274.0073, observed 274.0072.



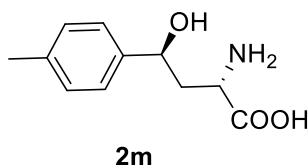
^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.54 (d, $J = 8.4$ Hz, 2H), 7.29 (d, $J = 8.4$ Hz, 2H), 4.77 (m, 1H, partially overlapped with solvent), 3.31 (m, 1H), 2.09 (ddd, $J = 14.0, 9.1, 5.0$ Hz, 1H), 1.80 (ddd, $J = 14.0, 8.3, 4.7$ Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $\text{pD} = 8.0$) δ 182.4, 142.8, 131.5, 131.5, 127.9, 127.9, 120.8, 70.6, 53.4, 42.8 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{BrNO}_3$ calculated 274.0073, observed 274.0072.



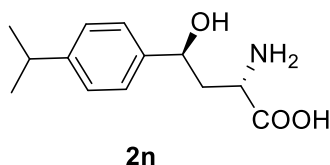
^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.45 (d, $J = 7.6$ Hz, 1H), 7.26 (m, 1H), 7.21 (m, 2H), 5.10 (dd, $J = 9.5, 3.9$ Hz, 1H), 3.36 (dd, $J = 8.7, 4.5$ Hz, 1H), 2.30 (s, 3H), 2.05 (ddd, $J = 14.2, 9.5, 4.5$ Hz, 1H), 1.76 (ddd, $J = 14.2, 8.7, 3.9$ Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , $\text{pD} = 8.0$) δ 184.4, 143.3, 136.6, 131.9, 129.0, 127.9, 126.6, 68.7, 55.1, 43.6, 19.5 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1122.



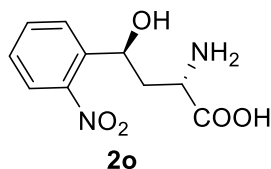
^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.30 (t, $J = 7.8$ Hz, 1H), 7.23 (s, 1H), 7.17 (t, $J = 7.8$ Hz, 2H), 4.76 (m, 1H, partially overlapped with solvent), 3.30 (q, $J = 5.0$ Hz, 1H), 2.31 (s, 3H), 2.10 (ddd, $J = 14.0, 8.8, 5.0$ Hz, 1H), 1.78 (ddd, $J = 14.0, 8.8, 5.0$ Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $\text{pD} = 8.0$) δ 182.9, 143.8, 138.8, 128.7, 128.4, 126.6, 123.0, 71.2, 53.5, 43.2, 20.4 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1124.



^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.28 (d, $J = 8.0$ Hz, 2H), 7.23 (d, $J = 8.0$ Hz, 2H), 4.76 (m, 1H, partially overlapped with solvent), 3.34 (dd, $J = 8.2, 4.9$ Hz, 1H), 2.29 (s, 3H), 2.12 (ddd, $J = 14.0, 9.1, 4.9$ Hz, 1H), 1.82 (ddd, $J = 14.0, 8.2, 4.7$ Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $\text{pD} = 8.0$) δ 182.0, 140.6, 138.0, 129.2, 129.2, 126.0, 126.0, 71.0, 53.4, 42.6, 20.1 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1119.

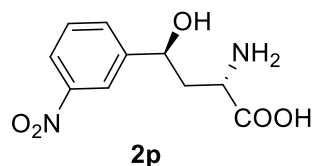


^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.32 (s, 4H), 4.79 (m, 1H, partially overlapped with solvent), 3.30 (dd, $J = 8.1, 5.1$ Hz, 1H), 2.89 (m, 1H), 2.10 (m, 1H), 1.78 (m, 1H), 1.20 (d, $J = 7.1$, 6H) ppm. ^{13}C NMR (125 MHz, D_2O , $\text{pD} = 8.0$) δ 183.0, 149.1, 141.4, 126.7, 126.7, 126.2, 126.2, 71.1, 53.5, 43.3, 33.3, 23.2, 23.2 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{13}\text{H}_{20}\text{NO}_3$ calculated 238.1438, observed 238.1438.

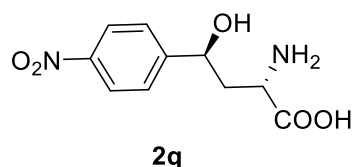


^1H NMR (400 MHz, D_2O , $\text{pD} = 8.0$) δ 7.95 (d, $J = 7.7$ Hz, 1H), 7.79 (d, $J = 7.7$ Hz, 1H), 7.72 (t, $J = 7.7$ Hz, 1H), 7.48 (t, $J = 7.7$ Hz, 1H), 5.40 (dd, $J = 9.6, 3.1$ Hz, 1H), 3.44 (dd, $J = 9.6, 4.2$ Hz, 1H), 2.07 (ddd, $J = 14.0, 9.6, 4.2$ Hz, 1H), 1.90 (ddd, $J = 14.0, 9.5, 3.1$ Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , $\text{pD} = 8.0$) δ 184.4, 148.6, 140.6, 135.5,

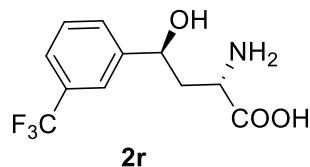
129.9, 129.2, 125.9, 67.6, 55.0, 44.0 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{10}H_{13}N_2O_5$ calculated 241.0819, observed 241.0812.



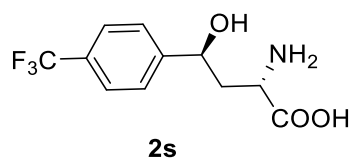
1H NMR (400 MHz, D_2O , pD = 8.0) δ 8.24 (s, 1H), 8.15 (d, J = 7.9 Hz, 1H), 7.77 (d, J = 7.9 Hz, 1H), 7.59 (t, J = 7.9 Hz, 1H), 4.94 (dd, J = 9.3, 4.4 Hz, 1H), 3.27 (m, 1H, partially overlapped with solvent), 2.12 (ddd, J = 14.0, 9.3, 4.9 Hz, 1H), 1.86 (ddd, J = 14.0, 8.5, 4.4 Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 182.7, 148.0, 145.9, 132.8, 129.8, 122.7, 120.9, 70.4, 53.4, 43.0 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{10}H_{13}N_2O_5$ calculated 241.0819, observed 241.0813.



1H NMR (400 MHz, D_2O , pD = 8.0) δ 8.15 (d, J = 8.5 Hz, 2H), 7.51 (d, J = 8.5 Hz, 2H), 4.88 (dd, J = 9.4, 4.4 Hz, 1H), 3.28 (dd, J = 8.6, 4.8 Hz, 1H), 2.02 (ddd, J = 14.0, 9.4, 4.8 Hz, 1H), 1.77 (ddd, J = 14.0, 8.6, 4.4 Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 184.2, 153.3, 148.4, 128.3, 128.3, 125.3, 125.3, 72.0, 64.0, 44.5 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{10}H_{13}N_2O_5$ calculated 241.0819, observed 241.0814.

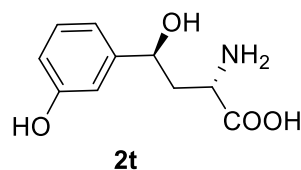


1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.71 (s, 1H), 7.62 (m, 2H), 7.54 (m, 1H), 4.89 (m, 1H), 3.35 (dd, J = 7.8, 4.8 Hz, 1H), 2.12 (m, 1H), 1.84 (m, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 182.4, 144.9, 130.1 (q, $^2J_{C-F}$ = 32.0 Hz), 129.7, 129.3, 124.5 (q, $^3J_{C-F}$ = 3.9 Hz), 124.2 (q, $^1J_{C-F}$ = 273.5 Hz), 122.7 (q, $^3J_{C-F}$ = 3.9 Hz), 70.8, 53.4, 42.9 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 8.0) δ -62.4 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{11}H_{13}F_3NO_3$ calculated 264.0842, observed 264.0844.

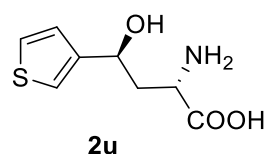


1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.79 (d, J = 8.1 Hz, 2H), 7.63 (d, J = 8.1 Hz, 2H), 5.05 (dd, J = 9.1, 4.1 Hz, 1H), 3.94 (dd, J = 6.8, 4.3 Hz, 1H), 2.24 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.0, 147.2, 129.3 (q, $^2J_{C-F}$ = 32.1 Hz), 126.2,

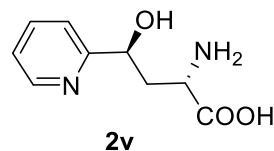
126.2, 125.6 (q, $^3J_{\text{C-F}} = 3.8$ Hz), 125.6 (q, $^3J_{\text{C-F}} = 3.8$ Hz), 124.2 (q, $^1J_{\text{C-F}} = 271.3$ Hz), 70.4, 52.9, 37.7 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 7.0) δ -62.3 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{13}\text{F}_3\text{NO}_3$ calculated 264.0842, observed 264.0840.



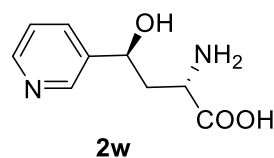
^1H NMR (500 MHz, D_2O , pD = 7.0) δ 7.28 (m, 1H), 6.94 (d, $J = 7.6$ Hz, 1H), 6.89 (s, 1H), 6.82 (d, $J = 8.1$ Hz, 1H), 4.83 (m, 1H, partially overlapped with solvent), 3.84 (d, $J = 6.2$ Hz, 1H), 2.23 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.3, 155.8, 145.1, 130.3, 117.7, 114.9, 112.6, 70.7, 52.9, 37.9 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{14}\text{NO}_4$ calculated 212.0917, observed 212.0915.



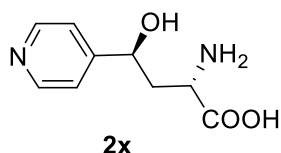
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.45 (m, 1H), 7.36 (s, 1H), 7.14 (d, $J = 5.3$ Hz, 1H), 4.99 (dd, $J = 9.2, 4.1$ Hz, 1H), 3.85 (m, 1H), 2.30 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.2, 144.1, 127.1, 125.5, 121.8, 67.1, 52.8, 37.2 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_8\text{H}_{12}\text{NO}_3\text{S}$ calculated 202.0532, observed 202.0530.



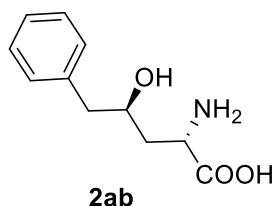
^1H NMR (600 MHz, D_2O , pD = 7.0) δ 8.44 (m, 1H), 7.86 (td, $J = 7.7, 1.8$ Hz, 1H), 7.51 (d, $J = 7.8$ Hz, 1H), 7.34 (m, 1H), 4.95 (t, $J = 6.5$ Hz, 1H), 3.88 (t, $J = 6.0$ Hz, 1H), 2.29 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.1, 160.5, 148.3, 138.4, 123.5, 120.9, 71.2, 52.7, 36.5 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_9\text{H}_{13}\text{N}_2\text{O}_3$ calculated 197.0921, observed 197.0919.



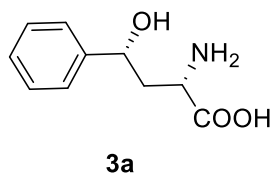
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 8.51 (s, 1H), 8.44 (d, $J = 5.0$, 1H), 7.86 (d, $J = 8.2$ Hz, 1H), 7.44 (dd, $J = 8.2, 5.0$ Hz, 1H), 4.95 (dd, $J = 9.3, 3.6$ Hz, 1H), 3.89 (dd, $J = 6.9, 4.2$ Hz, 1H), 2.26 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.0, 148.2, 146.4, 139.0, 135.0, 124.4, 68.7, 52.8, 37.5 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_9\text{H}_{13}\text{N}_2\text{O}_3$ calculated 197.0921, observed 197.0924.



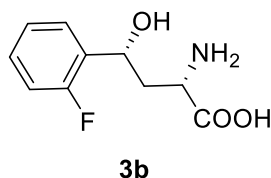
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 8.49 (d, J = 5.5, 2H), 7.45 (d, J = 5.5, 2H), 4.94 (t, J = 6.6 Hz, 1H), 3.89 (t, J = 5.7 Hz, 1H), 2.24 (m, 2H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) 174.0, 153.6, 148.8, 148.8, 121.2, 121.2, 69.5, 52.8, 37.3 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_9\text{H}_{13}\text{N}_2\text{O}_3$ calculated 197.0921, observed 197.0918.



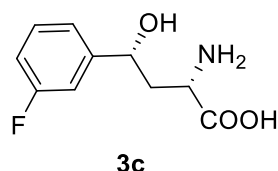
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.35 (m, 2H), 7.27 (m, 3H), 4.11 (m, 1H), 3.70 (m, 1H), 2.88 (dd, J = 13.8, 4.8 Hz, 1H), 2.72 (m, 1H), 2.11 (d, J = 14.9 Hz, 1H), 1.76 (m, 1H) ppm. ^{13}C NMR (150 MHz, D_2O , pD = 7.0) δ 196.2, 138.1, 129.5, 129.5, 128.6, 128.6, 126.6, 71.5, 54.3, 43.2, 37.2 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1128.



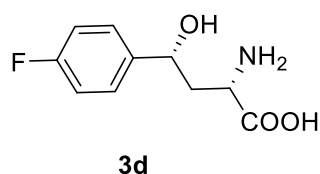
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.42 (m, 5H), 4.88 (dd, J = 8.4, 5.3 Hz, 1H), 3.48 (dd, J = 8.4, 5.3 Hz, 1H), 2.19 (dt, J = 14.4, 5.3 Hz, 1H), 2.03 (dt, J = 14.4, 8.4 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 178.9, 143.0, 128.8, 128.8, 128.0, 126.0, 126.0, 72.2, 54.0, 40.9 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{14}\text{NO}_3$ calculated 196.0968, observed 196.0965.



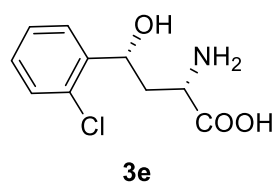
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.47 (m, 1H), 7.34 (m, 1H), 7.23 (t, J = 7.5 Hz, 1H), 7.13 (m, 1H), 5.15 (dd, J = 8.0, 5.9 Hz, 1H), 3.39 (m, 1H), 2.20 (m, 1H), 2.04 (dt, J = 14.9, 8.0 Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 180.2, 159.8 (d, $^1J_{\text{C-F}}$ = 244.5 Hz), 129.8 (d, $^3J_{\text{C-F}}$ = 8.1 Hz), 129.7 (d, $^2J_{\text{C-F}}$ = 12.6 Hz), 127.6 (d, $^3J_{\text{C-F}}$ = 4.6 Hz), 124.6 (d, $^4J_{\text{C-F}}$ = 3.5 Hz), 115.6 (d, $^2J_{\text{C-F}}$ = 21.8 Hz), 66.4 (d, $^3J_{\text{C-F}}$ = 2.9 Hz), 54.0, 40.6 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 8.0) δ -119.6 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{FNO}_3$ calculated 214.0874, observed 214.0872.



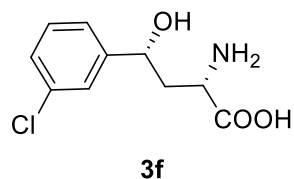
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.40 (m, 1H), 7.19 (m, 2H), 7.07 (t, J = 8.9 Hz, 1H), 4.99 (d, J = 9.4 Hz, 1H), 3.87 (m, 1H), 2.32 (m, 1H), 2.09 (dt, J = 16.5, 9.4 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 7.0) δ 174.2, 162.7 (d, $^1J_{\text{C-F}}$ = 234.1 Hz), 145.7 (d, $^3J_{\text{C-F}}$ = 7.0 Hz), 130.4 (d, $^3J_{\text{C-F}}$ = 8.3 Hz), 121.5 (d, $^4J_{\text{C-F}}$ = 2.6 Hz), 114.7 (d, $^2J_{\text{C-F}}$ = 21.2 Hz), 112.5 (d, $^2J_{\text{C-F}}$ = 22.3 Hz), 71.6, 53.8, 38.4 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 7.0) δ -113.4 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{FNO}_3$ calculated 214.0874, observed 214.0875. $[\alpha]_{\text{D}}^{20}$ = + 4.667 (c = 0.05, Methanol).



^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.40 (dd, J = 8.8, 5.6 Hz, 2H), 7.13 (t, J = 8.8 Hz, 1H), 4.85 (dd, J = 8.1, 5.7 Hz, 2H), 3.38 (m, 1H), 2.14 (dt, J = 14.3, 5.7 Hz, 1H), 2.00 (dt, J = 14.3, 8.1 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 179.9, 162.1 (d, $^1J_{\text{C-F}}$ = 234.1 Hz), 138.9 (s), 128.0 (d, $^3J_{\text{C-F}}$ = 8.4 Hz), 128.0 (d, $^3J_{\text{C-F}}$ = 8.4 Hz), 115.3 (d, $^2J_{\text{C-F}}$ = 21.6 Hz), 115.3 (d, $^2J_{\text{C-F}}$ = 21.6 Hz), 71.5, 54.0, 41.4 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 8.0) δ -115.3 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{ClNO}_3$ calculated 214.0874, observed 214.0868.

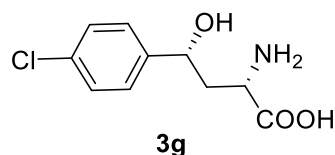


^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.56 (m, 1H), 7.40 (m, 2H), 7.29 (td, J = 7.8, 1.4 Hz, 1H), 5.31 (dd, J = 9.0, 4.0 Hz, 1H), 3.59 (t, J = 6.5 Hz, 1H), 2.22 (ddd, J = 14.5, 6.5, 4.0 Hz, 1H), 1.93 (m, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 178.6, 140.4, 131.4, 129.5, 129.1, 127.5, 126.9, 68.4, 53.9, 39.8 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{ClNO}_3$ calculated 230.0578, observed 230.0575.

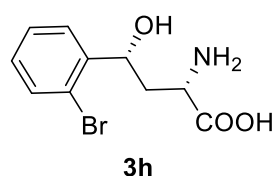


^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.43 (s, 1H), 7.34 (m, 3H), 4.79 (m, partially overlapped by solvent, 1H), 3.19 (m, 1H), 2.05 (dt, J = 14.0, 6.1 Hz, 1H), 1.92 (dt, J = 14.0, 7.8 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 182.5, 145.4, 133.8,

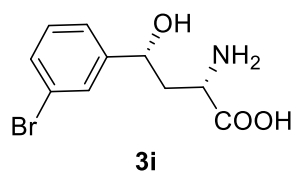
130.2, 127.8, 126.1, 124.6, 71.5, 54.0, 42.9 ppm. HRMS (ESI) (m/z) for [M+H]⁺ C₁₀H₁₃ClNO₃ calculated 230.0578, observed 230.0584.



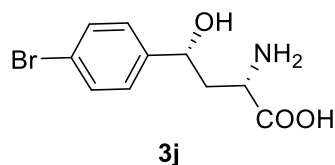
¹H NMR (400 MHz, D₂O, pD = 8.0) δ 7.41 (d, *J* = 8.3 Hz, 2H), 7.36 (d, *J* = 8.3 Hz, 2H), 4.85 (dd, *J* = 8.4, 5.5 Hz, 1H), 3.43 (m, 1H), 2.14 (dt, *J* = 14.0, 5.5 Hz, 1H), 1.99 (dt, *J* = 14.0, 8.4 Hz, 1H) ppm. ¹³C NMR (125 MHz, D₂O, pD = 8.0) δ 179.3, 141.8, 133.0, 128.7, 128.7, 127.7, 127.7, 71.6, 54.0, 41.2 ppm. HRMS (ESI) (m/z) for [M+H]⁺ C₁₀H₁₃ClNO₃ calculated 230.0578, observed 230.0580.



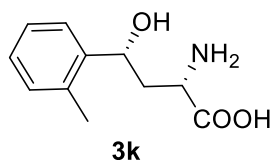
¹H NMR (400 MHz, D₂O, pD = 8.0) δ 7.60 (d, *J* = 8.0 Hz, 1H), 7.54 (m, 1H), 7.42 (t, *J* = 7.6 Hz, 1H), 7.22 (m, 1H), 5.23 (dd, *J* = 8.9, 4.4 Hz, 1H), 3.42 (t, *J* = 6.8 Hz, 1H), 2.13 (ddd, *J* = 14.2, 6.8, 4.4 Hz, 1H), 1.86 (m, 1H) ppm. ¹³C NMR (100 MHz, D₂O, pD = 8.0) δ 181.1, 142.1, 132.7, 129.4, 128.1, 127.2, 121.6, 70.6, 54.0, 41.5 ppm. HRMS (ESI) (m/z) for [M+H]⁺ C₁₀H₁₃BrNO₃ calculated 274.0073, observed 274.0074.



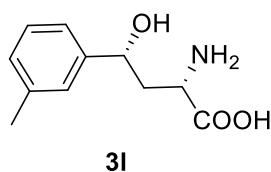
¹H NMR (400 MHz, D₂O, pD = 8.0) δ 7.59 (s, 1H), 7.51 (d, *J* = 7.7 Hz, 1H), 7.33 (m, 2H), 4.79 (m, 1H, partially overlapped with solvent), 3.24 (m, 1H), 2.07 (m, 1H), 1.93 (m, 1H) ppm. ¹³C NMR (100 MHz, D₂O, pD = 8.0) δ 181.9, 145.7, 130.8, 130.5, 129.1, 125.0, 122.0, 71.6, 54.0, 42.6 ppm. HRMS (ESI) (m/z) for [M+H]⁺ C₁₀H₁₃BrNO₃ calculated 274.0073, observed 274.0078.



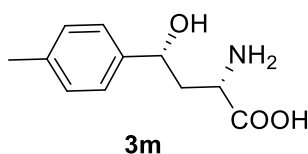
¹H NMR (400 MHz, D₂O, pD = 8.0) δ 7.56 (d, *J* = 8.8 Hz, 2H), 7.30 (d, *J* = 8.8 Hz, 2H), 4.75 (m, 1H, partially overlapped with solvent), 3.16 (dd, *J* = 7.7, 5.7 Hz, 1H), 2.04 (dt, *J* = 14.4, 5.7 Hz, 1H), 1.92 (dt, *J* = 14.4, 7.7 Hz, 1H) ppm. ¹³C NMR (125 MHz, D₂O, pD = 8.0) δ 182.7, 142.4, 131.6, 131.6, 128.2, 128.2, 121.0, 71.6, 54.1, 42.9. HRMS (ESI) (m/z) for [M+H]⁺ C₁₀H₁₃BrNO₃ calculated 274.0073, observed 274.0080.



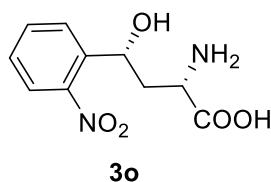
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.47 (d, J = 7.5 Hz, 1H), 7.25 (m, 3H), 5.19 (dd, J = 9.0, 3.3 Hz, 1H), 3.73 (m, 1H), 2.31 (s, 3H), 2.12 (m, 1H), 1.97 (dt, J = 15.2, 9.0 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 176.9, 141.4, 134.9, 130.5, 127.7, 126.4, 125.0, 68.4, 54.1, 39.3, 18.0 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1120.



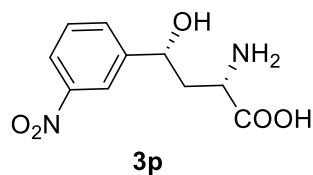
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.32 (t, J = 7.6 Hz, 1H), 7.25 (s, 1H), 7.20 (m, 2H), 4.75 (m, 1H, partially overlapped with solvent), 3.20 (t, J = 6.9 Hz, 1H), 2.33 (s, 3H), 2.06 (m, 1H), 1.95 (m, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 8.0) δ 182.4, 143.2, 138.8, 128.7, 128.5, 126.8, 123.2, 72.1, 54.0, 42.8, 20.4 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1122.



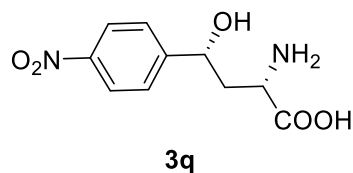
^1H NMR (400 MHz, D_2O , pD = 9.0) δ 7.30 (d, J = 8.1, 2H), 7.25 (d, J = 8.1 Hz, 2H), 4.76 (m, partially overlapped with solvent, 1H), 3.15 (dd, J = 8.0, 5.7 Hz, 1H), 2.31 (s, 3H), 2.05 (m, 1H), 1.94 (dt, J = 13.5, 8.0 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 9.0) δ 182.7, 140.0, 138.1, 129.2, 129.2, 126.3, 126.3, 71.9, 54.0, 42.9, 20.1 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{16}\text{NO}_3$ calculated 210.1125, observed 210.1130.



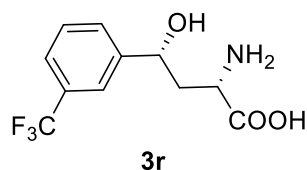
^1H NMR (400 MHz, D_2O , pD = 7.0) δ 8.01 (d, J = 8.3 Hz, 1H), 7.83 (m, 1H), 7.75 (m, 1H), 7.52 (m, 1H), 5.51 (d, J = 9.9 Hz, 1H), 3.96 (m, 1H), 2.46 (m, 1H), 2.08 (m, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 7.0) δ 174.1, 146.9, 138.8, 134.4, 128.9, 127.7, 124.7, 67.7, 54.2, 38.2 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{N}_2\text{O}_5$ calculated 241.0819, observed 241.0825.



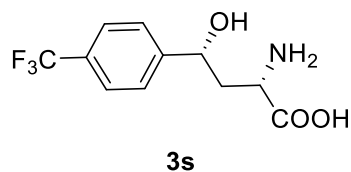
^1H NMR (400 MHz, D_2O , pD = 8.0) δ 8.26 (s, 1H), 8.17 (m, 1H), 7.79 (d, J = 7.7 Hz, 1H), 7.61 (t, J = 8.0 Hz, 1H), 5.01 (dd, J = 8.8, 4.9 Hz, 1H), 3.49 (m, 1H), 2.19 (dt, J = 14.3, 4.9 Hz, 1H), 2.02 (dt, J = 14.3, 8.4 Hz, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 179.5, 148.0, 145.3, 132.8, 129.8, 122.9, 121.0, 71.4, 54.0, 41.3 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{N}_2\text{O}_5$ calculated 241.0819, observed 241.0822.



^1H NMR (400 MHz, D_2O , pD = 9.0) δ 8.23 (d, J = 8.4 Hz, 2H), 7.59 (d, J = 8.4 Hz, 2H), 4.93 (m, 1H), 3.24 (t, J = 6.9 Hz, 1H), 2.07 (dt, J = 13.8, 6.9 Hz, 1H), 1.94 (dt, J = 13.8, 6.9 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 9.0) 182.5, 151.2, 147.0, 127.0, 127.0, 123.9, 123.9, 71.3, 54.0, 43.0 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{10}\text{H}_{13}\text{N}_2\text{O}_5$ calculated 241.0819, observed 241.0823.

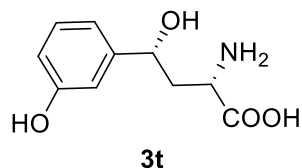


^1H NMR (400 MHz, D_2O , pD = 9.0) δ 7.72 (s, 1H), 7.64 (m, 2H), 7.56 (t, J = 7.8 Hz, 1H), 4.92 (t, J = 6.6 Hz, 1H), 3.36 (t, J = 6.6 Hz, 1H), 2.13 (m, 1H), 1.98 (m, 1H) ppm. ^{13}C NMR (125 MHz, D_2O , pD = 9.0) δ 180.7, 144.3, 130.1 (q, $^2J_{\text{C-F}}$ = 32.0 Hz), 129.8, 129.4, 124.7 (q, $^3J_{\text{C-F}}$ = 4.0 Hz), 124.2 (q, $^1J_{\text{C-F}}$ = 271.6 Hz), 122.8 (q, $^3J_{\text{C-F}}$ = 4.0 Hz), 71.7, 54.1, 42.0 ppm. ^{19}F NMR (376 MHz, D_2O , pD = 9.0) δ -62.4 ppm. HRMS (ESI) (m/z) for $[\text{M}+\text{H}]^+$ $\text{C}_{11}\text{H}_{13}\text{F}_3\text{NO}_3$ calculated 264.0842, observed 264.0846.

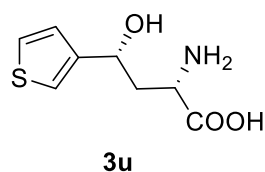


^1H NMR (400 MHz, D_2O , pD = 8.0) δ 7.72 (d, J = 8.0 Hz, 2H), 7.55 (d, J = 8.0 Hz, 2H), 4.88 (t, J = 7.0 Hz, 1H), 3.19 (t, J = 7.0 Hz, 1H), 2.06 (dt, J = 14.2, 7.0 Hz, 1H), 1.94 (dt, J = 14.2, 7.0 Hz, 1H). ^{13}C NMR (125 MHz, D_2O , pD = 8.0) δ 182.6, 147.5, 129.2 (q, $^2J_{\text{C-F}}$ = 32.1 Hz), 126.6, 126.6, 125.6 (q, $^3J_{\text{C-F}}$ = 3.9 Hz), 125.6 (q, $^3J_{\text{C-F}}$ = 3.9 Hz), 124.2 (q, $^1J_{\text{C-F}}$ = 271.3 Hz), 71.6, 54.1, 43.0 ppm. ^{19}F NMR (376 MHz, D_2O , pD =

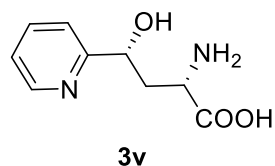
8.0) δ -62.3 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{11}H_{13}F_3NO_3$ calculated 264.0842, observed 264.0845.



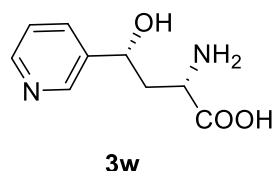
1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.28 (t, J = 7.9 Hz, 1H), 6.95 (d, J = 7.9 Hz, 1H), 6.89 (s, 1H), 6.82 (m, 1H), 4.90 (m, 1H), 3.84 (dd, J = 8.6, 4.7 Hz, 1H), 2.28 (dt, J = 15.0, 4.7 Hz, 1H), 2.08 (dt, J = 15.0, 8.6 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 7.0) δ 174.2, 155.6, 145.1, 130.2, 117.7, 114.9, 112.5, 71.9, 53.8, 38.5 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_{10}H_{14}NO_4$ calculated 212.0917, observed 212.0913.



1H NMR (400 MHz, D_2O , pD = 7.0) δ 7.47 (m, 1H), 7.39 (s, 1H), 7.17 (m, 1H), 5.06 (dd, J = 9.1, 3.7 Hz, 1H), 3.84 (m, 1H), 2.41 (m, 1H), 2.18 (dt, J = 14.9, 9.1 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 7.0) δ 174.2, 144.0, 127.1, 125.5, 121.9, 68.1, 53.7, 37.8 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_8H_{12}NO_3S$ calculated 202.0532, observed 202.0530.

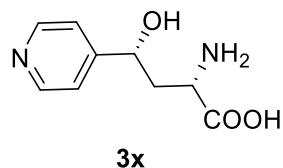


1H NMR (400 MHz, D_2O , pD = 7.0) δ 8.46 (d, J = 4.9 Hz, 1H), 7.88 (td, J = 7.9, 2.0 Hz, 1H), 7.54 (d, J = 7.9 Hz, 1H), 7.37 (m, 1H), 5.04 (m, 1H), 3.92 (dd, J = 9.1, 4.4 Hz, 1H), 2.40 (dt, J = 14.6, 4.4 Hz, 1H), 2.12 (dt, J = 14.6, 9.1 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 7.0) δ 174.2, 160.7, 148.2, 138.6, 123.5, 120.9, 72.4, 53.7, 37.1 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_9H_{13}N_2O_3$ calculated 197.0921, observed 197.0919.



1H NMR (400 MHz, D_2O , pD = 7.0) δ 8.53 (s, 1H), 8.45 (d, J = 4.8 Hz, 1H), 7.89 (d, J = 8.0, 1H), 7.46 (dd, J = 8.0, 4.8 Hz, 1H), 5.07 (dd, J = 10.2, 4.0 Hz, 1H), 3.91 (dd, J = 8.5, 4.0 Hz, 1H), 2.33 (dt, J = 14.9, 4.0 Hz, 1H), 2.14 (ddd, J = 14.9, 10.2, 8.5 Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , pD = 7.0) δ 174.1, 148.4, 146.2, 139.1, 135.1, 124.3,

70.0, 53.9, 38.2 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_9H_{13}N_2O_3$ calculated 197.0921, observed 197.0918.



1H NMR (400 MHz, D_2O , $pD = 7.0$) δ 8.49 (d, $J = 4.9$ Hz, 2H), 7.45 (d, $J = 4.9$ Hz, 2H), 5.04 (m, 1H), 3.91 (dd, $J = 8.3, 5.2$ Hz, 1H), 2.33 (m, 1H), 2.04 (dt, $J = 14.8, 8.3$ Hz, 1H) ppm. ^{13}C NMR (100 MHz, D_2O , $pD = 7.0$) 174.1, 153.5, 148.8, 148.8, 121.1, 121.1, 70.8, 53.8, 38.1 ppm. HRMS (ESI) (m/z) for $[M+H]^+$ $C_9H_{13}N_2O_3$ calculated 197.0921, observed 197.0916.

Preparative scale of ApUstD

For preparative scale product of ApUstD, 62.5 ml reactions containing 0.4 mg/ml ApUstD, 100 mg L -Aspartic acid, 80 mM **1c**, 1 mM DTT, 100 μ M PLP and 50 mM sodium phosphate, 50 mM NaCl, at pH 7.4 were run at 25 $^{\circ}C$. After 72 hours, the reaction mixture was centrifugated (800 g), and then the supernatant loaded on C18 chromatography eluted with methanol in water in a stepwise manner (0%, 20%, 40%, 60%, 95%) and Fractions containing product were collected and the solvent was evaporated under vacuum to yield **2c** (134 mg, 84%).

Supplementary Figure S7-S88

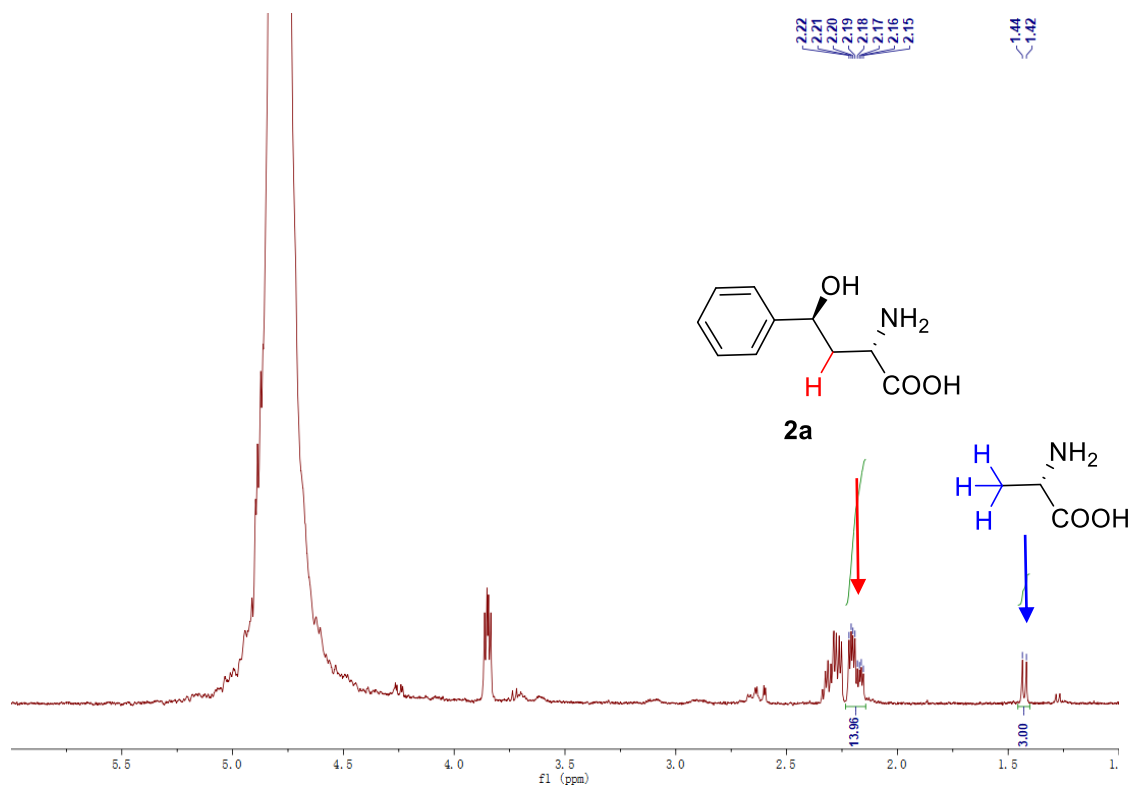


Figure S7. Conversion of 2a

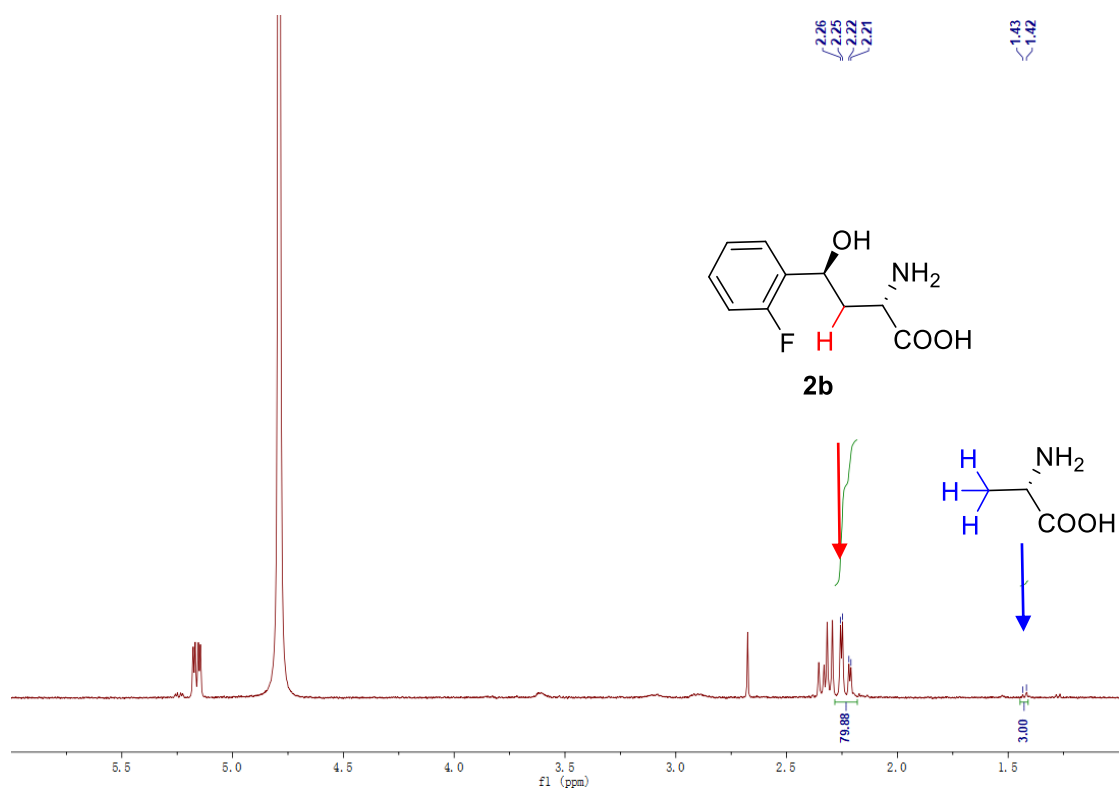


Figure S8. Conversion of 2b

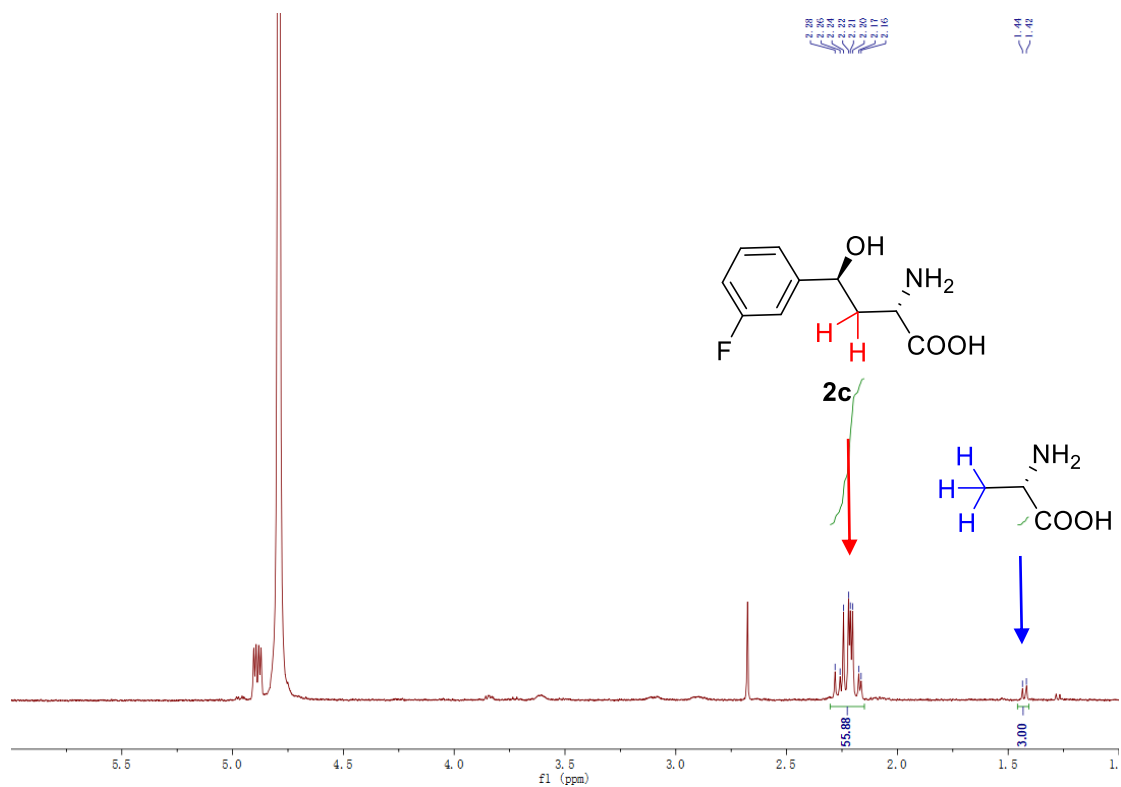


Figure S9. Conversion of 2c

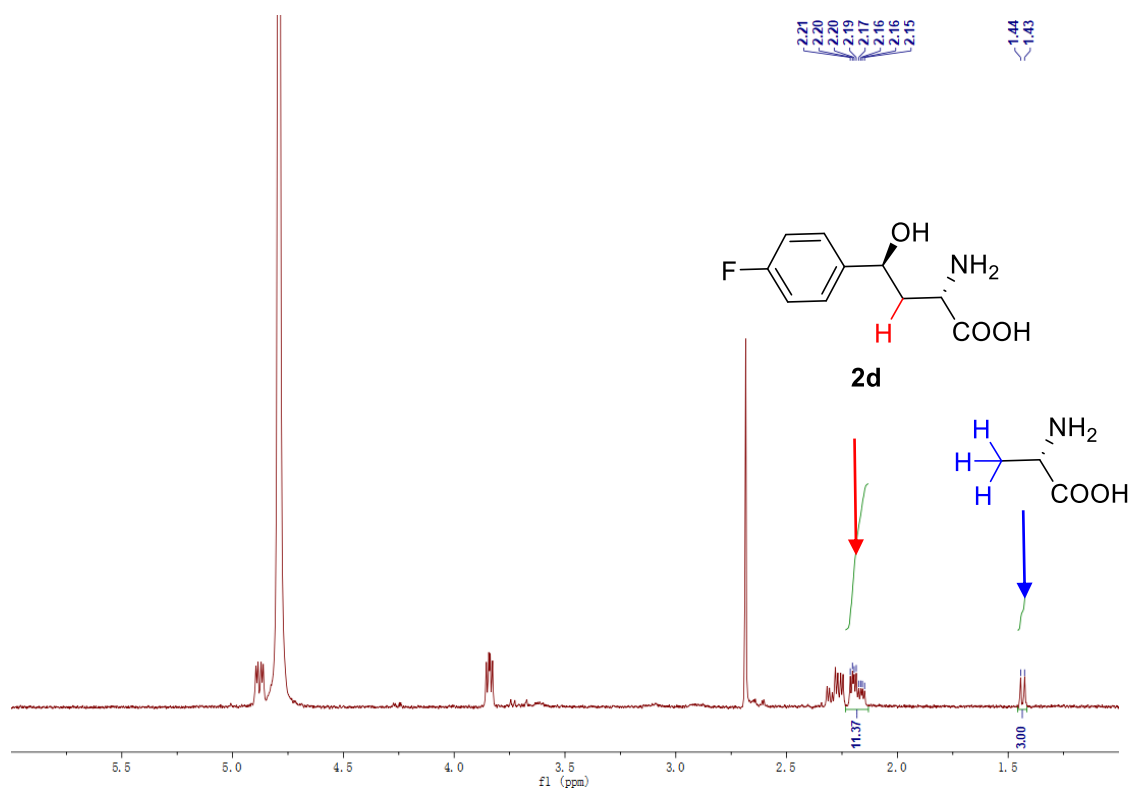


Figure S10. Conversion of 2d

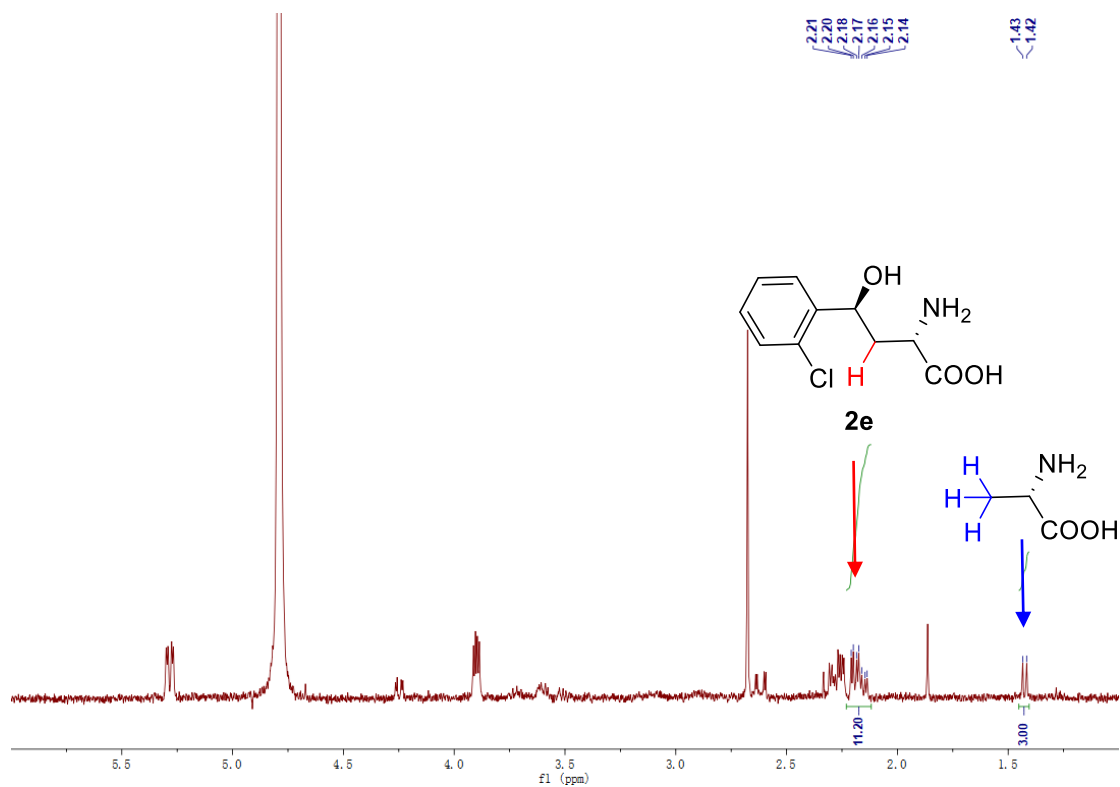


Figure S11. Conversion of **2e**

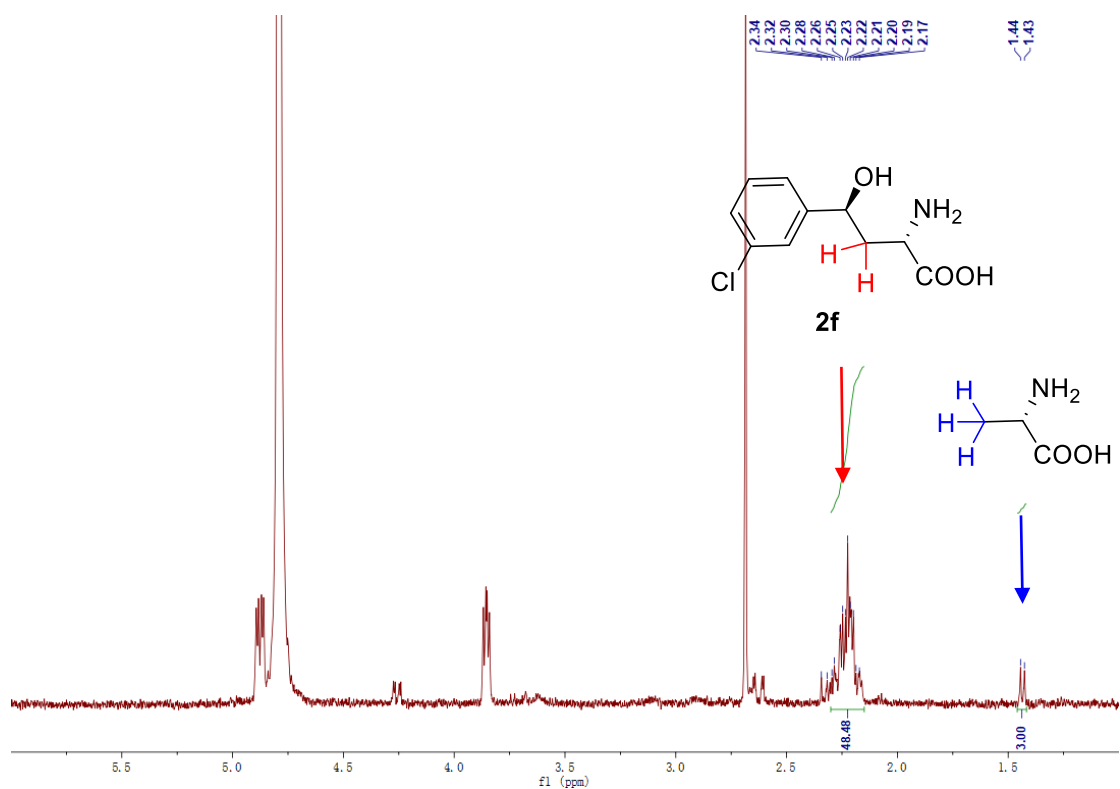


Figure S12. Conversion of **2f**

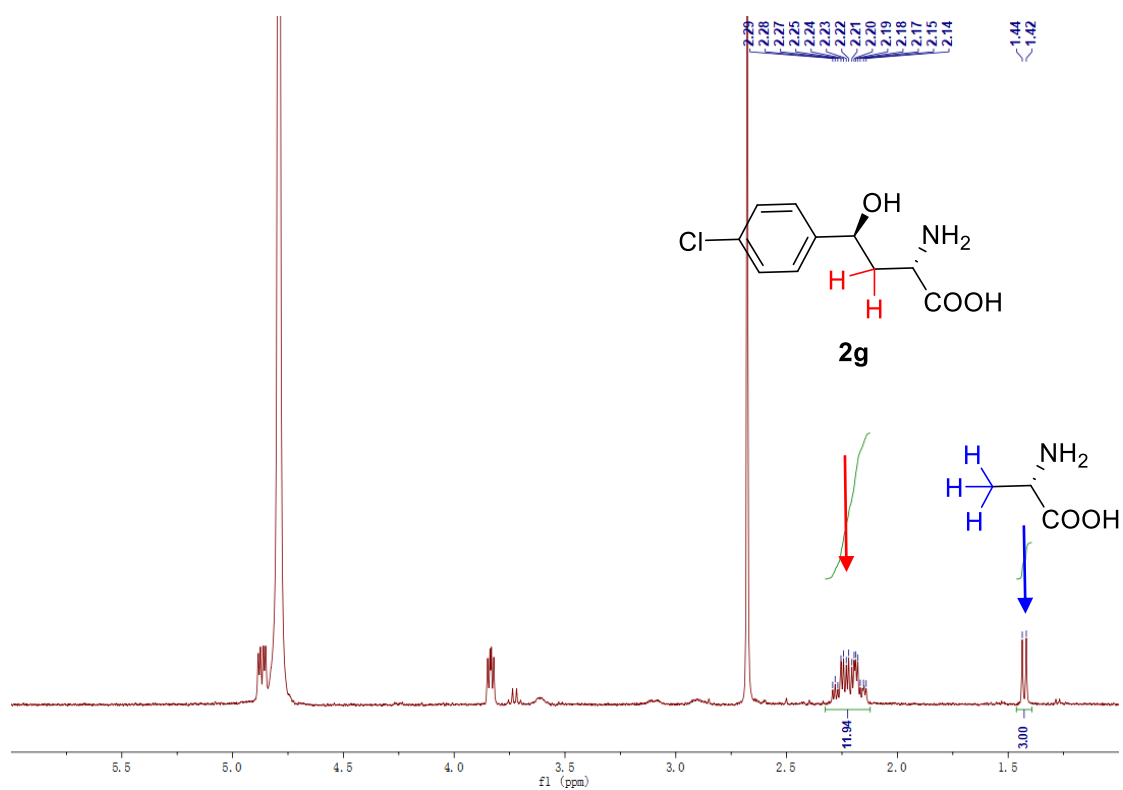


Figure S13. Conversion of **2g**

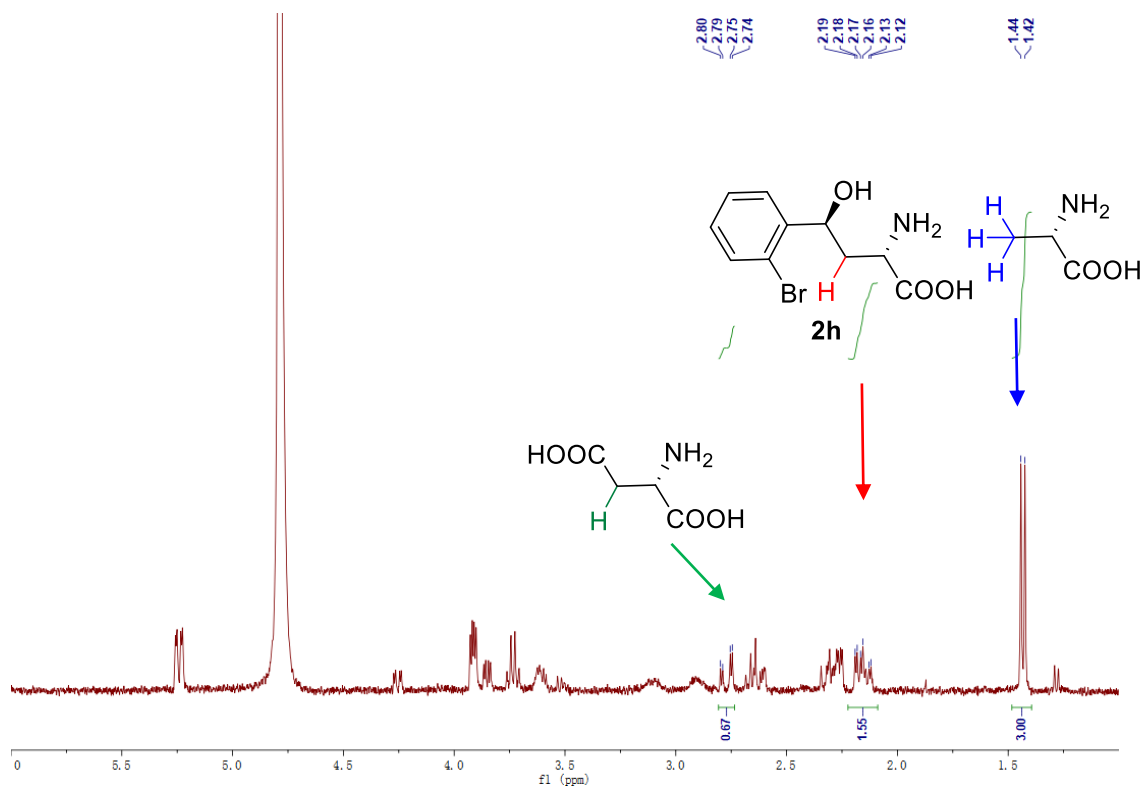


Figure S14. Conversion of **2h**

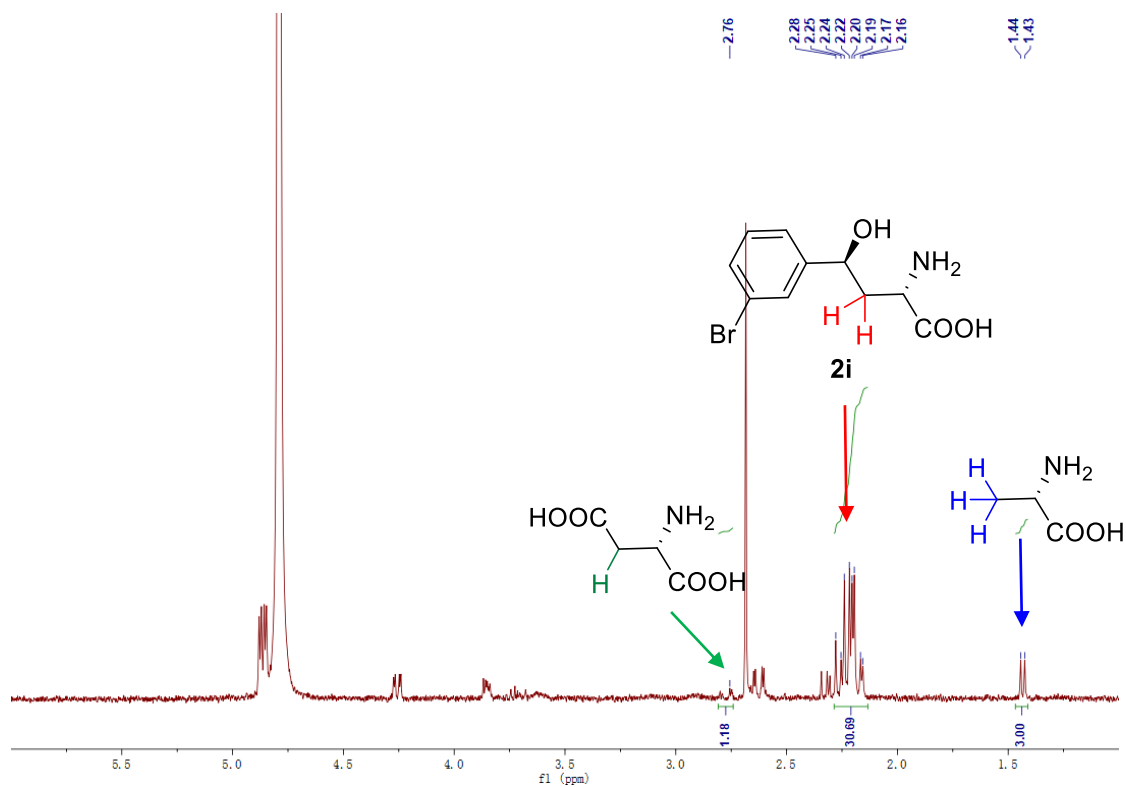


Figure S15. Conversion of **2i**

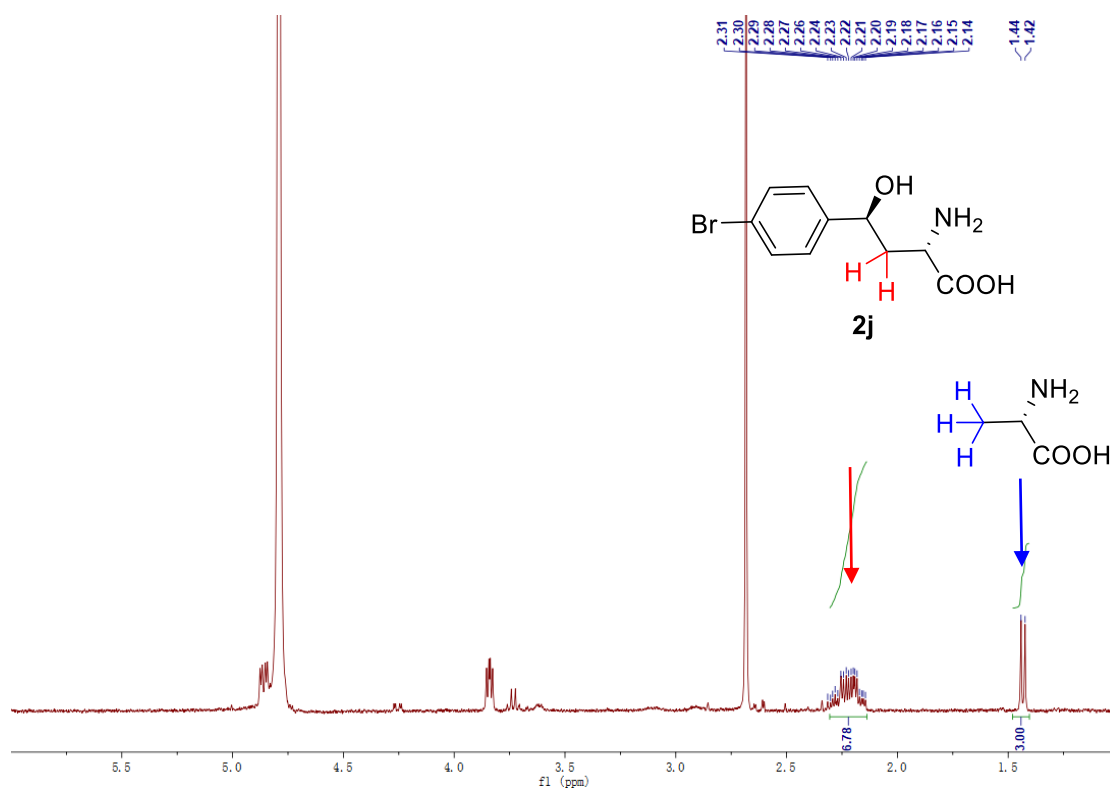


Figure S16. Conversion of **2j**

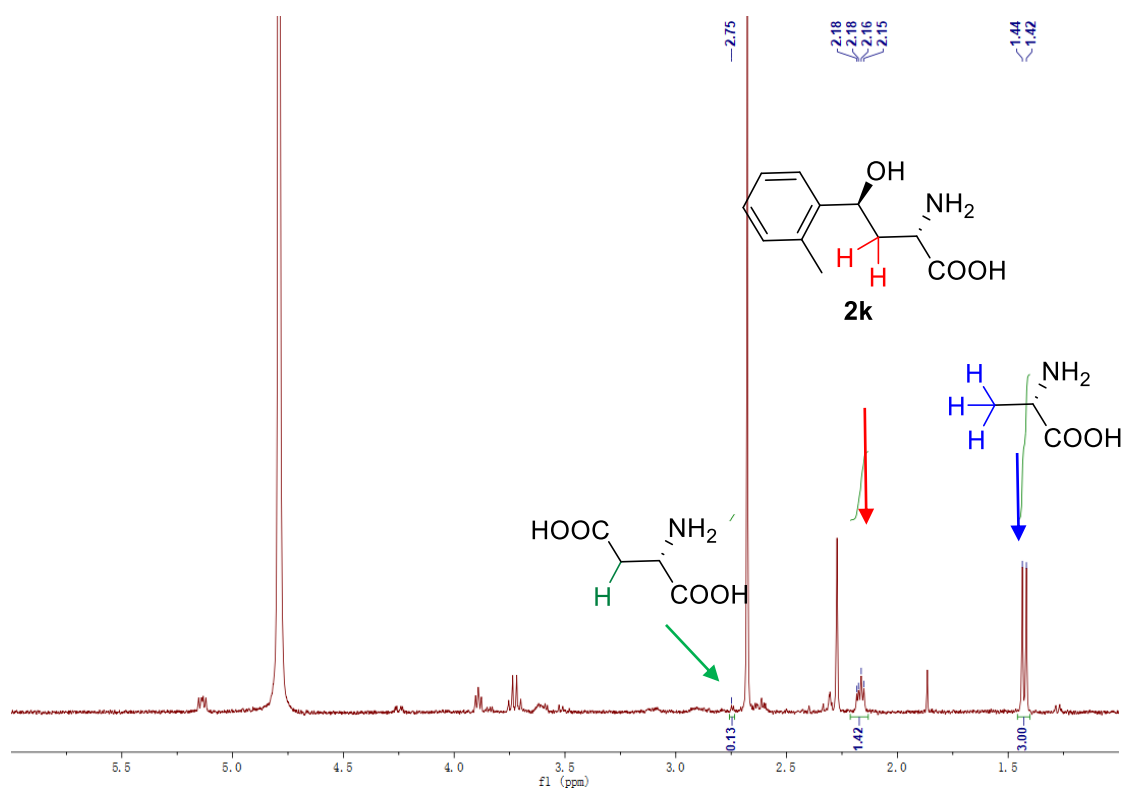


Figure S17. Conversion of **2k**

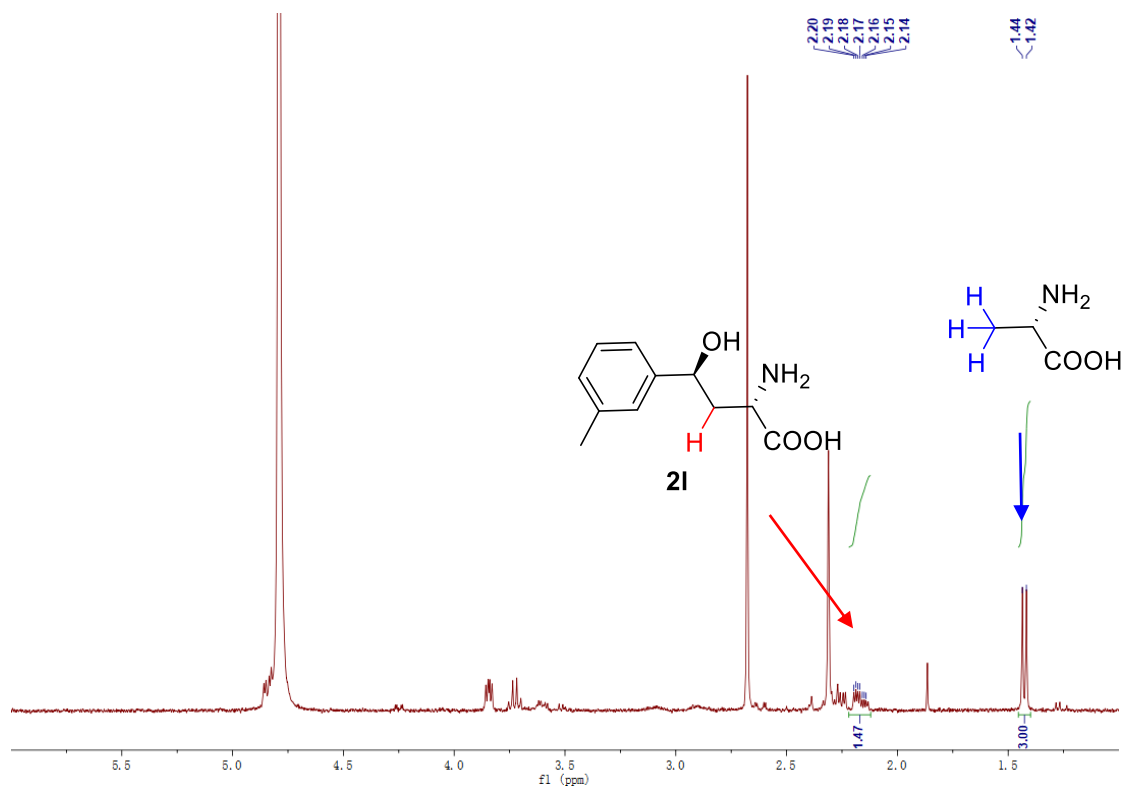


Figure S18. Conversion of **2l**

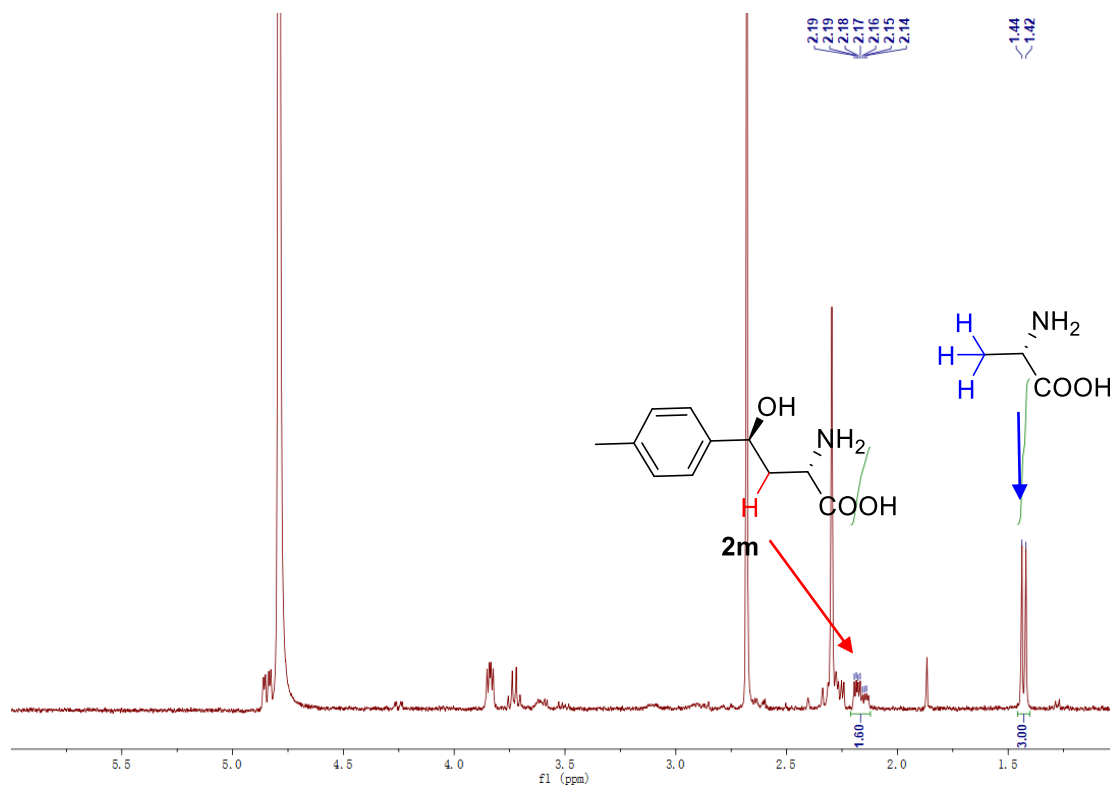


Figure S19. Conversion of **2m**

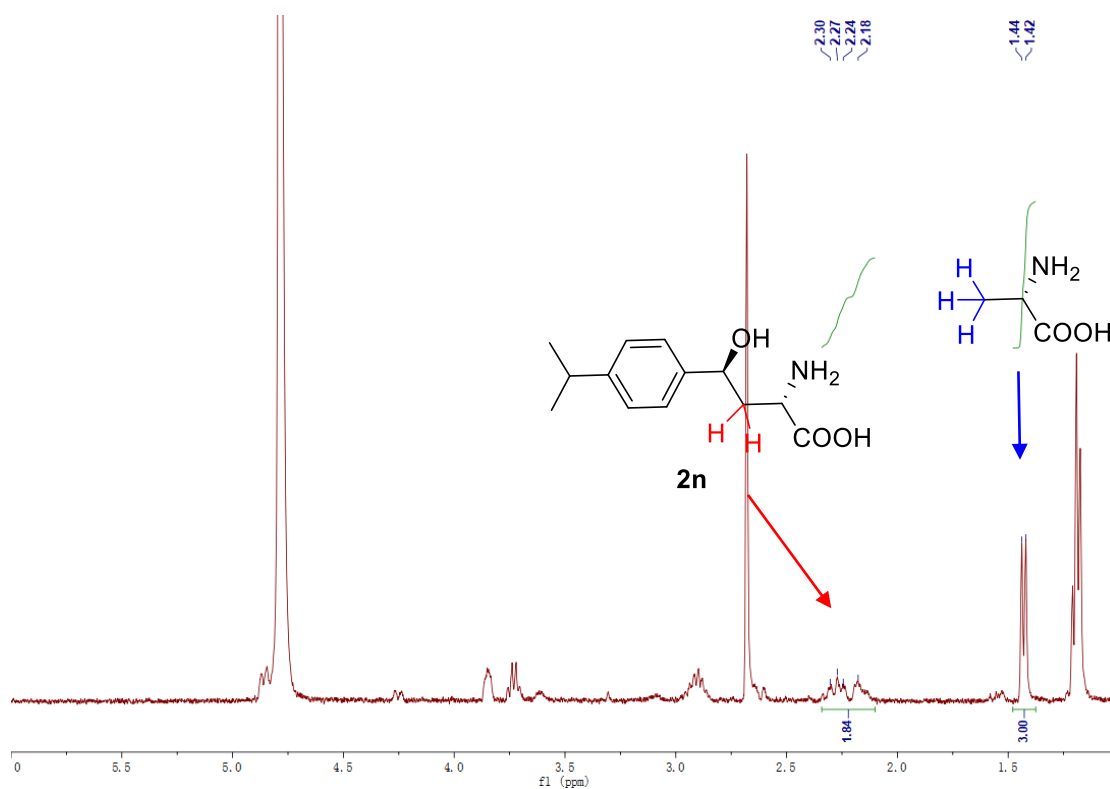


Figure S20. Conversion of **2n**

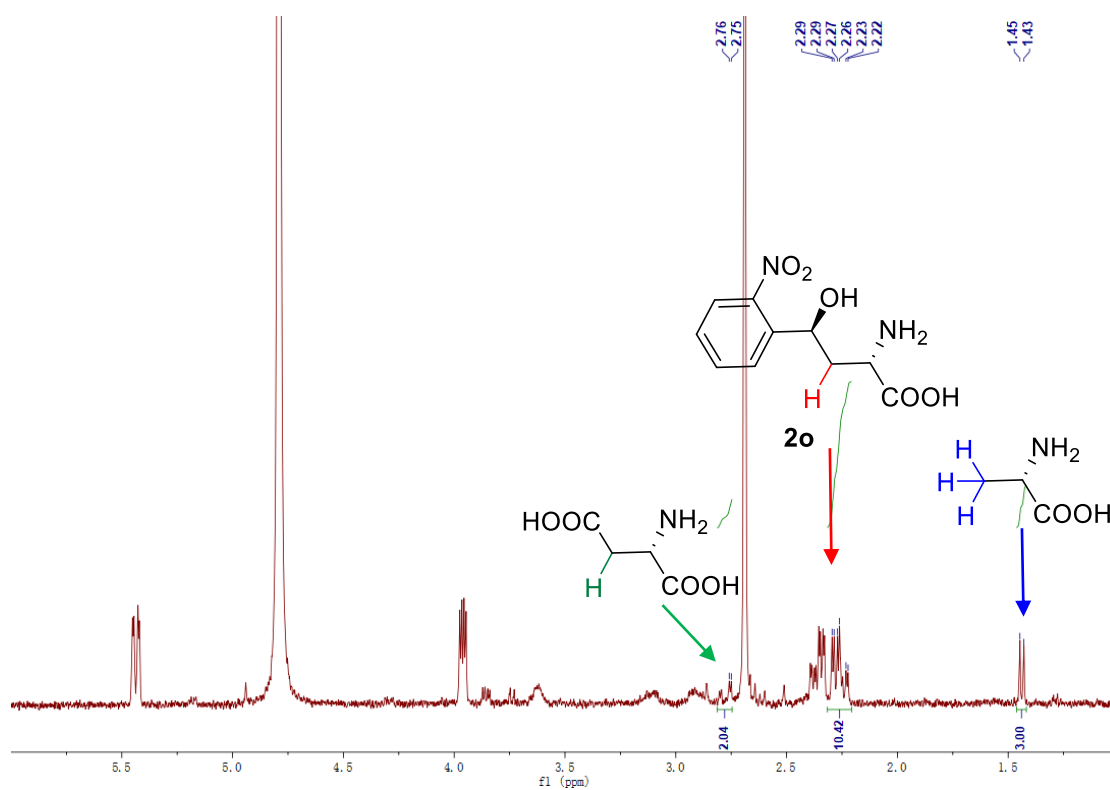


Figure S21. Conversion of **2o**

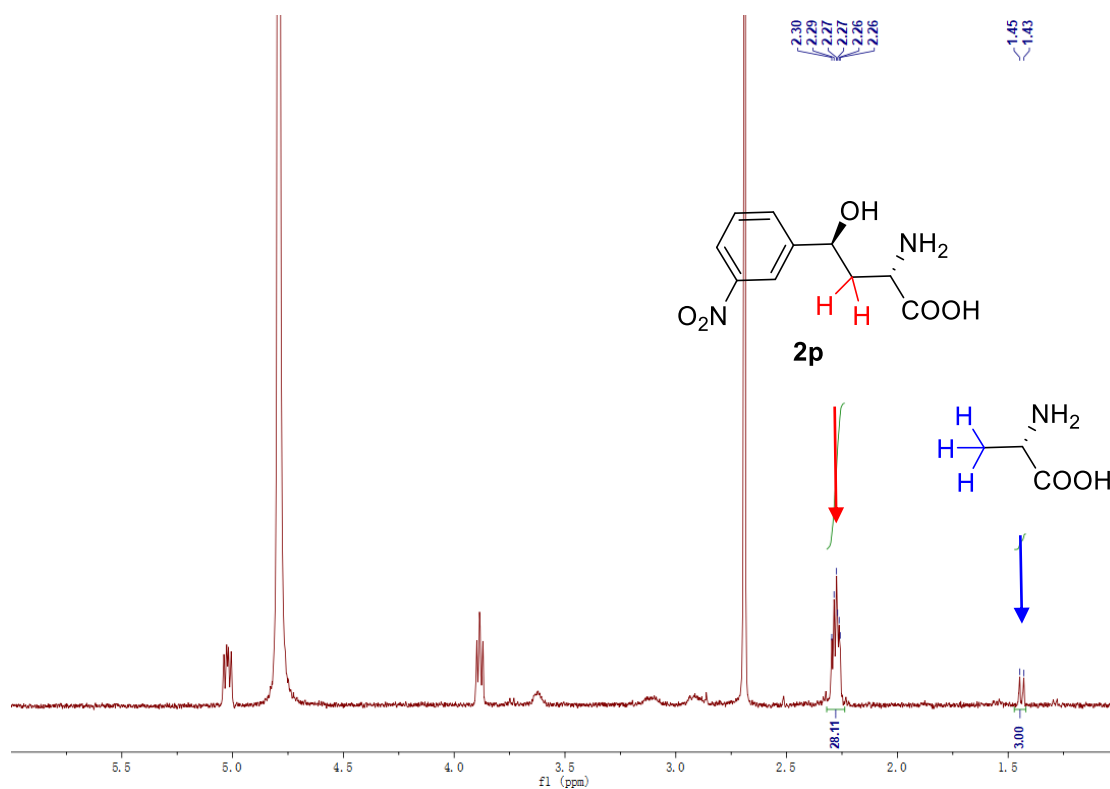


Figure S22. Conversion of **2p**

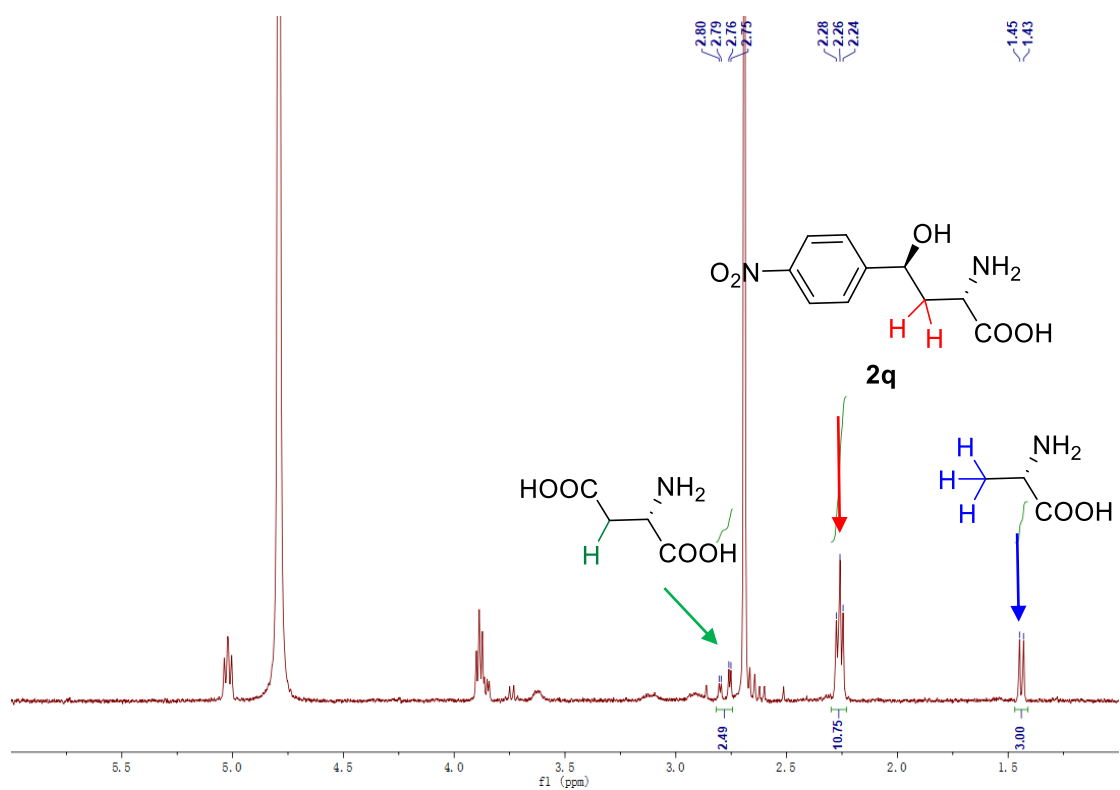


Figure S23. Conversion of 2q

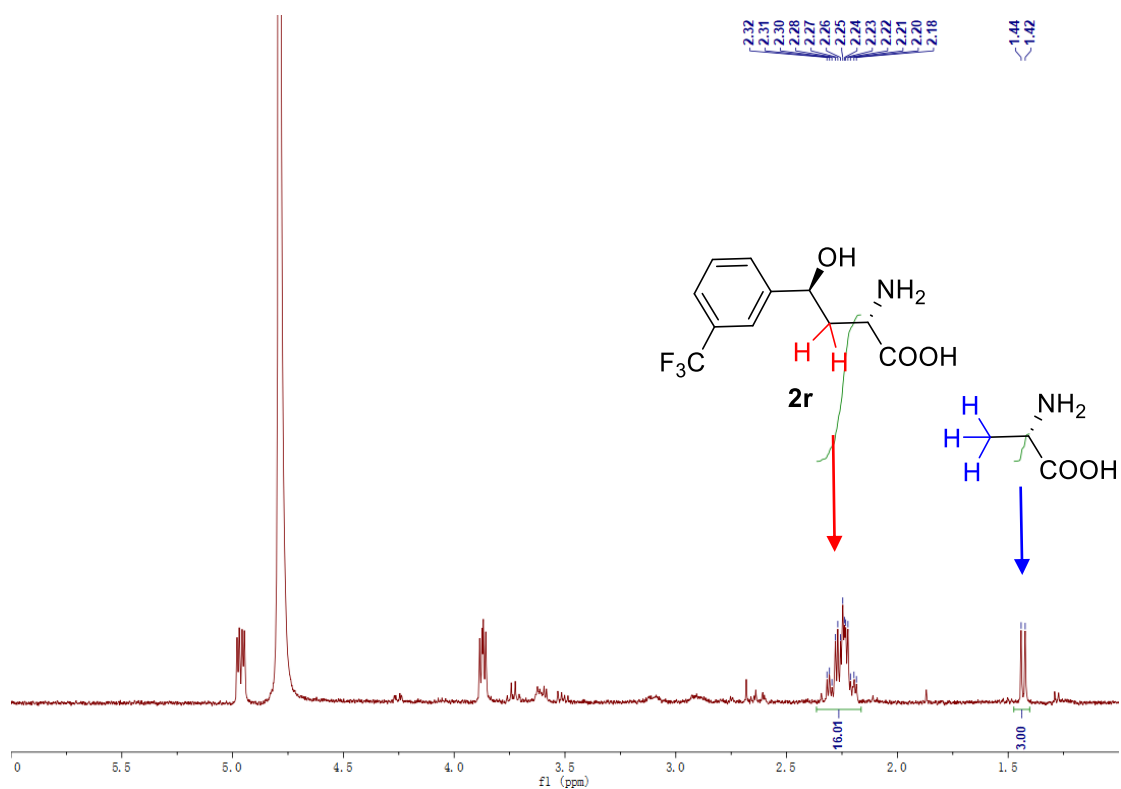
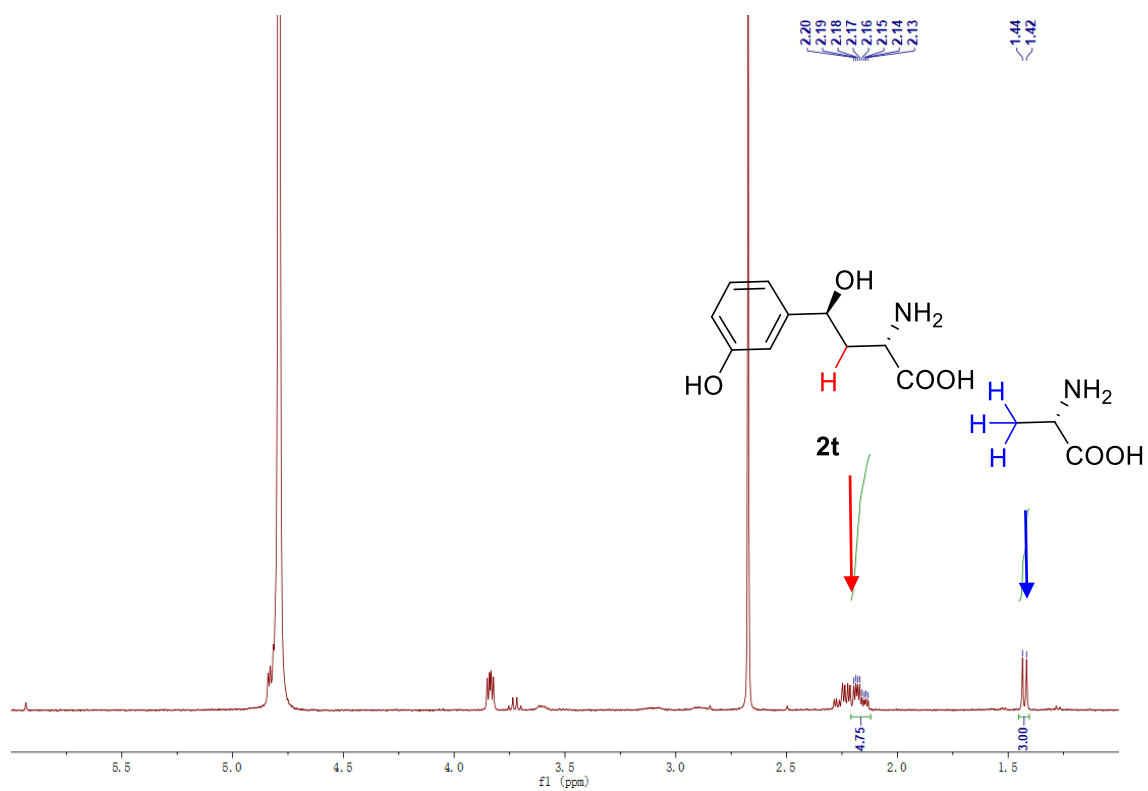
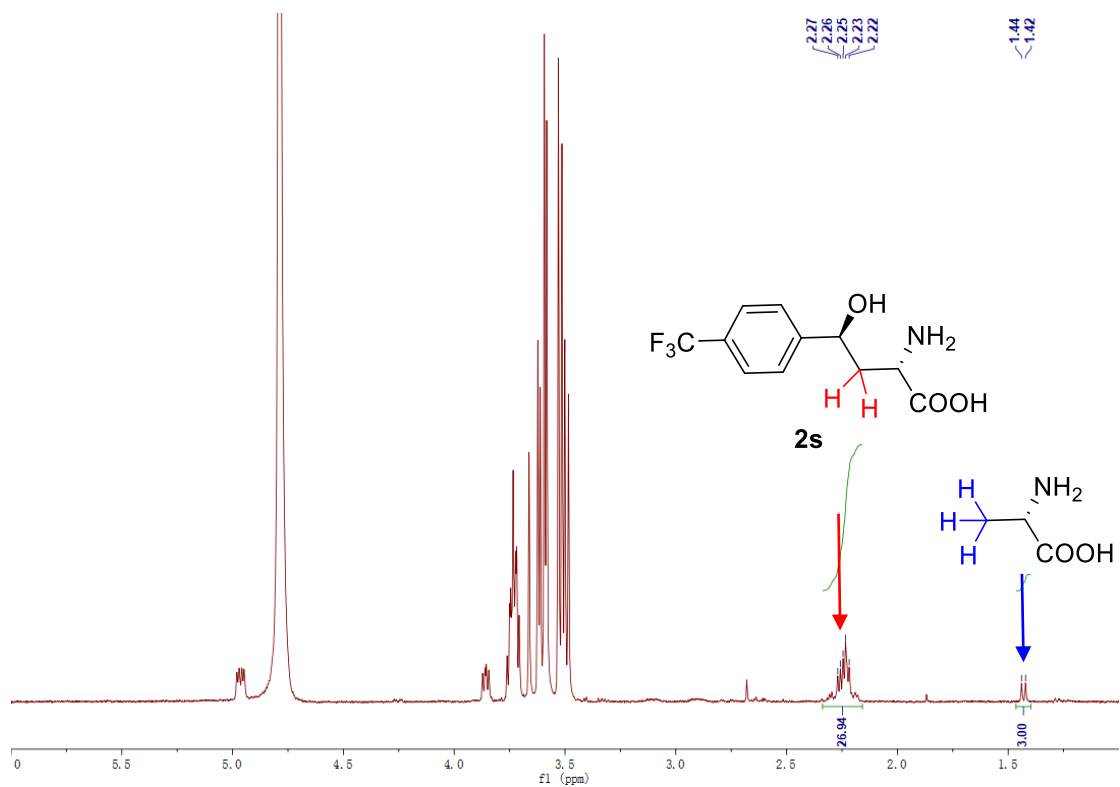


Figure S24. Conversion of 2r



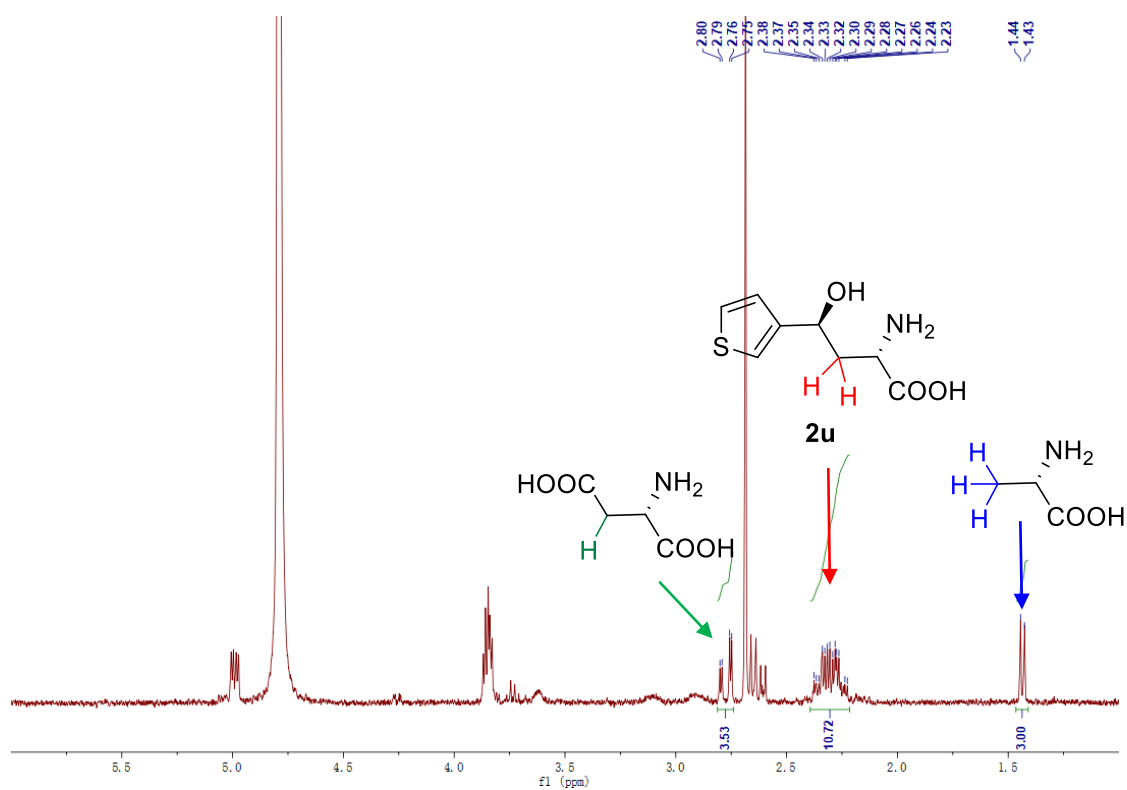


Figure S27. Conversion of **2u**

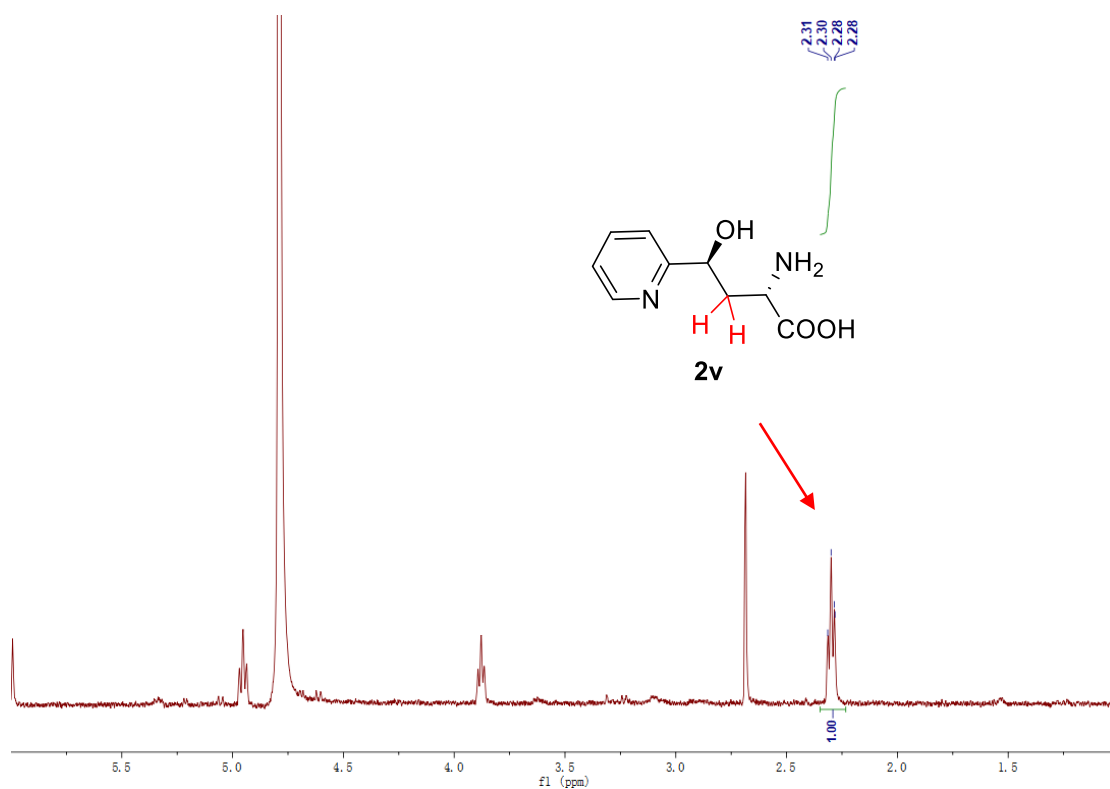


Figure S28. Conversion of **2v**

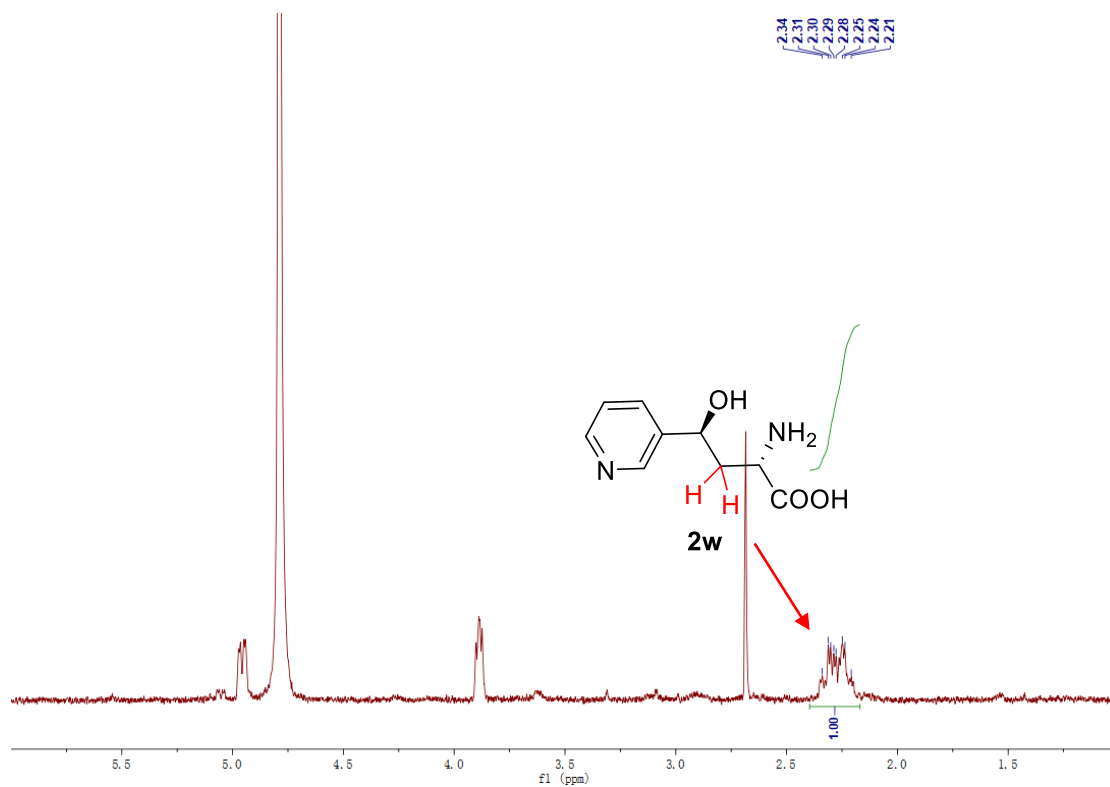


Figure S29. Conversion of **2w**

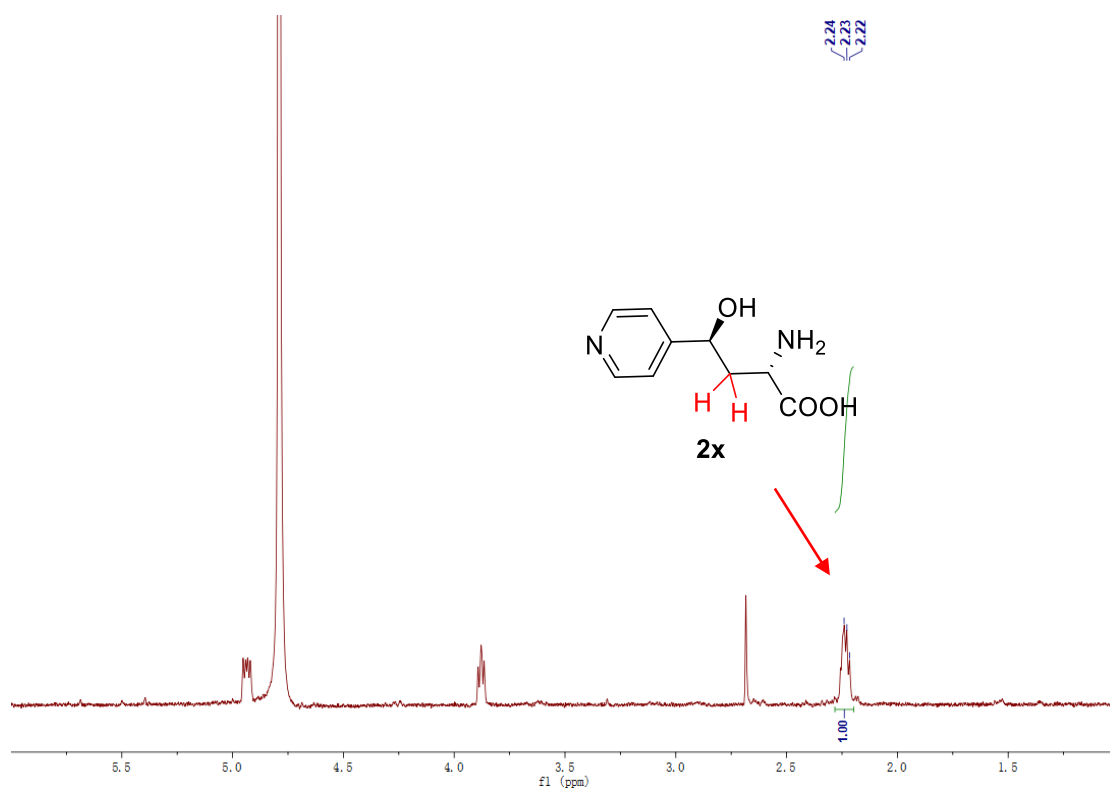


Figure S30. Conversion of **2x**

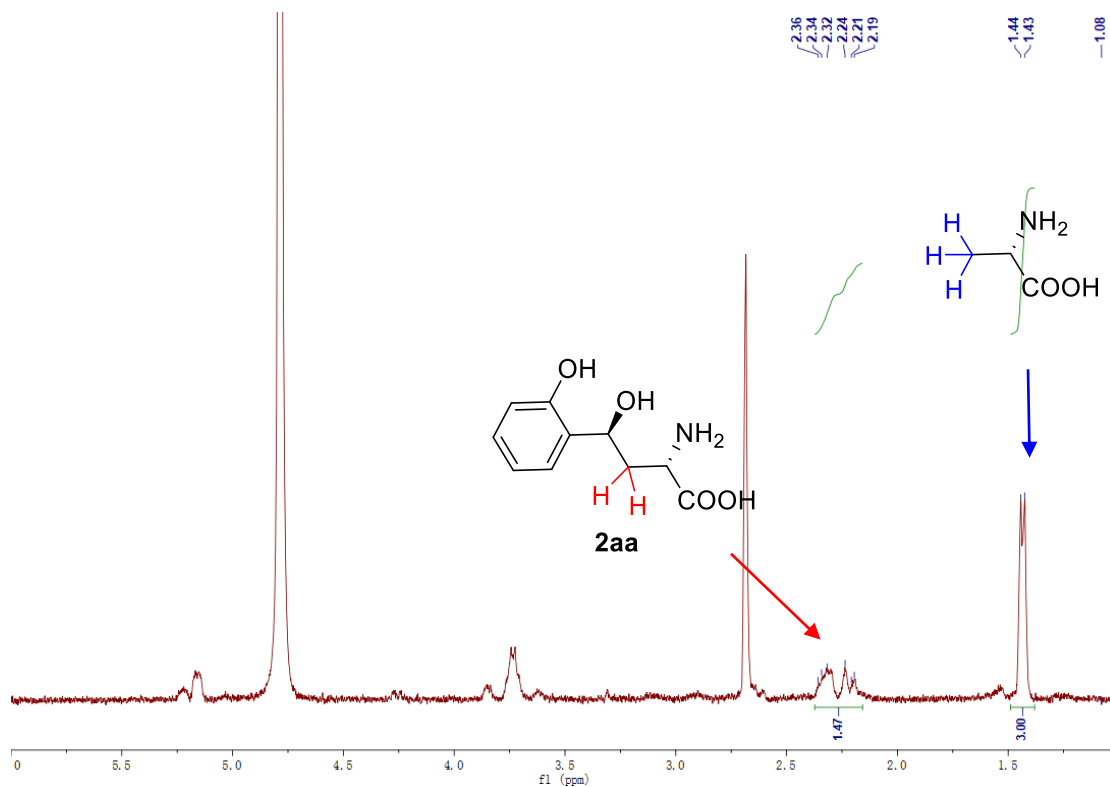


Figure S31. Conversion of **2aa**

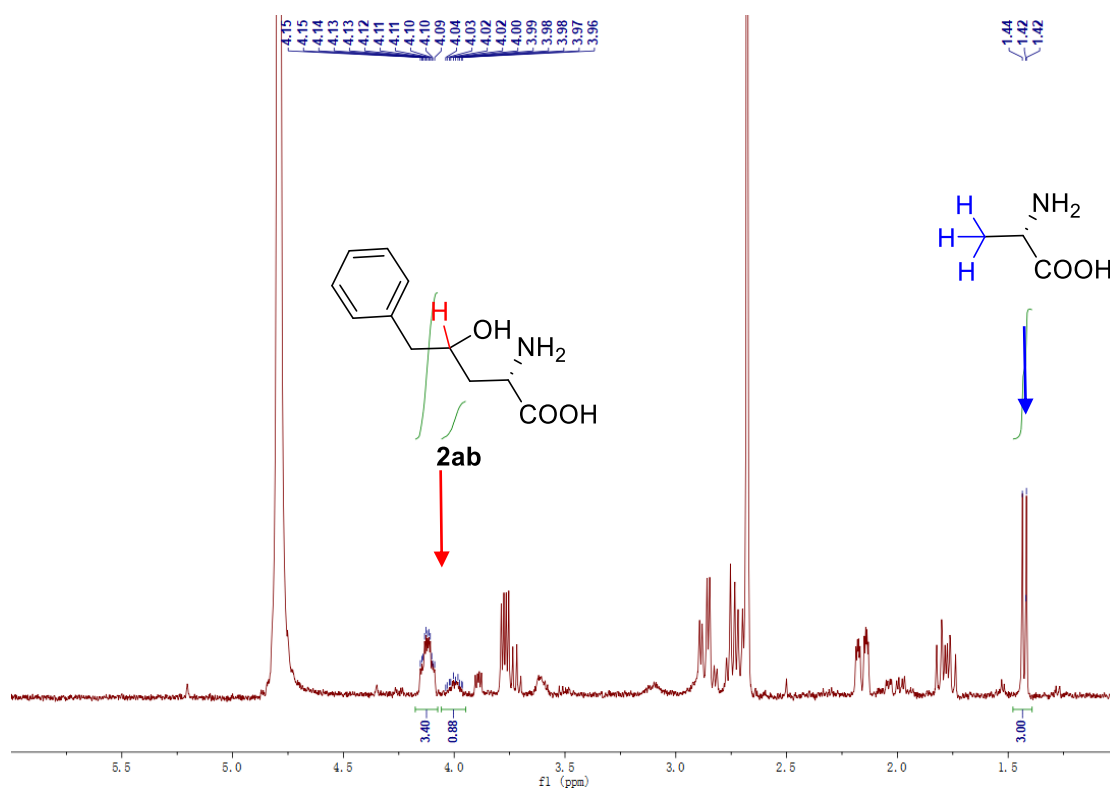


Figure S32. Conversion of **2ab**

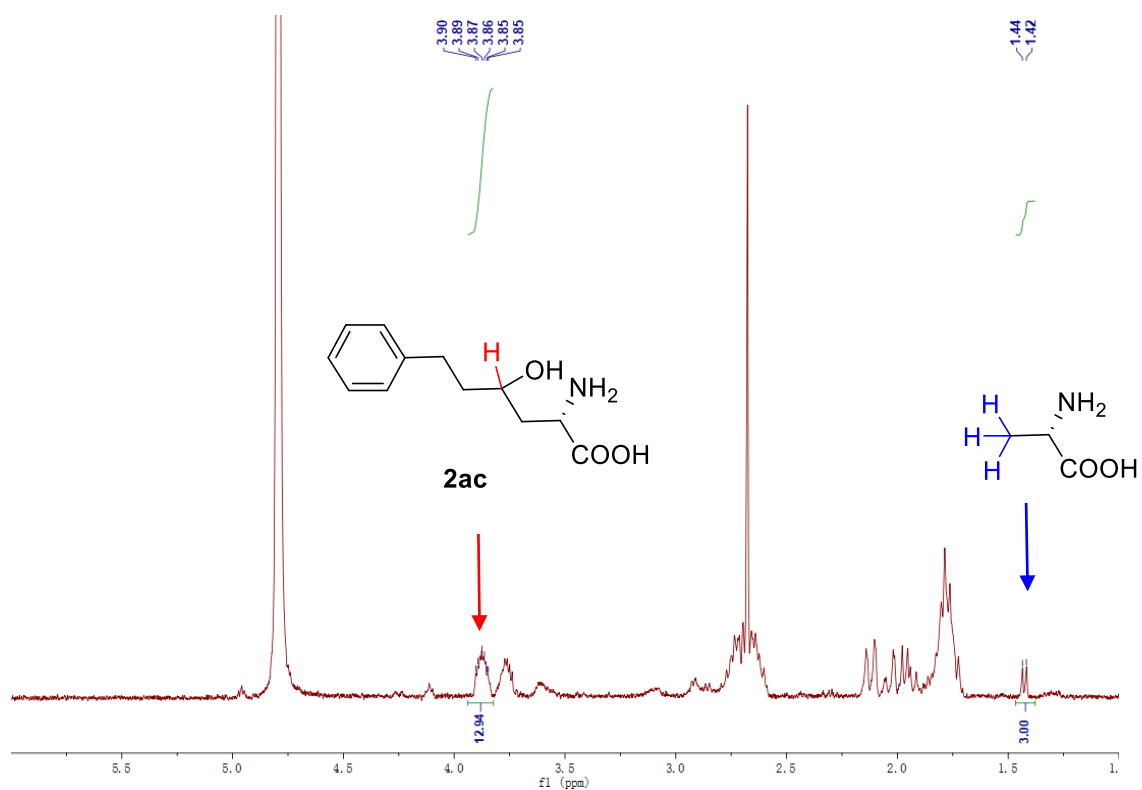


Figure S33. Conversion of 2ac

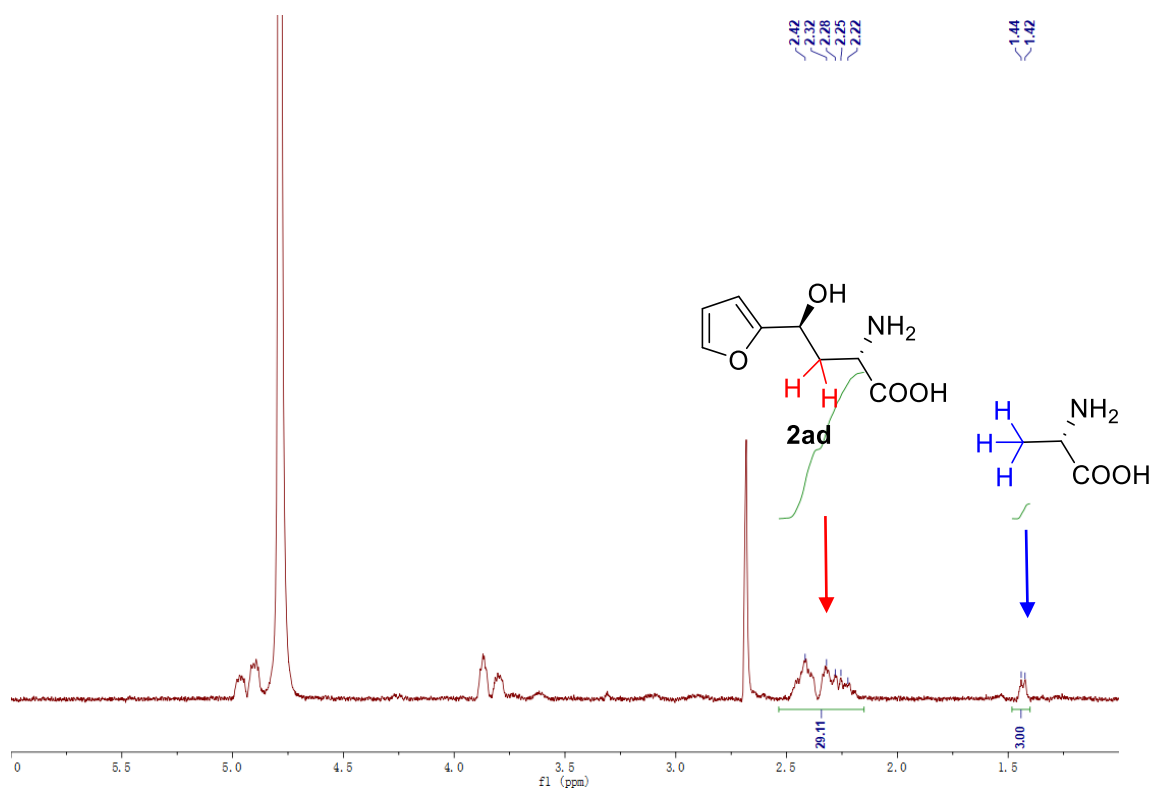


Figure S34. Conversion of 2ad

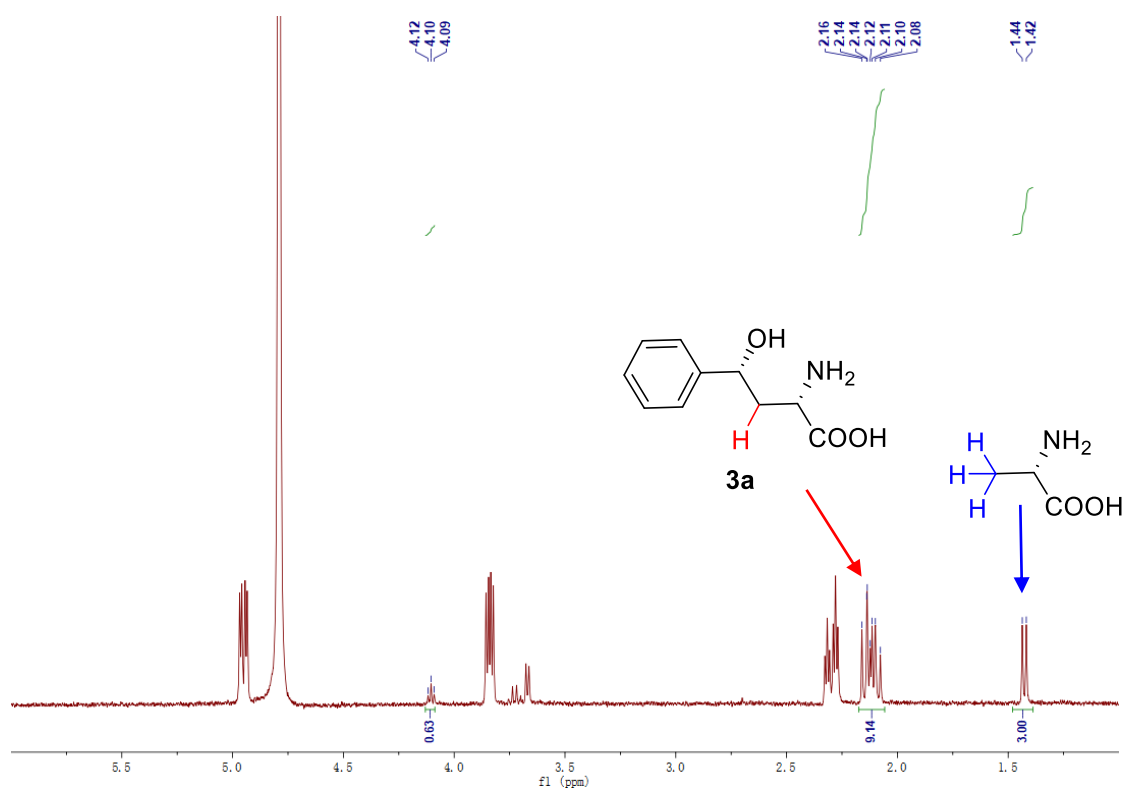


Figure S35. Conversion of **3a**

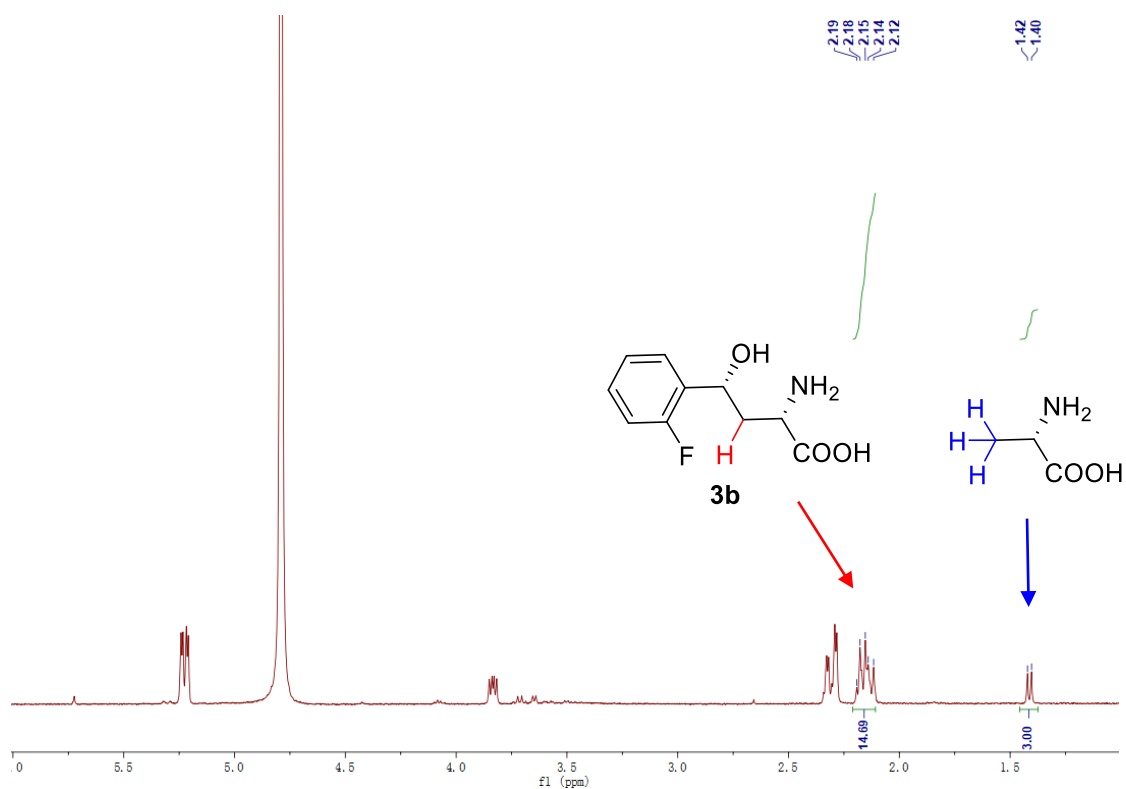


Figure S36. Conversion of **3b**

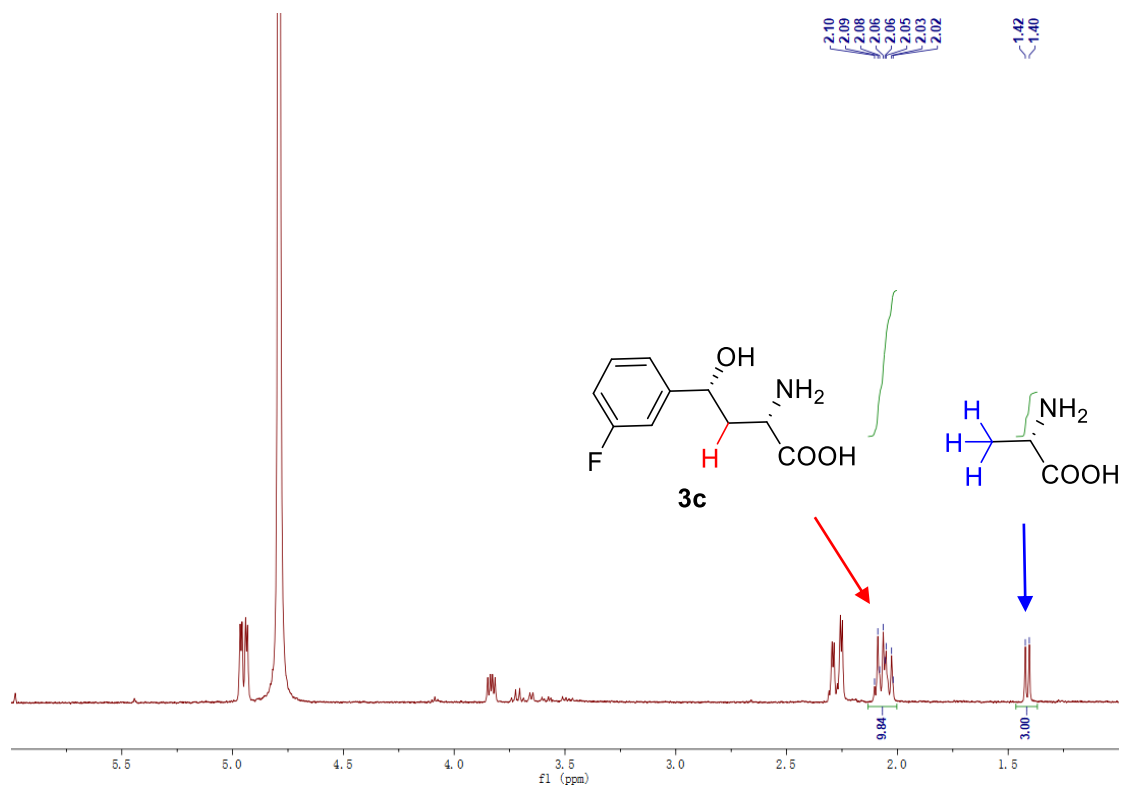


Figure S37. Conversion of **3c**

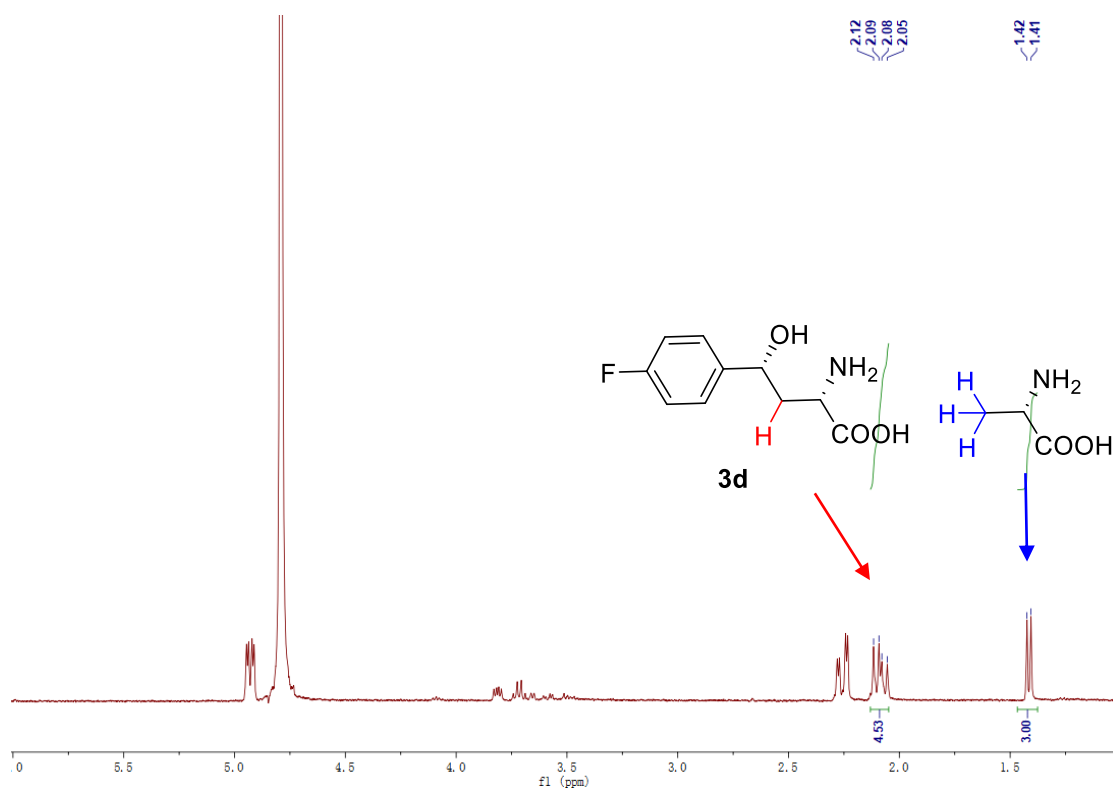


Figure S38. Conversion of **3d**

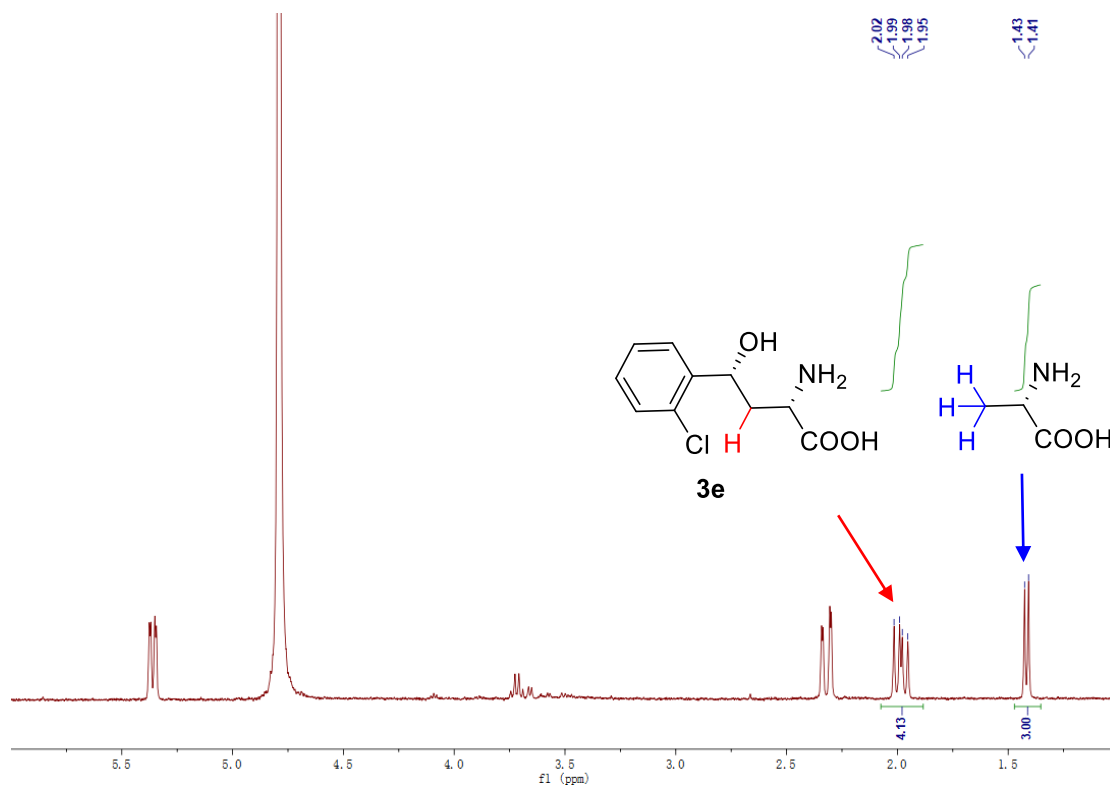


Figure S39. Conversion of **3e**

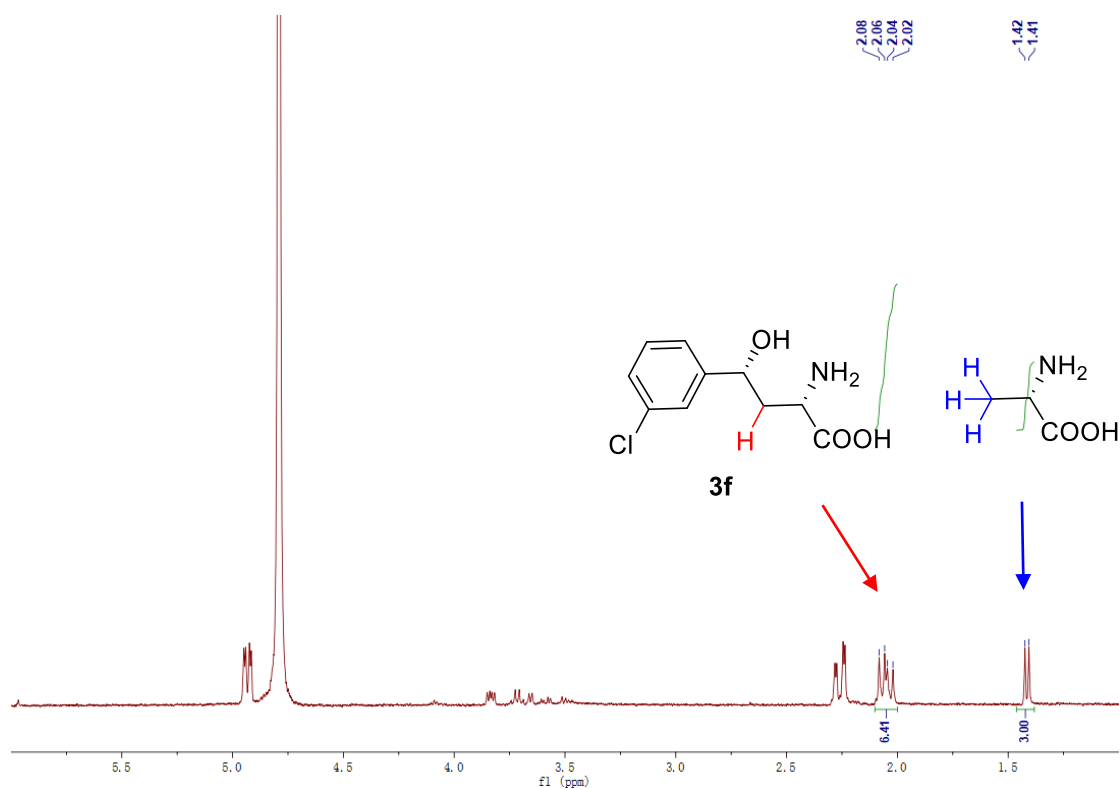


Figure S40. Conversion of **3f**

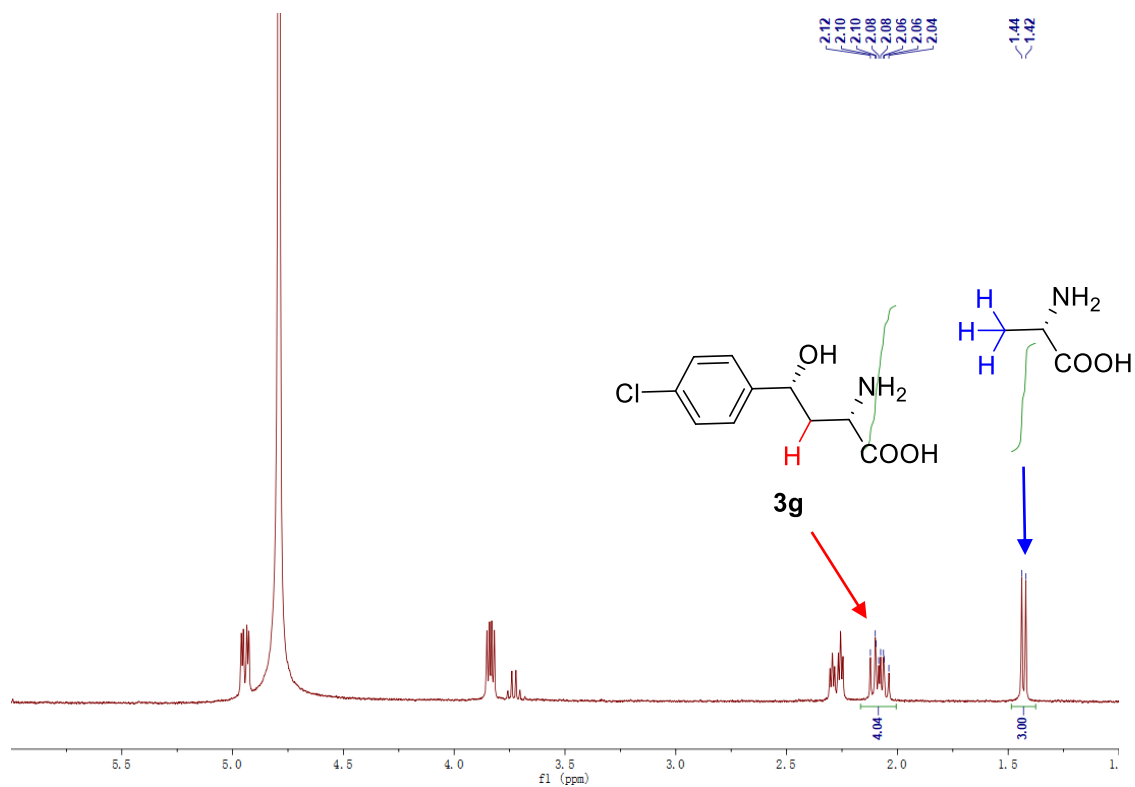


Figure S41. Conversion of **3g**

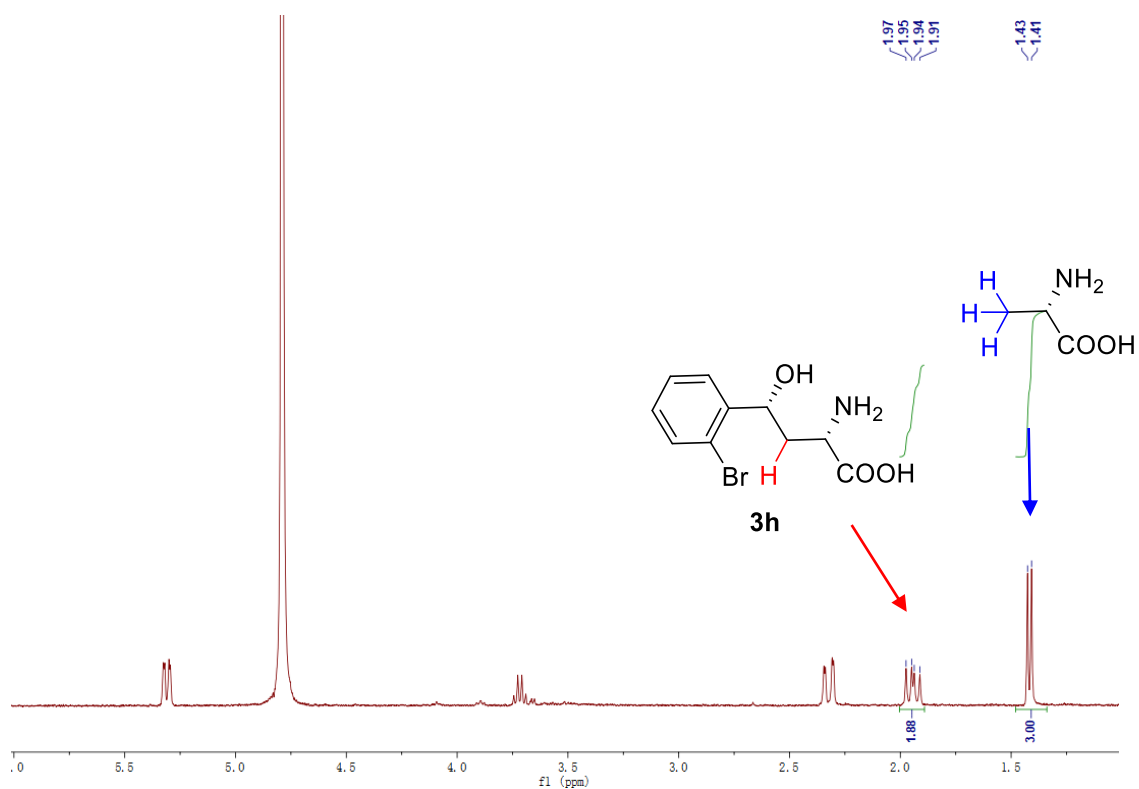


Figure S42. Conversion of **3h**

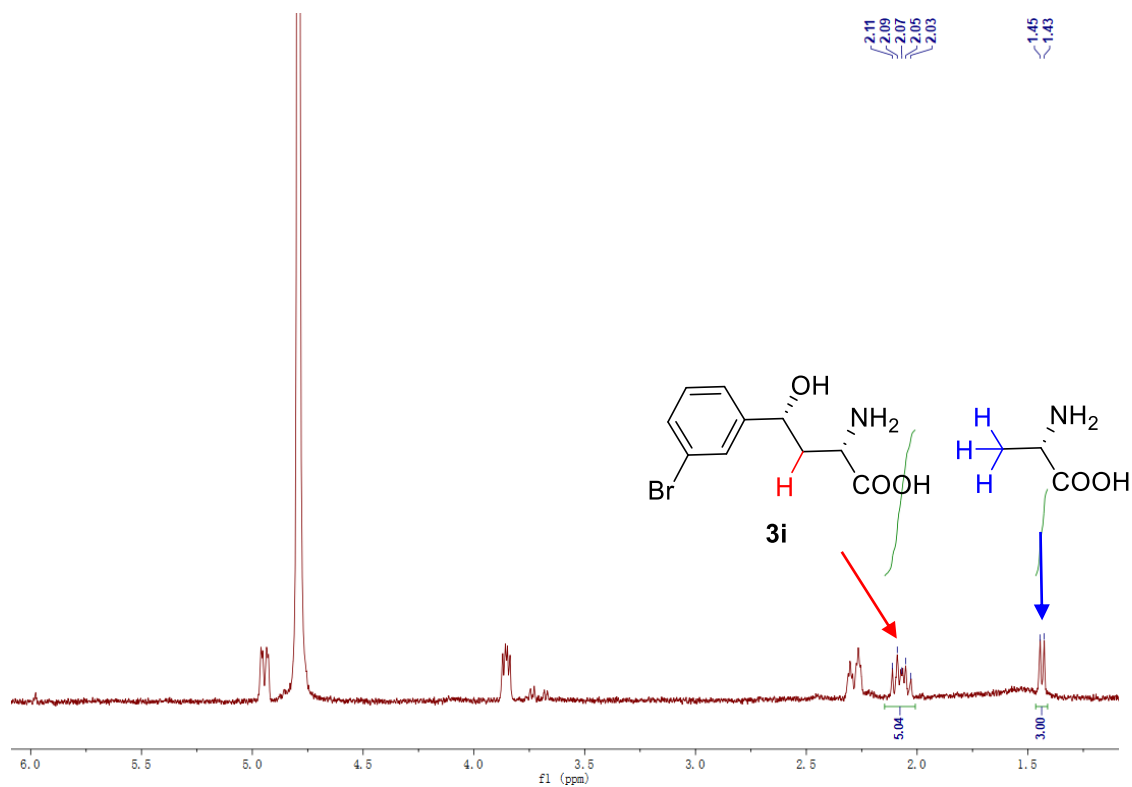


Figure S43. Conversion of **3i**

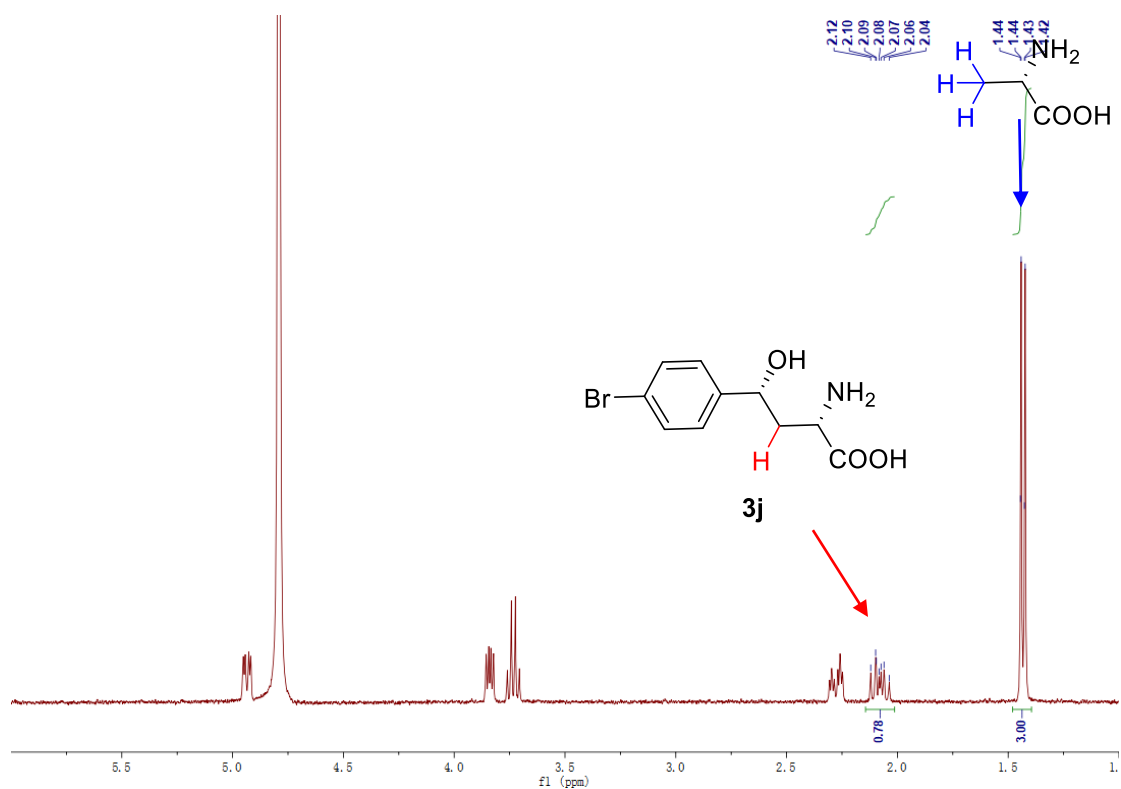


Figure S44. Conversion of **3j**

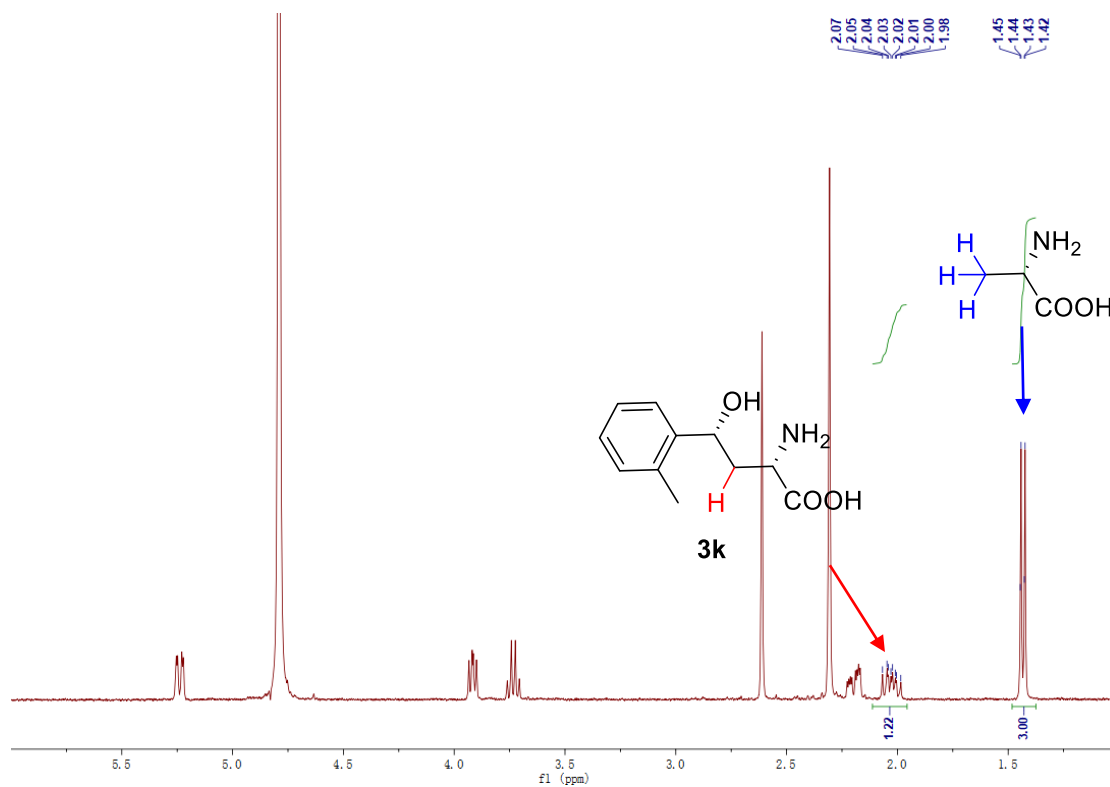


Figure S45. Conversion of **3k**

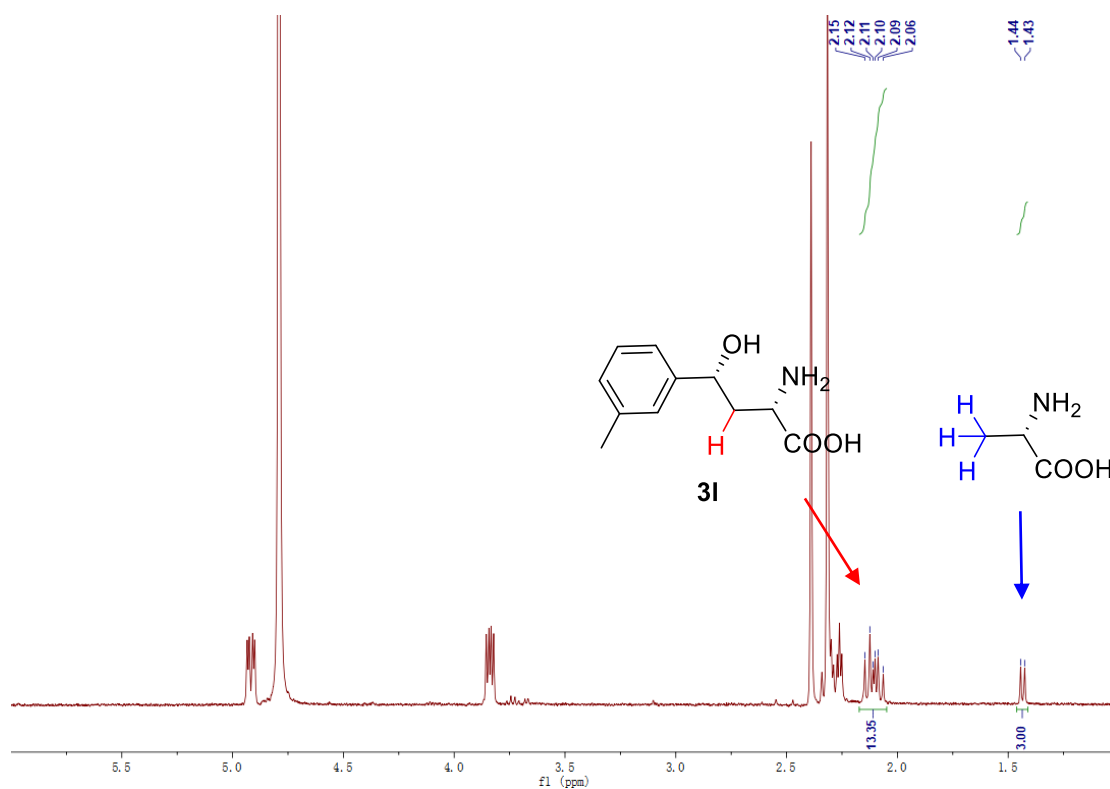


Figure S46. Conversion of **3l**

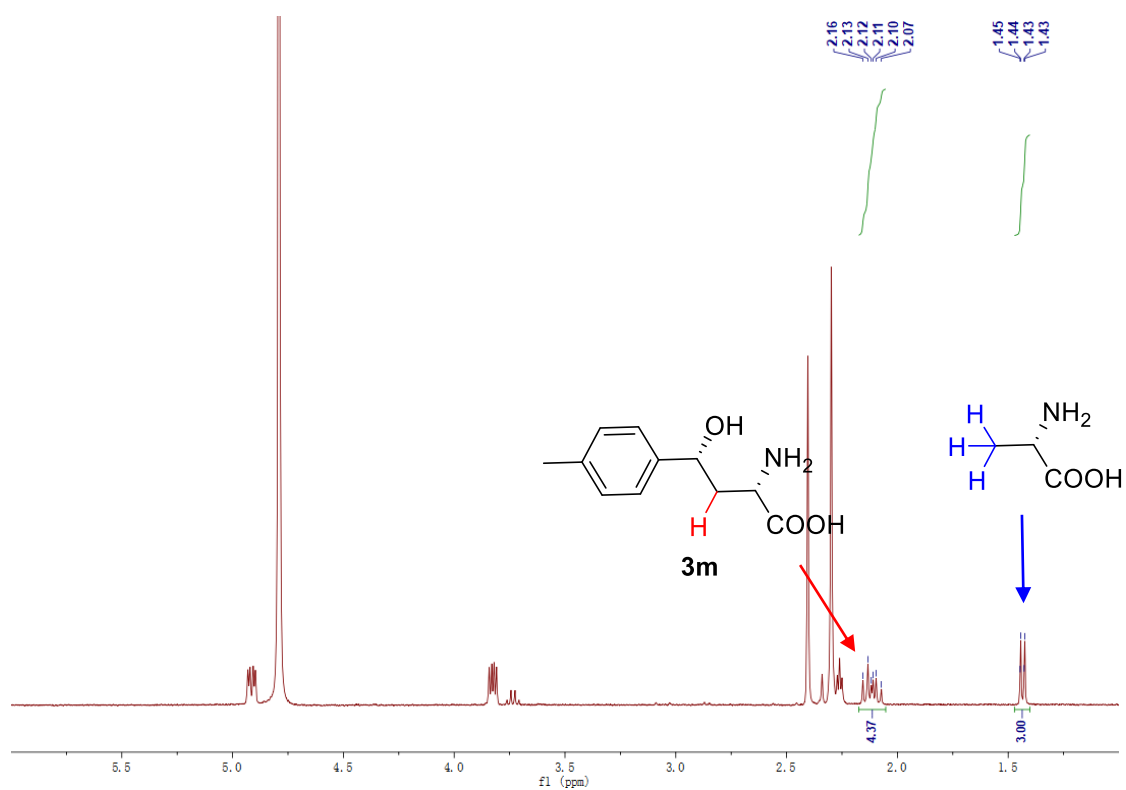


Figure S47. Conversion of **3m**

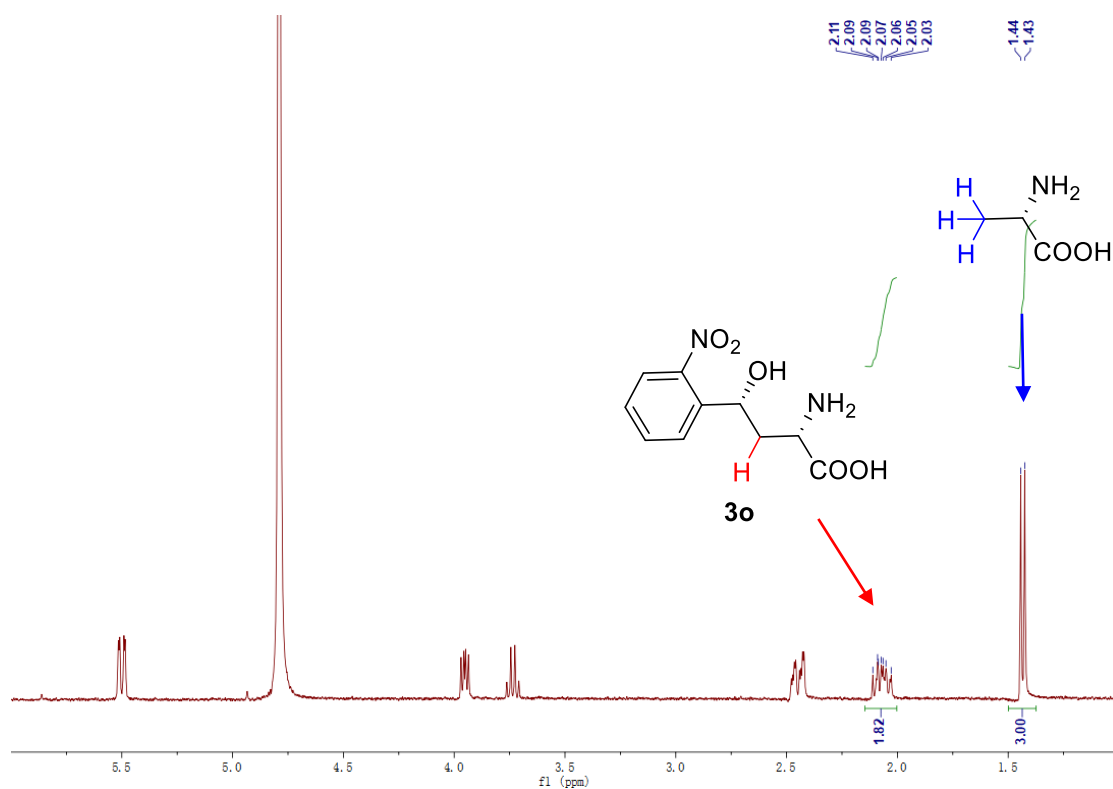


Figure S48. Conversion of **3o**

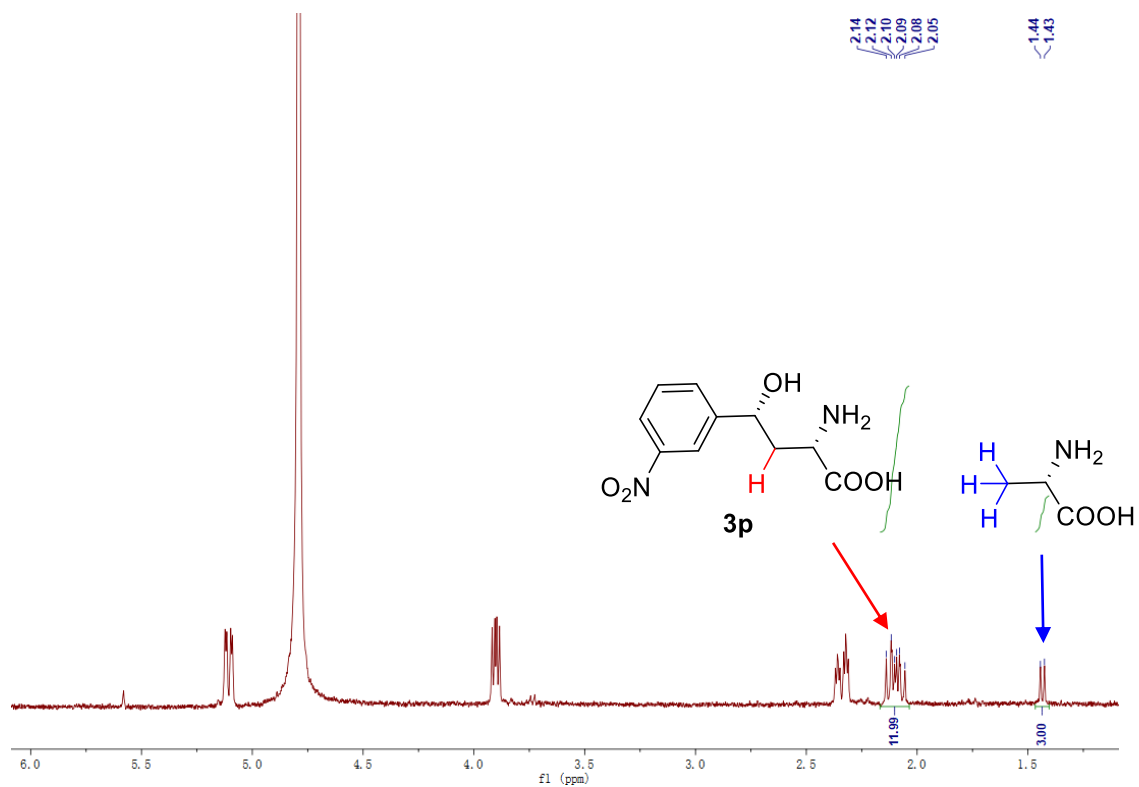


Figure S49. Conversion of **3p**

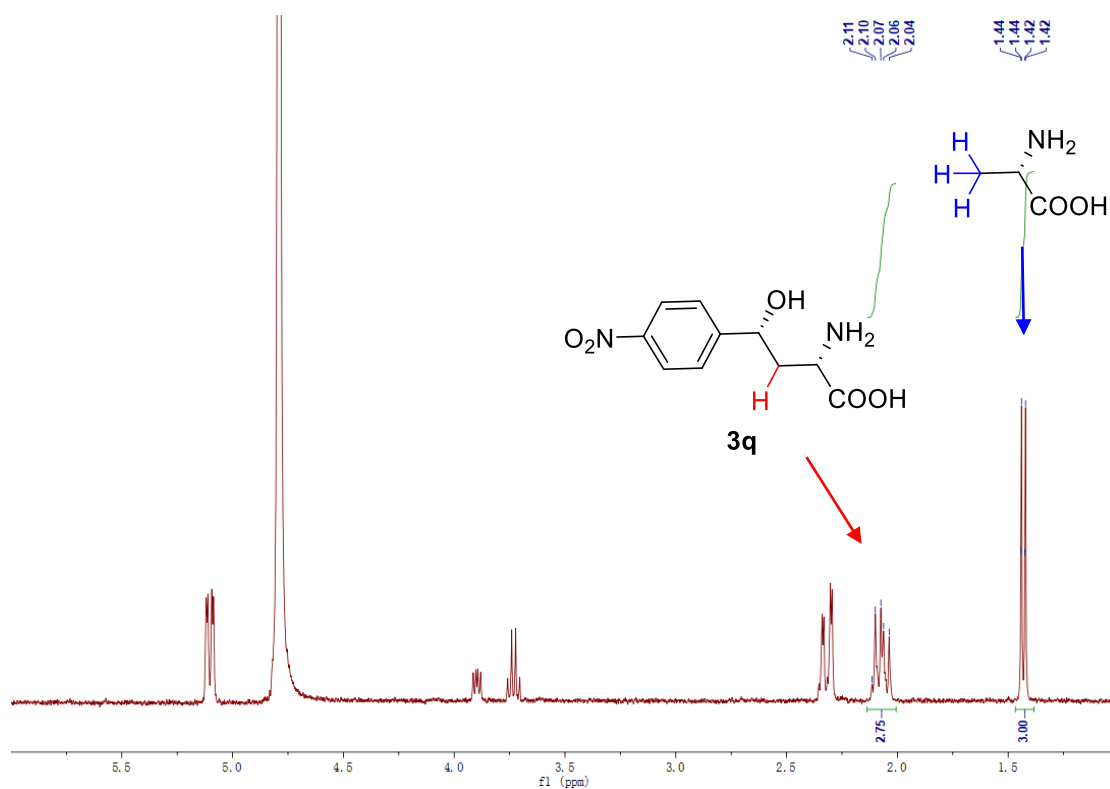


Figure S50. Conversion of **3q**

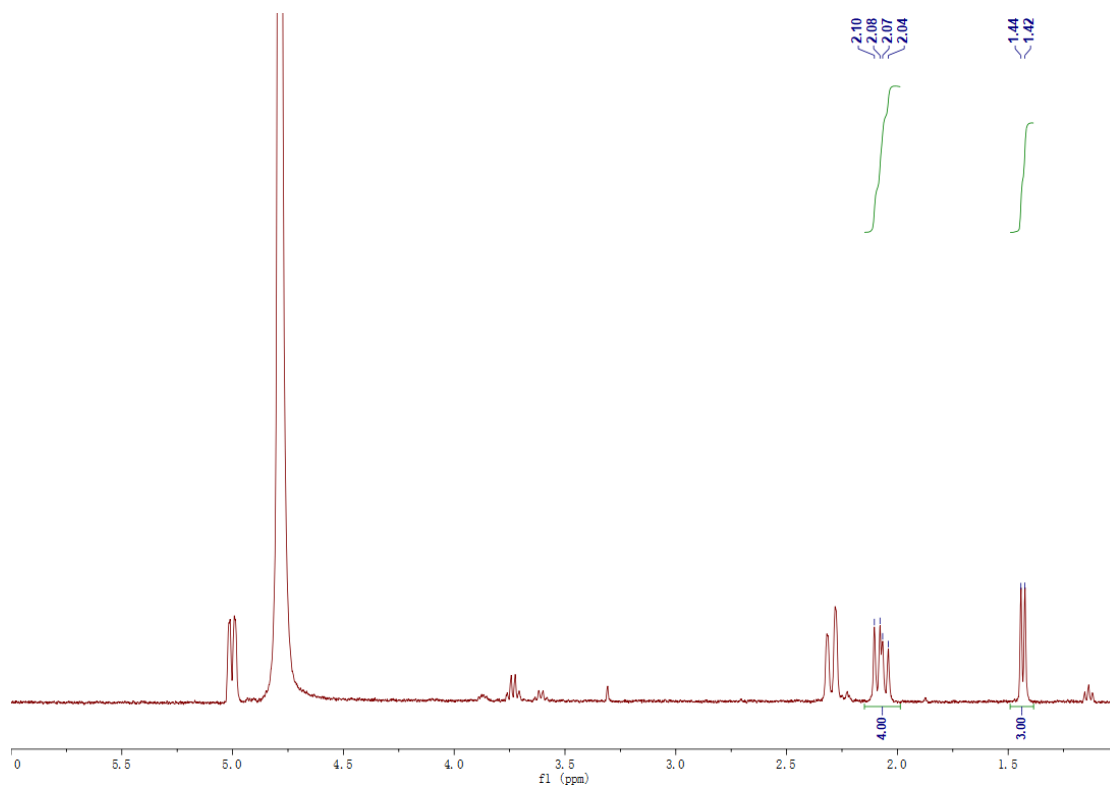


Figure S51. Conversion of **3r**

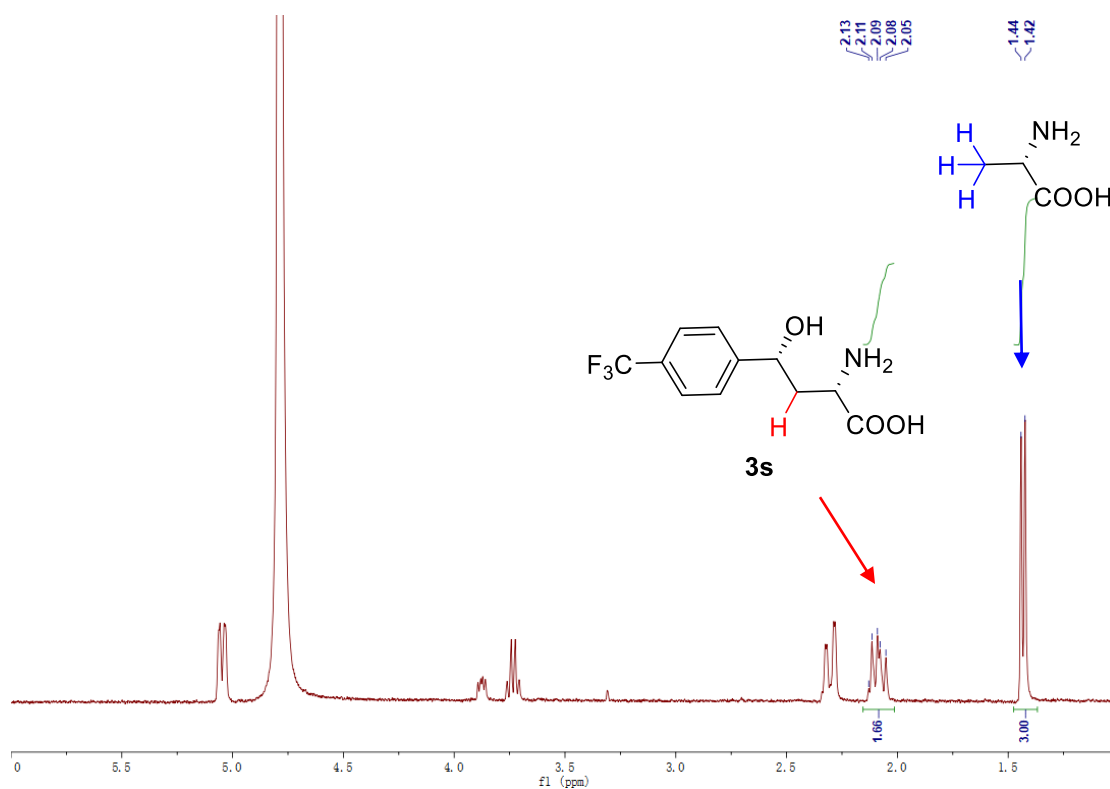


Figure S52. Conversion of **3s**

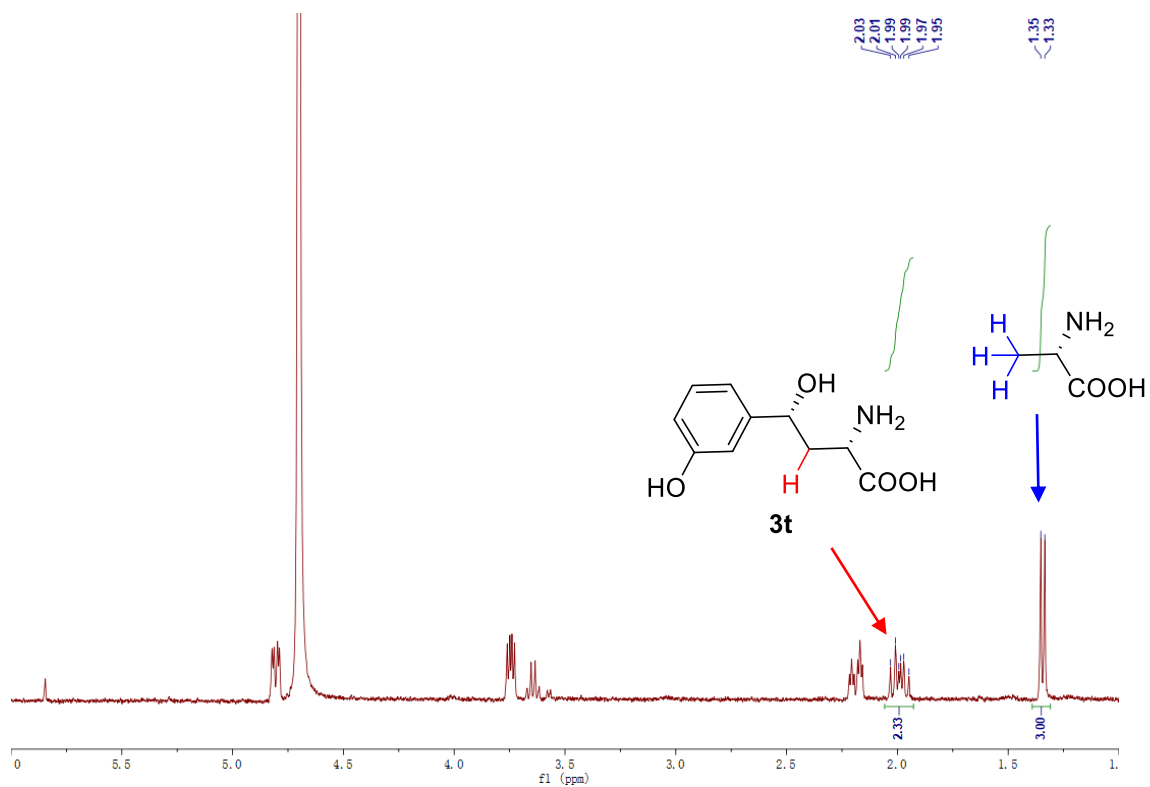


Figure S53. Conversion of **3t**

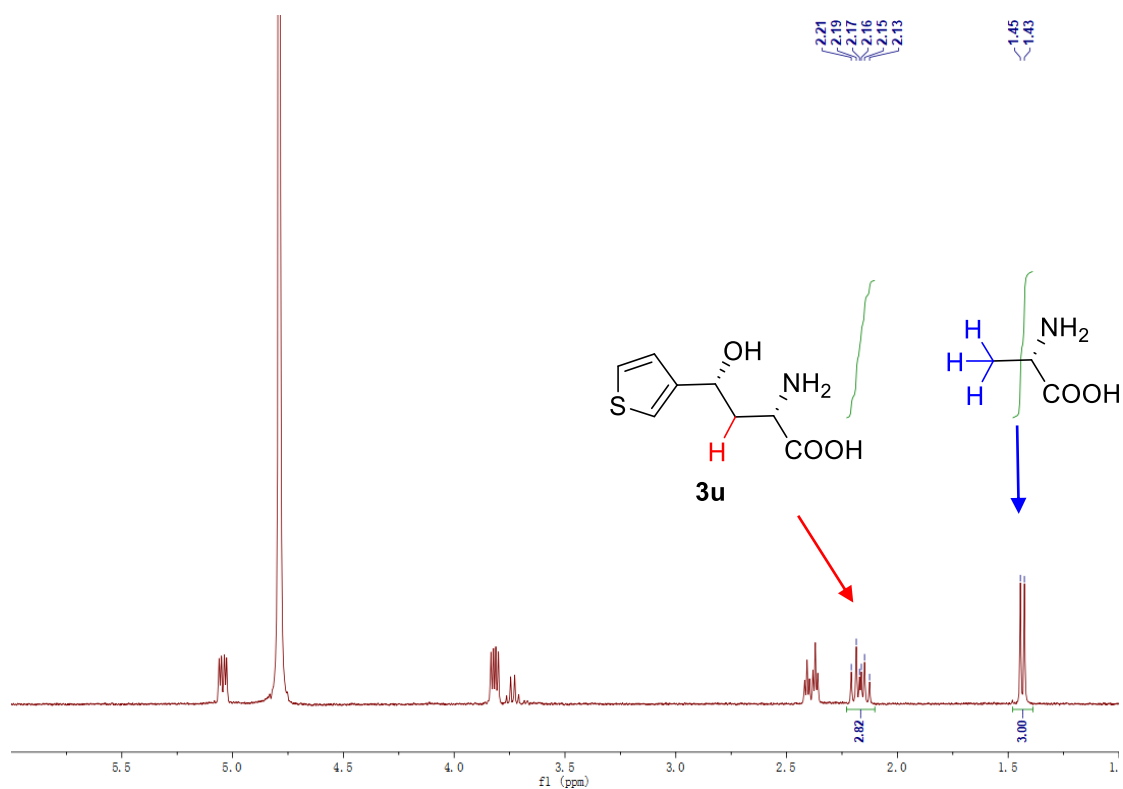


Figure S54. Conversion of **3u**

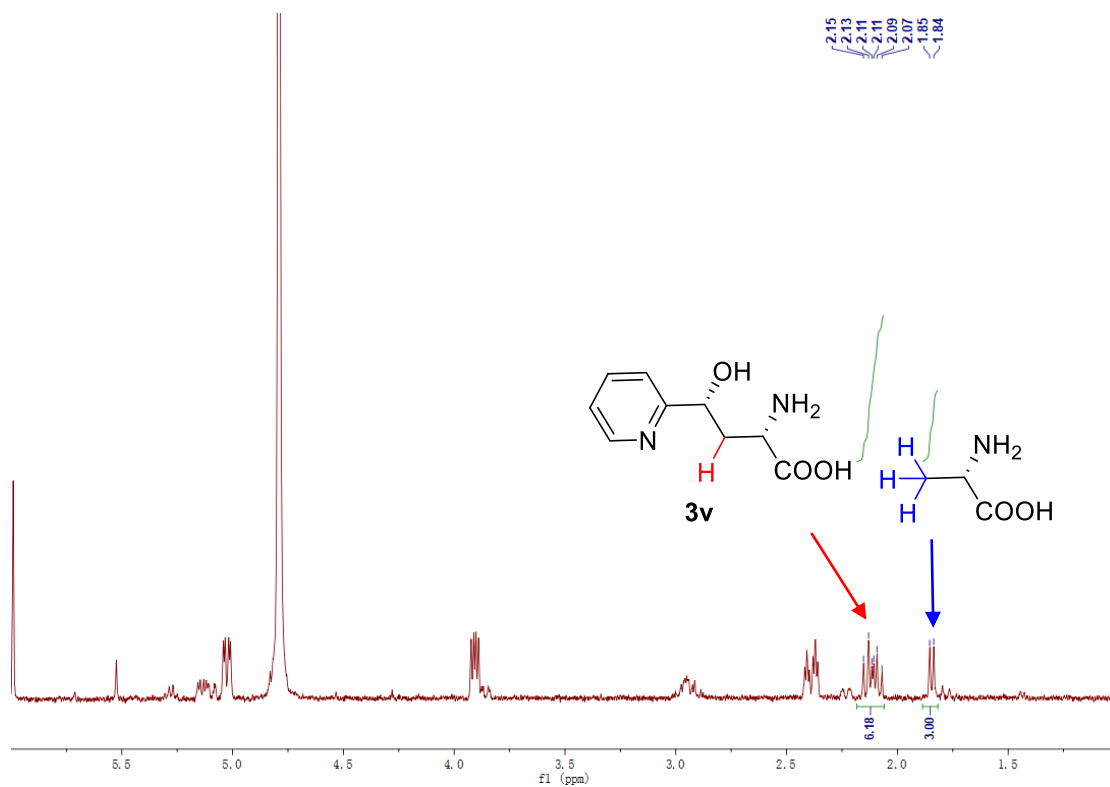


Figure S55. Conversion of **3v**

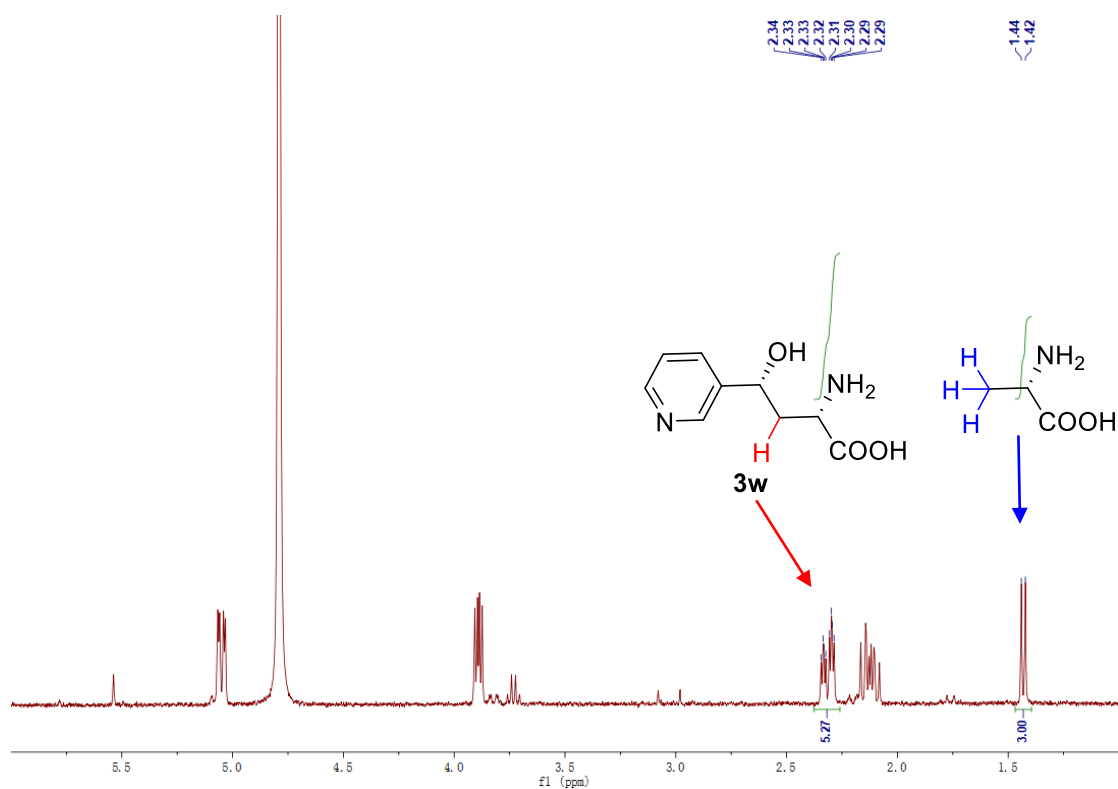


Figure S56. Conversion of **3w**

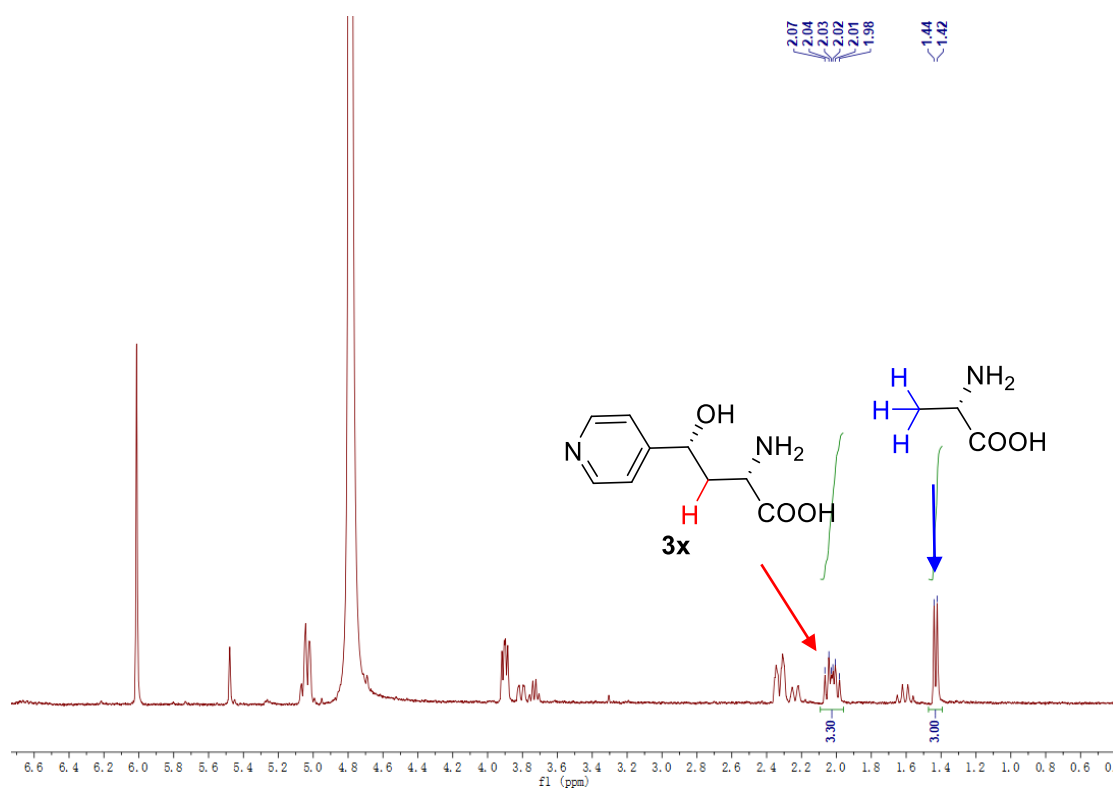


Figure S57. Conversion of **3x**

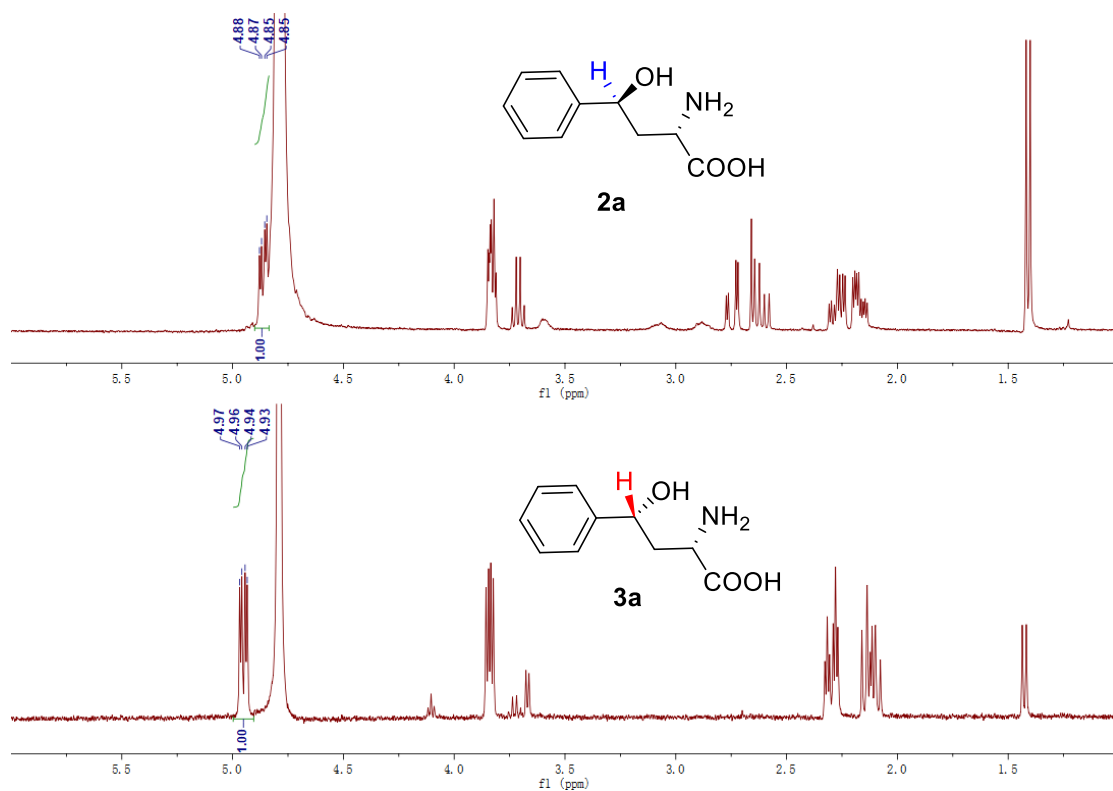


Figure S58. Diastereoisomeric ratio of **2a** and **3a**

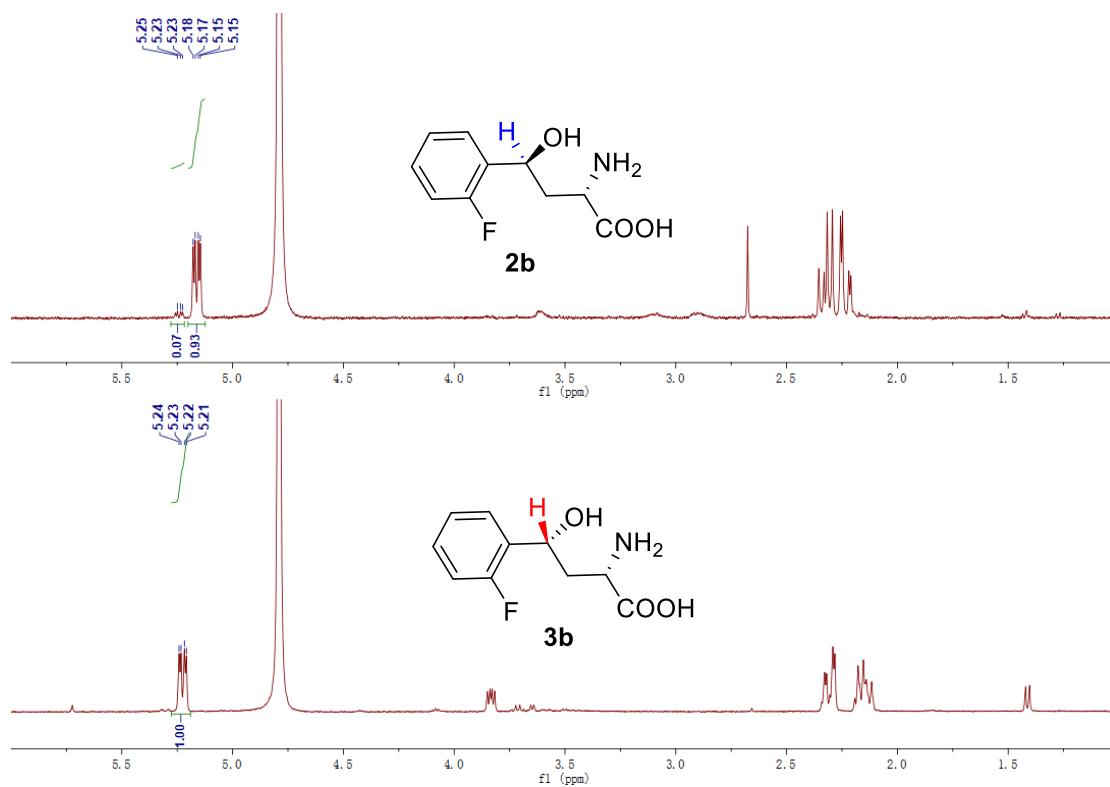


Figure S59. Diastereoisomeric ratio of **2b** and **3b**

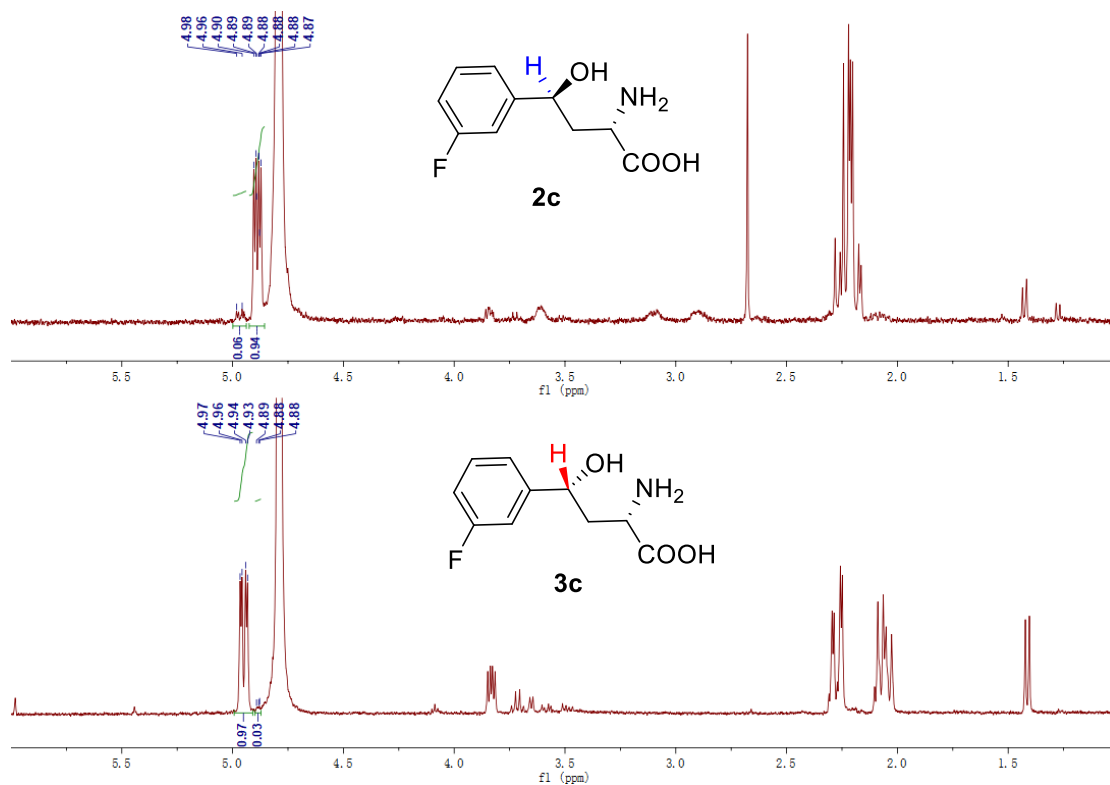


Figure S60. Diastereoisomeric ratio of **2c** and **3c**

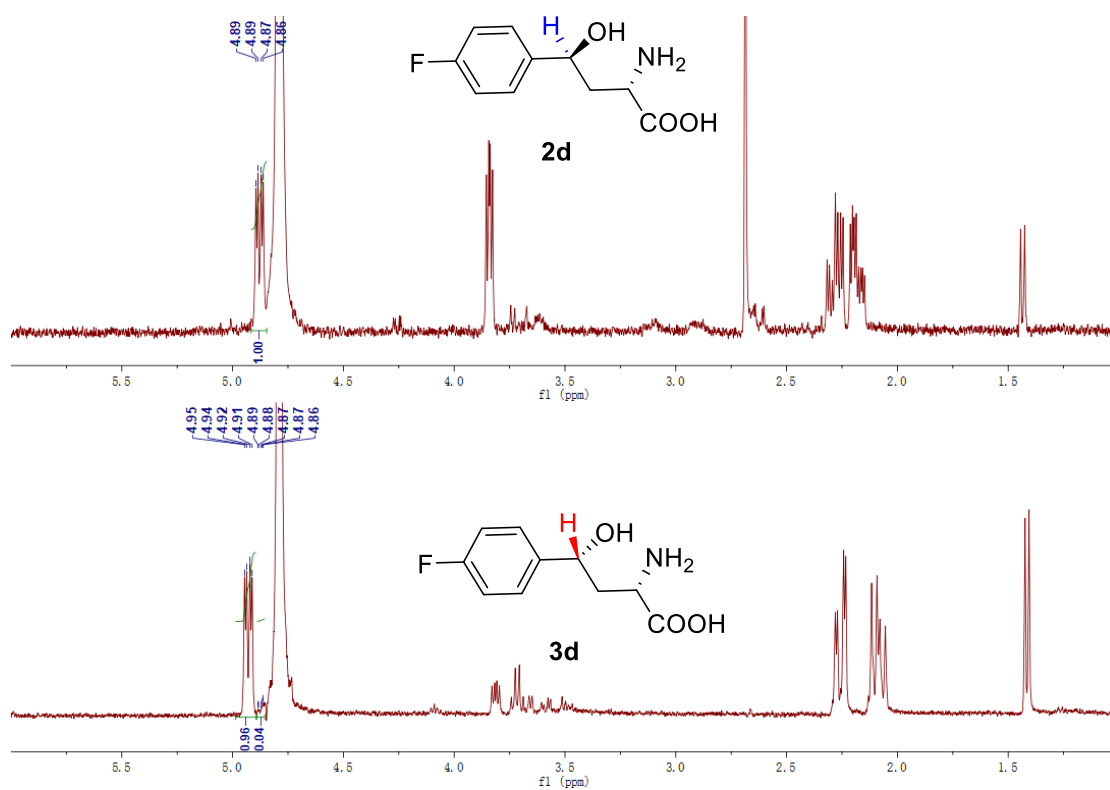


Figure S61. Diastereoisomeric ratio of **2d** and **3d**

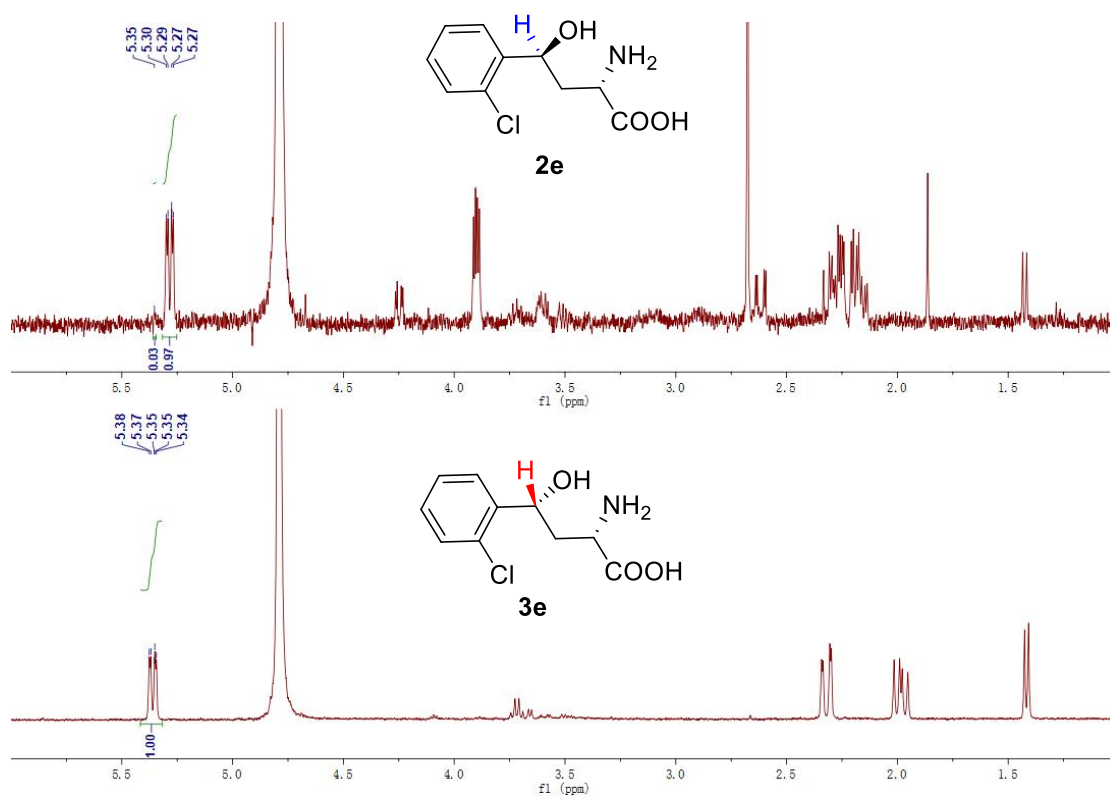


Figure S62. Diastereoisomeric ratio of **2e** and **3e**

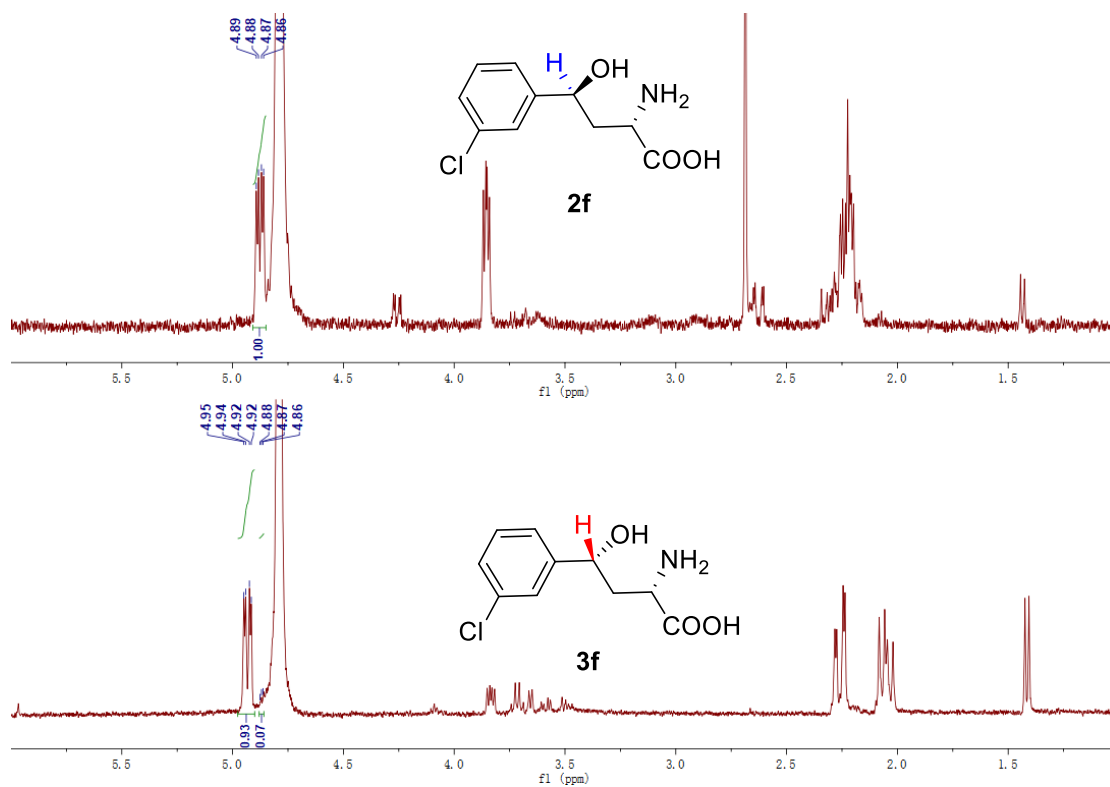


Figure S63. Diastereoisomeric ratio of **2f** and **3f**

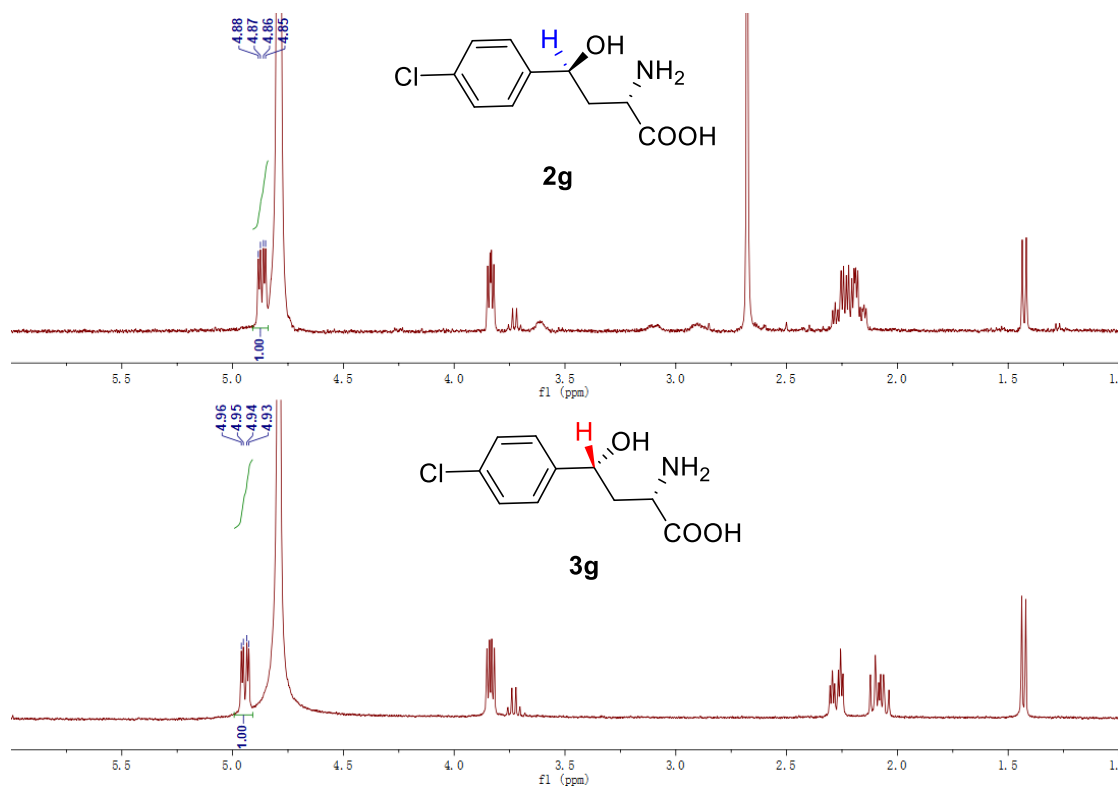


Figure S64. Diastereoisomeric ratio of **2g** and **3g**

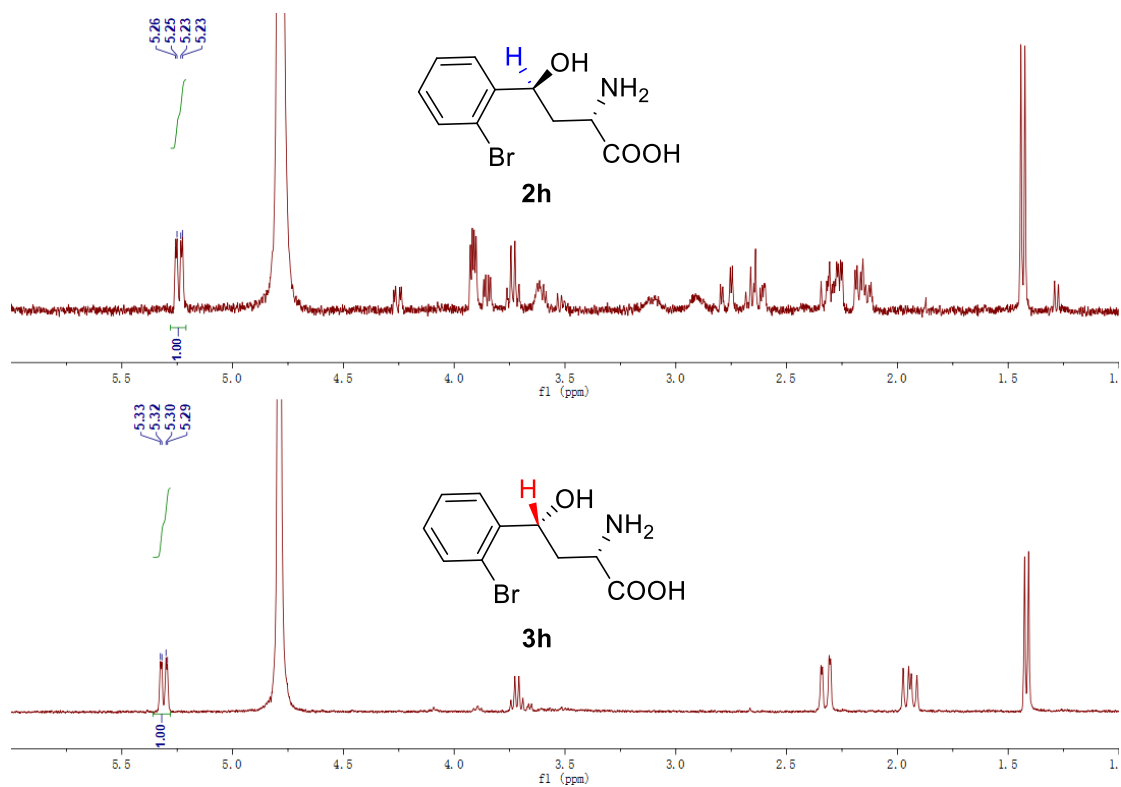


Figure S65. Diastereoisomeric ratio of **2h** and **3h**

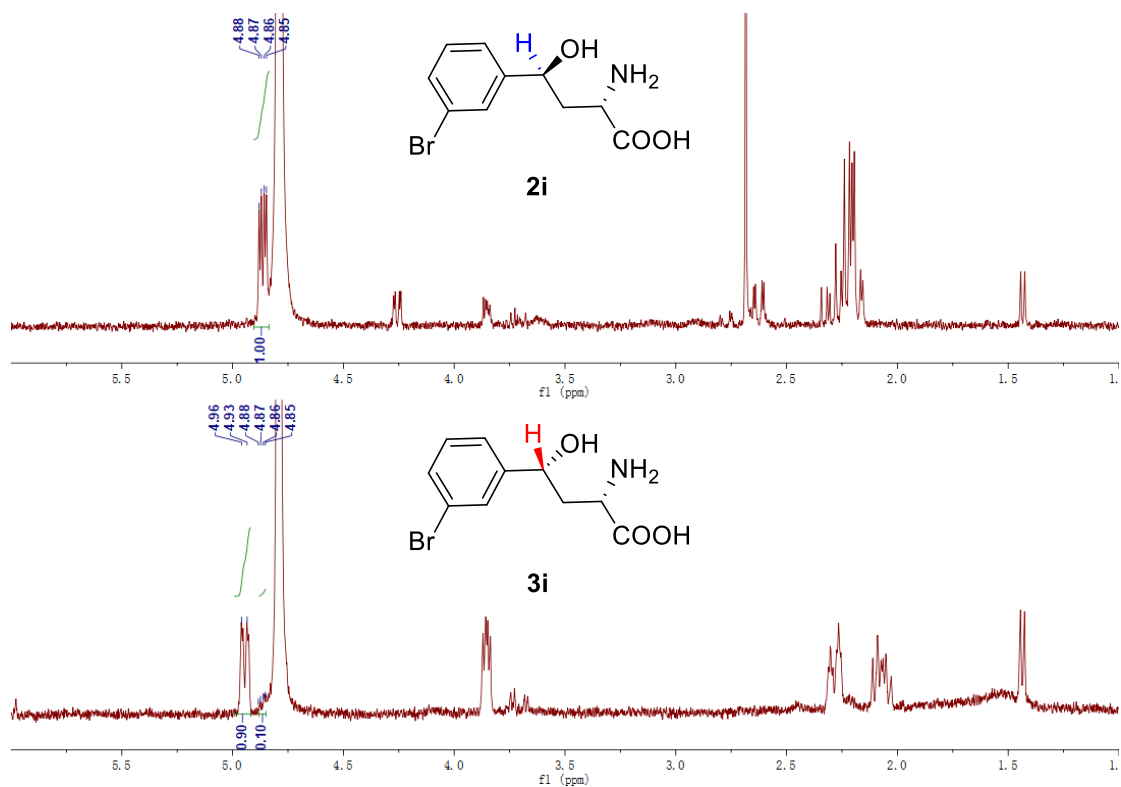


Figure S66. Diastereoisomeric ratio of **2i** and **3i**

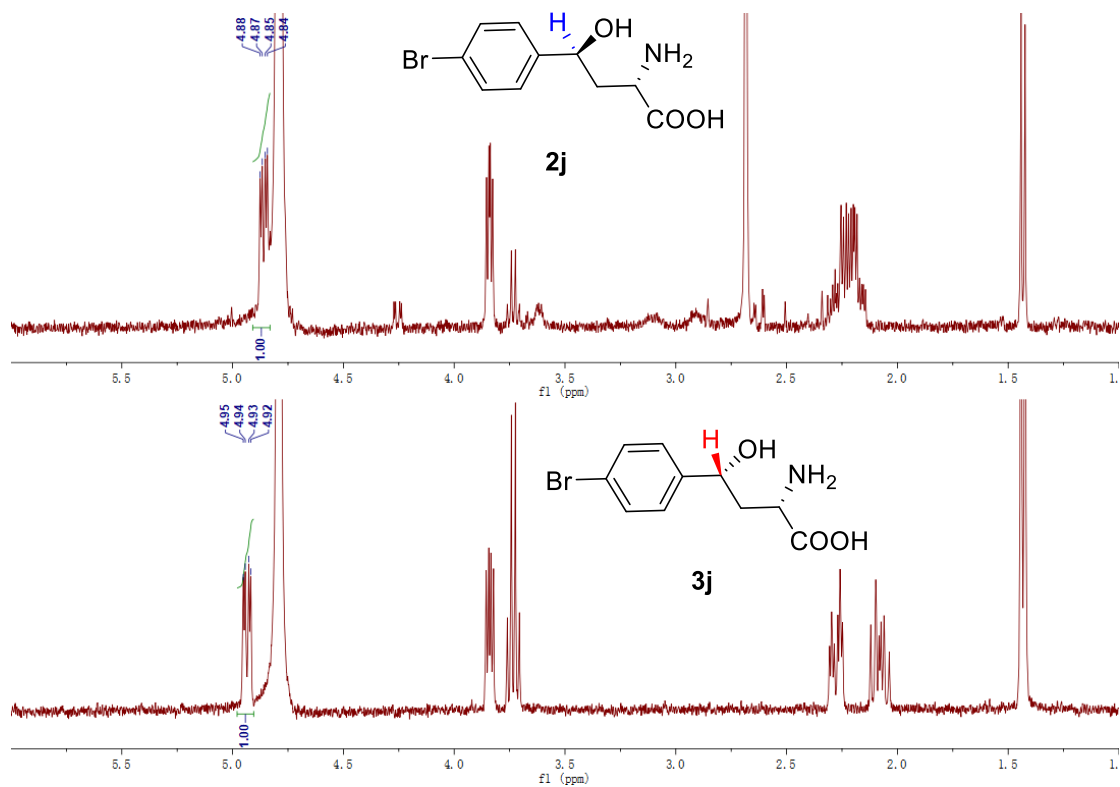


Figure S67. Diastereoisomeric ratio of **2j** and **3j**

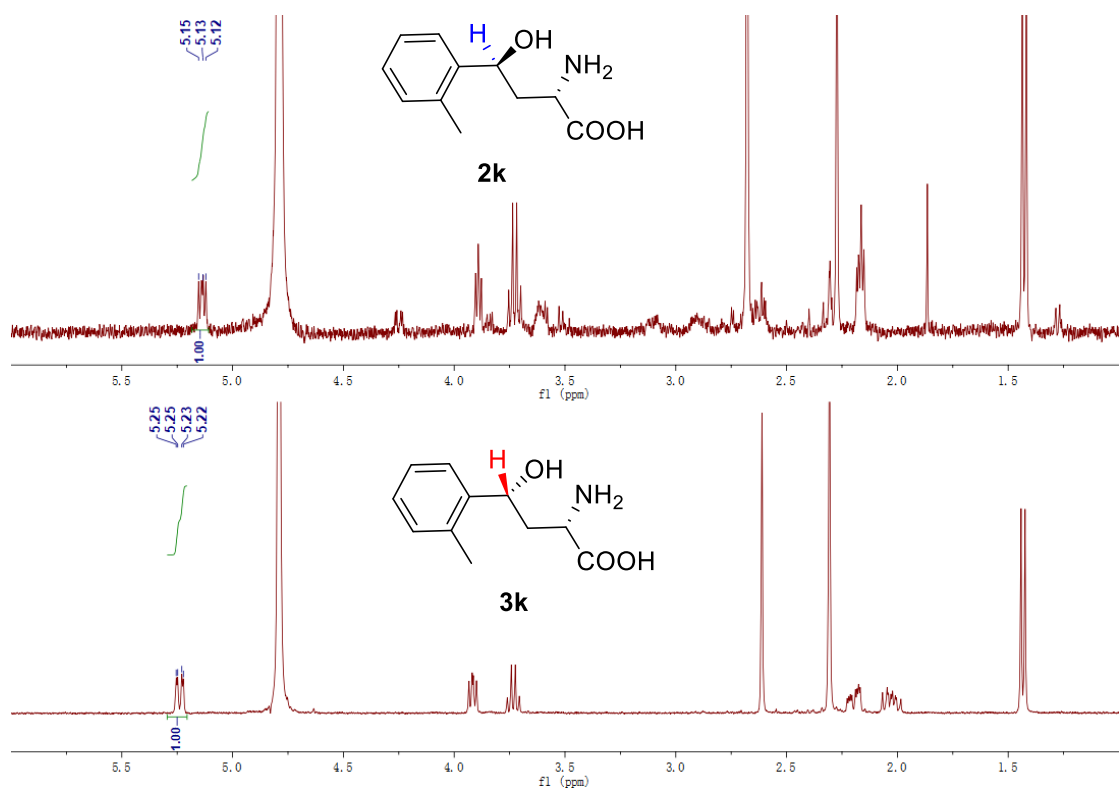


Figure S68. Diastereoisomeric ratio of **2k** and **3k**

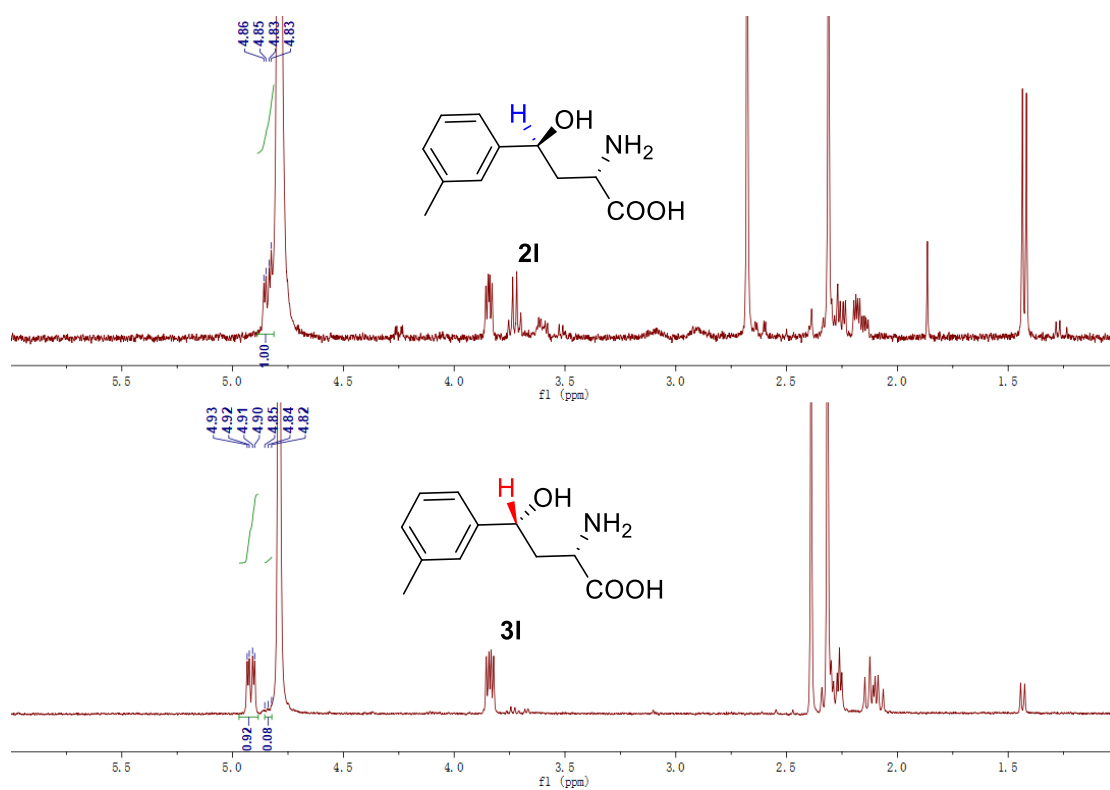


Figure S69. Diastereoisomeric ratio of **2l** and **3l**

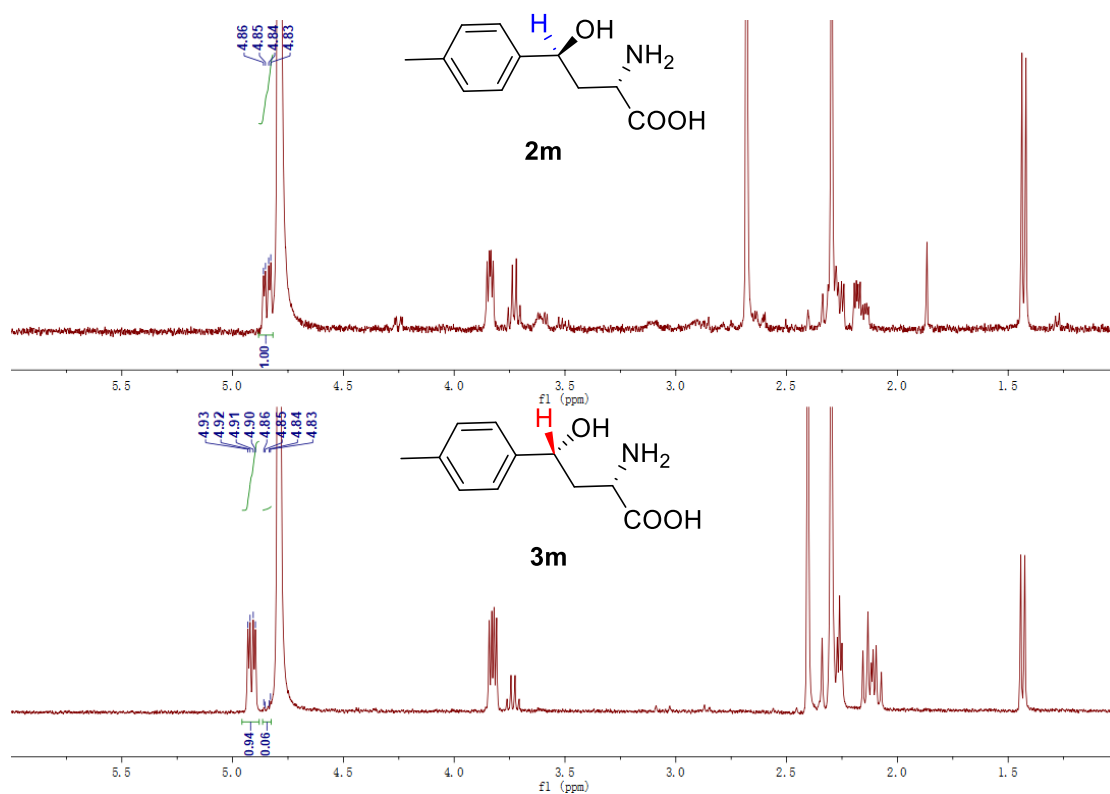


Figure S70. Diastereoisomeric ratio of **2m** and **3m**

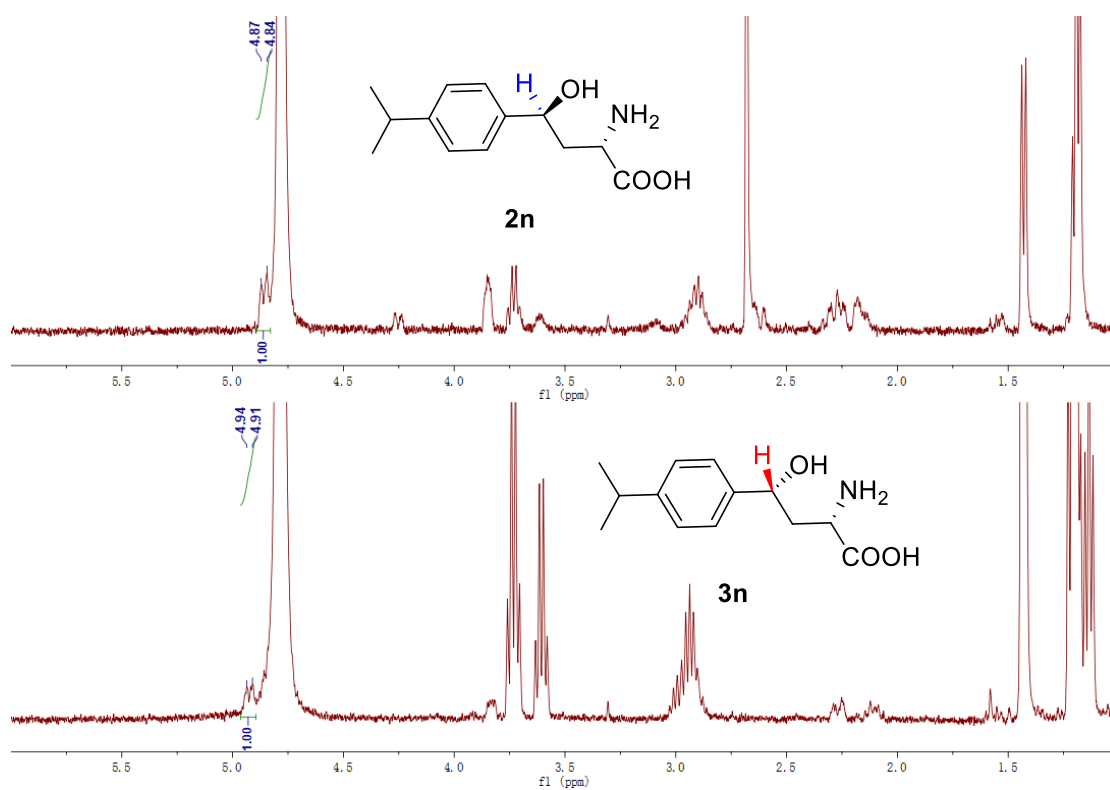


Figure S71. Diastereoisomeric ratio of **2n** and **3n**

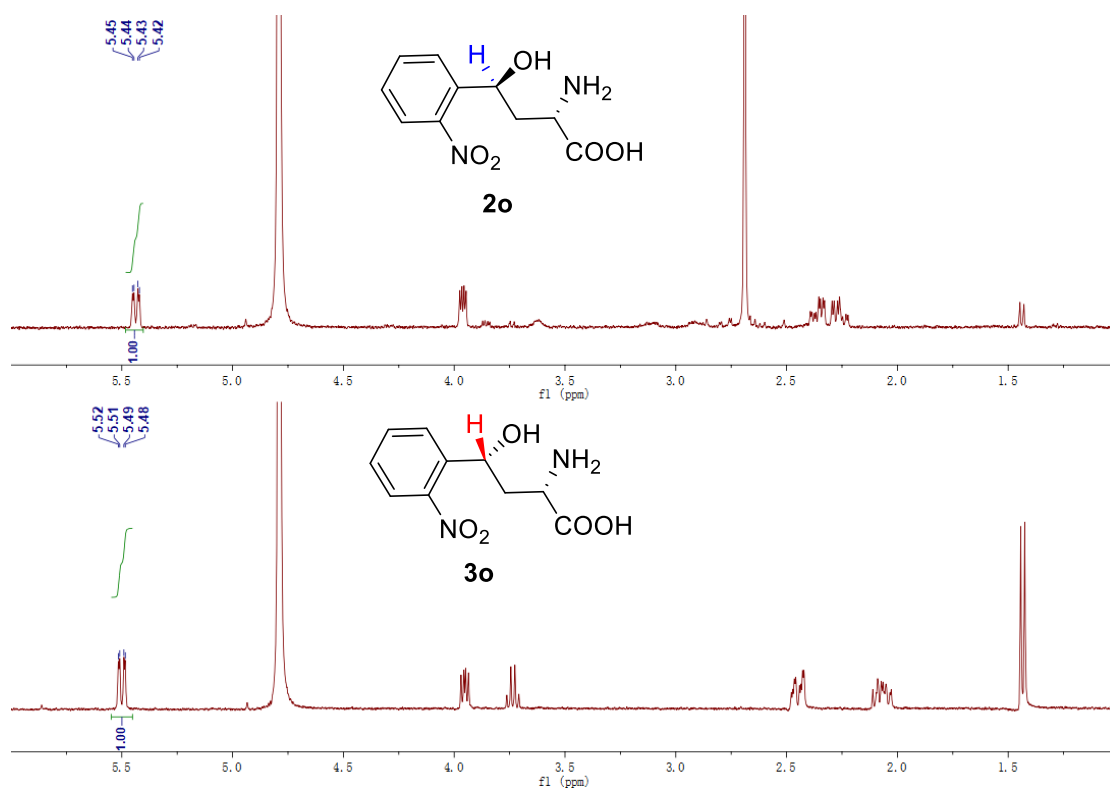


Figure S72. Diastereoisomeric ratio of **2o** and **3o**

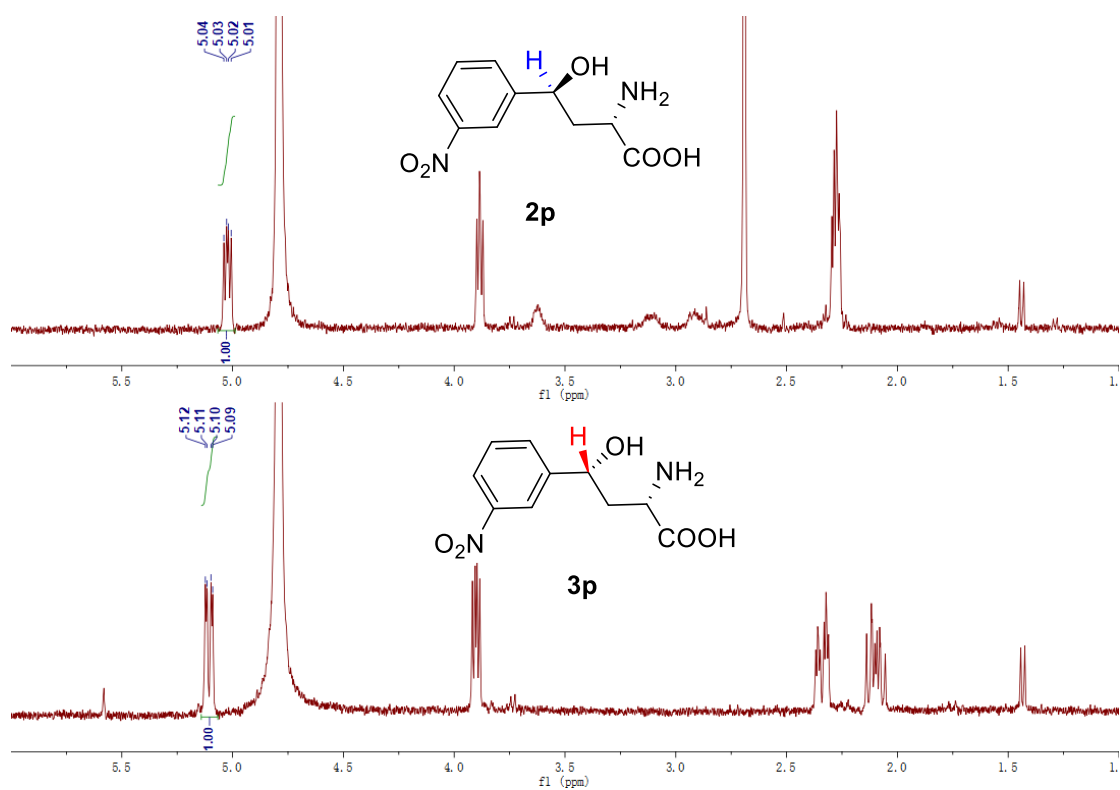


Figure S73. Diastereoisomeric ratio of **2p** and **3p**

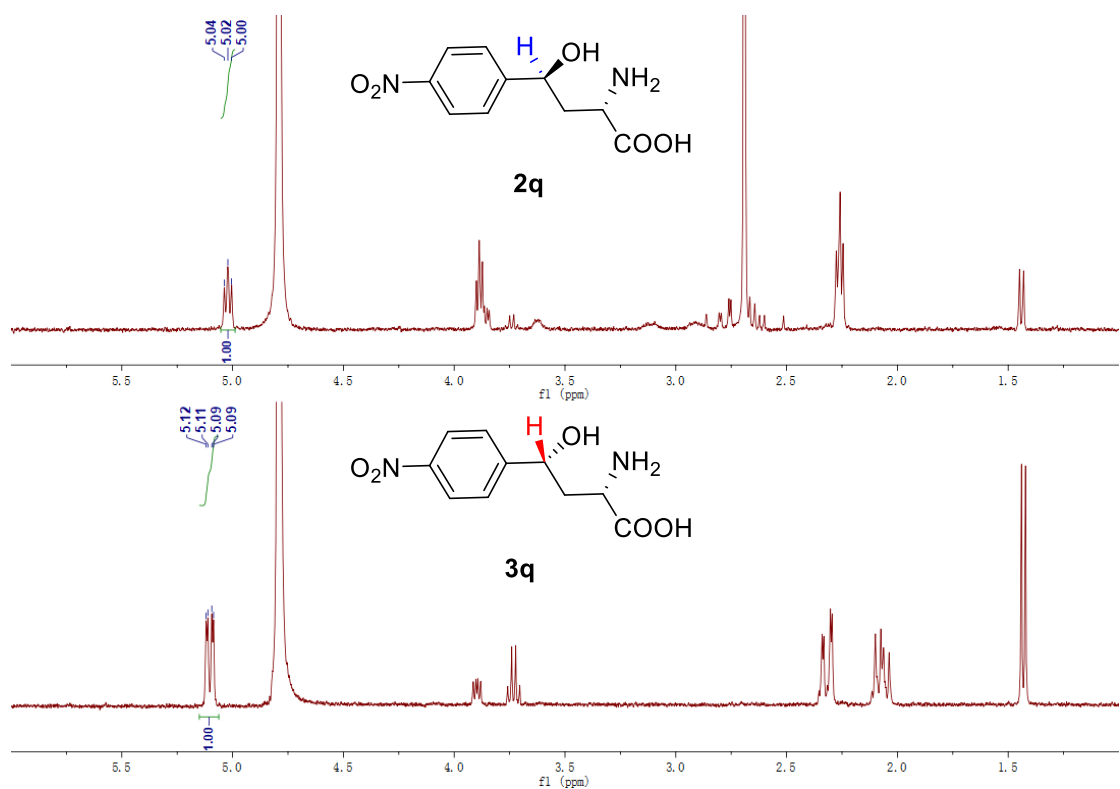


Figure S74. Diastereoisomeric ratio of **2q** and **3q**

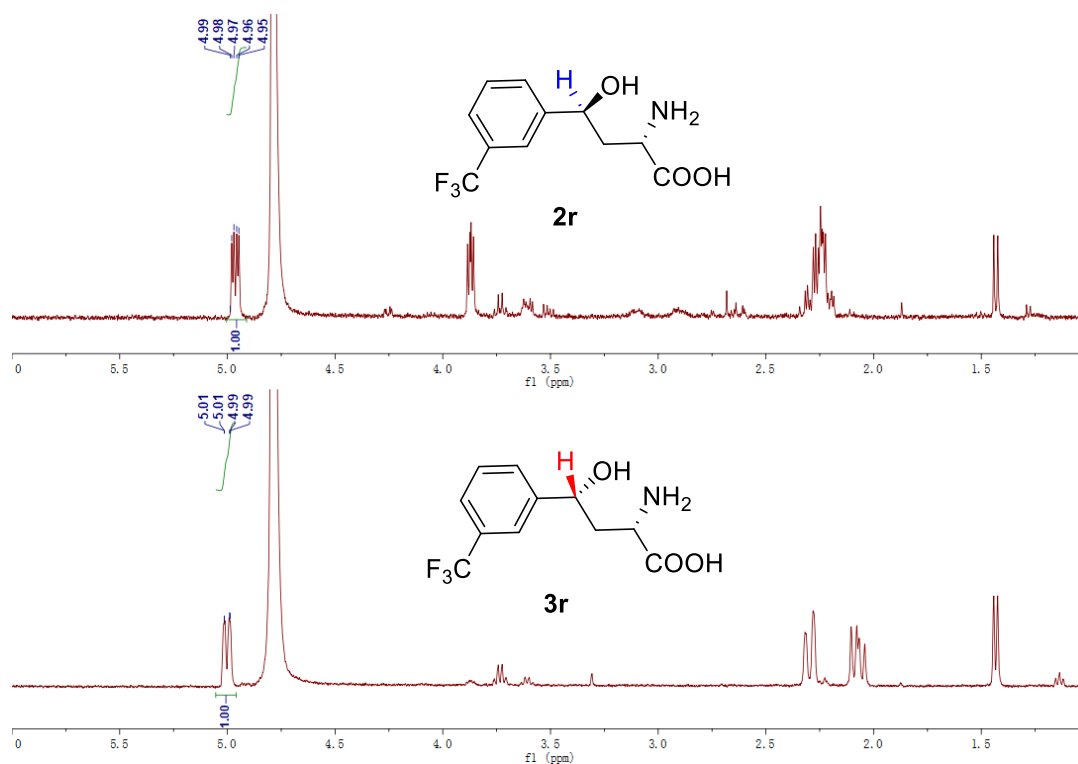


Figure S75. Diastereoisomeric ratio of **2r** and **3r**

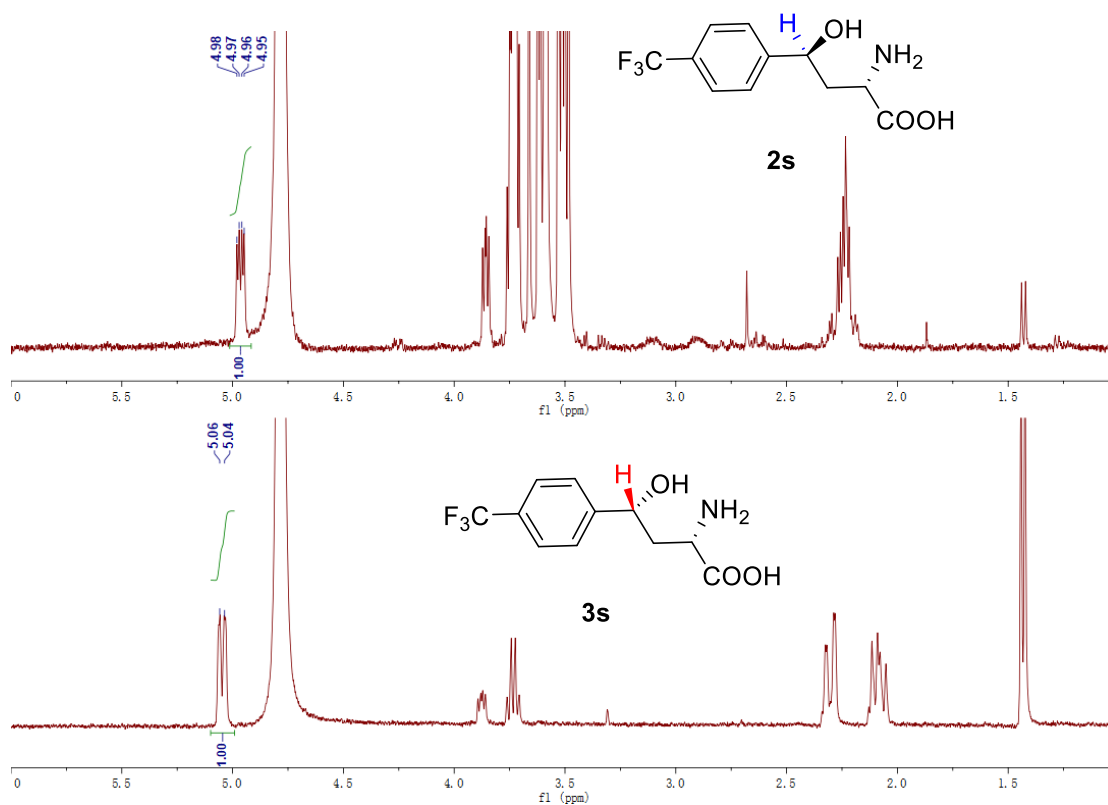


Figure S76. Diastereoisomeric ratio of **2s** and **3s**

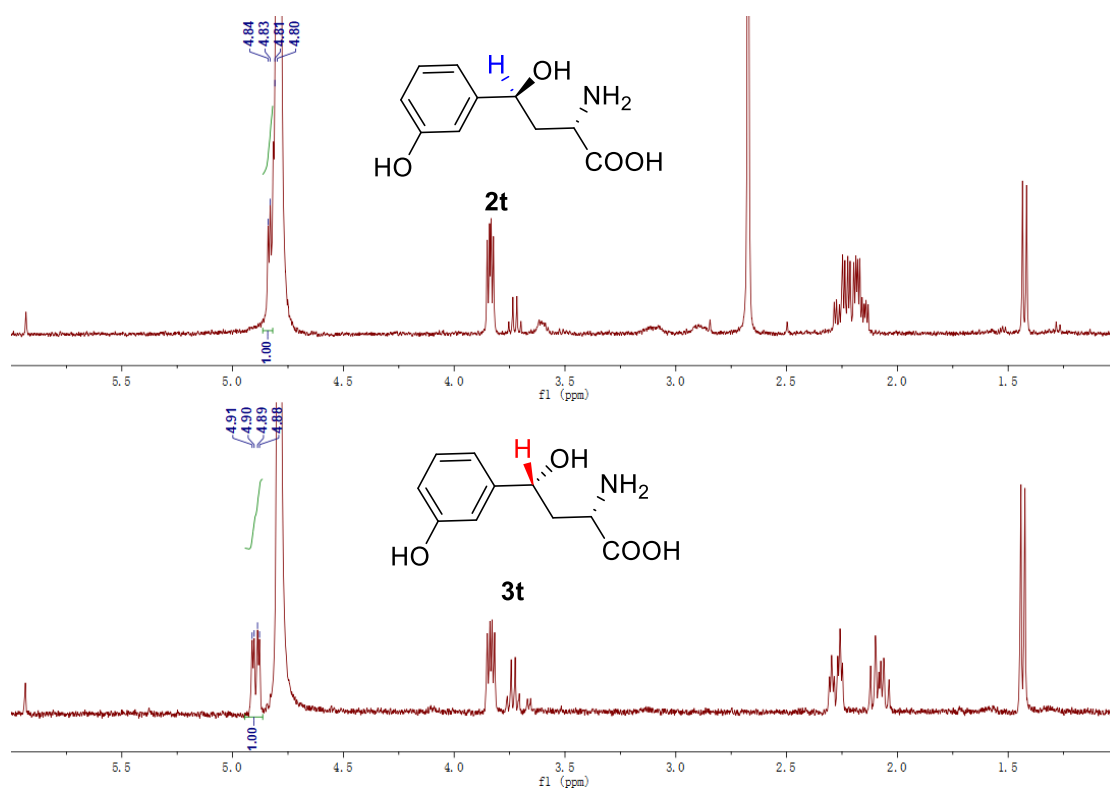


Figure S77. Diastereoisomeric ratio of **2t** and **3t**

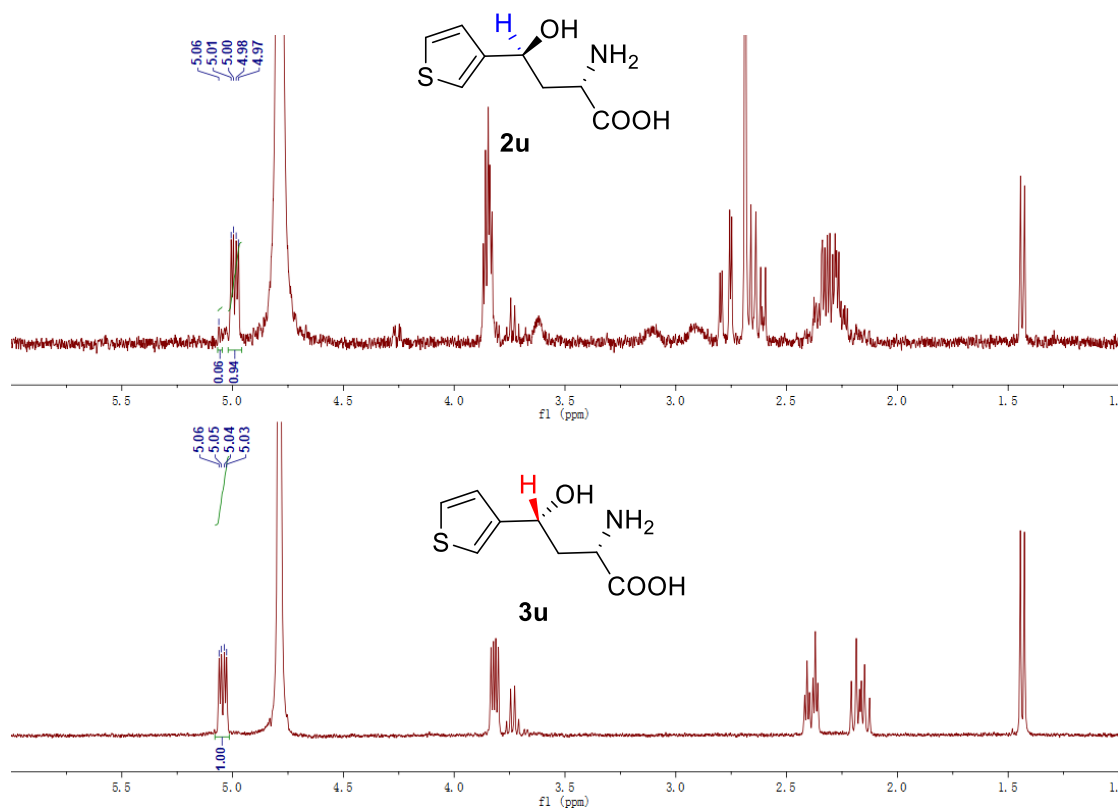


Figure S78. Diastereoisomeric ratio of **2u** and **3u**

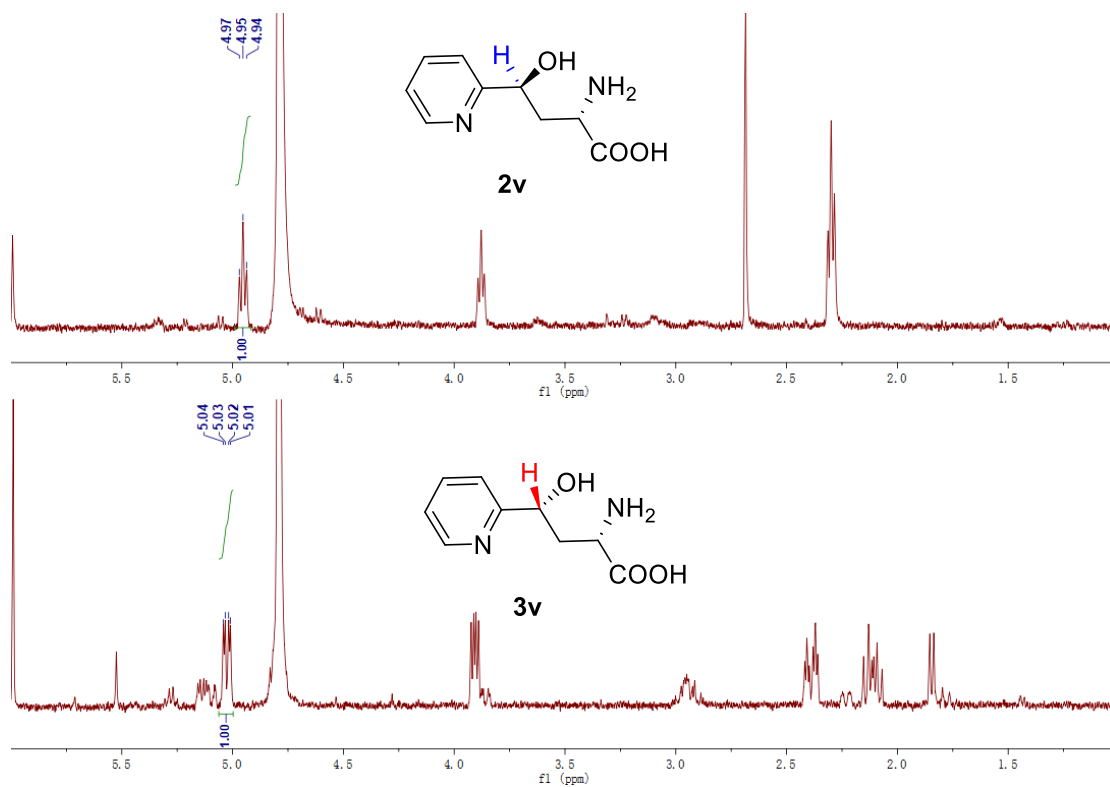


Figure S79. Diastereoisomeric ratio of **2v** and **3v**

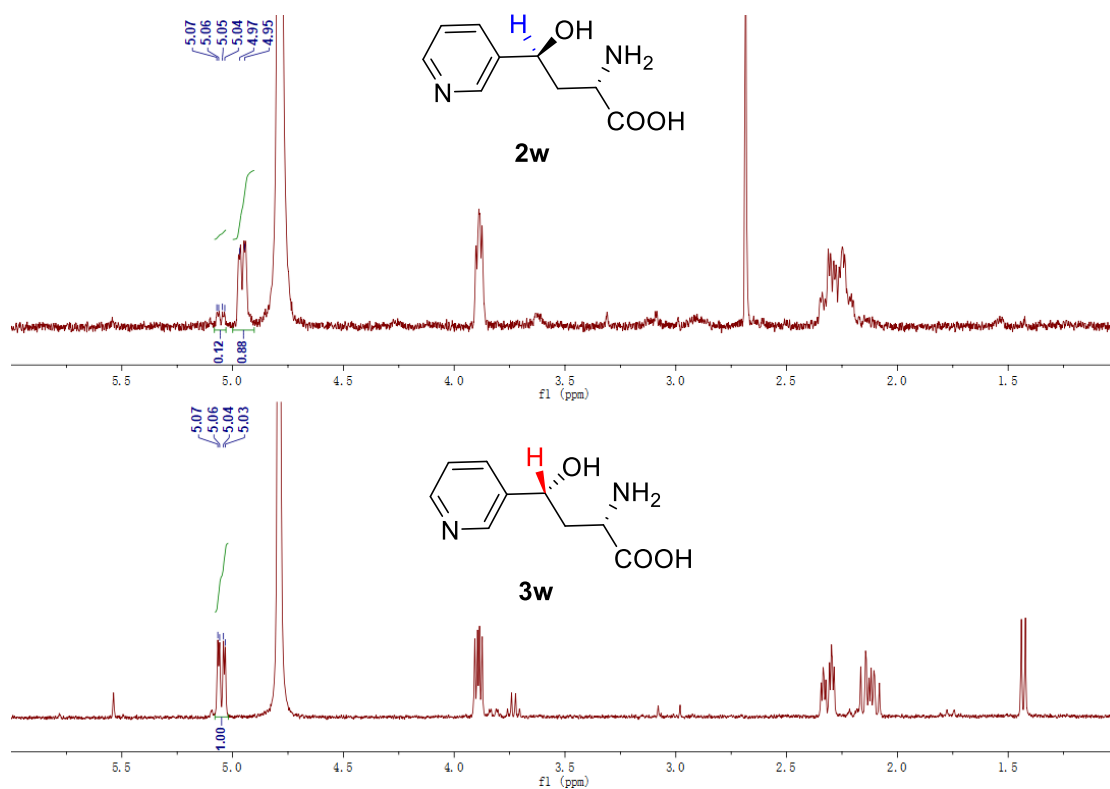


Figure S80. Diastereoisomeric ratio of **2w** and **3w**

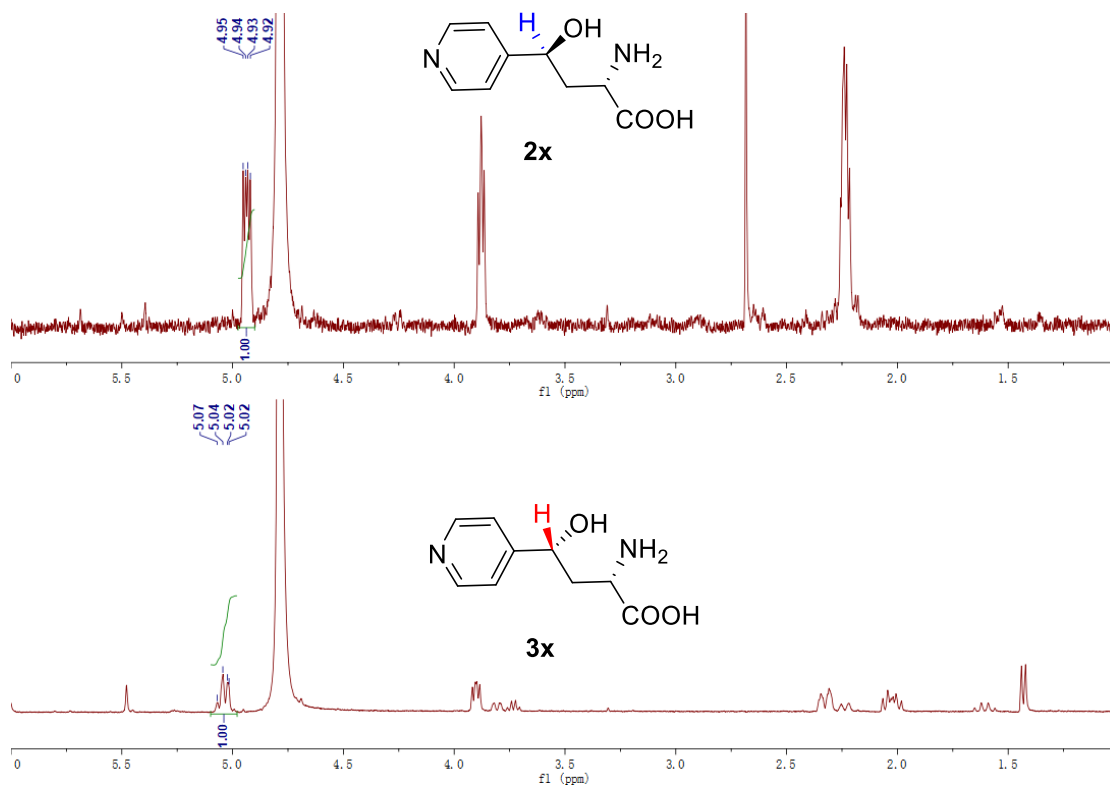


Figure S81. Diastereoisomeric ratio of **2x** and **3x**

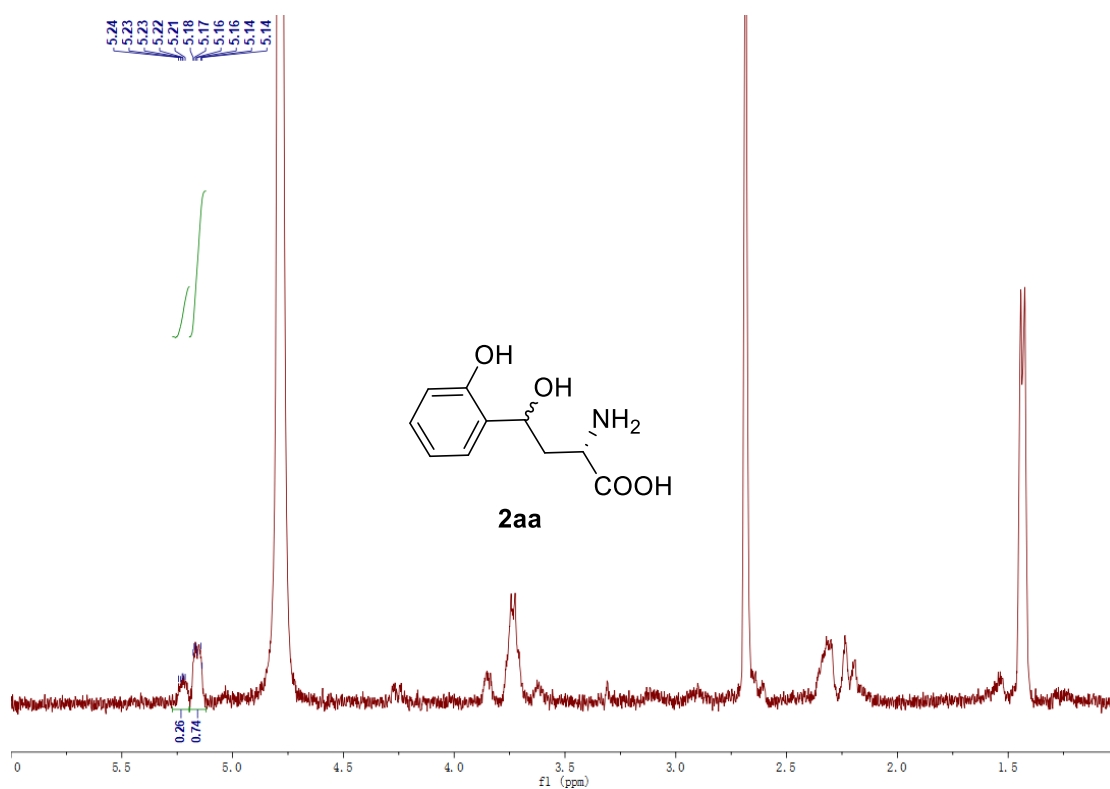


Figure S82. Diastereoisomeric ratio of **2aa**

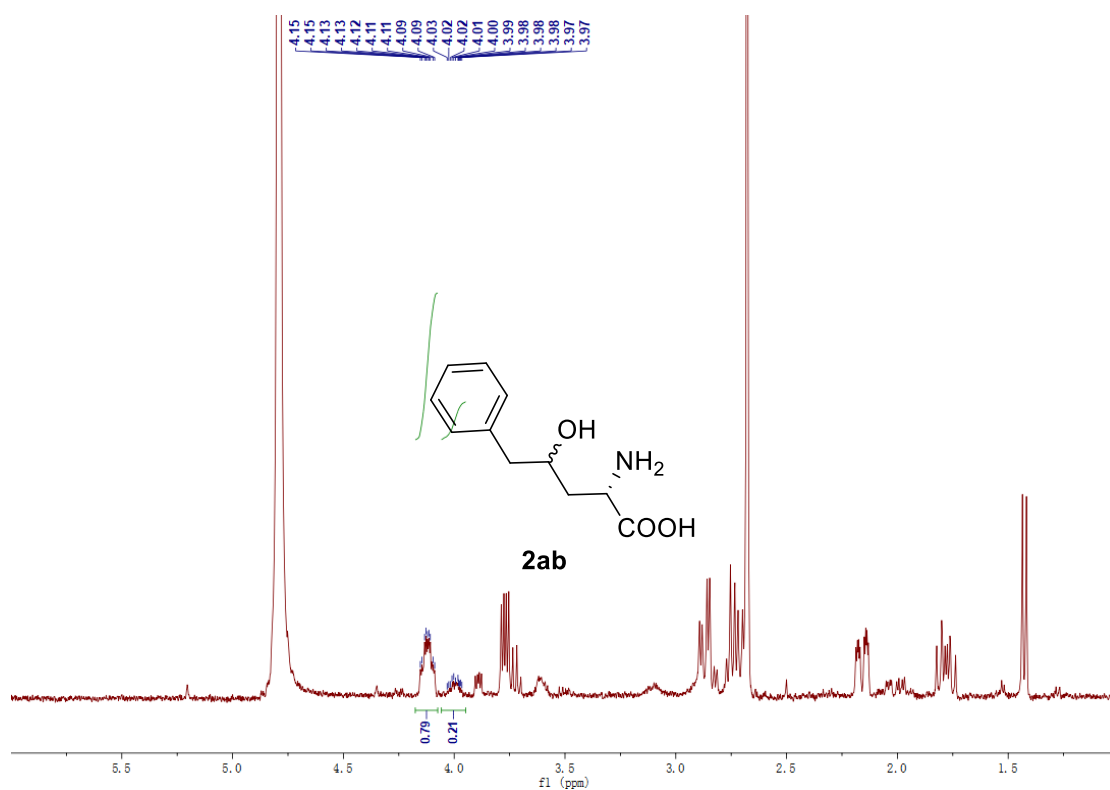


Figure S83. Diastereoisomeric ratio of **2ab**

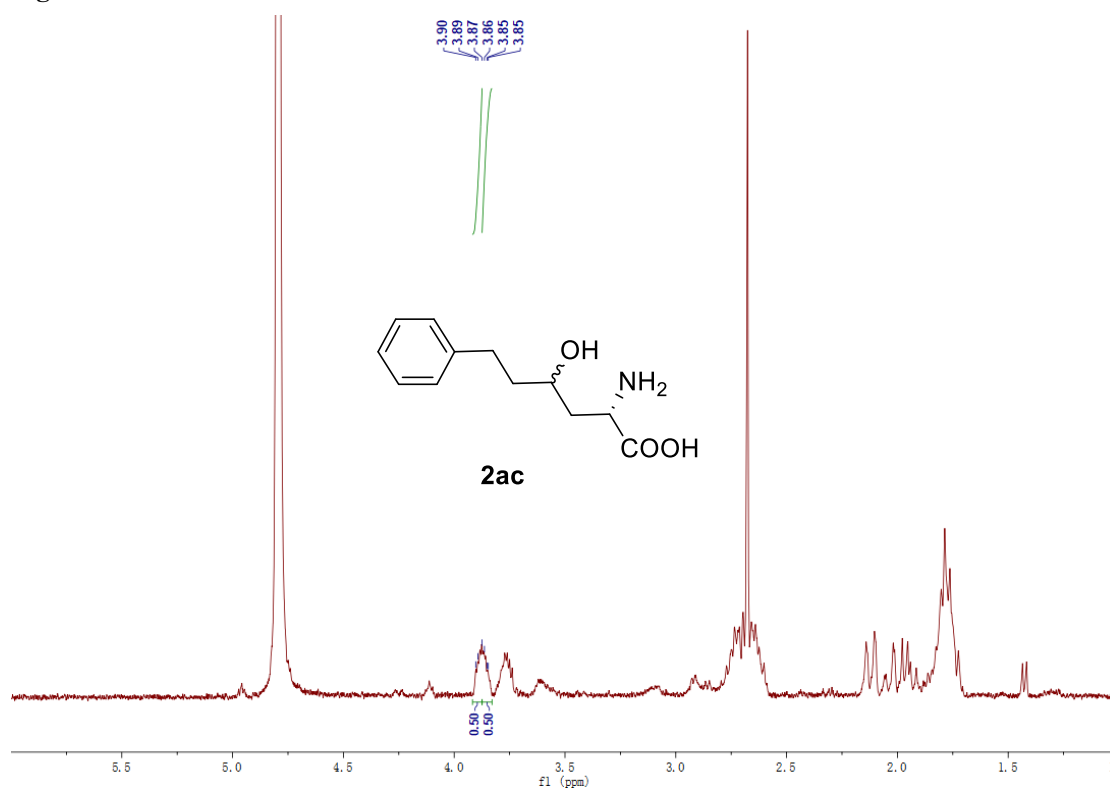


Figure S84. Diastereoisomeric ratio of **2ac**

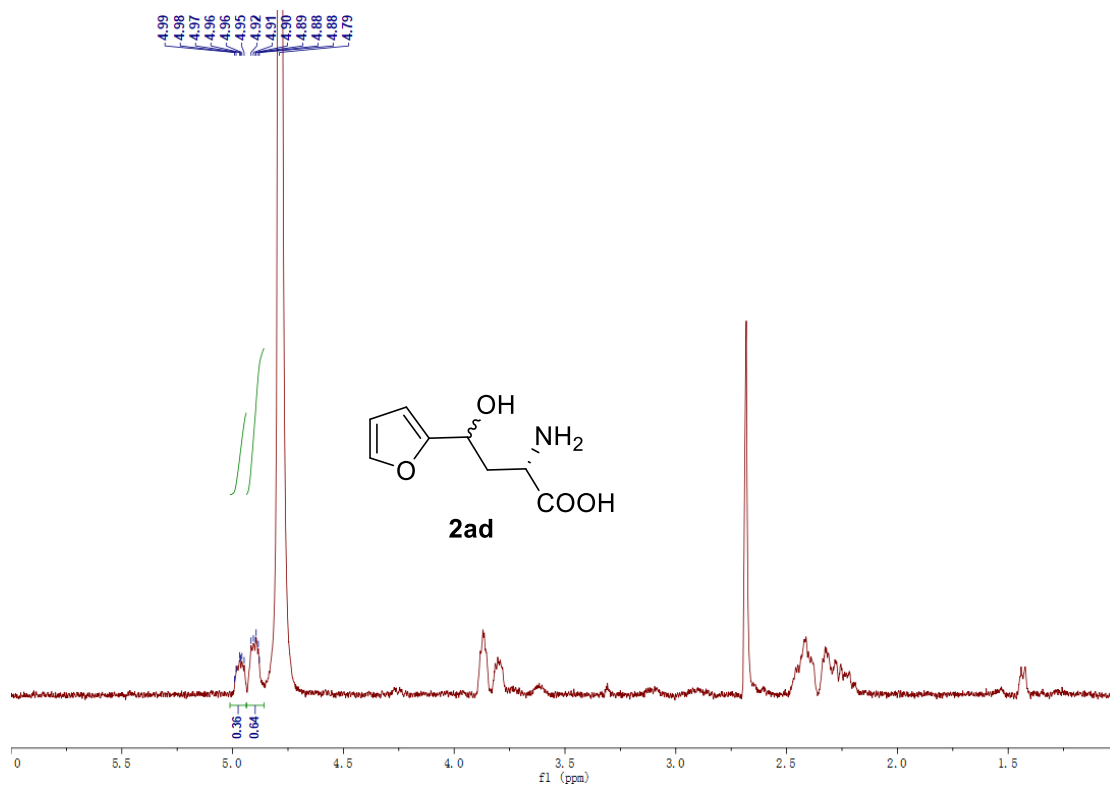


Figure S85. Diastereoisomeric ratio of **2ad**

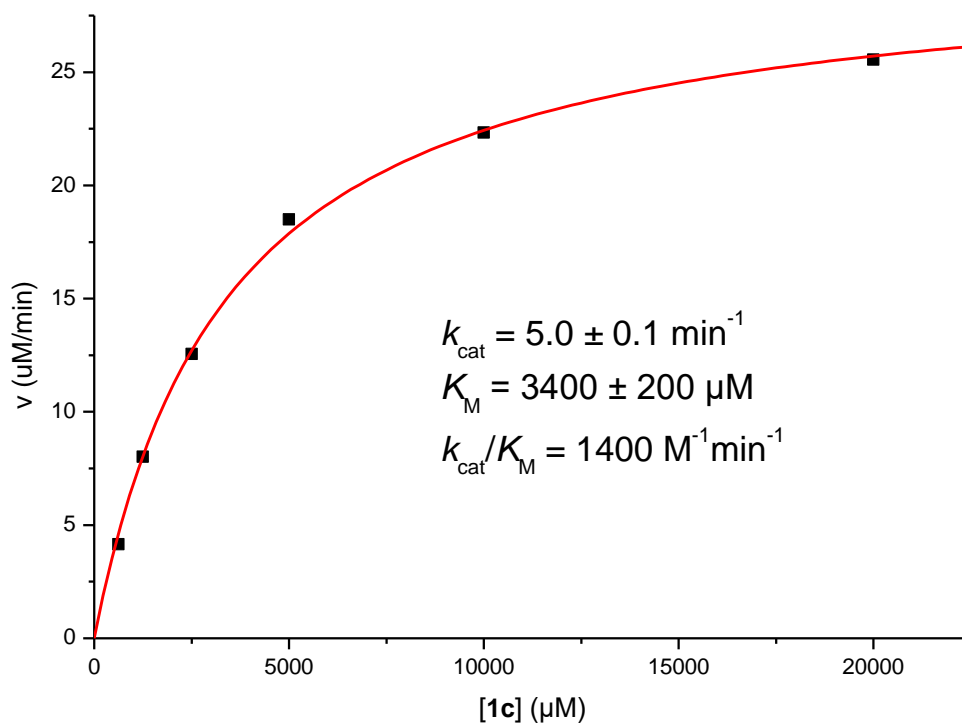


Figure S86. Michaelis-Menten curve and kinetic parameters of **1c**

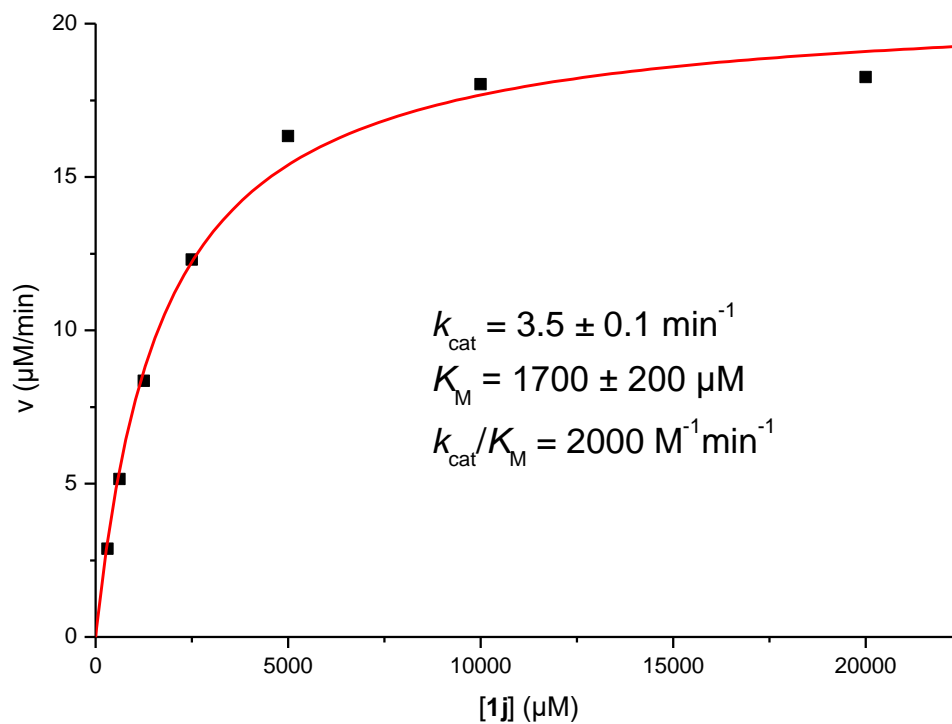


Figure S87. Michaelis-Menten curve and kinetic parameters of **1j**

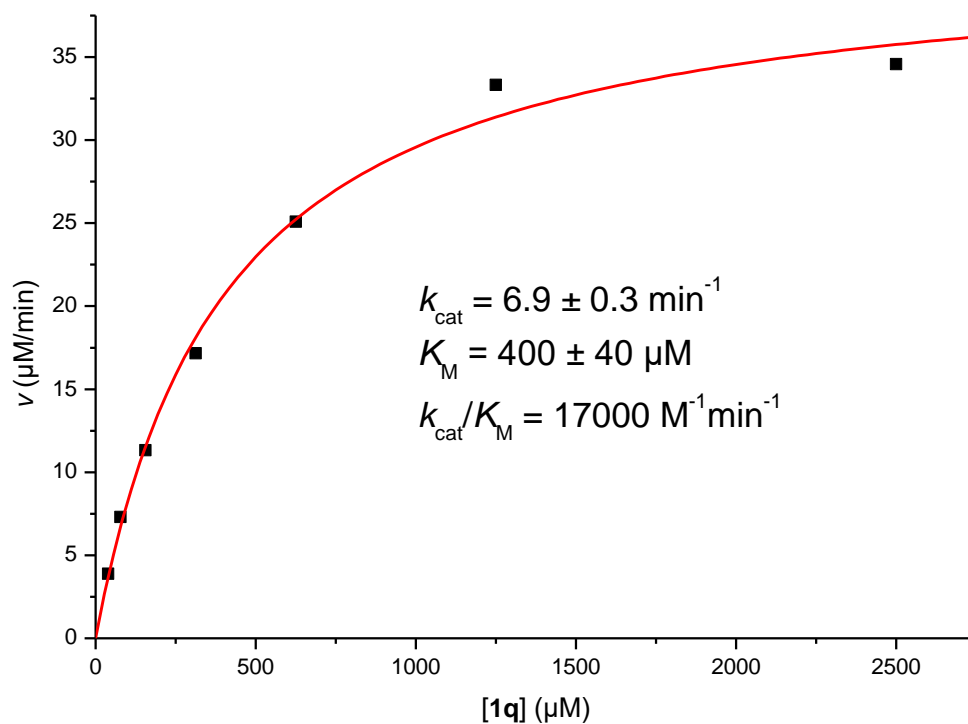


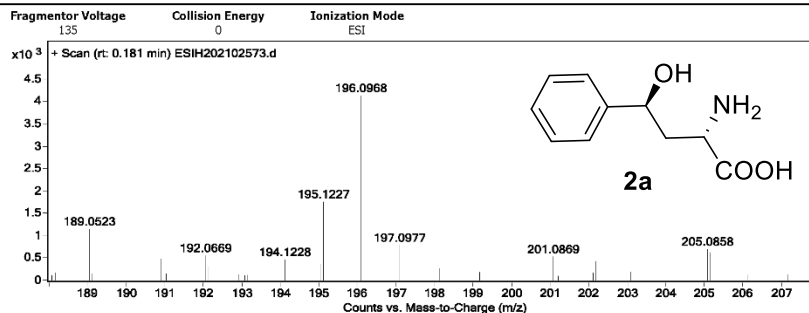
Figure S88. Michaelis-Menten curve and kinetic parameters of **1q**

HR-MS and NMR spectra

Qualitative Analysis Report

Data Filename	ESI202102573.d	Sample Name	G5-G5-BJQ
Sample ID		Position	P1-B6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:47:42	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESI2H by zhuzhenyun

User Spectra



Formula Calculator Results

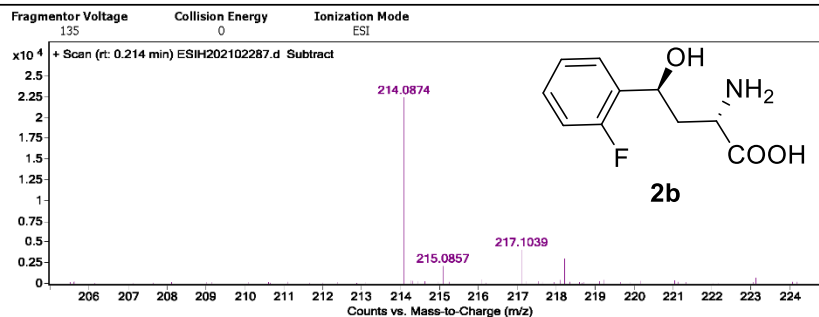
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
196.0968	196.0968	0.05	0.26	C10 H14 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102287.d	Sample Name	G5-AP-2F
Sample ID		Position	P2-A3
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Acquired Time	4/27/2021 18:12:47	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESI2H by zhuzhenyun

User Spectra



Formula Calculator Results

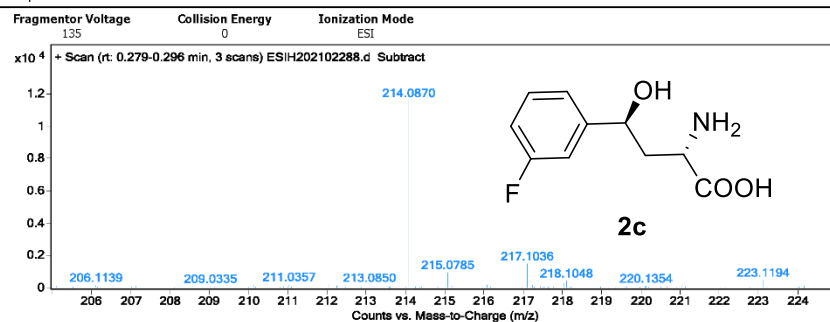
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.0874	214.0874	0.05	0.22	C10 H13 F N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102288.d	Sample Name G5-AP-3F
Sample ID	Position P2-A4
Instrument Name Agilent G6520 Q-TOF	Acq Method 20160322_MS_ESIH_POS_1min.m
Acquired Time 4/27/2021 18:14:14	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

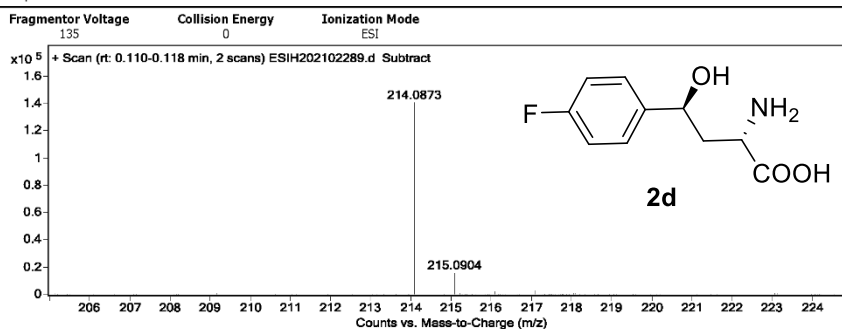
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.087	214.0874	0.38	1.77	C ₁₀ H ₁₃ F N O ₃	(M+H) ⁺

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102289.d	Sample Name G5-AP-4F
Sample ID	Position P2-A5
Instrument Name Agilent G6520 Q-TOF	Acq Method 20160322_MS_ESIH_POS_1min.m
Acquired Time 4/27/2021 18:15:42	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

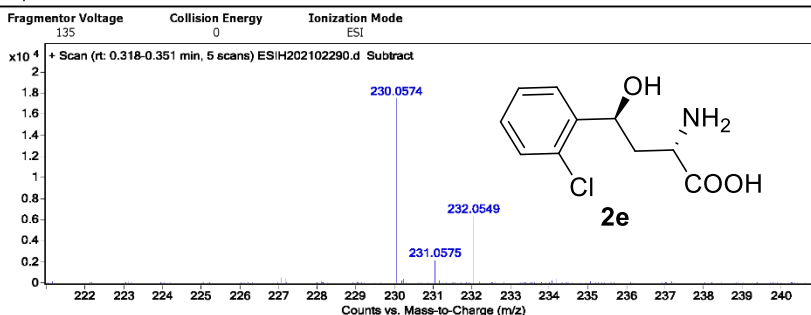
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.0873	214.0874	0.06	0.39	C ₁₀ H ₁₃ F N O ₃	(M+H) ⁺

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102290.d	Sample Name	G5-AP-2Cl
Sample ID		Position	P2-A6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:17:10	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

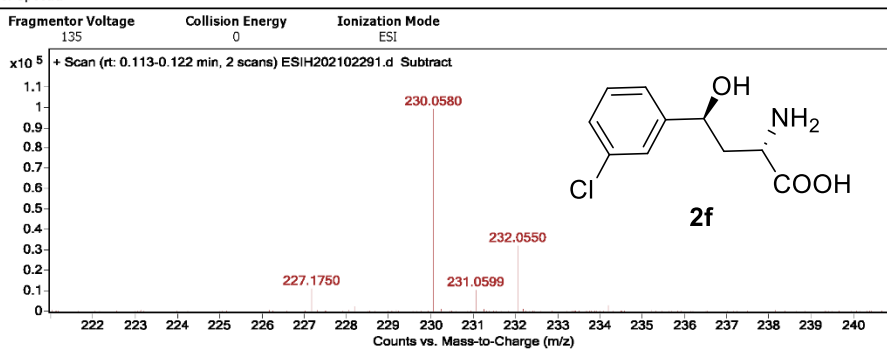
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.0574	230.0578	0.42	1.83	C10 H13 Cl N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102291.d	Sample Name	G5-AP-3Cl
Sample ID		Position	P2-A7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:18:37	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

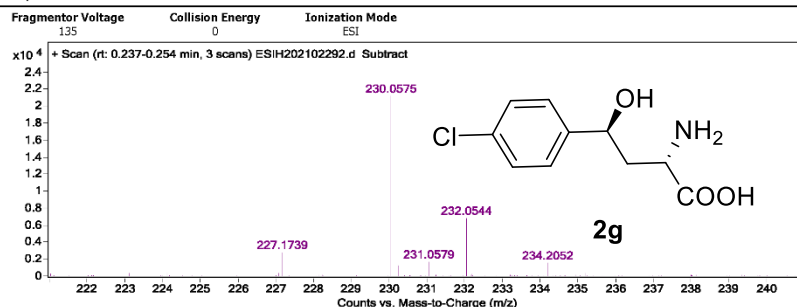
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.058	230.0578	-0.15	-0.64	C10 H13 Cl N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102292.d	Sample Name	G5-AP-4Cl
Sample ID		Position	P2-A8
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESI_H_POS_1min.m
Acquired Time	4/27/2021 18:21:17	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESI2H by zhuzhenyun

User Spectra



Formula Calculator Results

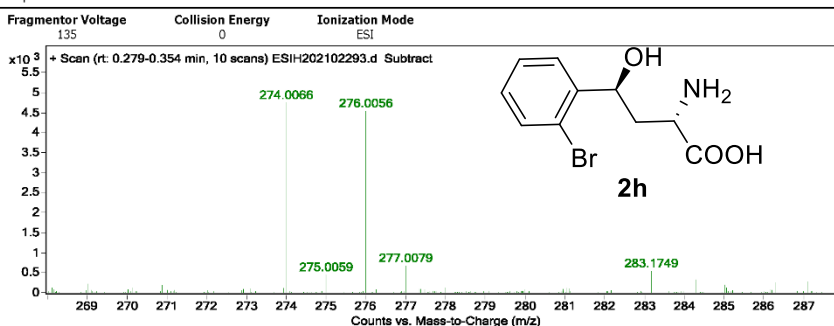
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.0575	230.0578	0.38	1.66	C10 H13 Cl N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102293.d	Sample Name	G5-AP-2Br
Sample ID		Position	P2-A9
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESI_H_POS_1min.m
Acquired Time	4/27/2021 18:22:45	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESI2H by zhuzhenyun

User Spectra



Formula Calculator Results

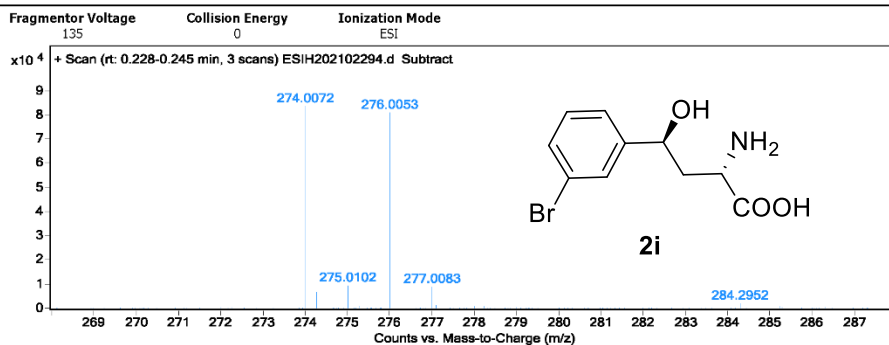
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.0066	274.0073	0.71	2.6	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102294.d	Sample Name G5-AP-3Br
Sample ID	Position P2-B1
Instrument Name Agilent G6520 Q-TOF	Acq Method Z0160322_MS_ESIH_POS_1min.m
Acquired Time 4/27/2021 18:24:14	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

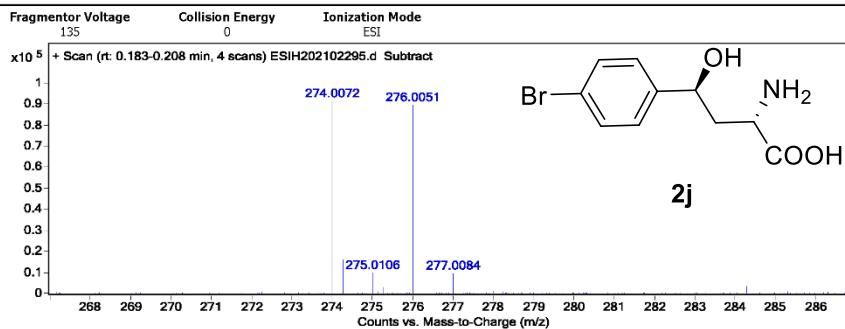
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.0072	274.0073	0.13	0.48	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102295.d	Sample Name G5-AP-4Br
Sample ID	Position P2-B2
Instrument Name Agilent G6520 Q-TOF	Acq Method Z0160322_MS_ESIH_POS_1min.m
Acquired Time 4/27/2021 18:25:42	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

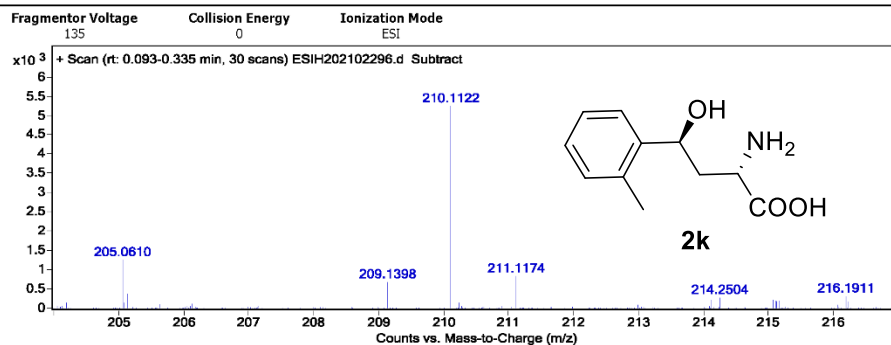
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.0072	274.0073	0.13	0.48	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102296.d	Sample Name	G5-AP-2JJ
Sample ID		Position	P2-B3
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:27:10	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

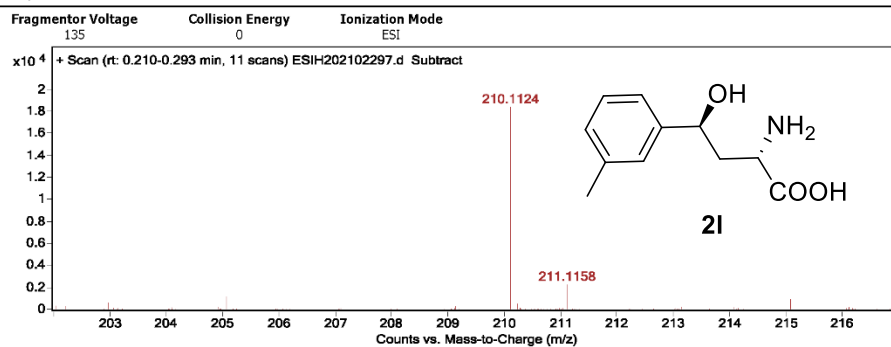
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.1122	210.1125	0.23	1.11	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102297.d	Sample Name	G5-AP-3JJ
Sample ID		Position	P2-B4
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:28:38	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

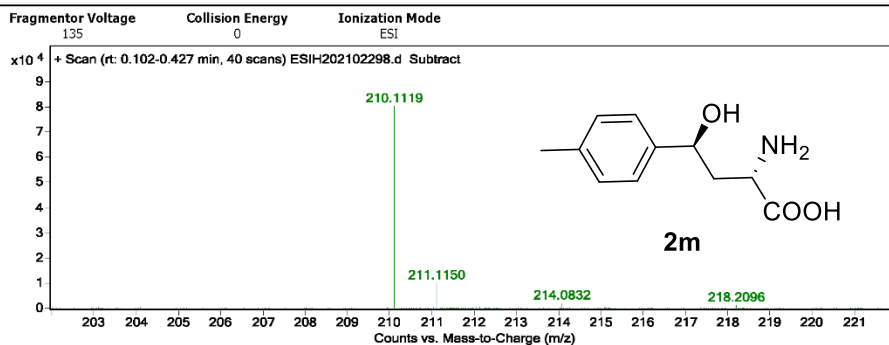
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.1124	210.1125	0.02	0.11	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102298.d	Sample Name	G5-AP-4J
Sample ID		Position	P2-B5
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:30:05	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

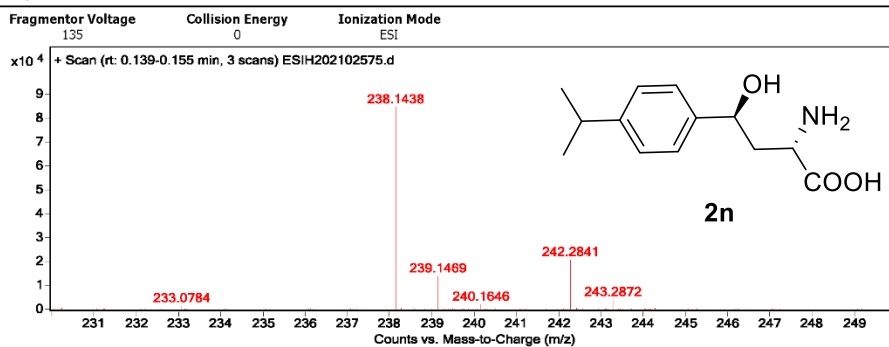
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.1119	210.1125	0.55	2.61	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102575.d	Sample Name	G5-G5-4YB
Sample ID		Position	P1-B8
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:50:16	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

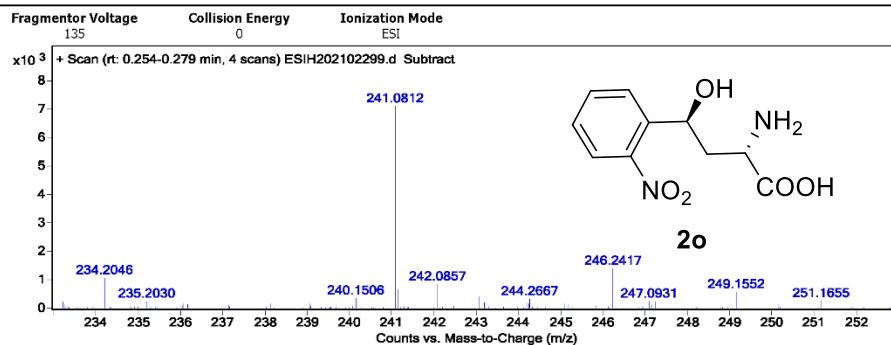
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
238.1438	238.1438	-0.04	-0.17	C13 H20 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102299.d	Sample Name	G5-AP-2NO2
Sample ID		Position	P2-B6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:31:32	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

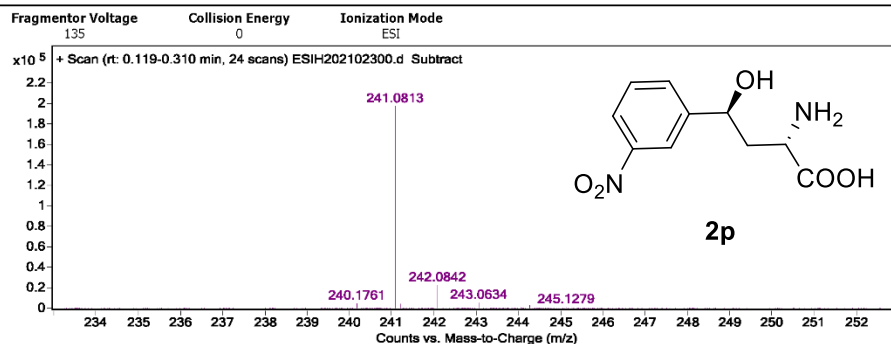
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0812	241.0819	0.67	2.79	C10 H13 N2 O5	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102300.d	Sample Name	G5-AP-3NO2
Sample ID		Position	P2-B7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:33:00	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

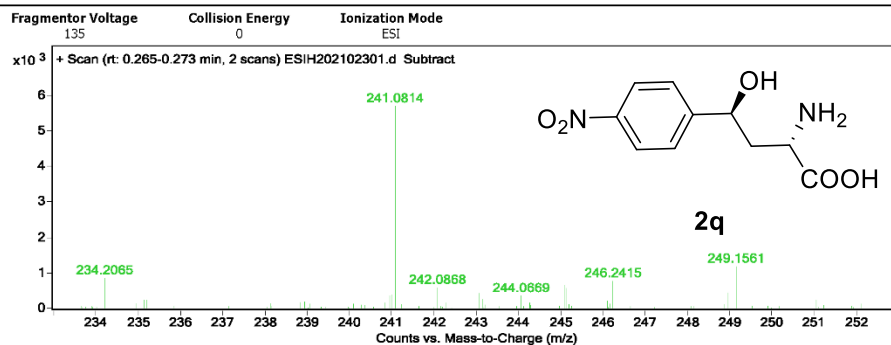
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0813	241.0819	0.63	2.61	C10 H13 N2 O5	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102301.d	Sample Name	G5-AP-4NQ2
Sample ID		Position	P2-B8
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:34:28	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

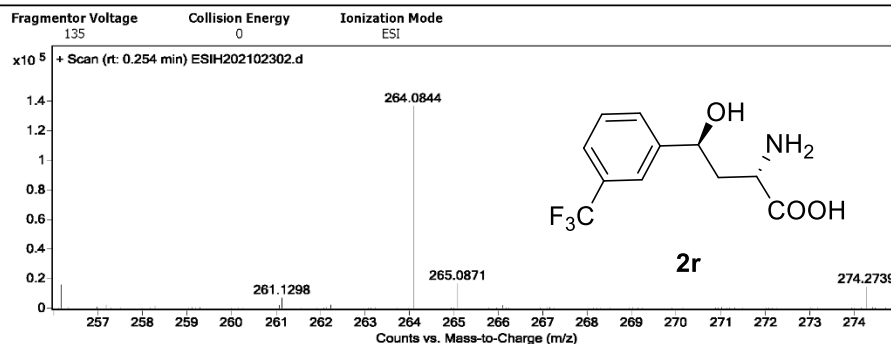
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0814	241.0819	0.45	1.88	C10 H13 N2 O5	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102302.d	Sample Name	G5-AP-3-3F
Sample ID		Position	P2-B9
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:35:55	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

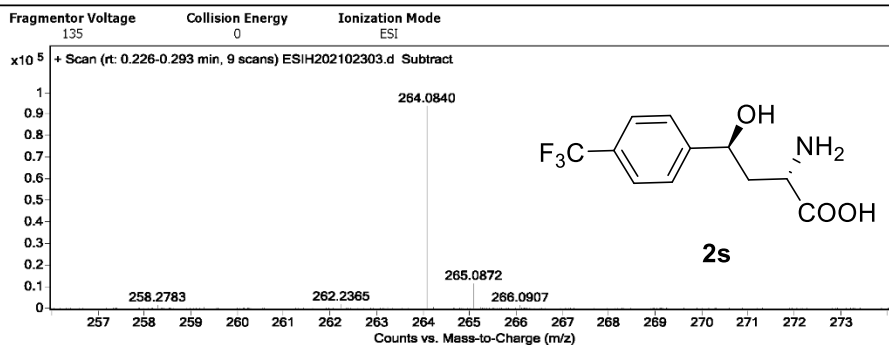
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
264.0844	264.0842	-0.17	-0.66	C11 H13 F3 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102303.d	Sample Name	G5-AP-4-3F
Sample ID		Position	P2-C1
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	4/27/2021 18:37:20	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

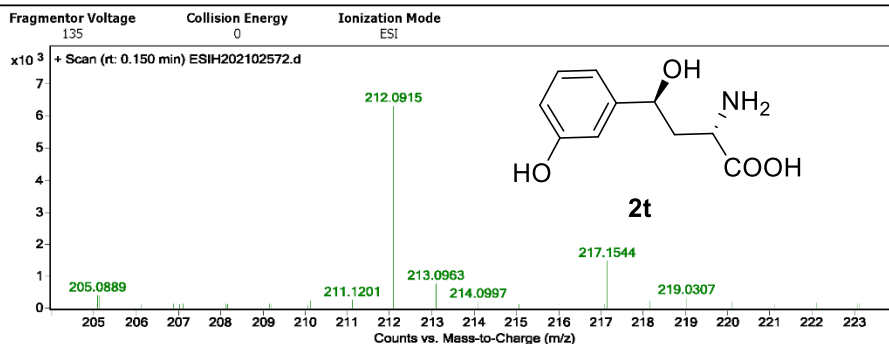
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
264.084	264.0842	0.18	0.69	C11 H13 F3 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102572.d	Sample Name	G5-G5-3QJ
Sample ID		Position	P1-B5
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:46:25	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

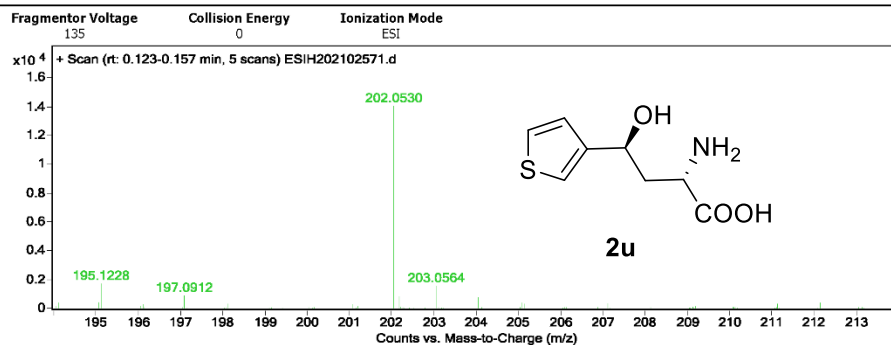
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
212.0915	212.0917	0.23	1.07	C10 H14 N O4	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102571.d	Sample Name	G5-G5-3SF
Sample ID		Position	P1-B4
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:45:08	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

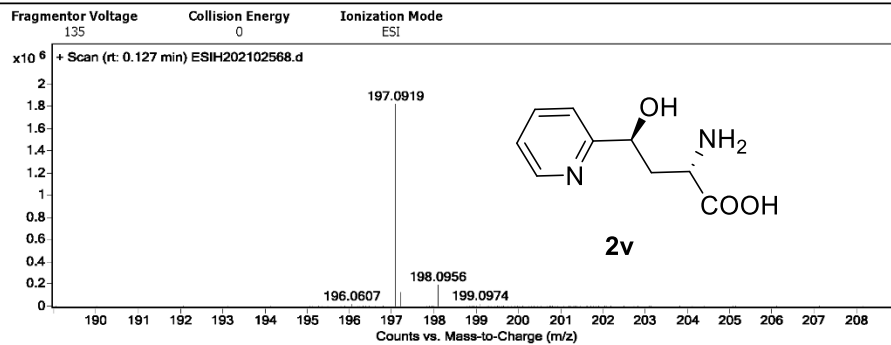
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
202.053	202.0532	0.19	0.96	C8 H12 N O3 S	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102568.d	Sample Name	G5-2BD
Sample ID		Position	P1-B1
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:41:17	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0919	197.0921	0.12	0.61	C9 H13 N2 O3	(M+H)+

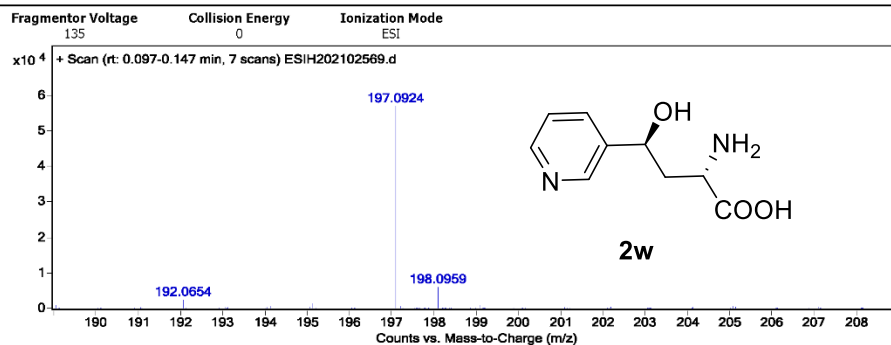
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102569.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/13/2021 11:42:34
DA Method small molecular data analysis method.m

Sample Name G5-3BD
Position P1-B2
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0924	197.0921	-0.37	-1.88	C ₉ H ₁₃ N ₂ O ₃	(M+H) ⁺

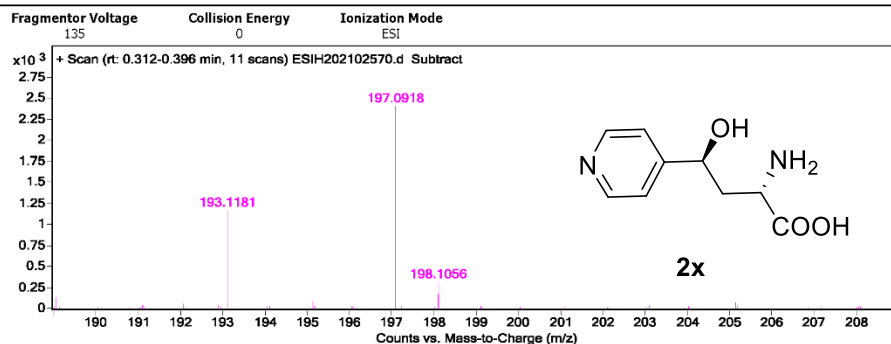
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102570.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/13/2021 11:43:51
DA Method small molecular data analysis method.m

Sample Name G5-4BD
Position P1-B3
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

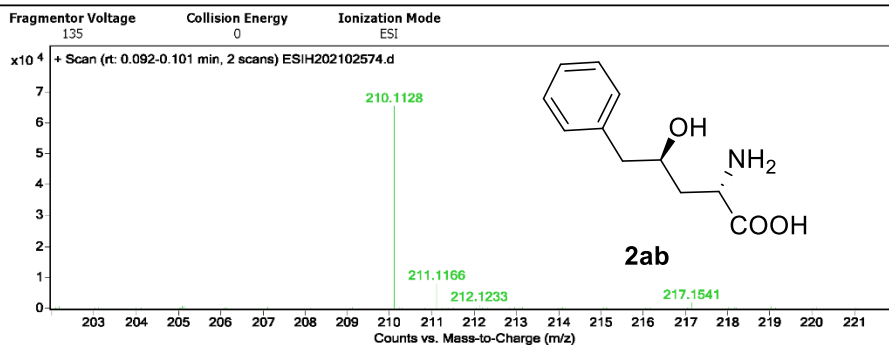
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0918	197.0921	0.23	1.18	C ₉ H ₁₃ N ₂ O ₃	(M+H) ⁺

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102574.d	Sample Name	G5-G5-BYQ
Sample ID		Position	P1-B7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/13/2021 11:49:00	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

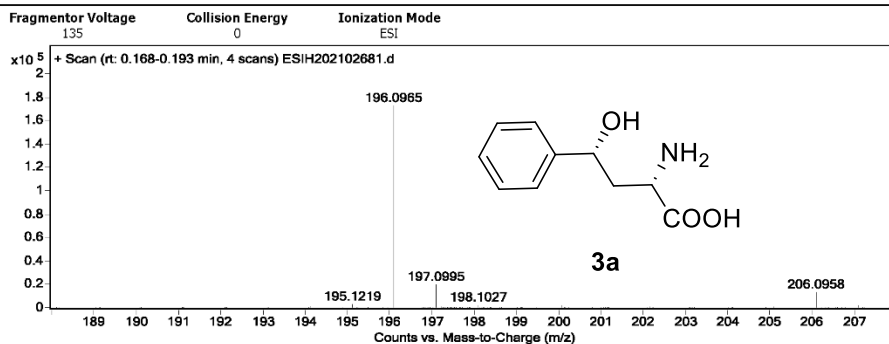
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.1128	210.1125	-0.32	-1.54	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102681.d	Sample Name	G5-G5-PFK-BJQ
Sample ID		Position	P1-B5
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:29:24	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

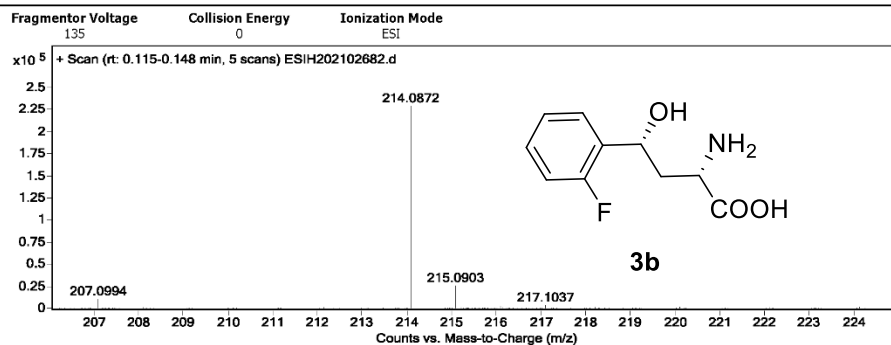
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
196.0965	196.0968	0.34	1.73	C10 H14 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102682.d	Sample Name	G5-G5-PFK-2F
Sample ID		Position	P1-B6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:30:41	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

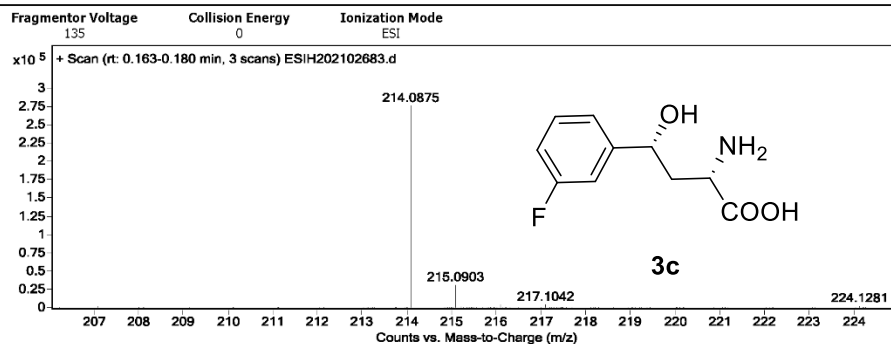
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.0872	214.0874	0.19	0.87	C10 H13 F N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102683.d	Sample Name	G5-G5-PFK-3F
Sample ID		Position	P1-B7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:31:58	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

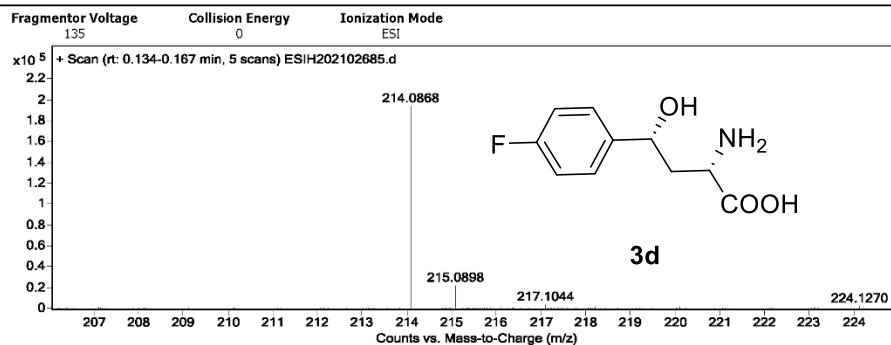
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.0875	214.0874	-0.14	-0.63	C10 H13 F N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102685.d	Sample Name	G5-G5-PFK-4F
Sample ID		Position	P1-B9
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:34:30	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

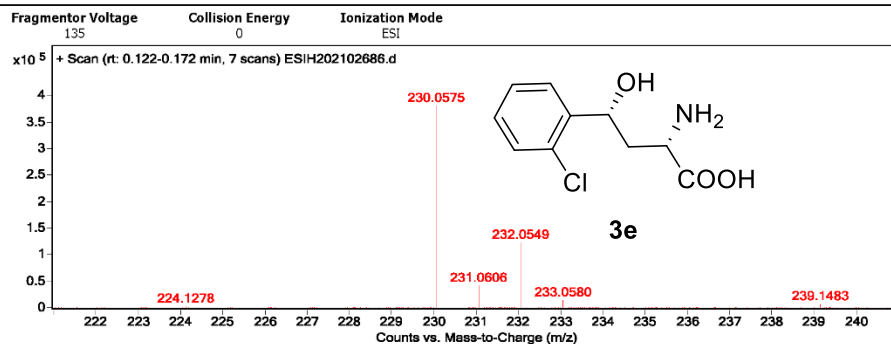
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
214.0868	214.0874	0.62	2.89	C10 H13 F N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102686.d	Sample Name	G5-G5-PFK-2CL
Sample ID		Position	P1-C1
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:35:44	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.0575	230.0578	0.31	1.36	C10 H13 Cl N O3	(M+H)+

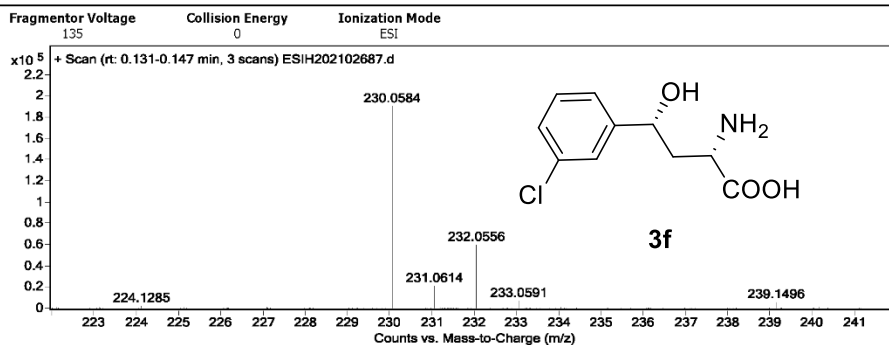
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102687.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/17/2021 19:37:01
DA Method small molecular data analysis method.m

Sample Name G5-G5-PFK-3CL
Position P1-C2
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.0584	230.0578	-0.57	-2.47	C10 H13 Cl N O3	(M+H)+

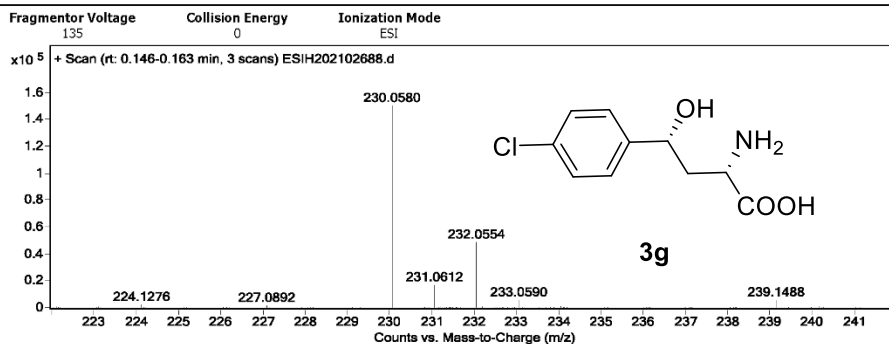
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102688.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/17/2021 19:38:17
DA Method small molecular data analysis method.m

Sample Name G5-G5-PFK-4CL
Position P1-C3
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

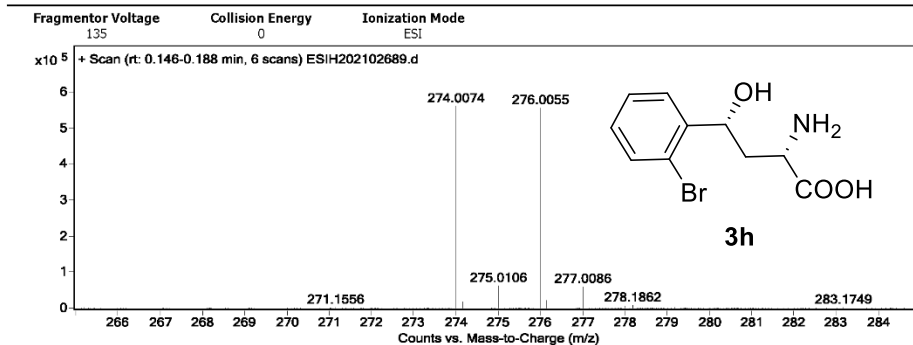
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
230.0580	230.0578	-0.18	-0.8	C10 H13 Cl N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102689.d Sample ID Instrument Name Agilent G6520 Q-TOF Acquired Time 5/17/2021 19:39:34 DA Method small molecular data analysis method.m	Sample Name G5-G5-PFK-2BR Position P1-C4 Acq Method 20160322_MS_ESIH_POS_1min.m IRM Calibration Status Success Comment ESIH by zhuzhenyun	
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User Spectra



Formula Calculator Results

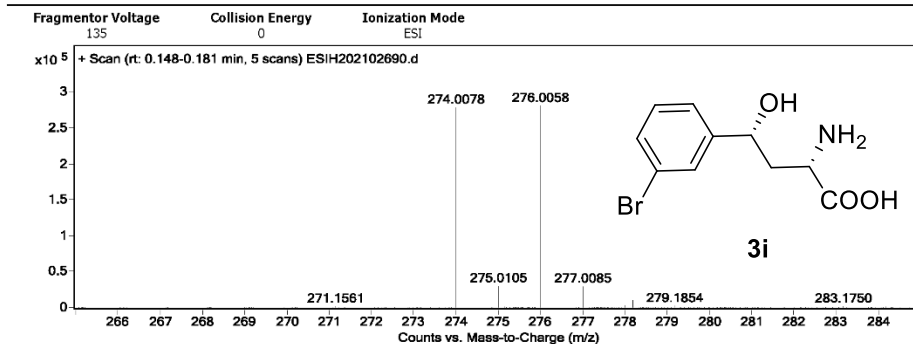
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.0074	274.0073	-0.08	-0.28	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102690.d Sample ID Instrument Name Agilent G6520 Q-TOF Acquired Time 5/17/2021 19:40:51 DA Method small molecular data analysis method.m	Sample Name G5-G5-PFK-3BR Position P1-C5 Acq Method 20160322_MS_ESIH_POS_1min.m IRM Calibration Status Success Comment ESIH by zhuzhenyun	
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User Spectra



Formula Calculator Results

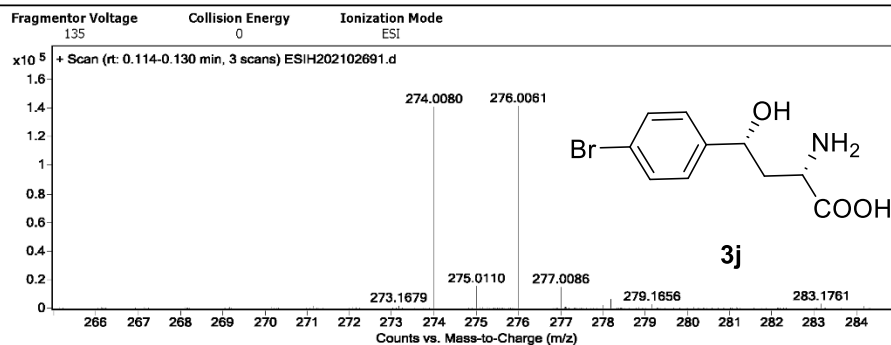
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.0078	274.0073	-0.42	-1.53	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102691.d	Sample Name	G5-G5-PFK-4BR
Sample ID		Position	P1-C6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:42:08	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

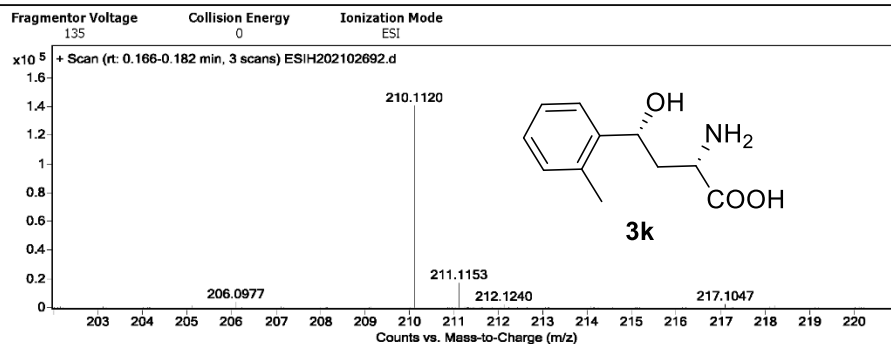
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
274.008	274.0073	-0.66	-2.42	C10 H13 Br N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102692.d	Sample Name	G5-G5-PFK-2JJ
Sample ID		Position	P1-C7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:43:26	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

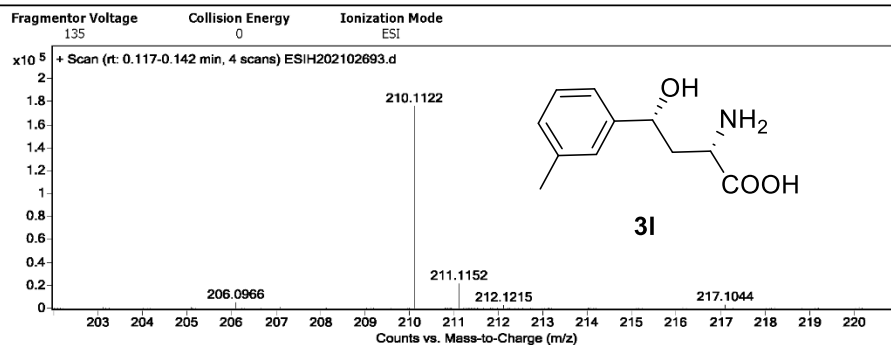
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.112	210.1125	0.52	2.46	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102693.d	Sample Name G5-G5-PFK-3JJ
Sample ID	Position P1-C8
Instrument Name Agilent G6520 Q-TOF	Acq Method 20160322_MS_ESIH_POS_1min.m
Acquired Time 5/17/2021 19:44:43	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

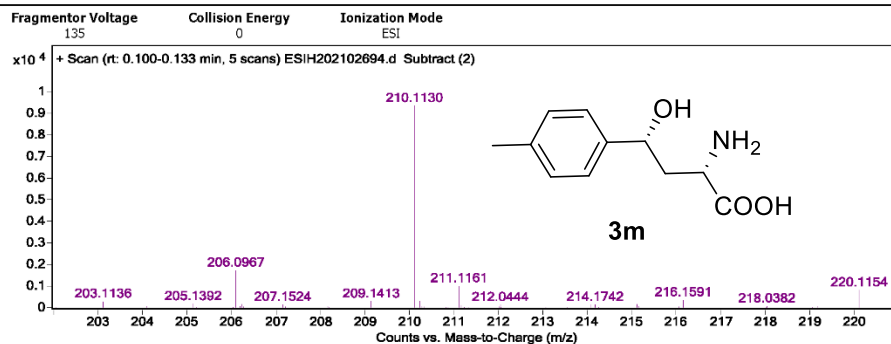
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.1122	210.1125	0.22	1.05	C11 H16 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102694.d	Sample Name G5-G5-PFK-4JJ
Sample ID	Position P1-C9
Instrument Name Agilent G6520 Q-TOF	Acq Method 20160322_MS_ESIH_POS_1min.m
Acquired Time 5/17/2021 19:46:00	IRM Calibration Status Success
DA Method small molecular data analysis method.m	Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
210.113	210.1125	-0.48	-2.31	C11 H16 N O3	(M+H)+

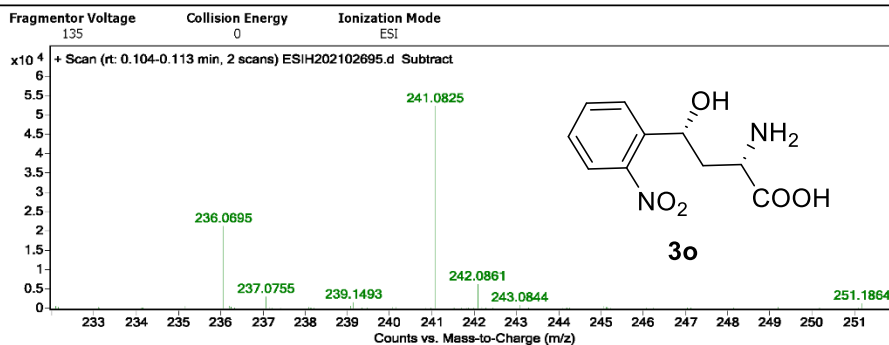
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102695.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/17/2021 19:47:16
DA Method small molecular data analysis method.m

Sample Name G5-G5-PFK-2XJ
Position P1-D1
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0825	241.0819	-0.65	-2.7	C10 H13 N2 O5	(M+H)+

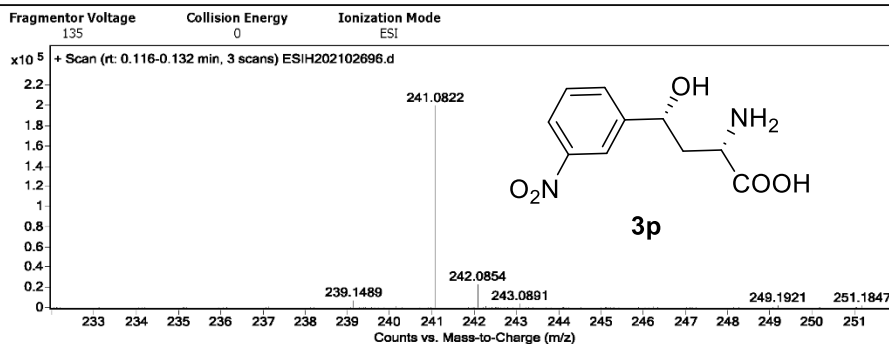
--- End Of Report ---

Qualitative Analysis Report

Data Filename ESIH202102696.d
Sample ID
Instrument Name Agilent G6520 Q-TOF
Acquired Time 5/17/2021 19:48:33
DA Method small molecular data analysis method.m

Sample Name G5-G5-PFK-3XJ
Position P1-D2
Acq Method 20160322_MS_ESIH_POS_1min.m
IRM Calibration Status Success
Comment ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

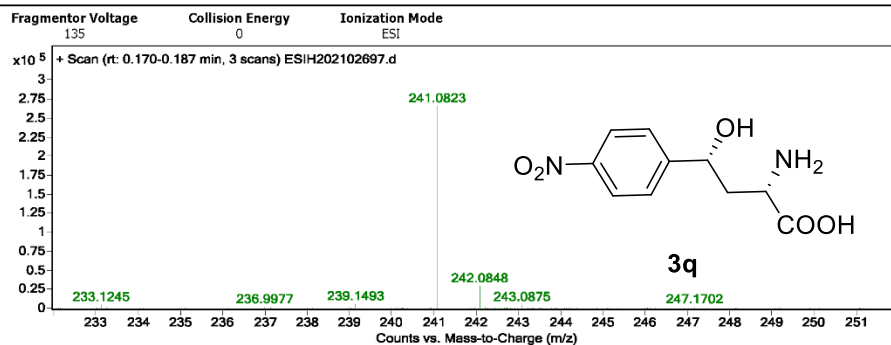
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0822	241.0819	-0.28	-1.16	C10 H13 N2 O5	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102697.d	Sample Name	G5-G5-PFK-4XJ
Sample ID		Position	P1-D3
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:49:49	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

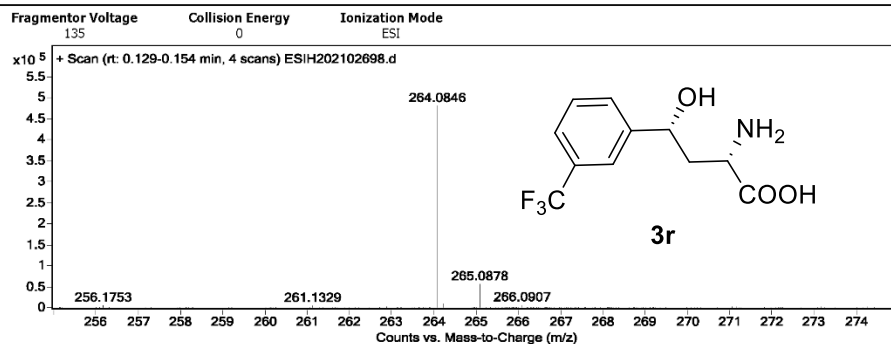
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
241.0823	241.0819	-0.38	-1.57	C10 H13 N2 O5	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102698.d	Sample Name	G5-G5-PFK-3-3F
Sample ID		Position	P1-D4
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:51:07	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

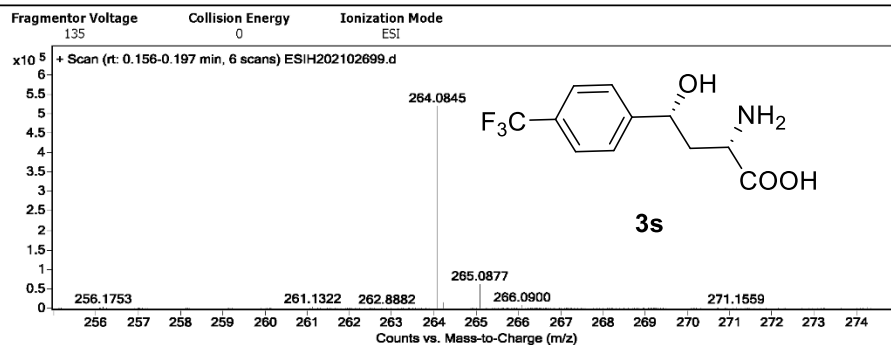
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
264.0846	264.0842	-0.37	-1.4	C11 H13 F3 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102699.d	Sample Name	G5-G5-PFK-4-3F
Sample ID		Position	P1-D5
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:52:24	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

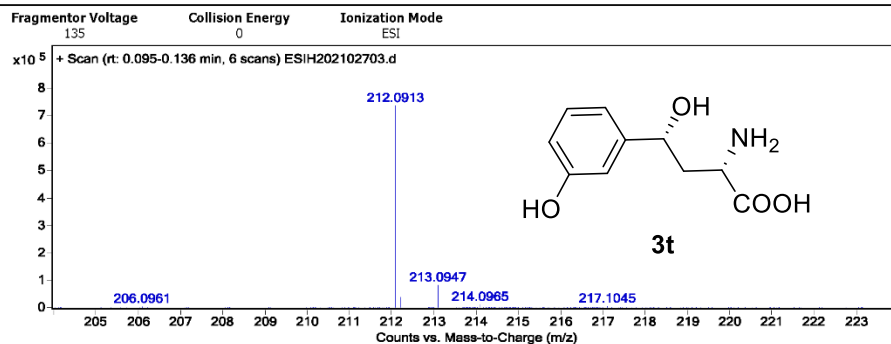
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
264.0845	264.0842	-0.25	-0.96	C11 H13 F3 N O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102703.d	Sample Name	G5-G5-PFK-2QJ
Sample ID		Position	P1-D9
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:57:32	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

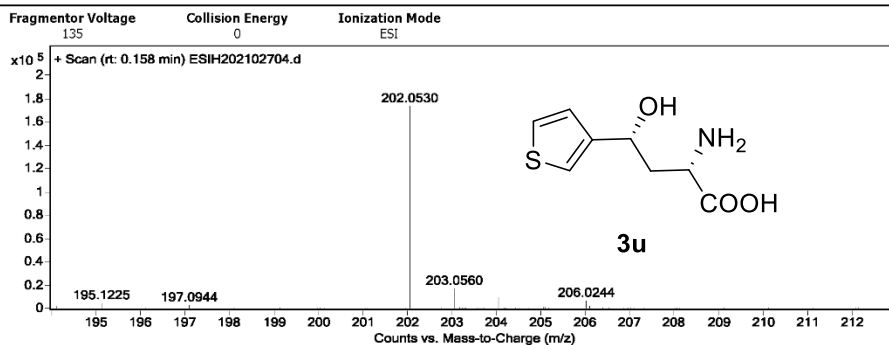
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
212.0913	212.0917	0.42	1.98	C10 H14 N O4	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102704.d	Sample Name	G5-G5-PFK-35F
Sample ID		Position	P1-E1
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:58:48	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

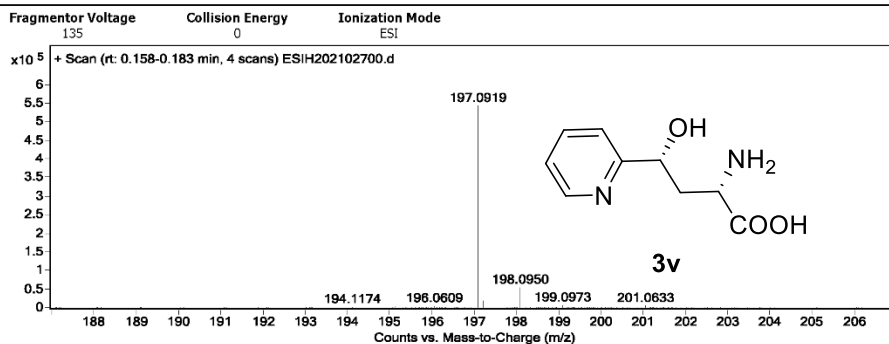
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
202.053	202.0532	0.2	0.98	C8 H12 N O3 S	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102700.d	Sample Name	G5-G5-PFK-2BD
Sample ID		Position	P1-D6
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:53:40	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

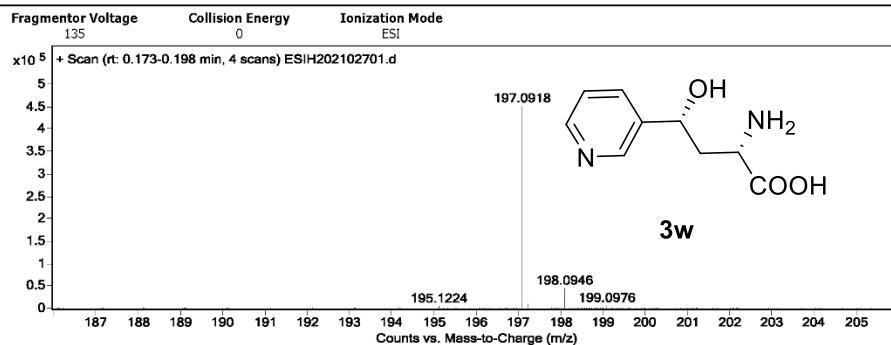
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0919	197.0921	0.13	0.66	C9 H13 N2 O3	(M+H)+

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102701.d	Sample Name	G5-G5-PFK-3BD
Sample ID		Position	P1-D7
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:54:58	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

User Spectra



Formula Calculator Results

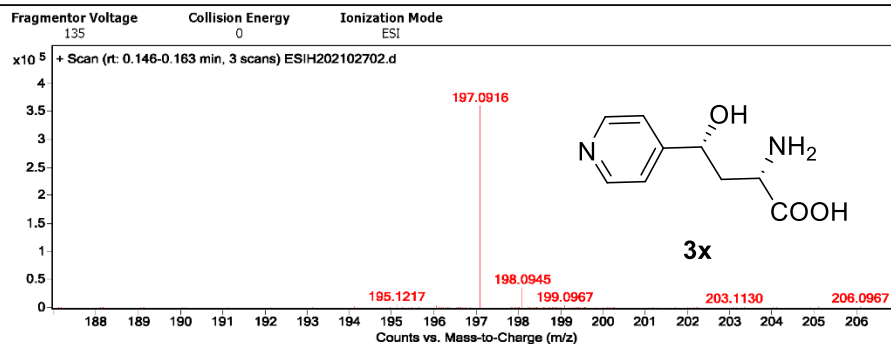
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0918	197.0921	0.28	1.42	C ₉ H ₁₃ N ₂ O ₃	(M+H) ⁺

--- End Of Report ---

Qualitative Analysis Report

Data Filename	ESI202102702.d	Sample Name	G5-G5-PFK-4BD
Sample ID		Position	P1-D8
Instrument Name	Agilent G6520 Q-TOF	Acq Method	20160322_MS_ESIH_POS_1min.m
Acquired Time	5/17/2021 19:56:15	IRM Calibration Status	Success
DA Method	small molecular data analysis method.m	Comment	ESIH by zhuzhenyun

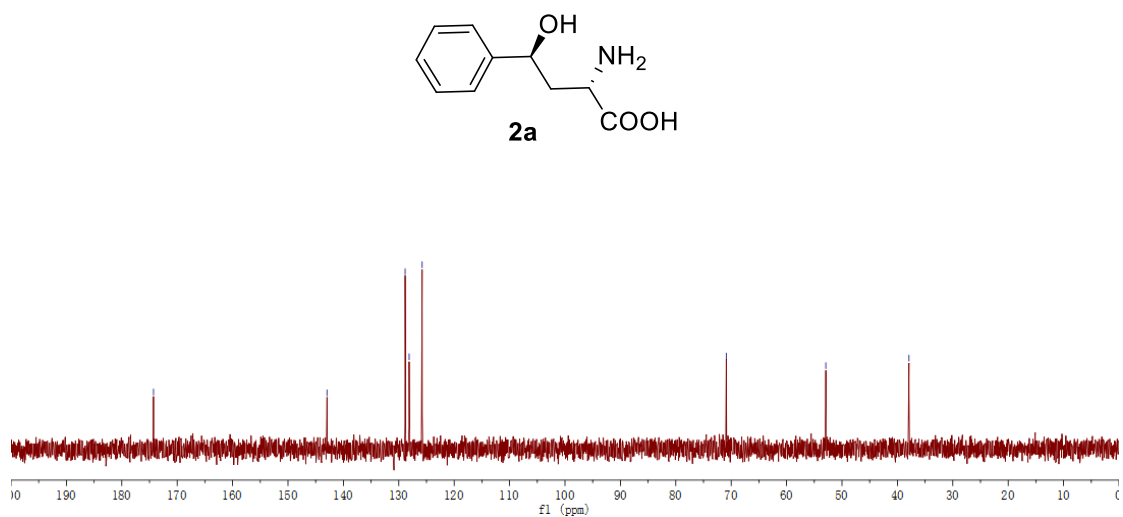
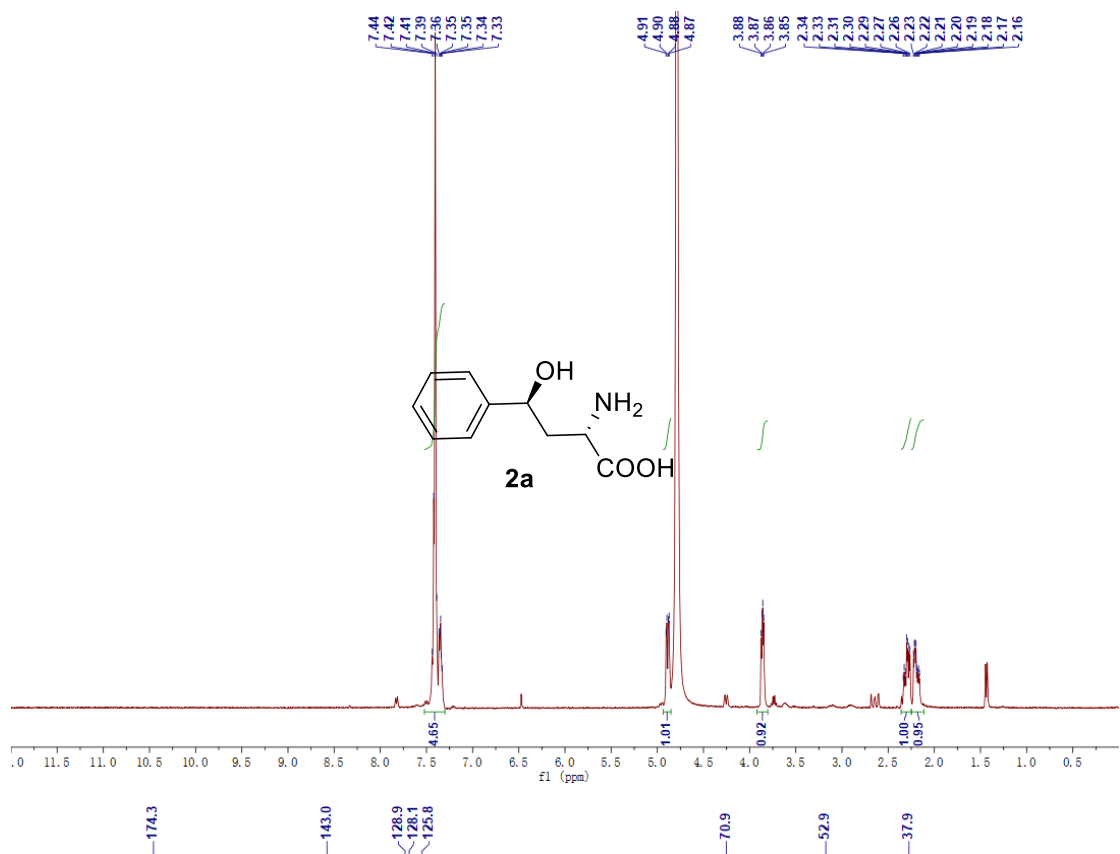
User Spectra

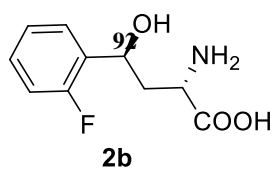
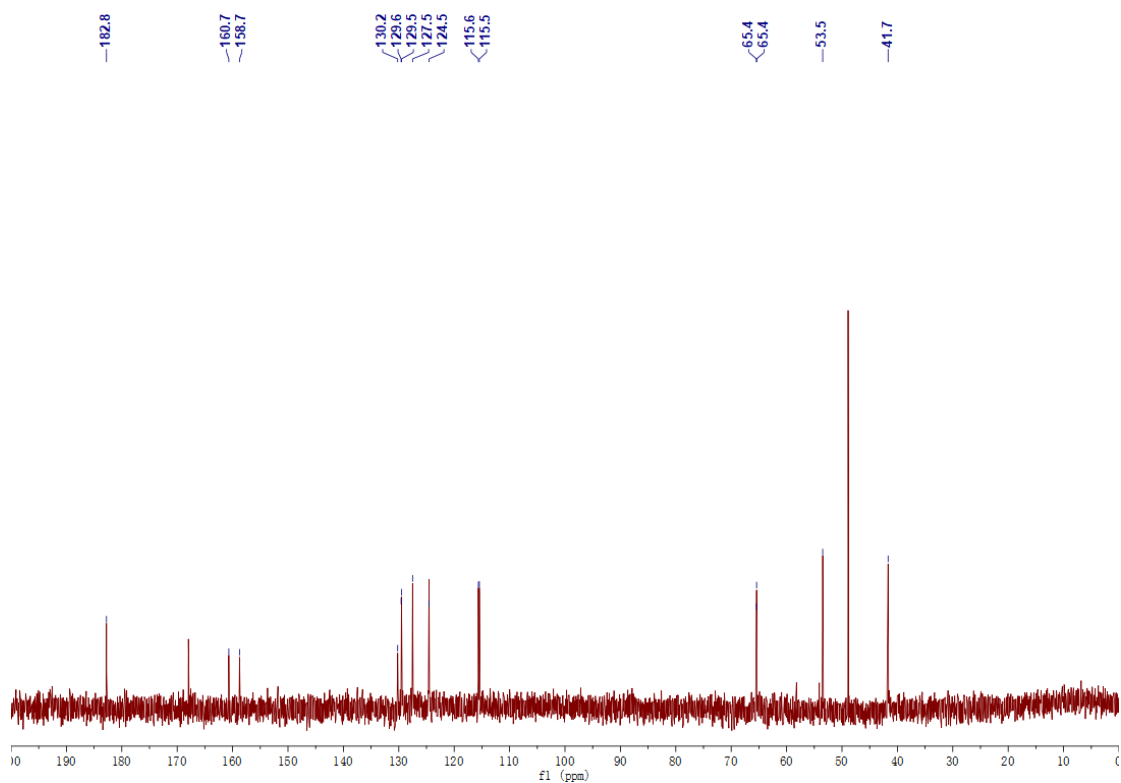
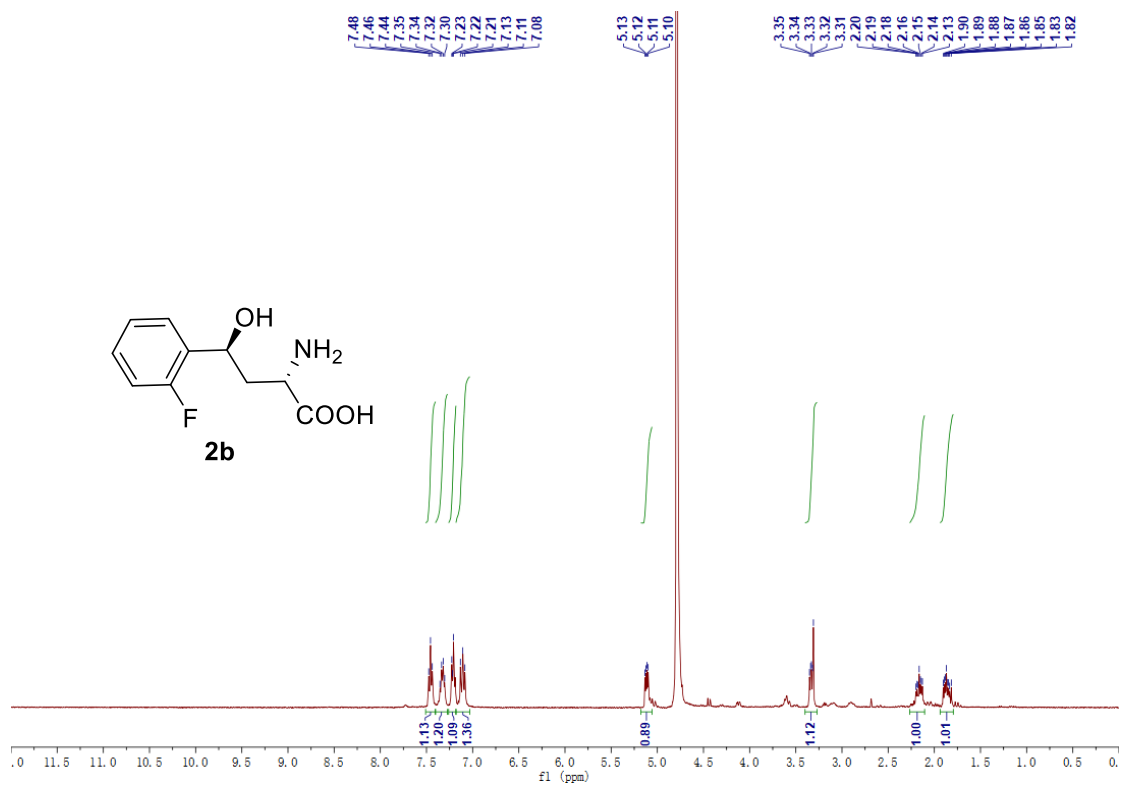


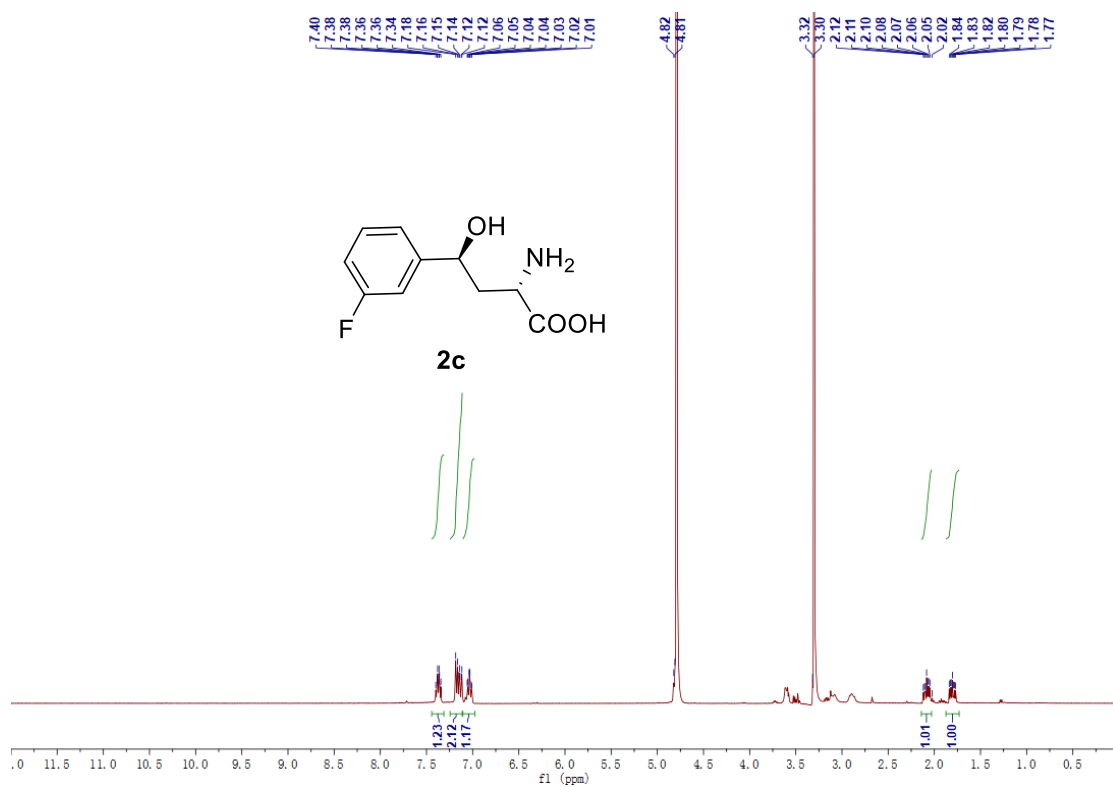
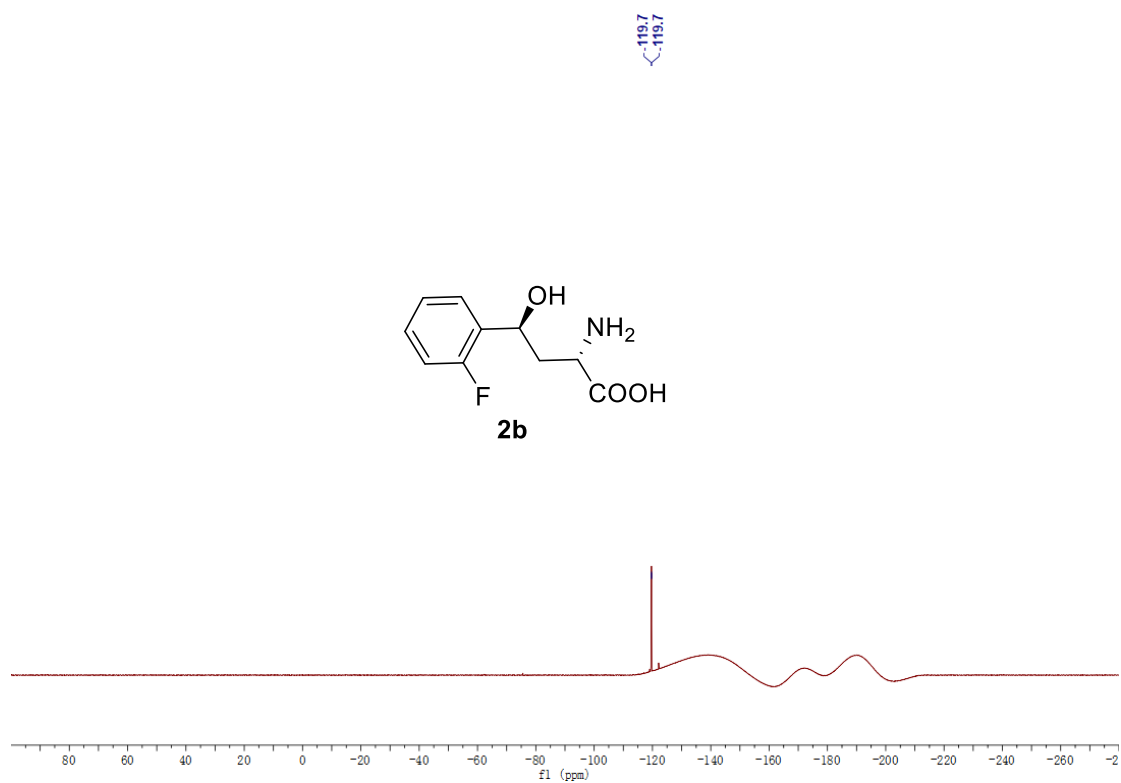
Formula Calculator Results

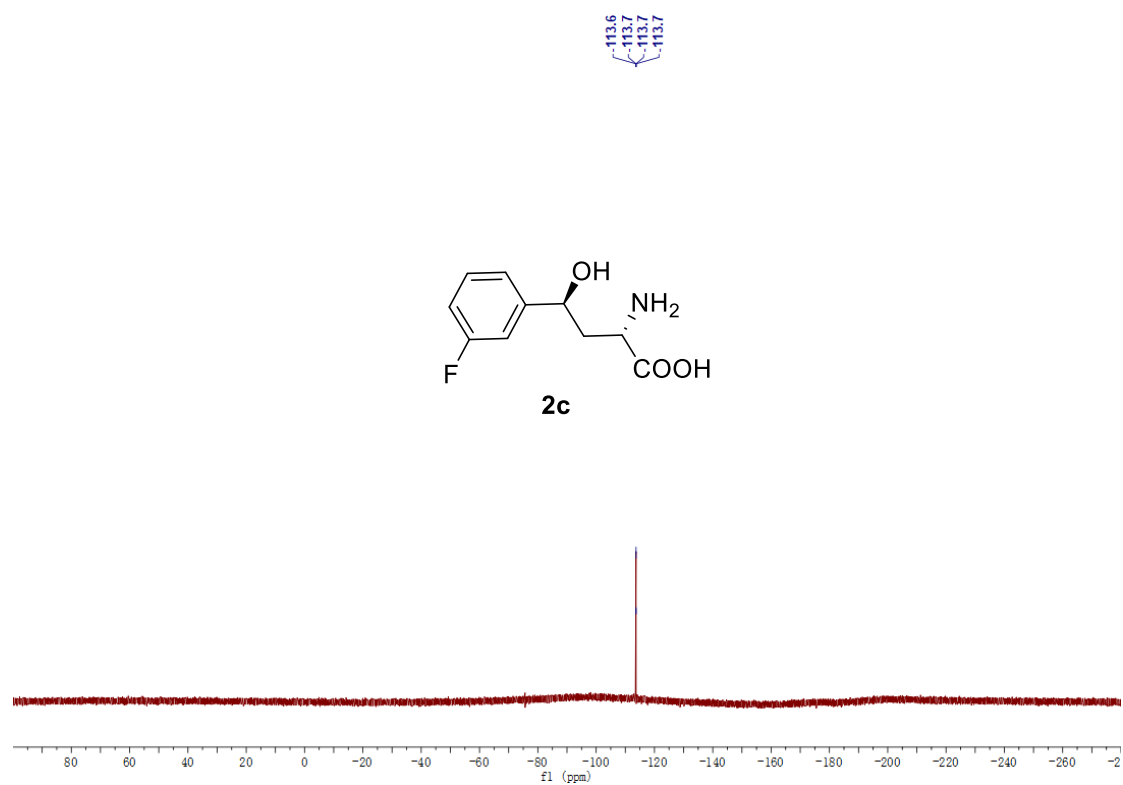
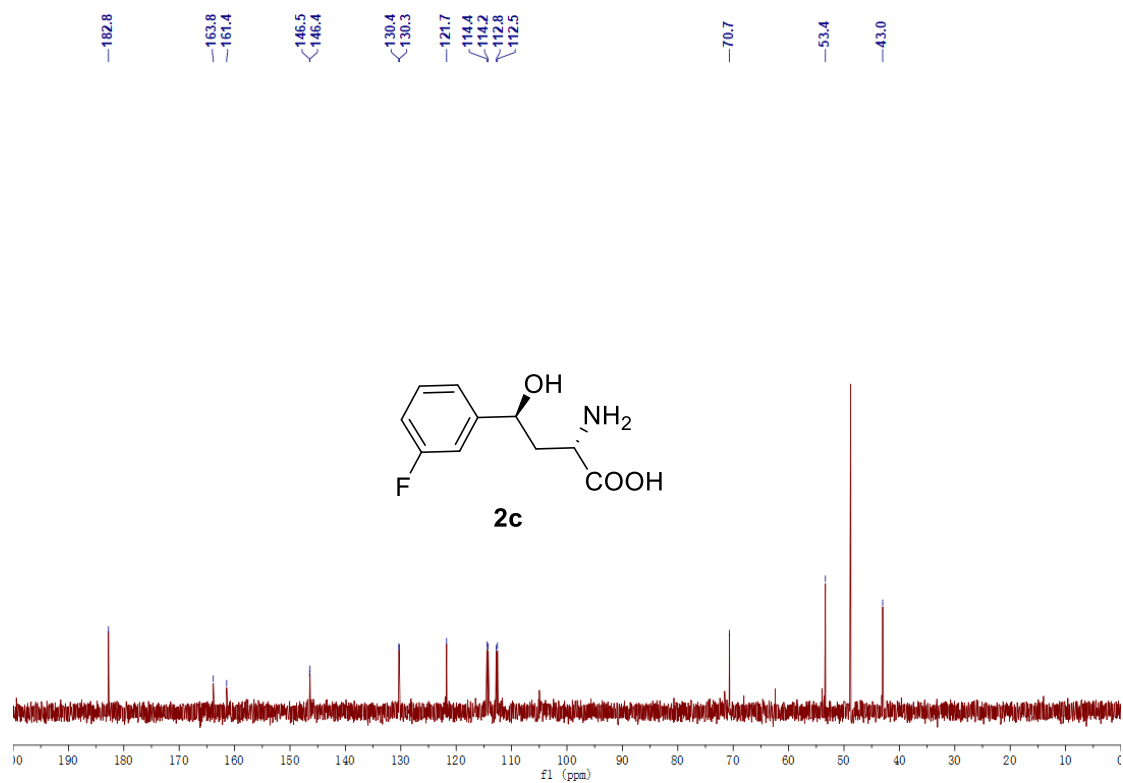
m/z	Calc m/z	Diff (mDa)	Diff (ppm)	Ion Formula	Ion
197.0916	197.0921	0.5	2.53	C ₉ H ₁₃ N ₂ O ₃	(M+H) ⁺

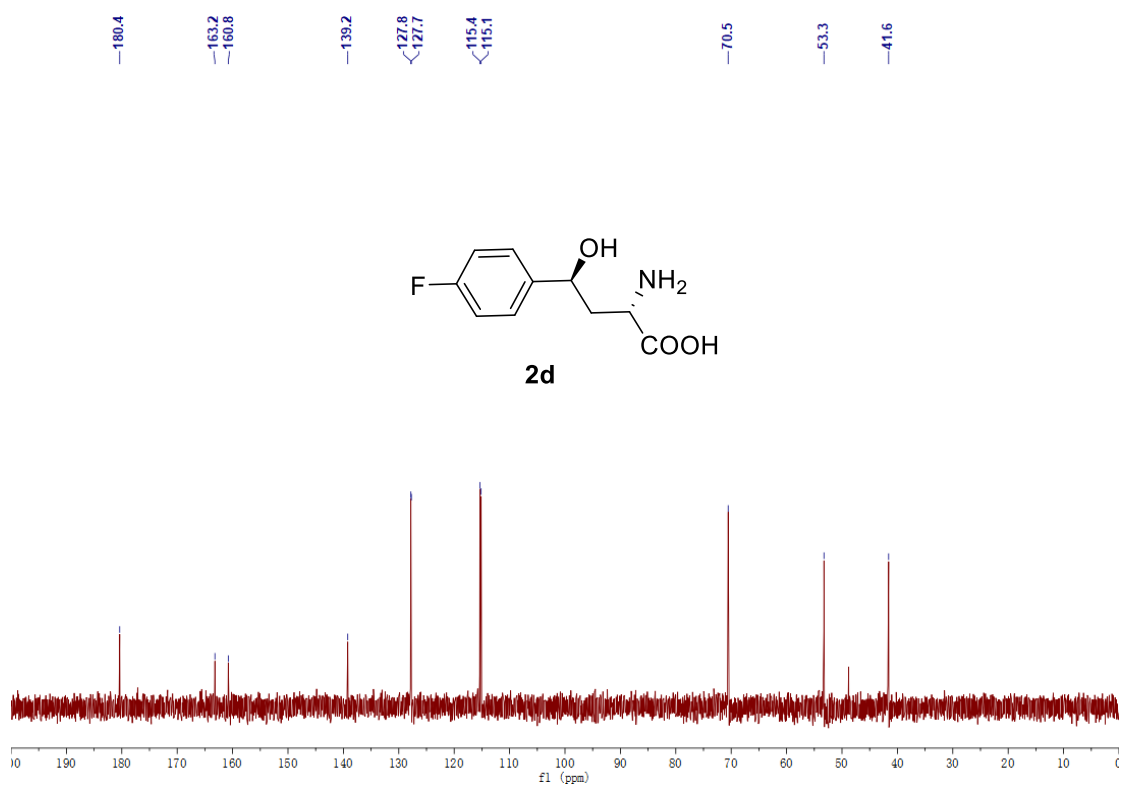
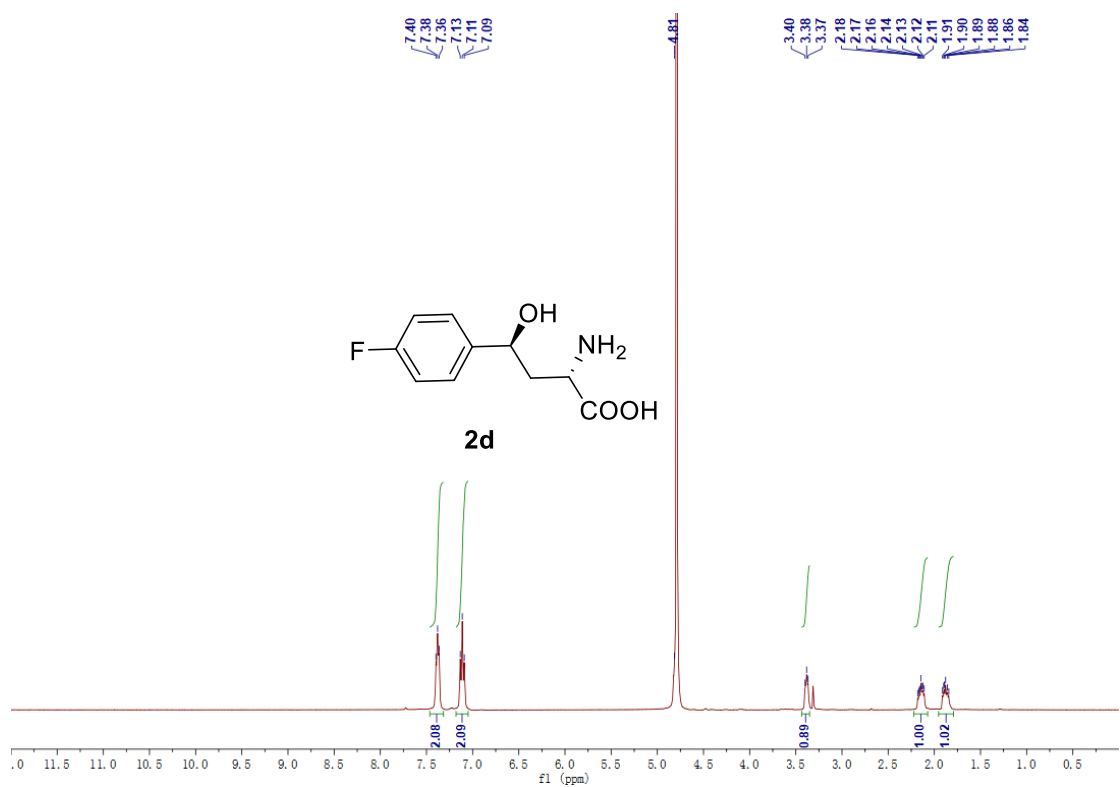
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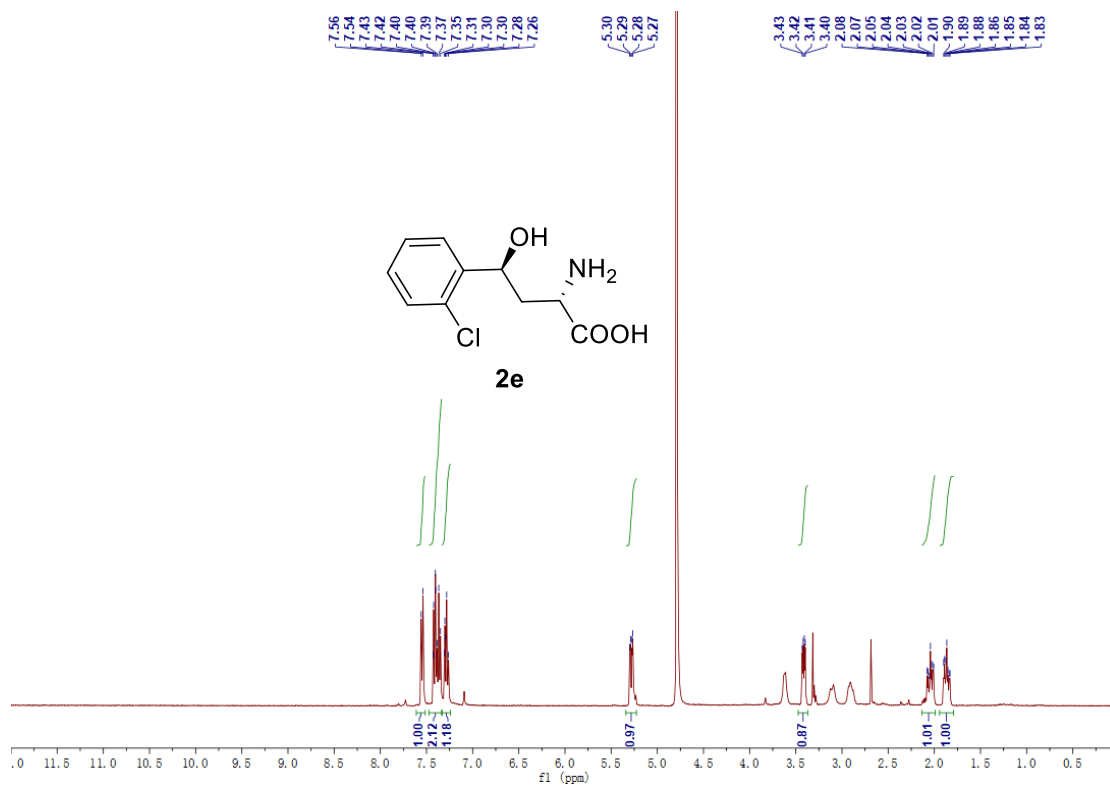
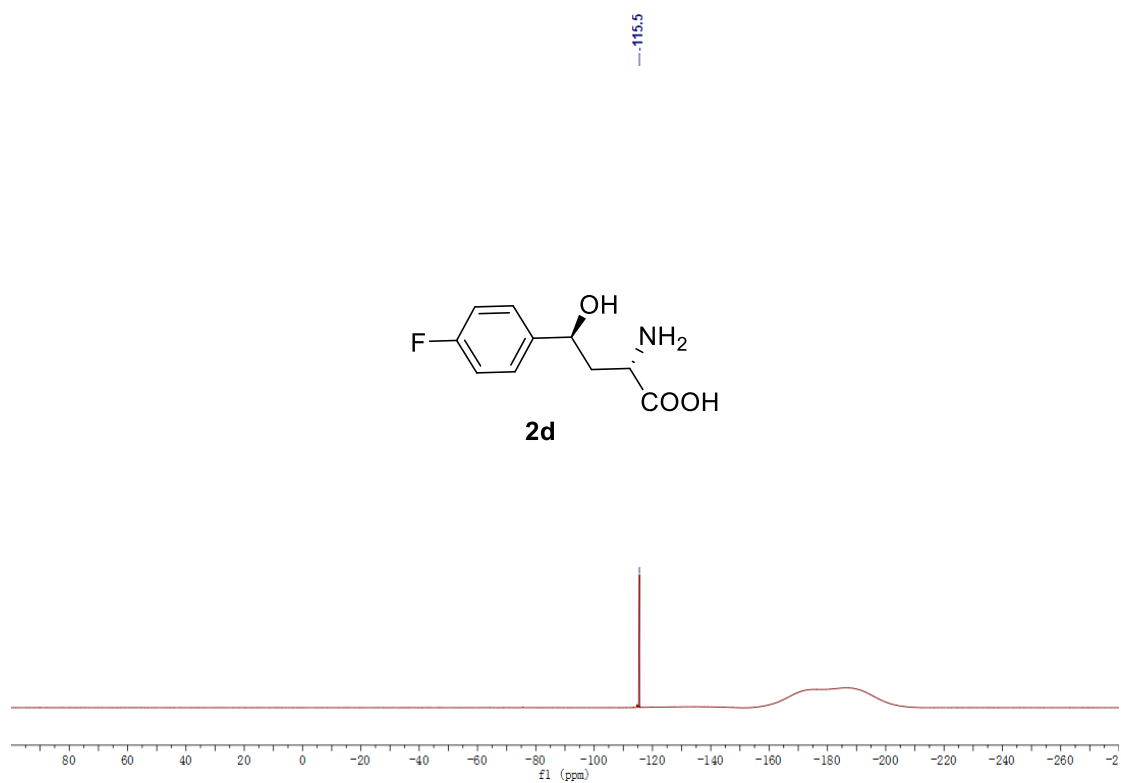


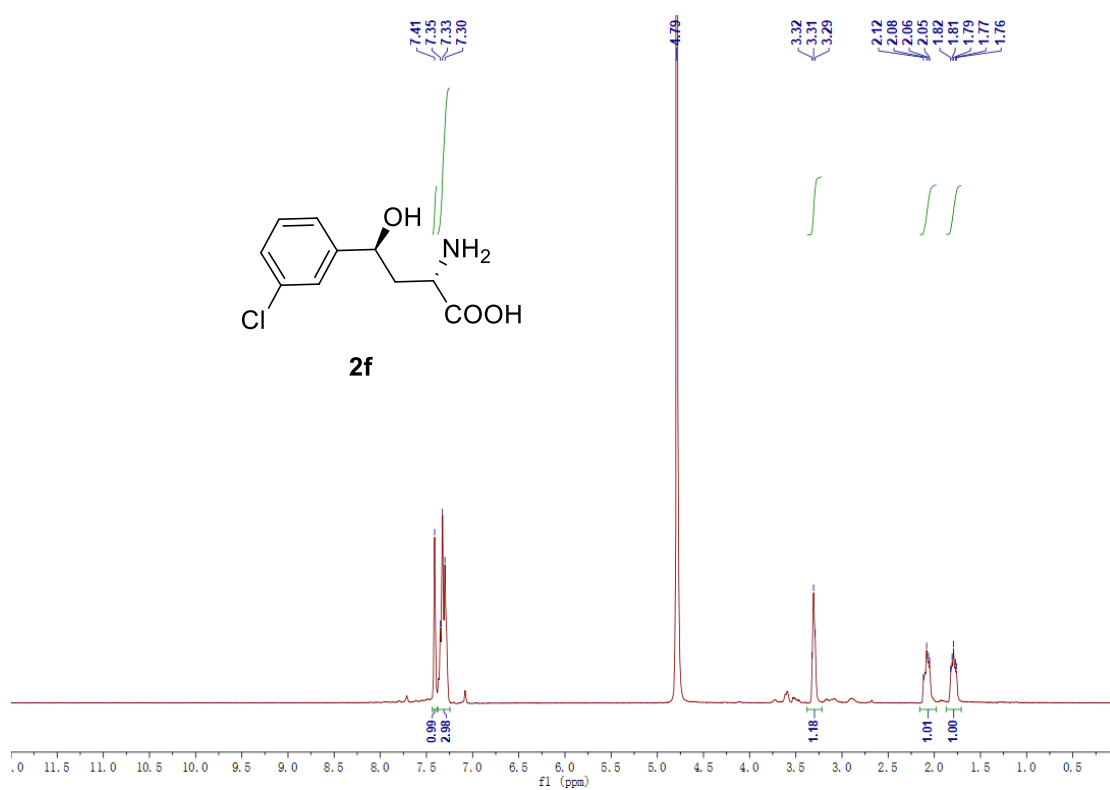
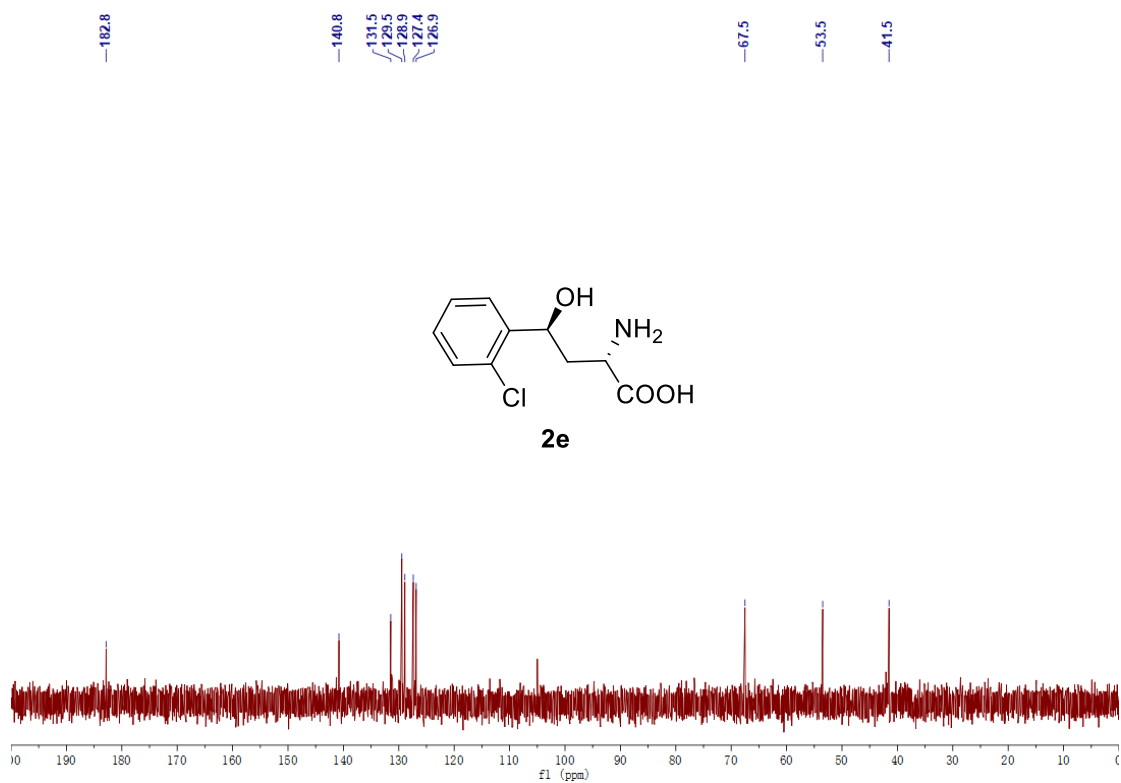


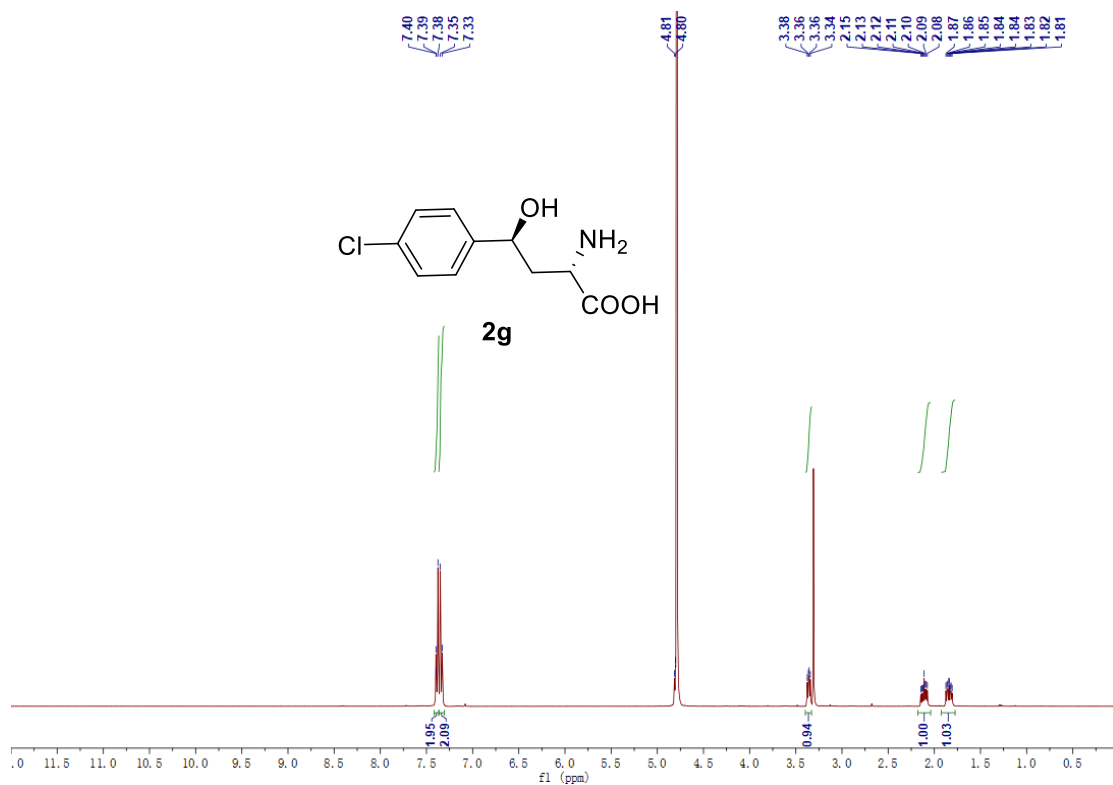
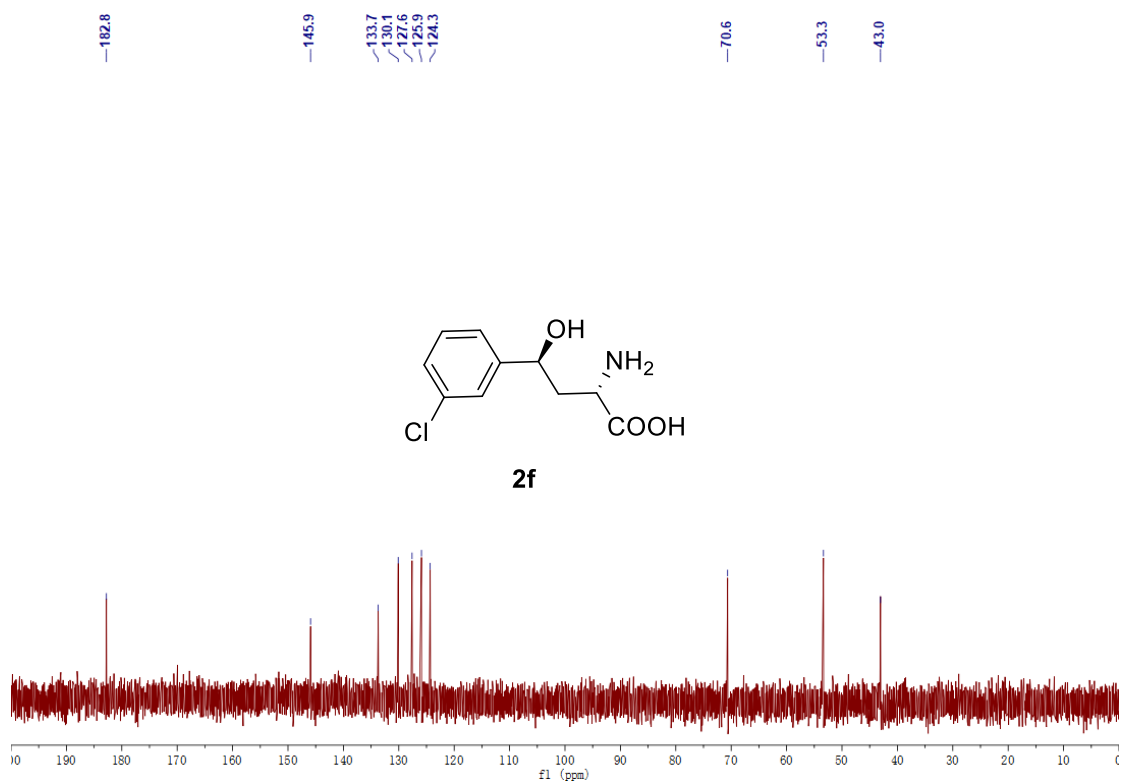


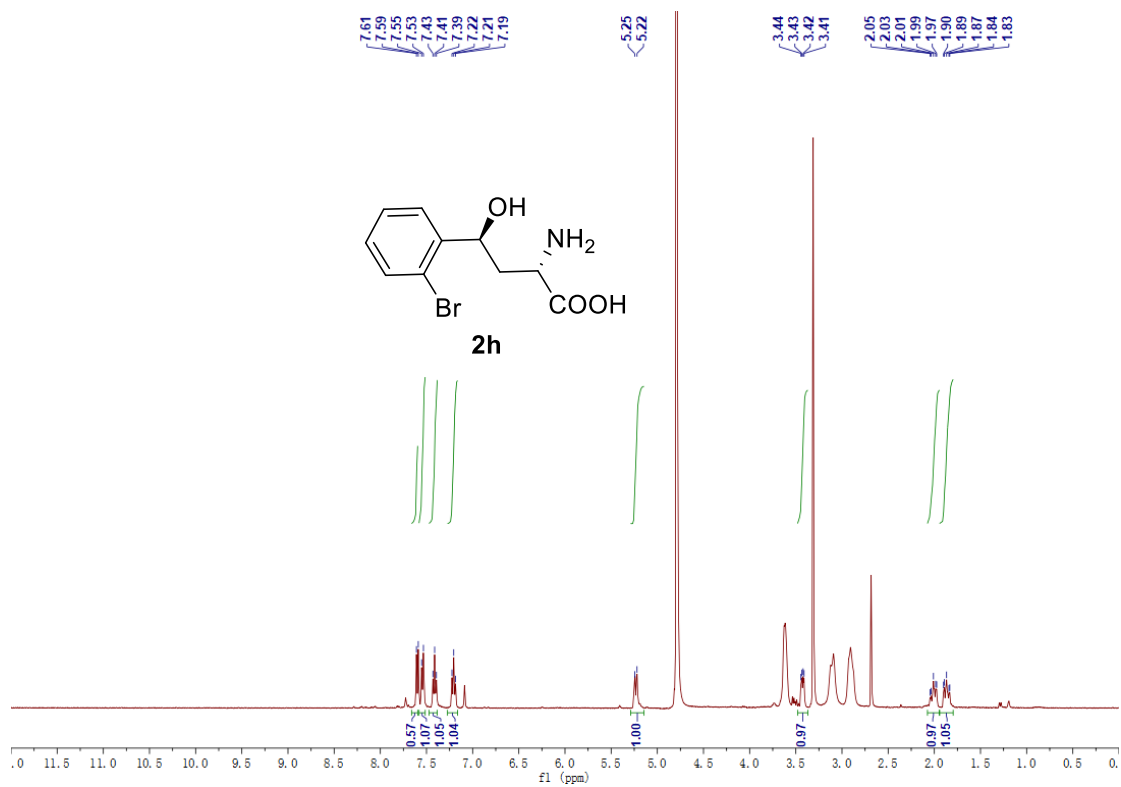
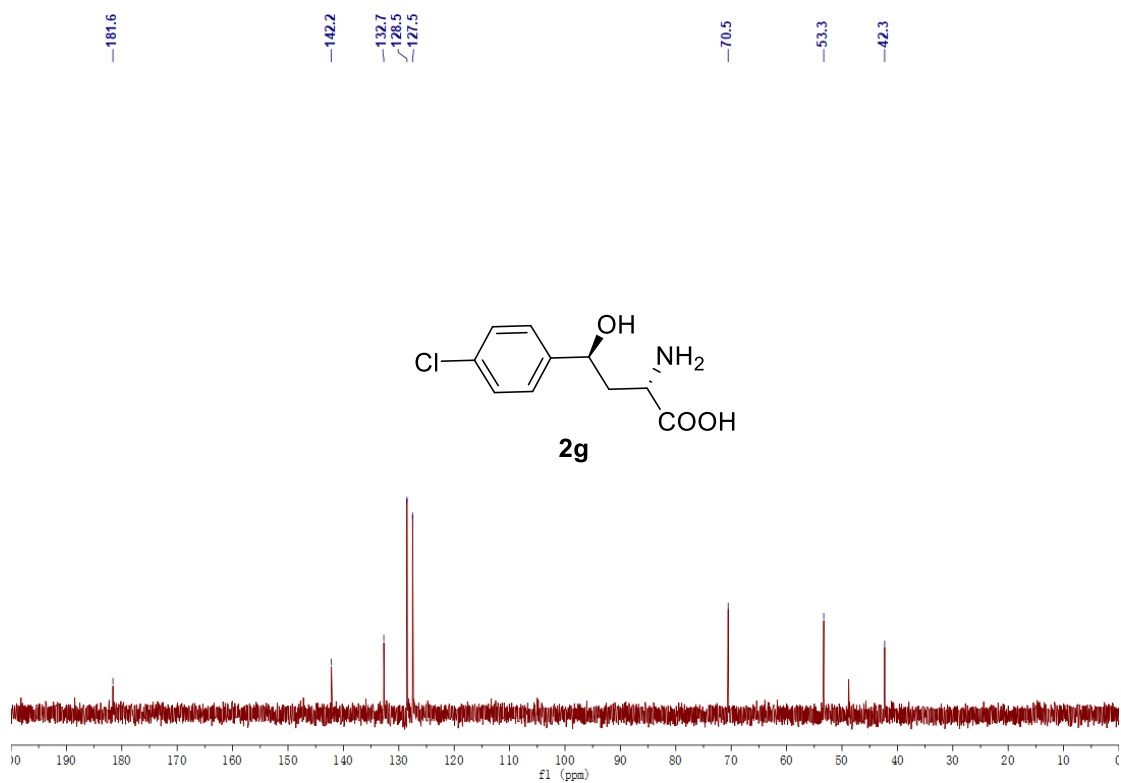


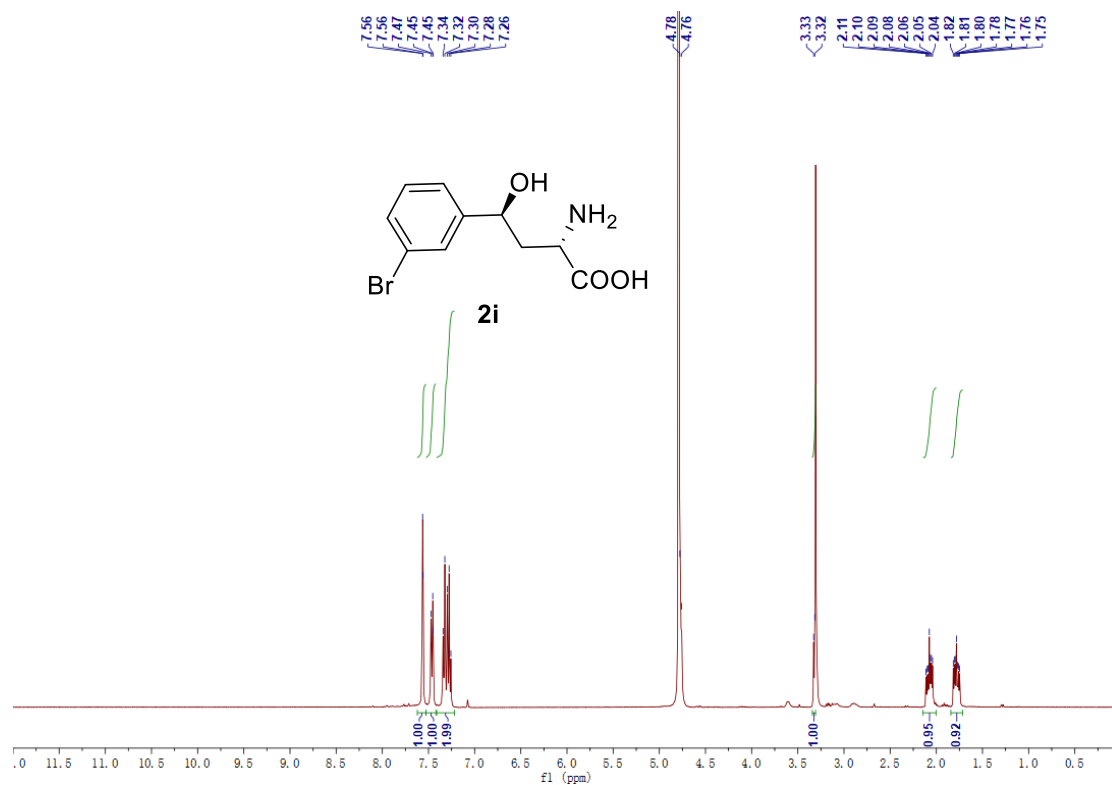
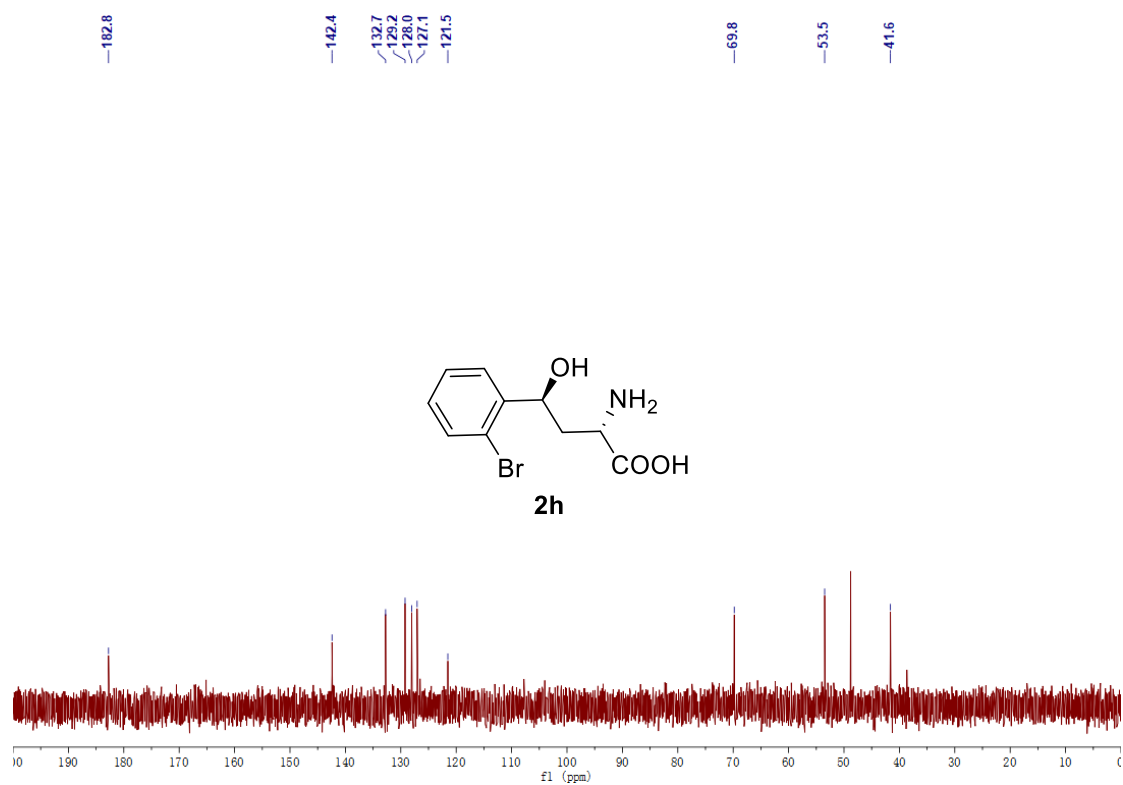


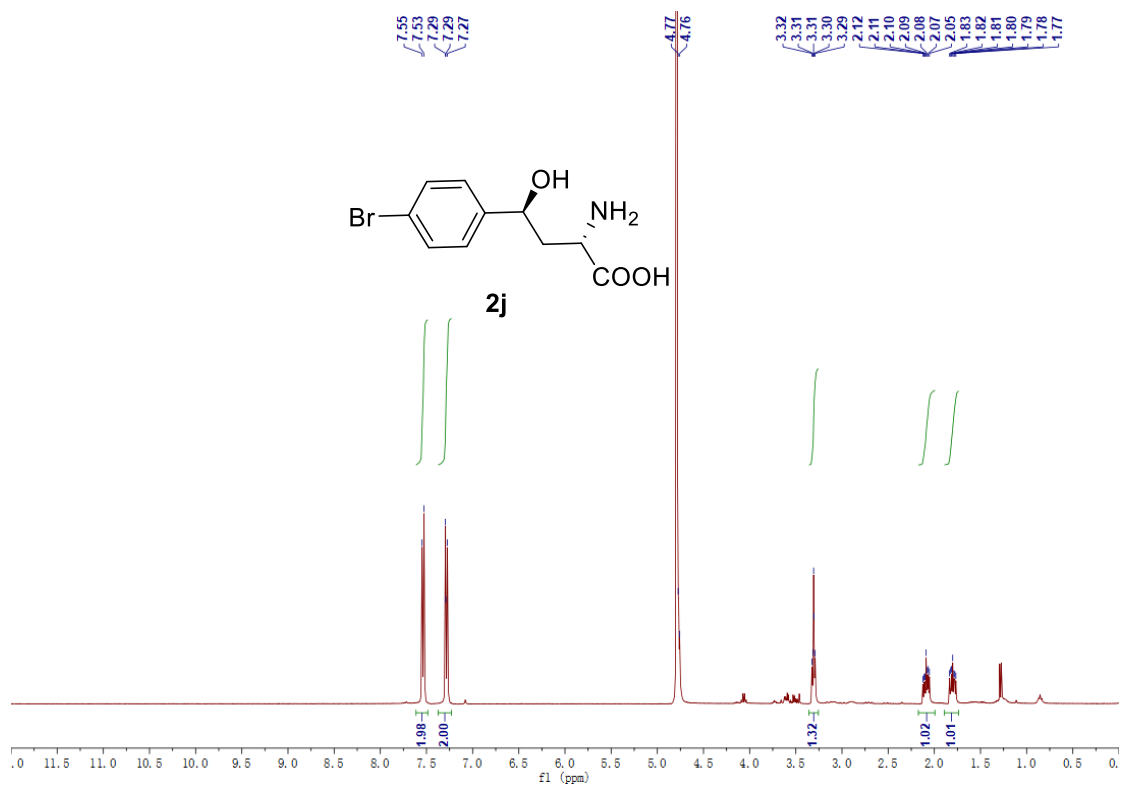
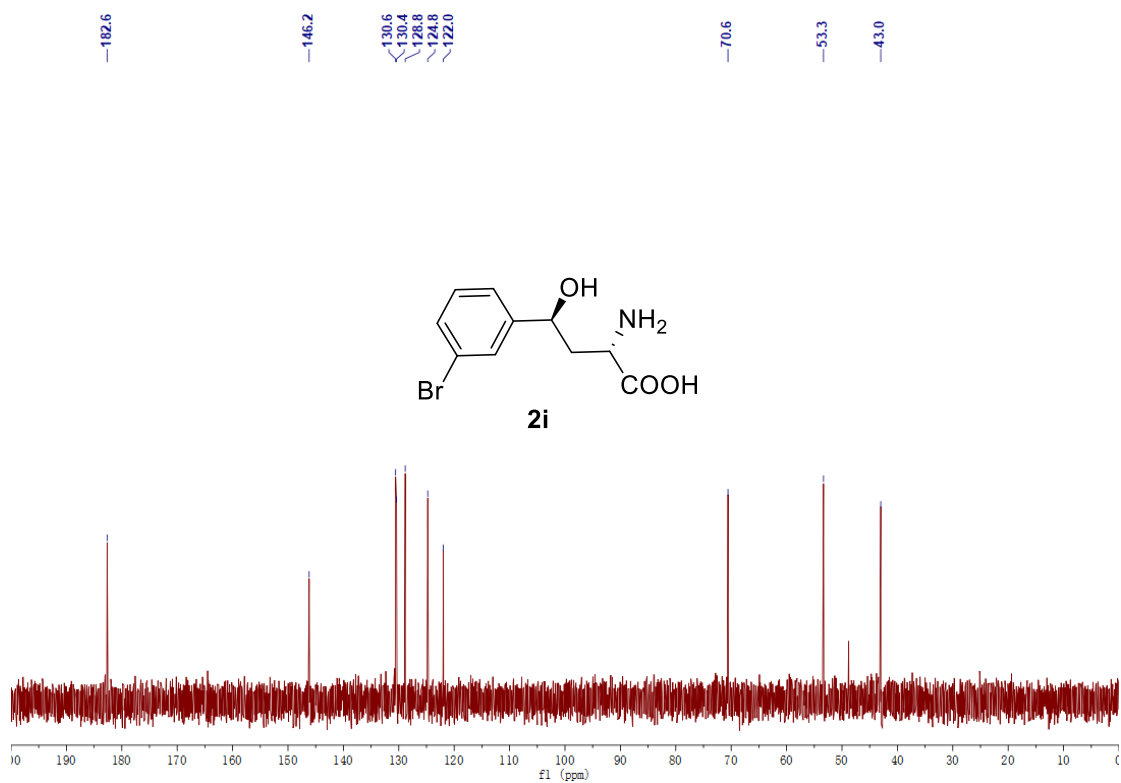


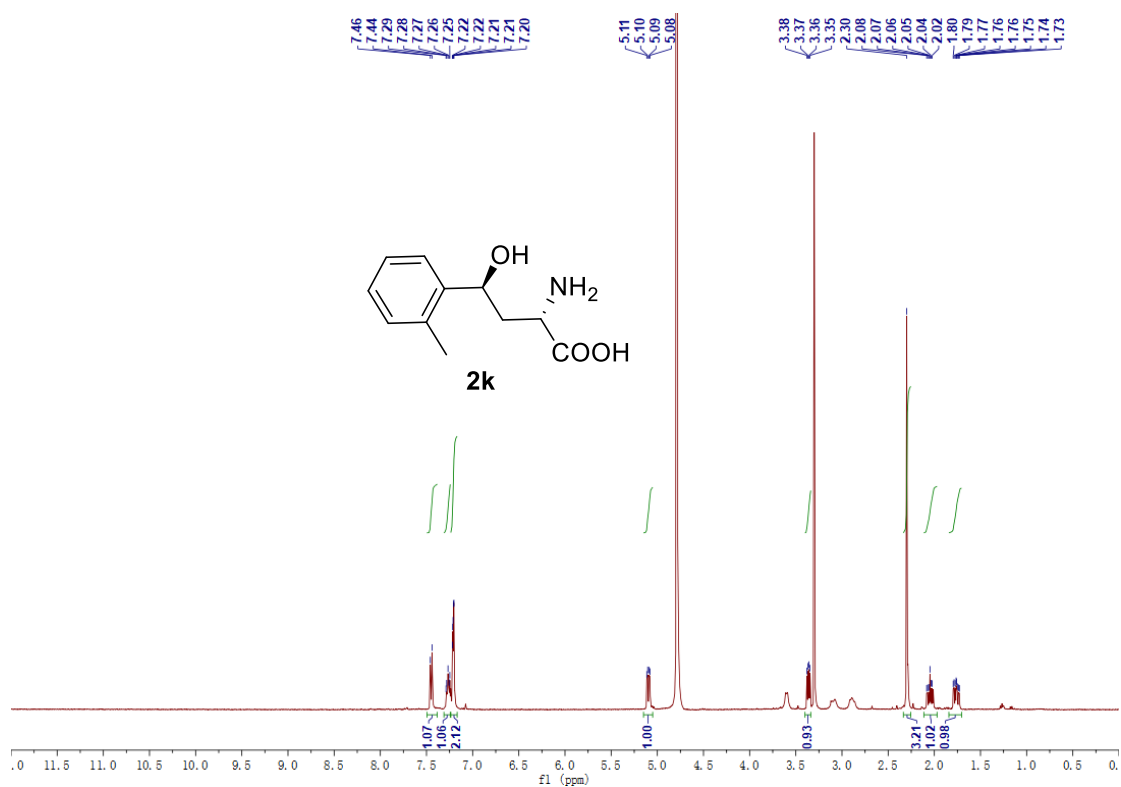
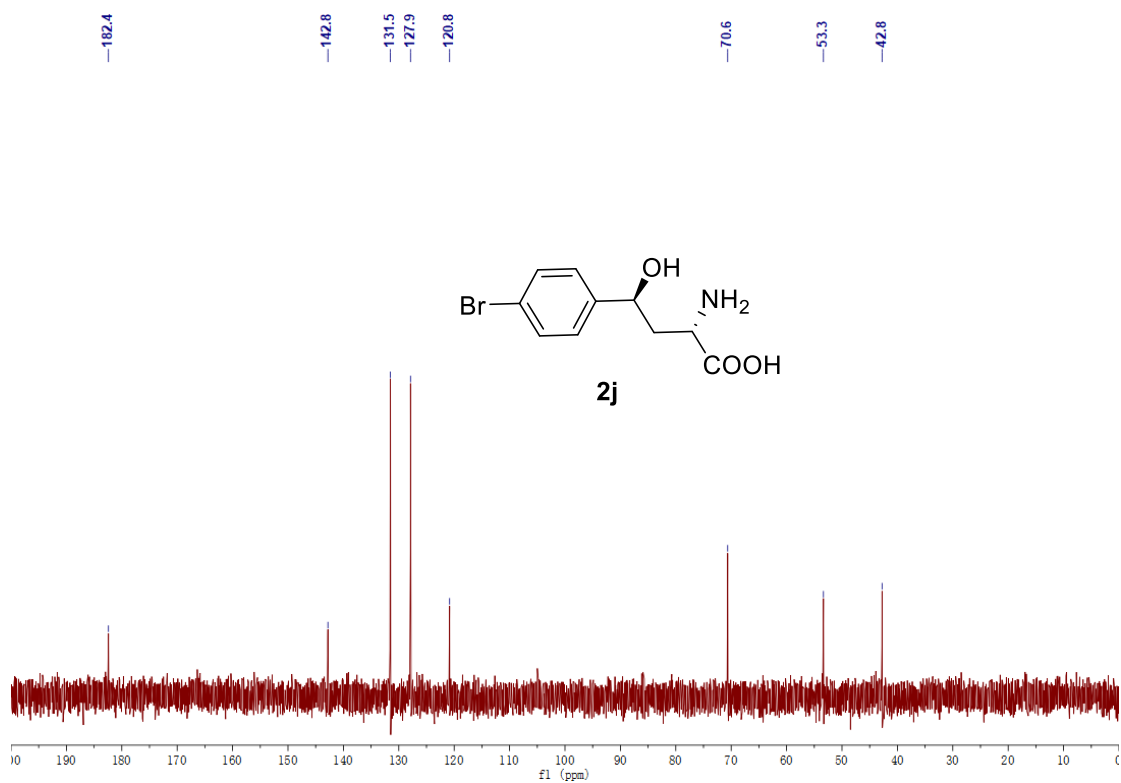


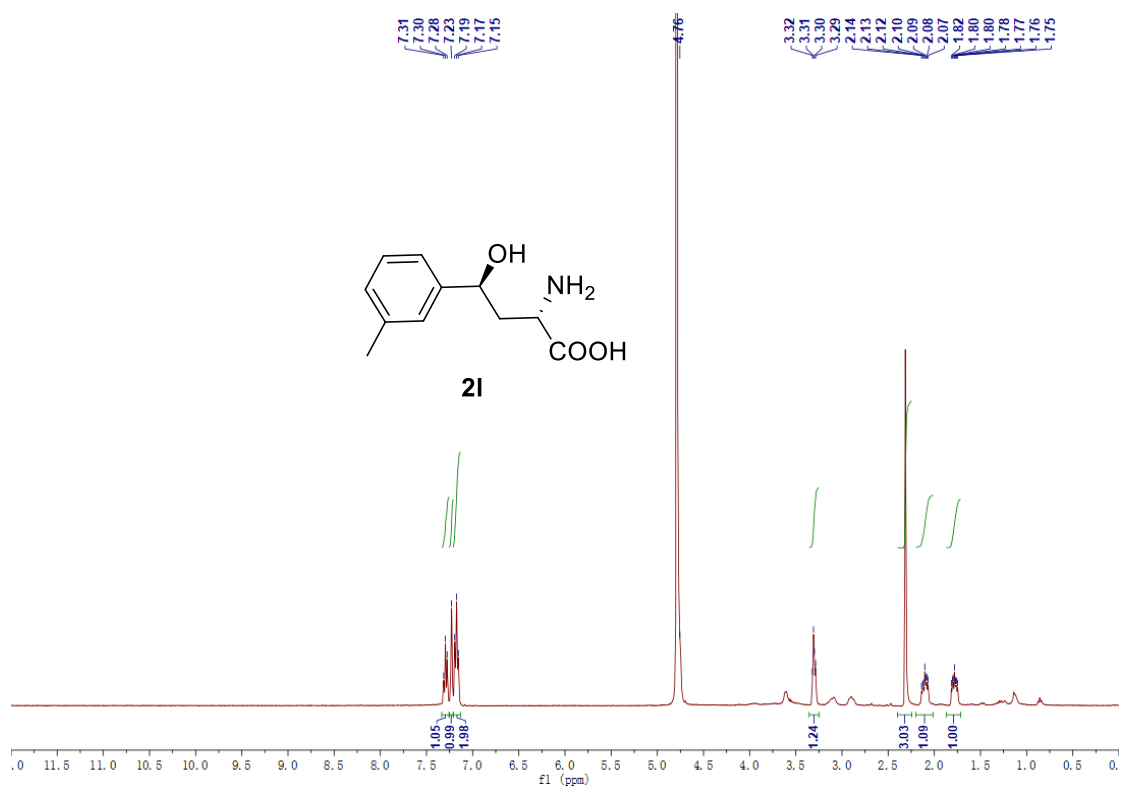
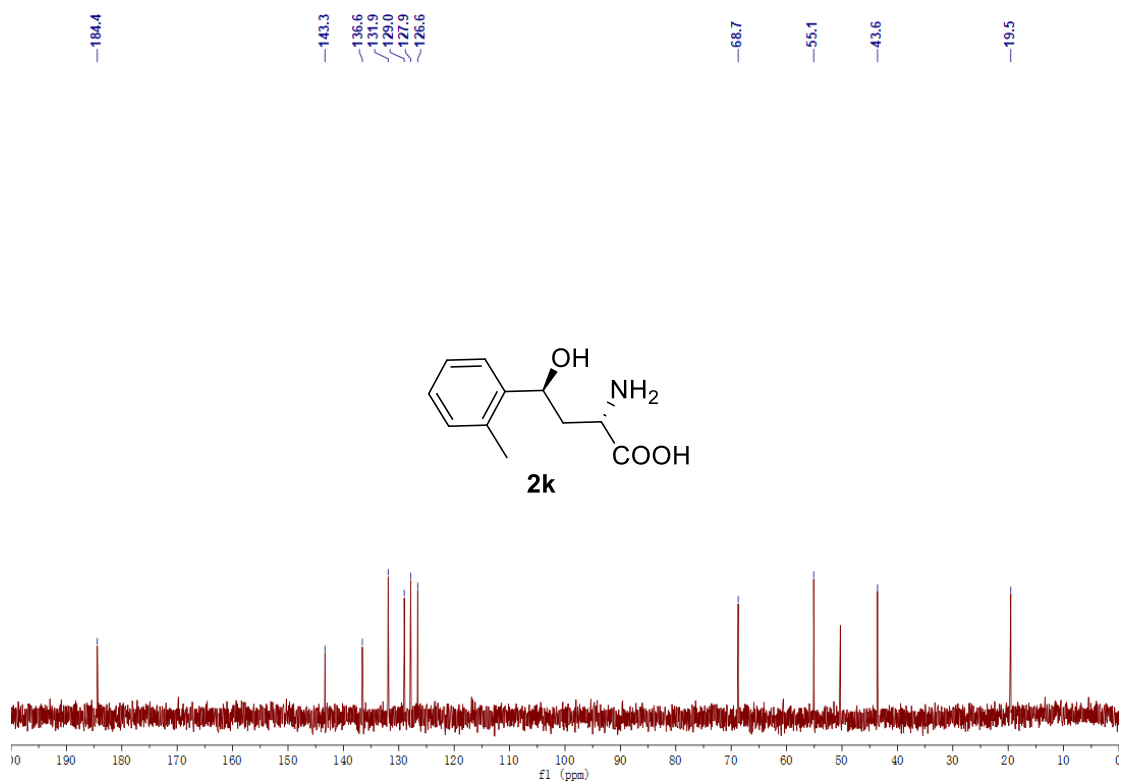


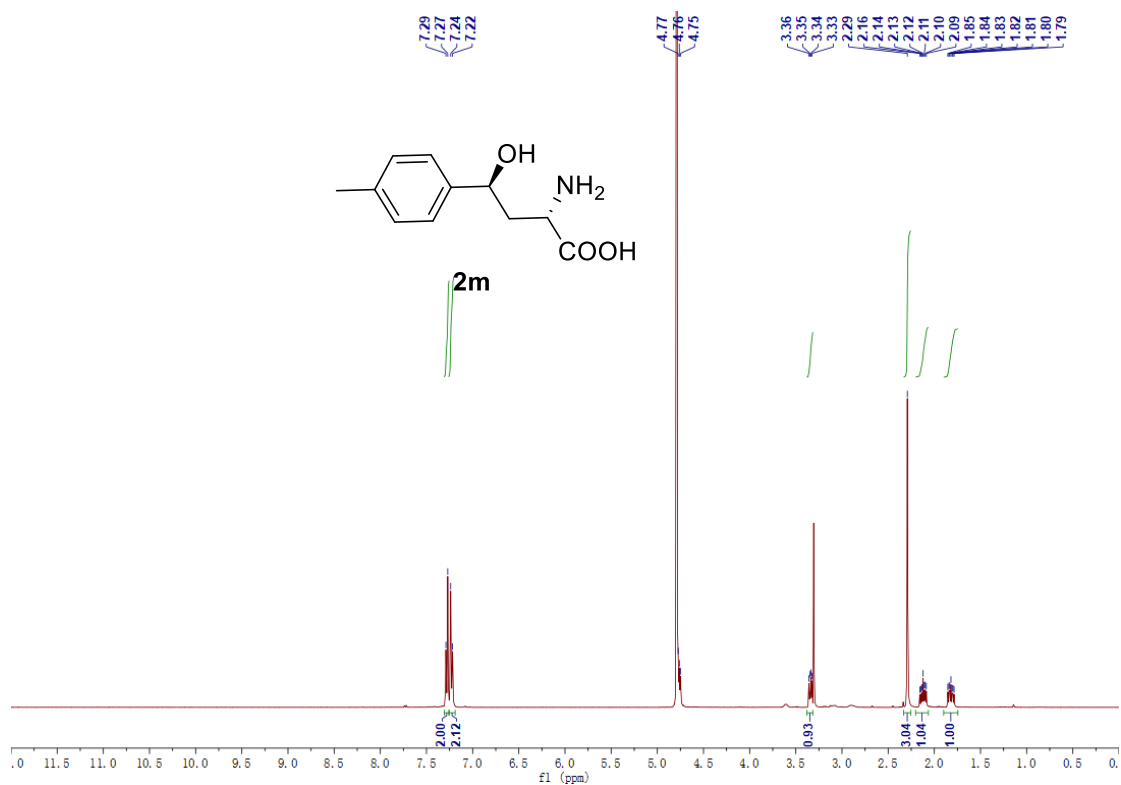
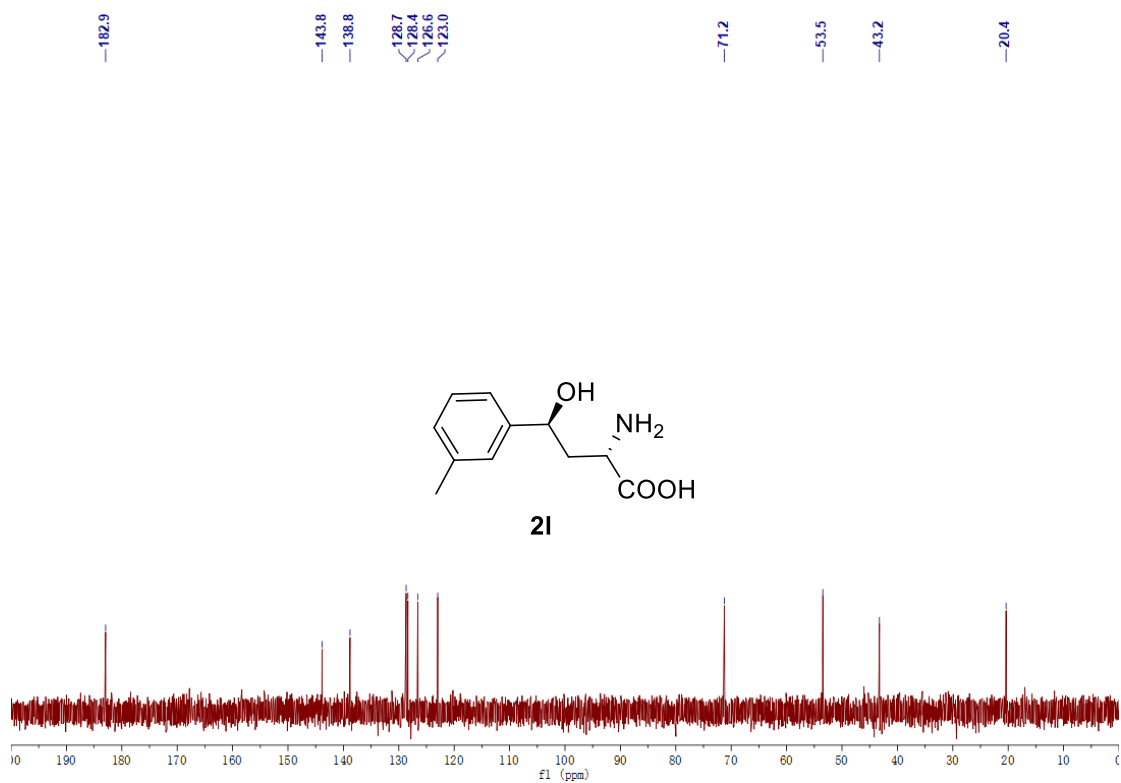


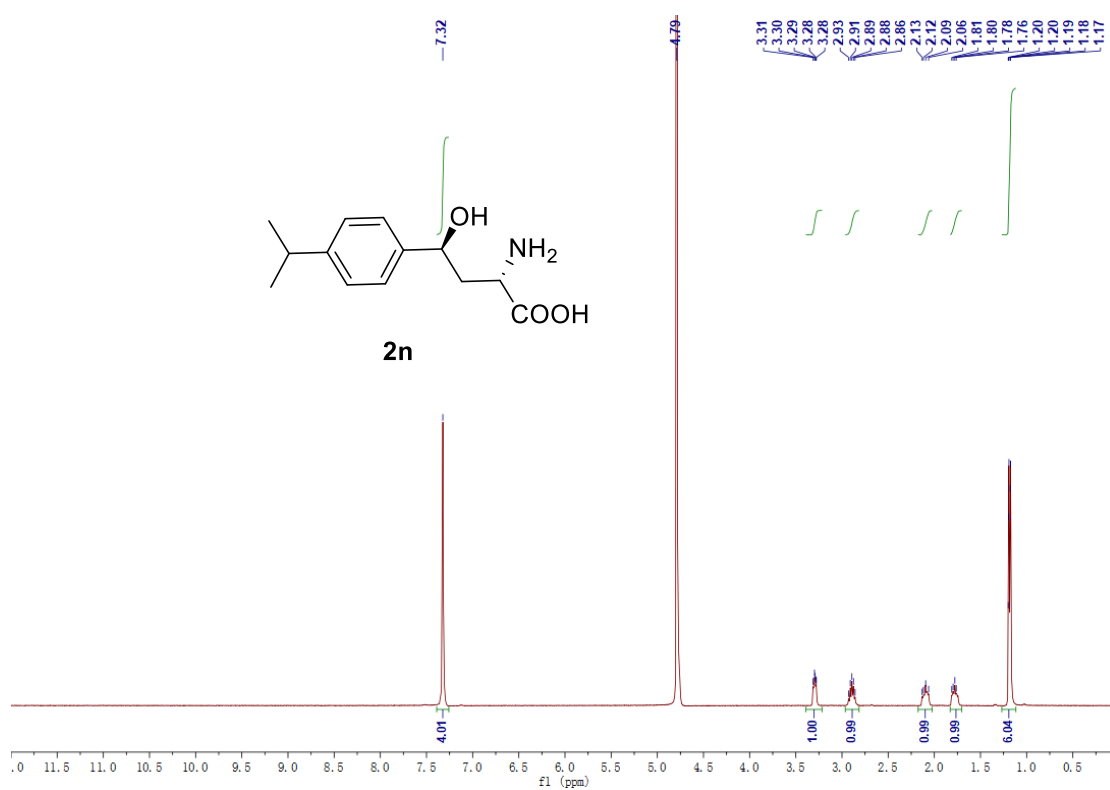
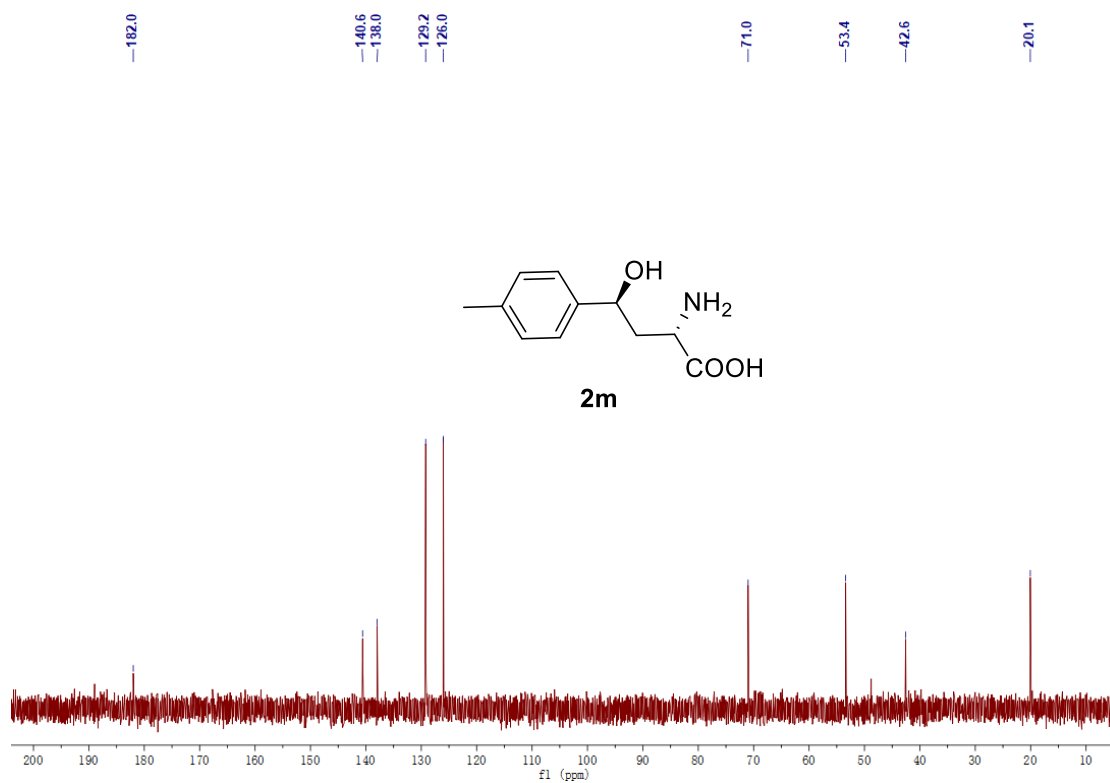


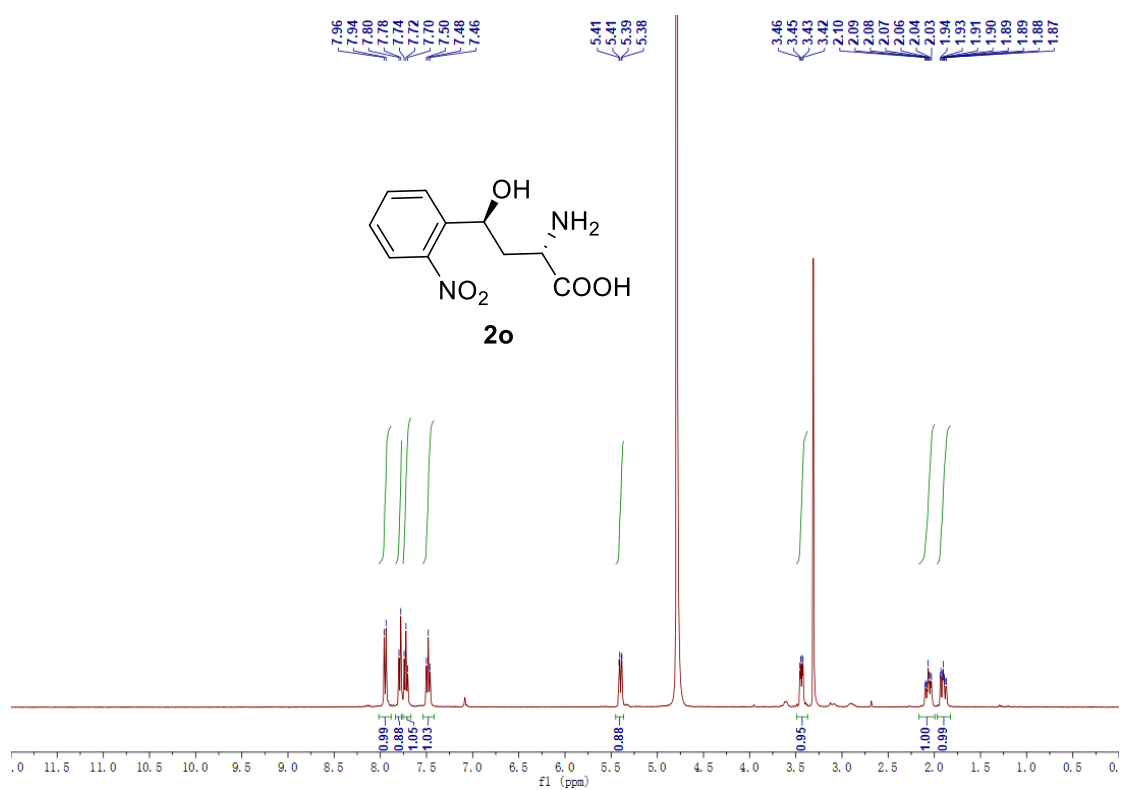
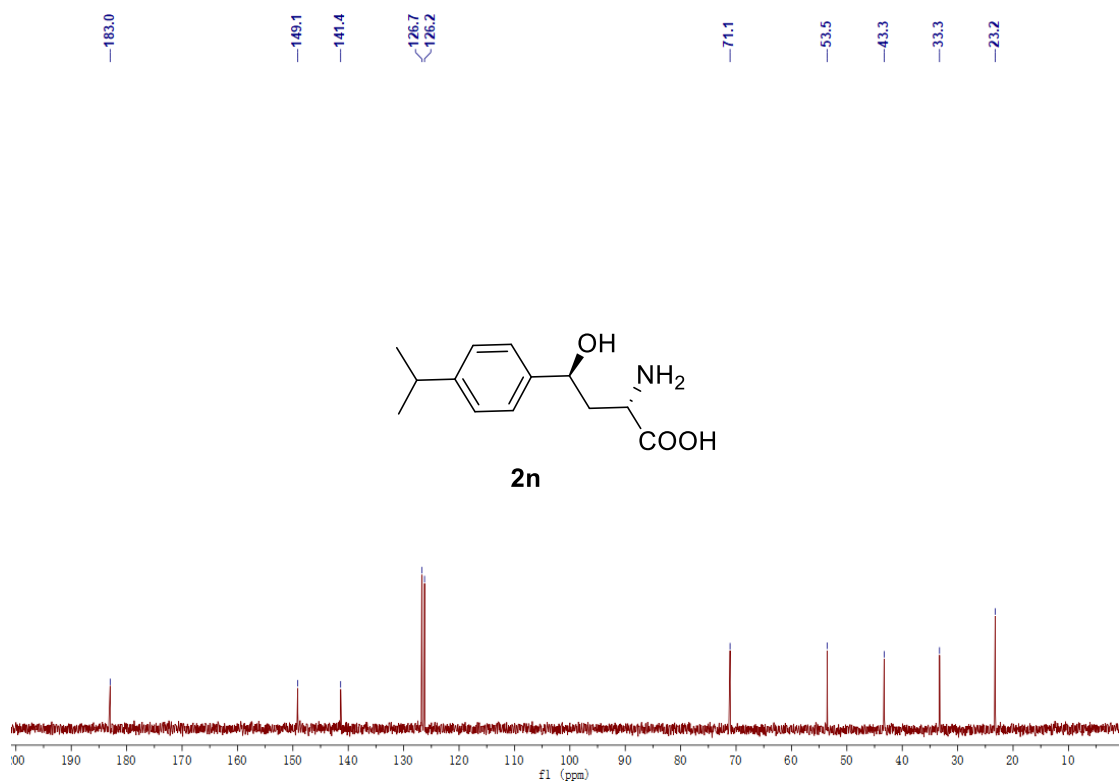


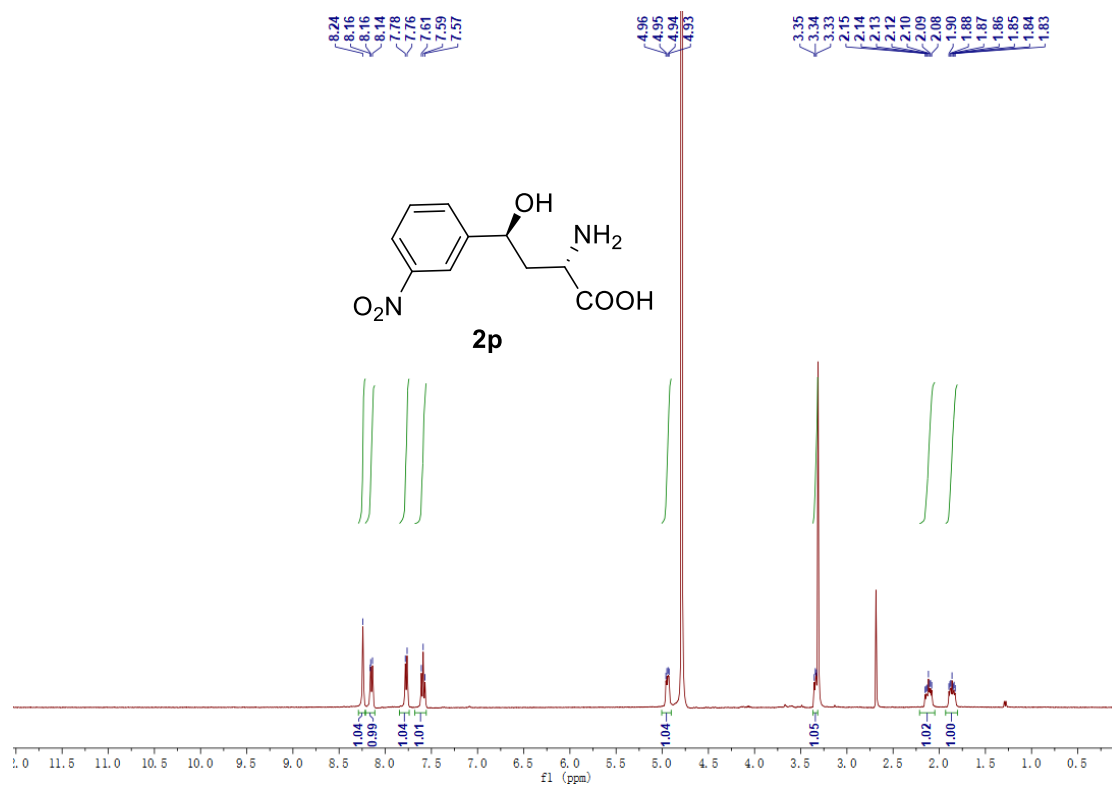
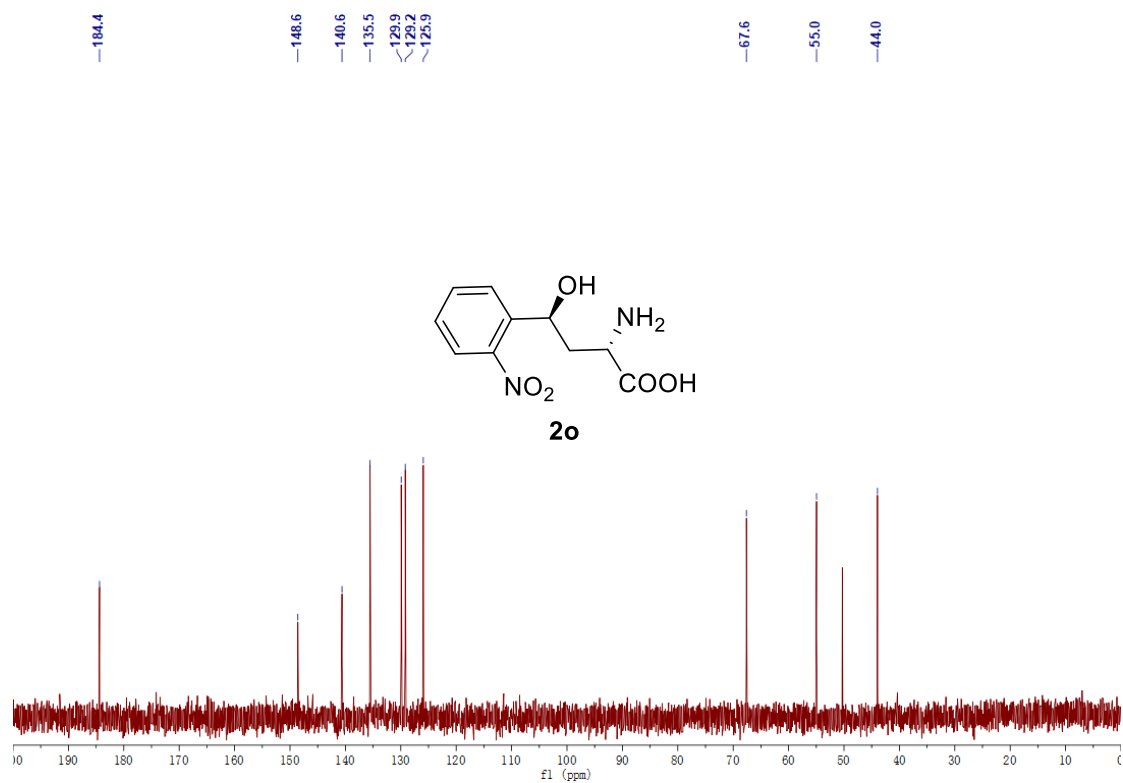


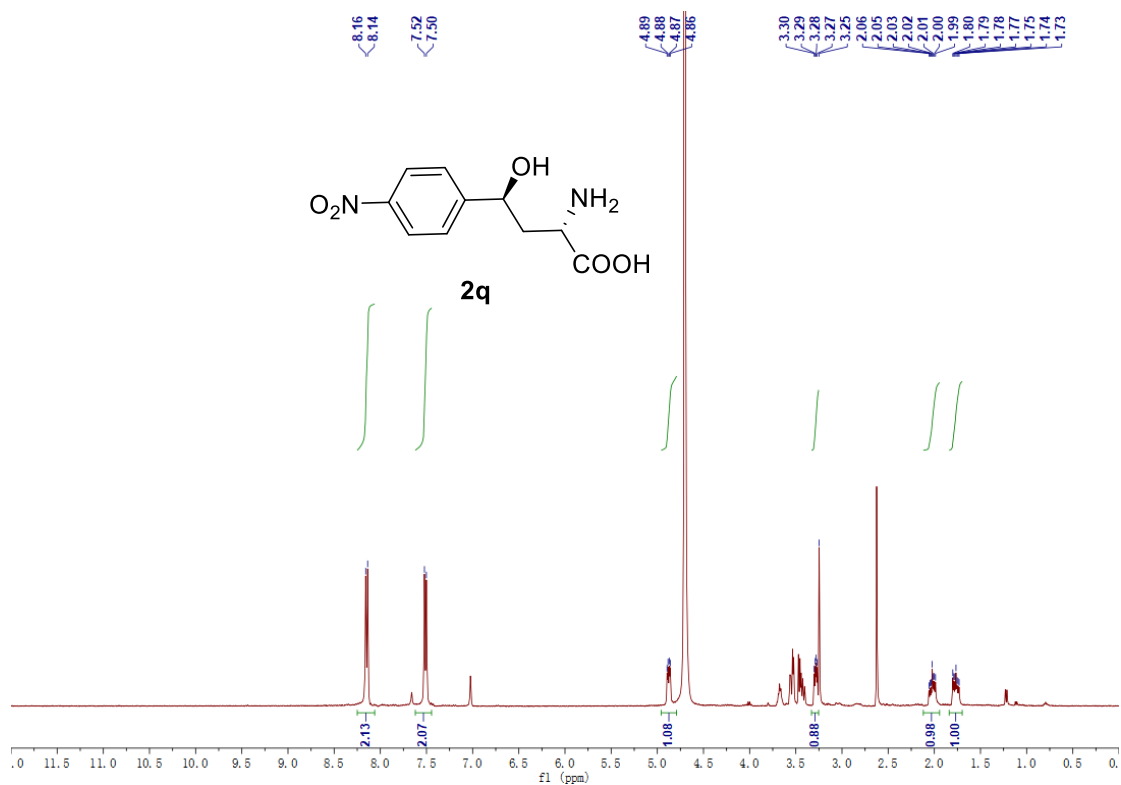
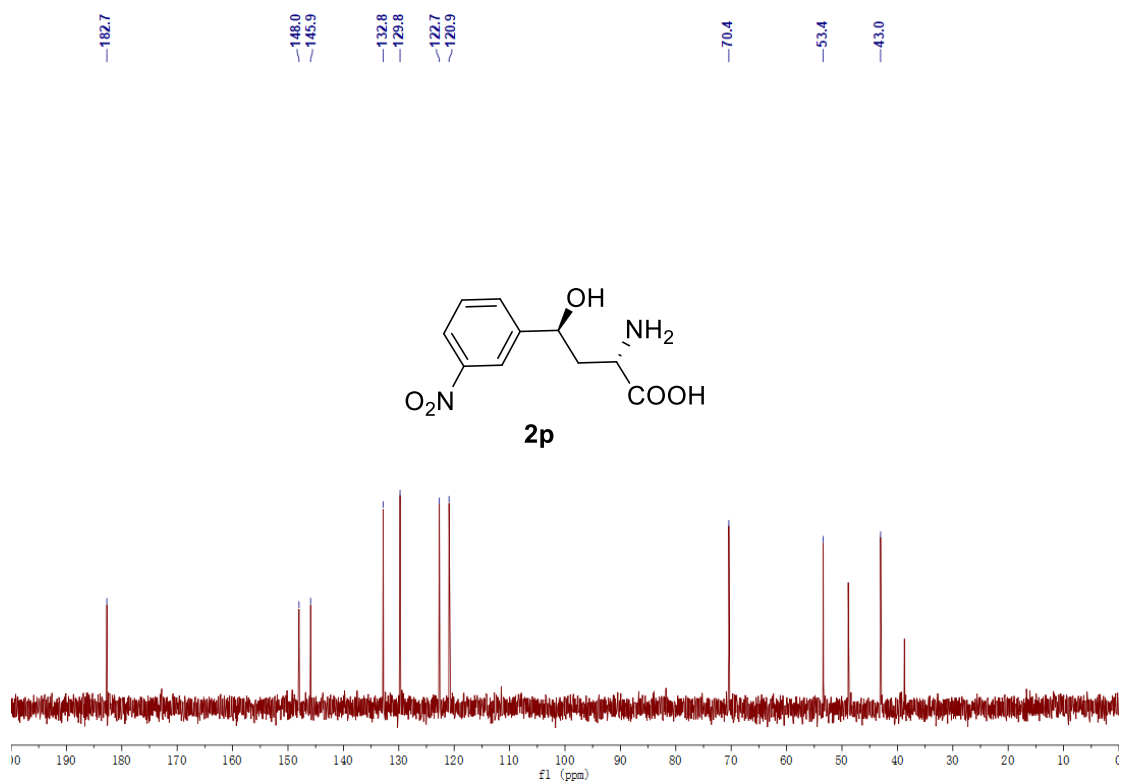


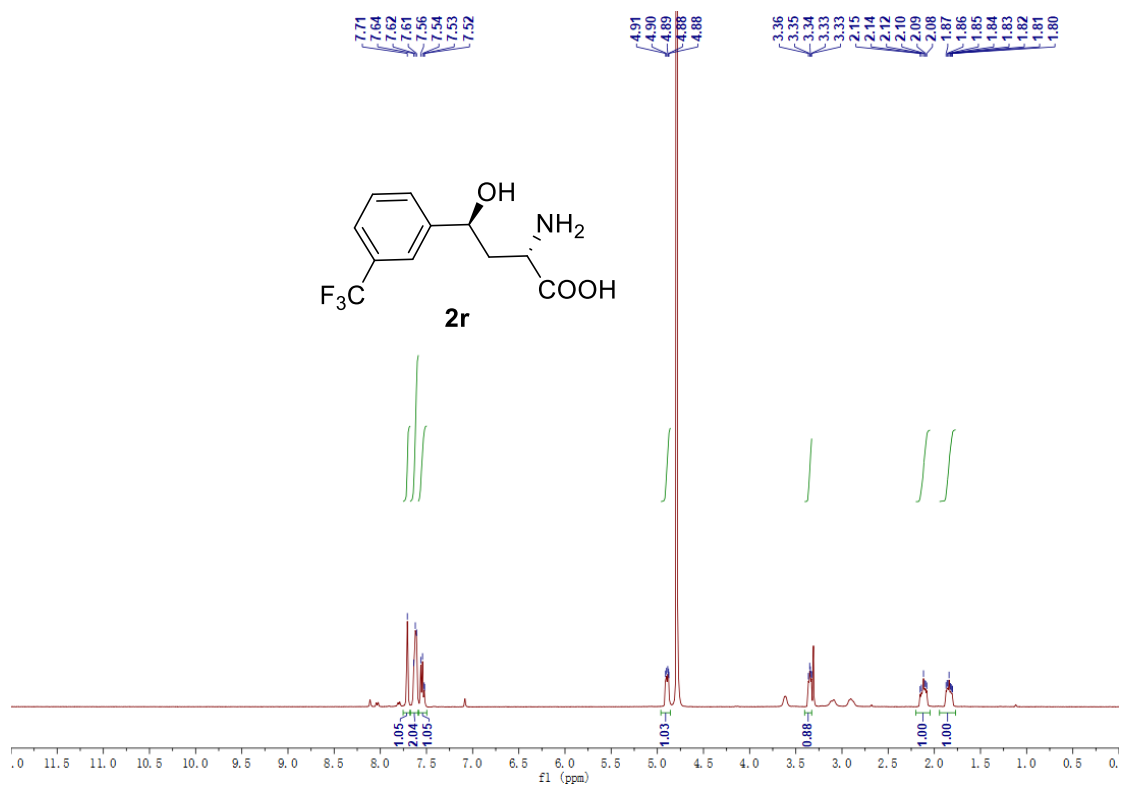
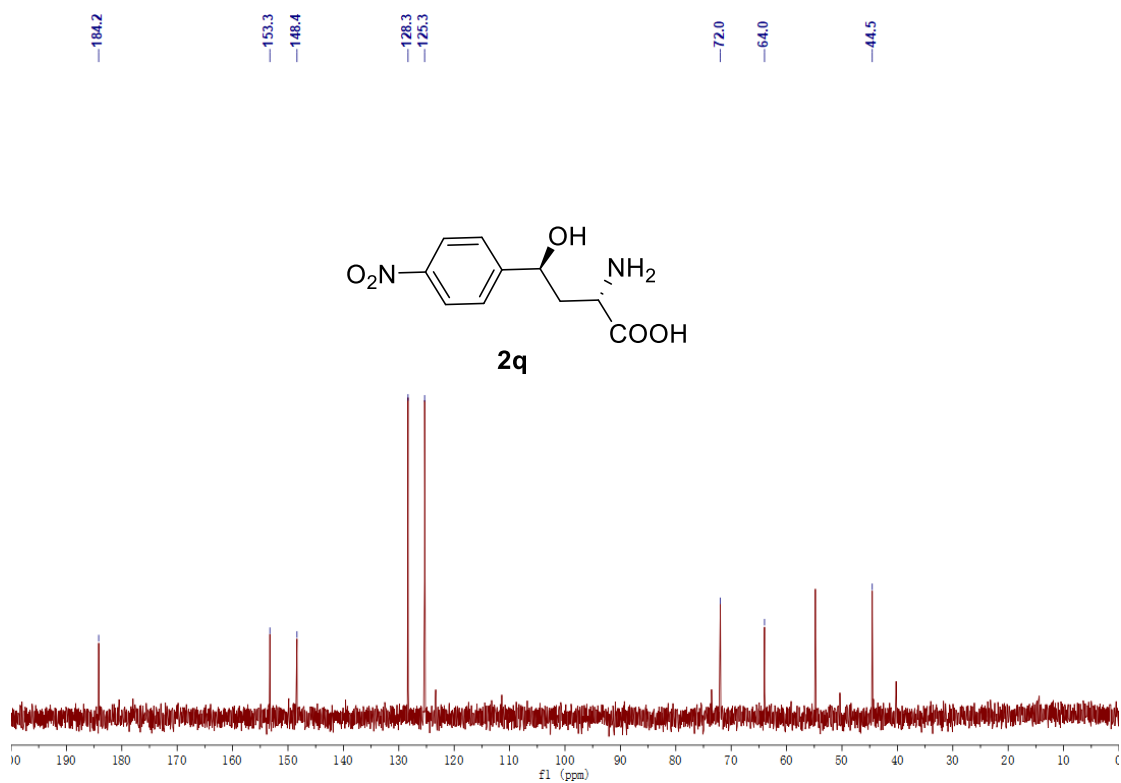


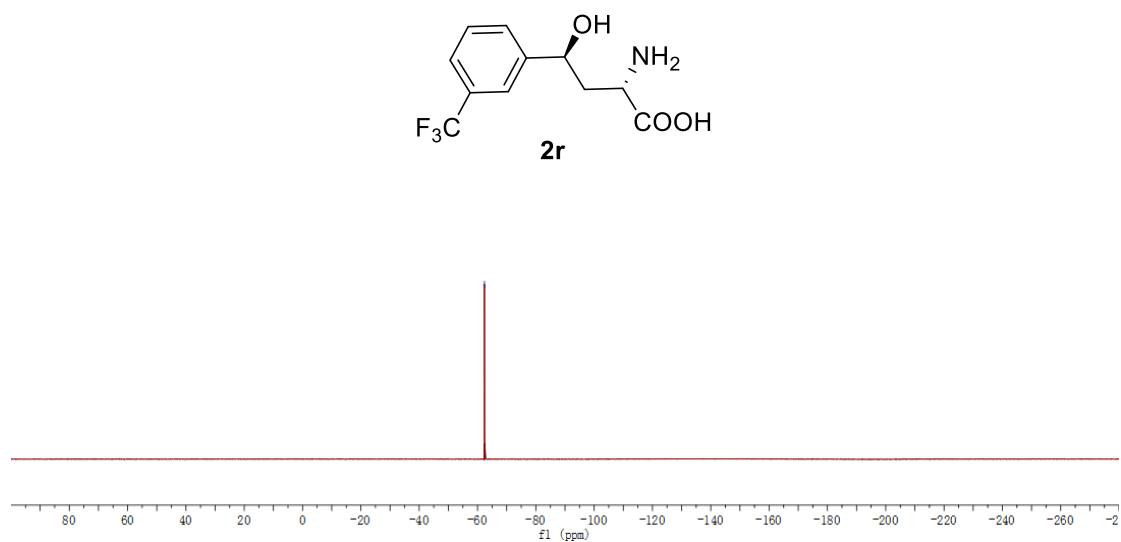
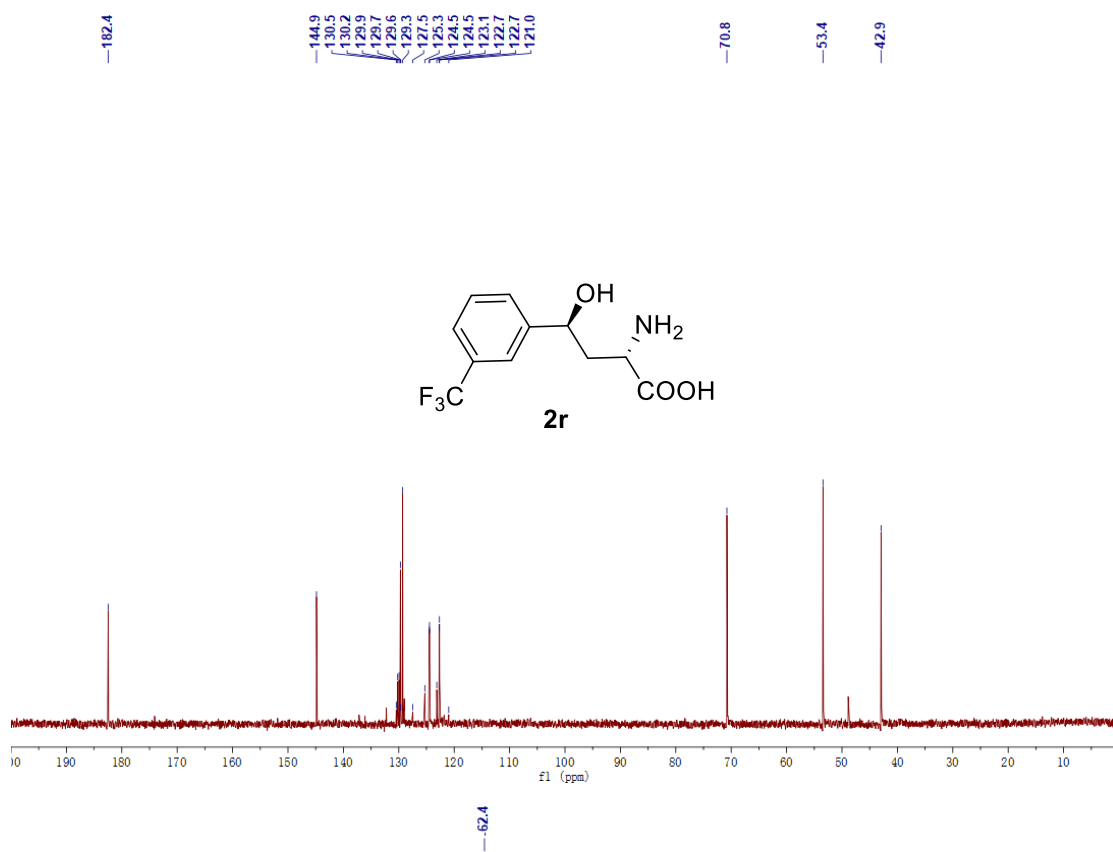


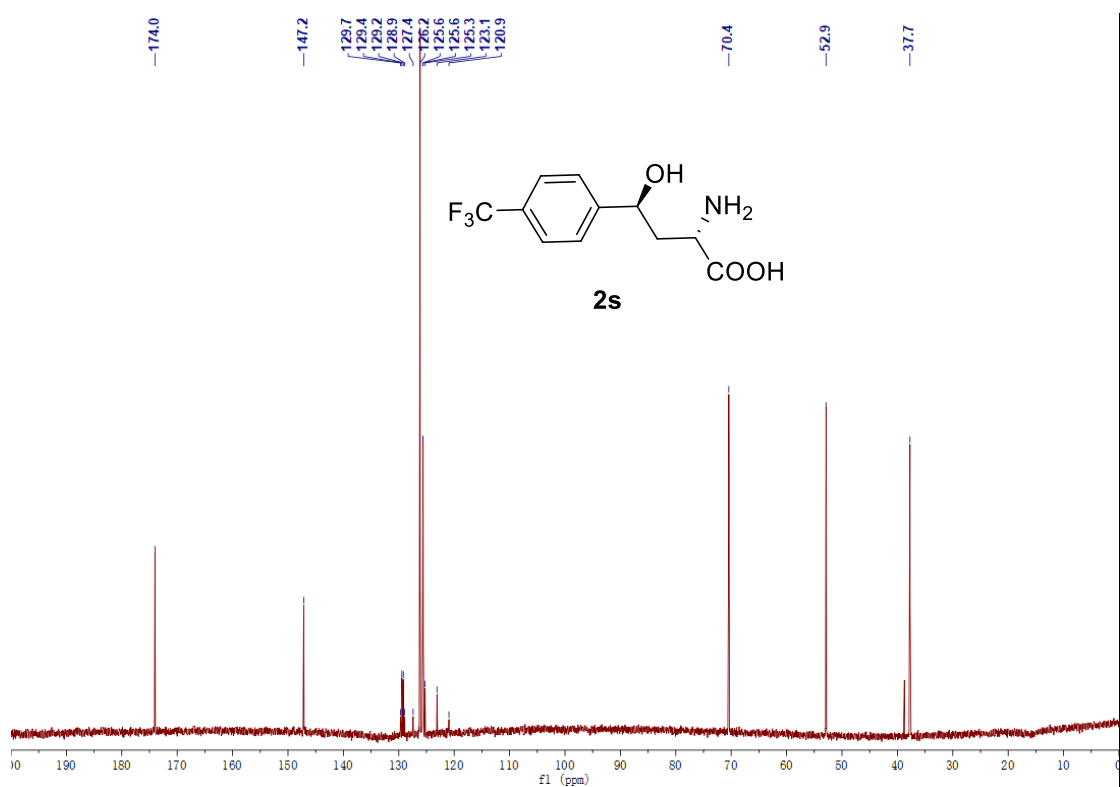
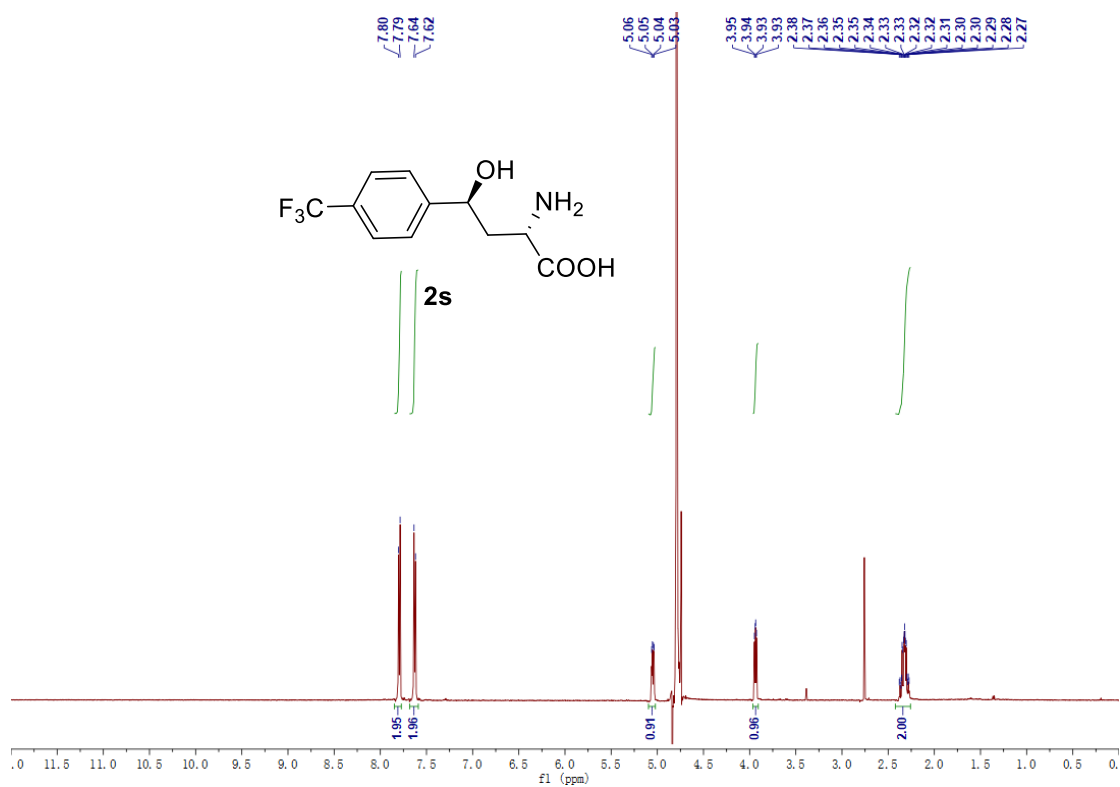


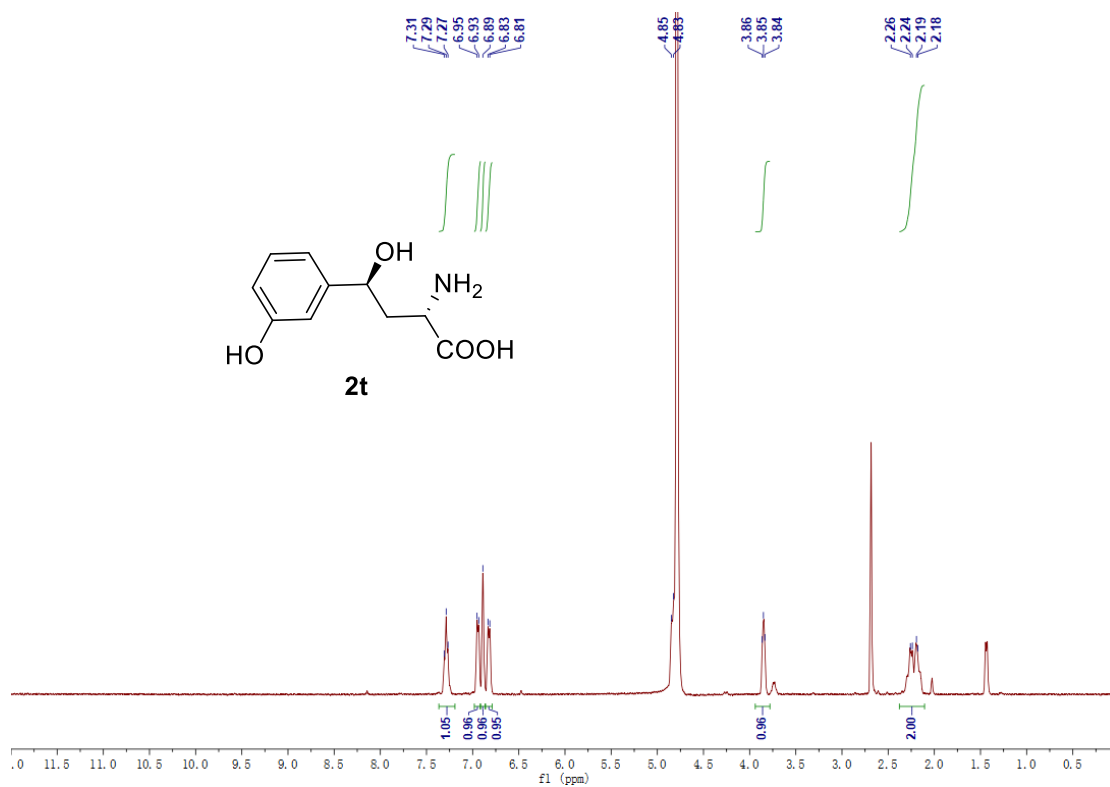
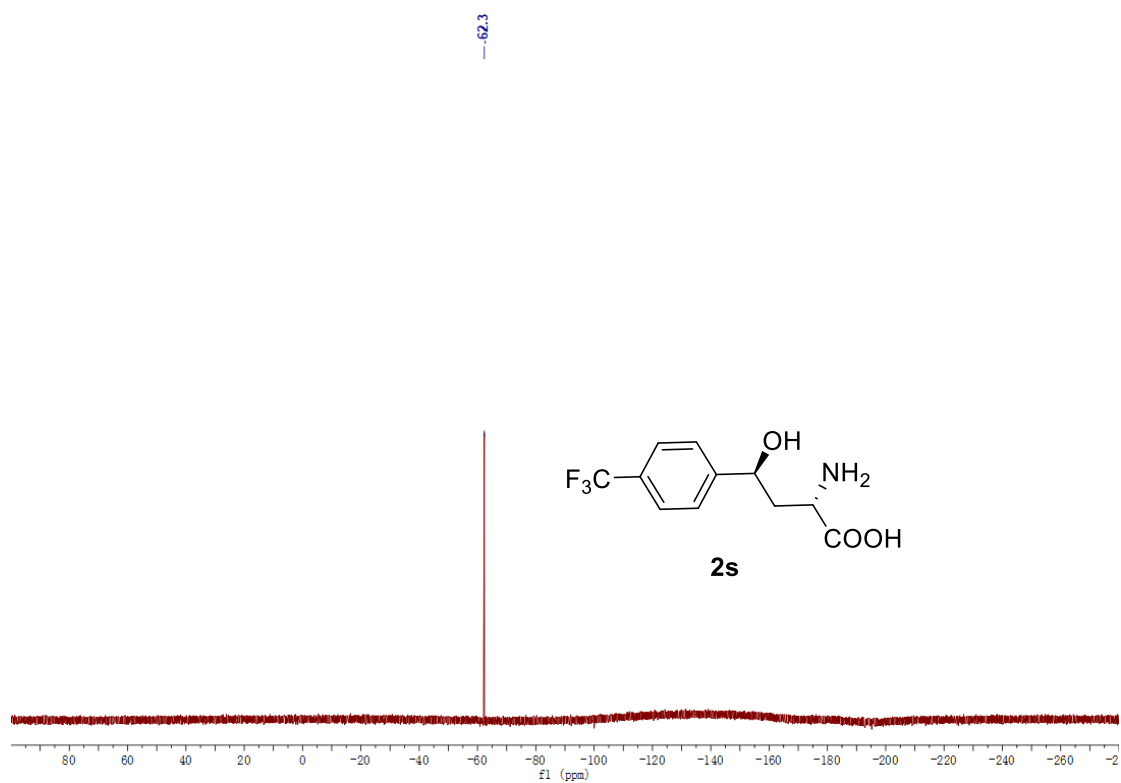




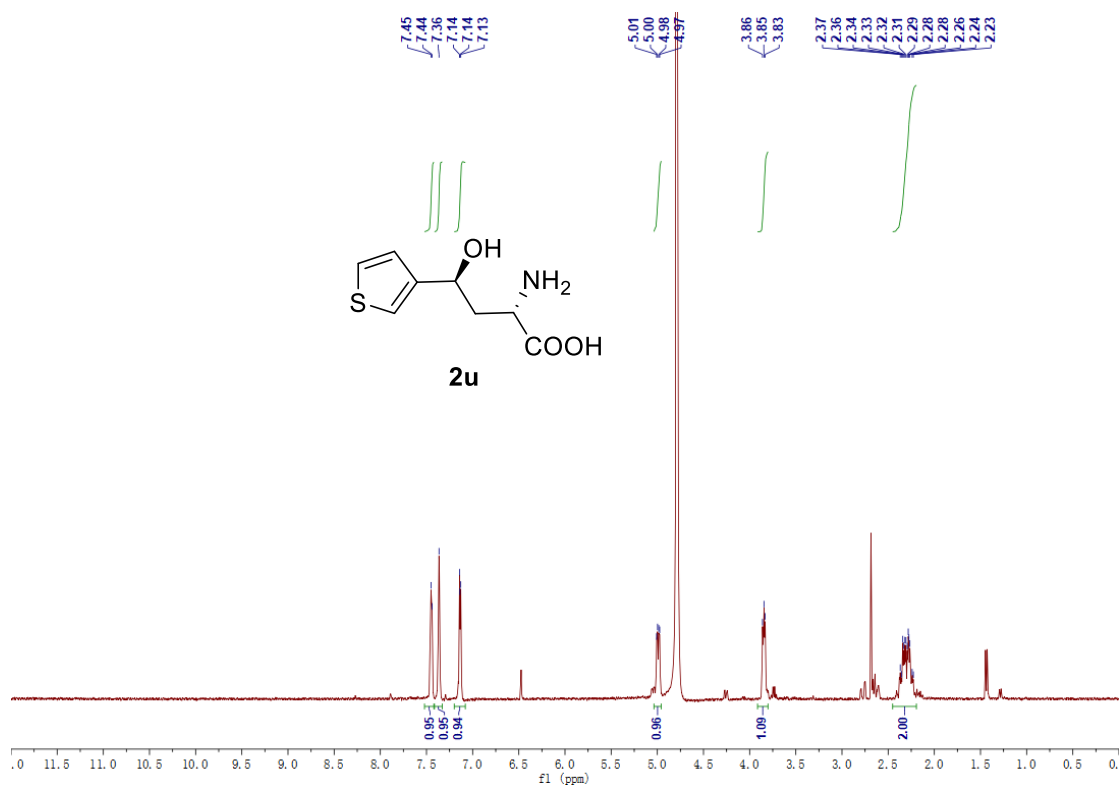
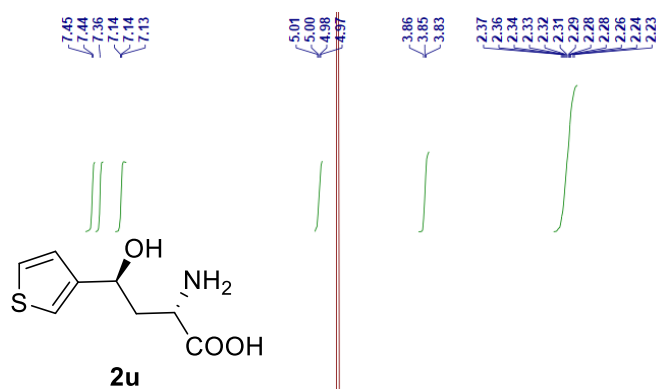
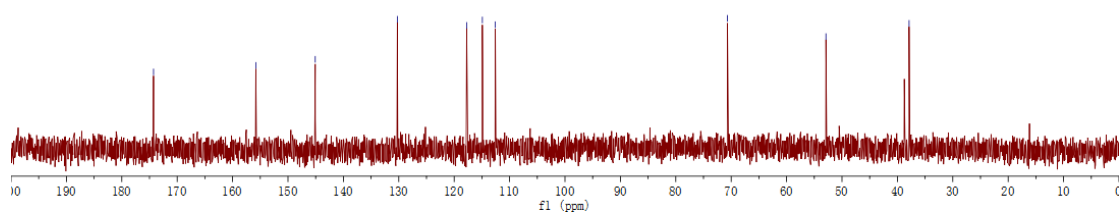
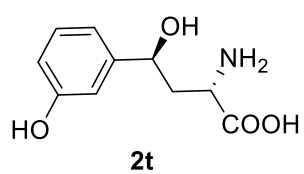


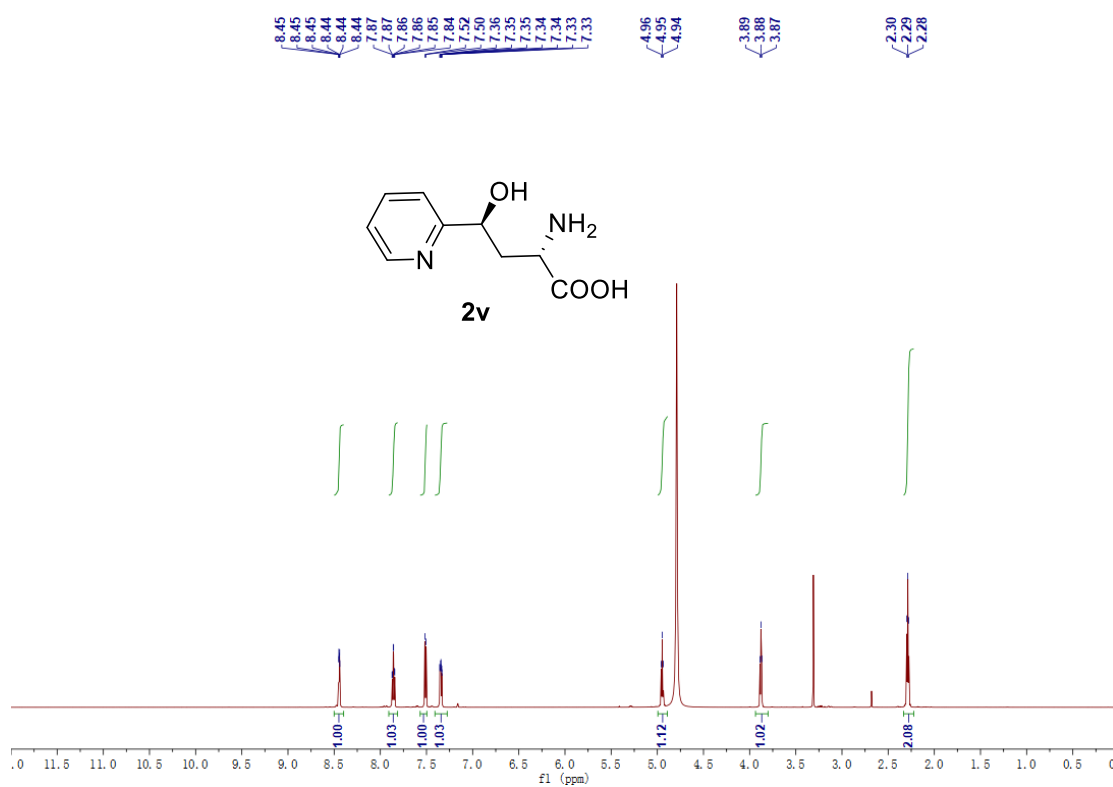
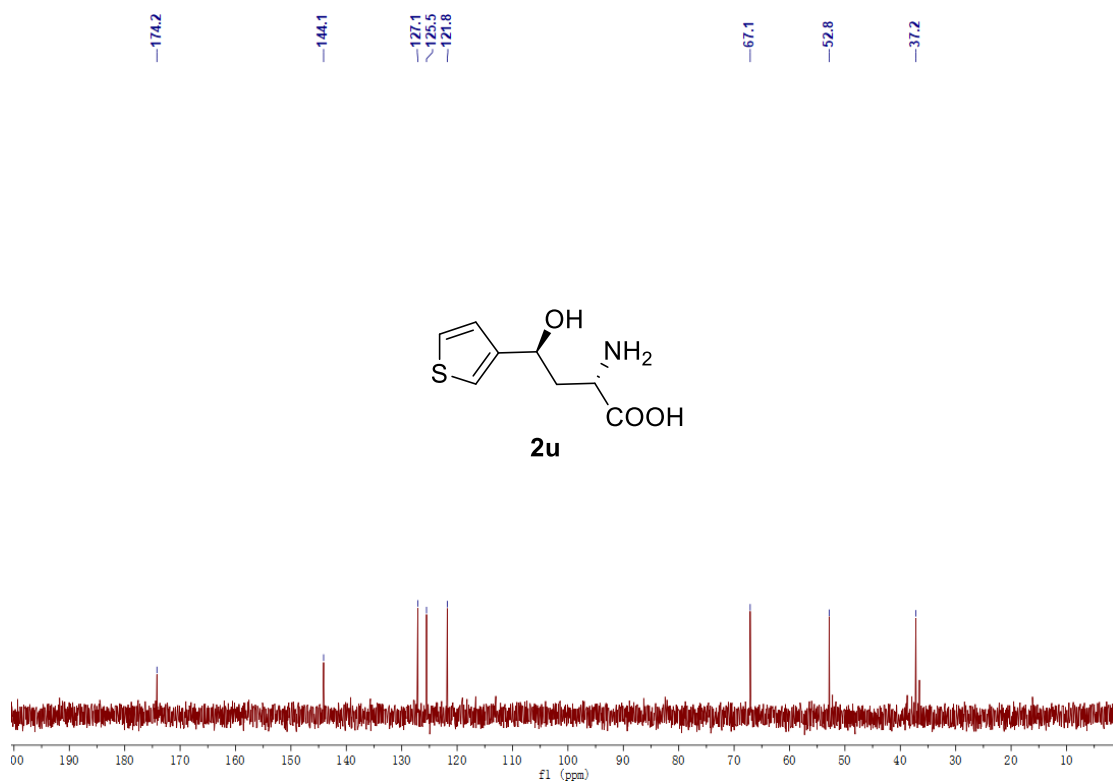


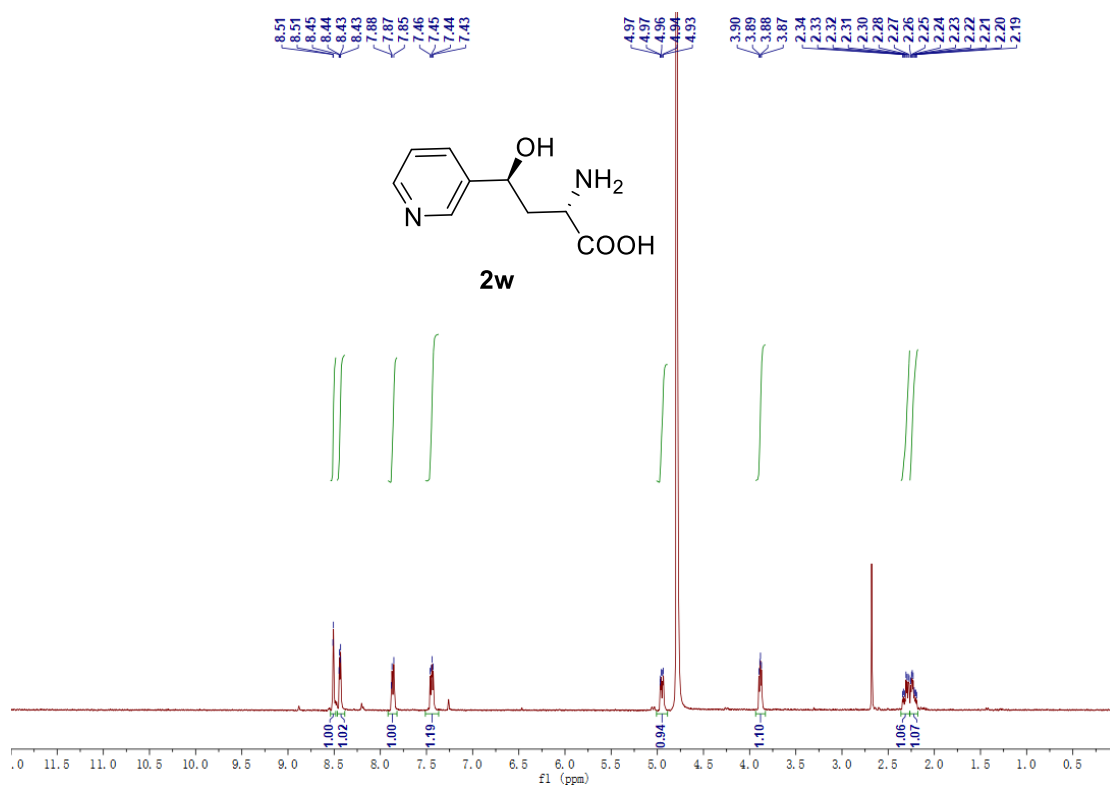
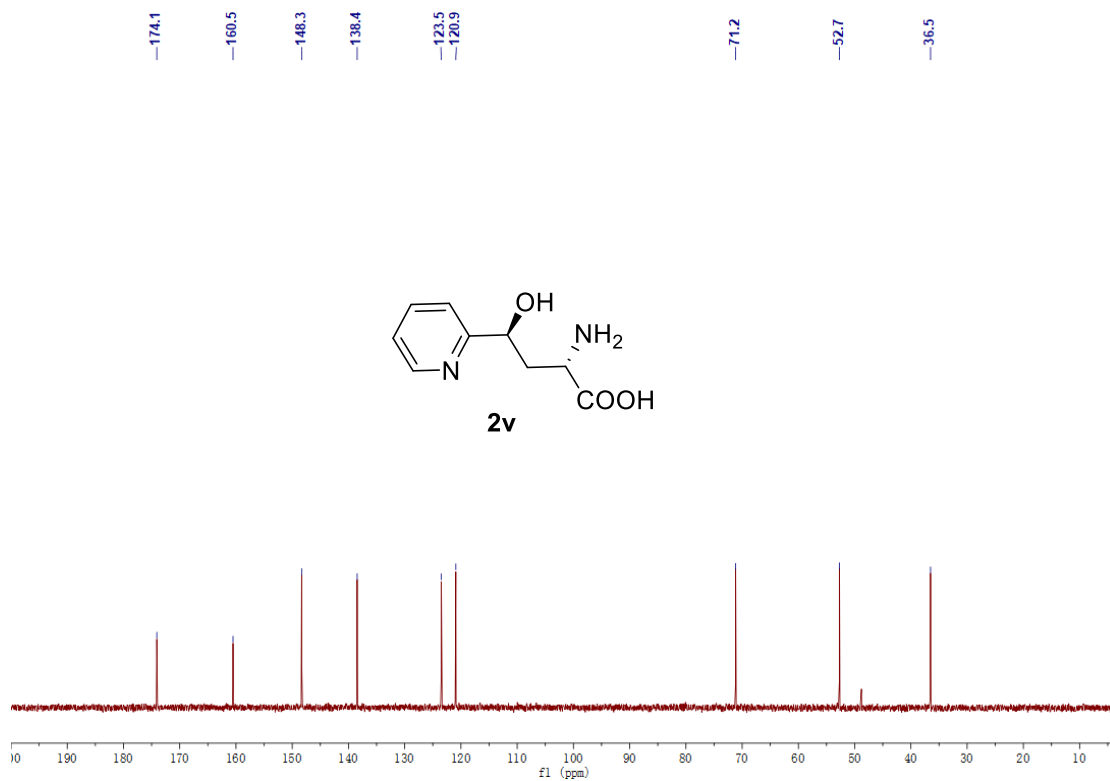


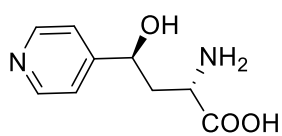
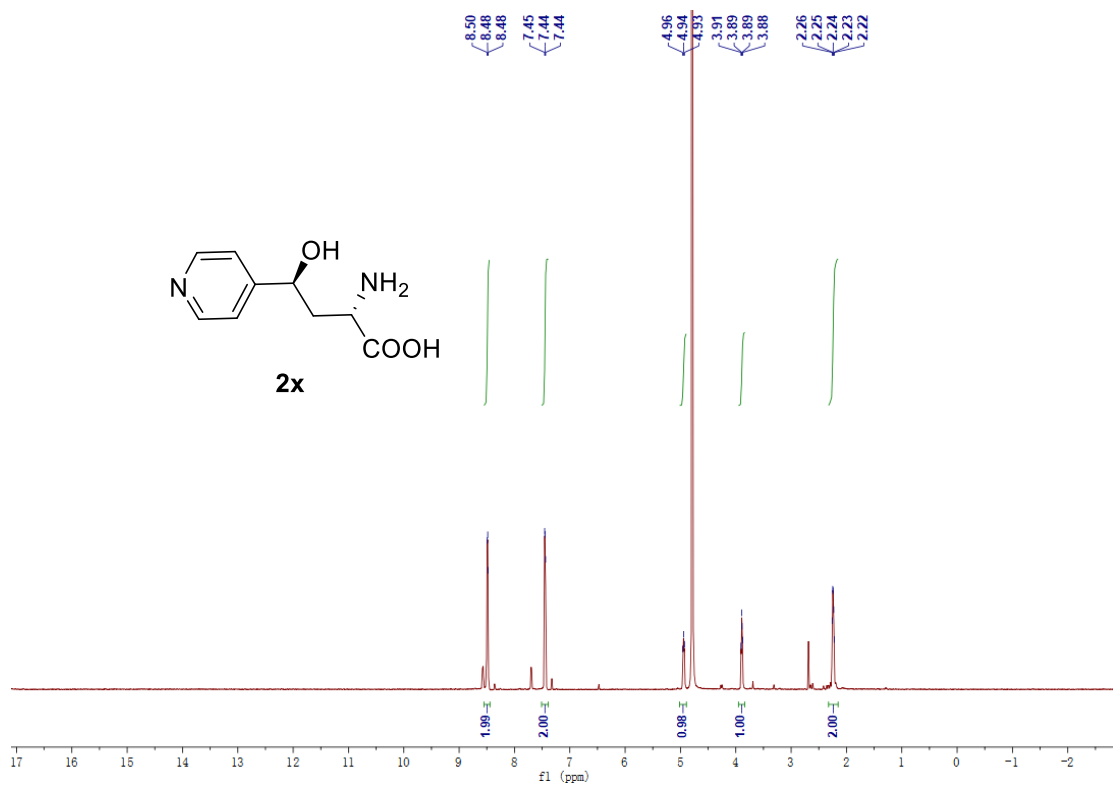
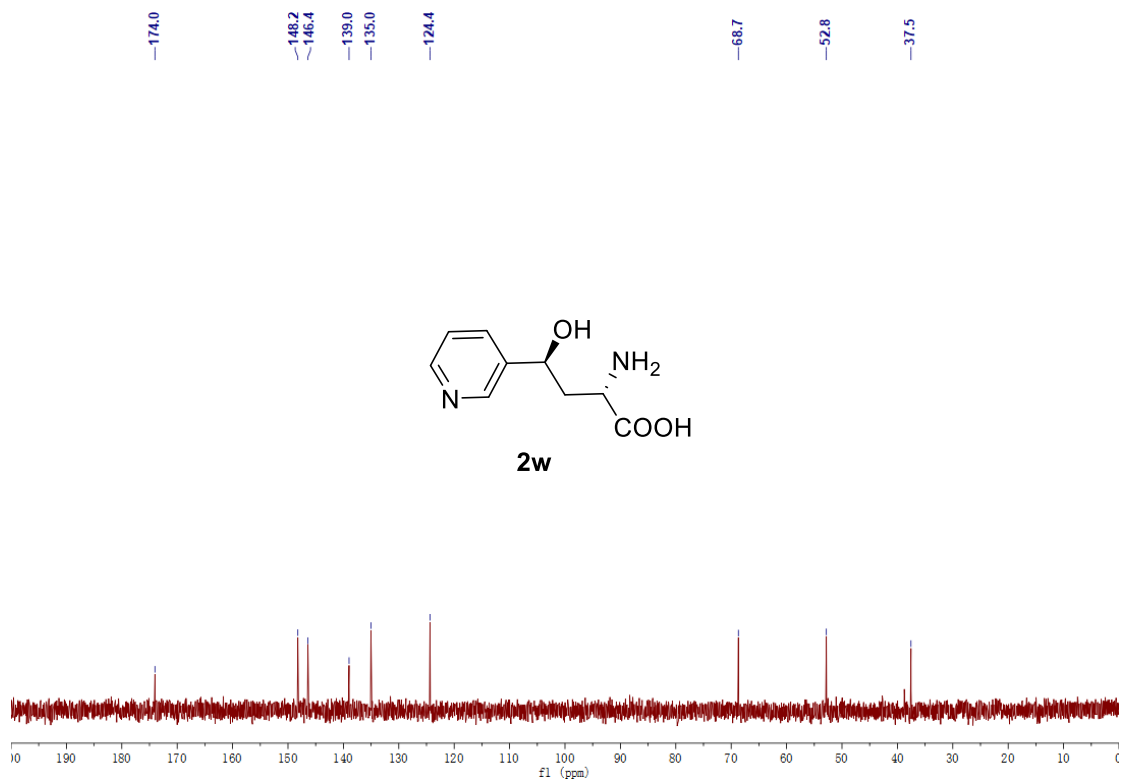


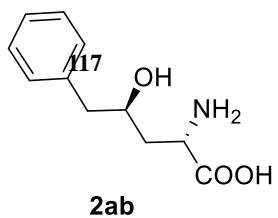
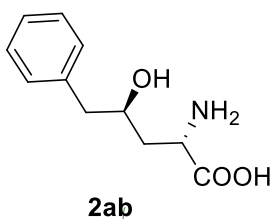
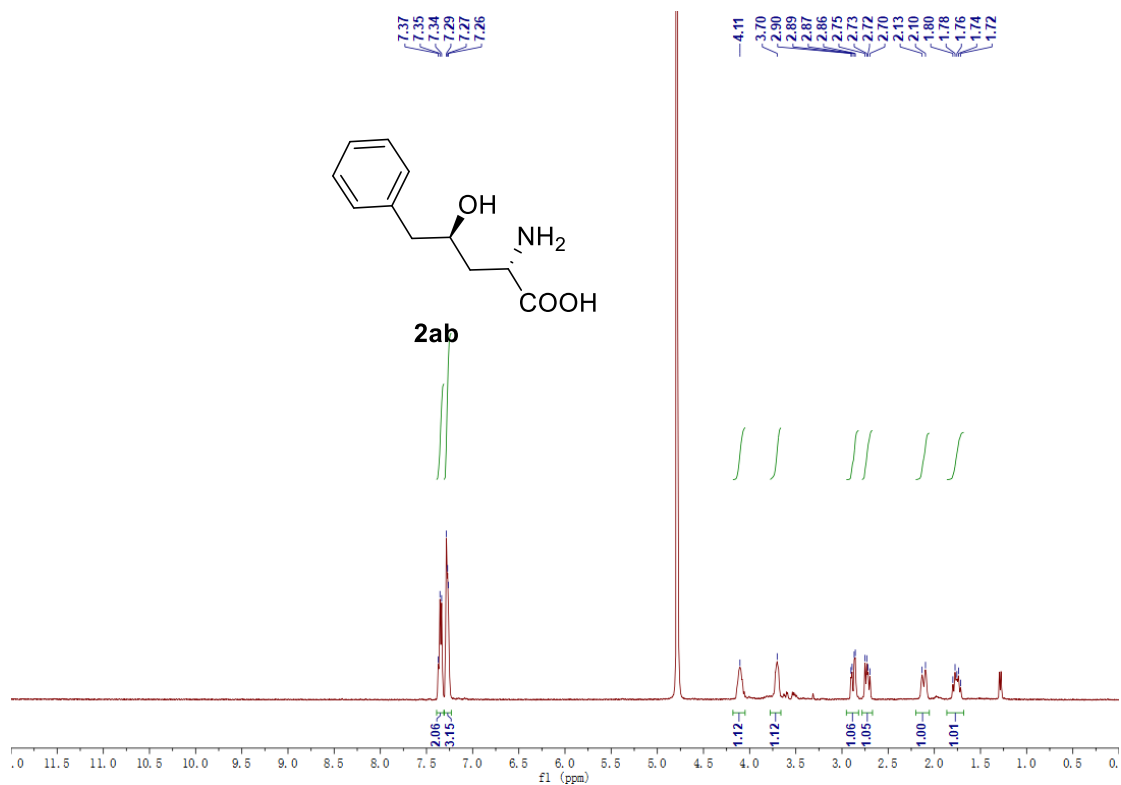
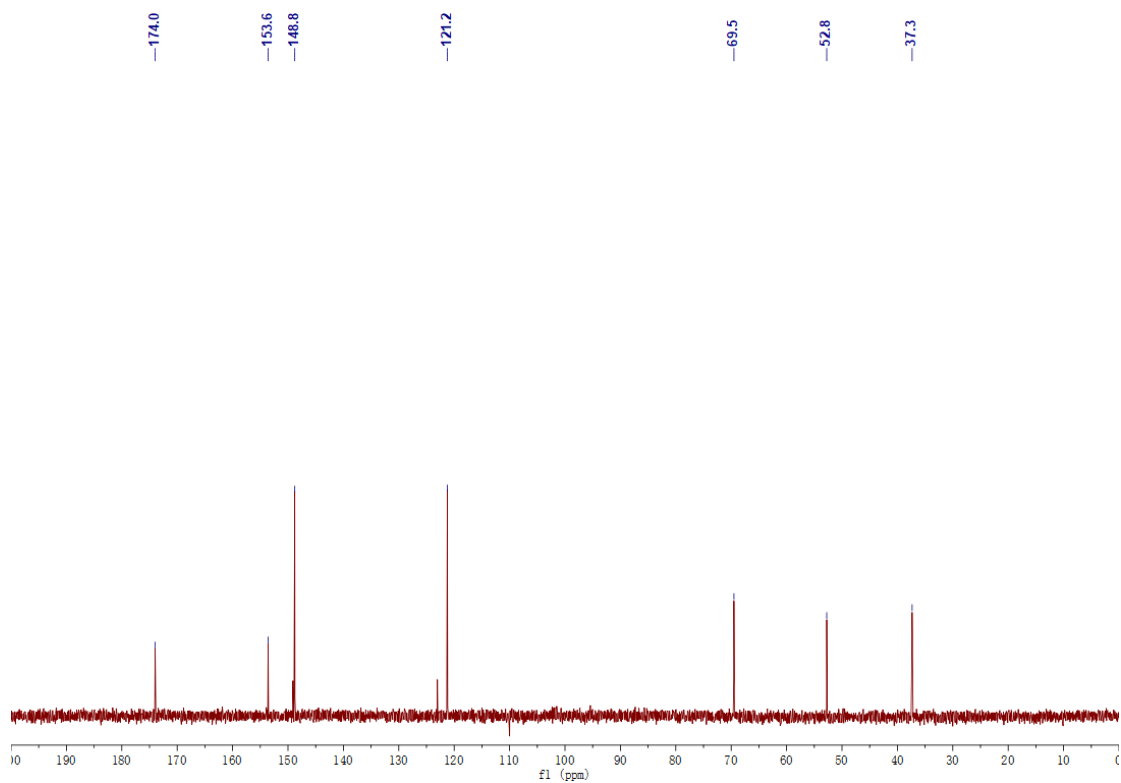
—174.3 —155.8 —145.1 —130.3 —117.7 —114.9 —112.6 —70.7 —52.9 —37.9

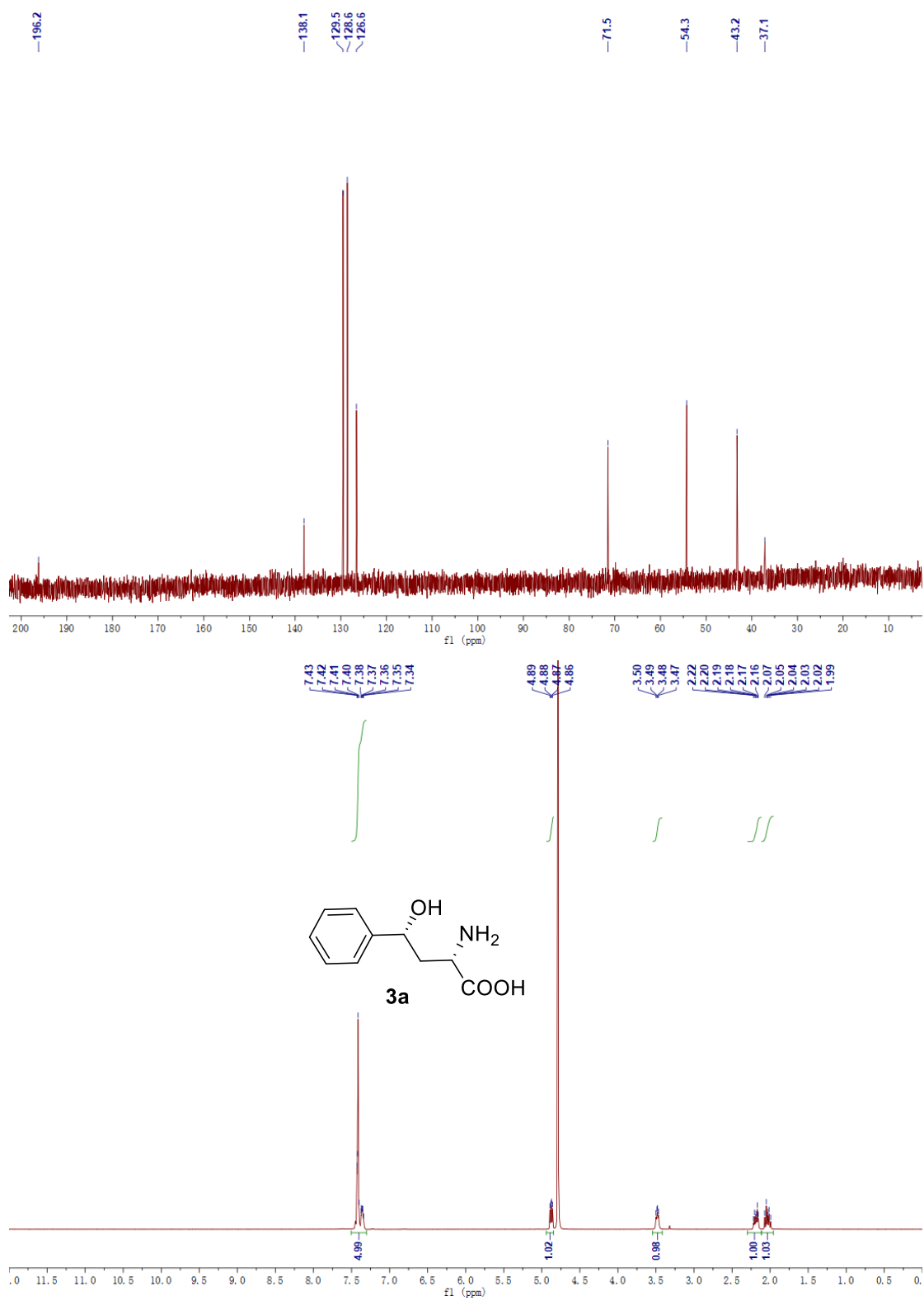


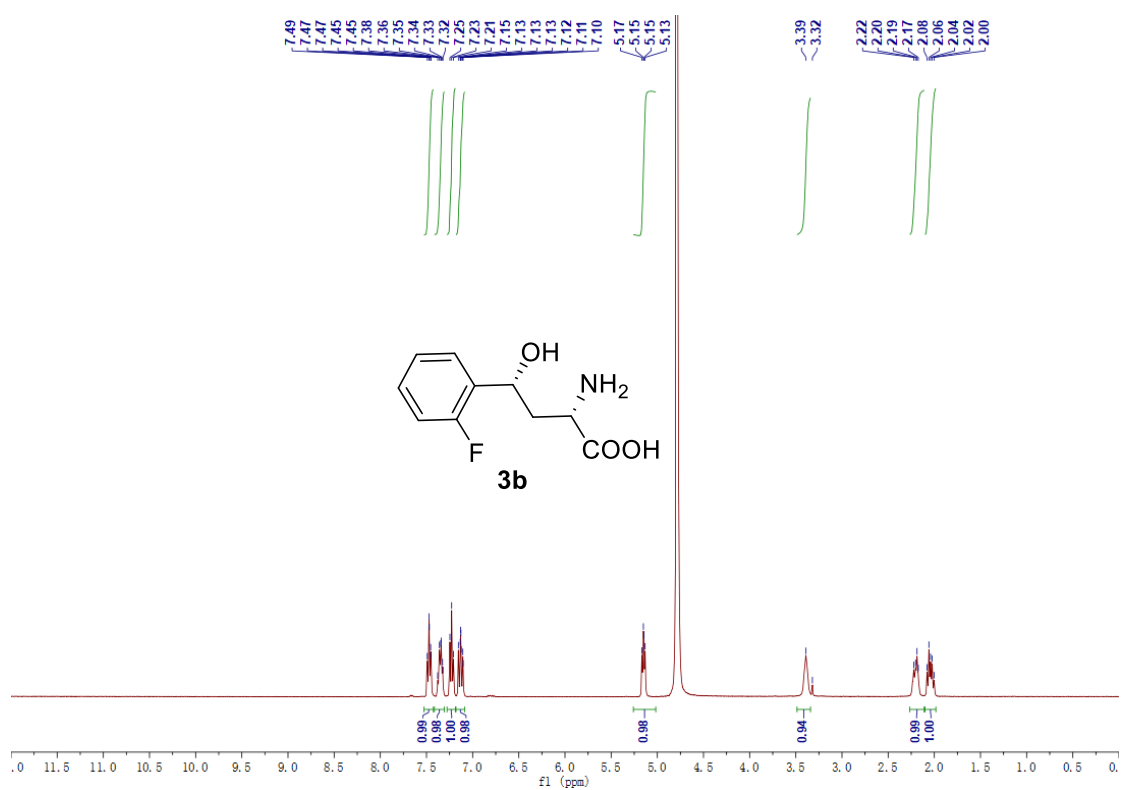
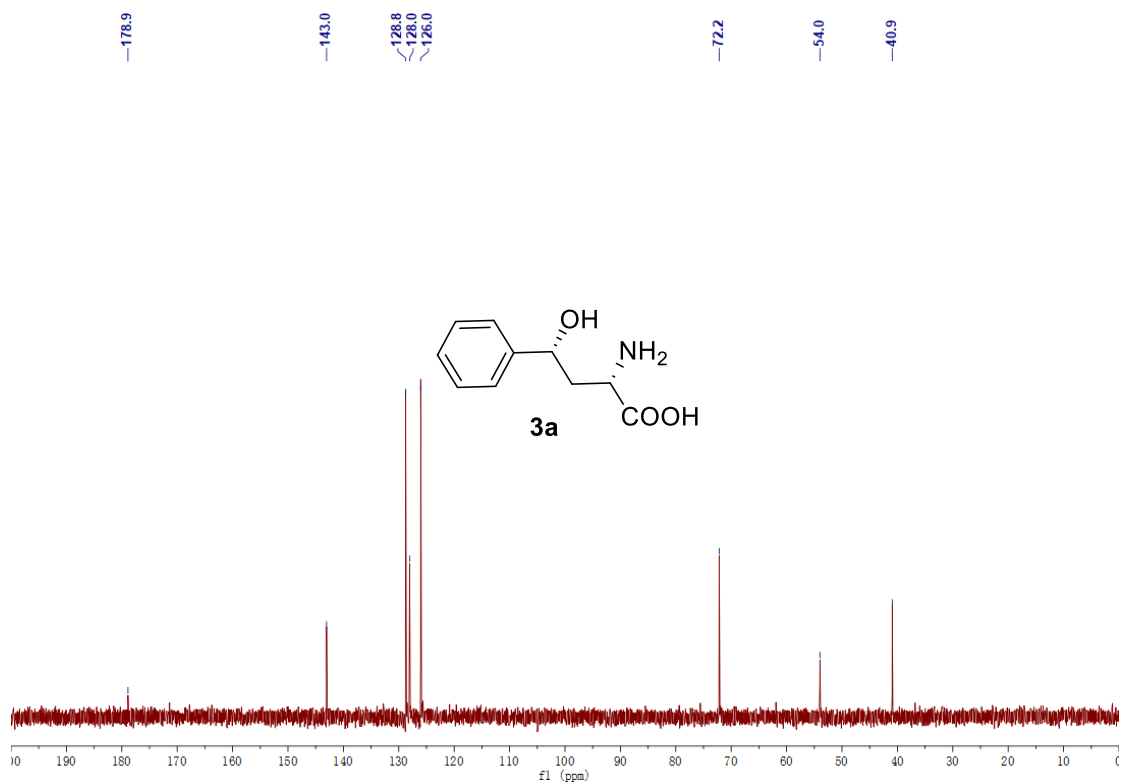


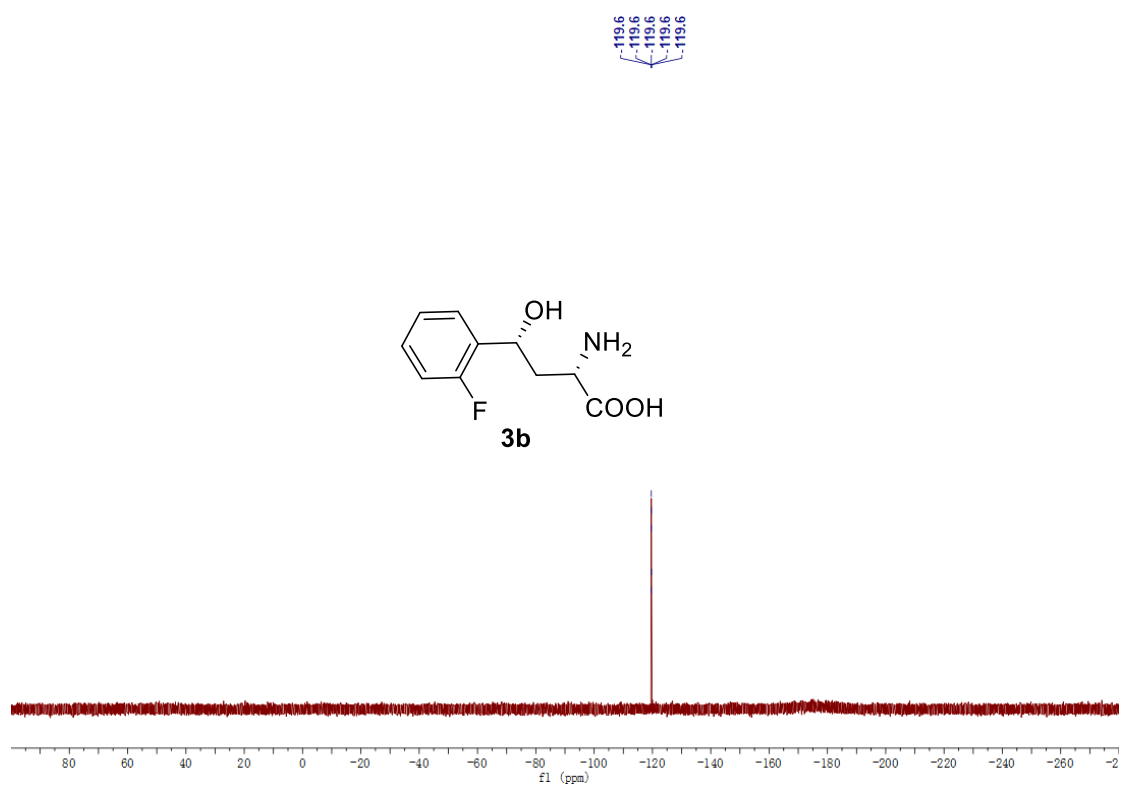
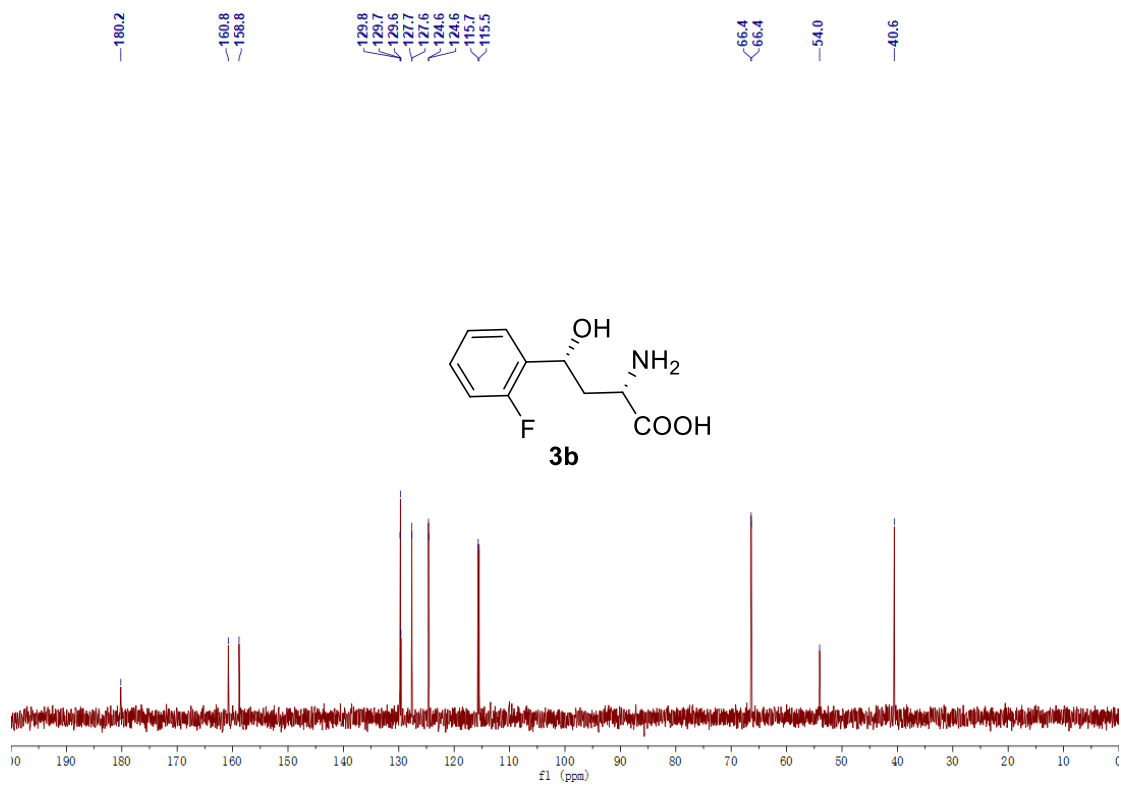


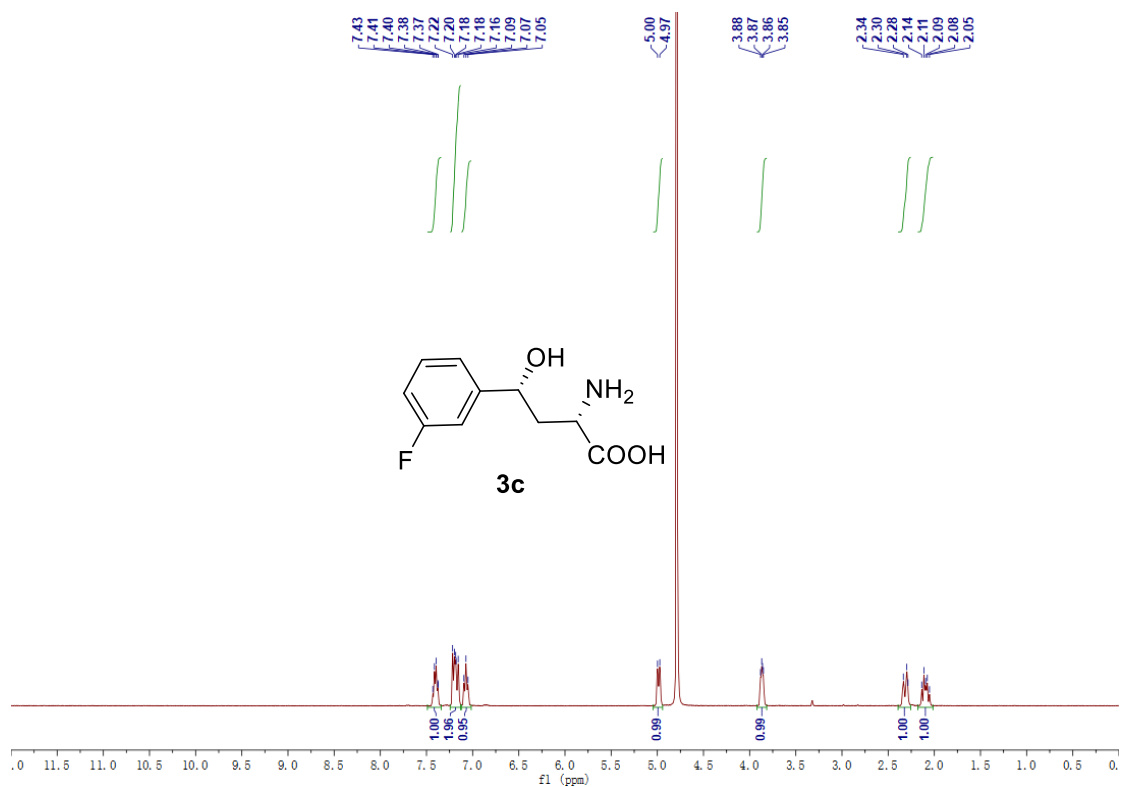












174.2

163.9
161.5

145.7

130.5
130.4

121.5

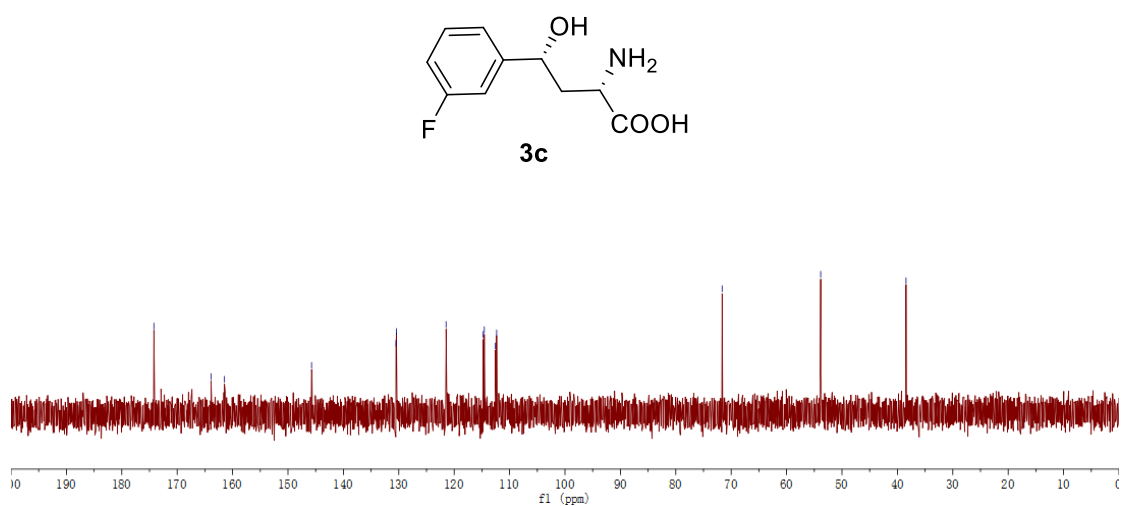
114.8
114.6

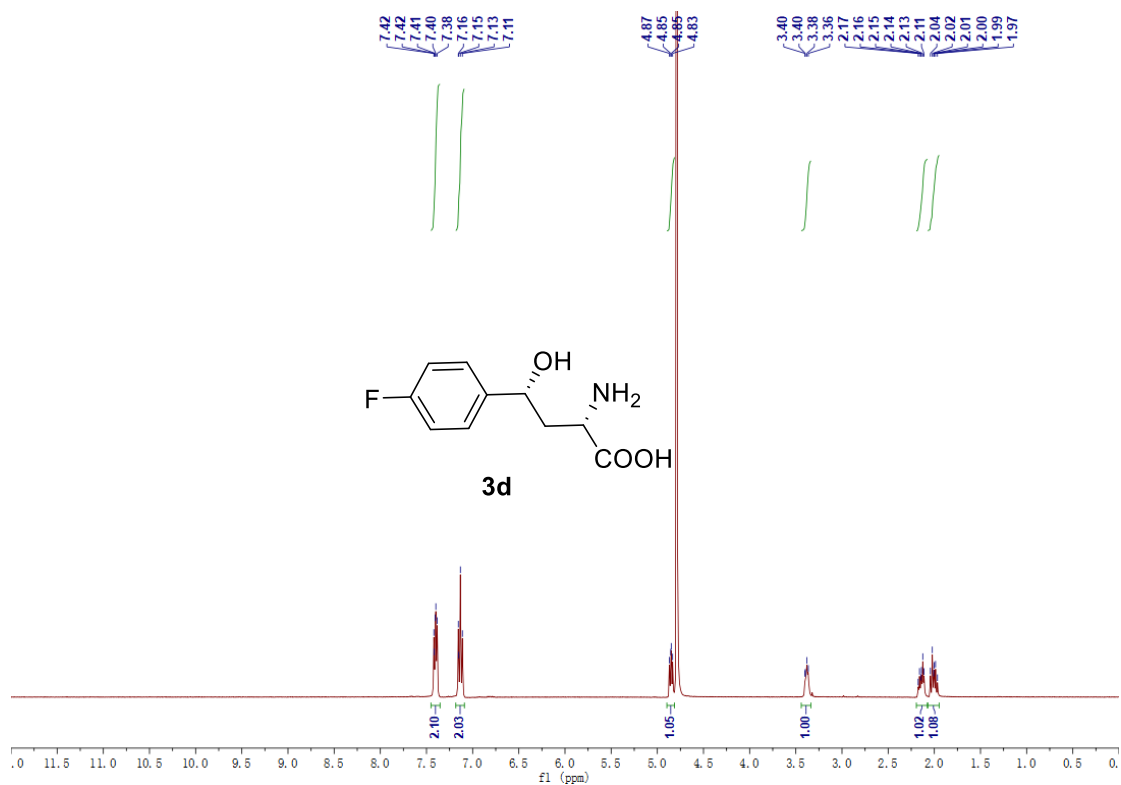
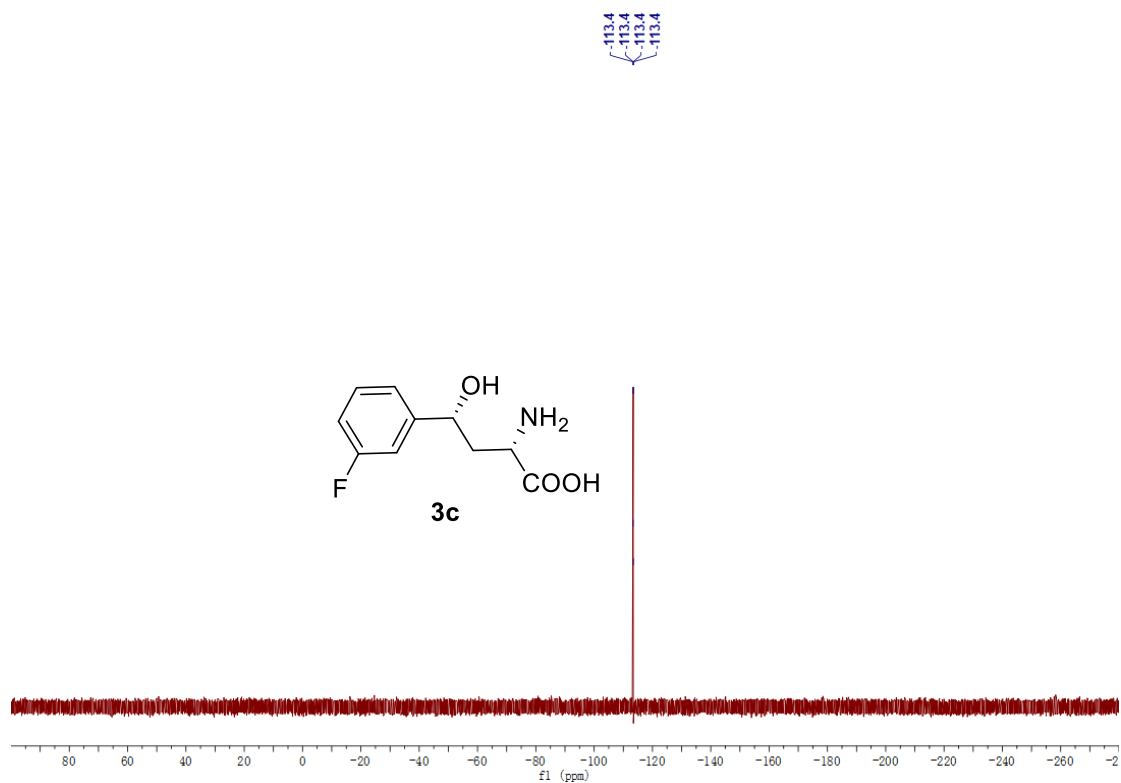
112.6
112.3

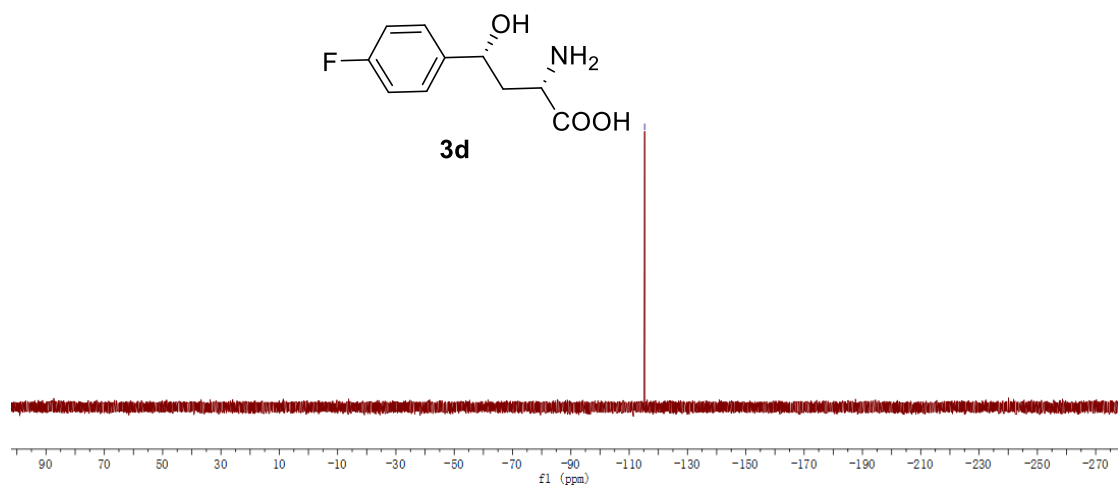
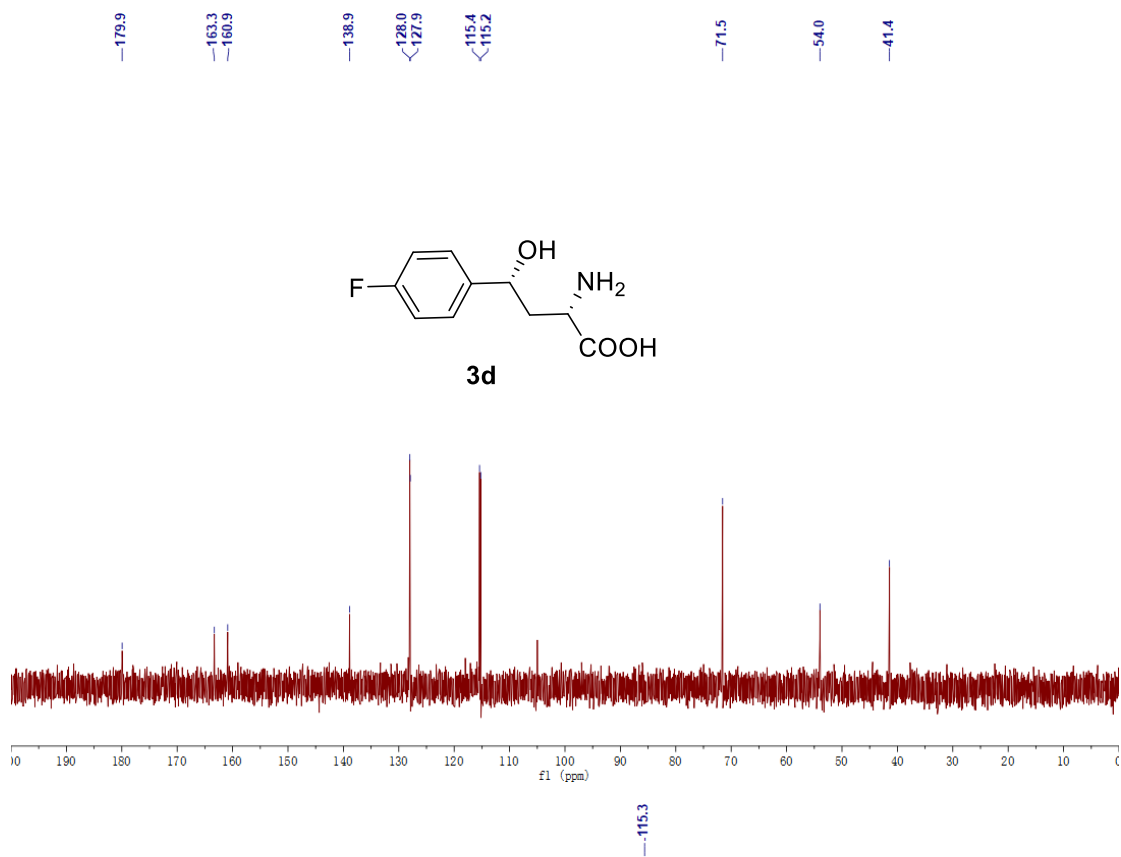
71.6

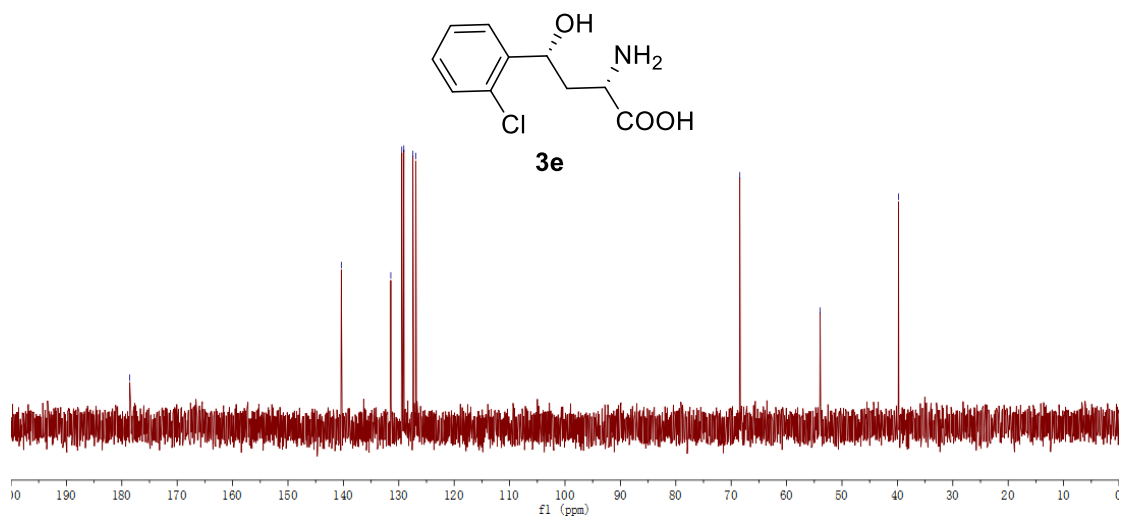
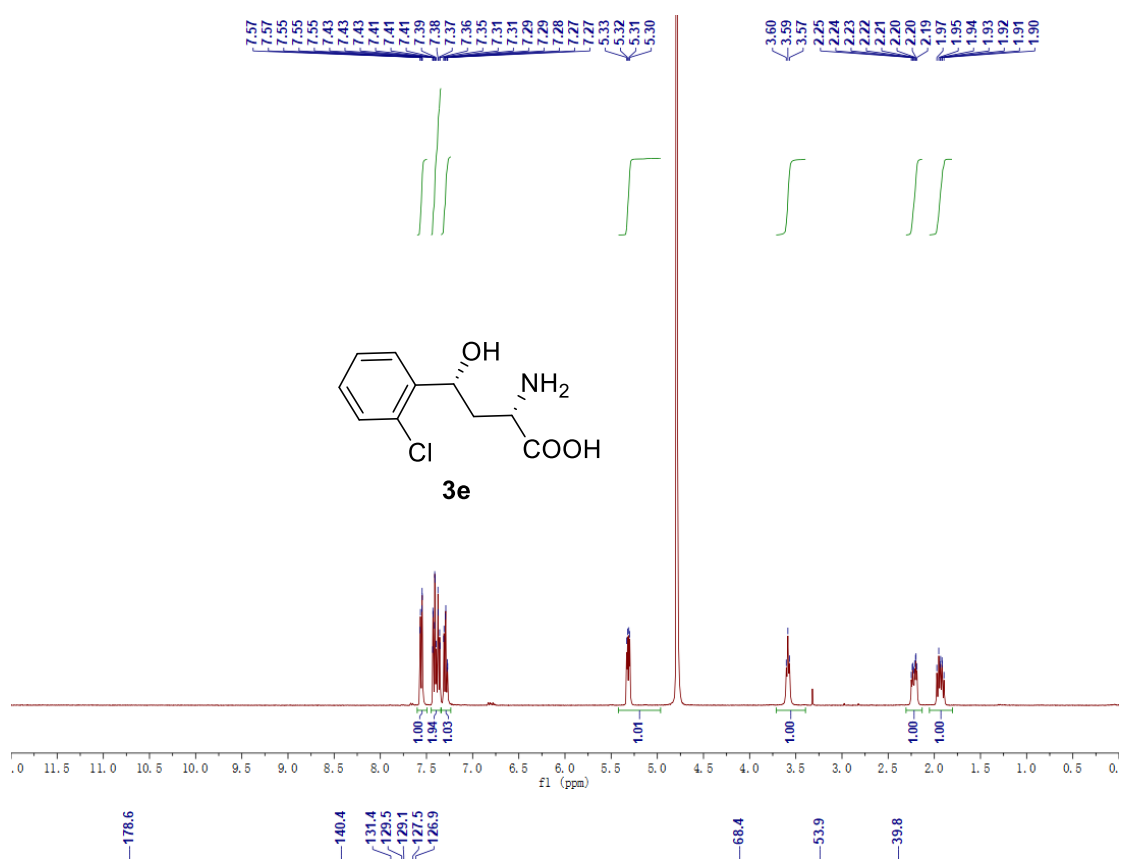
53.8

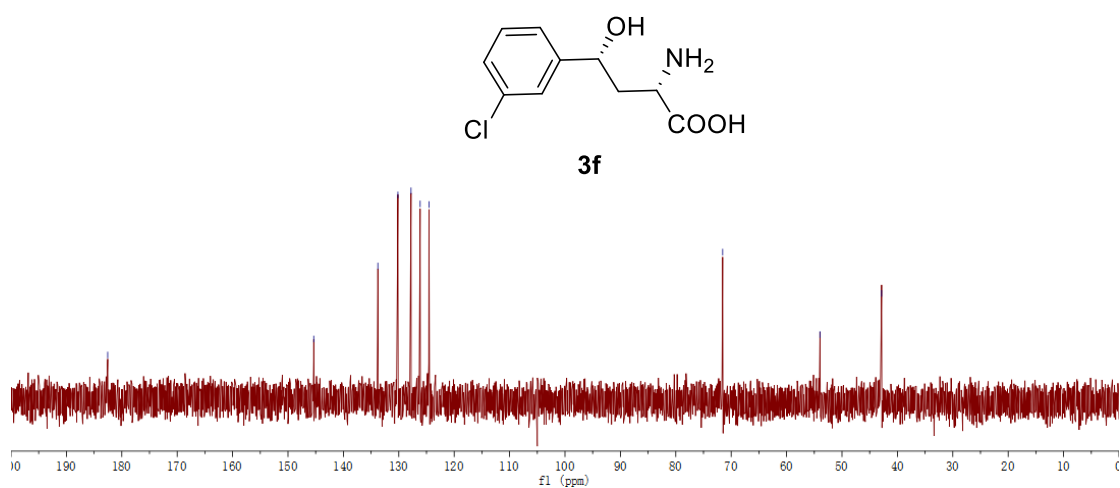
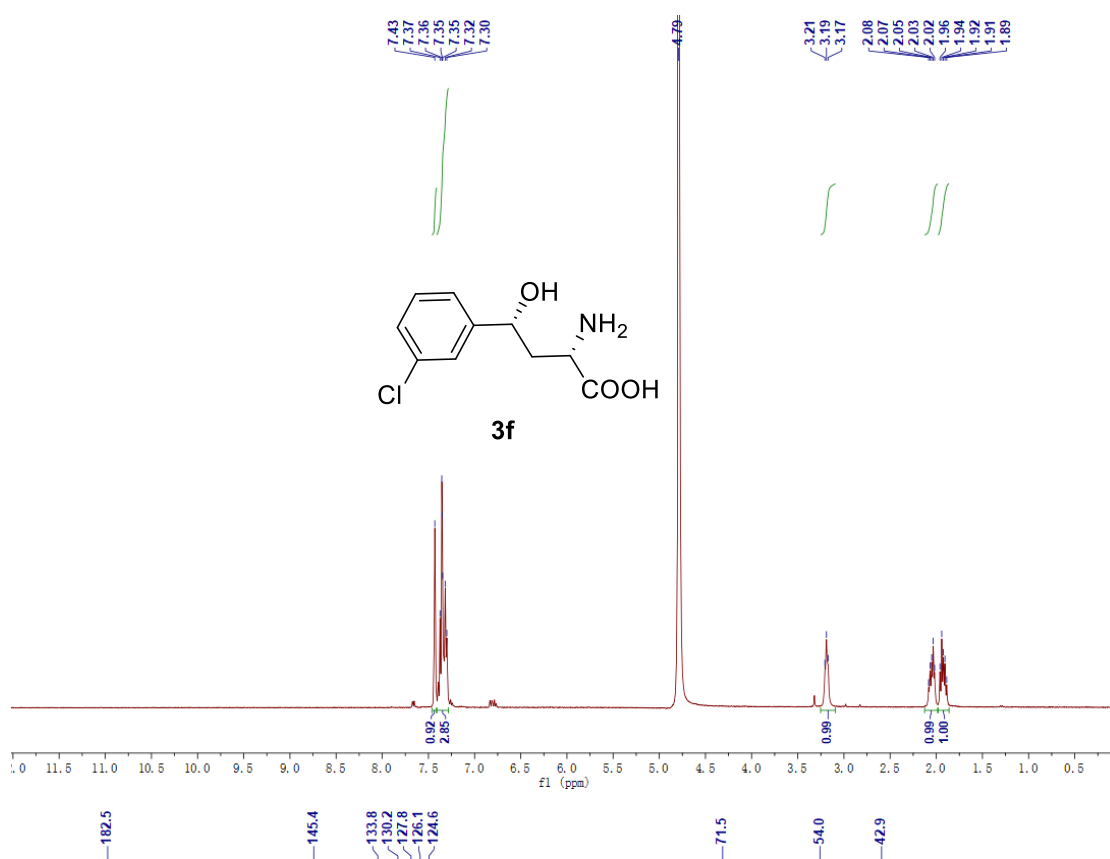
38.4

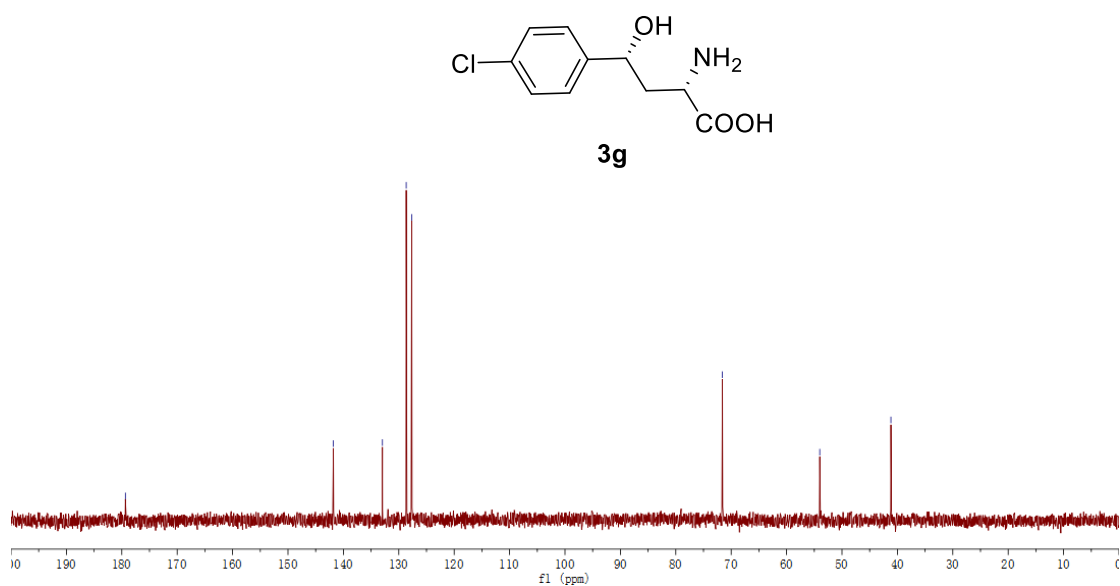
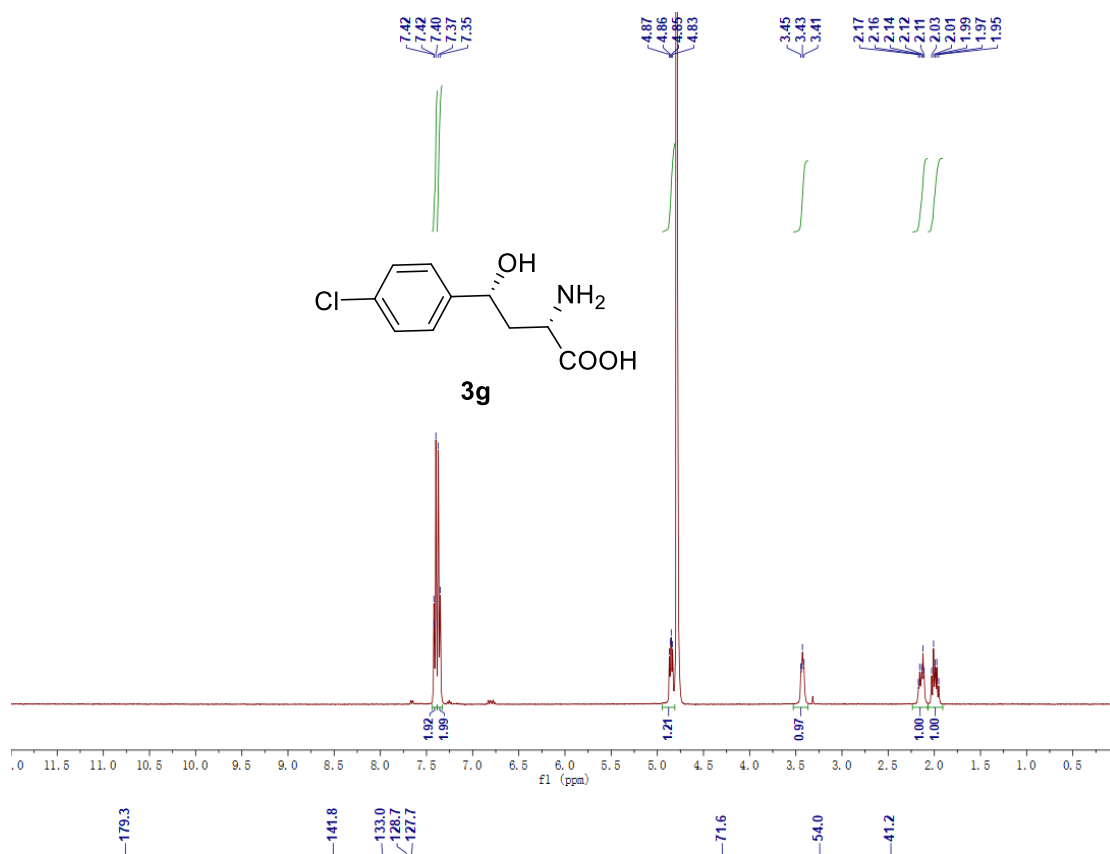


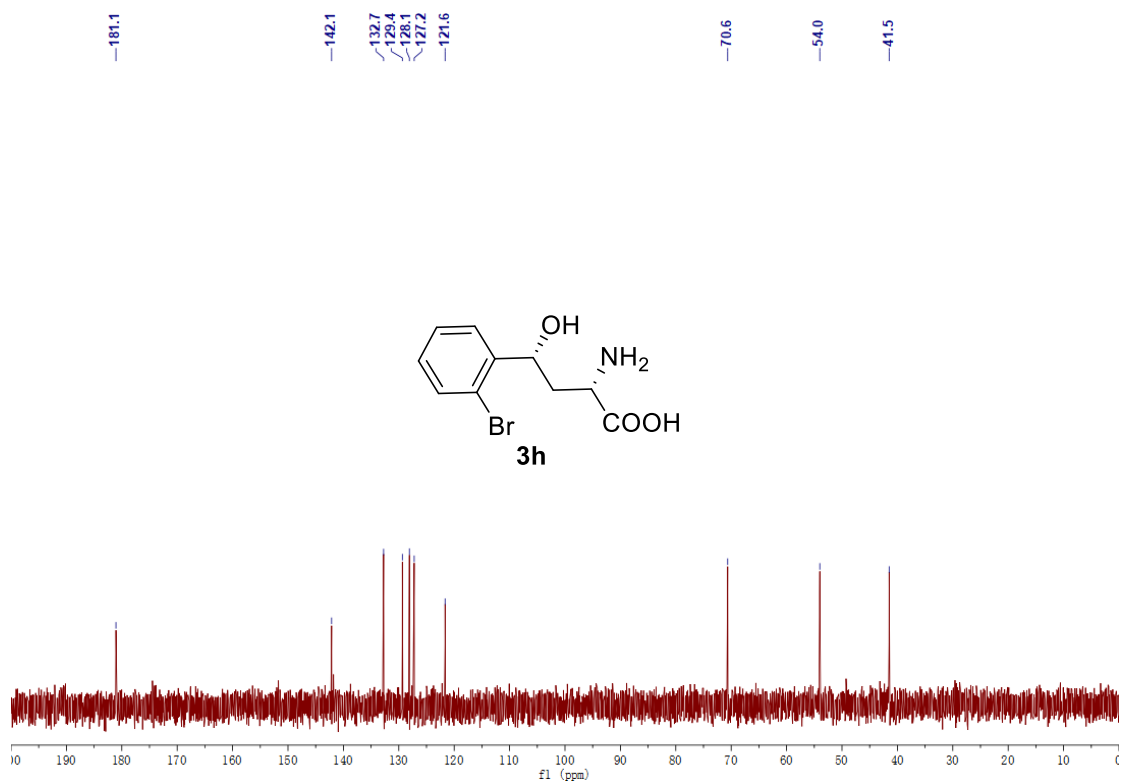
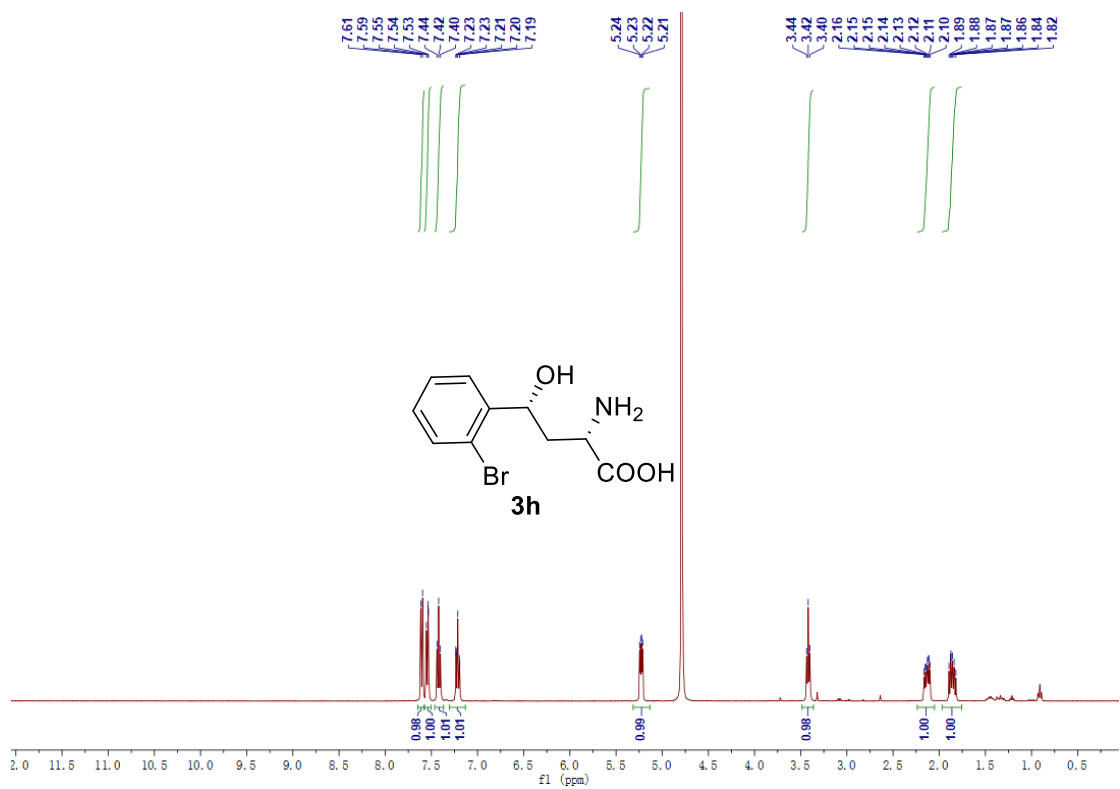


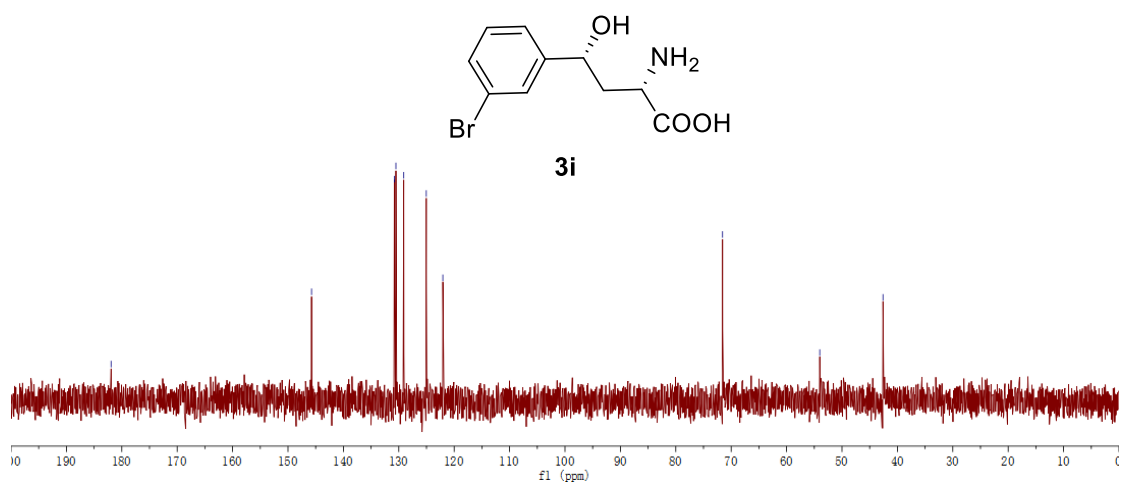
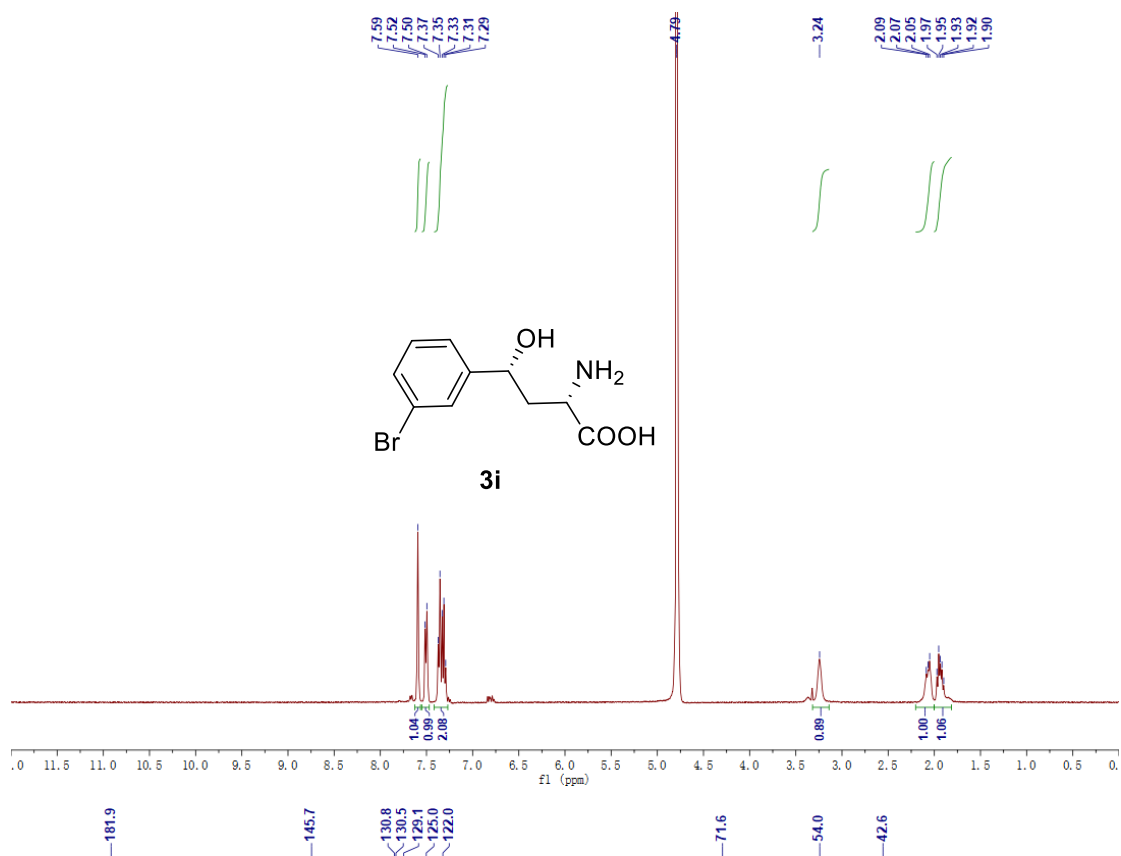


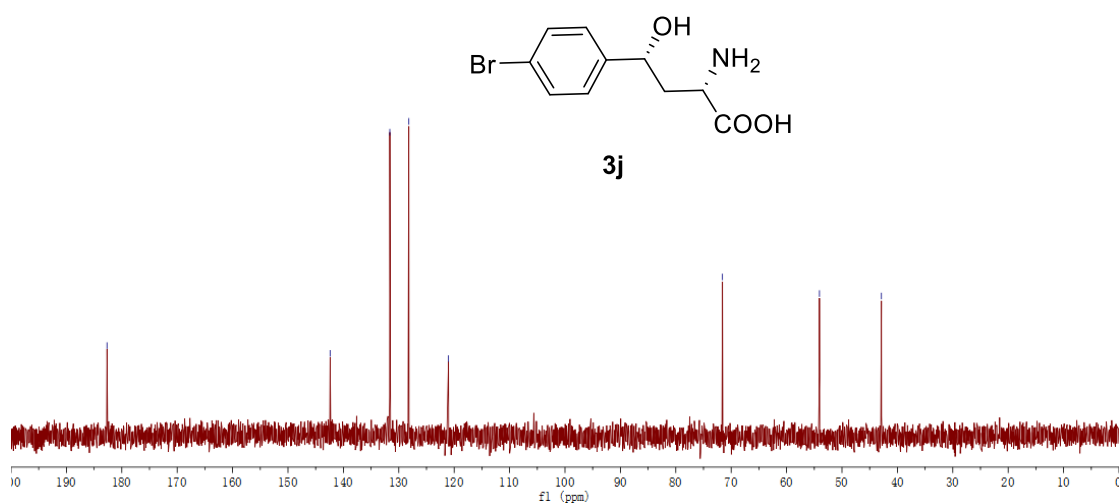
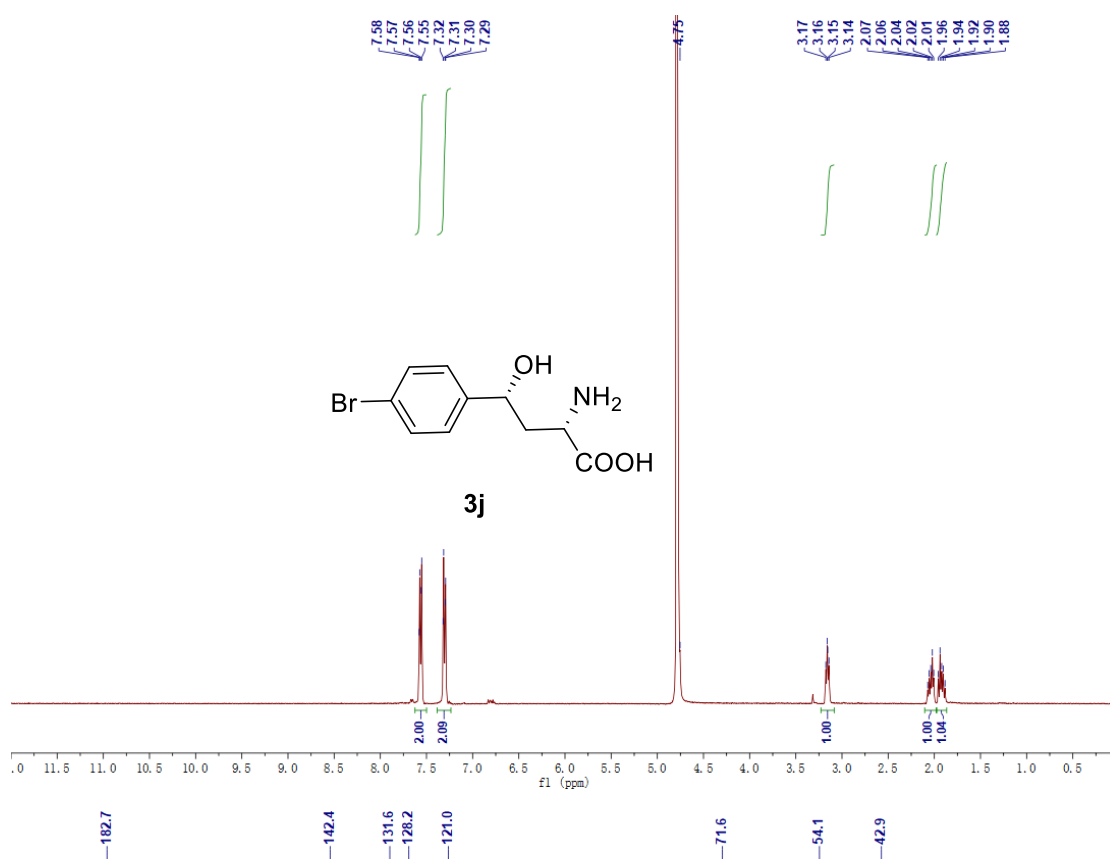


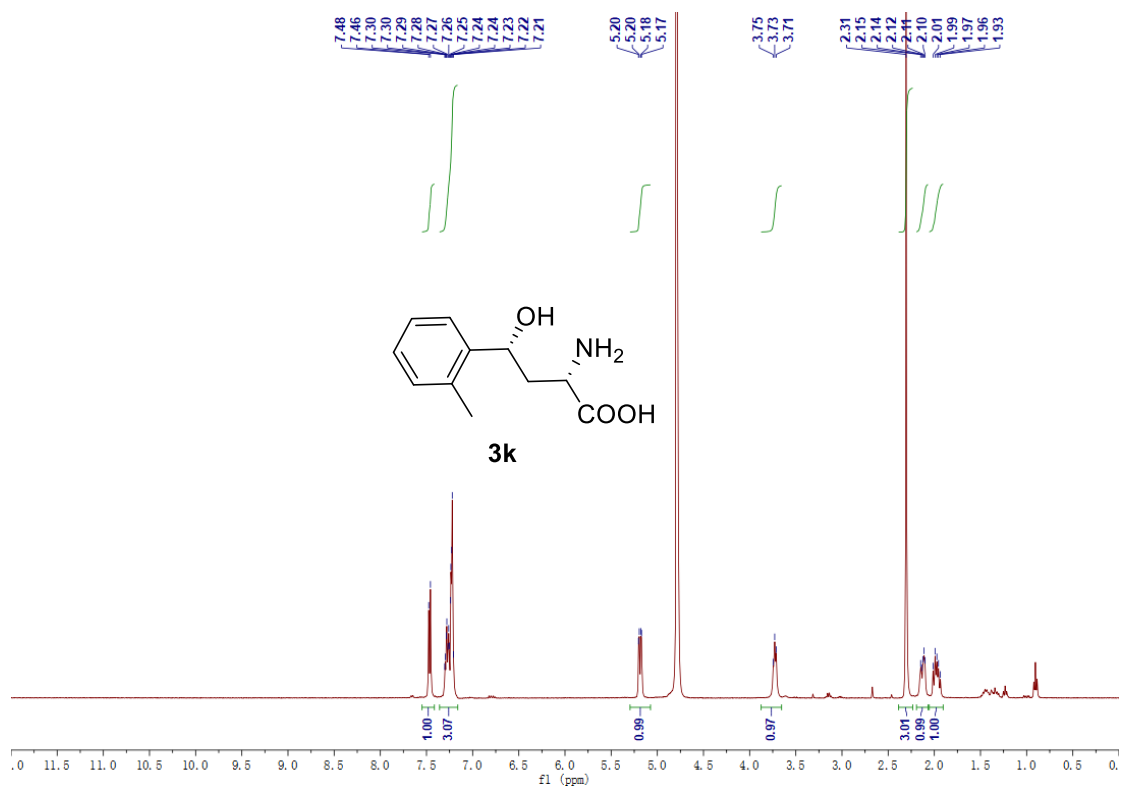












—176.9

—141.4

—134.9

—130.5

—127.7

—126.4

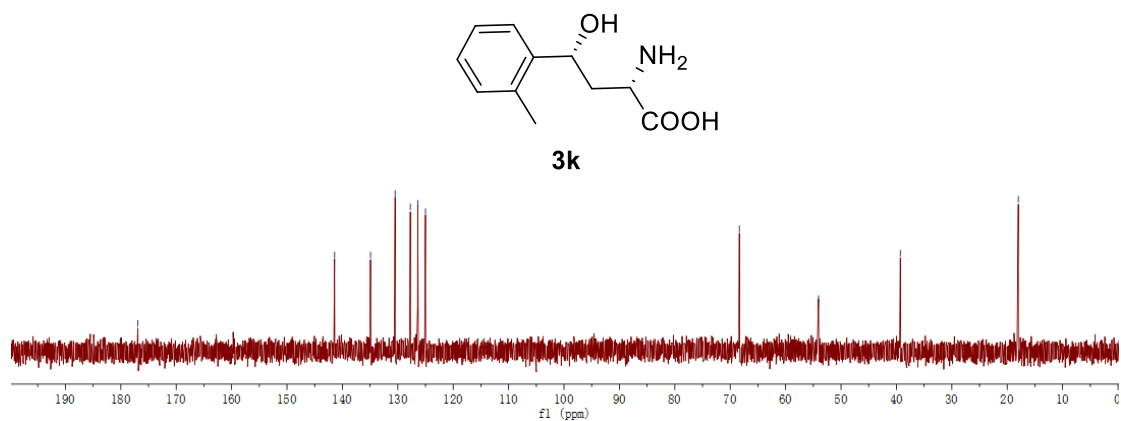
—125.0

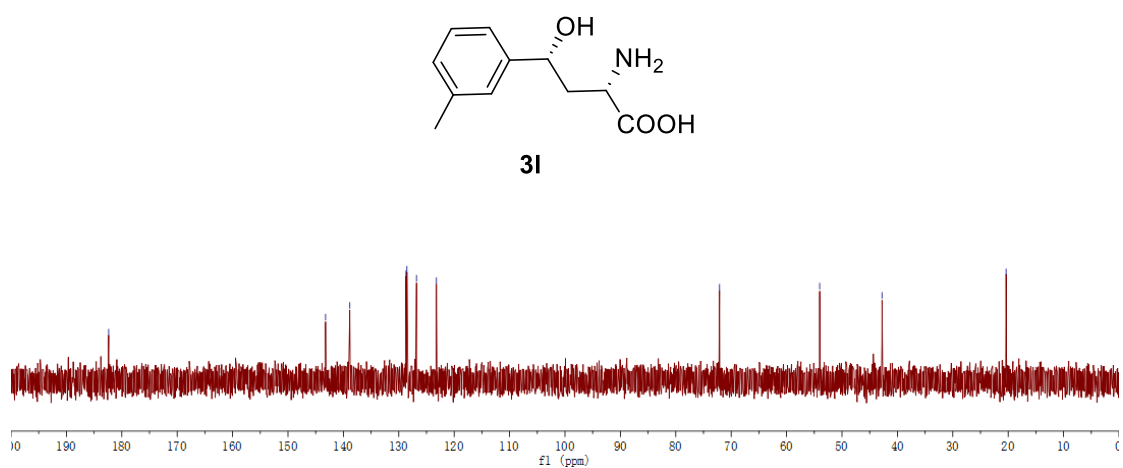
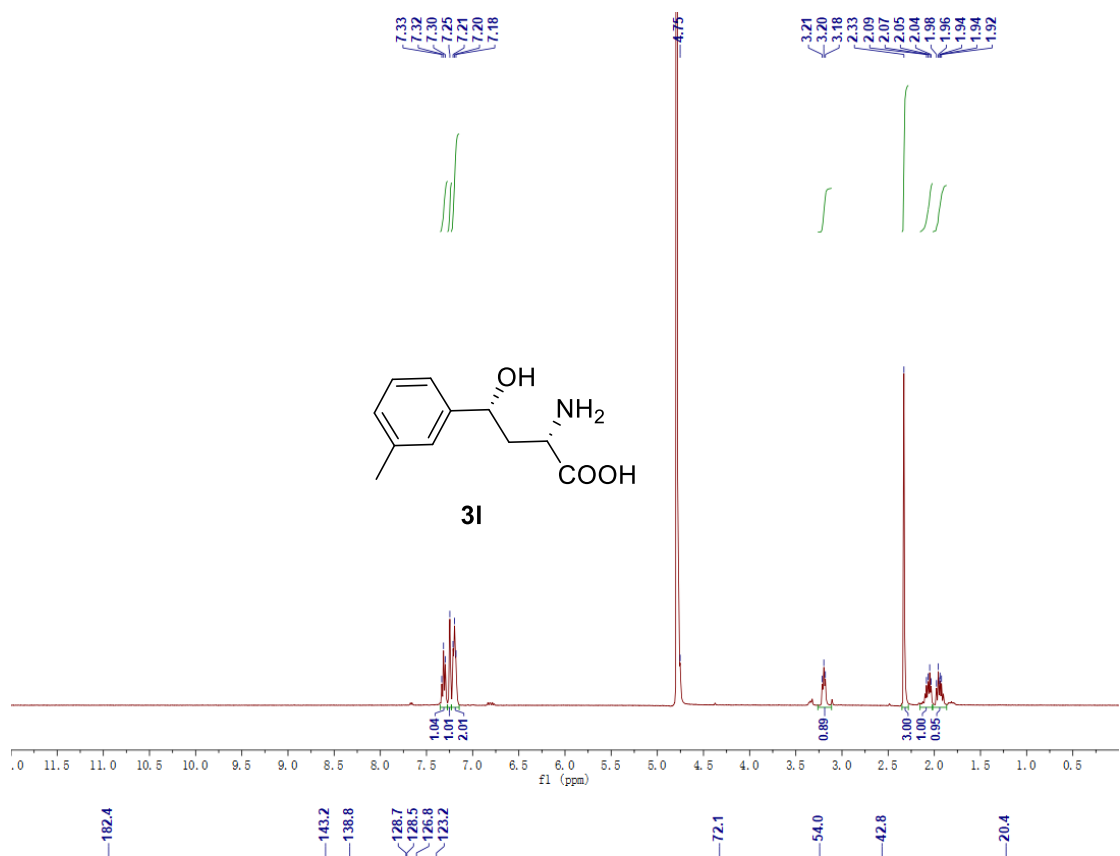
—68.4

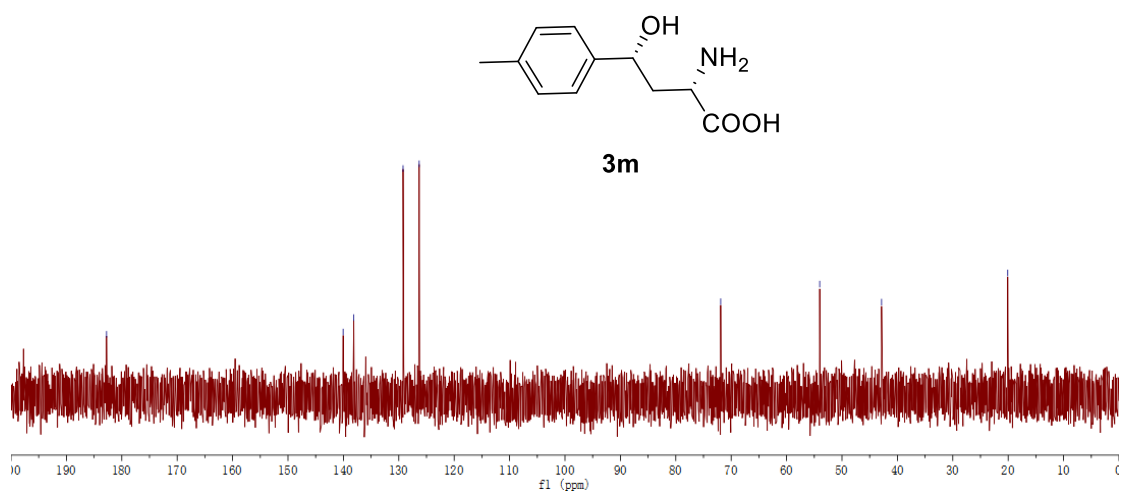
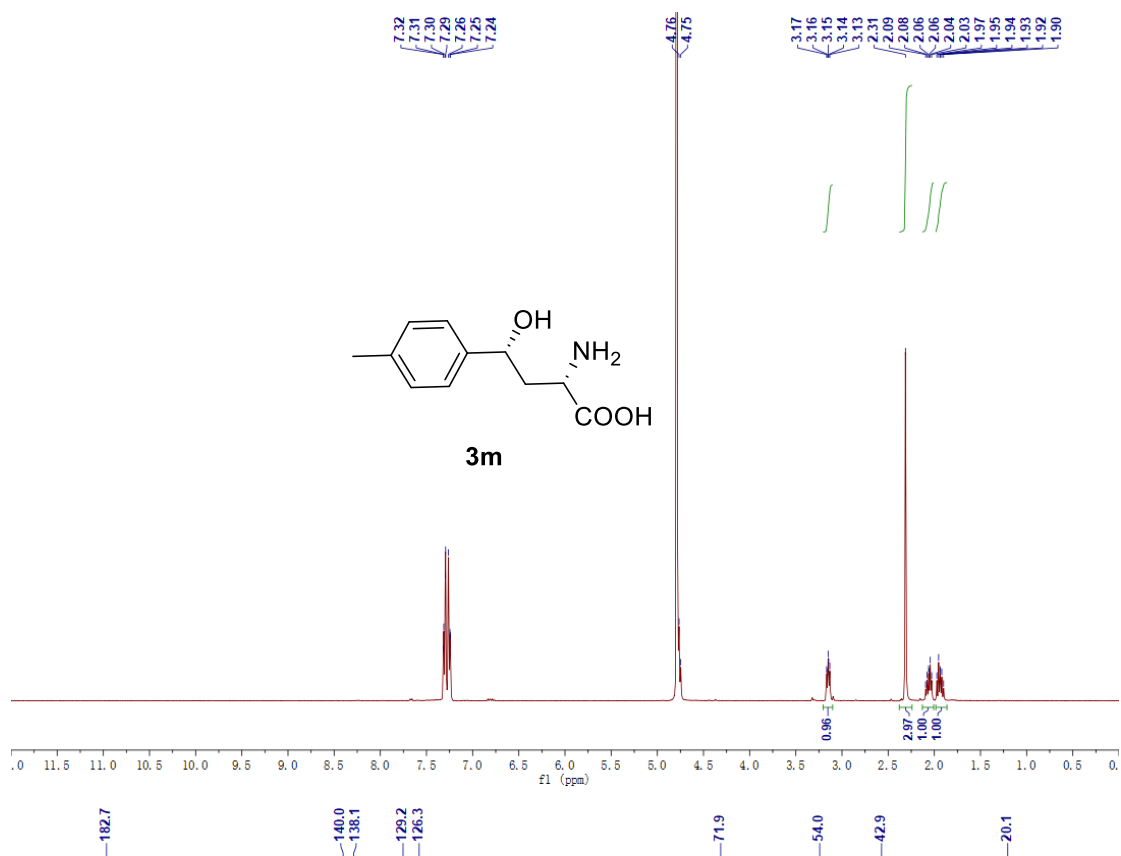
—54.1

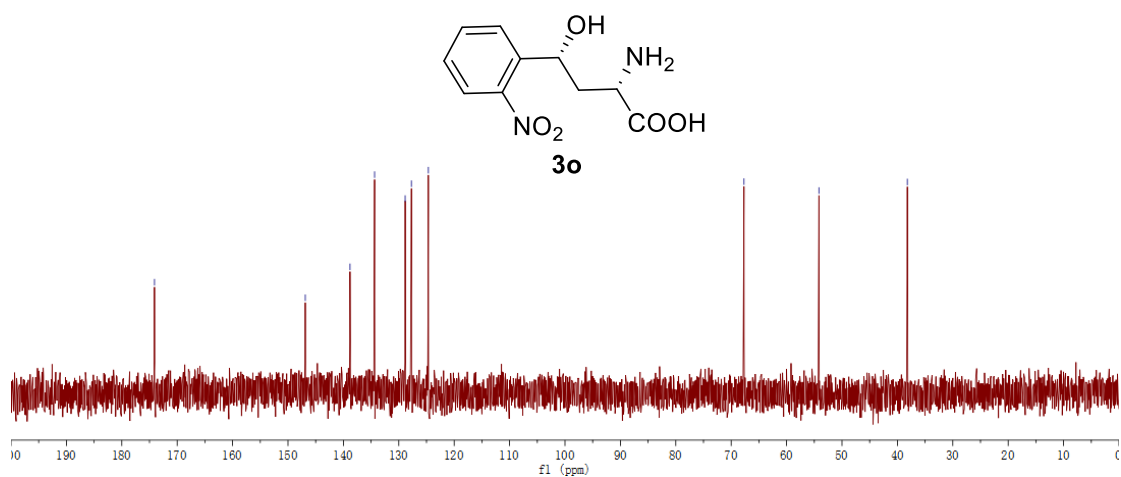
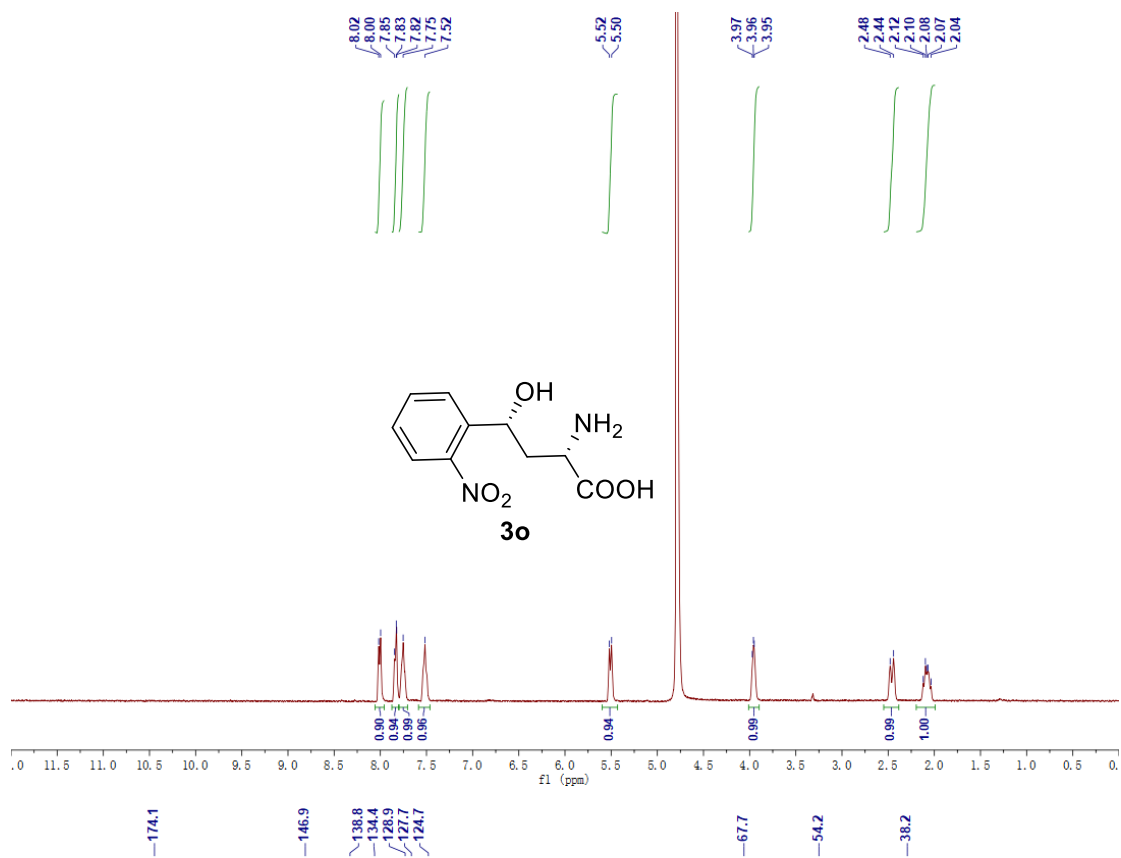
—39.3

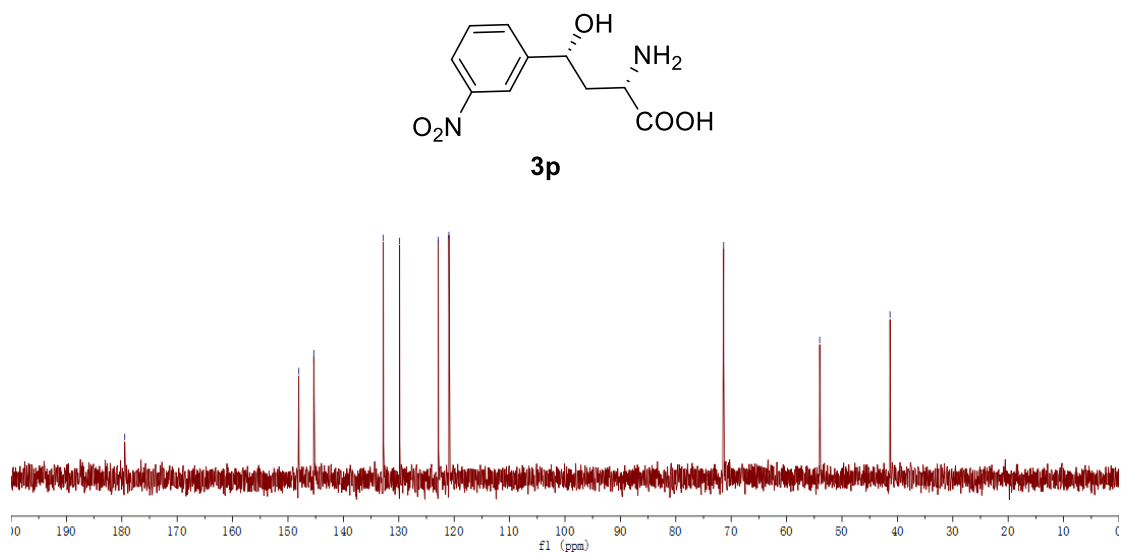
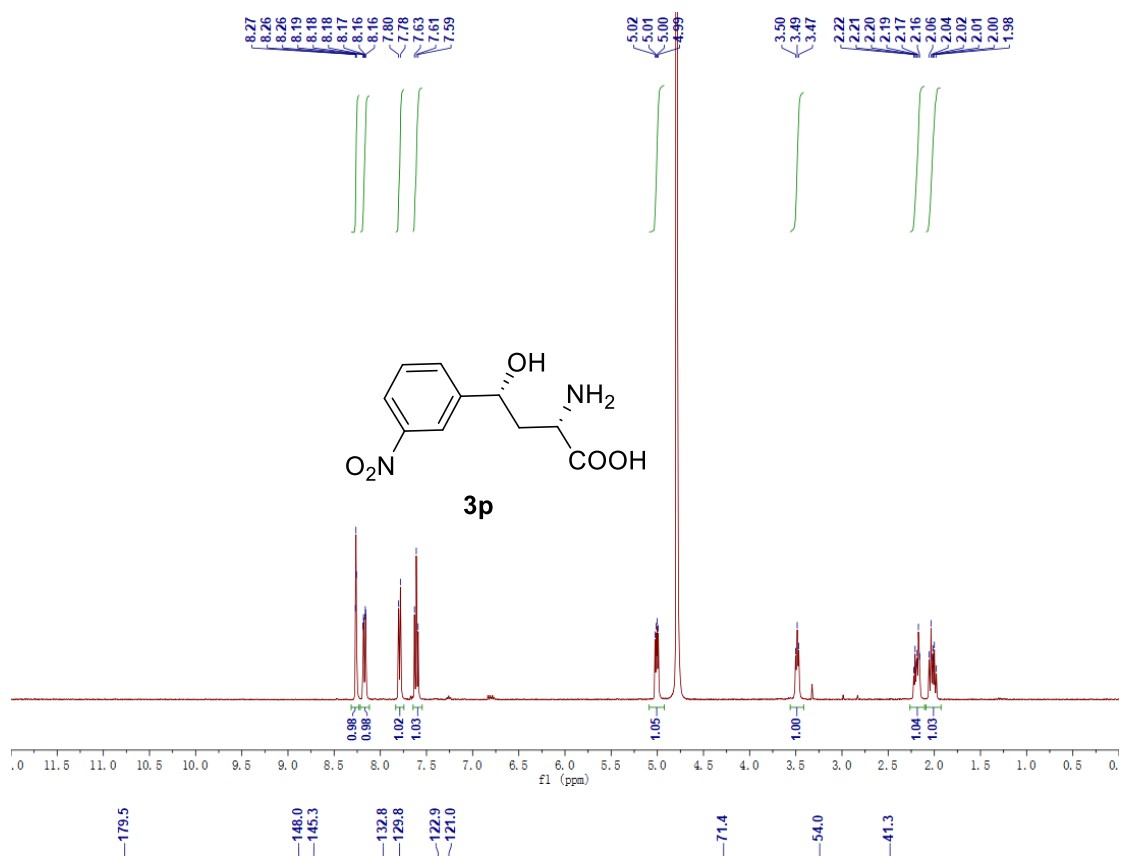
—18.0

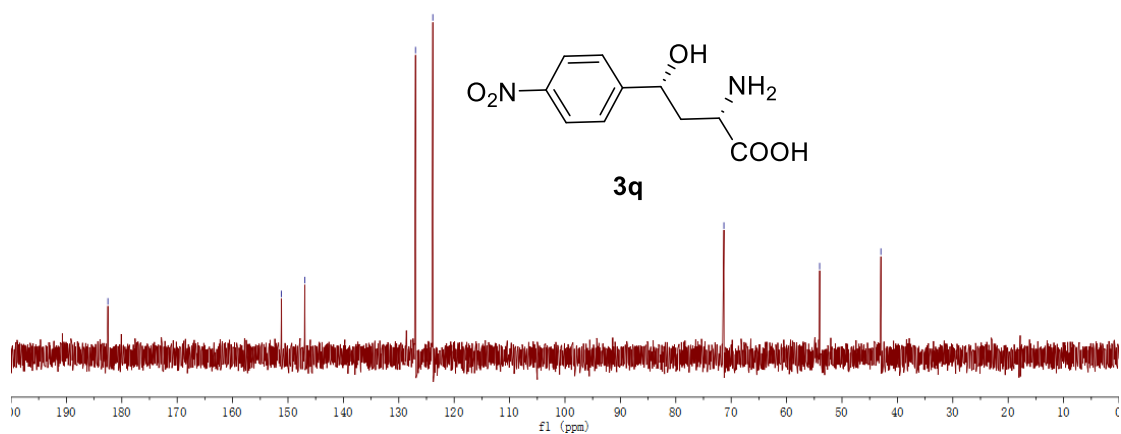
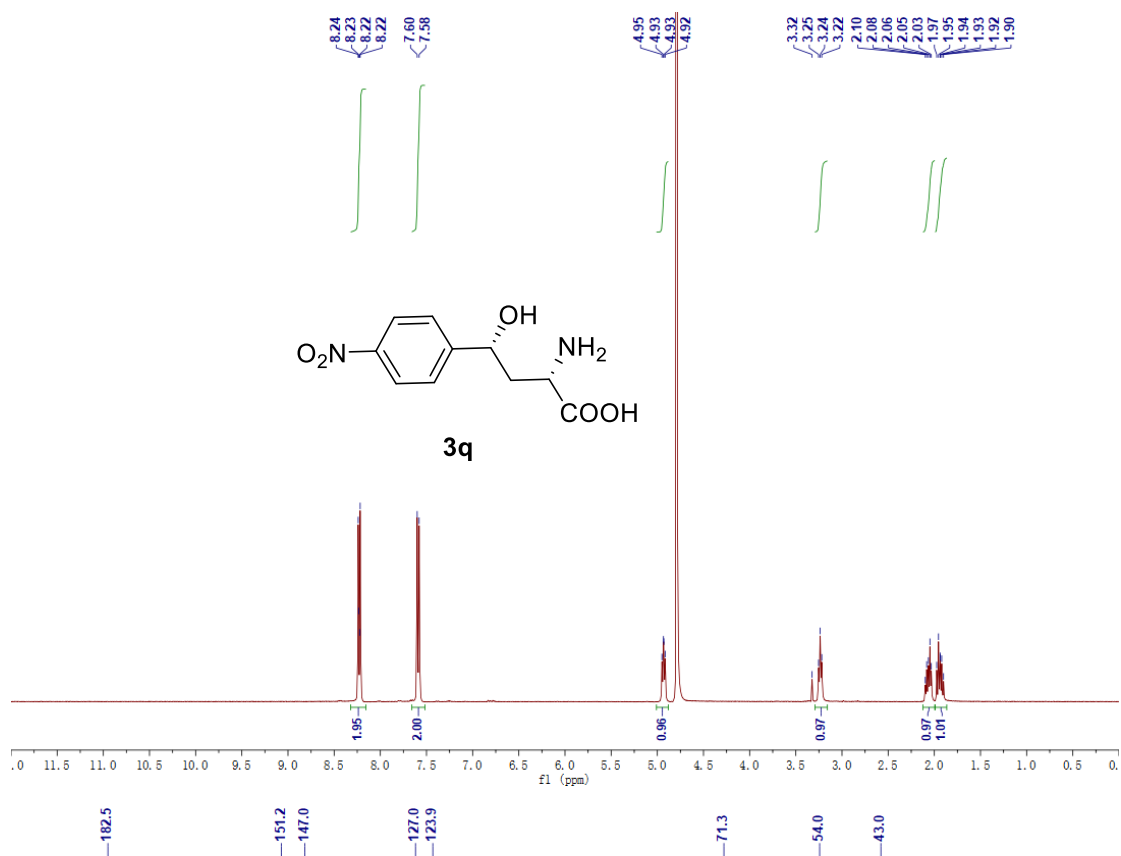


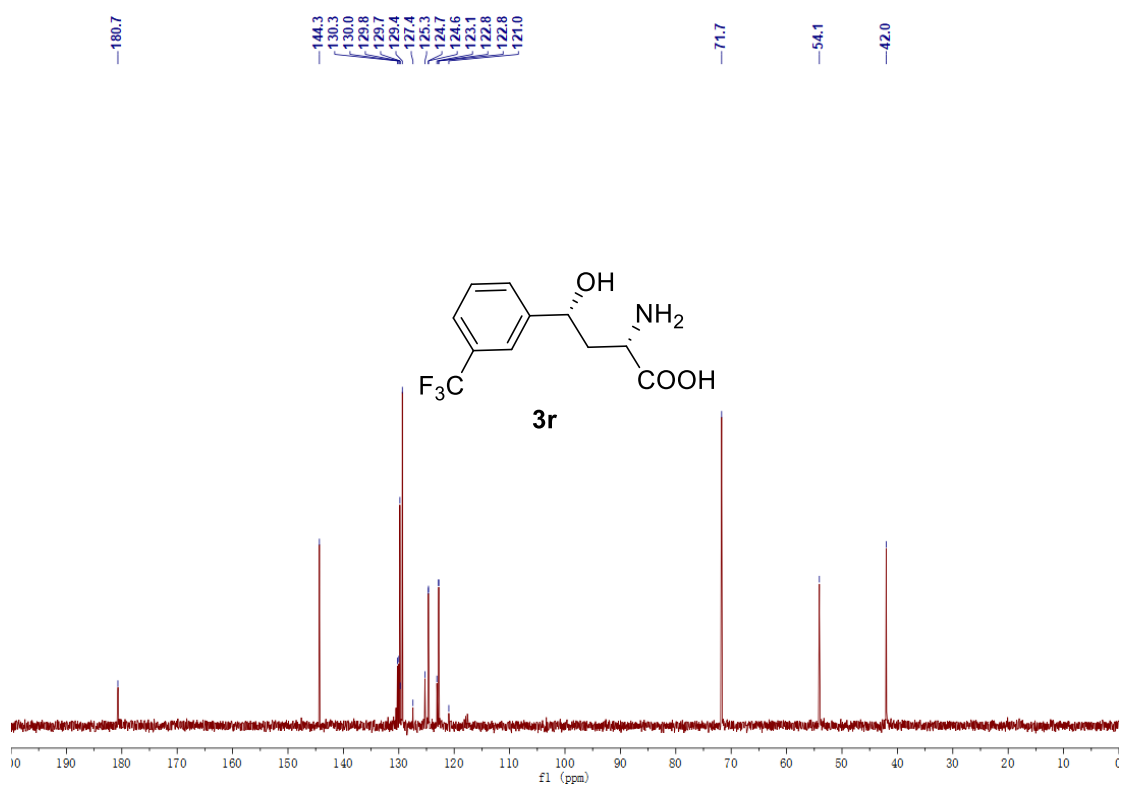
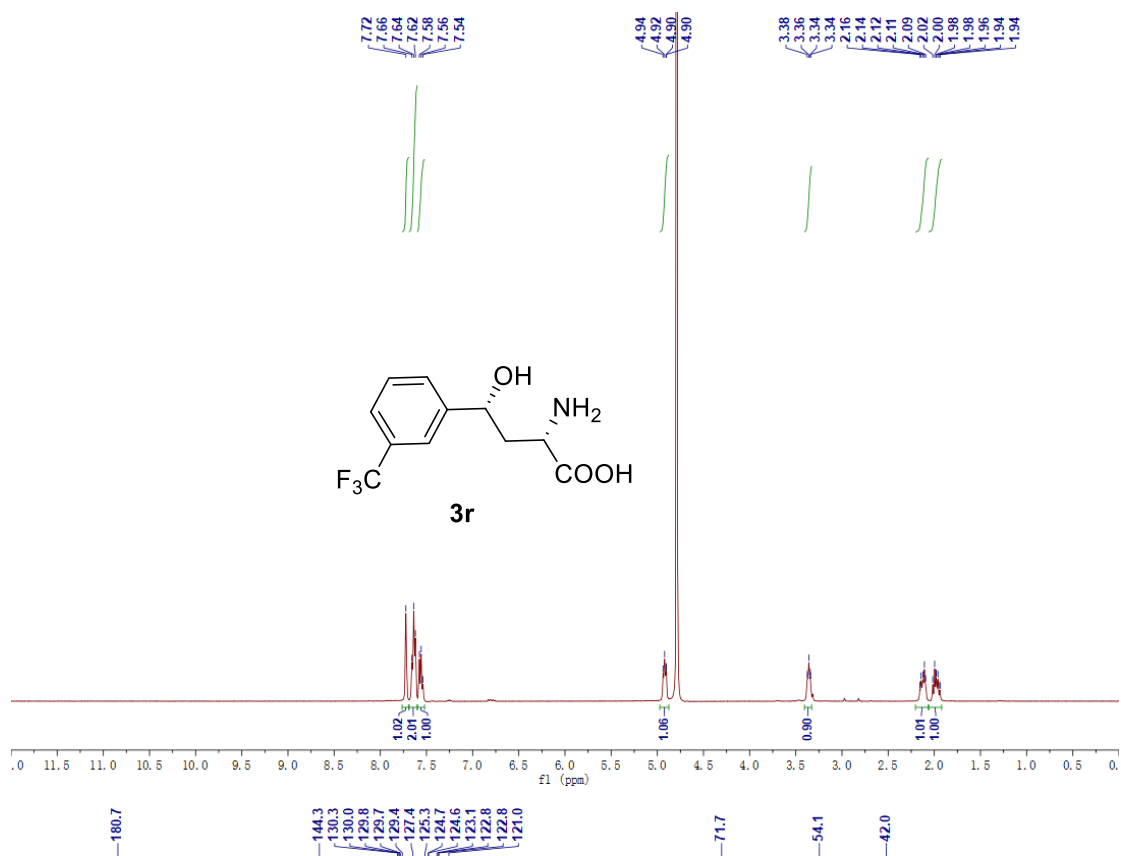


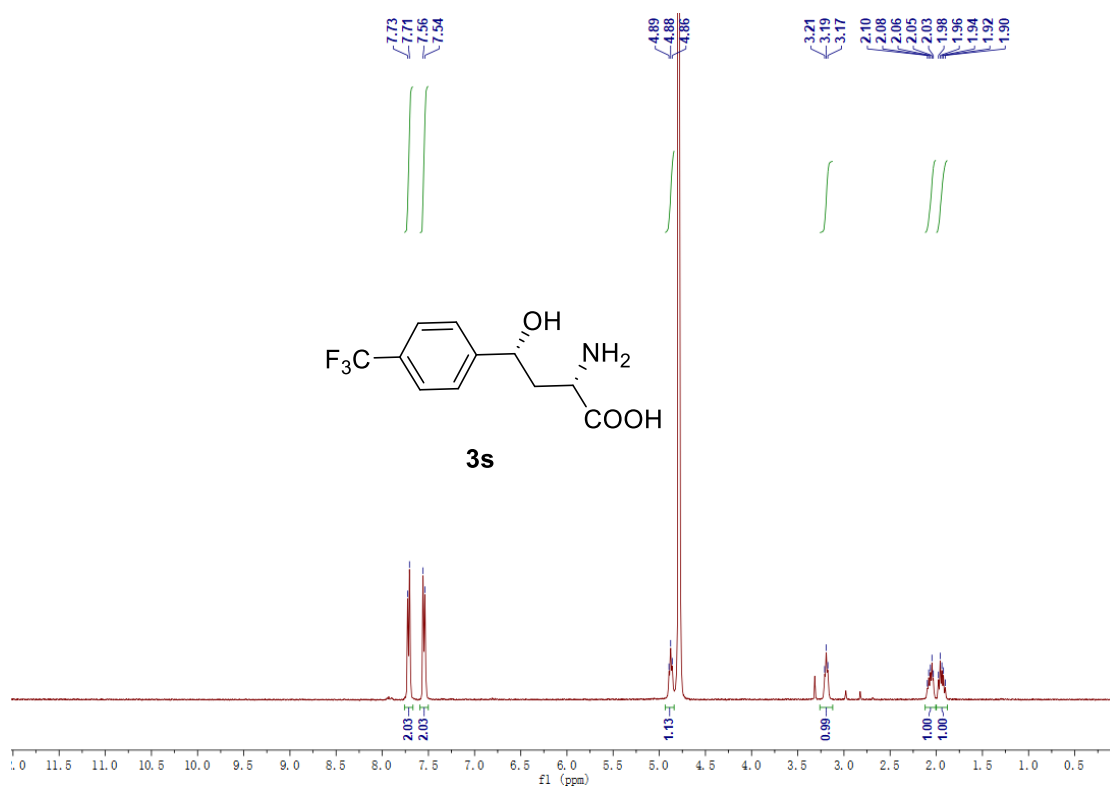
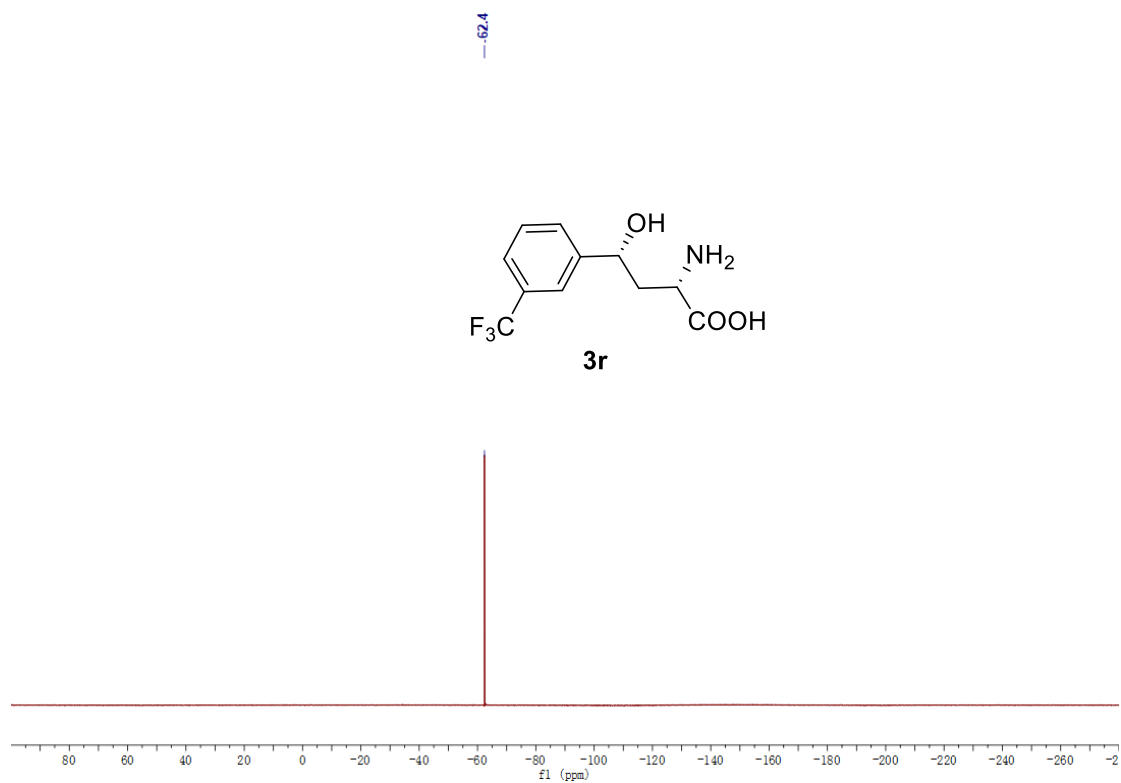


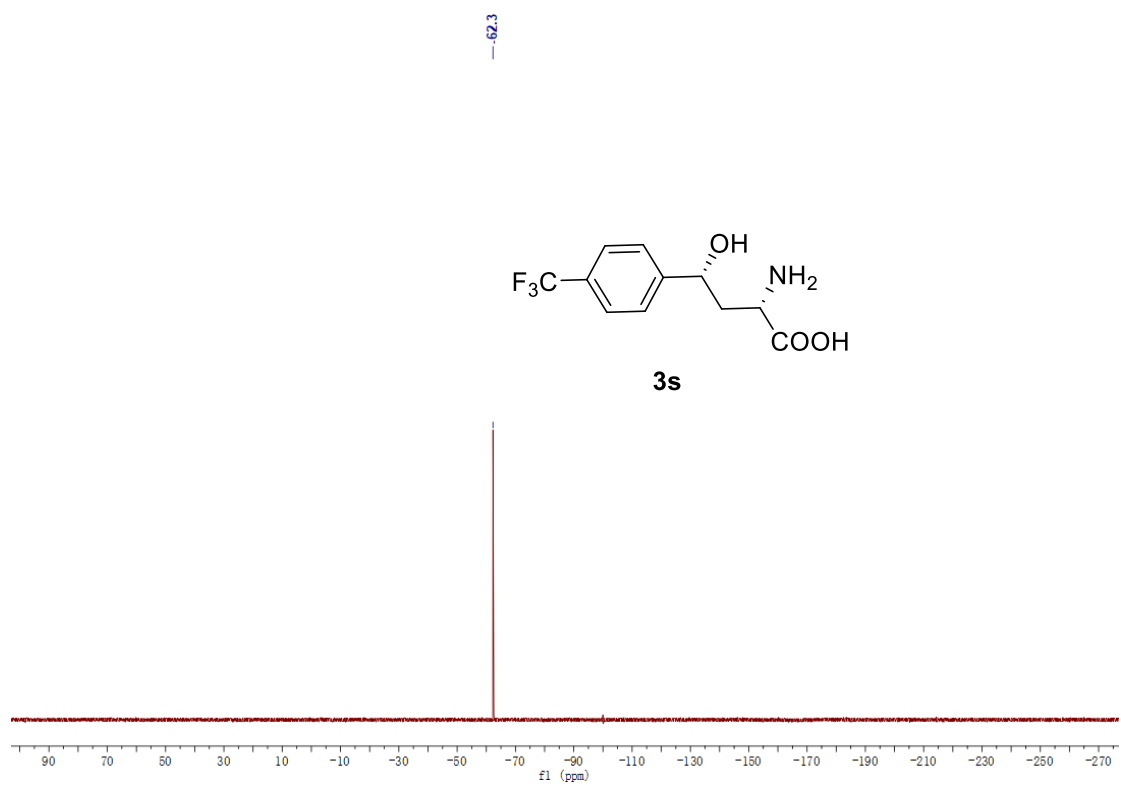
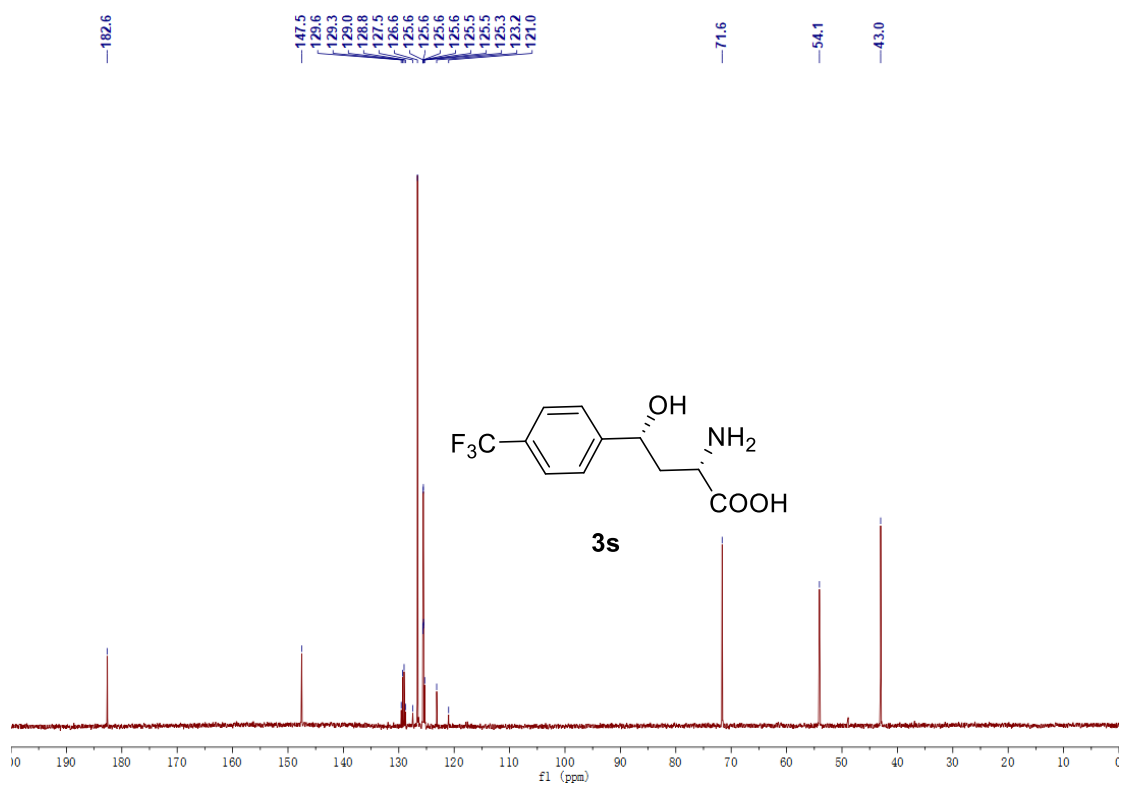


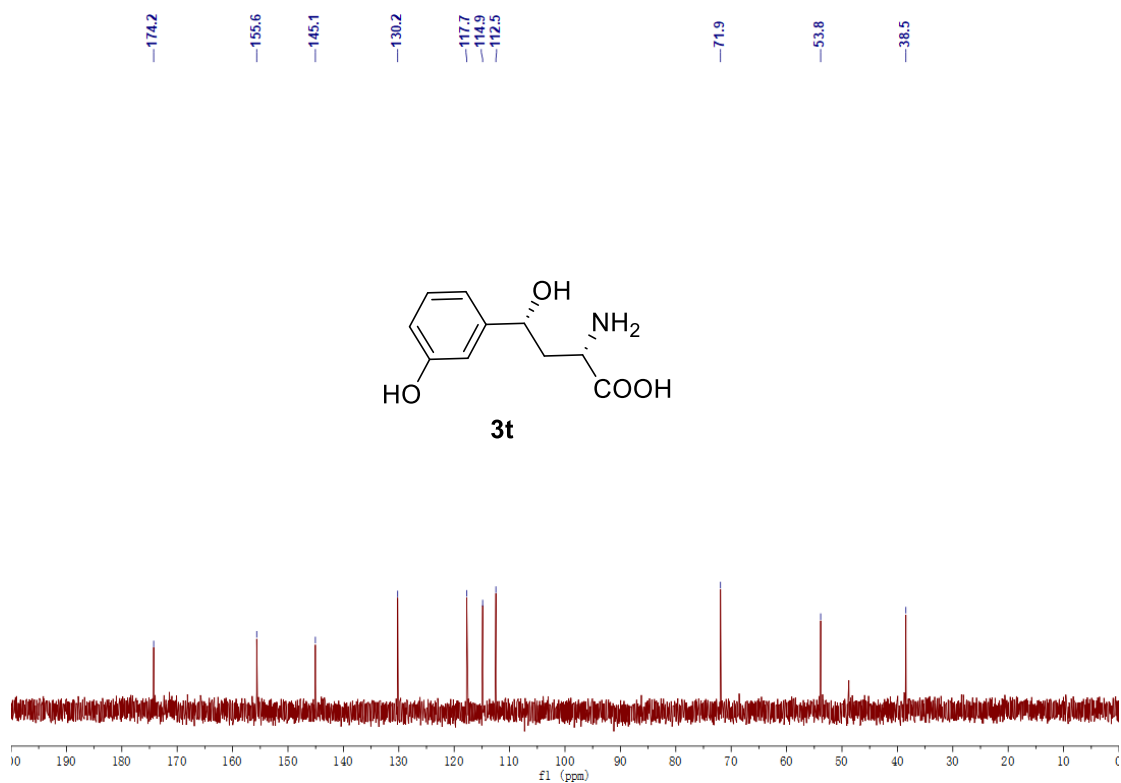
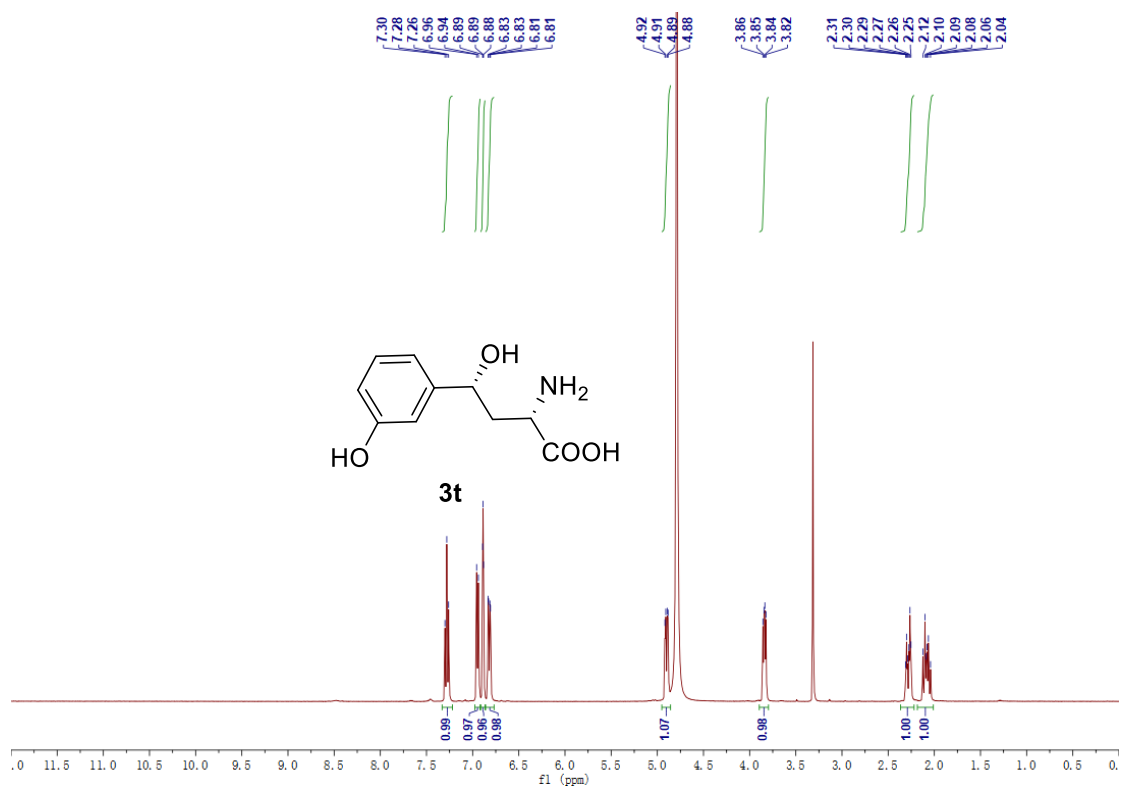


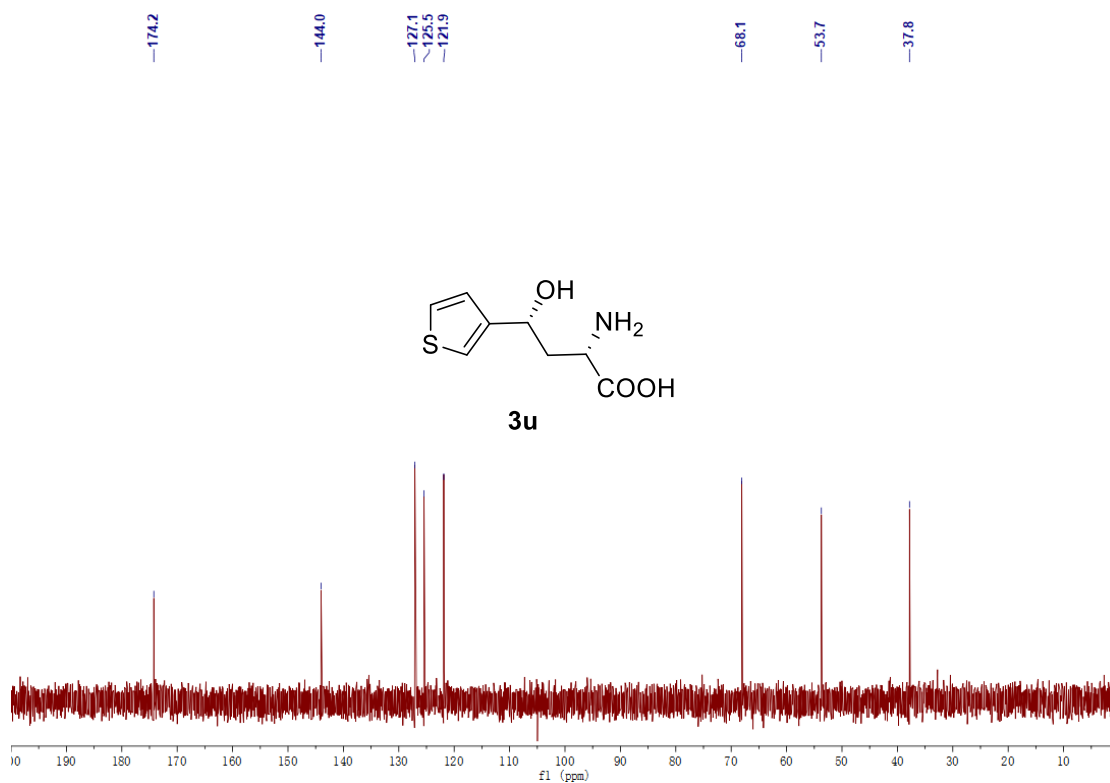
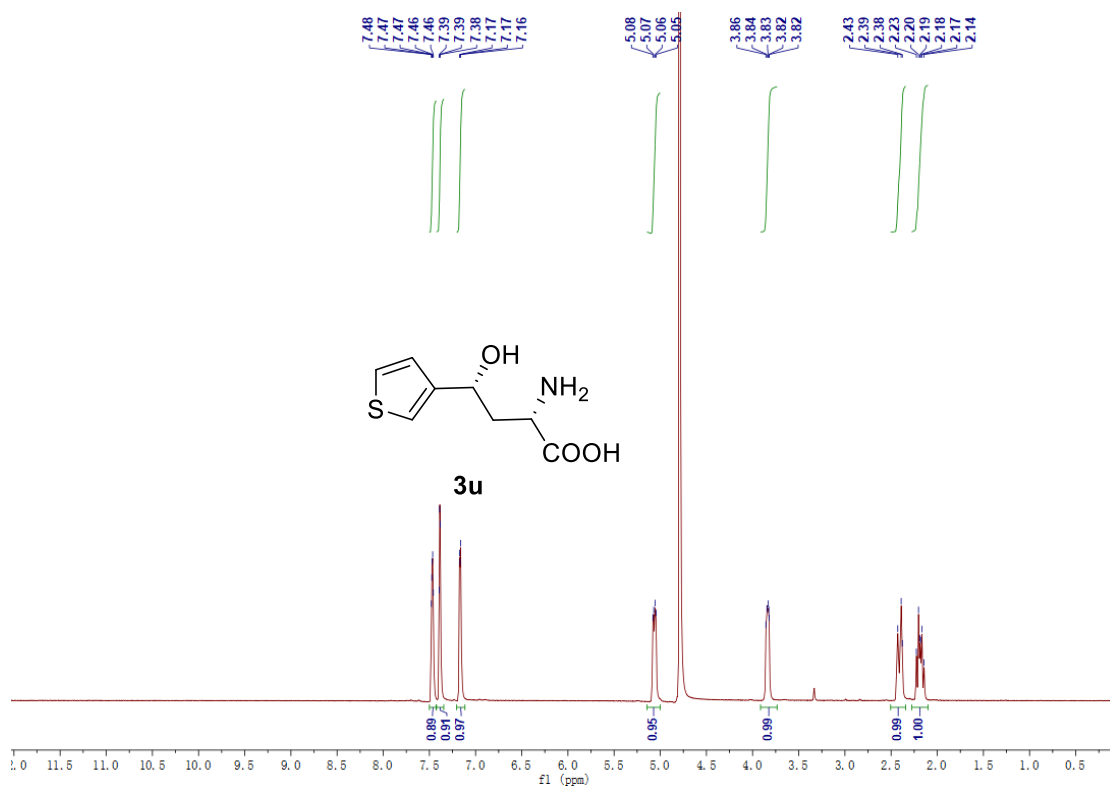


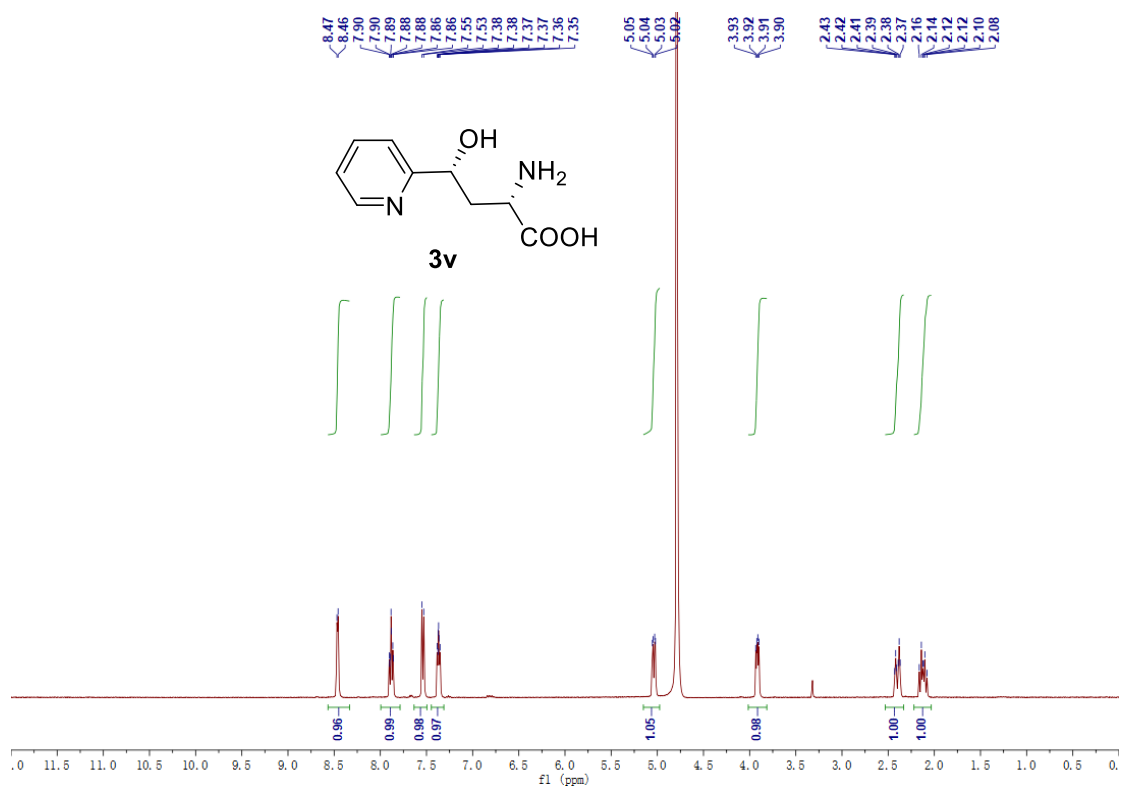












—174.2

—160.7

—148.2

—138.6

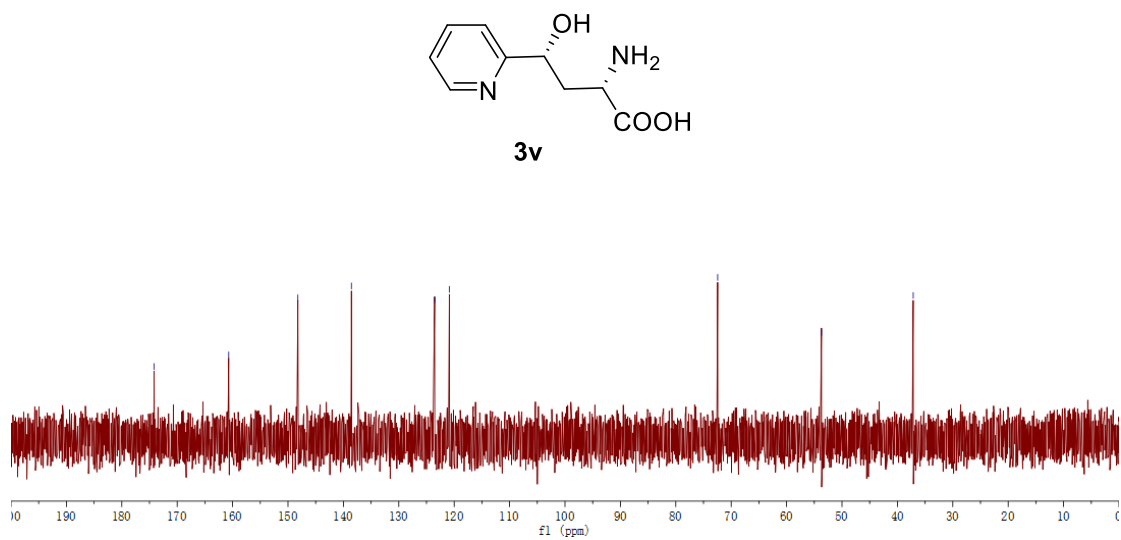
—123.5

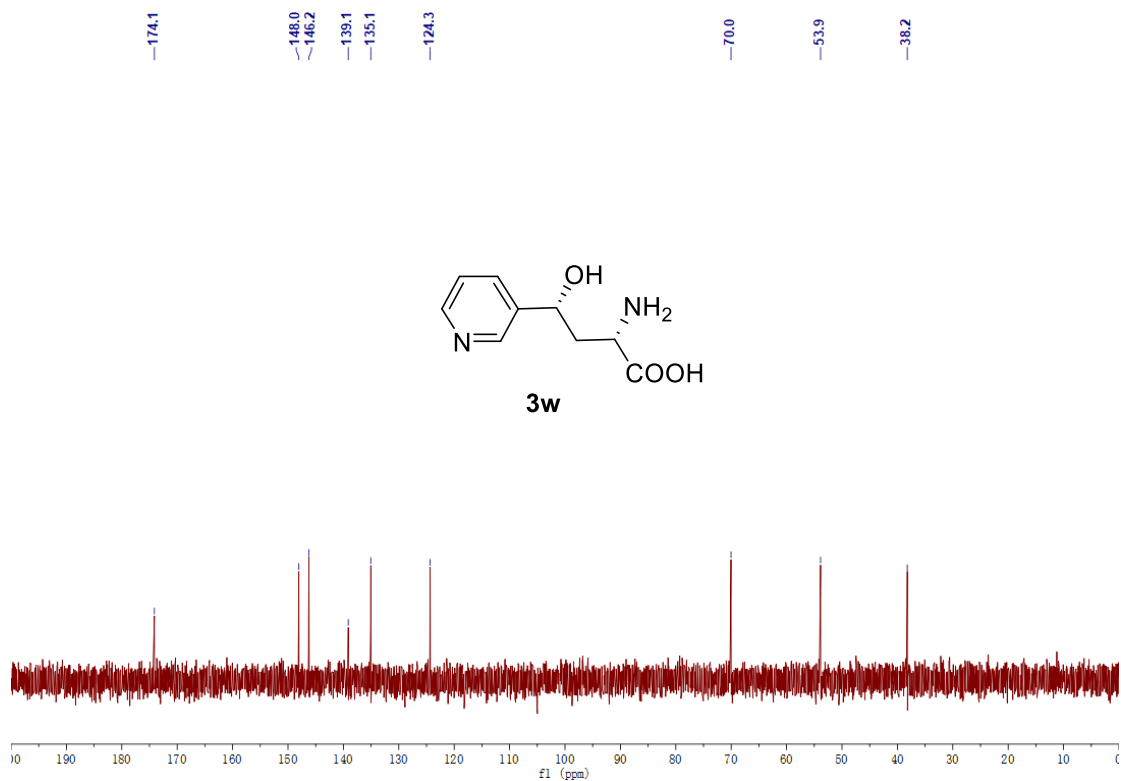
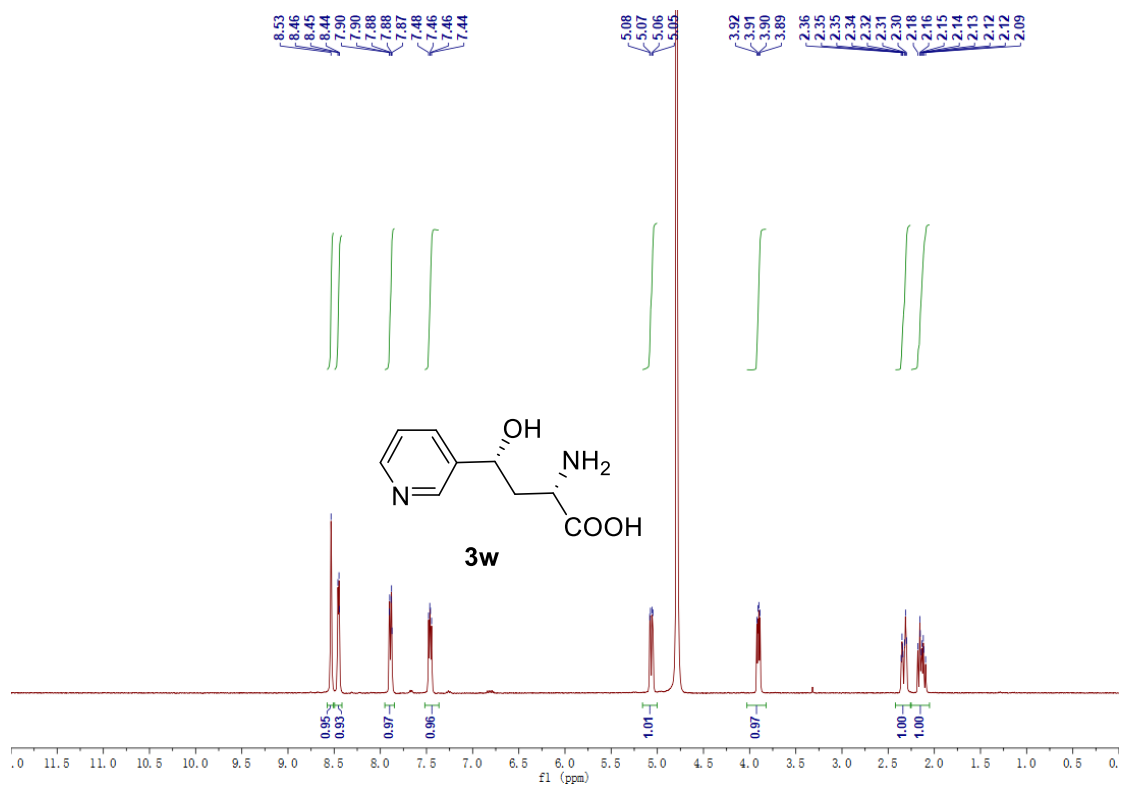
—120.9

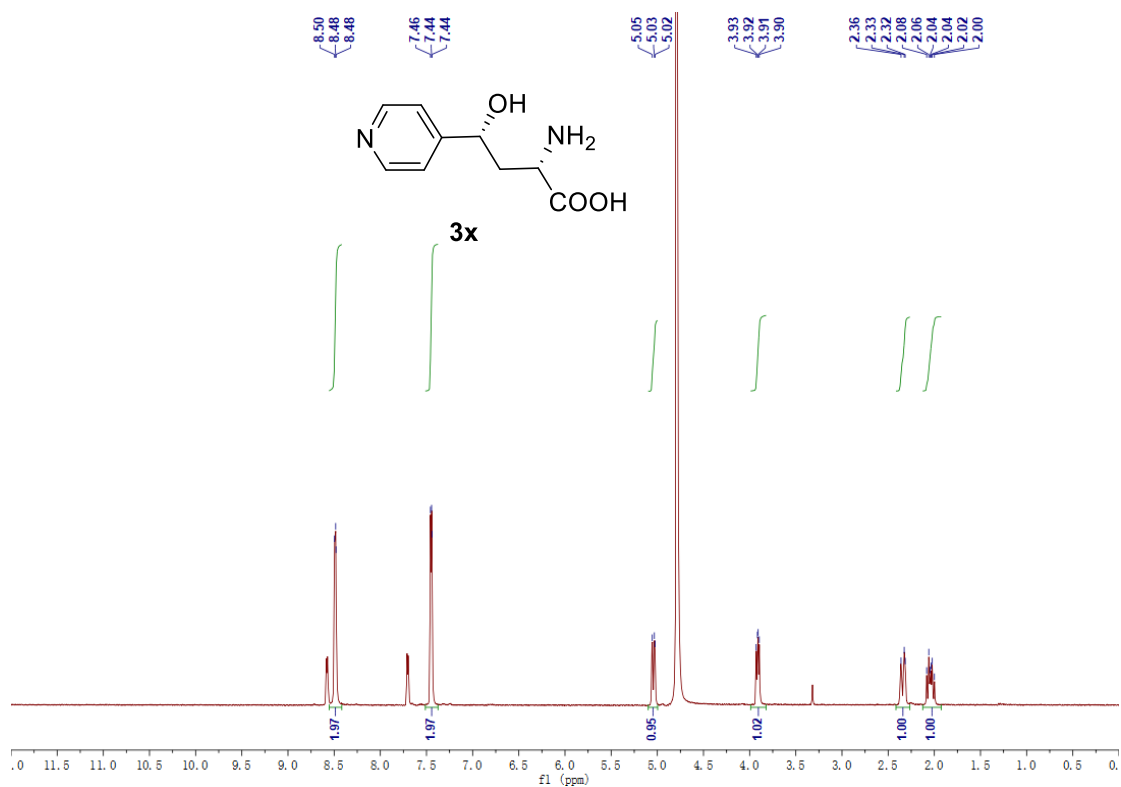
—72.4

—53.7

—37.1







—174.1

—153.5

—148.8

—121.1

—70.8

—53.8

—38.1

