Supplementary Information

Specific Lift-Up Behavior of Acetate-Intercalated Layered Yttrium Hydroxide Interlayer in Water: Application for Heterogeneous Brønsted Base Catalysts toward Knoevenagel Reactions

Takayoshi Hara*, Maoko Habe, Hikaru Nakanishi, Takuya Fujimura, Ryo Sasai, Chikako Moriyoshi, Shogo Kawaguchi, Nobuyuki Ichikuni, and Shogo Shimazu

General

All chemical compounds were purified by standard procedures before use.^{S1} X-Ray diffraction (XRD) measurements were performed on a Rigaku MiniFlex 600 using monochromatic Cu K α radiation ($\lambda = 0.15418$ nm), and the diffraction data were collected with D/teX Ultra 250 1D detector. The XRD was operated at 40 kV and 15 mA with divergence slit of 0.625°, scattering slit of 1.25°, and receiving slit of 0.30 mm, a step width of 0.02° and a scan speed of 10°/min. Catalytic activity was determined by gas chromatography with a flame ionization detector (FID) equipped with Shimadzu CPB-20 widebore-capillary column (25 m×0.53 mm×1 µm). GC analysis was also performed by GC-2010 with a flame ionization detector equipped with a Shimazu Rtx-5MS capillary column (30 m×0.25 mm×0.25 µm). Gas chromatograph-mass spectroscopy (GC-MS) was performed by Shimadzu GCMS-QP2010 Plus with a thermal conductivity detector equipped with a RESTEK Rtx®-5MS capillary column (30 m×0.25 mm×0.25 μm), and ionizing by electron impact. Products were confirmed by use of GC-MS, FT-IR, and ¹H and ¹³C NMR. ¹H and ¹³C NMR spectroscopy were performed using a JNM-AL400 spectrometer at 400 MHz in CDCl₃ with TMS as an internal standard. Fourier transform infrared (FT-IR) analysis was conducted using Shimadzu IRAffinity-1S, equipped with the diffuse reflectance measurement instrument DRS-8000. The sample was diluted with KBr powder and measured at room temperature. TG-DTA was performed using a Rigaku Thermo plus EVOII TG8120 and STAR System TGA/DSC 2 under air flow. Ion chromatography was performed by ICA-2000 (DKK-TOA) equipped with PCI-230 column (DKK-TOA) and chemical suppressor system (SeQuant® SAMSTM Standard Suppressor). 3.2 mM of Na₂CO₃ and 1.0 mM of NaHCO₃ was selected for the eluent. 10 mM of H₂SO₄ was also used for the eluent of chemical suppressor.

Gram-scale Knoevenagel reaction between ethylcyanoacetate (1) and benzaldehyde (2)

CH₃COO⁻/Y-LRH catalyst (0.05 g) was placed in a reaction vessel, followed by addition of **1** (10 mmol), **2** (10 mmol), acetonitrile (2 mL), and H₂O (8 mL). The reaction mixture was stirred at 333 K. After 3 h, CH₃COO⁻/Y-LRH was separated from the reaction mixture by a centrifugation. The supernatant was extracted with diethyl ether and then dried over MgSO₄. Removal of diethyl ether under the reduced pressure, followed by recrystallization with *n*-hexane and ethyl acetate, afforded analytically pure (2*E*)-2-

cyano-3-phenyl- 2-propenoic acid ethyl ester (**3**) as a white powder (93% isolated yield): CAS registry number [2025-40-3]; mp 52 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.45 (t, 3H, J = 7.1 Hz), 4.42 (q, 2H, J = 7.1 Hz), 7.48-7.64 (m, 3H), 8.02 (m, 2H), 7.98-8.08 (s, 1H); ¹³C NMR (105 MHz, CDCl₃) δ 14.2, 62.9, 103.7, 115.3, 128.3, 130.3, 132.3, 132.7, 153.6, 162.3; IR (KBr) 1200, 1610, 2240 cm⁻¹.

Recycling test

After the Knoevenagel reaction, acetonitrile (3 mL) was added to the reaction mixture, then, all of the reaction mixture was transferred into 10 mL of sample tube. The liquid phase and the solid catalyst were separated by simple centrifugation (3500 rpm for 5 min). By use of the same procedure, the solid catalyst was washed with 3 mL of acetonitrile 3-5 times and dried under vacuum overnight. For the next catalytic reaction, the water solvent (5 mL) was added to this sample tube, and all of the aqueous dispersion containing the recovered catalyst was moved into Schlenk tube.

References

S1) W. L. F. Armarego and C. L. L. Chai, *Purification of Laboratory Chemicals*, Elsevier, 2003.





Fig. S1 Thermogravimetric-differential thermal analysis (TG-DTA) for (a) Cl⁻/Y-LRH, (b) NO₃⁻/Y-LRH, (c) HCOO⁻/Y-LRH, (d) CH₃COO⁻/Y-LRH, (e) C₂H₅COO⁻/Y-LRH, (f) C₃H₇COO⁻/Y-LRH, (g) C₅H₁₁COO⁻/Y-LRH, (h) C₇H₁₅COO⁻/Y-LRH, and (i) C₉H₁₉COO⁻/Y-LRH catalyst.

Black curve: TG line, blue line: temperature, and red curve: DTA line.

Entry	Anion	n	Y ³⁺ amount in 0.05 g (10 ⁻² mmol)	
1	CF	1.43	7.71	
2	NO ₃ ⁻	1.60	7.07	
3	HCOO-	1.35	7.53	
4	CH ₃ COO ⁻	1.40	7.20	
5	C ₂ H ₅ COO ⁻	1.56	6.87	
6	C ₃ H ₇ COO ⁻	1.30	6.70	
7	C ₅ H ₁₁ COO ⁻	1.44	6.41	
8	C7H15COO	1.22	6.25	
9	C ₉ H ₁₉ COO ⁻	1.04	6.09	

Table S1. The *n* value and Y^{3+} amount in various Anion/Y-LRH, $Y_2(OH)_5(Anion) \cdot nH_2O$

^a Determined by TG-DTA and ion-chromatography



Fig. S2 FT-IR spectra for (a) HCOO⁻/Y-LRH, (b) CH₃COO⁻/Y-LRH, (c) C₂H₅COO⁻/Y-LRH, (d) C₃H₇COO⁻/Y-LRH, (e) C₅H₁₁COO⁻/Y-LRH, (f) C₇H₁₅COO⁻/Y-LRH, and (g) C₉H₁₉COO⁻/Y-LRH catalyst.



Fig. S3 (A) 2D mapping plots of the *t*-SXRD patterns of Y-LRH during anion-exchange reaction from Cl^{-} into $CH_{3}COO^{-}$. (B) Time profile of the disappeared peak area at 5.51° in *t*-SXRD. (C) The peak position and FWHM values of newly appeared peak at 3.45°.



Fig. S4 2D mapping plots of the *t*-SXRD patterns of Cl^{-}/Y -LRH. After 5 sec, 2 mL of water was added in the slurry of Cl^{-}/Y -LRH (0.05 g) and water (0.5 mL).



Fig. S5 (A) 2D mapping plots of the *t*-SXRD patterns of Y-LRH during anion-exchange reaction from NO_3^- into CH₃COO⁻. (B) Time profile of the newly appeared peak area at 3.43° in *t*-SXRD. (C) Time profile of the disappeared peak area at 5.10° in *t*-SXRD. (D) The peak position and FWHM values of newly appeared peak at 3.43°.



Fig. S6 XRD profiles for (a) NO₃⁻/Y-LRH and (b) Cl⁻/Y-LRH catalyst



Fig. S7 XRD profiles for (a) Cl⁻/Y-LRH, (b) HCOO⁻/Y-LRH, (c) CH₃COO⁻/Y-LRH, (d) $C_2H_5COO^-/Y$ -LRH, (e) $C_3H_7COO^-/Y$ -LRH, (f) $C_5H_{11}COO^-/Y$ -LRH, (g) $C_7H_{15}COO^-/Y$ -LRH, and (h) $C_9H_{19}COO^-/Y$ -LRH catalyst.



Fig. S8 (A) Photoimages of CH₃COO⁻/Y-LRH catalyst: (A) during reaction and (B) after centrifugation. (C) Hot filtration experiment. reaction conditions were as follows: CH₃COO⁻/Y-LRH catalyst (0.05 g), 1 (1.5 mmol), 2 (1 mmol), H₂O (5 mL), 303 K. Yield was based on 2, determined by GC.



Fig. S9 XRD profiles for (a) fresh CH₃COO⁻/Y-LRH in dry state and (b) recovered CH₃COO⁻/Y-LRH in dry state. Values in graph were d_{00l} space, calculated by the Bragg equation



Fig. S10 XRD profiles for (a) dry CH₃COO⁻/Gd-LRH, (b) wet CH₃COO⁻/Gd-LRH, (c) dry CH₃COO⁻/Yb-LRH, and (d) wet CH₃COO⁻/Yb-LRH. Values in graph were d_{00l} space, calculated by the Bragg equation.



Fig. S11 (A) 2D mapping plots of the *t*-SXRD patterns of CH_3COO^-/Y -LRH during the interlayer lift-up process. Arrows indicated the start point of addition of water into solid dispersion in acetonitrile solution. (B) Time profile of the newly appeared peak area at 3.47° in *t*-SXRD. (C) Time profile of the disappeared peak area at 4.70° in *t*-SXRD. (D) The peak position and FWHM values of newly appeared peak at 3.47°.



Fig. S12 XRD profiles for (a) dry Cl⁻/Y-LRH, (b) wet Cl⁻/Y-LRH, (c) dry NO₃⁻/Y-LRH, (d) wet NO₃⁻/Y-LRH, (e) dry HCOO⁻/Y-LRH, and (f) wet HCOO⁻/Y-LRH,

T (K)	T ⁻¹ (10 ⁻³ K ⁻¹)	<i>k</i> (M ⁻¹ ·min ⁻¹)	ln <i>k</i> (-)
303	3.300	0.2547	-1.3677
313	3.195	0.3220	-1.1332
323	3.096	0.3947	-0.9296
333	3.003	0.4987	-0.7160
343	2.915	0.5947	-0.5197

Table S2. Kinetic rate constants for the Knoevenagel condensation with CH₃COO⁻/Y-LRH catalyst^a

^a CH₃COO⁻/Y-LRH catalyst (0.01 g), **1** (1 mmol), **2** (1 mmol), H₂O (5 mL), 5 min.



Fig. S13 XRD profiles for (a) dry $C_2H_5COO^-/Y$ -LRH, (b) wet $C_2H_5COO^-/Y$ -LRH, (c) dry $C_3H_7COO^-/Y$ -LRH and (d) wet $C_3H_7COO^-/Y$ -LRH catalyst

Ethyl (*E*)-2-cyano-3-phenyl-2-propenoate (CAS: 2169-69-9) A white solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.41 (t, *J* = 7.2 Hz, 3H), 4.39 (q, *J* = 7.2 Hz, 2H), 7.49-7.57 (m, 3H), 8.00 (d, *J* = 7.2 Hz, 2H), 8.26 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.1, 62.7, 103.0, 115.5, 129.3, 131.0, 131.5, 133.3, 155.0, 162.5.



Ethyl (*E*)-2-cyano-3-(4-chlorophenyl)-2-propenoate (CAS: 2169-68-8)

A white solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.40 (t, J = 7.2 Hz, 3H), 4.39 (q, J = 7.2 Hz, 2H), 7.48 (d, J = 8.8 Hz, 2H), 7.94 (d, J = 8.4 Hz, 2H), 8.20 (s, 1H).

¹³C NMR (100 MHz, CDCl₃) *δ* ppm: 14.1, 62.6, 103.5, 115.2, 129.7, 129.8, 132.2, 139.6, 153.4, 162.2.





Ethyl (E)-2-cyano-3-(4-methylphenyl)-2-propenoate (CAS: 2017-88-1)

A white solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.40 (t, J = 7.6Hz, 3H), 2.44 (s, 3H), 4.38 (q, J = 7.6 Hz, 2H), 7.31 (d, J = 8.0 Hz, ^{H₃C CN}

¹³C NMR (100 MHz, CDCl₃) *δ* ppm: 14.2, 21.8, 62.6, 101.5, 115.8, 128.8, 130.0, 131.2, 144.6, 155.0, 162.8.





Ethyl (E)-2-cyano-3-(4-methoxyphenyl)-2-propenoate (CAS: 2017-87-0)

A yellow crystalline solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.39 (t, *J* = 8.0 Hz, 3H), 3.90 (s, 3H), 4.37 (q, *J* = 7.2 Hz, 2H), 7.00 (d, *J* H_3CO H_3CO

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.2, 55.6, 62.4, 99.3, 114.7, 116.2, 124.3, 133.6, 154.4, 163.1, 163.7.





Ethyl (*E*)-2-cyano-3-(4-hydroxyphenyl)-2-propenoate (CAS: 42205-38-9)

A yellow crystalline solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.39 (t, *J* = 7.2 Hz, 3H), 4.38 (q, *J* = 7.2 Hz, 2H), 6.98 (d, *J* = 9.2 Hz, 2H), HO (N) 7.96 (d, *J* = 8.4 Hz, 2H), 8.19 (s, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: □14.2, 62.6, 99.2, 116.2, 116.4, 124.3, 133.9, 154.9, 160.7.





Ethyl (E)-2-cyano-3-(1-naphthyl)-2-propenoate (CAS: 29708-01-8)

A light yellow needle. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.42 (t, *J* = 7.2 Hz, 3H), 4.41 (q, *J* = 6.8 Hz, 2H), 7.55-7.64 (m, 2H), 7.87-7.95 (m, 3H), 8.17-8.20 (m, 1H), 8.37-8.38 (m, 1H), 8.39 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.2, 62.7, 102.6, 115.7, 125.2, 127.2, 127.8, 129.0,





Ethyl (*E*)-2-cyano-3-(2-thienyl)-2-propenoate (CAS: 62309-97-1)

A pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.39 (t, J = 7.2Hz, 3H), 4.37 (q, J = 6.8 Hz, 2H), 7.22-7.24 (m, 1H), 7.79 (d, J = 4.8 Hz, 1H), 7.84 (d, J = 3.6 Hz, 1H), 8.35 (s, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.2, 62.5, 99.2, 115.7, 128.6, 135.1, 136.0, 137.2, 146.7, 162.7.



Ethyl (E)-2-cyano-3-(2-furanyl)-2-propenoate (CAS: 67449-75-6)

An orange pillar. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.39 (t, J = 7.6 Hz, 3H), 4.36 (q, J = 7.2 Hz, 2H), 6.66-6.68 (m, 1H), 7.40 (d, J = 3.6 Hz, 1H), 7.77 (d, J = 1.6 Hz, 1H), 8.02 (s, 1H);

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.1, 62.5, 98.6, 113.8, 115.3, 121.7, 139.4, 148.2, 148.7, 162.5.



Ethyl (E)-2-cyano-3-(pyridin-2-yl)2-propenoate (CAS: 68752-87-4)

A yellow solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.41 (t, J = 7.2 Hz, 3H), 4.41 (q, J = 7.2 Hz, 2H), 7.41-7.44 (m, 1H), 7.84-7.91 (m, 2H), 8.30 (s, 1H), 8.83 (d, J = 4.0 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.1, 62.9, 106.8, 114.7, 126.2, 126.5, 136.9, 150.0, 150.5, 153.4, 162.0.





Ethyl 2-cyano-5-phenyl-(2E, 4E)-pentadienoate (CAS: 41109-95-9)

A pale yellow pillar. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.38 (t, J = 7.2 Hz, 3H), 4.34 (q, J = 7.2 Hz, 2H), 7.26-7.29 (m, 2H), 7.41-7.44 (m, 3H), 7.58-7.60 (m, 2H), 7.99-8.02 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.1, 62.3, 104.4, 114.5, 123.0, 128.5, 129.1, 131.1, 134.6, 148.8, 155.4, 162.3.





Ethyl (*E*)-2-cyano-2-decenoate (CAS: 879496-08-9)

A colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 0.89 (t, J = 6.8 Hz, 3H), 1.24-1.39 (m, 11H), 1.52-1.59 (m, 2H), 2.56 (q, J = 7.6 Hz, 2H), 4.32 (q, J = 7.2 Hz, 2H), 7.66 (t, J = 8.0 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.0, 14.0, 22.5, 27.7, 28.8, 29.1, 31.5, 31.9, 62.3, 109.7, 113.7, 161.3, 163.9.



Ethyl (*E*)-2-cyano-3-cyclohexylacrylate (CAS: 90913-43-2) A colorless liquid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 1.18-1.42 (m, 8H), 1.68-1.82 (m, 5H), 2.67-2.76 (m, 1H), 4.31 (q, *J* = 6.8 Hz, 2H), 7.48 (d, *J* = 10.8 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 14.0, 24.7, 25.3, 31.0, 41.0, 62.3, 107.5, 113.7, 161.5, 167.8.



(*E*)-2-Cyano-3-phenyl-2-propenamide (CAS: 15795-18-3)

A white solid. ¹H NMR (400 MHz, CDCl₃) δ ppm: 6.40 (d, J = 15.6 Hz, 2H), 7.49-7.58 (m, 3H), 7.94-7.98 (m, 2H), 8.35 (s, 1H).

¹³C NMR (100 MHz, CDCl₃) δ ppm: 103.1, 117.0, 129.3, 130.8, 131.5, 133.1, 154.0, 162.1.

