Coordination of Trivalent Lanthanum and Cerium, and Tetravalent Cerium and Actinides (An = Th(IV), U(IV), Np(IV)) by a 4-Phosphoryl 1H-Pyrazol-5olate Ligand in Solution and the Solid State

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Experimental details

General Considerations: If not stated differently, all manipulations were performed with HPLC grade, analytical grade or technical grade reagents and solvents, which were used without further purification. The used starting materials were purchased from *SIGMA-ALDRICH*, *FLUKA*, *MERCK*, *VWR*, *TCI*, *ABCR CHEMICALS*, *ACROS*, *CARL ROTH* or *WAKO*.

Manipulations under dry, oxygen-free conditions were performed in a Glovebox MB Unilab or using *Schlenk* technique under an atmosphere of purified nitrogen. All glassware was oven-dried at 160°C prior to use. Dry, oxygen-free solvents (CH₂Cl₂, CH₃CN, C₆F₆ (distilled from CaH₂), toluene, Et₂O (distilled from potassium)) were employed. Anhydrous deuterated acetonitrile (CD₃CN), dichloromethane (CD₂Cl₂), chloroform (CDCl₃) and methanol (CD₃OD) were purchased from Sigma-Aldrich or Deutero. All distilled and deuterated solvents were stored over molecular sieves (4 Å: CH₂Cl₂, CD₂Cl₂, CDCl₃, CD₃OD, toluene, Et₂O, C₆F₆; 3 Å: CH₃CN, CD₃CN)). Dry, oxygen-free CH₂Cl₂ was obtained by distillation from CaH₂. The obtained anhydrous solvents were stored over 4 Å molecular sieves.

NMR spectra were measured on a Bruker AVANCE III HD Nanobay, 400 MHz UltraSield (¹H (400.13 MHz), ¹³C (100.61 MHz), ³¹P (161.98 MHz)) or on a Bruker AVANCE III HDX, 500 MHz Ascend (¹H (500.13 MHz), ¹³C (125.75 MHz), ³¹P (202.45 MHz)), or on a Varian Inova 400, 400 MHz Oxford magnet (¹H (399,89 MHz), ¹³C (100.56 MHz), ³¹P (161.87 MHz)) installed in a radioactive controlled laboratory. All ¹³C NMR spectra were exclusively recorded with composite pulse decoupling. Reported numbers assigning atoms in the ¹³C spectra were indirectly deduced from the cross-peaks in 2D correlation experiments (HMBC, HSQC). Chemical shifts were referenced the respective solvent to $\delta = 7.26$ ppm (¹H), 77.16 ppm (¹³C) for CDCl₃, $\delta = 5.32$ ppm (¹H), 53.84 ppm (¹³C) for CD₂Cl₂, $\delta = 3.31$ ppm (¹H), 49.00 ppm (¹³C) for CD₃OD)¹ and $\delta_{\text{H3PO4(85\%)}} = 0.00$ ppm (³¹P, externally). Chemical shifts (δ) are reported in ppm. Coupling constants (J) are reported in Hz. Infrared (IR) and Raman spectra were recorded at ambient temperature using a Bruker Vertex 70 instrument equipped with a RAM II module (Nd-YAG laser, 1064 nm). The Raman intensities are reported in percent relative to the most intense peak and are given in parenthesis. An ATR unit (diamond) was used for recording IR spectra. The intensities are reported relative to the most intense peak and are given in parenthesis using the following abbreviations: vw = very weak, w = weak, m = medium, s = strong, vs = very strong. Elemental analyses were performed on a Vario MICRO cube Elemental Analyzer by Elementar Analysatorsysteme GmbH in CHNS modus. For the mass spectrometry experiments a waters ACQUITY UPLC H-Class system in combination with an ACQUITY TQ Detector V4.1 SCN849 SCN896 was used. MassLynx V4.1 SCN849 SCN896 served as evaluation software. The required ionization was provided by the electrospray method (ESI). As diluent for the samples served an acetonitrile water mixture containing both solvents in a 70:30 (acetonitrile: water) ratio and additionally 0.1% of formic acid.

Synthesis of Diisopropyl (5-hydroxy-3-methyl-1-(p-tolyl)-1H-pyrazol-4-yl)phosphonate (HL)

The 4-phosphoryl pyrazolone ligand HL has been synthesised in 3 steps by adoption of procedure reported in the literature (Scheme S1).²⁻⁴



Scheme S1: Synthesis of the 4-phosphoryl pyrazolone HL; i) 1.05 eq methyl bromoacetate, 120 °C, neat, 4 h 98% with a purity of 92%; ii) 1.05 eq. MgCl₂, 3.5 eq. Et₃N, 1.95 eq. acetyl chloride, CH_2Cl_2 , 20°C, 66%; iii) 1.1 eq. *p*-tolylhydrazine hydrochloride, 2.0 eq. K₂CO₃, H₂O, 2 h reflux, r.t. 12 h, 67% in a 91% purity; II: purification of HL *via* **3** and subsequent back extraction; i) LaCl₃·7H₂O, NaOH, CH₃CN/water, 96%; ii) CHCl₃/H₂O, HCl (0.05 M), 99%.

Diisopropyl (5-hydroxy-3-methyl-1-(p-tolyl)-1H-pyrazol-4-yl)phosphonate (HL):



The ligand was synthesized according to a procedure reported by Modranka and coworkers.² To a suspension of 2.76 g, 8.25 mmol of methyl 2-(diisopropoxyphosphoryl)-3-oxobutanoate (**2**) in water (15 mL), 1.1 eq (1.47 g; 9.08 mmol in 18 mL H₂O) of *p*-tolylhydrazine hydrochloride (98%) were added. The suspension was heated up to 100-110 °C and refluxed for 2 h to yield a brown oil in a yellowish solution. After cooling down to r.t. 2.28 g (16.51 mmol, 2 eq) of potassium carbonate were slowly added. The resulting suspension was heated again to reflux for another 2 h. After cooling down the reaction mixture was stirred at r.t. overnight. The resulting mixture was transferred into a separation funnel and washed with Et₂O (3 x 20 ml). The combined aqueous phase

were collected and acidified with a 0.5 M HCl to pH = 2, affording a pale yellow oily suspension, which was extracted with EtOAc (3 x 20 ml). The combined organic layers were washed with brine, dried over Na₂SO₄ and the solvent was evaporated under reduced pressure to yield the product as a brown oil in 91% purity and a yield of 2.14 g, 67%. All classical methods to purify this ligand, including chromatography and recrystallization, failed. However, HL readily forms the stable La(III) complex [LaL₃HL]·3CH₃CH·3H₂O (**3**·3CH₃CH·3H₂O) upon reaction with LaCl₃·7H₂O at 80°C in an aqueous CH₃CN solution in the presence of NaOH (Scheme S1). Therefore, the reaction mixture stirred for 1 h at 80 °C, cooled down to r.t. and put in the fridge for another 2 h. The light yellow powder was collected by filtration giving **3**·3CH₃CH·3H₂O in 96 % yield. The extraction of a CHCl₃ solution of the complex **3**·3CH₃CH·3H₂O with 0.05 M HCl (pH = 1.3) enables the stripping of La(III) into the aqueous phase, releasing HL after removing the solvent in high purity > 99%.

Raman (255 mW, in cm⁻¹): *v* = 3083 (27), 2983 (51), 2923 (100), 2871 (47), 2734 (31), 1616 (78), 1579 (22), 1570 (22), 1531 (28), 1516 (29), 1452 (35), 1427 (20), 1402 (25), 1384 (22), 1369 (29), 1346 (28), 1315 (22), 1292 (20), 1215 (18), 1184 (17), 1055 (24), 842 (24), 785 (13), 684 (16), 77 (49); **IR (ATR, in cm⁻¹)**: *v* = 2980 (w), 2928 (vw), 1614 (vw), 1531 (m), 1514 (m), 1483 (vw), 1466 (w), 1450 (w), 1416 (w), 1387 (w), 1375 (w), 1348 (vw), 1273 (vw), 1180 (m), 1155 (m), 1103 (w), 1055 (vw), 978 (vs), 939 (w), 887 (m), 841 (vw), 818 (m), 75 (m), 750 (m), 714 (w), 683 (m), 648 (w), 636 (w), 611 (w), 577 (s), 565 (s), 548 (w), 528 (w), 507 (m), 494 (w), 442 (w), 422 (w); ¹**H NMR (CDCl₃, in ppm)**: *δ* = 1.25 (6H, d, ³*J*_{HH} = 6.3 Hz, H6a), 1.37 (6H, d, ³*J*_{HH} = 6.2 Hz, H5b), 2.25 (3H, d, ⁴*J*_{HP} = 0.7 Hz, H4), 2.34 (3H, s, H12), 4.59 (2H, d sept, ³*J*_{HP} = 8.2 Hz, ³*J*_{HH} = 6.2 Hz, H5), 7.20 (2H, d, ³*J*_{HH} = 8.3 Hz, H10), 7.61-7.64 (2H, m, H9); ¹³C{¹H} **NMR (CDCl₃, in ppm)**: *δ* = 13.9 (1C, s, C4), 21.0 (1C, s, C12), 23.8 (2C, d, ³*J*_{CP} = 5.2 Hz, C6a), 24.1 (2C, d, ³*J*_{CP} = 3.9 Hz, C6b), 71.4 (2C, d, ²*J*_{CP} = 4.9 Hz, C5), 85.0 (1C, d, ¹*J*_{CP} = 218.3 Hz, C2), 121.4 (2C, s, C9), 129.7 (2C, s, C10), 135.4 (1C, s, C8), 136.3 (1C, s, C11), 149.2 (1C, d, ²*J*_{CP} = 10.1 Hz, C3), 158.9 (1C, d, ²*J*_{CP} = 23.2 Hz, C1); ³¹P{¹H} **NMR (CDCl₃, in ppm)**: *δ* = 15.3 (s); ³¹P{ **NMR (CDCl₃, in ppm)**: *δ* = 15.3 (t, ³*J*_{PH} = 8.3 Hz); **elemental analysis (in %)**: calculated for C₁₇H₂₅N₂O₄P: C: 57.95, H: 7.15 N: 7.95, found: C: 58.09, H: 6.96, N: 8.14.

Methyl 2-(diisopropoxyphosphoryl)acetate (1):



The synthesis of **1** was performed according to the procedure reported by Hubbard and Miller³ in dried N₂ atmosphere. Trisisopropyl phosphite (86.5 ml, 0.35 mol) was heated to 120 °C and 35.5 ml (0.36 mol, 1.05 eq) methyl bromoacetate were slowly added over a period of 2 h while keeping the temperature at 120 °C. Simultaneously, the generated 2-bromopropane was distilled from the reaction mixture at about 35 - 50 °C inner

temperature in the distillation apparatus. After the addition of methyl bromoacetate was completed the amount of evolving 2-bromopropan decreased and the reaction mixture was kept at a temperature of 120 - 130 °C for additional 2 h. After cooling down residual volatiles were removed under reduced pressure to yield in 87.82 g (98% yield) of a colourless oil containing the crude produce **1** in 92 % purity. The product was used without further purification.

Raman (255 mW, in cm⁻¹): v = 3047 (20), 3029 (21), 3022 (23), 2985 (62), 2943 (100), 2929 (89), 2885 (35), 2877 (36); **IR (ATR, in cm⁻¹)**: v = 2982 (vw), 2955 (vw), 2937 (vw), 1740 (m), 1468 (vw), 1454 (vw), 1437 (vw),

1387 (w), 1375 (w), 1254 (s), 1213 (w), 1178 (w), 1142 (w), 1119 (w), 1103 (m), 976 (vs), 905 (w), 889 (m), 820 (w), 764 (w), 716 (vw), 617 (w), 501 (m), 419 (w); ¹H NMR (CDCl₃, in ppm): $\delta = 1.27$ (12H, d, ³*J*_{HH} = 6.2 Hz, H4), 2.86 (2H, d, ²*J*_{HP} = 21.8 Hz, H2), 3.66 (3H, s, H5), 4.68 (2H, d sept, ³*J*_{HP} = 7.6 Hz, ³*J*_{HH} = 6.2 Hz, H3); ¹³C{¹H} NMR (CDCl₃, in ppm): $\delta = 23.8$ (2C, d, ³*J*_{CP} = 4.9 Hz, C4a), 24.0 (2C, d, ³*J*_{CP} = 4.2 Hz, C4b), 35.3 (1C, d, ¹*J*_{CP} = 135.3 Hz, C2), 52.3 (1C, s, C5), 71.5 (2C, d, ²*J*_{CP} = 6.5 Hz, C3), 166.4 (1C, d, ²*J*_{CP} = 6.3 Hz, C1); ³¹P{¹H} NMR (CDCl₃, in ppm): $\delta = 17.3$ (s); ³¹P{ NMR (CDCl₃, in ppm): $\delta = 17.3$ (tt, ²*J*_{PH} = 21.5 Hz, ³*J*_{PH} = 12.1 Hz).

Methyl 2-(diisopropoxyphosphoryl)-3-oxobutanoate (2):



The synthesis of the precursor **2** was performed on the basis of a procedure reported by *Corbel* and coworkers⁴ under an atmosphere of dried N_2 in dry solvents. To a 25 mL CH₂Cl₂ suspension of MgCl₂ (98%, 6.1176 g, 62.97

mmol) a solution (20 mL CH₂Cl₂) of 15.003 g (57.88 mmol) methyl 2-(diisopropoxyphosphoryl)acetate (1) (92% purity) was added, followed by the addition of 2 eq. Et₃N (99.5%, 17.6 mL, 125.92 mmol). The reaction mixture was stirred for 30 min at room temperature. Subsequent the temperature of the reaction mixture was reduce to 20 °C using an external water cooling and 1.1 eq acetyl chloride (4.94 mL, 69.25 mmol) were added slowly. The colour of the reaction mixture turned yellow. After 15 min of stirring a part of the solution was taken, acidified with 1 M HCl and the organic phase was separated, dried over Na₂SO₄ and monitored by ³¹P {}-NMR spectroscopy. The reaction was brought to completeness by sequential addition of an excess of 1 eq. (8.81 mL, 62.97 mmol) and then 0.5 eq. (4.41 mL, 31.48 mmol) of Et₃N and 0.55 eq. (2.47 mL, 34.63 mmol) and 0.275 .eq (1.24 mL, 17.31 mmol) of the acetyl chloride at intervals of 15 min. The resulting red orange suspension was quenched by 1 M HCl (2 eq.) and extracted 3 times with CH₂Cl₂. The combined organic phases were dried over Na₂SO₄, filtered and the solvent was evaporated under reduced pressure. The crude product was purified by column chromatography (EtOAc: *n*-hexane, 4:1) to yield a yellow oil containing a mixture of the different keto-enol-tautomer's of the target molecules. The product degrades slowly if not stored dry and in an inert atmosphere.

Yield: 10.6633 g, 66% Raman (255 mW, in cm⁻¹): v = 2983 (49), 2939 (100), 2927 (100), 2875 (31), 2738 (9), 1712 (10), 1587 (6), 1454 (16), 1355 (7), 887 (7), 734 (7), 129 (6), 69 (25); **IR (ATR, in cm⁻¹)**: v = 2982 (vw), 2937 (vw), 1744 (vw), 1709 (m), 1591 (w), 1466 (vw), 1437 (w), 1412 (w), 1385 (w), 1375 (m), 1329 (w), 1242 (s), 1180 (w), 1144 (w), 1103 (m), 1080 (m), 982 (vs), 899 (w), 887 (w), 851 (vw), 812 (vw), 781 (w), 771 (w), 737 (vw), 625 (w), 598 (w), 534 (m), 449 (w), 422 (w), 413 (w); ¹H NMR (CDCl₃, in ppm): $\delta = 1.22 - 1.26$ (6H, m, ${}^{3}J_{HH} = 6.2$ Hz, H6a/H6'a/H6'a), 1.31-1.34 (6H, m, ${}^{3}J_{HH} = 6.2$ Hz, H6b/H6'b/H6'b), 2.39 (3H, s, H4), 2.43 (3H, d, ⁴*J*_{HP} = 0.7 Hz, H4'), 2.51 (3H, d, ⁴*J*_{HP} = 0.8 Hz, H4''), 3.69 (3H, s, H7'), 3.76 (3H, s, H7), 3.79 (3H, s, H7''), 4.18 (1H, d, ${}^{2}J_{HP} = 23.8$ Hz, H2), 4.57 (2H, d sept, ${}^{3}J_{HP} = 8.4$ Hz, ${}^{3}J_{HH} = 6.2$ Hz, H5'), 4.60 (2H, m, H5''), 4.80 (2H, m, H5), 13.81 (1H, s, H8'), 14.55 (1H, s, H8''); ${}^{13}C{}^{1}H$ NMR (CDCl₃, in ppm): $\delta = 22.6$ (1C, s, C4''), 23.0 (1C, s, C4'a), 23.1 (1C, s, C4'b), 23.6-24.2 (4C, m, C6/C6'/C6''), 30.0 (1C, s, C4), 51.1 (1C, s, C7'), 52.1 ${}^{2}J_{CP} = 5.2 \text{ Hz}, \text{ C5'}$, 72.8 (1C, d, ${}^{2}J_{CP} = 6.9 \text{ Hz}, \text{ C5a}$), 72.9 (1C, d, ${}^{2}J_{CP} = 6.8 \text{ Hz}, \text{ C5b}$), 89.5 (1C, d, ${}^{1}J_{CP} = 178.7 \text{ Hz}$) Hz, C2'), 92.0 (1C, d, ${}^{1}J_{CP} = 206.0$ Hz, C2''), 164.9 (1C, d, ${}^{2}J_{CP} = 5.5$ Hz, C1), 166.8 (1C, d, ${}^{2}J_{CP} = 9.2$ Hz, C1'), 173.8 (1C, d, ${}^{2}J_{CP} = 9.2$ Hz, C1^{''}), 187.9 (1C, d, ${}^{2}J_{CP} = 6.0$ Hz, C3[']), 189.6 (1C, d, ${}^{2}J_{CP} = 21.7$ Hz, C3^{''}), 196.6 $(1C, d, {}^{2}J_{CP} = 5.0 \text{ Hz}, C3); {}^{31}P{}^{1}H} \text{ NMR (CDCl}_{3}, \text{ in ppm}): \delta = 11.2 (s, P), 14.5 (s, P''), 23.0 (s, P'); {}^{31}P{} \text{ NMR}$ (CDCl₃, in ppm): $\delta = 11.2$ (td, ${}^{2}J_{PH} = 23.7$ Hz, ${}^{3}J_{PH} = 7.6$ Hz, P), 14.5 (t, ${}^{3}J_{PH} = 8.3$ Hz, P''), 23.0 (t, ${}^{3}J_{PH} = 8.3$ Hz, P'); ESI-MS (in m/z): 281.3 [M+H]⁺, 303.3 [M+Na]⁺

Synthesis of [LaL₃HL]·3CH₃CN·3H₂O (3·3CH₃CN·3H₂O)



To prepare the La(III) complex 3.3CH₃CN·3H₂O ([LaL₃HL]·3CH₃CN·3H₂O) 377.4 mg (1.00 mmol, 4 eq) of HL (91%) dissolved in 8 mL CH₃CN, 30.0 mg (0.75 mmol, 3 eq) of NaOH (99%) and 94.4 mg (0.25 mmol, 1 eq.) of LaCl₃·7H₂O (98%) were dissolved in 1.5 ml and 2.0 ml H₂O, respectively. The NaOH solution was added to HL under stirring. Subsequent, the LaCl₃·7H₂O solution was added into the HL and NaOH mixture dropwise at 80 °C. During the addition of LaCl₃·7H₂O, some light yellow solid precipitated. After the addition was completed, the reaction mixture was stirred for 1 h at 80 °C, cooled down to room

temperature, put it in fridge for another 2 h. The light yellow powder was collected after filtration. Suitable crystals for single crystal X-ray diffraction analysis were recrystallized from the acetonitrile and H_2O (6:1) mixture.

Yield: 416.2 mg, 96%, **Raman (80 mW, in cm⁻¹)**: v = 3080 (15), 3039 (14), 3029 (14), 3014 (17), 2979 (32), 2921 (62), 2871 (27), 2730 (16), 1614 (61), 1566 (13), 1516 (38), 1452 (32), 1415 (19), 1375 (34), 1355 (49), 1311 (18), 1294 (31), 1215 (19), 1182 (21), 1143 (13), 1109 (13), 1060 (20), 854 (26), 788 (14), 721 (24), 704 (12), 626 (13), 596 (11), 270 (8), 258 (9), 208 (14), 185 (15), 79 (100); **IR (ATR, in cm⁻¹)**: v = 2976 (w), 2926 (w), 2870 (vw), 1634 (vw), 1616 (w), 1585 (m), 1556 (s), 1514 (s), 1464 (w), 1450 (w), 1421 (m), 1371 (m), 1352 (m), 1310 (w), 1294 (w), 1177 (s), 1142 (w), 1103 (m), 1059 (vw), 972 (vs), 885 (m), 820 (m), 775 (s), 762 162 (m), 739 (m), 717 (w), 704 (w), 644 (w), 625 (m), 590 (vs), 548 (w), 530 (m), 509 (m), 442 (w), 420 (m); ¹**H NMR (CD₃OD, in ppm**): $\delta = 1.06$ (24H, d, ³*J*_{HP} = 7.7 Hz, ³*J*_{HH} = 6.3 Hz, H5), 6.95 (8H, s(br), H9), 7.73 (8H, d, ³*J*_{HH} = 7.7 Hz, H8); ¹³**C**{¹**H**} **NMR (CD₃OD, in ppm)**: $\delta = 14.1$ (4C, s, C4), 20.9 (4C, s, C11), 24.0 (8C, d, ³*J*_{CP} = 4.8 Hz, C6a), 24.4 (8C, d, ³*J*_{CP} = 4.1 Hz, C6b), 71.7 (8C, d, ²*J*_{CP} = 4.6 Hz, C5), 84.8 (4C, d, ¹*J*_{CP} = 237.9 Hz, C2), 121.8 (8C, s, C8), 130.0 (8C, s, C9), 135.3 (4C, s, C10), 137.6 (4C, s, C7), 151.4 (4C, d, ²*J*_{CP} = 14.2 Hz, C3), 166.6 (4C, s(br)), C11; ³¹P{} **NMR (CD₃OD, in ppm**): $\delta = 16.6$ (s(br)); **Elemental analysis** for C₇₄H₁₁₂LaN₁₁O₁₉P4 (*[LaL₃HL]-3CH₃CN·3H₂O)*, calculated: C 51.60, N 8.94, H 6.55; found: C 51.15, N 8.56, H 6.35; **ESI-MS (in m/z)**: 1193.3 [M - HL+ H]⁺.



Fig. S1 ¹H NMR spectrum of La(III) complex (CD₃OD, 300 K).



Fig. S2 ¹³C{¹H} NMR spectrum of La(III) complex (CD₃OD, 300 K).



Fig. S3 ³¹P NMR spectrum of La(III) complex (CD₃OD, 300 K).

Synthesis of $[CeL_3]$ (4)



To prepare the Ce(III) complex [CeL₃] (4) 335.6 mg (0.90 mmol, 3 eq:) of HL (91%) were dissolved in 8 ml acetonitrile and 40.0 mg (0.99 mmol, 3 eq:) of NaOH (99%) in 6 ml degassed H₂O were added under argon atmosphere. The reaction mixture was heated to 80 °C and 87.1 mg (0.15 mmol, 0.5 eq.) Ce₂(SO₄)₃ (99.9%) in 6 ml degassed H₂O were added dropwise under stirring. During the addition of Ce₂(SO₄)₃, a yellow precipitate is generated which indicates the formation of complex. The reaction mixture was stirred for 1 h at 80 °C, cooled down to room temperature, put in the fridge for 2 h. The yellow precipitate was filtrated, washed with a water/acetonitrile mixture (3:2) and dried overnight.

Yield: 170.5 mg, 49%, **Raman (100 mW, in cm⁻¹)**: *v* = 3078 (17), 3038 (13), 3016 (14), 2978 (37), 2921 (89), 2871 (27), 2732 (8), 1615 (100), 1516 (48), 1452 (30),

1416 (11), 1377 (60), 1361 (55), 1311 (18), 1298 (40), 1214 (13), 1180 (17), 1143 (7), 1102 (7), 1061 (24), 889 (5), 856 (23), 791 (12), 758 (5), 717 (17), 650 (6), 627 (10), 595 (7), 422 (6), 390 (5), 327 (6); **IR (ATR, in cm**⁻¹): v = 3649 (vw), 2976 (vw), 2926 (vw), 1615 (vw), 1586 (vw), 1542 (s), 1514 (s), 1423 (m), 1373 (m), 1297 (vw), 1152 (m), 1094 (m), 1060 (vw), 1003 (m), 975 (vs), 888 (w), 854 (vw), 820 (w), 777 (m), 736 (w), 715 (vw), 625 (w), 592 (s), 577 (m), 551 (w), 529 (w), 512 (w), 495 (vw), 421 (vw); ¹H NMR (**CD**₂**Cl**₂, **in ppm**): $\delta = 0.25$ (18H, s, H6a), 1.01 (18H, s, H6b), 1.92 (9H, s, H4), 3.77 (9H, s, H11), 5.21 (6H, s, H5), 6.54 (6H, s, H9), 8.16 (6H, s, H8); ¹³**C**{¹H} NMR (**CD**₂**Cl**₂, **in ppm**): $\delta = 16.7$ (3C, s, C4), 20.6 (3C, s, C11), 23.3 (6C, s, C6a), 23.8 (6C, s, C6b), 71.8 (6C, s, C5), 91.2 (3C, d, ¹J_{CP} = 228.5 Hz, C2), 121.4 (6C, s, C8), 129.2 (6C, s, C9), 133.9 (3C, s, C10), 139.9 (3C, s, C7), 153.3 (3C, s, C3), 187.7 (3C, s (br), C1); ³¹P{¹H} NMR (**CD**₂**Cl**₂, **in ppm**): $\delta = 41.0$ (s); **31**P{} NMR (**CD**₂**Cl**₂, **in ppm**): $\delta = 41.0$ (s); **Elemental analysis** for C₅₁H₇₆CeN₆O₁₄P₃ ([*CeL*₃]·2H₂O), calculated: C 49.79, N 6.83, H 6.23; found: C 49.87, N 6.81, H 5.99; **ESI-MS (in m/z)**: 1194.4 [M+H]⁺.



Fig. S4 ¹H NMR spectrum of $[CeL_3]$ complex (CD₂Cl₂, 300 K).



Fig. S5 ${}^{13}C{}^{1}H$ NMR spectrum of [CeL₃] complex (CD₂Cl₂, 300 K).



Fig. S6 ³¹P NMR spectrum of [CeL₃] complex (CD₂Cl₂, 300 K).

Suitable crystals for single crystal X-ray diffraction analysis were obtained by recrystallization in the glove box involving diethyl ether diffusion into a solution of the complex in acetonitrile with the composition [CeL₃CH₃CN] (CH₃CN \simeq 4).

Synthesis of $[CeL_3HL]$ (5)



To prepare the complex **5** ([CeL₃HL]) 776.7 mg (2.0 mmol, 4 eq.) of HL (91%) were dissolved in 16 ml acetonitrile and 59.9 mg (1.5 mmol, 3 eq.) of NaOH (99%) dissolved in 8 ml degassed H₂O were added under argon atmosphere. The reaction mixture was heated to 80 °C and 142.1 mg (0.25 mmol, 0.5 eq.) Ce₂(SO₄)₃ (99.9%) dissolved in 8 ml degassed H₂O were added dropwise to the stirred solution. Upon the addition of Ce₂(SO₄)₃, light yellow precipitate was formed. The reaction mixture

was stirred for another 2 h at 80 °C to complete the reaction, cooled down to room temperature and put in the fridge for another 2 h. The yellow precipitate was filtrated, washed with a water acetonitrile (1:1) mixture and dried overnight.

Yield: 659.4 mg, 85%, **Raman (100 mW, in cm⁻¹)**: v = 3080 (12), 2979 (42), 2922 (100), 2871 (29), 2730 (8), 1615 (91), 1516 (40), 1452 (26), 1416 (6), 1372 (32), 1356 (52), 1310 (11), 1294 (28), 1213 (15), 1180 (18), 1143 (6), 1107 (8), 1059 (18), 853 (25), 788 (9), 760 (5), 719 (20), 644 (6), 626 (11), 595 (8); **IR (ATR, in cm⁻¹)**: v = 2976 (vw), 2925 (vw), 1615 (vw), 1585 (w), 1555 (w), 1513 (w), 1422 (w), 1371 (w), 1352 (w), 1309 (vw), 1296 (vw), 1175 (w), 1142 (vw), 1104 (w), 973 (s), 886 (w), 819 (w), 775 (w), 739 (vw), 716 (vw), 644 (vw), 625 (w), 591 (m), 548 (vw), 531 (vw), 510 (vw), 422 (vw); ¹**H NMR (CD₂Cl₂, in ppm)**: $\delta = 0.01$ (24H, s (br), H6a), 0.80 (24H, s (br), H6b), 2.13 (12H, s, H4), 2.46 (12H, s (br), H11), 3.52 (8H, s (br), H5), 6.89 (8H, d, ³*J*_{HH} = 5.4 Hz, H9), 8.40 (8H, s, H8); ¹³C{¹H} NMR (CD₂Cl₂, in ppm): $\delta = 15.1$ (4C, s, C4), 20.9 (4C, s, C11), 22.8 (8C, s, C6a), 23.2 (8C, s, C6b), 70.5 (8C, s, C5), 84.7 (4C, d, ¹*J*_{CP} = 229.6 Hz, C2), 121.5 (8C, s, C8), 129.4 (8C, s, C9), 134.6 (4C, s, C10), 138.4 (4C, s, C7), 150.3 (4C, s, C3), 173.2 (4C, s (br), C1); ³¹P{¹H} NMR (CD₂Cl₂, in ppm): $\delta = 32.1$ (s(br)); ³¹P{} NMR (CD₂Cl₂, in ppm): $\delta = 32.1$ (s(br)); **Elemental analysis** for C₆₈H₉₇CeN₈O₁₆P₄ (CeL₃HL), calculated: C 52.81, N 7.25, H 6.32; found: C 52.65, N 7.11, H 5.98; **ESI-MS (in m/z)**: 1194.6 [M-HL+H]⁺(ESI⁺), 1544.6 [M-H]⁻(ESI⁻).





Recrystallization in the glove box involved diethyl ether diffusion into a solution of the complex in acetonitrile to give suitable crystals for single crystal X-ray diffraction analysis as the acetonitrile and diethyl ether solvate of the composition $[CeL_3HL]$ ·CH₃CN·(C₂H₅)₂O (5·CH₃CN·(C₂H₅)₂O).

As depicted in **Fig. S9**, a single ³¹P resonance of **5** is observed at $\delta = 32.1$ ppm, which is 8.9 ppm shifted to higher field compared to the single resonance observed for **4** at $\delta = 41.0$ ppm (**Fig. S6**). The latter is the complex isolated from the reaction of Ce(III) with three equivalents of HL. Presumable the change in coordination number of the

metal centre results in the obtained shift of the ³¹P resonance. In order to proof this hypothesis a solution of **4** was reacted with 1 eq. of HL and monitored by ³¹P NMR spectroscopy. The obtained results are displayed in **Fig. S10** and **Fig. 7** showing that the change in the Ce(III) to ligand ration in the solution leads to shift of about 8 ppm of the resonance in the ³¹P NMR spectra, presumable cause by the change in the coordination of the Ce(III) metal centre.



Fig. S10 Stack of the ³¹P NMR spectra of 4, the reacting of 4 with 1 eq. HL and 5 (CD₂Cl₂, 300 K).

Synthesis of the complexes of Ce(IV), Th(IV), U(IV), and Np(IV)

The synthesis of the 4*f* and 5*f* block metal(IV) complexes [ML₄] were performed in acetonitrile or methanol water mixtures with moderate yields. The ligand HL (4 eq.) was dissolved in acetonitrile or methanol and a 1 eq. water solution of the corresponding metal salt (Ce(SO₄)₂ ·4H₂O, Th(NO₃)₄·5H₂O, NpCl₄, and UCl₄, respectively) was added. For the synthesis of the Ce(IV) and U(IV) complex NaOH was added to promote the deprotonation of HL, suggesting that the *Lewis* acidity of Th(IV) and Np(IV) is strong enough to induce the deprotonation of the ligand without further addition of a base under the present conditions. Due to radiation safety not elemental analysis were conducted for the obtained Th(IV) and Np(IV) complex.

Synthesis of [CeL₄] (6)



For the synthesis of the Ce(IV) complex 385.3 mg (1.0 mmol, 4 eq.) of HL (91%) were dissolved in 8 ml acetonitrile and 40.6 mg (1.0 mmol, 4 eq.) of NaOH (99%) were dissolved in 6 ml H₂O. Ce(SO₄)₂·4H₂O (99.8%, 102.3 mg, 0.25 mmol, 1 eq.) was suspended in 4 ml H₂O. After the addition of the NaOH solution to the solution of HL the Ce(SO₄)₂ suspension was added dropwise at 80 °C to the stirred solution. After some time a colour changed to dark violet on the surface of the residual Ce(SO₄)₂ particles hint to a start of the complex formation. The reaction mixture was stirred for another hour at 80 °C, cooled down to room temperature, put it in fridge for another 2h. The dark violet powder was collected after filtration. The obtained ¹H NMR and ³¹P NMR are shown in **Fig. S11** and **Fig. S12**.





Fig. S12 ³¹P NMR spectrum of Ce(IV) complex (CD₂Cl₂, 300 K).

From ¹H NMR and ³¹P NMR spectra, we concluded that there are two species of the cerium complex in a 1:2 ratio present. From the analytical data of **5** the broad resonance at $\delta = 32.3$ ppm may be assigned to the complex of Ce(III) with the metal centre coordinated by four ligands (³¹P NMR of [CeL₃HL]: $\delta = 32.1$ ppm, **Fig. S9**). Thus, the resonance at $\delta = 16.2$ ppm can be assigned to the desired Ce(IV) complex. To further prove this assumption, 5 mM KMnO₄ solution was used as oxidant to wash the above NMR sample (1:1 ratio). The obtained ¹H, ³¹P and ¹³C{¹H} NMR spectra are shown in **Fig. S13** to **Fig. S15**.



Fig. S13 ¹H NMR spectrum of Ce(IV) complex after washing with 5 mM KMnO₄ (CD₂Cl₂, 300 K).



Fig. S14 ${}^{13}C{}^{1}H$ NMR spectrum of Ce(IV) complex after washing with 5 mM KMnO₄ (CD₂Cl₂, 300 K).



Fig. S15 ³¹P NMR spectrum of Ce(IV) complex after washing with 5 mM KMnO₄ (CD₂Cl₂, 300 K).

As shown in **Fig. S15**, after oxidation, in the ³¹P NMR only one resonance at $\delta = 16.1$ ppm is obtained, pointing at a complete oxidization to Ce(IV) species. Control experiments with pure H₂O were also performed. In the obtained ³¹P NMR spectra after washing with H₂O no change was observed, pointing at the presence of both complexes.

Yield: 294.3 mg, 75%; Raman (80 mW, in cm⁻¹): v = 3080 (22), 3064 (20), 3033 (24), 3010 (27), 2977 (44), 2923 (95), 2885 (52), 2871 (56), 2736 (28), 1649 (28), 1614 (78), 1587 (51), 1564 (100), 1516 (75), 1452 (51), 1357 (53), 1313 (33), 1294 (38), 1213 (29), 1180 (39), 1155 (30), 1141 (28), 1097 (31), 1060 (31), 891 (21), 854 (35), 790 (36), 760 (20), 717 (29), 640 (18), 626 (19), 598 (27), 434 (25), 426 (24), 412 (23), 405 (23), 362 (22), 227 (21), 220 (22), 77 (64); **IR (ATR, in cm⁻¹)**: v = 2976 (w), 2926 (w), 2870 (vw), 1645 (vw), 1614 (w), 1583 (m), 1558 (m), 1529 (m), 1512 (s), 1450 (w), 1423 (m), 1383 (m), 1373 (m), 1352 (m), 1308 (w), 1296 (w), 1221 (w), 1198 (w), 1178 (s), 1144 (m), 1101 (m), 1047 (w), 976 (vs), 887 (m), 845 (w), 820 (m), 777 (s), 739 (m), 717 (w), 685 (w), 642 (w), 627 (m), 592 (s), 577 (s), 569 (s), 548 (m), 530 (m), 509 (m), 453 (w), 422 (m); ¹H NMR (**CD₂Cl₂, in ppm**): $\delta = 0.97$ (24H, d, ³*J*_{HH} = 4.4 Hz, H6a), 1.13 (24H, s (br), H6b), 2.11 (12H, s, H4), 2.13 (12H, s (br), H11), 4.54 (8H, m, H5), 6.72 (8H, s (br), H9), 7.68 (8H, s (br), H8); ¹³C{¹H} NMR (**CD₂Cl₂, in ppm**): $\delta = 16.1$ (k, ³*J*_{CP} = 21.7 Hz, C1); ³¹P{¹H} NMR (**CD₂Cl₂, in ppm**): $\delta = 16.1$ (s); ³¹P{} NMR (**CD₂Cl₂, in ppm**): $\delta = 16.1$ (t, ³*J*_{PH} = 7.7 Hz); **Elemental analysis** for C₆₈H₉₈CeN₈O₁₇P₄ ([CeL₄]·H₂O), calculated: C 52.24, N 7.17, H 6.32; found: C 52.26, N 7.18, H 6.05; **ESI-MS (in m/z)**: 1194.54 [M-L]⁺.

To obtain single crystals suitable for X-ray diffraction analysis the dark violet powder was dissolved in acetonitrile/H₂O mixture and 10 ml ethanol were added. After slow evaporation of all volatiles, square dark violet (almost black) crystalline plates grow on the walls of the vial of the composition $[CeL_4]$ ·3CH₃CN·3H₂O (**6**·3CH₃CN·3H₂O).

Synthesis of [ThL₄]·3CH₃CN·3H₂O (7·3CH₃CN·3H₂O)

Caution! Thorium (²³²Th) is long-lived α -emitter with the half-lives of 1.41×10^{10} years. The radionuclide is also chemically toxic. Handling the radionuclide involves a serious risk to human health. Therefore, special precautions with appropriate lab equipment and facilities dedicated to radiation protection are required for handling this radioactive material.



50.33 mg (0.133 mmol, 4 eq) of HL (91%) were dissolved in 3 ml acetonitrile. Then 19.17 mg (0.033 mmol, 1 eq) of (Th(NO₃)₄·5H₂O) dissolved in 1 mL of deionized water were added to the solution. At the interphase of the aqueous and the organic phase the immediate formation of colourless precipitate, which immediately re-dissolves, could be observed. The reaction was thoroughly mixed by a vortex apparatus and after some seconds a colourless precipitate formed. The precipitate was recrystallized by heating the reaction mixture at 50 °C for 2 h in a microwave and slowly cooling down stepwise from 40 °C to r.t. without stirring, yielding in 29.81 mg of the target compound 7.



Fig. S16 ¹H NMR spectrum of Th(IV) complex (CDCl₃, 300 K).



Fig. S17 ${}^{13}C{}^{1}H$ NMR spectrum of Th(IV) complex (CDCl₃, 300 K).



17.8698

Fig. S18 ³¹P NMR spectrum of Th(IV) complex (CDCl₃, 300 K).

Synthesis of [UL₄]

Caution! Uranium consists of radioactive nuclides including long-lived α -emitters (²³⁵U and ²³⁸U; $T_{1/2} = 7.04 \times 10^8$, and 4.47×10^9 years). These radionuclides are also chemically toxic. Handling these radionuclides involves a serious risk to human health. Therefore, special precautions with appropriate lab equipment and facilities dedicated to radiation protection are required for handling these radioactive materials."



The manipulations were performed under N_2 atmosphere. The formed complex is stable in air. The ligand HL (48.6 mg, 0.130 mmol, 4 eq.) were dissolved in 2 mL CH₃OH and reacted with 1 eq. of UCl₄ (16.0 mg, 0.032 mmol) in H₂O in the presence of 4 eq. NaOH. The addition of water to the reaction mixture resulted in the precipitation of the formed complex. The obtained suspension was heated 80°C, stirred for 1 h, filtered and recrystallized upon cooling to room temperature to give 11.0 mg of **8**.

Yield: 11.0 mg, 21%, **IR (ATR, in cm⁻¹)**: *v* = 2978 (w), 2926 (w), 2872 (vw), 2869 (vw), 1615 (w), 1586 (w), 1532 (s), 1512 (vs), 1466 (m), 1444 (m), 1423 (m), 1374 (s), 1312 (w), 1299 (w), 1288 (w), 1275 (w), 1180 (m), 1142 (s), 1093 (s), 1061 (w), 977 (vs), 898 (m),

888 (s), 854 (w), 844 (w), 819 (s), 774 (s), 760 (s), 729 (m), 715 (m), 704 (m), 685 (m). ¹H NMR (CD₂Cl₂ in ppm): $\delta = -5.29$ (s (br)), -3.88 (s (br)), -0.85 (s (br)), -0.26 (s (br)), 0.19 (s (br)), 0.57 (s (br)), 1.23 (s (br)), 1.34 (s (br)), 2.19 (s (br)), 2.34 (s (br)), 4.22 (s (br)), 4.54 (s (br)), 6.25 (s (br)), 6.97 (s (br)), 7.20 (s (br)), 7.60 (s (br)), 8.94 (s (br)), 11.08 (s (br)); ¹³C{¹H} NMR (CD₂Cl₂, in ppm): $\delta = 15.0$ (s, (br)), 15.5 (s, (br)), 17.6 (t, (br)), 19.4 (s, (br)), 21.2 (s, (br)), 21.6 (m, (br)), 23.3 (s, (br)), 28.1 (s, (br)), 62.0 (s, (br)), 66.1 (s, (br)), 66.9 (s, (br)), 72.7 (s, (br)), 77.7 (s, (br)), 79.4 (s, (br)), 119.3 (s, (br)), 122.9 (s, (br)), 127.9 (s, (br)), 129.9 (s, (br)), 130.7 (s, (br)), 131.5 (s, (br)), 134.5 (s, (br)), 142.5 (s, (br)), 143.8 (s, (br)), 147.1 (s, (br)), 151.1 (s, (br)), 167.3 (s, (br)); ³¹P{H} NMR (CD₂Cl₂ in ppm): $\delta = -40.5$ (s (br)), -29.9 (s (br)). Elemental analysis for C₆₈H₉₆N₈O₁₆P₄U ([UL₄]), calculated: C 49.70, N 6.82, H 5.89; found: C 49.34, N 6.83, H 5.91.



Fig. S19 ¹H NMR spectrum of U(IV) complex (CD₂Cl₂, 300 K).







Fig. S21 ³¹P NMR spectrum of U(IV) complex (CD₂Cl₂, 300 K, * free ligand).

Employing different solvents results in the insolation of single crystals suitable for X-ray diffraction with an altered placement of the ligands coordinating the U(IV) metal centre. If CH_3CN is used during the crystallisation the complex with the composition $[UL_4]$ ·3 CH_3CN ·3 H_2O (8·3 CH_3CN ·3 H_2O) is obtained. From CH_2Cl_2 in the absence

of CH₃CN [UL₄]·CH₂Cl₂ (8·CH₂Cl₂) and from toluene [UL₄] (8) were obtained. In all cases has the metal centre a coordination number of eight formed by the coordination of the two O-donor atoms of four deprotonated ligands. Whereas 8·3CH₃CN·3H₂O exhibits a strictly square-antiprismatic coordination environment due to the tetragonal space group P4/n, the structures 8·CH₂Cl₂ and 8 are distorted towards a bicapped trigonal geometry (Fig. 5). The coordination polyhedron is best determined by measuring the angles δ and φ according to Porai-Koshits and Aslanov,⁵ indicating a square-antiprismatic arrangement for 8·CH₂Cl₂ but a bicapped trigonal geometry for 8. Furthermore, a different arrangement of the ligands is observable for all three isomers. 8·3CH₃CN·3H₂O adopts the *llll*-configuration with all ligands linking the two square faces of the polyhedron and thereby pointing all in the same direction. In contrast the ligands in solvent-free 8 occupy two edges each of quadrangle closest to square shape, which is described as *ssss*-configuration. In the bicapped trigonal geometry this configuration is described as *t₁t₁p₂p₂*. Potential pathways for interconversion between the three isomers are shown in Fig. S22.



Fig. S22 Potential pathways for interconversion between the three isomers $[UL_4]$ ·3CH₃CN·3H₂O (8·3CH₃CN·3H₂O), $[UL_4]$ ·CH₂Cl₂ (8·CH₂Cl₂) and $[UL_4]$ (8).

Extended ³¹P NMR studies of the U(IV) complex **8** were performed motivated by the presence of two broad resonances at $\delta = -40.5$ ppm (s (br)) and $\delta = -29.9$ ppm (s (br) (**Fig. 6**).

Variable temperature ³¹P NMR in CD₂Cl₂ were performed between 300 K and 190 K. The obtained spectra at 300 K, 280 K, 260 K, 240 K, 220 K, 200 K and 190 K are displayed in **Fig. S23** and reveal a broadening of the two resonances upon cooling. At 240 K an extremely broad signal appears, whereas at 220 K, 200 K and 190 K more complex spectra are observable. The two major resonances shift upon cooling to 190 K to $\delta = -2.3$ ppm (s (br)) and $\delta = -46.1$ ppm (s (br)). Presumable, the more complex spectra is a result of a reduced movement of the *iso*-propyl groups of the ligand at low temperature and hence a series of resonances can be observed. In contrast, the resonance attributed to the free ligand at $\delta = -15.9$ ppm (s (br) does not shift with the temperature variation.



Fig. S23 ³¹P NMR spectra of U(IV) complex at variable temperatures in CD₂Cl₂.

In order to investigate potential interaction between the present species ${}^{31}P{}^{-31}P{}^{-}$ exchange spectroscopy (EXSY) experiment were performed at 300 K (**Fig. 8**) and 190 K (**Fig. S24**). In both a mixing time of 0.025 seconds was employed. The presence of exchange peaks between the two resonances at $\delta = -40.5$ ppm and -29.9 ppm at 300 K indicates a dynamic exchange between the present species. Also at 190 K multiple exchange peaks are obtained showing an exchange of the different species. In contrast, no exchange peaks are observed involving the free ligand, pointing at an exchange between the complex species only.



Fig. S24 ³¹P-³¹P- EXSY NMR spectrum of a solution of the U(IV) complex at 190 K in CD_2Cl_2 with a mixing time of 0.025 seconds.

In a second set of experiments toluene- d_8 was used as solvent and the ³¹P NMR spectra of the U(IV) complex were recorded between 190 K and 360 K in increments of 10 K (**Fig. S25** – **Fig. S27**). In addition, the spectra were recorded in the presence of various amounts of CD₃CN at 300 K. As shown in **Fig. S28** the ratio between normalised integrals of the two resonance as $\delta = -28.2$ ppm and $\delta = -38.9$ ppm change from 0.68 to 0.32 in the absence of CD₃CN to 0.47 to 0.53 in the presence of an excess of CD₃CN. In ³¹P NMR experiments using hexafluorobenzene a comparable change of the integral ratios for the two resonances at $\delta = -23$ ppm and $\delta = -43$ ppm is observable (**Fig. S29**). In the absence of CD₃CN the ratio is approximately 0.70 to 0.30 upon consideration of the very broad resonance at 23 ppm, whereas in the presence of an excess of CD₃CN a ratio of 0.38 to 0.62 is obtained.



Fig. S25 ³¹P NMR spectra of U(IV) complex in dependence of the temperature in toluol- d_8 between 360 and 300 K.



Fig. S26 ³¹P NMR spectra of U(IV) complex in dependence of the temperature in toluol- d_8 between 300 and 240 K.



Fig. S27 ³¹P NMR spectra of U(IV) complex in dependence of the temperature in toluol- d_8 between 240 and 190 K.



Fig. S28 ³¹P NMR spectra of U(IV) complex at 300 K in toluene- d_8 with increasing amount of CD₃CN with the normalised integral to 1.0 of the resonances at -28.2 and -38.9 ppm..



Synthesis of [NpL₄]·3CH₃CN·3H₂O (9·3CH₃CN·3H₂O)

Caution! Neptunium (²³⁷Np) consists of radioactive nuclides including long-lived α -emitters (²³⁷Np; $T_{1/2} = 2.14 \times 10^{6}$ years). Special precautions as well as appropriate equipment and facilities for radiation protection are required for handling this material. All experiments were carried out in a controlled laboratory at the Institute of Resource Ecology, Helmholtz-Zentrum Dresden - Rossendorf.



The complex $[NpL_4]$ ·3CH₃CN·3H₂O (9·3CH₃CN·3H₂O) was prepared by the reaction of 4 eq. of HL (91%; 48.3 mg, 0.130 mmol), dissolved in 3 mL of acetonitrile with 1 eq. of NpCl₄ (12.5 mg, 0.032 mmol) dissolved in 1 mL deionized water (turquoise coloured solution) at room temperature. An immediate colour change and the formation of a microcrystalline precipitate indicate the starting complex formation. After filtration of the microcrystalline precipitate the residual liquor was collected and left without stirring. Overnight green yellowish coloured crystals of 9, suitable for X-ray single crystal analysis were grown from the mother liquor. After removing the crystals from their mother liquor they rapidly lose solvent molecules, resulting in a pale noncrystalline solid.

Yield: 17 mg, 30%, **IR (ATR, in cm⁻¹)**: v = 2978 (w), 2951 (vw), 2926 (vw), 2887 (vw), 2882 (vw), 2875 (vw), 2870 (vw), 2866 (vw), 1726 (vw), 1724 (vw), 1615 (w), 1586 (w), 1534 (s), 1513 (s), 1465 (w), 1422 (m), 1383 (m), 1374 (m), 1313 (w), 1300 (w), 1288 (w), 1179 (w), 1144 (s), 1138 (s), 1092 (s), 1060 (w), 1045 (m), 983 (vs), 936 (m), 897 (m), 889 (m), 855 (w), 834 (w), 821 (s), 789 (m), 772 (s), 758 (m), 727 (m), 716 (m), 704 (m); 1³C{¹H} NMR (CDCl₃, in ppm): $\delta = 15.2$ (s), 21.1 (s), 21.8 (s), 24.1 (s), 70.0 (s), 121.3, 129.5 (s), 129.9 (s), 134.4 (s), 142.3 (s (br)), 143.8 (s (br)); ³¹P{} NMR (CDCl₃ in ppm): $\delta = -53.4$ (s (br)).



Fig. S31 ³¹P NMR spectrum of Np(IV) complex 9 in CDCl₃ (300 K, * free ligand).

X-ray crystal structure analyses

Suitable single crystals were coated with Paratone-N oil or Fomblin Y25 PFPE oil and mounted using either a glass fiber or a nylon loop. In the diffractometer the mounted single crystals were exposed to a cold nitrogen stream. Crystal and data collection details are given in Table S1-S3. The single crystals analysis of $[NpL_4]\cdot 3CH_3CN\cdot 3H_2O \quad (\textbf{9}\cdot 3CH_3CN\cdot 3H_2O) \quad was \quad performed \quad at \quad the \quad Helmholz-Zentrum-Dresden-Rossendorf$ (HZDR) on a Bruker D8 VENTURE diffractometer with a PHOTON 100 CMOS detector at 100 K and microfocus Mo K α radiation ($\lambda = 0.71073$ Å). Suitable single crystals were selected under a polarizing optical microscope and mounted on a Micro-LoopTM (MiTiGen, USA) with mineral oil. The remaining X-ray diffraction data was collected on a Rigaku Oxford Diffraction SuperNova diffractometer using Cu K α radiation ($\lambda = 1.54184$ Å) generated by a Nova micro-focus source. For 9.3CH₃CN·3H₂O the data reduction was accomplished performed using SAINT (Version 8.37A).⁶ The data correction for absorption effects was performed by using the Numerical Mu Calculated method (SADABS-2016/2).⁷ The development of the structure was achieved by successive difference Fourier syntheses and the refinement was obtained by full-matrix least-squares on all F^2 data using SHELXL program suite (Version 2014-7)⁸ and ShelXle.⁹ Data reduction and absorption correction of the residual compounds was accomplished either with CrysaAlisPro¹⁰ software or Bruker SMART¹¹ or Bruker SADABS¹². Employing Olex2¹³, the structures were solved with SHELXT¹⁴ and refined with SHELXL⁸ by least-square minimization against F^2 using first isotropic and later anisotropic thermal parameters for all non-hydrogen atoms. Hydrogen atoms bonded to carbon atoms were added to the structure models on calculated positions using the riding model. All other hydrogen atoms were localized in the difference Fourier map. If necessary, disorders of solvent molecules were treated with appropriate restraints (SADI, SIMU, DELU, ISOR, DFIX, SUMP, SAME, FLAT, EADP, EXYZ, DFIX). In the structure of 3·3CH₃CN·3H₂O the solvent molecules are heavily disordered, probably due to their position in channels through the crystal structure. Similar to related structures of complexes 6, 7, 8 and 9 the unit cell contains 6 molecules acetonitrile and 6 molecules of water of which only 2 molecules of acetonitrile could be refined. The other solvent molecules have been treated as a diffuse contribution to the overall scattering without specific atom positions by SQUEEZE/PLATON.¹⁵ Images of the structures were created with Olex2¹³ software. All structures have been deposited with the Cambridge Crystallographic Data Centre (CCDC) and can be accessed free of charge under the numbers 1995660-1995663, 1997488 and 2020834-2020837.

	3 ·3CH ₃ CN·3H ₂ O	CH ₃ CN⊂4
formula	C ₇₄ H ₁₁₂ LaN ₁₁ O ₁₉ P ₄	C ₅₃ H ₇₅ CeN ₇ O ₁₂ P ₃
M_r in g mol ⁻¹	1722.53	1235.23
colour, habit	light yellow, block	clear colourless, block
crystal system	tetragonal	triclinic
space group	P4/n	<i>P</i> -1
a in Å	17.80376(5)	11.6391(3)
b in Å	17.80376(5)	13.3055(3)
c in Å	13.75097(7)	20.5728(3)
α in $^{\circ}$	90	91.504(2)
β in °	90	90.766(2)
γ in °	90	111.160(2)
V in Å ³	4358.70(3)	2969.24(12)
Z	2	2
T in K	100.0(3)	100.0
crystal size in mm ³	$0.257\times0.154\times0.104$	$0.143 \times 0.105 \times 0.033$
$ ho_{\rm c}$ in g cm ⁻³	1.312	1.382
F(000)	1804.0	1282.0
diffractometer	OD SuperNova	OD SuperNova
$\lambda_{XK\alpha}$ in Å	X = Cu 1.54184	X = Cu 1.54184
θ_{\min} in °	6.428	7.128
θ_{\max} in °	153.386	//.200
index range	$-22 \leq h \leq 20$	$-14 \leq h \leq 14$
	$-22 \le k \le 15$	$-16 \le k \le 16$
	$-10 \le 1 \le 17$	$-22 \le 1 \le 25$
μ in mm ⁻¹	5.058	7.208
abs. correction	gaussian	gaussian
reflections collected	48464	69638
P	4388	0.0565
\mathbf{K}_{int}	0.0249	0.0303
reflections obs. $[F \ge 26(F)]$		11/35
residual density in e A	0.227-0.52	1.58/-1.73
parameters	327	704
GOOF	1.079	1.071
$R_1[I>2\sigma(I)]$	0.0196	0.0377
wR_2 (all data)	0.0527	0.1009
CCDC	1995661	2020835

	5·CH ₃ CN·Et ₂ O	6·3CH ₃ CN·3H ₂ O
formula	C ₇₄ H ₁₁₀ CeN ₉ O ₁₇ P ₄	C ₇₄ H ₁₁₁ CeN ₁₁ O ₁₉ P ₄
M_r in g mol ⁻¹	1661.70	1722.73
colour, habit	clear colourless, block	dark violet, plate
crystal system	monoclininc	tetragonal
space group	<i>P</i> 2 ₁	P4/n
a in Å	13.16990(10)	17.60897(13)
b in Å	18.44880(10)	17.60897(13)
c in Å	17.90400(10)	13.59834(15)
α in $^{\circ}$	90	90
β in °	106.0730(10)	90
γ in °	90	90
V in Å ³	4180.07(5)	4216.52(8)
Z	2	2
T in K	100.0	99.98(16)
crystal size in mm ³	$0.149 \times 0.108 \times 0.078$	$0.245\times0.196\times0.016$
$ ho_{\rm c}$ in g cm ⁻³	1.320	1.357
F(000)	1742.0	1804.0
diffractometer	OD SuperNova	OD SuperNova
$\lambda_{XK\alpha}$ in Å	X = Cu 1.54184	$X = Cu \ 1.54184$
θ_{min} in °	5.136	6.5 153 672
θ_{\max} in °	76.835	155.072
index range	$-14 \le h \le 16$	$-22 \le h \le 21$
	$-23 \le k \le 21$	$-19 \le k \le 21$ $-17 \le 1 \le 16$
u in mm ⁻¹	$-22 \le 1 \le 22$	5 / 8/
μ in min	gaussian	Gaussian
reflections collected	200500	30443
reflections unique	15854	4427
R _{int}	0.0396	0.0384
reflections obs. $[F>2\sigma(F)]$	15680	4355
residual density in e $Å^{-3}$	0.93/-0.82	0.30 / -0.71
parameters	973	244
GOOF	1.046	1.075
$R_1[I>2\sigma(I)]$	0.0377	0.0240
wR_2 (all data)	0.0990	0.0640
CCDC	2020834	1995660

Table S2 Crystallographic Data of $[CeL_3HL] \cdot CH_3CN \cdot Et_2O$ (5 $\cdot CH_3CN \cdot Et_2O$) and $[CeL_4] \cdot 3CH_3CN \cdot 3H_2O$ (6 $\cdot 3CH_3CN \cdot 3H_2O$).

	7·3CH ₃ CN·3H ₂ O	8-3CH ₃ CN·3H ₂ O
formula	$C_{74}H_{111}N_{11}O_{19}P_4Th$	$C_{74}H_{111}N_{11}UO_{19}P_4$
M_r in g mol ⁻¹	1814.65	1820.64
colour, habit	yellowish, block	clear yellow, block
crystal system	tetragonal	tetragonal
space group	<i>P</i> 4/ <i>n</i>	<i>P</i> 4/ <i>n</i>
a in Å	17.74494(4)	17.68430(10)
b in Å	17.74494(4)	17.68430(10)
c in Å	13.70378(6)	13.7690(2)
α in $^{\circ}$	90	90
β in °	90	90
γ in °	90	90
V in Å ³	4315.09(3)	4306.04(8)
Ζ	2	2
T in K	100.0(5)	100.0
crystal size in mm ³	$0.094 \times 0.070 \times 0.056$	$0.107\times 0.065\times 0.025$
$\rho_{\rm c}$ in g cm ⁻³	1.397	1.404
F(000)	1868.0	1872.0
diffractometer	OD SuperNova	OD SuperNova
$\lambda_{XK\alpha}$ in Å	X = Cu 1.54184	X = Cu 1.54184
θ_{\min} in °	3.225	6.42
θ_{\max} in °	76.813	/6.916
index range	$-22 \le h \le 22$	$-14 \le h \le 22$
	$-21 \le k \le 22$ $-17 \le 1 \le 17$	$-21 \le k \le 20$ -17 < 1 < 16
u in mm ⁻¹	6 956	6 502
μ in this	multi coon	0.392
reflections collected	55244	20204
reflections unique	4563	4509
Rint	0.0721	0.0458
reflections obs. $[F>2\sigma(F)]$	4488	4509
residual density in e $Å^{-3}$	0.63 / -0.77	1.07 / -0.97
parameters	323	321
GOOF	1 087	1 061
$\mathbb{R}_1[I > 2\sigma(I)]$	0.0188	0.0343
wR_2 (all data)	0.0483	0.0928
CCDC	1995662	1997488

Table S3 Crystallographic Data of $[ThL_4]$ ·3CH₃CN·3H₂O (7·3CH₃CN·3H₂O) and $[UL_4]$ ·3CH₃CN·3H₂O(8·3CH₃CN·3H₂O).

	9·3CH ₃ CN·3H ₂ O	8·CH ₂ Cl ₂
formula	C ₇₄ H ₁₁₃ N ₁₁ NpO ₂₀ P ₄	$C_{69}H_{98}Cl_2N_8O_{16}P_4U$
M _r in g mol ⁻¹	1837.63	1728.36
colour, habit	clear dark orange, block	clear yellow, block
crystal system	tetragonal	monoclininc
space group	P4/n	$P2_{1}/c$
a in Å	17.6551(7)	13.69468(8)
b in Å	17.6551(7)	24.32535(12)
c in Å	13.7559(6)	23.65863(10)
α in °	90	90
β in °	90	92.9022(4)
γ in °	90	90
V in Å ³	4287.7(4)	7871.24(7)
Ζ	2	4
T in K	100.0	100.0
crystal size in mm ³	$0.334 \times 0.324 \times 0.278$	$0.138 \times 0.082 \times 0.035$
$ ho_{\rm c}$ in g cm ⁻³	1.423	1.458
F(000)	1894.0	3528.0
diffractometer	Bruker D8 VENTURE	OD SuperNova
$\lambda_{XK\alpha}$ in Å	X = Mo 0.71073	X = Cu 1.54184
θ_{\min} in °	4.614	5.214
θ_{\max} in °	54.206	77.024
index range	$-22 \le h \le 22$	$-17 \le h \le 17$
	$-22 \le k \le 22$ -17 < 1 < 17	$-30 \le k \le 28$ -23 < 1 < 29
u in mm ⁻¹	1 350	7 751
μ in min	aussion	agussian
reflections collected	903 <i>4</i> 7	02048
reflections unique	4740	16496
R _{int}	0.0264	0.0491
reflections obs. $[F>2\sigma(F)]$	4599	15673
residual density in e $Å^{-3}$	1.23 / -1.57	1.28/-2.53
parameters	315	966
GOOF	1.061	1.095
$R_1[I>2\sigma(I)]$	0.0219	0.0403
wR_2 (all data)	0.0585	0.1090
CCDC	1995663	2020836

 $\textbf{Table S4} Crystallographic Data of [NpL_4] \cdot 3 CH_3 CN \cdot 3H_2 O (\textbf{9} \cdot 3 CH_3 CN \cdot 3H_2 O) and [UL_4] \cdot CH_2 Cl_2 (\textbf{8} \cdot CH_2 Cl_2).$

	8
formula	$C_{68}H_{96}N_8O_{16}P_4U$
M_r in g mol ⁻¹	1643.43
colour, habit	clear green, block
crystal system	monoclininc
space group	$P2_{1}/n$
a in Å	32.05395(19)
b in Å	13.02519(5)
c in Å	36.6159(2)
α in $^{\circ}$	90
β in °	101.1117(6)
γ in $^{\circ}$	90
V in Å ³	15000.89(14)
Z	8
T in K	100.0
crystal size in mm ³	$0.413 \times 0.209 \times 0.097$
$ ho_{\rm c}$ in g cm ⁻³	1.455
F(000)	6720.0
diffractometer	OD SuperNova
$\lambda_{XK\alpha}$ in Å	$X = Cu \ 1.54184$
θ_{\min} in °	4.918
θ_{\max} in °	76.937
index range	$-40 \leq h \leq 40$
	$-11 \le k \le 16$ -46 < 1 < 45
μ in mm ⁻¹	$-40 \le 1 \le 43$ 7 463
abs correction	gaussian
reflections collected	170311
reflections unique	31455
R _{int}	0.0510
reflections obs. $[F>2\sigma(F)]$	30867
residual density in e Å ⁻³	3.75/-1.93
parameters	1794
GOOF	1.156
$R_1[I \ge 2\sigma(I)]$	0.0596
wR ₂ (all data)	0.1529
CCDC	2020837

Table S5 Crystallographic Data of $[UL_4]$ (8).

Selected bond lengths/ Å	3 ·3CH ₃ CN·3H ₂ O		
Lal –O1	2.4604(9)		
La1 –O2	2.5128(9)		
P1 -O2	1.4852(10)		
P1 -C2	1.7371(13)		
C1 -01	1.2606(15)		
OO _{chelate} distances/ Å			
01 02	3.0128(13)		
Dihedral angles (°) of selected planes			
Plane (1): O1, La1, O2	7.01(5)		
Plane (2): O1, C1, C2, P1,	02		
Hydrogen bonds			
D-H ··· A	$D \cdots A / Å$ D-H-A angle (°)		
N2-H2 ··· O5B	2.69(2) 148(9)		
O5A-H5AA ··· N2 ⁱ	2.926(10) 170.7(8)		
O5A-H5AB ··· O5B ⁱⁱ	2.86(3) 143.7(8)		
ⁱ (1/2+y, 1-x, 1-z); ⁱⁱ (3/2-x, 1/2-y, z)			

Table S6: Selected bond lengths (Å), $O^{-}O_{chelate}$ distances (Å), dihedral angles of the planes (°) and hydrogen bonds of the La(III) complex **3**·3CH₃CN·3H₂O.

The numeration of the atoms in the planes refers to the numeration assigned to the atoms in each complex species (see Fig. 1 and Fig. S32)



Fig. S32 Molecular Structure of 3.3CH₃CN·3H₂O. All hydrogen atoms and solvates are omitted for clarity; ellipsoids are drawn at 50% probability level.

Table S7: Selected bond lengths (Å), O^{\cdots}O_{chelate} distances (Å), dihedral angles of the planes (°), δ and ϕ ⁵ of the planes (°) and hydrogen bonds of Ce(III) complexes CH₃CN⊂4 ([CeL₃CH₃CN]) and 5·CH₃CN·Et₂O ([CeL₃HL]·CH₃CN·Et₂O).

S	Selected bond lengths/ Å	CH ₃ CN⊂4	Selected bond lengths/ Å	5. CH ₃ CN. Et ₂ O
	Ce1-O1	2.399(2)	Ce1-O4	2.408(3)
	Ce1-O2	2.4198(19)	Ce1-O1	2.536(3)
	Ce1-O5	2.3764(18)	Ce1-O8	2.395(3)
	Ce1-O6	2.416(2)	Ce1-O5	2.441(3)
	Ce1-O9	2.3724(18)	Cel-O12	2.490(3)
	Cel-Ol0	2.427(2)	Ce1-09	2.526(3)
	Ce1-N7	2.678(3)	Cel-O16	2.500(3)
			Ce1-O13	2.468(3)
	P1-O2	1.500(2)	P1-O1	1.489(3)
	P2-O6	1.495(2)	P2-O5	1.500(3)
	P3-O10	1.503(2)	P3-O9	1.488(3)
			P4-O13	1.492(3)
	P1-C2	1.737(3)	P1-C7	1.737(5)
	P2-C19	1.733(3)	P2-C24	1.733(5)
	P3-C36	1.731(3)	P3-C41	1.761(4)
			P4-C58	1.744(4)
	C1-O1	1.284(3)	С9–О4	1.286(5)
	C18-O5	1.282(3)	C27-O8	1.278(6)
	C35-O9	1.283(3)	C44-O12	1.251(5)
			C61-O16	1.268(5)
0	O chelate distances/ Å		OO chelate distances/ Å	
	01 02	2.918(3)	O1 ··· O4	2.926(4)
	O5 ··· O6	2.933(2)	O5 ··· O8	2.981(4)
	O9 ··· O10	2.917(3)	O9 ··· O12	2.983(4)
			O13 ··· O16	2.914(4)
Dih	edral angles (°) of selected planes		Dihedral angles (°) of selected planes	
	Plane (1): O1, Ce1, O2		Plane (1): O1, Ce1, O4	
1-2	Plane (2): O1, C1, C2, P1, O2	26.33(11)	1-2 Plane (2): O1, P1, C7, C9, O4	5.28(18)
	Plane (3): O5, Ce1, O6		Plane (3): O5, Ce1, O8	
3-4	Plane (4): O5, C18, C19, P2, O6	17.94(12)	3-4 Plane (4): O5, P2, C24, C27, O8	10.47(13)

	Plane (5): O9, Ce1, O10			Plane (5): O9, Ce1, O12	
5-6	Plane (6): O9, C35, C36, P3, O10	22.69(9)	5-6	Plane (6): O9, P3, C41, C44, O12	3.87(15)
				Plane (7): O13, Ce1, O16	
			7-8	Plane (8): O13, P4, C58, C61, O16	3.69(14)
			9-10	Plane (9): O1, O5, O6	
			δ_1	Plane (10): O1, O5, O13	10.39(15)
			11 12	Plane (11): O2, O10, O14	
			δ ₂	Plane (12): O9, O10, O14	20.15(14)
			12 14	Plane (13): O5, O6, O10	
			δ ₃	Plane (14): O5, O9, O10	42.01(14)
			15 16	Plane (15): O1, O2, O14	
			13-10 δ ₄	Plane (16): O1, O13, O14	43.90(14)
			17.18	Plane (17): O1, Ce1, O10	
			φ ₁	Plane (18): O9, Ce1, O13	15.88(13)
			10.20	Plane (19): O5, Ce1, O14	
			φ ₂	Plane (20): O2, Ce1, O6	16.18(11)
				Hydrogen bonds	
				D-H … A / Å	
				$N5-H5 \cdots N7^i$	2.754(5)
				D-H-A angle (°)	
				N5-H5 ··· N7	151.5(3)
ⁱ (1-x,	-1/2+y, 1-z)				

The numeration of the atoms in the planes refers to the numeration assigned to the atoms in each complex species (see **Fig. 2** and **Fig. S33**).



Fig. S33 Representation of hydrogen bond interactions between N5 and N7' present in the crystal structure of $5 \cdot CH_3CN \cdot Et_2O$ ([CeL₃HL]·CH₃CN·Et₂O). All carbon hydrogen atoms and solvates are omitted for clarity; ellipsoids are drawn at 50% probability level.

		6·3CH ₃ CN·	7·3CH ₃ CN·	8-3CH ₃ CN·	9·3CH ₃ CN·
Selecte	ed bond lengths/ A	$3H_2O$	3H ₂ O	$3H_2O$	$3H_2O$
	Ce1/Th1/U1/Np1-O1	2.3346(11)	2.3568(13)	2.304(2)	2.2990(12)
	Ce1/Th1/U1/Np1-O2	2.4171(12)	2.4311(13)	2.388(2)	2.3766(12)
	P1-O2	1.4929(13)	1.5016(14)	1.502(2)	1.4955(13)
	P1-C2	1.7327(18)	1.728(2)	1.724(4)	1.725(2)
	C1-O1	1.2807(19)	1.285(2)	1.293(4)	1.283(2)
OO _{ch}	elate distances/ Å				
	01 02	2.9587(17)	2.9328(19)	2.916(3)	2.9269(18)
Dihedu	ral angles (°) of selected planes				
	Plane (1):				
1-2	OI, Cel/IhI/UI/NpI, O2	0.22(6)	1.65(7)	1.53(12)	2.41(6)
	Plane (2) :				
	$\frac{01, 01, 02, 11, 02}{\text{Plane} (3)}$				
3-4	01, 01', 01''	0	0	0	0
δ_1	Plane (4):	0	0	0	0
	01, 01", 01""				
	Plane (5):				
5-6	02, 02', 02''	0	0	0	0
δ_2	Plane (6):				
	$02, 02^{\prime\prime}, 02^{\prime\prime\prime}$				
7-8	(7):				
, U S.	O1, O1, O2 Plane (8):	49.42(4)	48.10(5)	49.43(8)	50.09(3)
03	$\Omega^2 \Omega^2 \Omega^1$				
	Plane $(9)^{\circ}$				
9-10	01", 01", 02"	40.40(4)	40 10(7)	40,42(0)	50.00(2)
δ_4	Plane (10):	49.42(4)	48.10(5)	49.43(8)	50.09(3)
	01''', 02 ^{''} , 02'''				
	Plane (11):				
11-12	O1, Ce1/Th1/U1/Np1, O2	25 82(3)	26 98(3)	25 89(15)	25 35(2)
ϕ_1	Plane (12):	25.02(5)	20.90(5)	25.69(15)	25.55(2)
	O1"", Ce1/Th1/U1/Np1, O2"				
12 14	Plane (13):				
13-14	OI, Cel/Ihl/Ul/Npl, $O2^{\prime\prime\prime}$	23.01(3)	21.95(3)	22.94(6)	23.46(2)
ϕ_2	Plane (14): $O1^2 Co1/Tb1/U1/Mrs1 O2^2$	~ /			~ /
	<u>01</u> , Ce1/1n1/01/Np1, O2	0			

Table S8: Selected bond lengths (Å), O^{\cdots}O_{chelate} distances (Å), dihedral angles of the planes (°), and δ and ϕ^5 of the planes (°) of **6** – **9**.

The numeration of the atoms in the planes refers to the numeration assigned to the atoms in each complex species (see Fig. 3 and Fig. 4).

Selecte	ed bond lengths/ Å	8·2CH ₂ Cl ₂	
	U1-O1/O5/O9/O13	2.312(2) / 2.303(2) / 2.305(2) / 2.289(2)	
	U1-O2/O6/O10/O14	2.423(3) / 2.399(3) / 2.387(3) / 2.402(3)	
P1-O2		1.497(3)	
	P1-C2	1.728(4)	
	P2-O6	1.502(3)	
	P2-C19	1.729(4)	
	P3-O10	1.500(3)	
	P3-C36	1.723(4)	
	P4-O14	1.510(3)	
	P4-C53	1.738(3)	
	C1-O1	1.295(5)	
	C18-O5	1.284(5)	
	C35-O9	1.285(4)	
C52-O13		1.281(4)	
OO _{che}	elate distances/ Å		
	01 02	2.811(4)	
	O5 ··· O6	2.803(4)	
	O9 ··· O10	2.876(4)	
	O13 ··· O14	2.863(4)	
Dihedr	al angles (°) of selected planes		
1.2	Plane (1): O1, U1, O2	2 17(14)	
1-2	Plane (2): O1, C1, C2, P1, O2	2.17(14)	
	Plane (3): O5, U1, O6		
3-4	Plane (4): O5, C18, C19, P2, O6	11.75(14)	
5 (Plane (5): O9, U1, O10	15 29(12)	
5-6	Plane (6): O9, C35, C36, P3, O10	15.38(13)	
7.0	Plane (7): O13, U1, O14	12.45(12)	
/-8	Plane (8): O13, C52, C53, P4, O14	12.45(12)	
9-10	Plane (9): O1, O2, O9	2 61(14)	
δ_1	Plane (10): O1, O9, O13	2.01(14)	
11-12 δ ₂	Plane (11): O5, O6, O14	A 75/1 A)	
	Plane (12): O6, O10, O14	4.73(14)	

Table S9: Selected bond lengths (Å), O^{...}O_{chelate} distances (Å), dihedral angles of the planes (°), and δ and ϕ^5 of the planes (°) of the U(IV) complex 8·2CH₂Cl₂.

13-14 δ ₃	Plane (13): O1, O5, O6	
	Plane (14): O1, O2, O6	48.20(12)
15-16	Plane (15): O9, O13, O14	
δ_4	Plane (16): O9, O10, O14	46.35(12)
17-18	Plane (17): O10, U1, O2	
φ ₁	Plane (18): O1, U1, O14	20.93(9)
19-20 φ ₂	Plane (19): O6, U1, O9	
	Plane (20): O5, U1, O13	19.46(8)

The numeration of the atoms in the planes refers to the numeration assigned to the atoms in the complex species (see **Fig. S34**).



Fig. S34 Molecular structure of $8 \cdot 2 CH_2 Cl_2$ with selected atom labelled. All hydrogen atoms and solvates are omitted for clarity; ellipsoids are drawn at 50% probability level.

Selected bond lengths/ Å		8		
U1A-01A/05A/09A/013A 2		2.260(4) / 2.278(4) / 2.301(4) / 2.357(4)		
U	11B-O1B/O5B/O9B/O13B	2.279(4) / 2.347(4) / 2.284(4) / 2.327(5)		
U	IA-02A/06A/010A/014A	2.443(4) / 2.416(4) / 2.385(4) / 2.393(4)		
U	1B-O2B/O6B/O10B/O14B	2.370(4) / 2.379(4) / 2.364(4) / 2.389(4)		
	P1A-O2A / P1B-O2B	1.503(5) / 1.510(5)		
	P1A-C2A / P1B-C2B	1.730(7) / 1.731(7)		
	P2A-O6A / P2B-O6B	1.513(5) / 1.508(5)		
	P2A-C19A / P2B-C53B	1.728(6) / 1.733(7)		
	P3A-O10A / P3B-O10B	1.510(4) / 1.505(4)		
	P3A-C36A / P3A-C19B	1.729(7) / 1.717(7)		
	P4A- O14A / P4B-O14B	1.505(5) / 1.510(5)		
P4A-C53A / P4B-C36B		1.729(6) / 1.727(7)		
	C1A-O1A / C1B-O1B	1.301(7) / 1.299(8)		
	C18A-O5A / C18B-O5B	1.298(8) / 1.295(7)		
	C35A-O9A / C35B-O9B	1.291(7) / 1.299(8)		
C	C52A-O13A / C52B-O13B	1.299(8) / 1.288(8)		
0Oc	helate distances/ Å			
(01A ··· O2A / O1B ··· O2B	2.784(6) / 2.774(6)		
0	5A ··· O6A / O5B ··· O10B	2.786(6) / 2.773(6)		
09	9A ··· O10A / O9B ··· O14B	2.739(7) / 2.760(6)		
01	3A ··· O14A / O13B ··· O6B	2.827(7) / 2.757(6)		
Dihee	dral angles (°) of selected planes			
1.2	Plane (1): O1A, U1A, O2A	0.2/2)		
1-2	Plane (2): O1A, C1A, C2A, P1A, O2A	9.2(2)		
	Plane (3): O5A, U1A, O6A			
3-4	Plane (4): O5A, C18A, C19A, P2A, O6A	6.5(2)		
5-6	Plane (5): O9A, U1A, O10A			
	Plane (6): O9A, C35A, C36A, P3A, O10A	2.3(2)		
7-8	Plane (7): O13A, U1A, O14A	12.2(2)		
	Plane (8): O13A, C52A, C53A, P4A, O14A	12.2(2)		

Table S10: Selected bond lengths (Å), O^{...}O_{chelate} distances (Å), dihedral angles of the planes (°), and δ and ϕ^5 of the planes (°) of the U(IV) complex **8**.

9-10	Plane (9): O1B, U1B, O2B			
	Plane (10): O1B, C1B, C2B, P1B, O2B	2.2(2)		
11 12	Plane (11): O5B, U1B, O10B			
9-10 11-12 13-14 15-16 $\overline{17-18}_{\delta_1}$ 19-20 δ_2 21-22 δ_3 23-24 δ_4 $\overline{25-26}_{\phi_1}$ 27-28 ϕ_2 29-30 δ_5 31-32 δ_6 33-34 δ_7 35-36 δ_8 $\overline{37-38}_{\phi_3}$	Plane (12): O5B, C18B, C19B, P3B, O10B	11.2(2)		
12 14	Plane (13): O9B, U1B, O14B			
13-14	Plane (14): O9B, C35B, C36B, P4B, O14B	8.02(2)		
15 16	Plane (15): O13B, U1B, O6B			
13-10	Plane (16): O13B, C52B, C53B, P2B, O6B	0.9(2)		
17 10	Plane (17): O1A, O2A, O5A			
δ_1	Plane (18): O1A, O5A, O6A	17.6(2)		
19-20	Plane (19): O9A, O10A, O14A			
9-10 11-12 13-14 15-16 $\overline{17-18}_{\delta_1}$ 19-20 δ_2 21-22 δ_3 23-24 δ_4 25-26 φ_1 27-28 φ_2 29-30 δ_5 31-32 δ_6 33-34 δ_7 35-36 δ_8 37-38 φ_3 39-40	Plane (20): O10A, O13A, O14A	9.7(2)		
21-22	Plane (21):O5A, O9A, O10A			
δ_3	Plane (22): O5A, O6A, O10A	45.62(19)		
23-24	Plane (23): O1A, O2A, O14A			
δ_4	Plane (24): O1A, O13A, O14A	44.85(19)		
25-26	Plane (25): O5A, U1A, O14A			
$\begin{array}{c} 13-14 \\ 15-16 \\ \hline \\ 17-18 \\ \delta_1 \\ 19-20 \\ \delta_2 \\ 21-22 \\ \delta_3 \\ 23-24 \\ \delta_4 \\ \hline \\ 25-26 \\ \phi_1 \\ 27-28 \\ \phi_2 \\ \hline \\ 29-30 \\ \delta_5 \\ \hline \\ 31-32 \\ \delta_6 \\ 33-34 \\ \delta_7 \\ 35-36 \\ \delta_8 \\ \hline \\ 37-38 \\ \phi_2 \\ \hline \\ \end{array}$	Plane (26): O6A, U1A, O13A	20.08(19)		
27-28	Plane (27): O2A, U1A, O9A			
φ ₁ 27-28 φ ₂	Plane (28): O1A, U1A, O10A	16.20(15)		
29-30	Plane (29): O1B, O2B, O9B			
δ_5	Plane (30): O1B, O9B, O14B	12.9(3)		
31-32	Plane (31): O5B, O6B, O10B	10.720		
δ_6	Plane (32): O6B, O10B, O13B	18.6(3)		
33-34	Plane (33): O1B, O2B, O6B			
δ_7	Plane (34): O1B, O6B, O13B	43.7(2)		
35-36	Plane (35): O9B, O5B, O10B			
δ_8	Plane (36): O9B, O10B, O14B	40.3(2)		
37-38 φ ₃	Plane (37): O1B, U1B, O10B			
	Plane (38): O2B, U1B, O5B	11.09(18)		
39-40	Plane (39): O6B, U1B, O9B			
39-40 φ ₄	Plane (40): O13B, U1B, O14B	15.86(16)		

The numeration of the atoms in the planes refers to the numeration assigned to the atoms in the complex species (see **Fig. S35**).



Fig. S35 Representation of the molecule A of the molecular structure of **8** with selected atom labelled. All hydrogen atoms are omitted for clarity; ellipsoids are drawn at 50% probability level.

EXAFS

The Np and UL_{III} -edge (17610 eV (Np), 17166 eV (U)) extended X-ray absorption fine structure (EXAFS) spectra were collected at the Rossendorf beamline (ROBL, BM20) at the European Synchrotron Radiation Facility (ESRF)¹⁶ under dedicated ring operation conditions of an electron energy of 6 GeV and a beam current of 200 mA. Two Rh-coated mirrors and a water cooled Si(111) double crystal monochromator were used for rejection of higher order harmonics and to monochromatize the incident white X-ray beam. The signal of the $L\alpha_{1,2}$ fluorescence lines was recorded with a 13-element Ge detector. For each sample multiple energy scans were performed in order to reach a sufficient signal-to-noise ratio after averaging (Np: 18 scans, U: 13 scans). For each energy scan the Y Kedge (17038 eV) absorption spectrum of a Y metal foil was measured simultaneously. The incident photon flux and the absorption spectrum of the Y metal foil was measured with gas filled ionization chambers. The data treatment, which includes the energy calibration, statistical weighting of the 13 fluorescence channels and their dead-time correction, averaging of the multiple sample scans, extraction of the EXAFS signal, and the shell fitting was accomplished with the program suite of EXAFSPAK.¹⁷ In order to calculate the photoelectron kinetic energy the ionization potential (E₀) of the Np and UL_{III} –edge was set arbitrary to 17620 eV and to 17185 eV, respectively, and was defined as a free parameter as $\Delta E_0 = E_0 - E_t (\Delta E_0 - \text{shift in energy threshold}, E_t - \text{theoretical ionization}$ potential) in the shell fit procedure. The expected EXAFS radial resolution is 0.12 Å and 0.13, as given by the maximum available k-interval of 1.0 - 13.6 Å⁻¹ (U) and 1.7 - 13.6 Å⁻¹ (Np), respectively. Theoretical scattering phase and amplitude functions where calculated with the ab-initio scattering code FEFF8.20¹⁸ based on the XRD structural data of 8·3CH₃CN·3H₂O and 9·3CH₃CN·3H₂O in the case of the U(IV) complexes and the Np(IV) complex, respectively.

Shell fit procedure

In equation (1) the structural parameter S_0^2 , N, σ^2 , and r can be fitted by common algorithms like the Levenberg-Marquardt algorithm. All other scattering phase and amplitude functions are available by theoretical *ab-initio* calculations on structural models or XRD data. For inspection of structural features a *k*-interval (k_{\min} , k_{\max}) of the EXAFS signal is Fourier-transformed by equation (2), whereby commonly the power spectrum is used (equation (3)) for visualization.

$$\chi(k) = S_0^2 \sum_{j=1}^{types} N_j S_j(k,r_j) F_j(k,r_j) e^{-2\sigma_j^2 k^2} e^{-2r_j/\lambda_j(k,r_j)} \frac{\sin(2kr_j + \varphi_j(k,r_j) + \varphi_c(k))}{kr_j^2}$$
(1)

 $\chi(k)$ - experimental EXAFS spectrum; S_0^2 - amplitude reduction factor; j – number of the backscattering atom; N_j – coordination number; $S_i(k,r_j)$ - total atomic loss factor; $F_j(k,r_j)$ - effective backscattering amplitude; σ_j - Debye-Waller factor; r_j – distance between absorbing atom and backscattering atom; $\lambda_j(k,r_j)$ – effective free path length of the photoelectron; $\phi_j(k,r_j)$ – phase shift of the photoelectron due to the interaction with the potential of the backscattering atom; $\phi_c(k)$ – phase shift of the photoelectron due to the interaction with the potential of the absorbing atom.

$$p(r) = \frac{1}{2\pi} \int_{k_{min}}^{k_{max}} \chi(k) e^{i(2kr)} dk$$

$$|p(r)| = \left\{ [p_{real}(r)]^2 + [p_{imag}(r)]^2 \right\}^{1/2}$$
(3)

An important property of the Fourier transform (FT) is possibility that structural features, which fall in a given *r*-interval (r_1, r_2) , can be back transformed (equations (4) – (6)).

$$z(k) = \frac{1}{2\pi} \int_{r_1}^{r_2} p(r) e^{-i(2kr)} dr$$
⁽⁴⁾

$$A(k) = \left\{ \left[z_{\text{real}}(k) \right]^2 + \left[z_{\text{imag}}(k) \right]^2 \right\}^{1/2}$$
(5)

$$\Phi(k) = \arctan\left\{\frac{z_{\text{imag}}(k)}{z_{\text{real}}(k)}\right\}, \text{ with } \Phi(k) = 2kr + \phi(k)$$
(6)

The EXAFS signal is then reconstructed as:

$$\chi_{r1-r2}(k) = A(k)sin(\Phi(k)) \tag{7}$$

The EXAFS equation (equation (1)) has some general drawbacks. The radial particle distribution function n(r), describing the density of interatomic distances in matter, is approximated by pure Gaussians shapes. The full width at the half maximum (FWHM) of n(r) is given by σ , which measures the radial structural and thermal atomic disorder. In the case that n(r) is non-Gaussian strong deviations between $\chi(k)$ and the fit can occur so that an accurate determination of structural parameter (r, N) is spoiled. Although, common shell fit analysis enables the inclusion of single scattering (SS) events, but also higher order scattering events, i.e. multiple scattering (MS) of the electron wave on more than one backscattering atom can be included. However, due to the strong angular dependency of the scattering amplitude of the MS contributions the results of the shell fit would be strongly influenced by the structural model used for prior calculation of the scattering phases and amplitudes. If the structural model is inaccurate, a shell fit will probably result in a poor description of the experimental $\chi(k)$, while unknown deviations between the true structure, probed by EXAFS, and the structural parameter gained by the shell fit must be considered.

Monte-Carlo (MC) simulation of EXAFS spectra

Basing on equation (1) the EXAFS oscillation of a backscattering atom *i* of the type *j* at the radial distance $r_{j,i}$ can be rewritten as:

$$\chi_{j}(k,r_{j,i}) = S_{0}^{2}S_{j}(k,r_{j,i})F_{j}(k,r_{j,i})e^{-2r_{j}/\lambda_{j}(k,r_{j,i})}\frac{\sin(2kr_{j,i}+\varphi_{j}(k,r_{j,i})+\varphi_{c}(k))}{kr_{j,i}^{2}}$$
(8)

were the radial absorber-backscatter distance is given by:

$$r_{j,i} = \left(dx_{j,i}^2 + dy_{j,i}^2 + dz_{j,i}^2\right)^{1/2}$$
(9)

For more than one molecule ($N_{molecule}$) and with N_{atoms}^{j} being the total number of atoms of type *j*, the total EXAFS signal becomes:

$$\chi(k) = \frac{1}{N_{molecule}} \sum_{j=1}^{types} \sum_{i=1}^{N_{atoms}} \chi_j(k, r_{j,i})$$
(10)

The radial particle distribution function $n_i(r)$ can be calculated for each type of backscattering atoms by

$$n_j(r) = \frac{1}{N_{molecule}} \sum_{j=1}^{types} \sum_{i=1}^{N_{atoms}} \delta(r - \bar{r}_{j,i})$$
(11)

where δ is the Kronecker delta function and $\overline{r}_{j,i} = r$ if $r - dr/2 < r_{j,i} < r + dr/2$, while dr is the predefined bin-width used for the histogram of $n_i(r)$.

By inspection of equations (8) and (9) it is obvious that for each type of an atom and for each set of Cartesian coordinates a theoretical EXAFS spectrum can be calculated, owing the possibility to calculate for a large number of molecules the total EAXFS signal (equation (10)). Moreover, opposite to shell fit methods, for each arbitrary atomic configuration the scattering phases and amplitudes can be precisely calculated for SS and MS events by *ab-initio* calculations. At the moment, only MC approaches enables both, the calculation of n(r) without assumptions about its shape and the inclusion of MS, while their scattering phase and amplitude functions can be updated in dependence on the structural changes during the simulation. The MC approach does not include gradient descent algorithms, like the Levenberg-Marquardt algorithm as used in shell fit procedures, which does not guaranty the convergence to a global minimum in the case of an extensive multi-shell fit scenario.

MC starts with a statistical ensemble of metal complexes which is gained by the replication of the initial complex structure. For the whole set of backscattering atoms the theoretical $\chi(k)_{theo}$ is calculated (equation (10)) and compared with the experimental $\chi(k)$. If a randomly picked atom is moved by a random distance from its origin the new $\chi(k)_{theo}$ will result in a better or poorer fit. If only the proposed movements, which lead to an improvement of the fit would be accepted, then an over-fitting will take place, while n(r) become unstable in respect to the experimental error. Moreover, the risk of a convergence into a local minimum could not be avoided. Here we use the Markov chain Monte-Carlo (MCMC) algorithm¹⁹ where the Markov chain is accomplished by introducing the Metropolis algorithm,²⁰ which measures for each proposed move of an atom its relative probability to be accepted or rejected. As an acceptance criterion, the relative probability of a move is calculated according the Boltzmann factor for which the difference in energy is replaced by the change of the sum of squared errors, caused by the particular move, and by replacing $k_b T$ (k_b - Boltzmann constant, T - temperature) by a multiple of the expected experimental error. Various studies have shown that this kind of *importance sampling* is well suited for analysing XAS spectra and data from other spectroscopic techniques.²¹⁻²⁶ After numerous atomic movements the Markov chain converges to a stationary distribution for which $\chi(k)_{theo}$ should fluctuate around a constant average in the size of the expected experimental error.

The Metropolis algorithm was included in our MCTFA code ²⁷ as proposed by Gurman et al.²⁶ and McGreevy²⁴ For SS paths the $\chi_j(k,r_{j,i})$ is calculated according our procedure as described in the literature.²⁸ MS paths up to the 4th order are calculated and updated during the simulation with a pre-defined spatial precision by using FEFF8.20.¹⁸

Shell Fit

Figure S36 displays the most important scattering paths for shell fit, which are also summarized in Table S11.



Fig. S36 Structural fragment taken from XRD structure of **8**·3CH₃CN·3H₂O and **9**·3CH₃CN·3H₂O. Ellipsoids are drawn at 50% probability level.

Path	r / Å	n
U/Np-O1	2.30/2.30	4
U/Np-O2	2.39/2.38	4
U/Np-C1	3.37/3.34	4
U/Np-O1-C1	3.48/3.46	8
U/Np-O1-C1-O1	3.60/3.58	4
U/Np-P	3.66/3.63	4
U/Np-O2-P	3.78/3.75	8
U/Np-O2-P-O2	3.89/3.87	4
U/Np-C2	3.93/3.91	4

Table S11 Single scattering (SS) and multiscattering (MS) paths used for the shell fit.

r – radial distance, n – coordination number.

Since the three mixed isomers of the U(IV) complex ($8 \cdot 3CH_3CN \cdot 3H_2O$, $8 \cdot CH_2Cl_2$, 8) are structurally similar with 9 we fitted the spectra (**Fig. S37**) of both samples with identical settings for the EXAFS structural parameter. All coordination numbers were fixed according to the structural model (**Table S11**). For paths with r > r_{Np/U-O2} several parameter are highly correlated and were kept constant or linked during the fit to avoid physically unrealistic results (**Table S12**). For the first shell (maximum at 1.8 Å in the FT), which consists of O1 and O2, all parameter,

except for *n*, could be fitted freely and resulted in physically reliable σ^2 for both samples. In the case of both samples the resulting *r* agrees with the average of the XRD structural data up to a maximum deviation of 0.035 Å and 0.031 Å for the U-P and the Np-P interaction, respectively. Note that the common uncertainty in determination of radial distances by EXAFS is 0.02 Å²⁹, hence the shell fit radial distances support the XRD structural data.



Fig. S37 Experimental (black) k³-weighted EXAFS spectra of the U(IV) and Np(IV) complexes in toluene solution with shell fit (red) and corresponding FT (right). Level of experimental error (green line). The three marked MS peaks in the FT are FT-filtered in range of 3.94 Å - 5.40 Å and FT back transformed (blue).

Path/sample	п	r /Å	$\Delta r/\AA$	σ^2/\AA^2	ΔE_0 /Ev
8 ·3CH ₃ CN·3H ₂ O, 8 ·	·CH ₂ Cl ₂ , 8				
U01	4	2.30(1)	0.00 ^a	0.0059(9)	1.9(3)
U–O2	4	2.39(1)	0.00 ^a	0.008(2)	1.9/
U–C1	4	3.361(8)	0.022 ^a	0.0034(5)	1.9/
U01C1	8	3.480/		0.0034/	1.9/
U-01-C1-01	4	3.598/		0.0034/	1.9/
U–P	4	3.647(5)	0.035ª	0.0064(8)	1.9/
U–O2–P	8	3.72(1)		0.0064/	1.9/
UO2PO2	4	3.83/		0.0064/	1.9/
U–C2	4	3.97(1)	0.00 ^a	0.005	1.9/
9·3CH ₃ CN·3H ₂ O					
Np-O1	4	2.278(7)	0.020	0.0052(9)	9.87(4)
Np-O2	4	2.369(9)	0.008	0.006(1)	9.87/
Np-C1	4	3.337(5)	0.007	0.0017(4)	9.87/
NpO1C1	8	3.455/		0.0017/	9.87/
Np-O1-C1-O1	4	3.572/		0.0017/	9.87/
Np-P	4	3.605(6)	0.031	0.0060(9)	9.87/
Np-O2-P	8	3.69(2)		0.0060/	9.87/
Np-O2-P-O2	4	3.809/		0.0060/	9.87/
Np-C2	4	3.94(1)	-0.03	0.005	9.87/

Table S12 EXAFS shell fit parameter for the U(IV) and Np(IV) complexes in solution.

/-linked parameter. n – coordination number, r – radial distance, σ^2 – Debye-Waller factor, ΔE_0 - shift in energy threshold, Δr - modulus of the difference between shell fit distances and the corresponding distances supplied by the XRD structure. ^a – calculated with average distances taken from XRD structural data of 8·3CH₃CN·3H₂O, 8·CH₂Cl₂, and 8, respectively. Estimated standard deviations of the variable parameter as given from EXAFSPAK in parenthesis.

The shell fitting might suffer from several drawbacks so that slight differences between the fit and the experiment can be caused by an intrinsic property of the shell fit method. However, the XRD structural data are not only supported by the shell fitted structural parameter, since the spectra of both systems are very similar and since they show the same strong structural features, which were taken into account for shell fitting. Especially the back transformation of the FT interval in the range of 3.94 Å - 5.40 Å (**Fig. S37**) contains a negative oscillation, which is responsible for the fine structure observed at 6.8 Å^{-1} and 6.6 Å^{-1} in the case of both systems, respectively. Following the numbering of the MS peaks as given in **Fig. S37** and with **Fig. S35**, the back transformed EXAFS signal consists of: peak 1: U/Np–O1–N2; peak 2: U/Np–O1–C4, U/Np–C2–C3; peak 3: U/Np–C1–N1. In total and with inclusion of SS and all related MS paths, twelve additional paths would be necessary for the description of these spectral features. Note that all these MS paths show focusing effect, hence a strong spectral amplitude due

to the small scattering angle and are therefore of high relevance. However, the scattering amplitudes and phases of these paths are very similar so that the fitted structural parameter would have strong correlations, thus a shell fit would be not conclusive.

Monte-Carlo simulation

The starting structural model for U(IV) complex was constructed by using the XRD structural data of the three isomers 8·3CH₃CN·3H₂O, 8·CH₂Cl₂ and 8 (Fig. S38), while for the Np(IV) complex we used the XRD structural data of 9.3CH₃CN·3H₂O (Fig. S39). Hydrogen atoms where removed due to their small EXAFS scattering amplitude, and only atoms in radial distances less than 7 Å were included, so that the models contain 258 and 87 backscattering atoms in the case of the U(IV) complexes and the Np(IV) complex, respectively. To get a statistically proper distribution of n(r) the models were replicated 200 times, hence in total the models for the U(IV) complexes and the Np(IV) complex consist of 51600 and 17400 backscattering atoms, respectively. For all types of backscattering atoms all SS paths where calculated with a resolution of 0.02 Å up to 6 Å, while the MS paths up to the fourth order with scattering angles $\geq 125^{\circ}$ and $r \leq 6$ Å were included and automatically updated by ab-inito calculations performed by FEFF8.20.¹⁸ The backscattering atoms were moved successively along a random vector with a random length of 0.03 Å. The convergence of the Markov chain was reached after $\sim 1 \times 10^7$ $\sim 4x10^6$ atomic movements in the case of the U(IV) and the Np(IV) system, respectively. For receiving a smooth n(r) and a stable average structure the MC was performed for the U(IV) and the Np(IV) system further up to 1.6×10^7 and 6×10^6 atomic movements, respectively, for which all atomic coordinates were stored 6 times in between. In total about 1.1×10^7 and 1.3×10^6 MS events were automatically calculated and updated for the simulation of the U(IV) complexes and the Np(IV) complex, respectively.

The MC resulted in the 3D particle distribution shown in **Fig. S38** and **Fig. S39** and in the fit of the experimental EXAFS spectra shown in **Fig. S40**. According equation (9) and equation (11) the radial pair distributions for all scattering atoms, inclusively for all MS paths are available in 2- and 3D Euclidean space (not shown here). The quality of the MC fit outperforms the shell fit procedure (**Fig. S37**) as visible in the low residual, which mainly consists in contributions stemming from effects of background subtraction, hence visible in the lower *r*-range of the FT (**Fig. S40**). As expected, the influence of the MS contributions above ~2.5 Å is very strong as shown in **Fig. S40**.



Fig. S38 3D Particle distribution gained by MC simulation of the U(IV) complexes solved in toluene solution. **8**·3CH₃CN·3H₂O (upper left), **8**·CH₂Cl₂ (upper right), and **8** (lower middle). CH₃CN entity in **8**·3CH₃CN·3H₂O omitted for better visualization.



Fig. S39 3D Particle distribution gained by MC simulation of the Np(IV) complex ($9 \cdot 3CH_3CN \cdot 3H_2O$) solved in toluene solution.

The fine structure at 6.8 Å⁻¹ and 6.6 Å⁻¹ in the k-space, which is typical for both systems, is well reproduced by the MC fit (vertical line **Fig. S40**), hence supports again that the structures in the liquid phase are up to 6 Å in accordance with the solid phase structures provided by the XRD measurements.



Fig. S40 Experimental (black) k^3 -weighted EXAFS spectra of the U(IV) and Np(IV) complexes acquired as toluene solution with MC-fit (red), corresponding FT (right) and MS contributions (blue). Residual between MC fit and the experimental shown in the FT (green).

Moreover, the average atomic positions gained by the MC simulations agree very well with the XRD structures (**Fig. S41**), thus MC supports the findings of NMR. The analysis of the first and second moments of n(r), i.e. r and σ^2 , are given in **Table S13**. The structural parameter gained by the MC simulation are in favourable agreement with the shell fit results (**Table S12**) and they match the XRD structural data within the common error in determination of radial distances of 0.02 Å by EXAFS ²⁹, except for the U–P interaction. Only for this interaction small structural deviations from the XRD structural data are observed. In the case of U(IV) the presence of the three isomers is supported by the decreased amplitude of the MS events above ~2.5 Å in the FT (**Fig. S40**), since the complex structures of 8·CH₂Cl₂ and 8 are structurally more disordered than the complex structures of the complexes 8·3CH₃CN·3H₂O and 9·3CH₃CN·3H₂O, respectively.



Fig. S41 Structure of $8 \cdot 3$ CH₃CN $\cdot 3$ H₂O (upper left), $8 \cdot$ CH₂Cl₂ (upper right), and 8 (lower middle) provided by XRD structural data (coloured) and the determined average atomic positions by using MC simulation (yellow). CH₃CN entity in $8 \cdot 3$ CH₃CN $\cdot 3$ H₂O omitted for better visualization.

Path/sample	n	$r_{\rm MC}/r_{\rm XRD}$ /Å	$\Delta r/Å$	$\sigma^2/\text{\AA}^2$	$\Delta E_0/\mathrm{Ev}$]
$8 \cdot 3 CH_3 CN \cdot 3H_2 O$,	8·CH ₂ Cl ₂ ,	8				n
U-O1/2	8.0(8)	2.346(2)/2.347 ^a	0.001	0.0064(3)	1.9	
U-C1	4.0(7)	3.3747(4)/3.3832 ^a	0.0085	0.007(2)	1.9	
U-P	4.0(7)	3.6416(5)/3.6820a	0.0404	0.0039(7)	1.9	1 10 m
U–C2	4.2(6)	3.9830(8)/3.9694 ^a	-0.0136	0.0052(8)	1.9	
9-3CH ₃ CN-3H ₂ O				•		$\left \begin{array}{c} v \\ r \end{array} \right $
Np-O1/2	8(1)	2.329(7)/2.338a	0.009	0.0066(4)	9.87	
Np-C1	4.0(9)	3.353(2)/3.344	-0.009	0.006(2)	9.87	
Np-P	4.0(7)	3.5894(4)/3.6364	0.0470	0.0039(9)	9.87	
Np-C2	4.4(9)	3.938(3)/3.910	-0.028	0.004(1)	9.87	ra

Table S13 Structural parameter of the U(IV) and Np(IV) complexes in toluene solution determined by the MC simulation.

l distances determined by MC simulation and XRD measurements, $\Delta r = r_{XRD} - r_{MC}$, ^a – calculated with average distances from XRD structural data of **8**·3CH₃CN·3H₂O, **8**·CH₂Cl₂, and **8**, respectively. σ^2 – Debye-Waller factor, ΔE_0 - shift in energy threshold.

Fourier filtering algorithm

As a third method we used a Fourier filtering algorithm (FFA), which was recently developed with the aim to compare experimental EXAFS spectra directly with theoretical EXAFS spectra calculated by FEFF using structures as gained by quantum chemical calculations or supplied by XRD measurements ³⁰. However, a direct comparison of the theoretical with the experimental spectrum is spoiled due to the fact that the amplitude function of the theoretical spectrum is not comparable with the experimental amplitude function since the σ^2 are unknown *a priory*.

In the first step of the FFA theoretical EXAFS spectra are calculated with the FEFF code basing on XRD structural data. In the case of the three mixed U(IV) isomers and with the XRD structural data of $8 \cdot 3 \text{CH}_3 \text{CN} \cdot 3 \text{H}_2 \text{O}$, $8 \cdot \text{CH}_2 \text{Cl}_2$, and **8** the resulting three theoretical spectra are averaged and weighted by a factor of 1/3, while for the Np(IV) complex the theoretical spectrum is calculated by using the XRD structural data from $9 \cdot 3 \text{CH}_3 \text{CN} \cdot 3 \text{H}_2 \text{O}$. In order to adjust the theoretical EXAFS spectra in respect to the experimental Fermi level, the FEFF code allows the implementation of ΔE_0 provided by the shell fit.

By taking equation (5) and (6) into account, the phase and amplitude functions are available for the experimental (exp.) and the theoretical (theo.) EXAFS spectrum basing on the XRD structural data. Applying

$$\chi_{r1-r2}^{\text{theo.}*}(k) = \frac{A_{exp.}(k)}{A_{\text{theo.}}(k)} A_{\text{theo.}}(k) \sin(\Phi_{\text{theo.}}(k))$$
(12)

the "experimental" EXAFS spectrum based on the XRD structural data $(\chi_{r_1-r_2}^{theo.*}(k))$ can be calculated, where equation (13) represents a simplification of equation (12).

$$\chi_{r_1 - r_2}^{\text{theo.}\,*}(k) = A_{exp.}(k) \sin(\Phi_{\text{theo.}}(k))$$
(13)

Thus, equation (13) enables the direct comparison between a theoretical and an experimental EXAFS spectrum, since the amplitude function of the theoretical spectrum is replaced with the amplitude function of the experimental EXAFS spectrum. The FT filtered *r*-range was 0.0 Å \leq r \leq 5.5 Å for both theoretical EXAFS spectra.



Fig. S42 Experimental (black) k³-weighted EXAFS spectra of the U(IV) complexes and the Np(IV) complex acquired as toluene solution and the modified theoretical spectra (red (left)). Corresponding Fourier transform (FT) (right).

The result of the FFA is shown in **Fig. S42**. For both complexes in solution the theoretical EXAFS spectra are in very good agreement with the experimental EXAFS spectra. Moreover, the negative oscillation at 6.8 Å⁻¹ and at 6.6 Å⁻¹, stemming from the typical MS paths U/Np–O1–N2 (1), U/Np–O1–C4 (2) and U/Np–C2–C3 (3) (FT in **Fig. S42**), is well reproduced. Thus, as a third independent method the FFA demonstrates again that the molecular structures of the complexes in solution match the XRD structural data.

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