# Electronic Supporting Information (ESI)

# Heterobimetallic Pd/Mn and Pd/Co Complexes as Efficient and Stereoselective Catalysts for the Sequential Cu-free Sonogashira Coupling–Alkyne Semi-Hydrogenation Reactions

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### 1. Ligands

1.1 **L2** (Molecular Structure in the Solid State; NMR Spectra (<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>31</sup>P{<sup>1</sup>H}, <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C HMBC, <sup>1</sup>H-<sup>13</sup>C HSQC, <sup>1</sup>H-<sup>1</sup>H NOESY))





Figure S1. Molecular structure of ligand L2 (hydrogen atoms are omitted for clarity; thermal ellipsoids are set at the 50% probability level).



Figure S2. <sup>1</sup>H NMR spectrum of ligand L2 in CD<sub>3</sub>CN at 25 °C.



Figure S3.  $^{13}C{^1H}$  NMR spectrum of ligand L2 in CD<sub>3</sub>CN at 25 °C.



Figure S4.  $^{31}P\{^{1}H\}$  NMR spectrum of ligand L2 in CD\_3CN at 25 °C.



Figure S6.  $^{1}H$ - $^{13}C$  HSQC spectrum of ligand L2 in CD<sub>3</sub>CN at 25 °C.



Figure S7. <sup>1</sup>H-<sup>13</sup>C HMBC spectrum of ligand L2 in CD<sub>3</sub>CN at 25 °C.



Figure S8. <sup>1</sup>H-<sup>1</sup>H NOESY spectrum of ligand L2 in CD<sub>3</sub>CN at 25 °C.





Figure S9. <sup>1</sup>H NMR spectrum of ligand L3 in CDCl<sub>3</sub> at 25 °C.





40

60

80

100

20 ppm

ò

-20

-40

-60



Figure S12. <sup>1</sup>H-<sup>1</sup>H COSY spectrum of ligand L3 in CDCl<sub>3</sub> at 25 °C.



Figure S13.  $^{1}$ H- $^{13}$ C HSQC spectrum of ligand L3 in CDCl<sub>3</sub> at 25 °C.







Figure S15.  $^{1}H^{-1}H$  NOESY spectrum of ligand L3 in CDCl<sub>3</sub> at 25 °C.

1.3 L5 (Molecular Structure in the Solid State; NMR Spectra (<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>31</sup>P{<sup>1</sup>H}))



Figure S16. Molecular structure of L5 (hydrogen atoms are omitted and *tert*-butyl groups are drawn as wireframes for clarity; thermal ellipsoids are set at the 50% probability level).

N1



Figure S17. <sup>1</sup>H NMR spectrum of ligand L5 in CDCl<sub>3</sub> at 25 °C.



Figure S19.  $^{31}\text{P}\{^{1}\text{H}\}$  NMR spectrum of ligand L5 in CDCl3 at 25 °C.

### 2. Monometallic Pd Complexes

2.1 **PdL2** (NMR Spectra (<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>31</sup>P{<sup>1</sup>H}, <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C HMBC, <sup>1</sup>H-<sup>13</sup>C HSQC))



L2

PdL2

Scheme S4. Synthesis of PdL2.



Figure S20. <sup>1</sup>H NMR spectrum of PdL2 in DMSO-d<sub>6</sub> at 25 °C.







Figure S22.  ${}^{31}P{}^{1}H$  NMR spectrum of PdL2 in DMSO-d<sub>6</sub> at 25 °C.



**Figure S23**. <sup>1</sup>H-<sup>1</sup>H COSY spectrum of **PdL2** in DMSO-d<sub>6</sub> at 25 °C.



Figure S24.  $^{1}$ H- $^{13}$ C HSQC spectrum of PdL2 in DMSO-d<sub>6</sub> at 25 °C.



Figure S25.  $^1\text{H-}{^{13}\text{C}}$  HMBC spectrum of PdL2 in DMSO-d\_6 at 25 °C.

2.2 **PdL3'** (Molecular Structure of **PdL3'** and [PdCl(**L3'**)]·HCl in the Solid State; NMR Spectra (<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>31</sup>P{<sup>1</sup>H}, <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C HMBC, <sup>1</sup>H-<sup>13</sup>C HSQC, <sup>1</sup>H-<sup>1</sup>H NOESY))



Scheme S5. Synthesis of PdL3'.



**Figure S26**. Molecular structure of **PdL3'**·CHCl<sub>3</sub>·2H<sub>2</sub>O (solvent molecules and hydrogen atoms are omitted and *tert*-butyl groups are drawn as wireframes for clarity; thermal ellipsoids are set at the 50% probability level).



**Figure S27**. Molecular structure of [PdCl(**L3**')]·HCl (hydrogen atoms (except H1) are omitted and *tert*-butyl groups are drawn as wireframes for clarity; thermal ellipsoids are set at the 50% probability level).



Figure S28. <sup>1</sup>H NMR spectrum of PdL3' in CDCl<sub>3</sub> at 25 °C.



Figure S30.  $^{31}\text{P}\{^{1}\text{H}\}$  NMR spectrum of PdL3' in CDCl3 at 25 °C.



Figure S31. <sup>1</sup>H-<sup>1</sup>H COSY spectrum of PdL3' in CDCl<sub>3</sub> at 25 °C.



**Figure S32**. <sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **PdL3'** in CDCl<sub>3</sub> at 25 °C.







**Figure S34**. <sup>1</sup>H-<sup>1</sup>H NOESY spectrum of **PdL3'** in CDCl<sub>3</sub> at 25 °C.

# 2.3 PdL5' (NMR Spectra (<sup>1</sup>H, <sup>13</sup>C{<sup>1</sup>H}, <sup>31</sup>P{<sup>1</sup>H}))







Figure S37.  $^{31}\text{P}\{^{1}\text{H}\}$  NMR spectrum of PdL5' in CDCl3 at 25 °C.

3. Monometallic Mn and Co Complex

3.1 MnL4 (Synthesis; NMR Spectra (<sup>1</sup>H, <sup>19</sup>F{<sup>1</sup>H}))



Scheme S7. Synthesis of MnL4.

MnCl<sub>2</sub> (148 mg, 1.18 mmol, 1.00 eq.) and L4 (250 mg, 1.18 mmol, 1.00 eq.) were dissolved in EtOH (30 mL). The orange solution was stirred for 3 h at room temperature forming an orange precipitate. The solid was filtered off, washed with EtOH (2 x 20 mL) and dried *in vacuo* to give [MnCl<sub>2</sub>(L4)] as an orange solid (304 mg, 76%). Ag(OTf) (464 mg, 1.81 mmol, 2.01 eq.) and CH<sub>3</sub>CN (20 mL) were added to [MnCl<sub>2</sub>(L4)]. The mixture was stirred for 12 h at room temperature. The orange solution was filtered over Celite, and the solvent was removed *in vacuo*. The orange solid was washed with Et<sub>2</sub>O (10 mL) and *n*-pentane (10 mL). The solid was dried under vacuum at 40 °C for 3 h to give MnL4 as an orange solid (410 mg, 81%). Elemental analysis: C<sub>14</sub>H<sub>12</sub>F<sub>6</sub>MnN<sub>4</sub>O<sub>6</sub>S<sub>2</sub>, calculated (%): C 29.74, H 2.14, N 9.91, found (%): C 29.66, H 1.82, N 10.08. HRMS (ESI pos., CH<sub>3</sub>CN): *m/z* calculated for [M–CF<sub>3</sub>SO<sub>3</sub>]<sup>+</sup>: 415.996, found: 415.993 (100%); calculated for [[M]<sub>2</sub>–CF<sub>3</sub>SO<sub>3</sub>]<sup>+</sup>: 980.944, found: 980.940 (50%). Selected ATR-IR:  $\tilde{v}$  (cm<sup>-1</sup>) = 3045 (w, vC–H), 1604 (m, vC=N/vC=C), 1476 (m, vC=C), 1440 (m, vC=C), 1210 (s, v<sub>5</sub>CF<sub>3</sub>), 1163 (s, v<sub>as</sub>CF<sub>3</sub>), 1027 (s, v<sub>5</sub>SO<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN):  $\delta$  (ppm) = 14.9 (br). Due to the paramagnetic Mn<sup>II</sup> centre, some protons were not observed in the <sup>1</sup>H NMR spectrum. <sup>19</sup>F{<sup>1</sup>H} NMR (377 MHz, CD<sub>3</sub>CN):  $\delta$  (ppm) = -55.7 (br).



Figure S39.  $^{19}F{^{1}H}$  NMR spectrum of MnL4 in CD<sub>3</sub>CN at 25 °C.

#### 3.2 CoL4 (Synthesis; NMR Spectra (<sup>1</sup>H, <sup>19</sup>F{<sup>1</sup>H}))



Scheme S8. Synthesis of CoL4.

A solution of CoCl<sub>2</sub>·6H<sub>2</sub>O (710 mg, 3.00 mmol, 1.00 eq.) in EtOH (30 mL) was added dropwise to a solution of L4 (640 mg, 3.00 mmol, 1.00 eq.) in EtOH (30 mL). The dark green solution was stirred for 3 h at room temperature. The green precipitate was filtered, washed with EtOH (2 x 20 mL) and dried *in vacuo* to give [CoCl<sub>2</sub>(L4)] as a dark green solid (739 mg, 72%). Ag(OTf) (150 mg, 0.29 mmol, 2.01 eq.) was added to a suspension of [CoCl<sub>2</sub>(L4)] (100 mg, 0.29 mmol 1.00 eq.) in CH<sub>3</sub>CN (10 mL). The mixture was stirred for 12 h at room temperature. The orange solution was filtered over Celite, and the solvent was removed *in vacuo*. The dark red solid was washed with CH<sub>2</sub>Cl<sub>2</sub> (5 mL), Et<sub>2</sub>O (5 mL) and *n*-pentane (10 mL). The solid was dried under vacuum at 40 °C for 3 h to give CoL4 as a dark red, hygroscopic solid (134 mg, 81%). Elemental analysis: C<sub>14</sub>H<sub>12</sub>CoF<sub>6</sub>N<sub>4</sub>O<sub>6</sub>S<sub>2</sub>·2CH<sub>3</sub>CN·0.4C<sub>5</sub>H<sub>12</sub>, calculated (%): C 35.31, H 3.38, N 12.35, found (%): C 35.62, H 3.12, N 12.48. HRMS (ESI pos., CH<sub>3</sub>CN): *m/z* calculated for [M–CF<sub>3</sub>SO<sub>3</sub>]<sup>+</sup>: 419.991, found: 419.990 (100%). Selected ATR-IR:  $\tilde{v}$  (cm<sup>-1</sup>) = 3068 (w, vC–H), 2963 (w, vC–H), 1601 (m, vC=N/vC=C), 1470 (m, vC=C), 1440 (m, vC=C), 1238 (s, v<sub>as</sub>SO<sub>3</sub>), 1223 (s, v<sub>s</sub>CF<sub>3</sub>), 1145 (s, v<sub>as</sub>CF<sub>3</sub>), 1026 (s, v<sub>s</sub>SO<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN):  $\delta$  (ppm) = 11.53 (s, 1H), 10.90 (s, 3H), 3.54 (s, 1H). Due to the paramagnetic Co<sup>II</sup> centre, some protons were not observed in the <sup>1</sup>H NMR spectrum. <sup>19</sup>F{<sup>1</sup>H} NMR (377 MHz, CD<sub>3</sub>CN):  $\delta$  (ppm) = -78.5 (s).



Figure S40.  $^{1}$ H NMR spectrum of CoL4 in CD<sub>3</sub>CN at 25 °C.



Figure S41.  $^{19}\text{F}\{^{1}\text{H}\}$  NMR spectrum of CoL4 in CD\_3CN at 25 °C.

## 4. Heterobimetallic Pd Complexes

4.1 **CoPdL2** (NMR Spectra ( ${}^{1}H$ ,  ${}^{19}F{}^{1}H$ },  ${}^{31}P{}^{1}H$ }))



Figure S43.  ${}^{19}F{}^{1}H$  NMR spectrum of CoPdL2 in CD<sub>3</sub>CN at 25 °C.







4.2 **MnPdL3'** (NMR Spectra ( ${}^{1}H$ ,  ${}^{19}F{}^{1}H$ },  ${}^{31}P{}^{1}H$ }))

PdL3'

MnPdL3'











# 4.3 CoPdL3' (NMR Spectra ( ${}^{1}H$ , ${}^{19}F{}^{1}H$ }, ${}^{31}P{}^{1}H$ }))











Figure S49.  $^{19}F\{^{1}H\}$  NMR spectrum of CoPdL3' in CD<sub>3</sub>CN at 25 °C.



Figure S50.  $^{31}P\{^{1}H\}$  NMR spectrum of CoPdL3' in CD<sub>3</sub>CN at 25 °C.

#### 4.4 Dihedral Angle

Complex	Dihedral angle [deg]
$[MnPdCl_2(OTf)_2(L1)]$	1.2
$[CoPdCl_2(OTf)(L1)]_2(OTf)_2$	10.0
$[CoPdCl_2(CH_3CN)(L2)]_2(OTf)_4$	22.8
$[MnPd(L3')(OTf)_3(CH_3CN)]$	26.7
[CoPd( <b>L3'</b> )(OTf) <sub>3</sub> (CH <sub>3</sub> CN)]	23.8

Table S1. Dihedral angles [deg] between two planes formed by N1,N2,N4,M1 (M = Mn<sup>II</sup>, Co<sup>II</sup>) and N5,N7/C15,P1,Pd1.



**Figure S51**. Molecular structure of complex [MnPdCl<sub>2</sub>(OTf)<sub>2</sub>(L1)] (only the oxygen atoms of the triflate anions are shown; solvent molecules and hydrogen atoms are omitted for clarity). The dihedral angle (1.2°) was determined between the two planes of N, N, N, Mn (red) and N, N, P, Pd (blue).



**Figure S52**. Molecular structure of complex  $[CoPdCl_2(OTf)(L1)]_2(OTf)_2$  (only the monomer of the dimeric structure and oxygen atoms of the coordinating triflate anions are shown; solvent molecules, noncoordinating triflate anions, and hydrogen atoms are omitted for clarity). The dihedral angle (10.0°) was determined between the two planes of *N*,*N*,*N*,*Co* (red) and *N*,*N*,*P*,*Pd* (blue).

### [CoPdCl<sub>2</sub>(CH<sub>3</sub>CN)(L2)]<sub>2</sub>(OTf)<sub>4</sub>



#### N,N,P,Pd plane (blue)

**Figure S53**. Molecular structure of complex  $[CoPdCl_2(CH_3CN)(L2)]_2(OTf)_4$  (only the monomer of the dimeric structure and nitrogen atom of the coordinating CH<sub>3</sub>CN molecule are shown; solvent molecules, noncoordinating triflate anions, and hydrogen atoms are omitted for clarity). The dihedral angle (22.8°) was determined between the two planes of *N*,*N*,*N*,*Co* (red) and *N*,*N*,*P*,*Pd* (blue).



**Figure S54**. Molecular structure of complex  $[MnPd(L3')(OTf)_3(CH_3CN)]$  (only the oxygen atoms of the coordinating triflate anion and nitrogen atom of the coordinating CH<sub>3</sub>CN molecule are shown; solvent molecules and hydrogen atoms are omitted for clarity). The dihedral angle (26.7°) was determined between the two planes of N,N,N,Mn (red) and N,C,P,Pd (blue).



**Figure S55**. Molecular structure of complex  $[CoPd(L3')(OTf)_3(CH_3CN)]$  (only the oxygen atoms of the coordinating triflate anion and nitrogen atom of the coordinating CH\_3CN molecule are shown; solvent molecules and hydrogen atoms are omitted for clarity). The dihedral angle (26.7°) was determined between the two planes of *N*,*N*,*N*,*Co* (red) and *N*,*C*,*P*,*Pd* (blue).

### 5. Catalysis

### 5.1 General Procedure

A mixture of bromobenzene (0.25 mmol, 1.0 eq.), phenylacetylene (0.38 mmol, 1.5 eq.), base (0.50 mmol, 2.0 eq.), precatalyst (0.015 mmol, 6 mol%) and dry, degassed solvent (1.0 mL) were heated under Ar for 4-24 h. The reaction process was monitored by GC-MS. After full conversion of bromobenzene,  $NH_3 \cdot BH_3$  (0.25 mmol, 1.0 eq.) was added, and the reaction was continued for 14-24 h at 50 °C. The yields have been determined by GC-MS using naphthalene as an internal standard (4  $\mu$ L of the reaction solution have been diluted with 1.000 mL of a naphthalene stock solution in acetone).

#### 5.2 Optimisation of the Reaction Conditions

**Table S2**. Optimisation of the reaction conditions for the Sonogashira cross-coupling reaction. A mixture of bromobenzene (0.25 mmol, 1.0 eq.), phenylacetylene (0.38 mmol, 1.5 eq.), base (0.50 mmol, 2.0 eq.), precatalyst **CoPdL3'** (0.015 mmol, 6 mol%) and dry, degassed solvent (1.0 mL) was heated under Ar for 4-24 h (DABCO = 1,4-diazabicyclo[2.2.2]octane; DIPEA = N,N-diisopropylethylamine).



Entry	Solvent	Catalyst loading	Base	Amount of Base [eq.]	Temperature [°C]	Yield <sup>[a]</sup> [%] after 4 h	Yield <sup>[a]</sup> [%] after 24 h
_		[mol%]					
1	CH₃CN	6	DABCO	2.0	90	99	99
2	CH₃CN	6	K <sub>2</sub> CO <sub>3</sub>	2.0	90	29	99
3	CH₃CN	6	Cs <sub>2</sub> CO <sub>3</sub>	2.0	90	5	17
4	CH₃CN	6	NEt₃	2.0	90	8	22
5	CH₃CN	6	DIPEA	2.0	90	8	19
6	Toluene	6	DABCO	2.0	110	23	68
7	Dioxane	6	DABCO	2.0	110	78	84
8	THF	6	DABCO	2.0	70	89	90
9	<sup>t</sup> BuOH	6	DABCO	2.0	90	62	62
10	DMF	6	DABCO	2.0	110	65	65
11	<sup>t</sup> BuOH	6	$K_2CO_3$	2.0	90	18	99
12	CH₃CN	3	DABCO	2.0	90	80	81
13	CH₃CN	6	DABCO	1.0	90	72	78

<sup>[a]</sup> The yields have been determined by GC-MS using naphthalene as an internal standard (4  $\mu$ L of the reaction solution have been diluted with 1.000 mL of a naphthalene stock solution in acetone).
**Table S3.** Comparison of monometallic and heterobimetallic  $Pd^{II}$  complexes as precatalyst for the transfer semihydrogenation. A mixture of diphenylacetylene (0.25 mmol, 1.0 eq.), NH<sub>3</sub>·BH<sub>3</sub> (0.25 mmol, 1.0 eq.), precatalyst (0.015 mmol, 6 mol%) and dry, degassed CH<sub>3</sub>CN (1.0 mL) was heated to 50 °C under Ar for 6-24 h.

3a	precatalyst NH <sub>3</sub> :E CH <sub>3</sub> CN, 50 <sup>c</sup>	(6 mol%) 3H <sub>3</sub> ℃, 6-24 h		
			Z/E/alkane ratio	
	Entry	Precatalyst	Yield [%] <sup>[a]</sup> after 6 h <i>Z/E/</i> alkane (after 24 h)	
	1	CoPdL3'	10/60/30	
	2	PdL3'	3/31/66	
	3	CoL4/PdL5'	32/68/-	
	4	CoL4	9/6/- (24/58/-)	
	5	PdL5'	21/50/28	
	6	-	-/-/- (-/3/-)	

<sup>[a]</sup> The yields have been determined by GC-MS using naphthalene as internal standard (4  $\mu$ L of the reaction solution have been diluted with 1.000 mL of a naphthalene stock solution in acetone).

#### 5.3 Reaction Process







Figure S57. Reaction process followed by GC-MS using naphthalene as an internal standard (Int. St.).



Figure S58. Reaction process followed over time by <sup>1</sup>H NMR spectroscopy in CD<sub>3</sub>CN at room temperature.

#### <sup>31</sup>P{<sup>1</sup>H} NMR (162 MHz, CD<sub>3</sub>CN, room temperature)



Figure S59. Reaction process followed over time by <sup>31</sup>P{<sup>1</sup>H} NMR spectroscopy in CD<sub>3</sub>CN at room temperature.

#### <sup>11</sup>B NMR (128 MHz, CD<sub>3</sub>CN, room temperature)



**Figure S60**. Reaction process of the transfer semi-hydrogenation followed over time by <sup>11</sup>B NMR spectroscopy in CD<sub>3</sub>CN at room temperature.

#### 5.4 Homogeneous or Heterogeneous?

Furthermore, we have been interested in the true nature of the catalyst. To exclude the formation of catalytically active nanoparticles, several poisoning and kinetic studies have been performed as well as analytical methods (dynamic light scattering (DLS) and high-resolution transmission electron microscopy/energy-dispersive X-ray spectroscopy (TEM/EDX)) to detect potential nanoparticles.<sup>1</sup> The reaction process was monitored over time (Figure S56). The reaction kinetics have not shown a sigmoidal curve indicating that there is no induction period to form catalytically active nanoparticles. In parallel, poisoning studies (mercury test, quantitative ligand poisoning, Crabtree's test<sup>2</sup>) have been carried out for the Sonogashira cross-coupling reaction (Figure S61). The mercury test resulted in a complete inhibition of the catalyst; therefore, a drop of Hg was added directly at the beginning and after 30 min. This result would mean that the precatalyst decomposes into catalytically active nanoparticles which would be inhibited by the amalgam formation. However, the reliability of the mercury test is doubtful for palladacycles.<sup>3</sup> Therefore, the stability of **CoPdL3'** was investigated by heating the complex with mercury for 2 d. No decomposition was observed but the addition of DABCO and mercury resulted in decomposition of the complex (Figure S62, S63). For this reason, the mercury test seems to be a false positive. The addition of 0.1 eq. PMe<sub>2</sub>Ph per CoPdL3' did not inhibit the catalytic performance. However, the addition of 0.5 eq. PMe<sub>2</sub>Ph per CoPdL3' caused a complete inhibition. This might underline the homogeneous nature of the catalytically active species. The limitations of this test are reaction temperatures over 50 °C because PMe<sub>2</sub>Ph might dissociate from the heterogeneous catalyst surface. On the other hand, the Crabtree's test is used to poison a homogeneous catalyst. However, a possible homogeneous catalyst was not inhibited by the addition of dibenzo[a,e]cyclooctatetraene (DCT) even if the reaction mixture was stirred for 3 h at room temperature before continuing with heating at 90 °C. In general, the DCT did not react with CoPdL3' at room temperature for 24 h nor at 90 °C for 24 h (Figure S64, S65). Therefore, the Crabtree's test is not suitable for this complex. Moreover, the Maitlis' test was performed after completion of the Sonogashira cross-coupling reaction. The hot reaction mixture was filtered over Celite and fresh substrates and solvent were added. The filtrate was still catalytically active giving diphenylacetylene with a yield of 71% after 4 h. Since these tests could not fully prove the nature of the catalytically active species, DLS and TEM-EDX measurements were carried out. The DLS measurement has shown nanoparticles with a size of 10 nm and 26 nm though the count rate was relatively low (77 kcps, should be 100–500 kcps (kilo counts per second)) and the baseline index was 5.7 (should be  $\geq$ 8). Both values suggest a low quality of the DLS measurement and a possible contamination by dust (Chapter 5.4.2). Therefore, we tried to track potential nanoparticles by TEM-EDX measurements, but no nanoparticles have been found (<u>Chapter 5.4.3</u>). Finally, the reaction process was followed by  ${}^{1}H$  and  ${}^{31}P{}^{1}H$  NMR spectroscopy (Figure S58, S59) indicating the complex identity during the Sonogashira cross-coupling. However, the phosphorus signal shifts from 180.4 ppm to 101.6 ppm during the transfer semihydrogenation which is comparable to the monometallic complex PdL3' (100.2 ppm). This suggests the potential loss of Co from the heterobimetallic complex. It is worth mentioning, that the complexes CoPdL3', PdL3', CoL4 and PdL5' have been tested for the direct transfer hydrogenation of diphenylacetylene but none of the complexes have shown similar reactivity and selectivity as CoPdL3' after the Sonogashira cross-coupling reaction (Table S3). Therefore, we propose that the catalytically active species for the transfer semi-hydrogenation was formed during the Sonogashira cross-coupling reaction. However, there is no clear evidence if Co or Pd is catalysing the transfer semi-hydrogenation. Nevertheless, we conclude that the catalytically active species is homogeneous.

#### 5.4.1 Poisoning Studies



**Figure S61**. Catalyst poisoning during the Sonogashira cross-coupling reaction with Hg (330 eq. per **CoPdL3'**), PMe<sub>2</sub>Ph (0.1 eq. and 0.5 eq. per **CoPdL3'**) and DCT (6.0 eq. per **CoPdL3'**). The yields have been determined by GC-MS using naphthalene as internal standard.

#### Hg Poisoning



Figure S62. <sup>1</sup>H NMR spectra of the complex CoPdL3' reacting with Hg and DABCO.

#### <sup>31</sup>P{<sup>1</sup>H} NMR (162 MHz, CD<sub>3</sub>CN, room temperature)



Figure S63. <sup>31</sup>P{<sup>1</sup>H} NMR spectra of the complex CoPdL3' reacting with Hg and DABCO.

#### **DCT** Poisoning





Figure S64. <sup>1</sup>H NMR spectra of the complex CoPdL3' not reacting with DCT.

#### <sup>31</sup>P{<sup>1</sup>H} NMR (162 MHz, CD<sub>3</sub>CN, room temperature)



Figure S65. <sup>31</sup>P{<sup>1</sup>H} NMR spectra of the complex CoPdL3' not reacting with DCT.

### 5.4.2 DLS (dynamic light scattering)

Measurement	Parame	eters:							
Temperature = 25.0 deg.		deg. C	g. C		Runs Completed				
Liquid		= Uns	pecified		Run Dur	= 00	= 00:00:30 = 00:02:30		
Viscosity		= 0.33	4 cP			apsed Time			= 00
Ref.Index I	Fluid	= 1.34	4		Average	Count Rate	= 76	.9 kcp	)S
Angle		= 90.0	0		Ref.Inde	ex Real	= 1.5	590	
Wavelengt	h	= 659.	0 nm		Ref.Inde	ex Imag	= 0.0	000	
Baseline		= Auto	(Slope Analysis	)	Dust Filt	er Setting	= 5.	00	
RC_78_H12s	t_dust5_	100x dilute	d (Combined)			9 T T			
Effective	e Dia	meter	: <mark>16.4</mark> nm			-			
Polydispersity: 0.1			0.158		C(τ)				
Baseline Index: 5		5.7/ 37.5	0%		10 102	103 104	105	106	
Elapsed Time: 0		00:02:30		1.1	10 10-	τ (μ <b>S</b> )	100	100	
					Correlation Function			ction	
Run	Eff. D	iam. (nm)	Half Wid	ith (nm)	Po	olydispersity	Base	line Ir	ndex
1		16.0	6	. 6		0.171	2.4/	43	. 06%
2		16.0	6	. 6		0.171	8.9/	32	. 64%
3		16.6	7	7.2		0.189	6.3/	37	. 19%
5		16.4	6	. 3		0.146	5.5/	36	. 11%
Mean		16 3	6	7		0 170	6.4	/ 37	5.0%
Std. Error		0.1	0	.2		0.007	1.2/	1 1	.69
Combined		16.4	6	. 5		0.158	5.7/	37	. 50%
Elapsed Time Mean Diam. Rel. Var. Skew	<ul> <li>⇒ 00:02</li> <li>19.6</li> <li>0.182</li> <li>-0.23</li> </ul>	2:30 nm 7 99	100 75				Ì		
			50 ten			-	()		
			 25		1				
			20						
			0	and a second	- XII				
				5.0					50.0
					Multime	Diameter	r (nm)		
					wondifie		noation		
d(nm)	G(d)	C(d)	d(nm)	G(d)	C(d	)	d(nm) G	(d)	C(d)
7.3	0	0	12.6	0	43	3	21.7	0	43
7.7	0	0	13.2	0	43	5	22.8	0	43
0.1	0	U	13.9	0	43	2	23.9	21	47

1.1	0	0	10.2	0	40	22.0	U	40
8.1	0	0	13.9	0	43	23.9	27	47
8.5	0	0	14.6	0	43	25.2	62	58
8.9	23	4	15.4	0	43	26.4	100	76
9.4	57	14	16.1	0	43	27.8	82	90
9.9	81	28	17.0	0	43	29.2	47	98
10.4	58	38	17.8	0	43	30.6	9	100
10.9	24	43	18.7	0	43	32.2	0	100
11.4	0	43	19.7	0	43	33.8	0	100
12.0	0	43	20.6	0	43	35.5	0	100

5.4.3 Transmission Electron Microscopy/Energy-dispersive X-ray Spectroscopy (TEM/EDX analysis)



kV: 200; Mag: 80000; Takeoff: 12.4; Live Time (s): 42; Amp Time (μs): 3.84; Resolution (eV): 126.8.



Element	Weight %	Atomic %	Net Int.	Net Error%	6kAB Factor <sup>1</sup>
СК	21.7	36.3	2140.7	0.4	1.27
N K	9.2	13.1	1210.8	0.6	0.94
ОК	27.0	33.9	3364.7	0.5	1
FΚ	2.0	2.1	154.1	10.8	1.61
Mg K	0.9	0.7	96.3	4.2	1.13
S K	2.5	1.6	276.8	1.7	1.12
Pd L	1.9	0.4	69.9	2.0	3.36
Со К	34.9	11.9	2079.4	0.5	2.09

<sup>&</sup>lt;sup>1</sup> The number kAB, normally called the k-factor, relates the compositions of A and B. It is not a proper constant, but it is referred to as a sensitivity factor. It depends on the particular AEM system, the voltage, and analysis conditions in general.



kV: 200; Mag: 100000; Takeoff: 12.4; Live Time (s): 45.7; Amp Time (μs): 3.84; Resolution (eV): 126.8.



sec: 45.7	907 Cnts	11.920 keV	Det: Octane T Optima 30 Windowless	wless

Element	Weight %	Atomic %	Net Int.	Net Error%	6kAB Factor <sup>1</sup>
СК	20.2	38.6	1176.6	0.5	1.27
N K	7.6	12.5	594.0	0.9	0.94
ОК	21.5	30.8	1581.9	0.7	1
Mg K	0.0	0.0	2.4	100.0	1.13
S K	0.4	0.3	23.3	15.4	1.12
Cl K	2.7	1.7	166.9	2.2	1.19
Со К	23.7	9.2	834.4	0.9	2.09
Br K	23.9	6.9	420.6	1.4	4.2



kV: 200; Mag: 40000; Takeoff: 12.4; Live Time (s): 45.9; Amp Time (μs): 3.84; Resolution (eV): 126.8.



Lsec: 45.9	709 Cnts	11.920 keV	Det: Octane	T Optima	30 Windowless

Element	Weight %	Atomic %	Net Int.	Net Error%	6kAB Factor <sup>1</sup>
СК	20.1	38.5	1032.3	0.6	1.27
N K	6.9	11.3	473.8	1.1	0.94
ОК	21.9	31.5	1423.4	0.7	1
Mg K	0.0	0.0	0.0	100.0	1.13
S K	0.4	0.3	26.0	12.4	1.12
CI K	2.9	1.9	156.9	2.3	1.19
Со К	25.7	10.0	798.3	0.9	2.09
Br K	22.2	6.4	344.0	1.5	4.2

#### 5.5 Substrate Scope

General procedure for sequential Sonogashira cross-coupling reaction followed by transfer semihydrogenation. **4a-4m**:

Substituted bromobenzene derivative (0.5 mmol, 1 eq.), phenylacetylene (1.0 mmol, 110  $\mu$ l, 2 eq.), **CoPdL3'** (0.03 mmol, 30 mg, 6 mol%), DABCO (1.0 mmol, 112 mg, 2 eq.) and acetonitrile (2 mL) were added. The reaction mixture was degassed by freeze-pump-thaw and heated in an oil bath at 90 °C. After 4 hours, the progress of the reaction was checked using GC-MS which was followed by addition of ammonia borane (31 mg, 1.0 mmol, 2 eq.) mixed in 1:1 ratio with MgSO<sub>4</sub> to ease the process of weighing in the solid and additionally acetonitrile (2 mL) was added. The flask was covered completely in aluminium foil and was heated in a water bath at 50 °C for 15 hours. The reaction was monitored by GC-MS, the reaction mixture was passed over Celite and the precipitate was washed with dichloromethane (3 × 10 mL). The crude reaction mixture filtrate was purified using a Biotage Isolera (25 g SNAP Ultra cartridge, EtOAc (EA)/*n*-hexane (Hex) or *n*-pentane) to afford the corresponding compounds (**4a-4m**) and isolated yields are reported.

Procedure for synthesis of combretastatin A-4 (**4n**) by sequential Sonogashira cross-coupling reaction followed by transfer semi-hydrogenation:

In a Schlenk flask under argon, 5-iodo-2-methoxyphenol (125 mg, 0.5 mmol, 1 eq.), 5-ethynyl-1,2,3trimethoxybenzene (192 mg, 1.0 mmol, 2 eq.), **CoPdL3'** (0.03 mmol, 30 mg, 6 mol%), DABCO (1.0 mmol, 112 mg, 2 eq.) and acetonitrile (2 mL) were added. The reaction mixture was degassed by freeze-pump-thaw and heated in an oil bath at 90 °C. After 5 hours, ammonia borane (31 mg, 1.0 mmol, 2 eq.) was weighed in and additionally acetonitrile (2 mL) was added. The flask was covered completely in aluminium foil and was heated in a water bath at 50 °C for 24 hours. The reaction mixture was cooled down and passed over Celite and washed with dichloromethane (3 × 10 mL). The crude reaction mixture was purified using a silica gel column eluted with *n*-pentane/ethyl acetate (7:3) to afford the combretastatin A-4 as a viscous oil that solidified on cooling (82 mg, 26 mmol, 52%). 4a: (Z)-1,2-diphenylethene4



Product was isolated via column chromatography (Hex) as a colourless liquid (71 mg, 0.39 mmol, 79%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  = 7.28 – 7.13 (m, 10H), 6.59 (s, 2H).

 $^{13}\text{C}\{^1\text{H}\}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  = 137.2, 130.2, 128.9, 128.2, 127.1.

#### 4b: (Z)-1-methyl-4-styrylbenzene<sup>4</sup>



Product was isolated via column chromatography (Hex/EA – 200:3) as a colourless liquid (64 mg, 0.33 mmol, 66%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.29 – 7.16 (m, 5H), 7.16 – 7.12 (m, 2H), 7.02 (d, J=7.9, 2H), 6.55 (s, 2H), 2.30 (s, 3H).

 $^{13}C\{^{1}H\}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 137.6, 137.0, 134.4, 130.3, 129.7, 129.0, 129.0, 128.9, 128.3, 127.1, 21.4.

4c: (Z)-1-methyl-3-styrylbenzene<sup>4</sup>



Product was isolated via column chromatography (Hex/EA – 50:1) as a colourless liquid (69 mg, 0.35 mmol, 71%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.26 - 7.16 (m, 5H), 7.11 - 6.98 (m, 4H), 6.56 (s, 2H), 2.25 (s, 3H).

 $^{13}C\{^{1}H\}$  NMR (101 MHz, CDCl\_3)  $\delta$  (ppm) = 137.9, 137.5, 137.3, 130.5, 130.2, 129.7, 129.0, 128.3, 128.2, 128.0, 127.2, 126.0, 21.5.

4d: (Z)-1-methyl-2-styrylbenzene<sup>4</sup>



Product was isolated via column chromatography (Hex/EA – 50:1) as a colourless liquid (65 mg, 0.33 mmol, 67%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.22 - 6.98 (m, 9H), 6.62 (m, 2H), 2.26 (s, 3H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 137.1, 137.0, 136.1, 130.5, 130.0, 129.5, 128.9, 128.8, 128.0, 127.2, 127.0, 125.7, 19.8.

4e: (Z)-1-chloro-4-styrylbenzene<sup>4</sup>

Product was isolated via column chromatography (Hex/EA – 49:1) as a colourless liquid (82 mg, 0.38 mmol, 76%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.28 – 7.10 (m, 9H), 6.62 (d, J = 12.2 Hz, 1H), 6.52 (d, J = 12.2 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 136.8, 135.6, 132.7, 130.9, 130.2, 128.9, 128.8, 128.4, 128.3, 127.3.

4f: (Z)-1-fluoro-4-styrylbenzene<sup>4</sup>



Product was isolated via column chromatography (Hex/EA – 100:3) as a colourless liquid (62 mg, 0.31 mmol, 63%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.28 – 7.15 (m, 7H), 6.90 (t, J = 8.8 Hz, 2H), 6.59 (d, J = 12.2 Hz, 1H), 6.54 (d, J = 12.2 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 162.0 (d,  $J_{CF}$  = 246.6 Hz), 137.2, 133.3 (d,  $J_{CF}$  = 3.5 Hz), 130.7 (d,  $J_{CF}$  = 7.9 Hz), 130.4 (d,  $J_{CF}$  = 1.3 Hz), 129.2, 129.0, 128.4, 127.3, 115.3 (d,  $J_{CF}$  = 21.4 Hz).

<sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>)  $\delta$  = -114.7 (tt, *J*<sub>FH</sub> = 9.0, 5.4 Hz).

4g: (Z)-trimethyl(4-styrylphenyl)silane<sup>5</sup>



Product was isolated via column chromatography (Hex/EA – 100:3) as a colourless liquid (81 mg, 0.32 mmol, 64%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 7.32 – 7.26 (m, 2H), 7.23 – 7.07 (m, 7H), 6.56 – 6.44 (m, 2H), 0.16 (s, 9H).

 $^{13}C\{^{1}H\}$  NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 139.5, 137.7, 137.5, 133.3, 130.5, 130.4, 129.0, 128.4, 128.3, 127.2, –1.0.

4h: (Z)-ethyl 4-styrylbenzoate<sup>6</sup>



Product was isolated via column chromatography (Hex/EA – 100:3) as a white solid (93 mg, 0.37 mmol, 74%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 7.89 (d, *J* = 8.4 Hz, 1H), 7.30 (d, *J* = 8.2 Hz, 1H), 7.24 - 7.19 (m, 2H), 6.71 (d, *J* = 12.2 Hz, 1H), 6.61 (d, *J* = 12.3 Hz, 1H), 4.36 (q, *J* = 7.1 Hz, 1H), 1.38 (t, *J* = 7.1 Hz, 2H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 166.4, 142.0, 136.7, 132.1, 129.5, 129.3, 128.9, 128.8, 128.3, 128.3, 127.5, 60.9, 14.3.

4i: (Z)-1-styrylnaphthalene<sup>7</sup>



Product was isolated via column chromatography (*n*-pentane) as a colourless oil (79 mg, 0.34 mmol, 69%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 8.11 – 8.06 (m, 1H), 7.92 – 7.84 (m, 1H), 7.83 – 7.71 (m, 1H), 7.57 – 7.41 (m, 2H), 7.41 – 7.30 (m, 2H), 7.13 – 7.02 (m, 6H), 6.84 (d, *J* = 12.2 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 136.7, 135.2, 133.6, 131.9, 131.5, 129.0, 128.4, 128.3, 127.9, 127.4, 127.0, 126.4, 125.9, 125.9, 125.5, 124.8.

4j: (Z)-2-styrylthiophene<sup>8</sup>



Product was isolated via column chromatography (*n*-pentane) as a colourless oil (59 mg, 0.32 mmol, 63%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 7.40 – 7.29 (m, 5H), 7.11 (dd, *J* = 5.1, 1.2 Hz, 1H), 6.99 (d, *J* = 3.6 Hz, 1H), 6.90 (dd, *J* = 5.1, 3.6 Hz, 1H), 6.72 (d, *J* = 11.9 Hz, 1H), 6.60 (d, *J* = 11.9 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 139.8, 137.3, 128.9, 128.8, 128.5, 128.1, 127.5, 126.4, 125.5, 123.3.

**4k**: (*Z*)-5-styrylpyrimidine<sup>4</sup>



Product was isolated via column chromatography (Hex/EA – 7:3) as a yellow oil (67 mg, 0.37 mmol, 74%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 9.01 (s, 1H), 8.56 (s, 2H), 7.32 – 7.17 (m, 6H), 6.89 (d, *J* = 12.1 Hz, 1H), 6.47 (d, *J* = 12.1 Hz, 1H).

 $^{13}C{^{1}H} NMR (101 MHz, CDCl_3) \delta (ppm) = 156.9, 156.7, 135.9, 135.1, 131.3, 129.0, 128.6, 128.2, 122.8.$ 

41: (Z)-2-styrylpyridine9



Product was isolated via column chromatography (Hex/EA – 8:2) as a yellow oil (61 mg, 0.33 mmol, 67%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 8.58 – 8.55 (m, 1H), 7.44 – 7.38 (m, 1H), 7.27 – 7.19 (m, 5H), 7.16 – 7.11 (m, 1H), 7.09 – 7.03 (m, 1H), 6.81 (d, J = 12.4 Hz, 1H), 6.67 (d, J = 12.4 Hz, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 156.5, 149.7, 136.8, 135.8, 133.4, 130.7, 129.0, 128.4, 127.7, 124.0, 121.9.

4m: (E)-4-styrylaniline<sup>[1]</sup>



Product was isolated via column chromatography (*n*-pentane/EA – 7:3) as a yellow solid (61 mg, 0.31 mmol, 62%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) = 7.50 – 7.42 (m, 2H), 7.36 – 7.28 (m, 4H), 7.25 – 7.16 (m, 1H), 7.02 (d, *J* = 16.3 Hz, 1H), 6.91 (d, *J* = 16.3 Hz, 1H), 6.70 – 6.60 (m, 2H), 3.71 (br, 1H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 146.1, 137.9, 128.7, 128.6, 128.0, 127.7, 126.9, 126.1, 125.1, 115.2.

4n: (Z)-2-methoxy-5-(3,4,5-trimethoxystyryl)phenol - Combretastatin A-4<sup>[10]</sup>



Product was isolated via column chromatography (*n*-pentane/EA-7:3) as a viscous oil that solidified on cooling (82 mg, 26 mmol, 52%).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ (ppm) = 6.92 (d, *J* = 2.0 Hz, 1H), 6.80 (dd, *J* = 8.4, 2.0 Hz, 1H), 6.73 (d, *J* = 8.4 Hz, 1H), 6.53 (s, 2H), 6.44 (dd, *J* = 12.2 Hz, 2H), 5.51 (s, 1H), 3.87 (s, 3H), 3.84 (s, 3H), 3.70 (s, 6H).

<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>) δ (ppm) = 153.0, 145.9, 145.4, 137.3, 132.9, 130.8, 129.6, 129.2, 121.3, 115.2, 110.5, 106.2, 61.1, 56.1.

4a: (Z)-1,2-diphenylethene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)





4b: (Z)-1-methyl-4-styrylbenzene



### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



4c: (Z)-1-methyl-3-styrylbenzene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)



60

4d: (Z)-1-methyl-2-styrylbenzene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)







<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)





4e: (Z)-1-chloro-4-styrylbenzene



### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)

136.8 132.7 132.7 130.9 130.9 130.9 128.9 128.8 128.3 128.3 128.3



4f: (Z)-1-fluoro-4-styrylbenzene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)





<sup>19</sup>F NMR (377 MHz, CDCl<sub>3</sub>)





4g: (Z)-trimethyl(4-styrylphenyl)silane



### <sup>1</sup>H NMR (400 Hz, CDCl<sub>3</sub>)



4h: (Z)-ethyl 4-styrylbenzoate



### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



4i: (Z)-1-styrylnaphthalene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

### 8.10 8.10 8.10 8.10 8.10 8.10 8.11 8.11 8.12 8.12 8.13 8.14 8.15 9.15</l



<sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)



4j: (Z)-2-styrylthiophene



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

### 



# <sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)



4k: (Z)-5-styrylpyrimidine



### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



# $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl<sub>3</sub>)



4I: (Z)-2-styrylpyridine



#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

### 



### <sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)



4m: (E)-4-styrylaniline



# <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



4n: (Z)-2-methoxy-5-(3,4,5-trimethoxystyryl)phenol - Combretastatin A4



#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



### <sup>13</sup>C{<sup>1</sup>H} NMR (101 MHz, CDCl<sub>3</sub>)

![](_page_70_Figure_5.jpeg)

### 6. Electrochemistry

**Table S4**. Electrochemical potentials of compounds L3, PdL3', MnPdL3' and CoPdL3' were determined by cyclic voltammetry (CV). L3 was measured in THF/0.1 mol·L<sup>-1</sup> [N(*n*Bu)<sub>4</sub>]PF<sub>6</sub> and the complexes PdL3', MnPdL3' and CoPdL3' were measured in CH<sub>3</sub>CN/0.1 mol·L<sup>-1</sup> [N(*n*Bu)<sub>4</sub>]PF<sub>6</sub> with a scan rate of 100 mV·s<sup>-1</sup> at room temperature under N<sub>2</sub> atmosphere. The scan range was from 1 V to -2 V. Ferrocene was used as internal standard at the end of the CV experiment to reference the reported potentials to the FcH/[FcH]<sup>+</sup> couple.

Compound	E <sub>PC</sub> in V (I in μA)	E <sub>PA</sub> in V (I in μA)	$E_{1/2}$ in V
L3	–1.60 (–5.2); –1.75 (–5.6)	0.71 (22.1); 0.81 (21.5)	
PdL3'		0.78 (24.0)	
MnPdL3'	-1.51 (-22.2);		
CoPdL3'	-1.41 (-16.6); -1.93 (-26.8)		
	-1.14 (-20.0)	-1.04 (5.6)	-1.09

![](_page_71_Figure_3.jpeg)

Figure S66. Cyclic voltammograms of the ligand L3 in THF/0.1 mol·L<sup>-1</sup> [N(nBu)<sub>4</sub>]PF<sub>6</sub> and the complexes PdL3', MnPdL3' and CoPdL3' in CH<sub>3</sub>CN/0.1 mol·L<sup>-1</sup> [N(nBu)<sub>4</sub>]PF<sub>6</sub> with a scan rate of 100 mV·s<sup>-1</sup>. The arrow represents the respective starting potential and scan direction.


**Figure S67**. Cyclic voltammogram of **CoPdL3'** in [N(nBu)<sub>4</sub>]PF<sub>6</sub>/CH<sub>3</sub>CN showing the assumed Co<sup>II</sup>/Co<sup>I</sup> quasi-reversible process at different scan rates. The arrow represents the respective starting potential and scan direction.

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