

### Supplemental Data

TITLE: Joint Effects of Antibiotics and Quorum Sensing Inhibitors  
On Resistance Development in Bacteria

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JOURNAL: Environmental Science: Processes & Impacts

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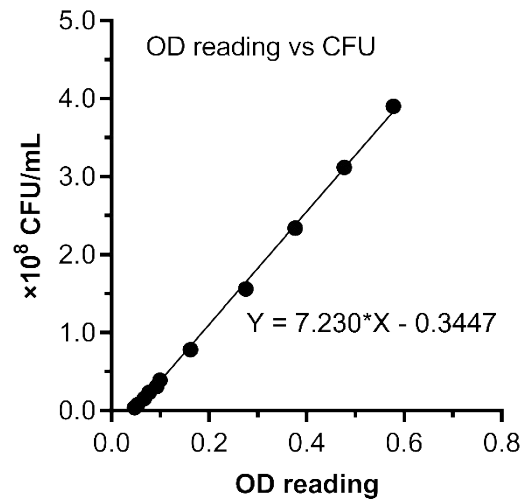


Fig. S1 Linear relationship between the optical density (OD) readings and CFU of *E. coli* in the present study. OD readings were recorded a Multiskan™ FC Microplate Photometer (ThermoFisher Scientific, USA), while CFU was determined by serially diluting the bacterial suspensions in LB medium and plating them on LB agar plates.

REF	TCGGGTGTACGTCTCGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGGA <sup>302</sup>
Control	TCGGGTGTACGTCTCGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGAA
REF	TACACGGACAGAGAGTTGATCAGACCGATGTTTCGGACCTTCA <sup>401</sup> GGGGTTTCGATTGGACATACGC
TET/CHL	TACACGGACAGAGAGTTGATCAGACCGATGTTTCGGACCTTCAAGGGTTTCGATTGGACATACGC
REF	ACACGGACAGAGAGTTGATCAGACCGATGTTTCGGACCTTCAGGGG <sup>397</sup> TTTCGATTGGACATACGCG
CIN	ACACGGACAGAGAGTTGATCAGACCGATGTTTCGGACCTTCAGGGGGTTTCGATTGGACATACGCG
REF	AATCTCAGACAGCGGGTTGTTCTGGTCCATAAACT <sup>247</sup> GAGACAGCTGGCTGGAACCGAAGA ACTCT
NPO/TET+NPO	AATCTCAGACAGCGGGTTGTTCTGGTCCATAAACTCAGACAGCTGGCTGGAACCGAAGA ACTCT
REF	CGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGGAGATAC <sup>295</sup> GACGTT
TET+CIN	CGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGGAGATACCACGTT
REF	CGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGG <sup>302</sup> GAGATACGACGTT
CHL+CIN	CGAACTTCGAAGCCTGCACGTTACGGGTCAGACCGCCTGGGCCGAGTGCGTAGATACGACGTT
REF	GCGTAATCTCAGACAGCGGGTTGTTCTGG <sup>257</sup> TCCATAAACT <sup>248</sup> TGAGACAGCTGGCTGGAACCGAAGAA
CHL+NPO	GCGTAATCTCAGACAGCGGGTTGTTCTGGCCATAAAACAGAGACAGCTGGCTGGAACCGAAGAA

Fig. S2 Comparisons between DNA sequence of reference (1057 bp of  $\beta$  subunit in RpoB) and samples. The red letters and figures mean the base changes and mutation sites, respectively. REF: reference, TET: tetracycline, CHL: chloramphenicol, CIN: cinnamaldehyde, NPO: 4-Nitropyridine-N-oxide.

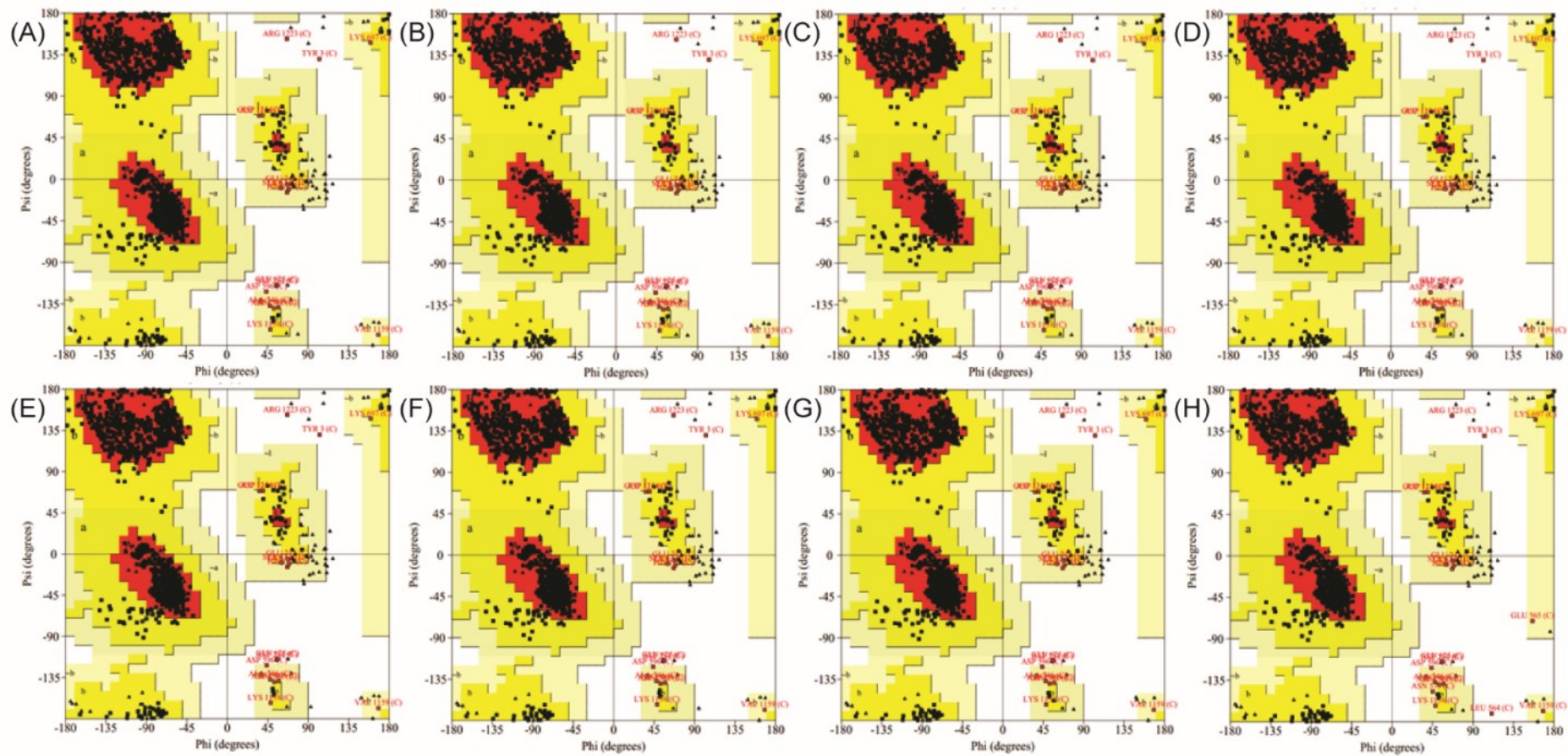


Fig. S3. Ramachandran plots of predicted RpoB mutants. (A) Wild type, (B) Q513E, (C) Q513L & D516G, (D) R529G, (E) S531F, (F) S531Y, (G) T563P (H) P564L

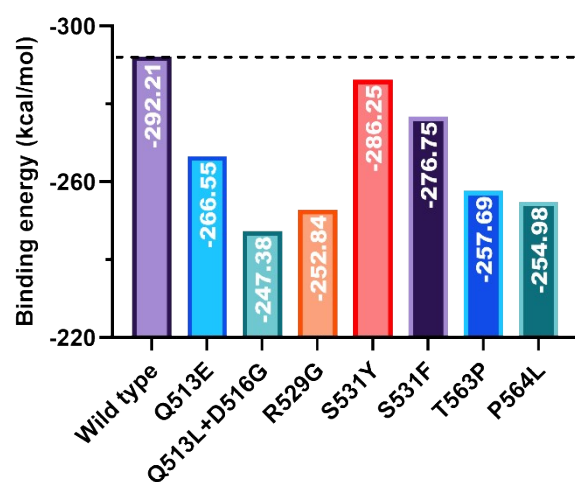


Fig. S4. Binding energies between the rifampicin molecule and the wild type and mutated RpoB.



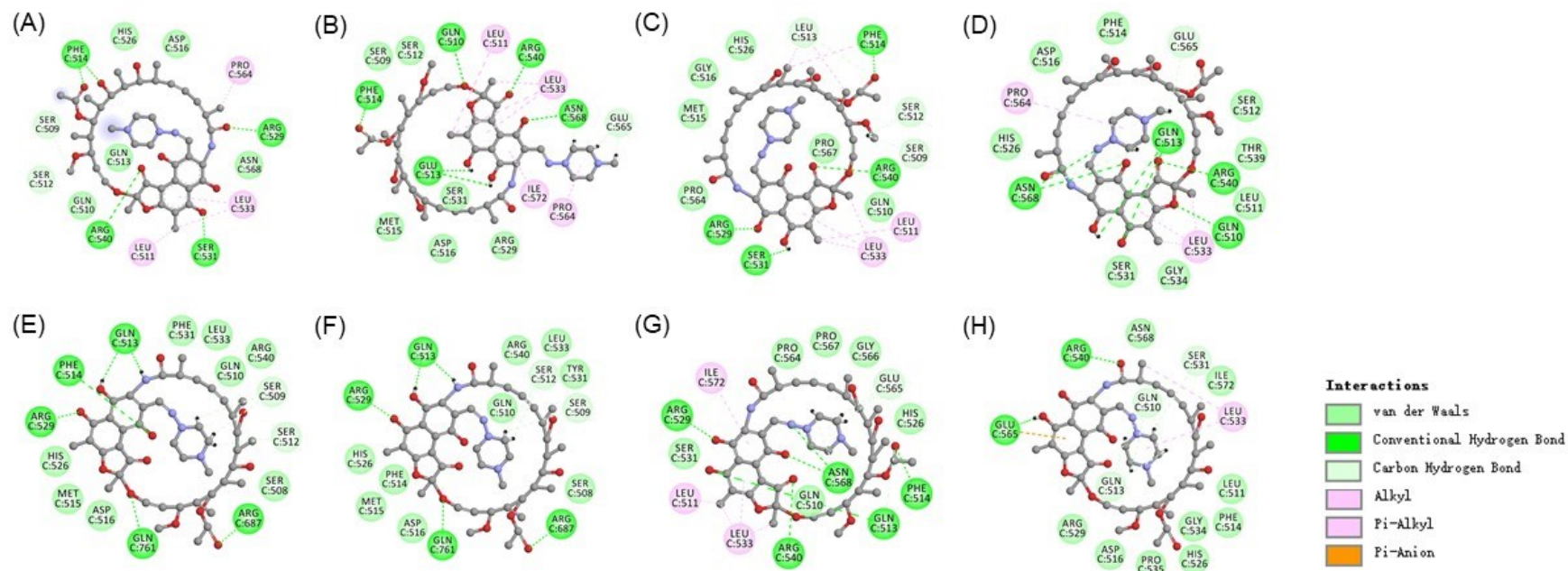


Fig. S5 Diagrams of docking interactions of rifampicin with RpoB protein in wild type and predicted mutants. (A) Wild type, (B) Q513E, (C) Q513L & D516G, (D) R529G, (E) S531F, (F) S531Y, (G) T563P, (H) P564L

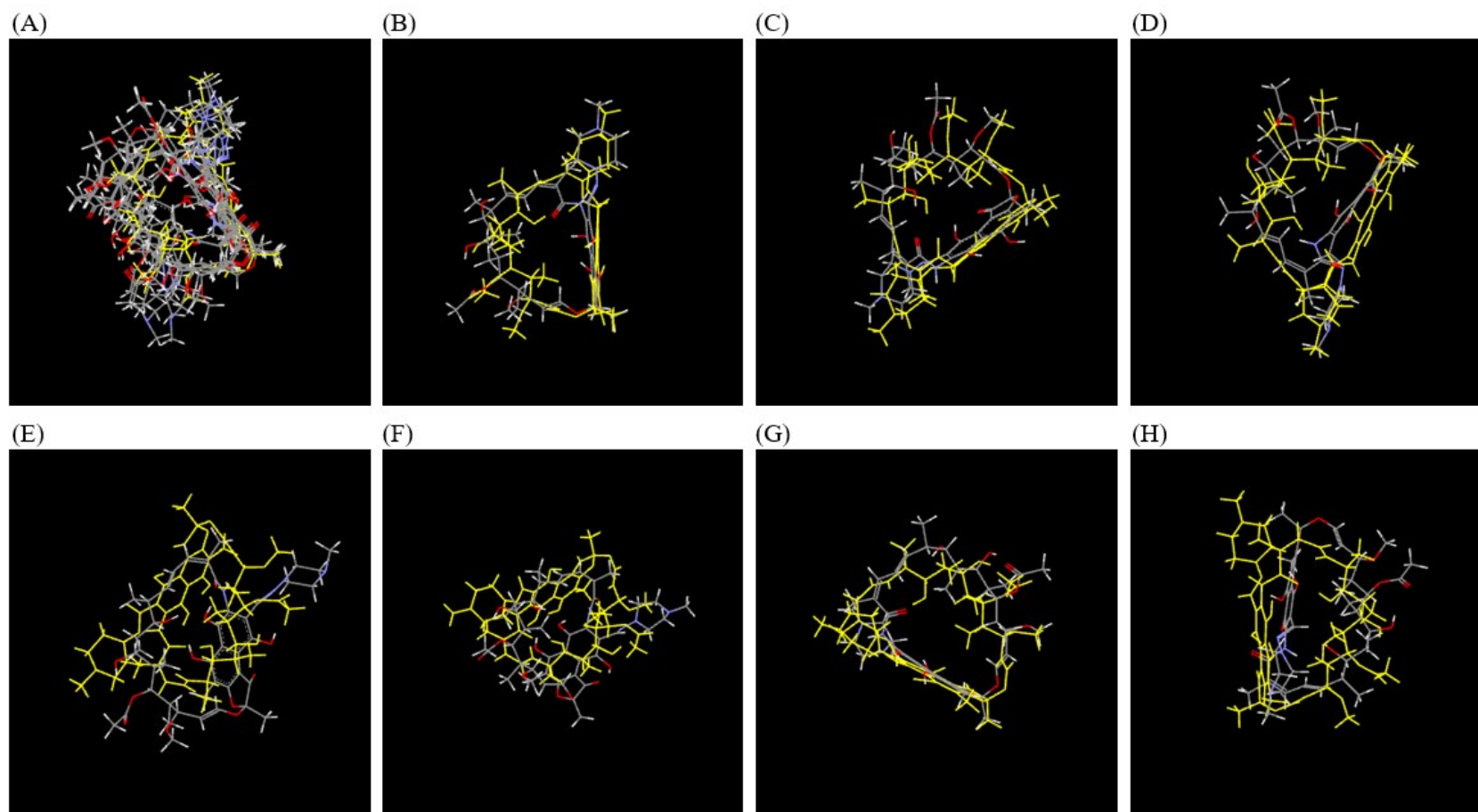


Fig. S6 The differences of docking poses of ligand between wild type and mutant RpoB, the ligand in yellow in all graphs represent the ligand pose after docking with wild type. (A) All poses docking with mutants and wild type, (B) Q513E, (C) Q513L & D516G, (D) R529G, (E) S531F, (F) S531Y, (G) T563P, (H) P564L.

Table S1. Information of the test chemicals, including their CAS number, structures and abbreviations in this work.

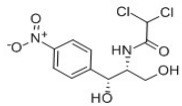
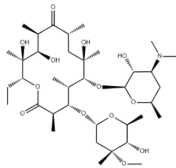
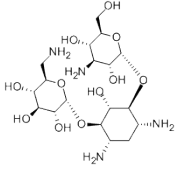
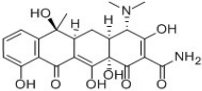
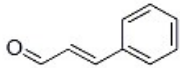
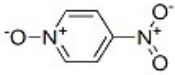
Class	CAS/Name	Abbreviation	Structure
Antibiotics	56-75-7 Chloramphenicol	CHL	
	114-07-8 Erythromycin	ERY	
	59-01-8 Kanamycin	KAN	
	60-54-8 Tetracycline	TET	
QSIs	104-55-2 Cinnamaldehyde	CIN	
	1124-33-0 4-Nitropyridine-N-oxide	NPO	



Table S2. Experimental setup for bacterial resistance mutation screening. Conc. is the abbreviation for concentration. The unit for conc. is mg/L. CHL: chloramphenicol, ERY: erythromycin, KAN: kanamycin, TET: tetracycline; CIN: cinnamaldehyde, NPO: 4-Nitropyridine-N-oxide.

Single exposure			Mixture exposure (QSIs in the mixtures were added at a fixed concentration).	
	Group	Antibiotic conc.	CIN conc.	NPO conc.
CHL	CHL1	0.006	42	1.0
	CHL2	0.06	42	1.0
	CHL3	0.3	42	1.0
	CHL4	0.6	42	1.0
ERY	ERY1	0.16	42	1.0
	ERY2	1.6	42	1.0
	ERY3	8	42	1.0
	ERY4	16	42	1.0
KAN	KAN1	0.01	42	1.0
	KAN2	0.1	42	1.0
	KAN3	0.5	42	1.0
	KAN4	1	42	1.0
TET	TET1	0.001	42	1.0
	TET2	0.01	42	1.0
	TET3	0.05	42	1.0
	TET4	0.1	42	1.0
CIN	CIN1	0.42		
	CIN2	4.2		
	CIN3	21		
	CIN4	42		
NPO	NPO1	0.01		
	NPO2	0.1		
	NPO3	0.5		
	NPO4	1.0		

Table S3. Detailed information of primers of tested genes in *E. coli*.

Genes	Primer sequence (5'-3')		Length (bp)
16S	Forward	GGACGCACGATAGAGAGTGG	141
	Reverse	TTCGGTCCGCGATCCTAATG	
<i>uvrD</i>	Forward	CTACGGTAATCCGGTGGAGC	143
	Reverse	GTTGCAGGATATGCGGCTTG	
<i>rpoS</i>	Forward	TGTCCAGCAACGCTTTTTTCG	189
	Reverse	AGCTGAACGTTTACCTGCGA	
<i>recA</i>	Forward	CTGCTACGCCTTCGCTATCA	231
	Reverse	TCTACGGCGAACTGGTTGAC	
<i>mutS</i>	Forward	ACGGGCCTATAACCCTGAACT	124
	Reverse	CAGCGGGTTGGCGATAAATG	
<i>lexA</i>	Forward	TGACTTGCTGGCAGTGCATA	175
	Reverse	GAAGCTCTGCTGACGAAGGT	
<i>dinB</i>	Forward	CGTTGTCACTGGATGAGGCT	278
	Reverse	TTTTGCCGCTGAGACTTTGC	

Table S4. The best models of predicted mutants using three different evaluation methods.

Mutants	3D-profile Score	DOPE Score	PROCHECK: Most Favored and Allowed Residues Percentage
S531F	559.63	-141,849	99.6%
P564L	558.21	-141,613	99.6%
T563P	559.58	-141,563	99.6%
Q513E	558.56	-141,563	99.6%
S531Y	559.28	-141,809	99.6%
Q513L & D516G	558.27	-141,718	99.7%
R529G	557.94	-141,445	99.6%

Table S5. Protein stability prediction of mutants by calculating unfolding Gibbs free energy changes ( $\Delta\Delta G$ , kcal/mol) using PremPS. The positive sign of  $\Delta\Delta G$  value corresponds to destabilizing mutations.

Mutant	$\Delta\Delta G$ (kcal/mol)	Prediction
Q513E	0.86	Destability
Q513L & D516G	0.51	Destability
R529G	1.55	Destability
S531F	0.52	Destability
S531Y	0.53	Destability
T563P	0.74	Destability
P564L	0.66	Destability