

Supplementary Information

Human CYP2A6 catalyzes the oxidation of 6:2 fluorotelomer alcohol

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Spiked concentration (ng/mL)	Recovered concentration (ng/mL)	Recovery (%)
1000	1020 ± 20	102 ± 2.0
4000	3800 ± 41	95 ± 1.0
8000	7120 ± 31	89 ± 0.4
15000	9900 ± 100	66 ± 0.7
50000	27500 ± 81	55 ± 0.2

Table S1. Spike and recovery data for 6:2 FTOH. Recovery values were obtained in triplicate after 2 hour incubations in 100 mM phosphate buffer and the NADPH recombinant system.

Treatment	Rate relative to control (%)
Uninhibited control	100.0 ± 5.3
Chloramphenicol (CYP2C19), 30 µM	87.5 ± 8.3
Diallyl disulfide (CYP2E1), 68 µM	97.2 ± 12.7
Sulfaphenazole (CYP2C9), 30 µM	95.2 ± 7.0
Quinidine (CYP2D6), 30 µM	84.1 ± 10.2
Ketoconazole (CYP3A4), 18 µM	58.4 ± 7.3
Tranlycypromine, HCl (CYP2A6), 50 µM	40.7 ± 5.3
1-Aminobenzotriazole (General CYP inhibitor), 75 µM	62.9 ± 1.0

Table S2. Relative metabolic rates of 6:2 FTOH conversion in the presence of competitive CYP inhibitors. The starting concentration of 6:2 FTOH is 10000 ng/mL. Standard deviations are derived from n=3 replicates.

Treatment	Rate relative to control (%)
Uninhibited control	100.0 ± 2.4
Chloramphenicol (CYP2C19), 30 µM	120.7 ± 0.9
Diallyl disulfide (CYP2E1), 68 µM	100.2 ± 0.3
Sulfaphenazole (CYP2C9), 30 µM	101.7 ± 1.8
Quinidine (CYP2D6), 30 µM	99.0 ± 4.0
Ketoconazole (CYP3A4), 18 µM	60.4 ± 7.3
Tranlycypromine, HCl (CYP2A6), 50 µM	37.6 ± 4.1

1-Aminobenzotriazole (General CYP inhibitor), 75 μ M

66.1 \pm 11.2

Table S3. Relative metabolic rates of 6:2 FTOH conversion in the presence of competitive inhibitors. The starting concentration of 6:2 FTOH is 1000 ng/mL. Standard deviations are derived from n=3 replicates.

Treatment	6:2 FTOH levels over time (ng/mL)			
	17 hr	20 hr	25 hr	31 hr
Active CYP2A6	4811 \pm 11	4709 \pm 20	4520 \pm 10	4464 \pm 15
Tranlycypromine, HCl, 17.5 μ M	4523 \pm 5	4510 \pm 25	4502 \pm 13	4497 \pm 13
Tranlycypromine, HCl, 35 μ M	4493 \pm 17	4492 \pm 13	4493 \pm 15	4488 \pm 20
Blank	4497 \pm 20	4500 \pm 19	4499 \pm 20	4493 \pm 17

Table S4. 6:2 FTOH conversion by human recombinant CYP2A6 with and without the inhibitor tranlycypromine. The initial concentration of 6:2 FTOH is 10000 ng/mL. The concentration of CYP2A6 is 1.5 μ g/mL. Standard deviations are acquired from n = 3 replicates.

Treatment	6:2 FTOH loss rate
Active CYP2A6	282.2 \pm 56.0
Tranlycypromine, HCl, 17.5 μ M	21.2 \pm 5.2
Tranlycypromine, HCl, 35 μ M	3.9 \pm 1.9
Blank	2.0 \pm 6.5

Table S5. Metabolic rates of inhibited and uninhibited 6:2 FTOH conversion in the presence of human recombinant CYP2A6. The initial concentration of 6:2 FTOH is 10000 ng/mL. The concentration of CYP2A6 is 1.5 μ g/mL. Standard deviations are acquired from n = 3 replicates.

Treatment	6:2 FTOH levels over time (ng/mL)			
	17 hr	20 hr	25 hr	31 hr
Active CYP2A6	948 \pm 3	919 \pm 11	875 \pm 5	822 \pm 3
Tranlycypromine, HCl, 35 μ M	1090 \pm 3	1090 \pm 6	1085 \pm 5	1083 \pm 3
Blank	1111 \pm 15	1109 \pm 20	1110 \pm 10	1107 \pm 13

Table S6. 6:2 FTOH conversion by human recombinant CYP2A6 with and without the inhibitor tranylcypromine. The initial concentration of 6:2 FTOH is 1000 ng/mL. The concentration of CYP2A6 is 1.5 μ g/mL. Standard deviations are acquired from n = 3 replicates.

Treatment	6:2 FTOH loss rate
Active CYP2A6	100.8 \pm 3.3
Tranylcypromine, HCl, 35 μ M	6.0 \pm 5.3
Blank	2.0 \pm 6.0

Table S7. Metabolic rates of inhibited and uninhibited 6:2 FTOH conversion in the presence of human recombinant CYP2A6. The initial concentration of 6:2 FTOH is 1000 ng/mL. The concentration of CYP2A6 is 1.5 μ g/mL. Standard deviations are acquired from n = 3 replicates.

Treatment	Concentration (ng/mL)	6:2 FTOH loss rate (ng/mL min)	Specific rate (ng/min mg)
Uninhibited control	1000	12.1 \pm 1.3	60.5
	2000	24.0 \pm 0.2	119.9
	4000	37.8 \pm 1.6	188.8
	7000	35.9 \pm 1.5	179.6
	8000	43.7 \pm 1.5	218.6
	9000	43.8 \pm 3.3	218.9
	15000	52.3 \pm 3.9	261.5
	20000	60.5 \pm 1.9	302.4
	50000	64.0 \pm 2.3	320.0
	Tranylcypromine, HCl	1000	6.7 \pm 1.5
2000		10.5 \pm 1.0	52.4
4000		21.6 \pm 1.1	108.1
7000		21.1 \pm 0.8	105.5
8000		30.1 \pm 1.5	150.8
9000		35.8 \pm 3.0	178.9
15000		46.5 \pm 2.5	232.7
20000		51.9 \pm 1.9	259.3
50000		56.1 \pm 3.0	280.4

Table S8. Rate and specific rate of 6:2 FTOH transformation in human liver microsomes, in presence and absence of the CYP2A6 selective inhibitor tranylcypramine (50.0 μ M). Standard deviations are acquired from n = 3 replicates.

Treatment	[6:2 FTOH] (ng/ml)	6:2 FTOH levels over time (ng/ml)					Loss Rate (ng/ml/min)
		5 min	30 min	60 min	90 min	120 min	
Active CYP3A4	10000	5780 \pm 129	5164 \pm 188	4571 \pm 149	4755 \pm 93	4113 \pm 79	12.8
Heat inactivated control	10000	6340 \pm 125	5866 \pm 35	5459 \pm 110	4983 \pm 77	4756 \pm 130	13.9
Active CYP3A4	1000	852 \pm 52	747 \pm 9	894 \pm 21	842 \pm 9	832 \pm 19	0.21
Heat inactivated control	1000	846 \pm 93	866 \pm 68	823 \pm 40	835 \pm 41	811 \pm 20	0.35

Table S9. 6:2 FTOH conversion by human recombinant CYP3A4. The initial concentration of 6:2 FTOH is 10000 or 1000 ng/mL. The concentration of CYP3A4 is 100 μ g/mL. Standard deviations are acquired from n = 3 replicates.

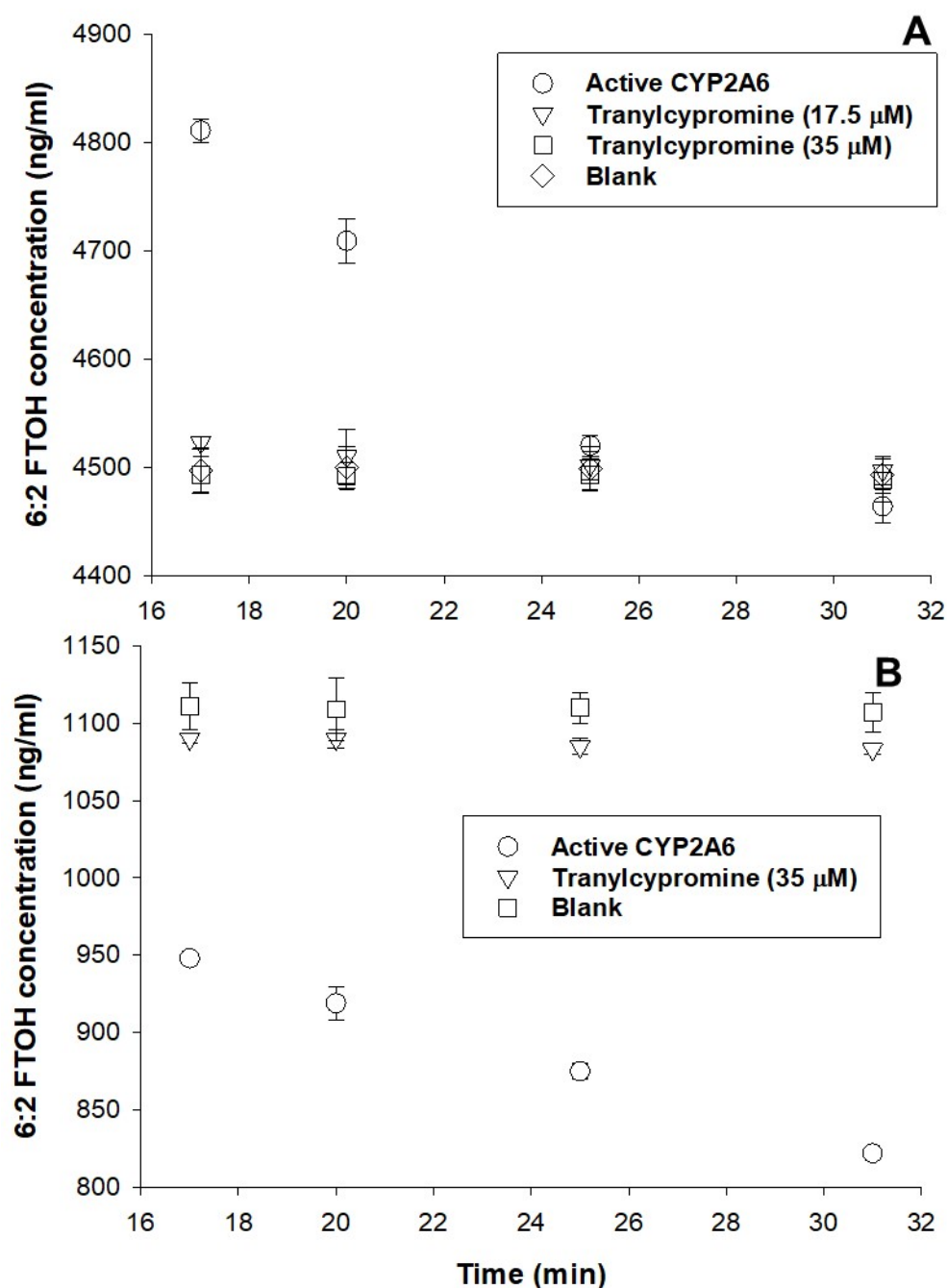


Figure S1. 6:2 FTOH metabolizes by human recombinant CYP2A6. 6:2 FTOH was incubated with human recombinant CYP2A6 (1.5 $\mu\text{g/mL}$) and a NADPH regeneration system in the presence and absence of the CYP2A6 selective inhibitor tranylcypromine. Panel A depicts the high concentration (10000 ng/mL) of 6:2 FTOH. Panel B depicts results with the low concentration (1000 ng/mL) of 6:2 FTOH. Blank assays were run in absence of enzyme. Standard deviations were derived from $n=3$ replicates. Raw data used for this figure are reported in supplemental Tables S4 and S6.

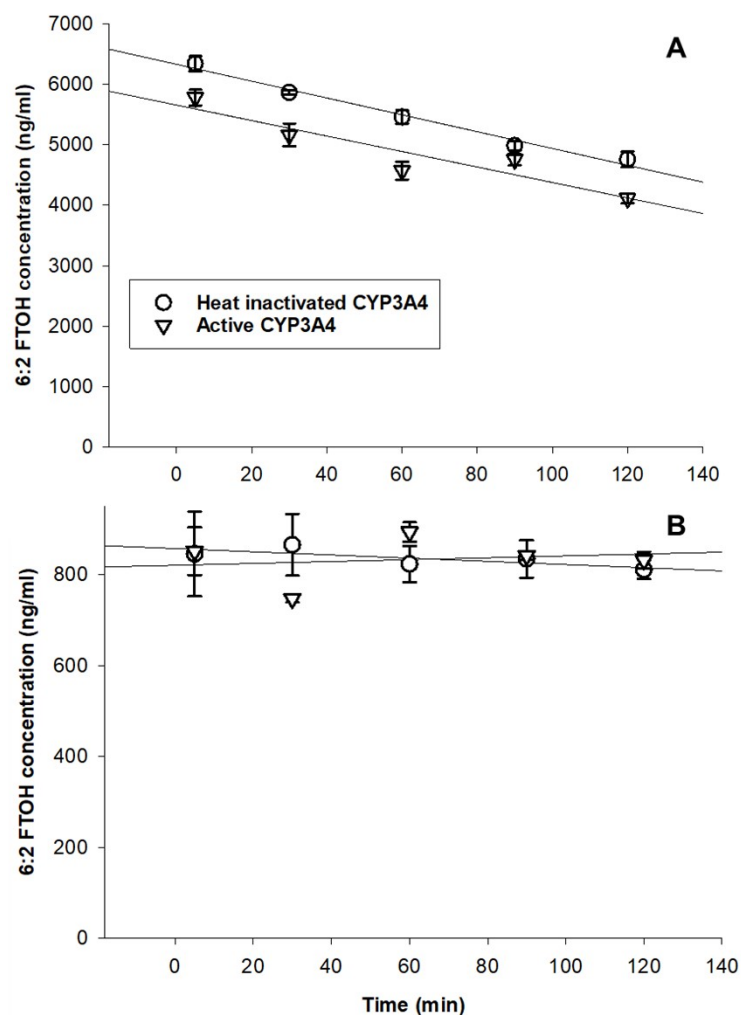


Figure S2. 6:2 FTOH does not metabolize by human recombinant CYP3A4. 6:2 FTOH was incubated with human recombinant CYP3A4 (100 $\mu\text{g}/\text{mL}$) and a NADPH regeneration system. As a negative control, the enzyme was heat inactivated. Panel A depicts the high concentration (10000 ng/mL) of 6:2 FTOH. Panel B depicts results with the low concentration (1000 ng/mL) of 6:2 FTOH. Standard deviations were derived from n=3 replicates. Raw data used for this figure are reported in supplemental Table S9.