## Supporting information for

# Nanoelicitors with prolonged retention and sustained release to produce beneficial compounds in wine

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#### S1. MeJ quantification by UV-Vis spectroscopy

MeJ ketone group strongly absorbs in the UV region near  $\lambda = 291$  nm and its ester group absorbs at  $\lambda = 214$  nm (Figure S1a).<sup>1</sup> We selected the former absorption band for the calibration curves. Taking into account the low solubility of MeJ, we carried out two calibration curves: (1) 100-800 ppm in ultrapure water (Figure S1b, green dots) and (2) 100-2000 ppm ultrapure water:ethanol (50:50, Figure S1b, blue dots). The fitting parameters for each curves are:

$$Abs = 1.73 \cdot 10^{-4} [MeJ] + 0.0078 \quad R^2 = 0.99 \quad (1)$$
$$Abs = 2.07 \cdot 10^{-4} [MeJ] - 0.0028 \quad R^2 = 0.99 \quad (2)$$



**Fig. S1** UV-Vis spectrum (a) and calibration curve (b) of methyl jasmonate in  $H_2O$  (green dots) and  $H_2O/EtOH$  (1:1, blue dots). Dashed lines represent the best fits of the experimental data according to equation 1 for  $H_2O$  and equation 2 for  $H_2O/EtOH$ .



**Fig. S2** FTIR spectrum of MeJ showing the most intense absorption band at 1740 cm<sup>-1</sup> corresponding to carbonyl (ketone) groups.<sup>2</sup>



**Fig. S3** TEM micrograph of nano-MeJ. The nanoparticles shows the same morphology than control ACP nanoparticles.<sup>3</sup> The amorphous nature of the particles is confirmed by the lack of diffraction spots in the selected-area electron diffraction (SAED) pattern (inset).



**Fig. S4** XRD patterns of nano-MeJ sample freeze-dried at time 0, 49 and after 363 days. XRD patterns after 49 and 363 days of storage show two broad Bragg peaks at around 26° and 32° (2θ) ascribed to hydroxyapatite (HA, ASTM card file No 09-432).



Fig. S5 MeJ release profile from nano-MeJ in ultrapure water. Dashed line represents the best fits of the experimental data to the first order equation:  $y(t) = a^*(1-e^{-kt})$ , being the rate constant, k = 0.05 h<sup>-1</sup>. The inset shows the linearized experimental data (symbols) and the first order equation (line).



**Fig. S6 (a)** *t*-piceid and **(b)** *c*-piceid concentration (mg L<sup>-1</sup>) in wines from grapes treated with MeJ (5 mM, MeJ5, and 10 mM, MeJ10) and nano-MeJ with a total concentration of 1 mM. Results of grapes treated with ACP nanoparticles (nano-Control) and non-treated grapes (control) are also shown. Data are expressed as mean with their corresponding standard deviation as error bars. Statistically significant differences between measurements are marked with \* (P-value < 0.05), \*\* (P-value < 0.01) or \*\*\* (P-value < 0.001).



**Fig. S7** Raman spectroscopy of 10 mM MeJ (red spectrum) and NanoMeJ (blue spectrum) after 24 hours at 50°C. Asterisks indicate the presence of MeJ.



**Fig. S8.** Images of vineyards leaves treated with water (Control), MeJ solution (10 and 2 mM) and NanoMeJ (2 mM) at time zero (a) and after 24 hours (b).

	Ca <sup>a</sup> (wt.%)	Pa (wt.%)	K <sup>a</sup> (wt.%)	Ca/P <sup>a</sup>	ζ <sup>b</sup> (mV)	MeJ <sup>c</sup> (wt.%)
Nano-MeJ	$18.02\pm4.3$	$8.79 \pm 1.3$	$0.27\pm0.01$	$1.57\pm0.13$	$-15.7\pm0.6$	$6.15 \pm 1.71$
Nano- Control	$14.7\pm0.14$	$8.06\pm0.1$	$0.46\pm0.01$	$1.41\pm0.03$	$-10.3 \pm 0.7$	-

**Table S1.** Chemical composition and  $\zeta$ -potential of nano-MeJ and ACP nanoparticles (nano-Control). Data are expressed as mean  $\pm$  standard deviation.

<sup>a</sup> Analysed by ICP-OES. <sup>b</sup>Analysed by Litesizer 500. <sup>c</sup>Estimated by UV-Vis spectroscopy

**Table S2.** Enological parameters of the must from grapes under each treatment. Statistically significant differences between measurements are marked with \*\* (P-value < 0.01) whereas ns means no statistical differences.

	Control	Nano-Control	MeJ	Nano-MeJ	р
°Baumé	$12.9\pm0.4$	$13.1\pm0.2$	$12.8\pm0.2$	$12.9\pm0.4$	ns
Total Acidity (g/L)	$2.8\pm0.2b$	$2.5\pm0.1\text{c}$	$3.2\pm0.2a$	$2.9\pm0.1b$	**
pH	$3.9 \pm 0.1$	$3.9\pm0.1$	$3.8\pm0.1$	$3.9\pm0.1$	ns

### References

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