

SUPPORTING INFORMATION

Nano-based soil conditioners eradicate micronutrient deficiency: Soil physico-chemical and plant molecular responses

***Corresponding authors:** Dr. Satya Sundar Bhattacharya, Tel : + 91-3712-275610

E-mail address: satya72@tezu.ernet.in; satyasundarb@yahoo.co.in

Dr. Sanjay Pratihar E-mail address: spratihar29@gmail.com

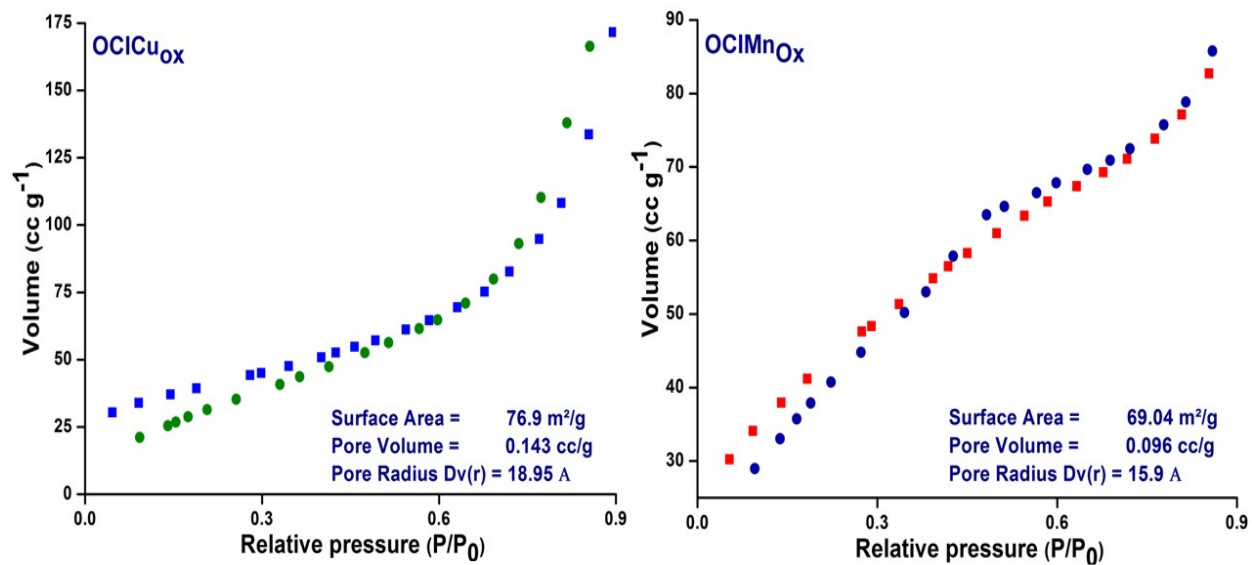


Figure 1S. BET plot for OCICu_{ox} and OCIMn_{ox}

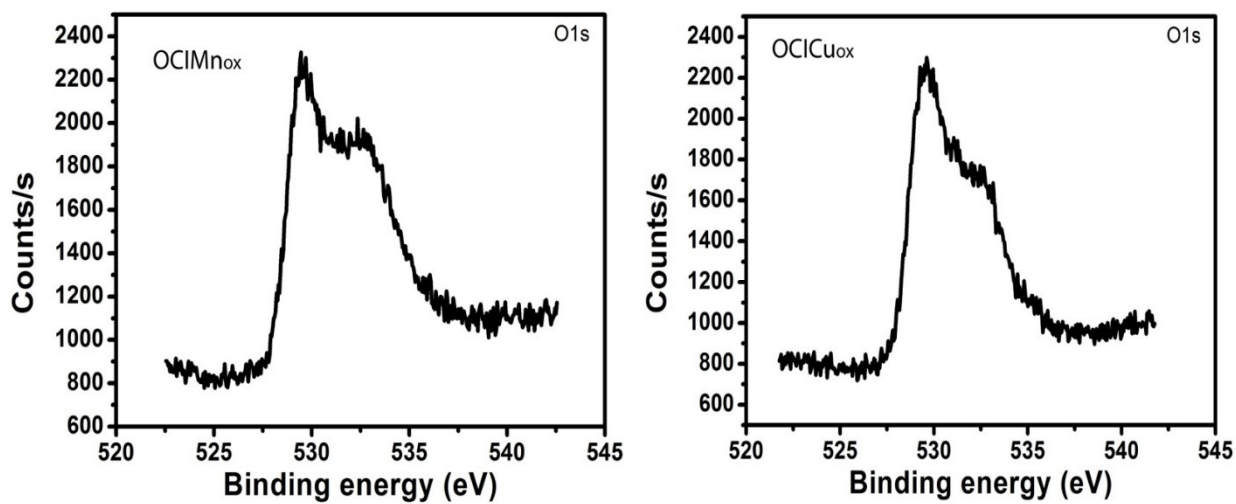


Figure 2S. XPS spectra broad peak of OCICu_{ox} and OCIMn_{ox}

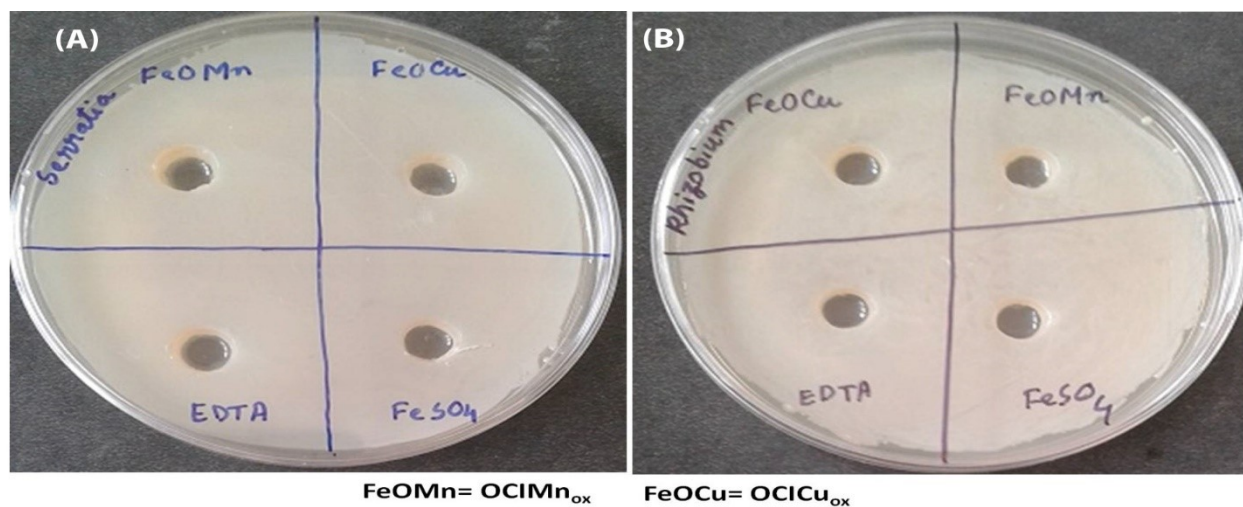


Figure 3S – Evidence of non-inhibitory effects on beneficial soil microbes

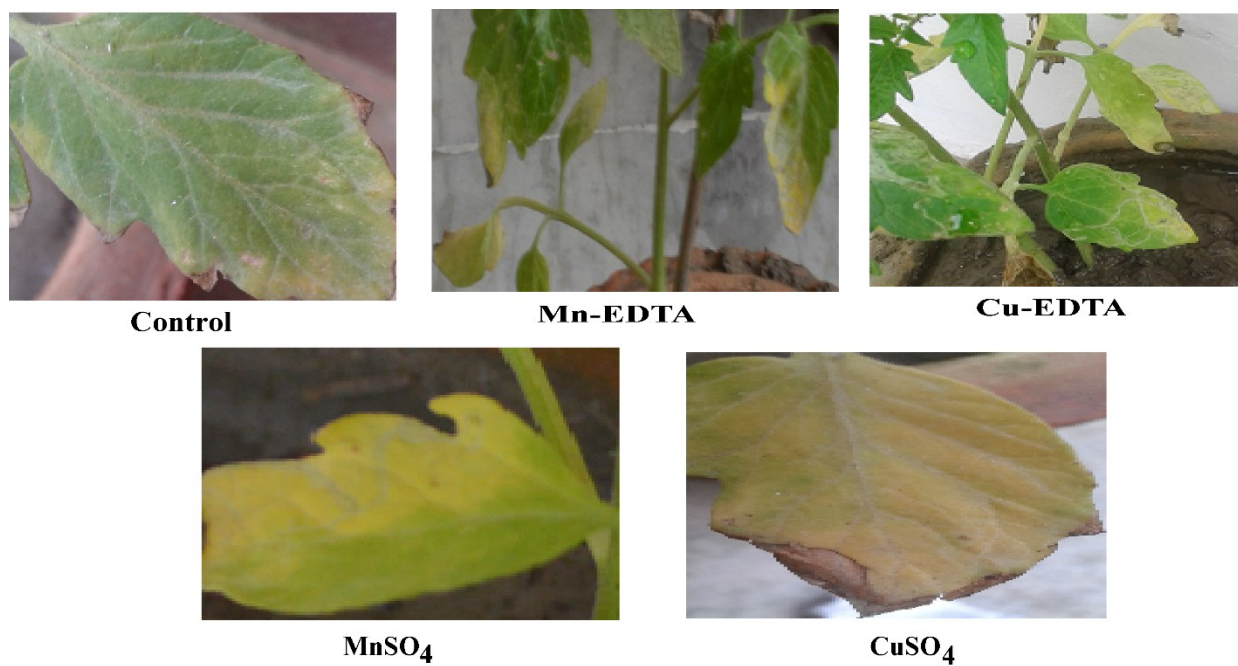


Figure 4S. Chlorotic leaves of control, Mn-EDTA, Cu-EDTA, MnSO₄, and CuSO₄ treatments.

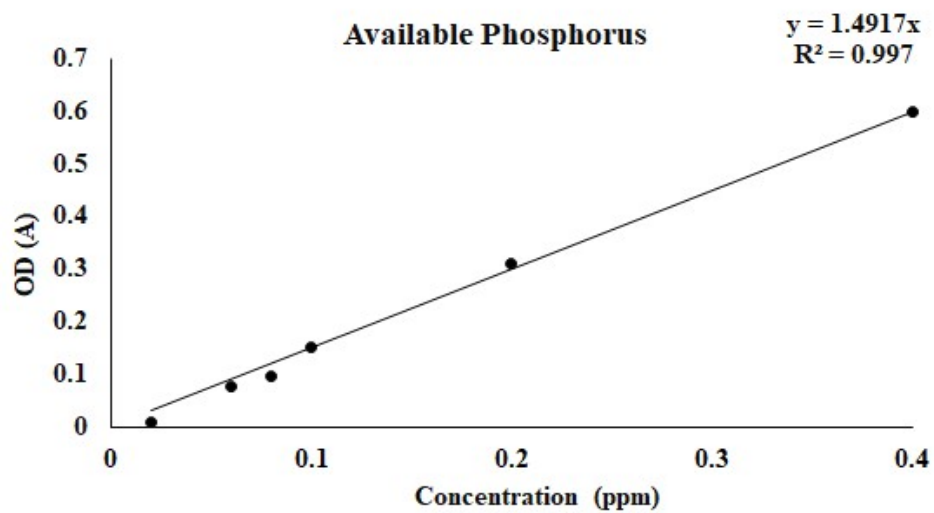


Figure 5S. Method detection limit (MDL) values for available phosphorus in UV-visible spectrophotometer

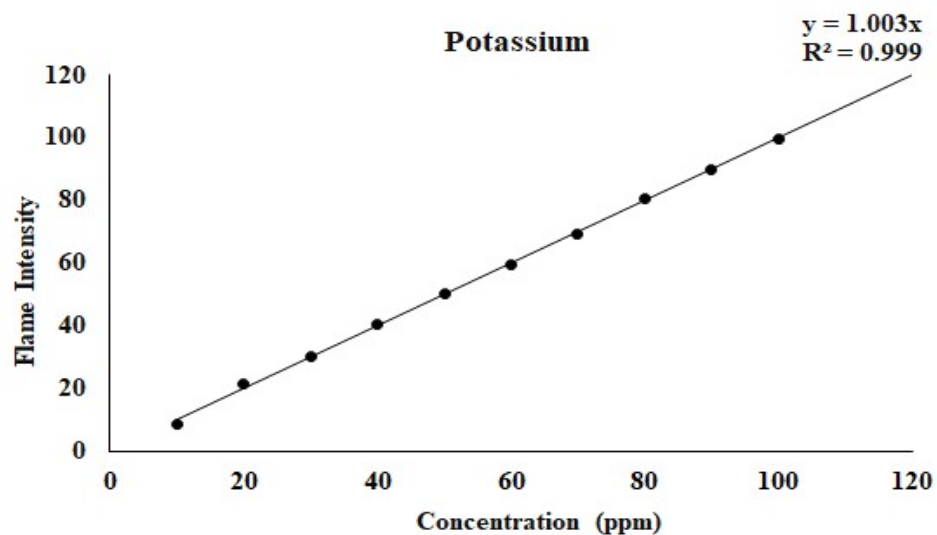


Figure 6S. Relative standard deviation (RSD) for potassium in flame photometer

Table 1S: Method detection limit (MDL) values for different instruments

Instrument	Parameter	Method Detection limit
UV-VIS spectrophotometer	Phosphorus (P)	0.5 ppm
Flame photometer	Potassium (K)	1 ppm
	Sodium (Na)	1 ppm
ICP-OES	Fe	0.01 ppm
	Mn	0.02 ppm
	Cu	0.01 ppm

Table 2S: Relative standard deviation (RSD) values for different attributes measured in various instruments

Instrument	Parameter/Attribute	Relative standard deviation
UV-VIS spectrophotometer	Phosphorus (P)	1.60%
Flame photometer	Potassium (K)	6.20%
	Sodium (Na)	4.70%
ICP-OES	Fe	5.20%
	Mn	3.80%
	Cu	4.60%

Table 3S. Physico-chemical characteristics of the test soil

Soil properties	Mean±stdev
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pH	5.49±0.1
BD (g cc ⁻¹)	1.4±0.5
WHC (%)	60.5±5.6
TOC (%)	0.38±0.01
Available N (mg kg ⁻¹)	270.5±5.1
Available P (mg kg ⁻¹)	21.4±2.1
Available K (mg kg ⁻¹)	89.15±5.6
Fe (mg kg ⁻¹)	110.5±2.4
Mn (mg kg ⁻¹)	47.1±1.1
Cu (mg kg ⁻¹)	17.19±1.1

Table 4S-A: Impact of OCIMn_{ox} and OCICu_{ox} on bulk density (BD) and water holding capacity (WHC) of soil

Treatment	BD (g cc ⁻¹)			WHC (%)		
	0 D	45D	90D	0 D	45D	90D
Control	1.41±0.5	1.38±0.1	1.42±0.2	65.87±1.3	65.87±2.4	68.33±1.2
(OCIMn _{ox}) ₁₀	1.4±0.2	1.36±0.1	1.31±0.2	71.07±1.7	73.2±1.3	78.5±1.5
(OCIMn _{ox}) ₅₀	1.41±0.1	1.37±0.5	1.33±0.1	70.1±2.5	72.1±5.1	75.2±1.8
(OCICu _{ox}) ₁₀	1.39±0.5	1.37±0.4	1.30±0.1	70.6±2.1	72.9±1.2	77.2±2.3
(OCICu _{ox}) ₅₀	1.41±0.01	1.38±0.04	1.35±0.2	68.3±1.7	72.07±1.7	75.2±1.4
Fe-EDTA ₁₀	1.39±0.01	1.38±0.02	1.43±0.01	60.8±0.6	55.6±0.3	52.09±0.5
Fe-EDTA ₅₀	1.41±0.01	1.4±0.01	1.44±0.01	60.5±0.9	56.7±0.4	52.8±0.6
Mn-EDTA ₁₀	1.39±0.01	1.38±0.02	1.43±0.01	60.9±0.6	56.06±0.4	52.1±0.7
Mn-EDTA ₅₀	1.41±0.01	1.4±0.01	1.44±0.01	60.5±0.1	56.7±0.1	50.2±0.2
Cu-EDTA ₁₀	1.4±0.1	1.38±0.1	1.44±0.3	62.56±1.6	53.15±2.1	51.5±1.6
Cu-EDTA ₅₀	1.4±0.04	1.41±0.03	1.44±0.01	63.4±1	61±1.9	50.4±1.6
FeSO ₄	1.41±0.01	1.37±0.02	1.46±0.02	61±1	55.5±1.5	54.3±1.5
MnSO ₄	1.39±0.02	1.39±0.03	1.43±0.01	62±1	56±1.6	53.4±1.4
CuSO ₄	1.38±0.2	1.4±0.2	1.45±0.4	60.4±0.2	55±0.2	51.4±0.2
P value	P(trt×day)<0.01			P(trt×day)<0.01		
LSD	LSD(day)=0.004			LSD (day)=0.37		
	LSD(trt)=0.01			LSD (trt)=0.79		

Table 4S-B: Impact of OCIMn_{ox} and OCICu_{ox} on pH, TOC , available N, and available P content in soil

	pH			TOC (%)			Available N (mg kg ⁻¹)			Available P (mg kg ⁻¹)		
	0 D	45D	90D	0 D	45D	90D	0 D	45D	90D	0 D	45D	90D
Control	5.49±0.1	5.5±0.1	5.51±0.2	0.44±0.06	0.62±0.05	0.69±0.02	279.07±4.2	286.5±1.8	294.7±2.4	24.77±0.8	30.95±1.1	26.5±1.1
(OCIMn _{ox}) ₁₀	5.5±0.1	5.58±0.1	5.64±0.1	0.5±0.04	0.83±0.05	1.1±0.06	285.6±9.6	378.6±7.8	385.3±5.6	26.2±1.1	48.1±1.2	78.8±1.4
(OCIMn _{ox}) ₅₀	5.53±0.1	5.59±0.1	5.63±0.1	0.53±0.02	0.81±0.03	1.03±0.05	281.9±8.3	373.6±9.4	378.8±10.4	25.1±1.1	43.9±1.3	64.9±1.4
(OCICu _{ox}) ₁₀	5.49±0.1	5.57±0.1	5.63±0.2	0.5±0.3	0.81±0.3	1.08±0.7	284.7±4.5	360.8±9.8	383.4±8.6	25.9±1.4	42.7±1.3	74.7±1.1
(OCICu _{ox}) ₅₀	5.52±0.5	5.59±0.5	5.62±0.4	0.49±0.02	0.78±0.01	1.03±0.03	281.9±4.7	356.5±4.7	377.6±4.9	24.98±1.8	39.7±1.9	59.2±1.4
Fe-EDTA ₁₀	5.43±0.2	5.4±0.5	5.46±0.4	0.38±0.12	0.61±0.12	0.71±0.1	278.6±4.7	276.1±5.4	285.9±6.1	21.19±0.5	22.9±0.4	25.6±0.5
Fe-EDTA ₅₀	5.46±0.2	5.43±0.3	5.48±0.4	0.43±0.14	0.59±0.1	0.65±0.05	282.6±11.7	284.7±6.7	279.8±5.7	21.7±1.1	20.64±0.8	24.85±0.9
Mn-EDTA ₁₀	5.40±0.2	5.42±0.3	5.43±0.1	0.41±0.14	0.6±0.15	0.60±0.08	273.7±8.5	278.3±11.4	279.5±8.1	20.1±0.8	21.54±0.9	23.64±0.8
Mn-EDTA ₅₀	5.45±0.3	5.47±0.1	5.5±0.2	0.4±0.15	0.56±0.1	0.61±0.06	275.7±8.1	271.2±4.6	266.1±11.1	21.71±1.1	19.56±0.8	23.9±0.9
Cu-EDTA ₁₀	5.41±0.4	5.45±0.4	5.43±0.2	0.41±0.16	0.60±0.12	0.58±0.08	278.6±4.7	285.8±7.4	286.8±10.5	23.86±0.8	21.58±0.8	18.97±0.9
Cu-EDTA ₅₀	5.41±0.1	5.43±0.2	5.45±0.5	0.40±0.07	0.6±0.1	0.59±0.06	270.9±4.5	275.2±9.8	272.8±10.4	21.6±0.8	22.5±0.9	24.4±0.7
FeSO ₄	4.61±0.1	4.38±0.1	4.33±0.1	0.45±0.06	0.63±0.1	0.61±0.06	271.2±4.8	275.2±5.7	278.9±5.6	21.9±1.1	21.11±0.8	22.28±.8
MnSO ₄	5.40±0.2	5.41±0.1	5.42±0.2	0.44±0.05	0.64±0.12	0.6±0.05	272.7±4.8	278±4.6	265.3±4.8	22.7±0.8	22.1±0.9	20.7±1.2
CuSO ₄	5.40±0.2	5.42±0.3	5.41±0.3	0.43±0.02	0.61±0.01	0.58±0.01	273.4±5.4	278.7±2.3	264.5±3.4	21.8±1.1	20.75±1.1	21.48±1.2
p value	P(trt×day)<0.01			P(trt×day)<0.01			P(trt×day)<0.01			P(trt×day)<0.01		
LSD	LSD(day)=0.022			LSD (day)=0.01			LSD (day)=1.92			LSD (day)=0.08		
	LSD(trt)=0.04			LSD (trt)=0.02			LSD (trt)=4.08			LSD (trt)=0.16		

Table 4S-C: Impact of OCIMn_{ox} and OCICu_{ox} on available K, DTPA extractable Fe, Mn, and Cu in soil

	Available K (mg kg ⁻¹)			Fe (mg kg ⁻¹)			Mn (mg kg ⁻¹)			Cu (mg kg ⁻¹)		
	0 D	45 D	90 D	0 D	45D	90D	0 D	45D	90D	0 D	45D	90D
Control	102.3±3.2	225±4.1	261.8±5	122.5±2	125.5±5.4	127.7±5.4	48.7±3.4	35.5±1.1	29.5±1.2	19.15±0.5	28.7±0.8	22.5±0.5
(OCIMn _{ox}) ₁₀	106.1±4	300.7±3.2	411±4	128.6±4.6	144.4±6.7	158.4±5.7	81.4±3.42	146.5±5.4	145±3.4	14.5±0.9	11.5±1	17.8±1.1
(OCIMn _{ox}) ₅₀	107.2±5.6	298.8±4	394.6±6	124.6±3.7	147.1±3.4	161.4±3.5	83.6±1.1	172.5±1.4	168.4±2.1	16.5±0.6	12.5±0.8	20.6±0.8
(OCICu _{ox}) ₁₀	108.6±4.3	300.4±3	408.4±5	126.2±2.8	141.2±2.5	156.22±2.6	17.4±0.2	25.8±0.8	22.4±0.9	58.15±1.1	107±2.4	114.1±3.4
(OCICu _{ox}) ₅₀	107.7±3	288±5	384±6	125.4±3.4	145.6±3.7	159.6±3.4	19.4±1.4	26.5±1.6	23.4±1.2	63.56±2.4	124.25±3.8	123.8±3.8
Fe-EDTA ₁₀	105.1±4	183.9±3.6	208.4±5	118.1±2.8	89.5±2.4	72.8±2.7	24.5±1.1	25.4±1.2	20.4±1.5	15.8±0.4	11.5±0.6	21.5±0.9
Fe-EDTA ₅₀	100.8±6	178.9±3.7	209.5±6	121.5±2.8	94.7±2.7	75.9±2.6	28.4±0.5	20.8±2.4	16.4±3.5	17.2±0.2	12.4±0.9	24.5±0.9
Mn-EDTA ₁₀	100.1±4	152.2±4	211.8±7	81.7±3.4	71.5±3.4	48.5±4.1	55.6 ±0.4	68.7±1.6	89.4±1.8	15.4±0.8	14.5±0.9	11.5±1.1
Mn-EDTA ₅₀	101.8±3	161±5	200.1±5	83.8±3.6	69.5±3.5	54.4±3.2	64.9±0.4	77.4±1.4	98±2.1	16.8±0.4	13.5±0.5	9.7±0.8
Cu-EDTA ₁₀	102.8±4	160.7±5	221.6±6	69.7±2.8	59.5±3.4	51.5±3.7	22.4±0.8	24.6±2.4	20.9±3.4	45.7±0.7	76.6±0.6	97.6±0.7
Cu-EDTA ₅₀	101.5±4	163.8±6	218.5±6	78.8±4.8	66.1±4.6	56.5±4.3	24.8±0.5	23.5±0.8	21.4±1.1	56.56±1.1	104.25±1.6	108.6±1.4
FeSO ₄	101.8±4	120.7±6	165.6±4	163.7±2.8	189.5±4	171.5±3.7	10.4±0.8	8.6±0.5	8.9±4	12.7±0.6	13.6±0.9	11.6±0.7
MnSO ₄	102±4	211.5±5	205.5±5	83.3±3.4	79.8±3.1	66.4±2.8	28.4±0.8	38.7±2.5	46.86±3.8	11.5±0.8	12.1±0.8	14.5±0.7
CuSO ₄	103.5±3	217.5±4	208.4±5	84.5±0.3	80.5±0.3	67.8±0.2	10.4±0.9	24.6±1.1	12.5±1.1	48.9±1	82.4±2.4	79.8±3.2
p value	P(trt×day)<0.01			P(trt×day)<0.01			P(trt×day)<0.01			P(trt×day)<0.01		
LSD	LSD (day)=0.10			LSD (day)=0.43			LSD (day)=0.32			LSD (day)=0.21		
	LSD (trt)=2.9			LSD (trt)=0.83			LSD (trt)=0.74			LSD (trt)=0.56		

Table 5S: Effect of OCIMn_{ox} and OCICu_{ox} on shift of pH, P, Fe, Mn, and Cu content in KH₂PO₄ solution

Attributes	Treatment	Days				
		24 hrs	48 hrs	7 day	14 day	21 day
pH	OCIMn _{ox}	6.84±0.05	6.83±0.05	6.85±0.04	6.86±0.05	6.87±0.5
	OCICu _{ox}	6.5±0.1	6.48±0.04	6.56±0.04	6.57±0.05	6.6±0.8
	Fe-EDTA	4.43±0.2	4.42±0.2	4.35±0.2	4.32±0.2	4.28±0.2
	Mn-EDTA	4.29±0.01	4.25±0.01	4.11±0.2	4.05±0.2	4.01±0.1
	Cu-EDTA	4.38±0.2	4.33±0.2	4.25±0.1	4.16±0.3	4.09±0.2
	FeSO ₄	2.64±0.15	2.56±0.15	2.41±0.15	2.32±0.12	2.05±0.11
	MnSO ₄	3.56±0.01	3.44±0.02	3.37±0.01	3.28±0.01	3.1±0.01
	CuSO ₄	2.68±0.01	2.51±0.01	2.45±0.02	2.41±0.01	2.35±0.01
Avl P (mg L ⁻¹)	OCIMn _{ox}	84.45±2.8	85.24±1.9	82.78±1.6	82.66±2.4	86.54±2.2
	OCICu _{ox}	68±1.1	65±2.3	72±1.8	75±2.4	78±2.5
	Fe-EDTA	2.51±0.01	2.31±0.05	1.98±0.05	1.86±0.01	1.67±0.02
	Mn-EDTA	5.86±1.2	4.17±1.1	3.54±0.5	2.19±0.25	0.89±0.4
	Cu-EDTA	2.65±2.4	4.82±3.1	3.11±2.8	2.55±2.4	1.11±2.7
	FeSO ₄	28.86±2.1	12.56±1.5	8.56±1.6	9.57±1.8	4.79±1.8
	MnSO ₄	18.7±0.7	11.4±1.2	10.5±0.8	8.9±0.2	6.8±0.1
	CuSO ₄	19.5±1.2	15.6±1.4	12.5±2.1	9.56±1.4	7.6±2.1
Fe (mg L ⁻¹)	OCIMn _{ox}	56.8±1.5	75±1.5	186.7±1.6	211.4±1.6	225±1.6
	OCICu _{ox}	34.2±1.2	46.8±1.2	71±1.6	93.8±1.5	115.6±8
	Fe-EDTA	22.4±1.5	61.85±1.8	47.8±2.1	80.64±2.8	87.21±2.7
	FeSO ₄	264.5±7.5	271±6.9	275±6.3	286±6.5	305±6.8
Mn (mg L ⁻¹)	OCIMn _{ox}	42.86±0.3	62.5±0.1	81.03±0.1	108.79±2.4	145.2±2.5
	Mn-EDTA	37.5±0.02	52.4±0.03	73.42±0.01	98.77±0.2	120.64±0.1
	MnSO ₄	34.18±0.2	47.07±0.9	56±0.16	68±0.07	75±0.5
Cu (mg L ⁻¹)	OCICu _{ox}	105.2±1.5	116.1±1.4	123.7±2.4	126.9±2.4	139.8±2.1
	CuSO ₄	126±1.3	117.8±1.4	133.5±1.7	137.8±2.1	138.5±2.1
	Cu-EDTA	63.13±1.2	77.05±1.14	83.95±1.1	95.13±1.1	85.06±1.1
pH		Avl P	Fe	Mn	Cu	
p(trt)<0.01		p(trt)<0.01	p(trt)<0.01	p(trt)<0.01	p(trt)<0.01	
p(day)<0.01		p(day)<0.01	p(day)<0.01	p(day)<0.01	p(day)<0.01	
p(trt×day)<0.01		p(trt×day)<0.01	p(trt×day)<0.01	p(trt×day)<0.01	p(trt×day)<0.01	
LSD(trt)=0.01		LSD(trt)=0.13	LSD(trt)=0.62	LSD(trt)=0.49	LSD(trt)=0.80	

Table 6S: Variation of Mn and Cu in different pH solutions of OCIMn_{ox} and OCICu_{ox}

Treatment	Mn (mg L ⁻¹)				Cu (mg L ⁻¹)			
	2 hours	24 hours	48 hours	72 hours	2 hours	24 hours	48 hours	72 hours
OCIMn _{ox} pH4	6.1±0.08	7.85±0.5	9.81±0.8	10.38±1.1	BDL	BDL	BDL	BDL
OCIMn _{ox} pH 7	10.12±0.23	14.89±1.2	21.46±1.4	22.45±1.2	BDL	BDL	BDL	BDL
OCIMn _{ox} pH 9	15.56±0.4	18.95±1.1	16.01±2.1	19.35±1.5	BDL	BDL	BDL	BDL
OCICu _{ox} pH4	BDL	BDL	BDL	BDL	2.5±0.01	3.4±0.01	4.2±0.05	5.87±0.5
OCICu _{ox} pH 7	BDL	BDL	BDL	BDL	6.1±0.2	8.74±0.5	9.26±1	9.98±0.8
OCICu _{ox} pH 9	BDL	BDL	BDL	BDL	5.4±0.11	6.67±0.5	7.78±0.5	8.74±0.8
Fe-EDTA pH4	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
Fe-EDTA pH 7	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
Fe-EDTA pH 9	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
Mn-EDTA pH4	2.6±0.2	4.18±0.4	6.2±0.5	8.34±0.05	BDL	BDL	BDL	BDL
Mn-EDTA pH 7	7.1±0.15	8.17±0.1	16.78±1.1	17.74±0.20	BDL	BDL	BDL	BDL
Mn-EDTA pH 9	7.65±0.18	9.64±0.1	12.76±0.8	15.74±0.05	BDL	BDL	BDL	BDL
Cu-EDTA pH 4	BDL	BDL	BDL	BDL	1.9±0.1	2.3±0.4	3.6±0.5	4.51±0.04
Cu-EDTA pH 7	BDL	BDL	BDL	BDL	5.9±0.2	7.89±0.8	8.28±0.8	10.47±0.8
Cu-EDTA pH 9	BDL	BDL	BDL	BDL	4.9±0.21	5.64±0.5	6.28±0.6	7.89±0.8
FeSO ₄ pH4	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
FeSO ₄ pH 7	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
FeSO ₄ pH 9	BDL	BDL	BDL	BDL	BDL	BDL	BDL	BDL
MnSO ₄ pH4	6.8±0.11	8.74±0.05	9.71±0.5	10.34±1.1	BDL	BDL	BDL	BDL
MnSO ₄ pH 7	15.6±0.1	17.5±1.1	20.41±1.1	21.45±1.4	BDL	BDL	BDL	BDL
MnSO ₄ pH 9	18.67±0.3	20.51±1.3	19.9±1.4	20.25±1.6	BDL	BDL	BDL	BDL
CuSO ₄ pH4	BDL	BDL	BDL	BDL	2.6±0.13	3.52±0.4	5.43±0.5	5.89±0.5
CuSO ₄ pH 7	BDL	BDL	BDL	BDL	6.5±0.11	8.89±0.8	9.4±1	10.14±0.8
CuSO ₄ pH 9	BDL	0.01±0.01	0.01±0.01	BDL	5.8±0.11	6.98±1	8.29±0.8	8.89±.7
P value	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01	P(trt)<0.01
LSD	LSD=1.05	LSD=1.5	LSD=1.63	LSD=1.01	LSD=0.46	LSD=0.56	LSD=0.53	LSD=0.49

*BDL: Below detection limit of the instrument

Table 7S: Correlation statistics between pH, Available N, Available P, Available K, Fe, Mn, and Cu of different types of soil; photosynthetic rate, chlorophyll content, and hill activity of tomato leaves under various treatment

	Soil in lab-scale study				
	pH	P	Fe	Mn	Cu
Aqueous media	0.72*	0.83**	0.67*	0.92**	0.95**
	Photosynthetic rate		Hill reaction		Chlorophyll
Photosynthetic rate			0.82**		0.95**
Hill reaction		0.82**			0.75*
Chlorophyll		0.95**	0.75*		

* p<0.05; ** p<0.01

Table 8S: Effect on Catalase activity, Lipid peroxidation, Super oxide dismutase, Photosynthetic rate, Hill activity, Chlorophyll content, total protein content and shelf life in tomato leaves treated with OCIMn_{ox} and OCICu_{ox}

Treatment	Attributes							
	Catalase (ml min ⁻¹ g ⁻¹)	Lipid peroxidation (μM g ⁻¹)	Super oxide dismutase (min ⁻¹ mg ⁻¹)	Photosynthetic rate (μM CO ₂ m ⁻² s ⁻¹)	Hill activity (μM hr ⁻¹)	Chlorophyll (mg g ⁻¹)	Total protein (mg ml ⁻¹)	Shelf life(g)
Control	6.2±0.5	0.073±0.011	34.08±0.12	20.59±1.5	4126±56	38.01±5.4	1.13±0.001	54.8±0.26
OCIMn _{ox}	5.3±1.2	0.013±0.005	11.17±0.31	45.83±4.5	5221±48	54.6±2.5	2.28±0.05	30.1±0.24
OCICu _{ox}	5.3±1.5	0.008±0.001	10.85±0.08	32.76±1.5	5035±78	45.2±2.4	1.99±0.011	28.5±0.64
Fe-EDTA	5.3±1.4	0.108±0.01	31.77±0.29	26.03±2.1	4136±48	40.15±1.8	1.71±0.002	45.5±0.45
Mn-EDTA	5.3±1.4	0.134±0.005	50.15±0.83	22±1.6	3897±38	39.66±1.9	1.25±0.001	47.5±0.53
Cu-EDTA	5.3±1.1	0.167±0.008	35.41±0.11	21±1.8	3884±37	41.6±2.9	1.18±0.001	49.5±0.45
FeSO ₄	8±1.1	0.128±0.005	21.66±0.15	23.79±2.7	4768±65	41.56±3.7	1.18±0.002	38±0.54
MnSO ₄	6.8±0.8	0.172±0.009	27.31±2.5	20±1.4	3564±41	39.56±3.7	1.11±0.002	34.9±0.33
CuSO ₄	6.2±1	0.144±0.012	48.97±0.86	21±1.6	3379±35	39.6±2.7	1.15±0.001	39.1±0.31
p value	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
LSD	0.58	0.007	0.76	0.61	26.8	0.17	0.03	0.75

Table 9S-A: qRT-PCR using *GS2*, *GOGAT*, *NR*, and *Fd* gene-specific primers where *gapdh* served as an internal control

Treatment	Attributes			
	<i>Fd</i>	<i>GOGAT</i>	<i>GS2</i>	<i>NR</i>
Control	1±0.1	1±0.1	1±0.01	1±0.01
OCIMn _{ox}	9.92±0.9	1.35±0.07	2.04±0.35	6.92±0.1
OCICu _{ox}	4.75±0.6	1.82±0.15	1.46±0.09	9.06±0.2
Fe-EDTA	0.05±0.001	0.005±0.001	0.55±0.05	0.38±0.3
FeSO ₄	0.57±0.07	0.26±0.03	0.55±0.05	0.88±0.3
p value	<0.01	<0.01	<0.01	<0.01
LSD	0.05	0.03	0.04	0.03

Table 9S-B: Activity of GS, GOGAT, and NR enzyme in tomato leaves under various treatments

Treatment	Attributes		
	GOGAT (nM min ⁻¹ mg ⁻¹)	GS (μM min ⁻¹ g ⁻¹)	NR (g hr ⁻¹)
Control	16.09±1.2	0.17±0.01	31.78±1.2
OCIMn _{ox}	16.2±0.5	0.195±0.02	68.34±2.4
OCICu _{ox}	18.49±0.5	0.178±0.01	55.9±1.1
Fe-EDTA	11.02±0.5	0.136±0.01	25±1.1
FeSO ₄	14.17±0.4	0.134±0.01	34±1.4
p value	<0.01	<0.01	<0.01
LSD	0.05	0.01	0.6

Table 10S: Effect of OCICuox and OCIMnox on uptake of N, P, Fe, Mn, Cu, and Yield of tomato plant grown in micronutrient leached and natural soil

Treatments	Uptake N (%)	Uptake P (mg kg ⁻¹)	Uptake Fe (mg kg ⁻¹)	Uptake Mn (mg kg ⁻¹)	Uptake Cu (mg kg ⁻¹)	Yield (g plant ⁻¹)
Control	2.44±0.2	28.5±4.5	3.3±0.6	0.48±0.01	0.12±0.01	14.9±3.4
OCIMn _{ox}	6.81±0.1	108.5±11.4	48.3±2.6	4.8±0.05	0.08±0.03	38.9±4.6
OCICu _{ox}	6.39±0.4	106.4±15.4	41.4±5.4	0.11±0.01	2.8±0.01	37.5±2.6
Fe-EDTA	2.45±0.3	41.52±1.7	28.54±1.1	0.15±0.01	0.11±0.01	11.08±1.3
Mn-EDTA	2.65±0.1	34.59±1.1	22.57±1.6	2.8±0.06	0.05±0.01	12.54±1
Cu-EDTA	3.03±0.2	39.09±1.4	25.17±1.2	0.11±0.05	1.5±0.01	10.25±1.1
FeSO ₄	3.11±0.1	34.58±1.8	53.18±3.4	0.21±0.05	0.08±0.01	8.97±1.1
MnSO ₄	3.03±0.4	39.19±2.1	28.6±1.2	1.5±0.09	0.06±0.01	10.41±1.1
CuSO ₄	2.98±0.2	33.56±1.5	32.2±1.7	0.16±0.01	1.2±0.05	11.5±1.1
p value	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
LSD	0.72	0.95	0.21	0.05	0.06	0.2

Table 11S: Effect of OCIMn_{ox} and OCICu_{ox} on relative seed germination (RSG), relative root growth (RRG), and germination index (GI) of tomato (Badshah *F1 hybrid*).

Treatment	Attributes		
	RSG (%)	RRG (%)	GI (%)
Control	91±7.5	68±5	61.88±4.8
(OCIMn _{ox}) ₁₀	166.67±24.5	131±15	218.3±36
(OCIMn _{ox}) ₂₀	135±17	134±27	180.9±30
(OCIMn _{ox}) ₅₀	134±18	101±31	135.34±24
(OCICu _{ox}) ₁₀	159±21	125±12	198.75±14
(OCICu _{ox}) ₂₀	131±18.54	105±14	137.55±15
(OCICu _{ox}) ₅₀	108±14	95±4.6	102.6±13
(FeSO ₄) ₅₀	63±10	45±4.5	28.35±5.2
(Fe-EDTA) ₅₀	45±8.5	36±3.4	16.2±5.3
p value	<0.01	<0.01	<0.01
LSD	0.86	0.82	1.08

Table 12S: Activity of catalase, reduced glutathione (GSH), glutathione S transferase (GST), super oxide dismutase (SOD), earthworm biomass and body length in earthworms exposed to OCIMn_{ox} and OCICu_{ox} (mean±stdev)

Treatment	Catalase (U mg ⁻¹ min ⁻¹)		GSH (nM mg ⁻¹)		GST (min ⁻¹ mg ⁻¹)		SOD (min ⁻¹ mg ⁻¹)		Earthworm biomass (g)		Body length (cm)	
	10 day	30 day	10 day	30 day	10 day	30 day	10 day	30 day	10 day	30 day	10 day	30 day
(OCIMn _{ox}) ₁₀	52.01±2.8	19.29±0.95	114.08±5.9	20.88±1	121.99±6.9	43.01±4.13	43.79±2.2	24.53±1.3	0.4±0.02	0.7±0.04	5.8±0.3	10±0.5
(OCIMn _{ox}) ₅₀	94.01±4.8	24.68±1.4	138.58±6.2	21.89±1.1	190.32±9.8	62.95±3.1	56.66±2.8	30.74±1.5	0.35±0.02	0.58±0.03	5.6±0.26	8.5±0.4
(OCICu _{ox}) ₁₀	36.69±1.9	15.85±0.8	119.01±6.3	20.58±1	185.47±10	52.71±2.4	46.01±2.3	15.02±0.8	0.37±0.02	0.67±0.03	5.6±0.28	7.5±0.4
(OCICu _{ox}) ₅₀	11.09±0.58	20.42±1	122.59±6.3	26.72±1.3	169.57±8.6	82.44±4.1	67.29±3.4	33.15±1.7	0.35±0.03	0.53±0.03	5.5±0.23	7.1±0.4
(FeSO ₄) ₁₀	581.48±28.5	314.08±16.1	206.46±9.8	168.91±8.6	1428.79±68	240.38±12	381.63±20.1	472.84±23.8	0.32±0.02	0.35±0.02	5.2±0.28	5.8±0.3
(FeSO ₄) ₅₀	765.32±37.1	346.97±17.6	370.65±19.2	176.44±8.8	2617.53±110	280.39±14	473.56±23.9	366.38±18.5	0.31±0.03	0.34±0.02	5.2±0.29	5.9±0.3
Control	211.48±11.4	57.32±2.9	83.31±4.3	42.06±2.1	154.04±7.7	72.78±3.5	69.23±3.5	66.91±3.5	0.33±0.03	0.39±0.03	5.1±0.22	6±0.3
P	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
LSD	0.162	8.85	0.469	0.907	15.78	1.99	7.594	9.26	0.0004	0.04	0.092	0.102

SI-A (for section 2.2.2.2.): Microbial analytical procedure

Soil microbial health was assessed by the growth of soil bacteria following serial dilution technique as reported by Parmer and Schimdt³⁸. At first, air dried soil samples (1 g) were thoroughly mixed in 10 ml of sterile deionized water for 15min with the help of a vortex machine and serially diluted from 10^{-1} to 10^{-6} . Then, 0.1 ml of aliquots was inoculated in previously prepared nutrient agar (NA) containing petriplates in a laminar air flow chamber to ensure aseptic condition. Subsequently, the petriplates were incubated in an incubator for 24h at 37 °C to promote bacterial growth. After incubation, the colonies appeared in the petriplates were enumerated by means of a colony counter and approximate number of bacterial load was estimated by the formula given below:

$$\text{No. of bacteria ml}^{-1} = \frac{\text{No. of colonies} \times \text{dilution}}{\text{Amount plated}}$$

Siderophore producing bacterial growth was also assessed. Siderophores are secondary metabolites produced by diverse groups of soil microorganisms; they play a vital in sustaining Fe availability in soils, specifically under nutrient deficient conditions⁴⁰. Hence, 1 g of soil samples under various treatments were aseptically submerged in 10 ml of sterilized distilled water and homogenized in vortex for 10-15 s and supernatants (100 µl) were collected. Eventually, the samples were serially diluted (10^{-3} to 10^{-4} times) and inoculated on CAS agar petri-dishes for isolating siderophore producing bacterial strains from different soil samples. The CAS agar plates were prepared according to the method of Loudon et al.⁴¹. We also added griseofulvin ($200 \mu\text{g ml}^{-1}$) with CAS agar plates to avoid fungal contaminations. After 48h of incubation at 28 °C, single colonies with maximum halo zones were identified, counted, and purified by

repeated streaking for 5 to 6 generations on CAS agar plates. These pure siderophore producing bacterial strains were again spotted on the CAS agar plates and incubated at 28 °C for 72h. Siderophore production potential of the isolates was qualitatively evaluated on the basis of the development of orange zones around the bacterial colonies.

SI-B for 2.2.2.3 (Plant analyses)

Details of qRT-PCR techniques

The qRT-PCR was conducted with gene specific primers of *Fd*, *NR*, *GS2*, and *GOGAT*. Initially, TRI reagent, Isopropanol and DEPC treated water were used for extraction of total RNA. From this total RNA, 2 µg of sample was used in qRT-PCR in a total volume of 20 µL with gene specific primers of the specified genes. The standard conditions maintained in qRT-PCR are as follows: initial activation step (95 °C, 15 min), cycling step (denaturation at 95 °C, 30 s; annealing at 55 °C, 30 s; extension at 72 °C, 30 s, 40 cycles), melt curve analysis (55–60 °C, 15 s, 40 cycles). GAPDH served as a house keeping gene.

The sequences of the primers are given below:

Fd (forward primer): AAGAAAGCCGGCTGTGACTA

Fd (reverse primer): ACTGGCCATGCAAGTAATCC

NR (forward primer): TTACAACTCCGTGCCATGAA

NR (reverse primer): TCCATGTCTCTCCTCCATCC

GS2 (forward primer): GAATCTATCCCTTCGCCACA

GS2 (reverse primer): CCTTGCCTTCCTTCTCAGTG

GOGAT (forward primer): ACGAGGAGCTCGGTAGTGAA

GOGAT (reverse primer): GCACCTGAGACAATGCAGAA

SI-C (for section 2.2.3): Formulas for deriving relative shoot growth (RSG), relative root growth (RRG), and germination index (GI) in treated tomato seeds

$$RSG (\%) = \frac{\text{Number of seeds germinated with treatments}}{\text{Number of seeds germinated in distilled water}} \times 100$$

$$RRG (\%) = \frac{\text{Mean root length of seeds receiving treated solutions}}{\text{Mean root length of seeds receiving distilled water}} \times 100$$

$$GI(\%) = \frac{RSG \times RRG}{100}$$
