

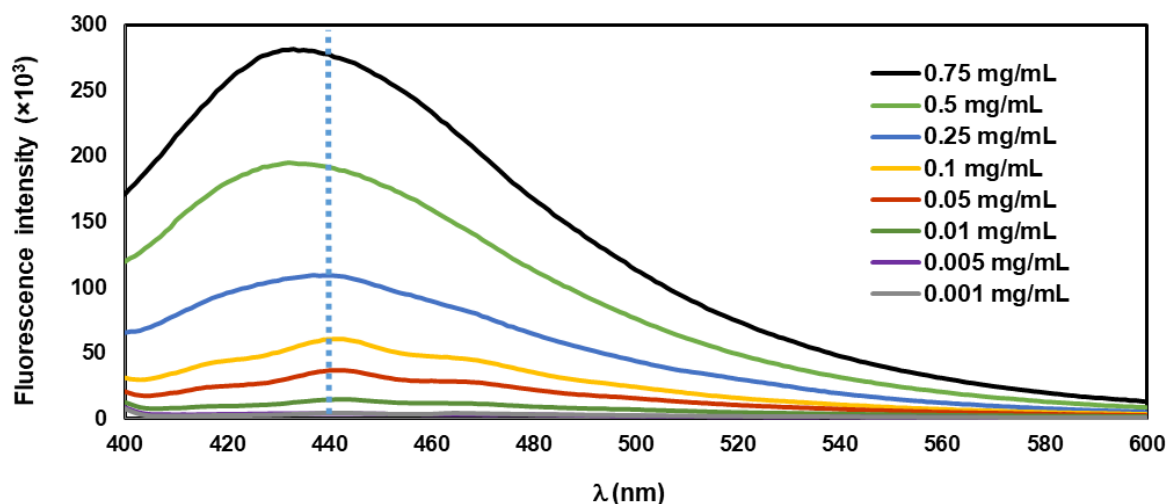
## Supporting Information

# Effect of Polycation Nanostructures on Cell Membrane Permeability and Toxicity

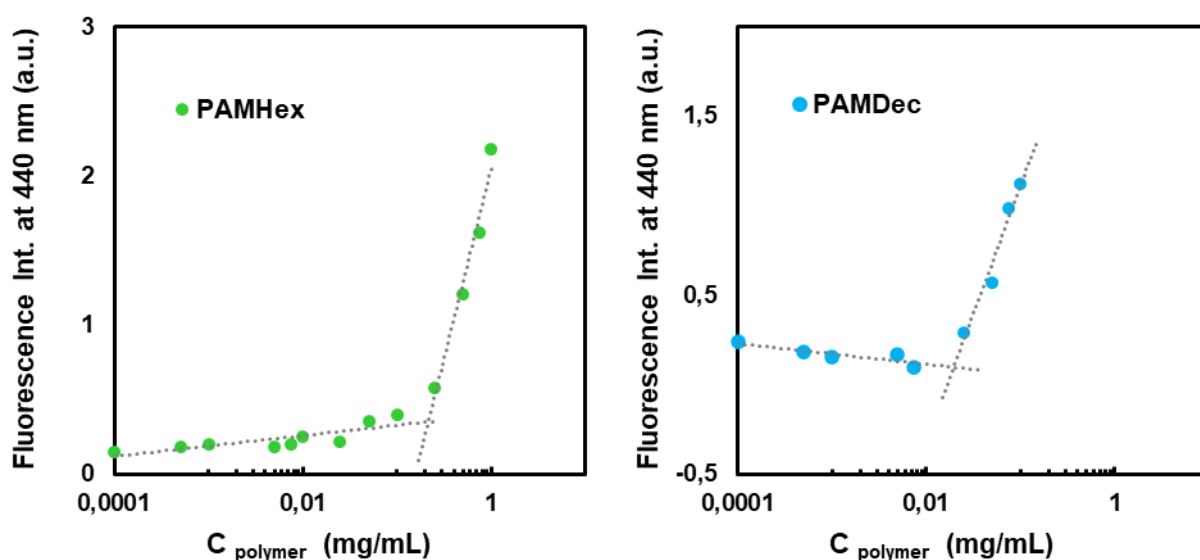
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### 1. Determination of the critical micellar concentration (CMC) for PAMAks.

The CMC value for the polycations was determined using a fluorescent molecular probe technique as described previously.<sup>1</sup> A stock solution of 1,6-diphenyl-1,3,5-hexatriene was prepared in methanol at a concentration of 0.4 mM by short sonication in a bath sonicator. A series of samples containing PAMAks at various concentrations (0 - 0.1 mg/mL) and a constant DPH concentration (4  $\mu$ M) were prepared and stirred for 4 hours in the dark. The fluorescence spectra of these samples were measured using the FS5 Spectrofluorometer (Edinburgh Instrument) (**Fig. S1**). The samples were excited at 350 nm and the fluorescence intensity at 440 nm in the presence of PAMAks was analyzed. Results are presented in **Table 1** and **Fig. 2** and **S2**.



**Fig. S1** Fluorescence spectra of DPH ( $c_{\text{DPH}} = 4 \mu\text{M}$ ,  $\lambda_{\text{ex}} = 350 \text{ nm}$ ) at selected concentrations of PAMBut.



**Fig. S2** The fluorescence intensity of DPH probe ( $c_{\text{DPH}} = 4 \mu\text{M}$ ,  $\lambda_{\text{ex}} = 350 \text{ nm}$ ,) at 440 nm as a function of the concentration of PAMHex and PAMDec.

## 2. Dynamic light scattering (DLS) and $\zeta$ -potential measurements of polymers

The distribution profiles of the hydrodynamic diameters were measured by light scattering experiments (Zetasizer, Malvern) for the aqueous solution of PAMAlks at the concentration of 1 mg/mL. Samples were filtrated by 0.45  $\mu\text{m}$  pores size filters. The mean hydrodynamic

diameter (dz), dispersity (PDI), and  $\zeta$ -potential for the observed objects are presented in Table 2.

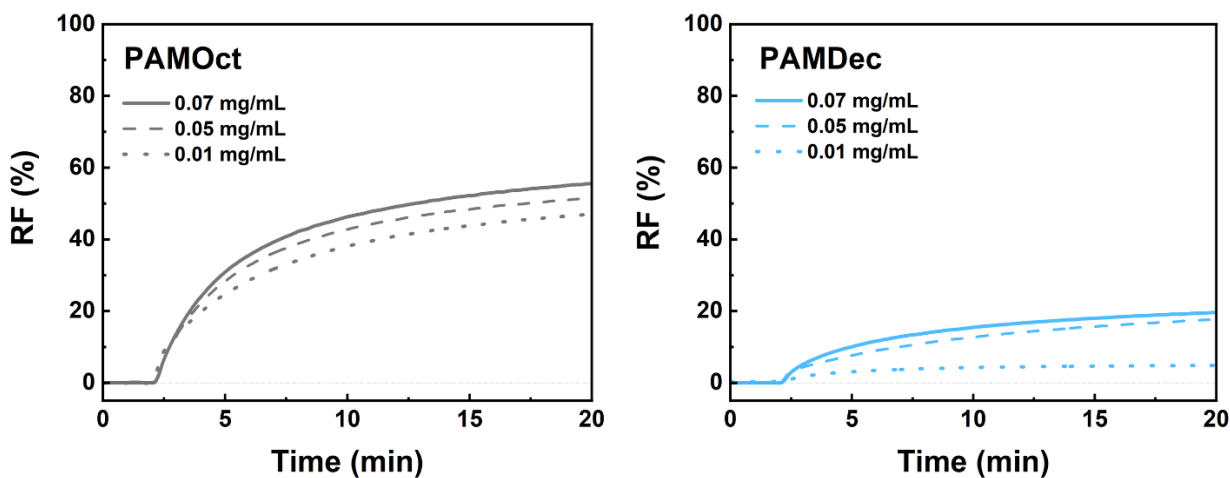
### **3. Dynamic light scattering (DLS) and $\zeta$ -potential - optimization of liposomes coating by polymers**

Polymer-coated anionic liposomes were prepared at various lipid/polymer weight ratios in a 1 mM NaCl solution. Different volumes of the PAM and PAMAks solutions (0.5 mg/mL) were mixed with the liposome suspension (2.5 mg in 0.5 mL) to the final volume of 1 mL. The optimal polymer content was determined from DLS and  $\zeta$ -potential measurements. The crucial criteria were: small size and PDI as well as overloading the  $\zeta$ -potential of the POPC/POPA liposomes to stable positive values (above 30 mV). The optimal polymer content is indicated in bold in **Table S1**.

**Table S1** The values of the mean hydrodynamic diameter (dz), polydispersity index (PDI), and the  $\zeta$ -potential at 298 K of the POPC/DOPA liposomes ( $C_{lipid} = 1.25$  mg/mL) dispersed in the 1 mM NaCl solution at pH 7.4 and treated with polycations (values are the mean  $\pm$  standard deviation). The chosen optimized polycation content was marked in bold.

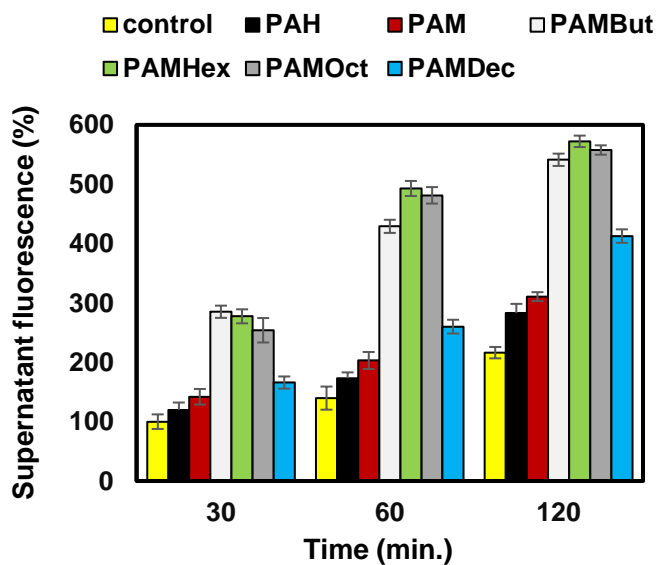
System	Polycation content (wt%)	dz (nm) (n = 5)	PDI (n = 5)	$\zeta$ -potential (mV) (n = 8)
Liposomes (POPC/DOPA)	0	112.0 $\pm$ 0.9	0.08 $\pm$ 0.02	-42.7 $\pm$ 2.8
+ PAM	1.6	> 10 <sup>5</sup>	0.92 $\pm$ 0.18	-17.4 $\pm$ 1.4
	2.4	318.5 $\pm$ 0.3	0.34 $\pm$ 0.09	19.1 $\pm$ 1.5
	3.2	175.6 $\pm$ 0.3	0.21 $\pm$ 0.04	32.9 $\pm$ 2.6
	<b>4.0</b>	<b>153.7 <math>\pm</math> 0.2</b>	<b>0.16 <math>\pm</math> 0.02</b>	<b>34.2 <math>\pm</math> 3.0</b>
	4.8	149.1 $\pm$ 0.2	0.21 $\pm$ 0.02	37.2 $\pm$ 2.9
	5.6	139.7 $\pm$ 0.1	0.13 $\pm$ 0.03	39.1 $\pm$ 1.1
+ PAMBut	3.2	160.2 $\pm$ 1.8	0.21 $\pm$ 0.03	19.9 $\pm$ 1.0
	4.0	153.5 $\pm$ 2.1	0.17 $\pm$ 0.02	22.4 $\pm$ 2.4
	4.8	150.1 $\pm$ 3.2	0.15 $\pm$ 0.05	27.0 $\pm$ 1.5
	<b>5.6</b>	<b>156.0 <math>\pm</math> 3.1</b>	<b>0.13 <math>\pm</math> 0.04</b>	<b>37.9 <math>\pm</math> 1.8</b>
	6.4	147.6 $\pm$ 2.0	0.13 $\pm$ 0.03	36.0 $\pm$ 1.9
	7.2	140.5 $\pm$ 0.1	0.13 $\pm$ 0.04	36.3 $\pm$ 2.2
+ PAMHex	2.4	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	-12.0 $\pm$ 1.0
	3.2	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	7.0 $\pm$ 1.3
	4.0	307.6 $\pm$ 108.8	0.38 $\pm$ 0.06	20.5 $\pm$ 0.7
	4.8	146.5 $\pm$ 1.4	0.12 $\pm$ 0.02	27.0 $\pm$ 1.8
	<b>5.6</b>	<b>142.8 <math>\pm</math> 1.2</b>	<b>0.14 <math>\pm</math> 0.01</b>	<b>32.3 <math>\pm</math> 2.4</b>
	6.4	138.0 $\pm$ 2.6	0.14 $\pm$ 0.02	33.2 $\pm$ 1.4
+ PAMOct	4.0	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	0.0 $\pm$ 0.0
	4.8	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	15.6 $\pm$ 0.9
	5.6	312.1 $\pm$ 10.2	0.40 $\pm$ 0.06	24.6 $\pm$ 1.4
	<b>6.4</b>	<b>209.8 <math>\pm</math> 6.8</b>	<b>0.34 <math>\pm</math> 0.04</b>	<b>33.3 <math>\pm</math> 0.9</b>
	7.2	198.4 $\pm$ 3.8	0.30 $\pm$ 0.02	35.2 $\pm$ 1.2
	8.0	193.0 $\pm$ 2.9	0.29 $\pm$ 0.03	38.4 $\pm$ 2.4
+ PAMDec	5.6	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	-7.2 $\pm$ 5.1
	6.4	> 10 <sup>5</sup>	1.00 $\pm$ 0.00	0.0 $\pm$ 0.0
	7.2	250.1 $\pm$ 10.3	0.28 $\pm$ 0.06	15.7 $\pm$ 2.6
	8.0	163.0 $\pm$ 2.4	0.14 $\pm$ 0.09	26.6 $\pm$ 3.1
	<b>8.8</b>	<b>157.5 <math>\pm</math> 4.6</b>	<b>0.16 <math>\pm</math> 0.08</b>	<b>31.9 <math>\pm</math> 2.5</b>
	9.6	151.4 $\pm$ 1.7	0.12 $\pm$ 0.02	30.2 $\pm$ 1.0

#### 4. Calcein release from liposomes



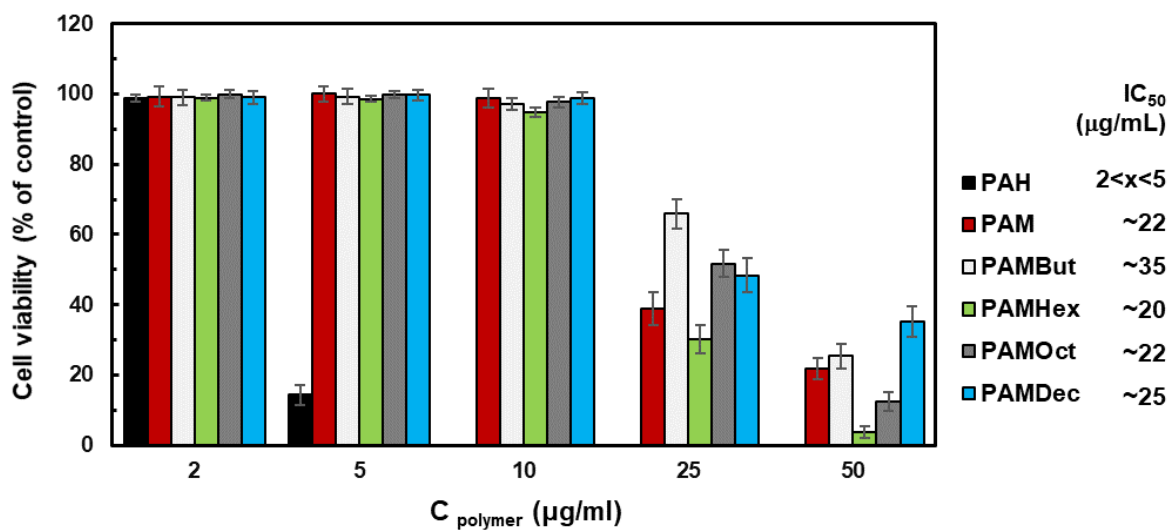
**Fig. S3.** Time-course of calcein leakage from the anionic SUVs treated with the polycations at 2 min. Liposomes were treated with PAMOct and PAMDec at above their CMC: 0.07 and 0.05 mg/mL and below CMC: 0.01 mg/mL. The dye was completely released by adding Triton X-100 and set as 100% leakage.

#### 5. Calcein release from HSFs



**Fig. S4** Fluorescence intensity of calcein in supernatants after 30, 60, and 120 min incubation of HSFs with polymer solutions (PAH, PAM, and PAMAiks - 10  $\mu$ g/mL). As a control, cells untreated with polymers were used. Average values  $\pm$  s.d. of two different experiments, each in triplicate, are shown.

## 6. Polymers cytotoxicity after 48h of HSFs culture.



**Fig. S5** Cytotoxicity of the polyelectrolytes to HSFs was measured by the FDA-EB assay. HSFs were incubated for 48 h in a medium containing the various polymers: PAH, PAM, and PAMAlks. The IC<sub>50</sub> values after 48 h of incubation assessed against HSFs are given on the right.

### References:

1. A. Chattopadhyay and E. London, Fluorimetric Determination of Critical Micelle Concentration Avoiding Interference from Detergent Charge, *Anal. Biochem.*, 1984, **139**, 408-412.