

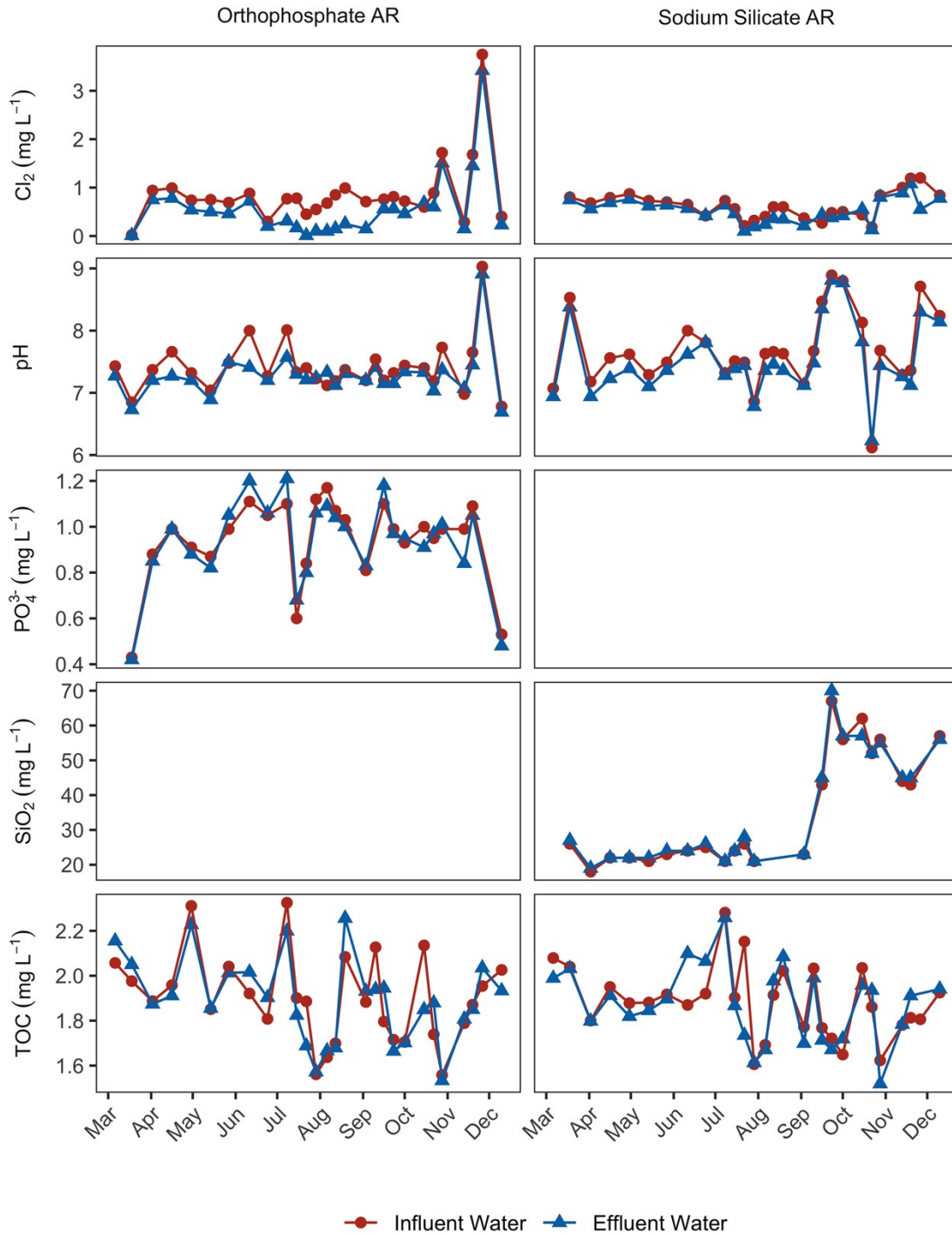
1 **Supplementary information for: *Effect of sodium silicate on***
2 ***drinking water biofilm development***

3 This document contains 6 figures, 2 tables, and 12 pages.

4 **Pilot-scale model distribution system**

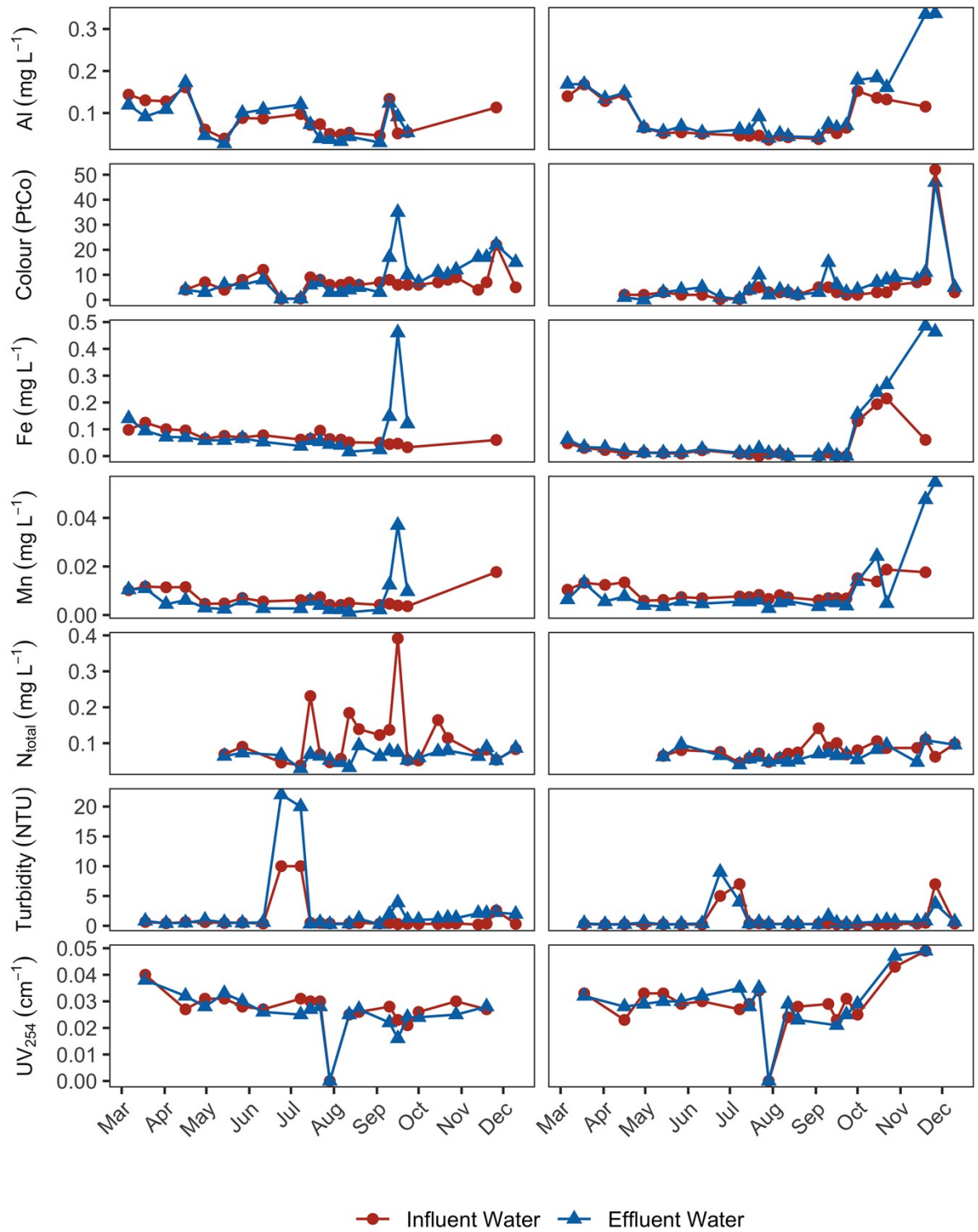
5 **General water quality**

6 Figure 1 summarizes free chlorine, pH, phosphate residual, silica residual, and TOC,
7 while Figure 2 summarizes aluminum, true colour, iron, manganese, total nitrogen,
8 turbidity, and UV₂₅₄. Both figures represent samples collected from the influent and
9 effluent of each AR.



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11 *Figure 1: Free chlorine, pH, phosphate residual, silica residual, and TOC measured in*
 12 *the influent (reservoir) and effluent water of each AR. The drop in pH in the*
 13 *influent/effluent of the silicate-treated AR represents an overcorrection (pH was*
 14 *adjusted in an attempt to maintain a stable target pH of 7.4 after doubling the sodium*
 15 *silicate dose).*



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17 Figure 2: Aluminum, true colour, iron, manganese, total nitrogen, turbidity, and UV₂₅₄
 18 measured in the influent (reservoir) and effluent water of each AR.

19 **Free chlorine**

20 During the full extent of the study median influent free chlorine concentrations in the
21 orthophosphate AR were 0.76 mg L⁻¹ (range: 0.02 - 3.75 mg L⁻¹, *n* = 26) and in the
22 sodium silicate AR were 0.63 L⁻¹ (range: 0.18 - 1.20 mg L⁻¹, *n* = 26). The
23 orthophosphate-treated system experienced a higher chlorine reduction across the AR,
24 especially during the July - September quarter (Q2). The corresponding mean free
25 chlorine reductions during Q2 were 69.6% in the orthophosphate AR, and 24.6% in the
26 sodium silicate AR.

27 **pH**

28 The pH of the influent water samples gathered during the study ranged from a median
29 of 7.35 (range: 9.03 - 6.78, *n* = 28) for the orthophosphate AR, to 7.63 (range: 8.89 -
30 6.12, *n* = 28) for the sodium silicate AR. The differences in pH between both systems
31 were not significantly different at the 95% confidence level. In the sodium silicate-
32 treated AR there was a noticeable increase in pH at the beginning of Q3 (September -
33 December) when the sodium silicate dose increased in the pipe loop systems
34 (September 05, 2019). This effect is explained by the alkaline nature of this type of
35 corrosion inhibitor.

36 **Phosphate and silica residual**

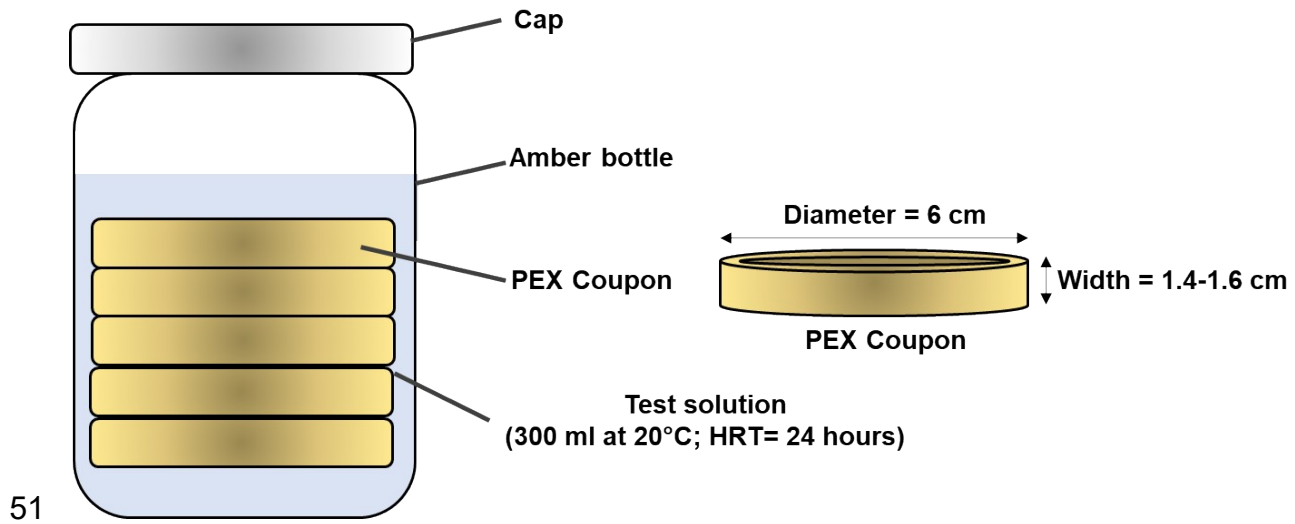
37 The median phosphate residual concentrations in the influent water were stable through
38 each quarter in contrast with the sodium silicate-treated system, mainly due to the
39 adjustment to a higher silica residual target from 24 to 48 mg L⁻¹ in September 05, 2019.
40 The median phosphate residual concentrations were 0.99 mg L⁻¹ (range: 1.17 - 0.43 mg
41 L⁻¹, *n* = 25), and the silica residual concentrations were 25.5 mg L⁻¹ (range: 67.00 -
42 18.00 mg L⁻¹, *n* = 22).

43 **Temperature**

44 Influent water temperatures recorded for each reactor system demonstrated seasonal
45 variation, as expected. Median water temperatures were higher during Q2 (July -
46 September) within each system, reaching median temperatures of 20.5 and 20.0 °C for

47 the orthophosphate and sodium silicate-treated systems, respectively. The median
48 influent water temperatures during this study were 18.0 °C (range: 23.00 - 12.00 °C, $n =$
49 29) and 17.5 °C (range: 22.50 - 12.00 °C, $n = 29$), respectively.

50 Batch Reactors



52 *Figure 3: Simplified schematic of the batch reactors.*

53 Microbial community structure

54 Taxonomic analysis

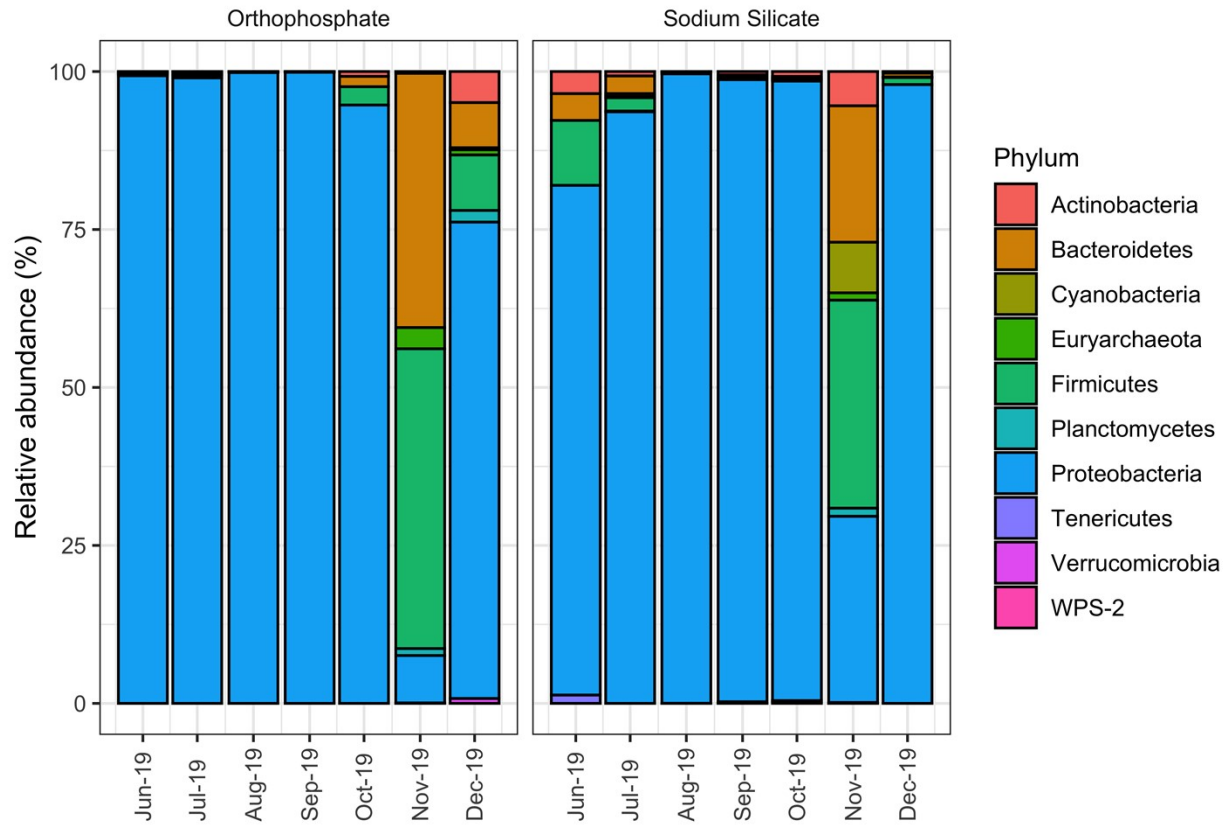
55 The mayor ASVs identified at the phylum level were *Proteobacteria*, *Bacteroidetes*,
56 *Firmicutes*, *Actinobacteria*, *Cyanobacteria*, and *Euryarchaeota* (Figure 2). At this phylum
57 level, the presence of *Proteobacteria* dominated the community structure in both
58 orthophosphate and sodium silicate-treated systems (> 74% for every month expect for
59 November). The presence of *Proteobacteria* was dominant from June to October until a
60 shift in the bacterial community structure occurred in November 2019, allowing the
61 identification of *Bacteroidetes*, *Firmicutes*, *Actinobacteria*, *Cyanobacteria*, and
62 *Euryarchaeota*. A similar presence of organisms at the phylum level have been reported
63 in drinking water biofilm studies using a variety of substrate materials, including ductile

64 cast-iron, stainless steel, tuberculated cast-iron, and PVC.¹⁻⁵ Generally, *Proteobacteria*,
65 *Firmicutes*, *Actinobacteria*, and *Bacteroidetes* have been associated with the culturable
66 portion of phosphate treated water on ductile cast-iron and stainless steel coupons;² the
67 same phyla were detected on both orthophosphate and sodium silicates treated cast-
68 iron systems, however their relative abundance (with the exception of *Proteobacteria*)
69 was higher in the orthophosphate-treated system.

70 The most noticeable difference at the phylum level between both orthophosphate and
71 sodium silicate-treated systems is the presence of *Cyanobacteria* from the sodium
72 silicate system in November, and the detection of *Euryarchaeota* from the
73 orthophosphate system, also in November. Douterelo *et al.*¹, reported that
74 *Cyanobacteria* were positively correlated with TOC levels and was present in plastic
75 pipes during the winter months (low water temperatures), which may suggest that with
76 an absence of phosphorus in the sodium silicate-treated system *Cyanobacteria* was
77 able to assimilate available carbon more effectively.

78 At the genus level the abundance of *Phreatobacter* was higher in the orthophosphate
79 system (identified from June through December, except for November), than in the
80 sodium silicate system (Figure 5). According to Perrin *et al.*⁵, the presence of
81 *Phreatobacter* in drinking water distribution systems is related to warm water
82 temperatures (>15 °C).

Phylum Composition



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84 *Figure 4: Relative abundance at the phylum level of bacterial community structure.*

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92 *Table 1: Relative abundance (%) summary of genera associated with MOB and*
 93 *pathogenic bacteria species identified from the sodium silicate and orthophosphate-*
 94 *treated AR coupons during the months of June - December 2019.*

Month	Inhibitor	Escherichia- Shigella	Halomonas	Hyphomicrobium	Legionella	Mycobacterium	Sphingomonas
Jun	P	-	-	-	-	-	1.6
Jul	P	-	0.1	0.2	-	0.2	3.2
Aug	P	-	0.0	0.0	-	0.0	10.2
Sep	P	0.0	-	0.0	0.0	0.0	15.8
Oct	P	0.2	0.6	0.2	-	0.4	9.6
Nov	P	-	7.6	-	-	-	-
Dec	P	0.1	0.5	0.9	0.1	3.3	7.9
Jun	Si	0.2	-	4.8	-	0.3	5.7
Jul	Si	-	-	2.8	-	0.6	14.4
Aug	Si	-	-	0.3	-	0.0	2.0
Sep	Si	-	0.0	0.2	0.0	0.0	0.9
Oct	Si	-	0.0	0.2	0.2	0.1	34.3
Nov	Si	0.6	1.9	-	0.2	1.7	7.8
Dec	Si	-	0.0	0.3	-	-	8.0

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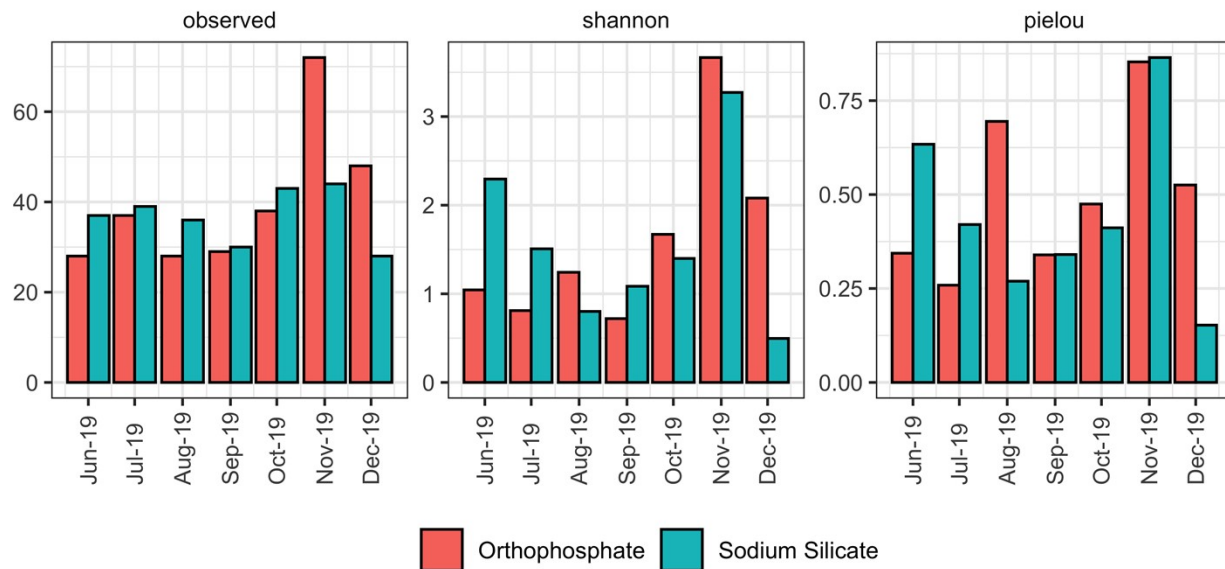
105 *Table 2: Absolute abundances of genera associated with MOB and pathogenic bacteria*
 106 *species identified from the sodium silicate and orthophosphate-treated AR coupons*
 107 *during the months of June - December 2019.*

Month	Inhibitor	Escherichia-Shigella	Halomonas	Hyphomicrobium	Legionella	Mycobacterium	Sphingomonas
Jun	P	-	-	-	-	-	843
Jun	Si	22	-	462	-	33	552
Jul	P	-	62	67	-	80	1449
Jul	Si	-	-	577	-	122	3001
Aug	P	-	21	13	-	3	17093
Aug	Si	-	-	414	-	26	2447
Sep	P	2	-	13	7	14	32735
Sep	Si	-	7	58	15	3	305
Oct	P	23	58	20	-	42	925
Oct	Si	-	4	116	93	74	20487
Nov	P	-	775	-	-	-	-
Nov	Si	14	43	-	5	39	175
Dec	P	6	39	65	9	239	574
Dec	Si	-	6	66	-	-	1591

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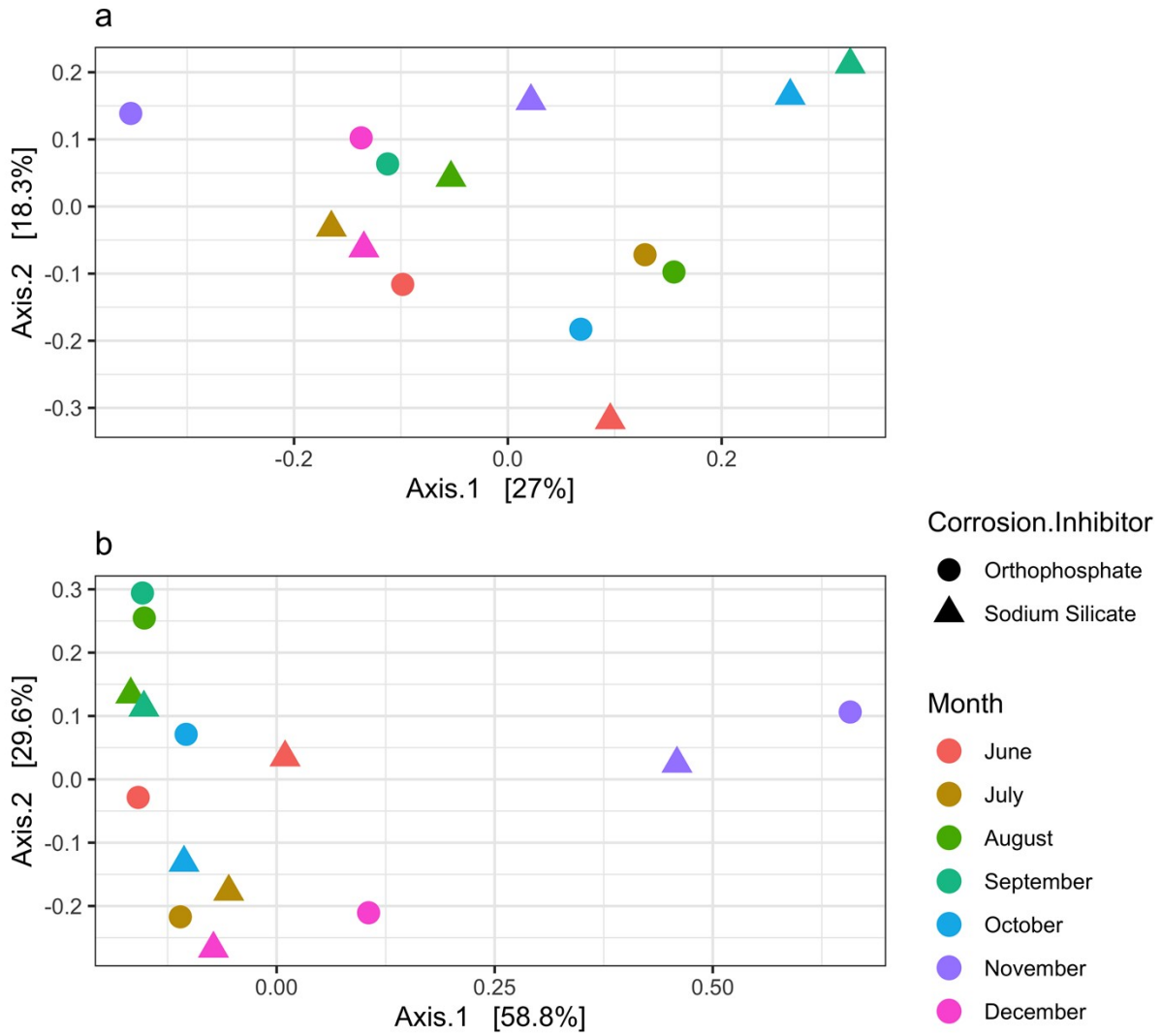
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110 Alpha and beta diversity analysis



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112 *Figure 5: Richness, Shannon diversity index, and Pielou evenness index in biofilm*
113 *samples collected from June to December 2019.*



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115 *Figure 6: (a) Principle coordinate analysis for unweighted and (b) weighted UniFrac*
 116 *distances.*

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120 References

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