2	crude fiber and may benefit the gut health without impacting the growth
3	performance in weaned pigs
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1 Wheat bran fermented by mixed fungal strains improves the digestibility of

25 1. Supplementary Materials and Methods

26 1.1 The fermentation of wheat bran

27 Wheat bran (WB) used in the current study was purchased from Xi'an Hangcheng Flour Company 28 Limited (Shanxi, China). The raw WB was sampled using a quarter method and crushed with a 29 0.45mm sample sieve. The crushed sample was then collected into an erlenmeyer flask and autoclaved 30 at 121°C for 20min for the following fermentation. Four fungal strains, Aspergillus niger GDM 3.576, 31 Neurospora crassa BNCC 336675, Trichoderma viride GDM 3.141 and Candida tropicalis GDM 2.6, 32 were used in the current study as inocula. The optimum fermentation parameters were determined 33 based on pre-trials. Briefly, a total of 5-g (NH₄)₂SO₄, 1-g KH₂PO₄, 0.6-g CaCl₂, 0.5-g MgSO₄·7H₂O, 34 0.5-g FeSO₄·7H₂O, 0.5-g ZnSO₄, 0.5-g MnSO₄ and 0.5-g CoCl₂·6H₂O were added into 1 L of 35 deionized water and autoclaved to prepare the mineral solution for each strain. The ratio of WB to mineral solution (g/v) was 1:1.2, and the ratio of inoculum to substrate was 5% (start culture 36 37 volume/wheat bran weight). The solid-state fermentation of WB was performed as follows: 38 (a) The autoclaved WB (1kg) with 5% inocula (compound fungi, Aspergillus niger, Neurospora crassa 39 and Trichoderma viride) and mineral solution (1.2 L) were well mixed and fermented for 2 days at 40 27°C to get the first-step product. 41 (b) The first-step product and 5% *Candida tropicalis* was well mixed and fermented for 1 day at 27°C

- 42 to get the second-step product.
- 43 (c) The second-step product was dried at 60°C to get the fermented WB (FWB).

44 The content of dry matter (DM), gross energy (GE), crude protein (CP), crude fiber (CF), neutral-

- 45 detergent fiber (NDF) and acid-detergent fiber (ADF) in FWB and feed samples were measured
- 46 according to the methods described by AOAC (1995). The concentration of soluble dietary fiber (SDF)

47 and Insoluble dietary fiber (IDF) in WB and FWB was analyzed using the Megazyme kit K-TDRF

- 48 (Wicklow, Ireland) based on the AOAC Method 991.43 and AACC Method 32-07.01 (Yin et al. 2004).
- 49 Both raw WB and FWB were sampled and sent to Wuhan Servicebio Technology Co., Ltd. (Hubei,
- 50 China) to observe the morphology using a scanning electron microscopy (SEM).

51 1.2 The determination of T lymphocyte subsets in blood and spleen

52 The determination of T lymphocyte subsets in jugular blood and spleen was described before (Liu et al., 53 2013). In the current study, the following four controls were used for each sample: a blank control 54 without any fluorescent antibodies and three positive controls with CD3 (FITC-Mouse Anti-Pig CD3, 55 Cat. nos. 559582), CD4 (PerCp-Cy5.5-mouse anti-pig CD4a, 561474) and CD8 (Alexa Fluor® 647-56 mouse anti pig CD8α, 561475) monoclonal fluorescent antibodies (BD Biosciences, Franklin Lakes, 57 NJ, USA). After fixing in phosphate buffer saline (PBS) buffer containing 2% formaldehyde for 30 min, 58 10,000 events (lymphocytes or Peripheral blood mononuclear cell) for each sample were collected 59 using a BD Verse flow cytometer (BD Biosciences, San Jose, CA) and data was analyzed using Flow 60 Jo software version 8.7 (Tree Star Inc., Ashland, OR, USA).

61 2. Supplementary Tables

02	Supplementary Tuble S1	The sequences of primers using in the current study	1
	Gene/Bacteria	Primer sequence (5'-3')	Reference
	0	F:TCTGGCACCACACCTTCT	1 1 2010
	<i>β-actin</i>	R:TGATCTGGGTCATCTTCTCAC	Luo et al. 2019
		F:TTTGCGTCAGTGTCATCG	Wenne (1.2010
	GAPDH	R:TGCTCTGCCTTGGGTAAT	wang <i>et al</i> . 2019
	10 014	F:CTGCCTTCCTTGGATGTG	Wana at al. 2012
	I8s rRNA	R:GCGGCTTTGGTGACTCTA	wang et al. 2012
		E-CTCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	

62 Supplementary Table S1 The sequences of primers using in the current study

	18s rRNA	T.CIUCCIICCIIUUAIUIU	Wang et al. 2012	
		R:GCGGCTTTGGTGACTCTA		
		F:GTGCCGCTGCCCACAACCTG	Luo et al. 2019	
	MUC-I	R:AGCCGGGTACCCCAGACCCA		
	pBD1	F:TGCCACAGGTGCCGATCT	Pu et al. 2018	
		R:CTGTTAGCTGCTTAAGGAATAAAGGC		
		F:CTACTCGTCCAACGGGAAAG	Luo <i>et al</i> . 2019	
	Occiuaing	R:ACGCCTCCAAGTTACCACTG		
	70.1	F:TGGCATTATTCGCCTTCATAC	Chap at $al 2012$	
	20-1	R:AGCCTCATTCGCATTGTTT	Chen <i>et al</i> . 2015	
		F:CAGATAAGCGAGGCCGTCATT	Doo at $al 2012$	
	ILK4	R:TTGCAGCCCACAAAAAGCA	Bao <i>ei ai</i> . 2012	
	M.D ⁰⁰	F:GATGGTAGCGGTTGTCTCTGAT	Chap at $al 2012$	
	МуД88	R:GATGCTGGGGGAACTCTTTCTTC	Chen <i>et al</i> . 2012	
	NF-kb	F:AGTACCCTGAGGCTATAACTCGC	Wang at al 2010	
		R:TCCGCAATGGAGGAGAAGTC	wang <i>et ut.</i> 2019	
	ΙL-1β	F:GCATGTGCTGAGCCTTTGTA	Char = (1, 2012)	
		R:CCTGGTCCTCCCAAGATTGT	Chen <i>et al</i> . 2015	
	IL-6	F:GGCTGCTTCTGGTGATGC	War a st r. 2010	
		R:AGAGATTTTGCCGAGGATGTA	wang <i>et ut.</i> 2019	
	II 10	F:GACCAGTGGGCGACTTCTT	Wang at al 2010	
	1L-10	R:ACGGCCTTGCTCTTGTTTTC	wang <i>et ut.</i> 2019	
т	atal hastaria	F:CGGTGAATACGTTCYCGG	Suzulci et al 2000	
1	otal bacteria	R:GGWTACCTTGTTACGACTT	Suzuki <i>ei ui</i> . 2000	
	Lastopacillus group	F:AGCAGTAGGGAATCTTCCA		
	Laciobacilius group	R:CACCGCTACACATGGAG	Kanno <i>et al</i> . 2009	
	Difidahaatavii	F:TCGCGTC(C/T)GGTGTGAAAG	Motoulci et al 2004	
	Булаовастегит	R:CCACATCCAGC(A/G)TCCAC		

	F:AAATGACGGTACCTGACTAA	Matsuki et al. 2002		
Clostridiumcluster XIVa	R:CTTTGAGTTTCATTCTTGCGAA			
	F:GCACA GCAGTGGAGT			
Clostridium cluster IV	R:CTTCCTCCGTTTTGTCAA	Matsuki <i>et al</i> . 2002		
	F:GCTACAATGGCGCATACAAA	1		
Escherichia coli	R:TTCATGGAGTCGAGTTGCAG	Lee <i>et al</i> . 2008		
Enterobacteriaceae	F:CATTGACGTTACCCGCAGAAGAAGC			
family	R:CTCTACGAGACTCAAGCTTGC Segata <i>et al.</i> 2			
с и	F:GTGAAATTATCGCCACGTTCGGGCA	N		
Samongella spp.	R:TCATCGCACCGTCAAAGGAACC	Xu et al. 2008		
a.	F:GAAGAATTGCTTGAATTGGTTGAA			
Streptococcus	R:GGACGGTAGTTGTTGAAGAATGG	Collado <i>et al</i> . 2009		
	F:ATGAAAAAGCTAATGTTGGC	Han <i>et al.</i> 2007		
Heat-stable toxin I(STa)	R:TACAACAAAGTTCACAGCAG			
	F:AATATCGCATTTCTTCTTGC	Han (1 2 007		
Heat-stable toxin II(S1b)	R:GCATCCTTTTGCTGCAAC	Han <i>et al</i> . 2007		
	F:TGCCATCAACACAGTATATCC	E 1 2005		
<i>astA</i>	R:TCAGGTCGCGAGTGACGGC	Ewers <i>et al.</i> 2005		
Facalihactarium prausnitzii	F: GGAGGATTGACCCCTTCAGT	Balamurugan et a		
recultoucler lum prausnitzli	R: CTGGTCCCGAAGAAACACAT	2008		
Fuhactarium ractala	F: AAGGGAAGCAAAGCTGTGAA	Balamurugan et a		
Eubucierium recluie	R: TCGGTTAGGTCACTGGCTTC	2008		
Clostridium hutoricum	F: TCAATTAGAAGGCAGAGTACC	Vi et al. 2020		
iosiriaiam Daiyricam	R: CTAAAACTGACTGTGGCATT	1101 11. 2020		
Angerostines caccae	F: GCGTAGGTGGCATGGTAAGT	Veiga et al 2010		
mucrosupes cuccue	R: CTGCACTCCAGCATGACAGT	• 01gu 01 ul. 2010		

Ingredients,%	WB	FWB	
DM	88.87	94.37	
GE, kcal/kg	3973.76	4119.08	
EE	4.31	4.46	
СР	17.95	19.33	
CF	9.42	7.11	
NDF	41.25	36.27	
ADF	29.05	22.18	
SDF	4.85	7.90	
IDF	43.20	39.76	

64 Supplementary Table S2 The level of main nutrients in WB and FWB

65 DM, dry matter; CP, crude protein; CF, crude fiber; NDF, neutral detergent fiber; ADF, acid

66 detergent fiber; SDF, soluble dietary fiber; IDF, insoluble dietary fiber; GE, gross energy; EE,

67 ether

extract.

Items ² C		WB	FWB	SE	<i>P</i> -value
IBW, kg	7.18	7.16	7.18	0.08	0.99
FBW, kg	24.34	24.65	24.44	0.49	0.75
ADFI, g/d	704.02	693.50	709.93	12.38	0.96
ADG, g/d	428.86	437.63	435.22	13.69	0.97
F/G	1.58	1.60	1.63	0.02	0.65

68 Supplementary Table S3 The growth performance of piglets in different groups¹

 69^{-1} Data are presented as means, n = 7. C, control; WB, wheat bran; FWB, fermented wheat bran;

70 SE, standard error of mean.

71 ² IBW, initial body weight; FBW, final body weight; ADG, average daily gain; ADFI, average

72 daily feed intake; F/G, feed/gain ratio.

73

74 Supplementary Table S4 The percentage of T lymphocyte subsets in the spleen and blood of piglets

75	in	different	groups	(%))
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It area?	Group			<i>P</i> -value				
Item ²	С	WB	FWB	SE	Group	C vs WB	C vs FWB	WB vs FWB
Spleen								
CD3 ⁺	60.61ª	53.74 ^b	58.37ª	1.13	0.02	0.01	0.6	0.04
$CD4^+$	27.49	24.45	28.1	1.21	0.47	NS	NS	NS
$CD8^+$	38.44	34.16	30.58	1.66	0.57	NS	NS	NS
CD4+/CD8+	0.62	0.62	1.03	0.13	0.94	NS	NS	NS
Blood								
CD3+	41.16 ^a	46.19ª	62.56 ^b	2.54	< 0.01	0.26	< 0.01	< 0.01
$CD4^+$	14.26 ^{ab}	11.47ª	18.55 ^b	1.05	0.02	0.22	0.07	< 0.01
$CD8^+$	43.22	40.11	34.25	1.57	0.38	NS	NS	NS
CD4+/CD8+	0.68	0.59	0.95	0.08	0.53	NS	NS	NS

76 ¹The different alphabetical superscripts in the same row indicate significant difference (P < 0.05, n 77 = 7). Normally distributed data are presented as means, while non-normally distributed data are 78 presented as median. Of all data, CD4⁺/CD8⁺ in the blood of piglets is non-normally distributed. C, 79 control; WB, wheat bran; FWB, fermented wheat bran; SE, standard error of mean; NS, not 80 significant. 81 ²CD3⁺, percentage of CD3⁺T lymphocytes; CD4⁺, percentage of CD4⁺T lymphocytes; CD8⁺,

82 percentage of CD8+T lymphocytes; CD4+/CD8+, ratio of CD4+T lymphocytes to CD8+T 83 lymphocytes.

84 3. Supplementary Figures



85

86 Supplementary Figure S1 The change of diarrhea rate in the piglets from different groups (n = 7). C, 87 control; WB, wheat bran; FWB, fermented wheat bran. Superscripts of lowercase letters represent 88 significant differences (P < 0.05).



8990 Supplementary Figure S2 The D-lactate and LPS concentration of blood in the different treatments.

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