

Metal-Free Regioselective Bromination of Imidazo-Heteroarenes: Dual Role of Organic Bromide Salt in Electrocatalysis

Partha Pratim Sen, Vishal Jyoti Roy and Sudipta Raha Roy*

Department of Chemistry, Indian Institute of Technology Delhi, Hauz Khas,
New Delhi, 110016, India

Phone number: (+91) 11-2659-7954; e-mail address: srr@chemistry.iitd.ac.in

Table of Content

S. No.	Content	Page No.
1	General information, materials, reaction setup	1
2	General procedure for the electrochemical bromination	2
3	General procedure for domino organo-electrochemical bromination	3
4	Resistance measurement of the reaction mixture	3
5	Voltage (V) vs Time (h) plot for the electrochemical bromination reaction	4
6	GC experiment to confirm exclusive regioselectivity	4-6
7	Reusability of the electrodes	7
8	Controlled experiment using sodium thiosulfate	7
9	Cyclic voltammetry experiments	8
10	Characterization data of the synthesized compounds	8-12
11	Reference	13
12	^1H and ^{13}C NMR Spectra	14-46

1. General information:

Electrochemical reactions were conducted using IKA Electrasyn 2.0 (at room temperature) and Keithley 2231A-30-3 triple channel DC power supply (at 50-60 °C). Commercially available reaction vials and electrodes were purchased from IKA. All the reactions were performed without taking any necessary precaution to exclude air or moisture. Cyclic voltammetry (CV) experiments were also conducted in IKA Electrasyn 2.0 using CV glassy carbon electrode, CV reference electrode and Platinum plated electrode purchased from IKA. Chromatographic purification of products was accomplished by Column chromatography on silica gel (230-400 mesh). For thin layer chromatography (TLC) analysis throughout this work, Merck pre-coated TLC plates (silica gel 60 GF₂₅₄, 0.25 mm) were employed, using UV light as the visualizing agent. Organic solutions were concentrated under reduced pressure on a Heidolph rotary evaporator. The products obtained were characterised using ¹H NMR, ¹³C NMR, FT-IR (ATR) and ESI-HRMS. NMR spectra were recorded at 400 MHz and 500 MHz for ¹H, 101 MHz and 126 MHz for ¹³C. The chemical shift (δ) for ¹H and ¹³C are given in ppm relative to internal standard/residual signals of the solvents (tetramethylsilane @ 0 ppm ¹H NMR and CDCl₃ @ 77.00 ppm ¹³C NMR). Coupling constants are given in Hertz. The following abbreviations are followed to indicate the multiplicity: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; dd, doublet of doublets; ddd, doublet of doublets of doublets. IR measurements were carried out on the Agilent ATR FT-IR spectrophotometer and the representative data are reported in terms of frequency (cm⁻¹) of absorption. High-resolution mass spectra (HRMS) were obtained from the High-Resolution Mass Spectrometry unit on MicroTOF Focus with electrospray ionization. Gas chromatography measurements were performed on a Shimadzu Chromatograph Nexis GC-2030 equipped with a Shimadzu SH-Rxi-5ms 30 meter capillary column and an FID detector.

Materials:

Synthesis grade solvents like acetonitrile (MeCN) and *N,N*-dimethylformamide (DMF) were used as purchased. Double distilled water was used as a solvent. 2-bromoacetophenones, 2-aminopyridines, tetra-*n*-butylammonium bromide (TBAB, $\geq 98\%$) and tetraethylammonium chloride (TEAC, $\geq 98\%$) were purchased from Sigma-Aldrich. All the other commercial grade reagents and solvents were purchased from Sigma-Aldrich at the highest commercial quality and used without further purification, unless otherwise stated. All the starting materials were prepared using the literature procedures.¹

Reaction setup:

The reaction setups are detailed in Figure S-01 and S-02. To perform electrolysis, Electrasyn vial of 5 mL capacity was used. Two Graphite SK-50 electrodes (0.8 cm \times 0.2 cm \times 5.2 cm) were inserted into the ElectraSyn vial cap. This vial cap was then fitted tightly to the reaction vial consisting of reaction mixture and a magnetic stirring bead.



Figure S-01: Reaction vial, vial cap and electrodes.

Electrasyn 2.0 was used for the electrochemical bromination at room temperature (Figure S-02a). However, for higher temperature Keithley 2231A-30-3 triple channel DC power supply was used as a source of DC supply (Figure S-02b).

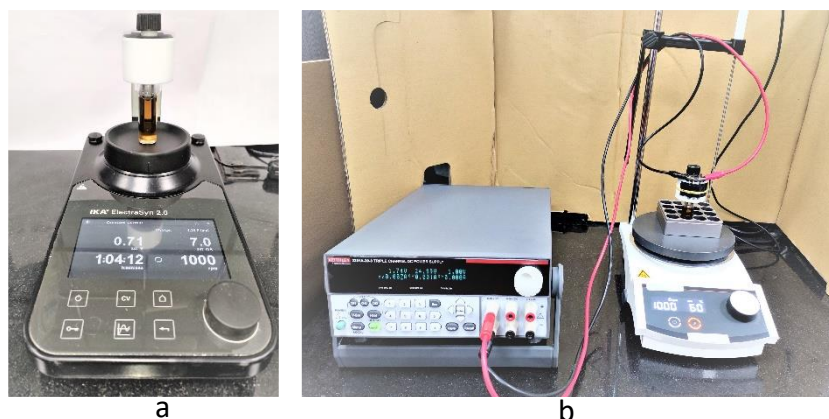


Figure S-02: (a) Reaction setup at room temperature, (b) Reaction setup at 50-60 °C.

2. General procedure for the electrochemical bromination:

(a) Electrochemical bromination at room temperature:

Without taking any precaution to remove air or moisture, an oven dried ElectroSyn vial (5 mL) equipped with magnetic stirring bar was charged with 0.2 mmol appropriate imidazo-heteroarenes (**1a-i**, **1k**, **1m-n**, **1r**, **1u-w**) and TBAB (3.0 equiv., 0.6 mmol, 193 mg). Exact 4 mL of MeCN (or in some cases a mixture of MeCN/DMF in a ratio of 3:1) was added to the ElectroSyn vial. An ElectroSyn vial cap equipped with Graphite SK-50 as an anode and the same as cathode was then fitted tightly to the reaction vial. The whole reaction mixture was stirred vigorously for 10 minutes to make the mixture homogenous. After 10 minutes of pre-stirring, the complete setup was connected to the vial holder of ElectroSyn 2.0. The reaction mixture was then electrolysed for 4 h by applying 7.0 mA constant current with 1000 rpm magnetic stirring (Figure S-02a). After completion, saturated sodium chloride solution (4.0 mL) was added to the reaction mixture and was extracted with ethyl acetate (3 × 5.0 mL), dried using anhydrous sodium sulfate, concentrated in vacuo. The residue was finally purified through silica gel (230-400 mesh) column chromatography using ethyl acetate/hexane (1:10) as eluent.

NOTE: For electrochemical chlorination reaction same procedure was followed, only TEAC was used instead of TBAB.

(b) Electrochemical bromination at 60 °C:

Without taking any necessary precaution to remove air or moisture, an oven dried ElectroSyn vial (5 mL) equipped with magnetic stirring bar was charged with 0.2 mmol appropriate imidazo-heteroarenes (**1j**, **1l**, **1o-q**, **1s-t**) and TBAB (3.0 equiv., 0.6 mmol, 193 mg). 4 mL of MeCN (or in some cases a mixture of MeCN/DMF in a ratio of 3:1) was added to the ElectroSyn vial. An ElectroSyn vial cap equipped with Graphite SK-50 as an anode and the same as cathode was then fitted tightly to the reaction vial. The vial was then placed into a heating block equipped with a temperature controlling probe. The reaction mixture was simultaneously stirred and heated for 10 minutes till the temperature of the mixture reaches 60 °C. With Keithley 2231A-30-3 triple channel DC power supply, the reaction mixture was next electrolysed for 4 h by applying 7 mA constant current with 1000 rpm magnetic stirring (Figure S-02b). After completion of the reaction, the mixture was left to cool down at room temperature. When the reaction mixture reached room temperature, saturated sodium chloride solution (4.0 mL) was added to the reaction mixture and was extracted with ethyl acetate (3 × 5.0 mL), dried using anhydrous sodium

sulfate, concentrated in vacuo. The residue was finally purified through silica gel (230-400 mesh) column chromatography using ethyl acetate/hexane (1:10) as eluent.

3. General procedure for domino organo-electrochemical bromination:

Without taking any necessary precaution to remove air or moisture, an oven dried ElectraSyn vial (5 mL) equipped with magnetic stirring bar was charged with 0.2 mmol of appropriate 2-bromoacetophenone (**4a-h**), 0.5 mmol (2.5 equiv.) of 2-aminopyridine (**5a-b**) and 30 mol% TBAB (0.06 mmol, 19 mg). 4 mL of mixed solvent system containing MeCN/water in the ratio of 3:1 was added to the reaction vial. After inserting a Electrasyn vial cap equipped with Graphite SK-50 as an anode and the same as cathode to the reaction vial, the whole system was placed in a heating block equipped with a temperature controlling probe for simultaneous pre-stirring and heating at 50 °C for 10 minutes. After that, the mixture was electrolysed at 50 °C for 7 h by applying a constant current supply of 6 mA from Keithley 2231A-30-3 triple channel DC power supply. After completion, the reaction mixture was left to settle down at room temperature. Then saturated sodium chloride solution (4.0 mL) was added to the mixture and was extracted with ethyl acetate (3 × 5.0 mL), dried using anhydrous sodium sulfate, concentrated in vacuo. The residue was finally purified using silica gel (230-400 mesh) column chromatography using ethyl acetate/hexane (1:10) as eluent.

4. Resistance measurement of the reaction mixture:

Resistance measurement was conducted using IKA Electrasyn 2.0. ElectraSyn vial (5 mL) equipped with magnetic stirring bar was charged with 0.2 mmol 6-phenylimidazo[2,1-b]thiazole (**1a**) and 4 mL MeCN. After inserting a Electrasyn vial cap equipped with Graphite SK-50 as an anode and the same as cathode to the reaction vial, the whole system was stirred for 10 minutes to attain homogeneity. Then the resistance of that solution was measured by applying a constant voltage of 2.0 V (Figure S-03a). Next, 0.2 mmol (1.0 equiv.) of TBAB was added to the same vial and resistance was measured at the same voltage after 5 minutes of pre-stirring (Figure S-03b). To the similar vial another 2.0 equivalent of TBAB was added and resistance was measured in a similar fashion (Figure S-03c).

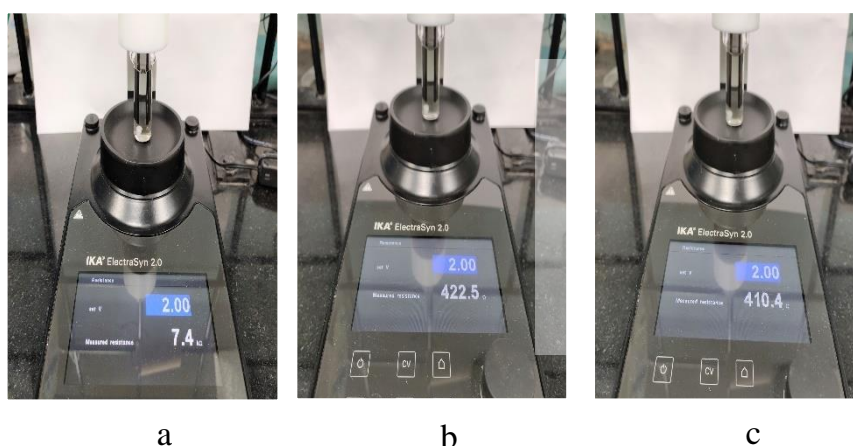
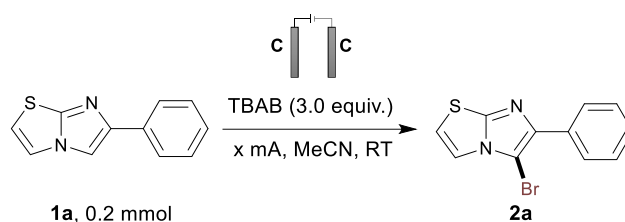


Figure S-03: Measured resistance of **1a** and TBAB in 4 mL MeCN at a constant voltage of 2.0 V, (a) resistance of the solution of 0.2 mmol **1a**, (b) resistance of the solution of 0.2 mmol **1a** + 0.2 mmol (1 equiv.) TBAB, (c) resistance of resistance of 0.2 the solution of mmol **1a** + 0.6 mmol (3.0 equiv.) TBAB.

5. Voltage (V) vs Time (h) plot for the electrochemical bromination reaction:



For the above reaction, change in the voltage was recorded at several time intervals during the electrolysis with constant current of 3 mA, 7 mA and 10 mA respectively. When we applied 3 mA current for the model reaction, after 4 h **2a** was formed in only 62% yield. On the other hand, when we have increased the current supply from 7 mA to 10 mA almost same amount (~90%) of the final product **2a** was formed in 3.5 h instead of 4 h. A plot of voltage (V) versus time (h) is shown below:

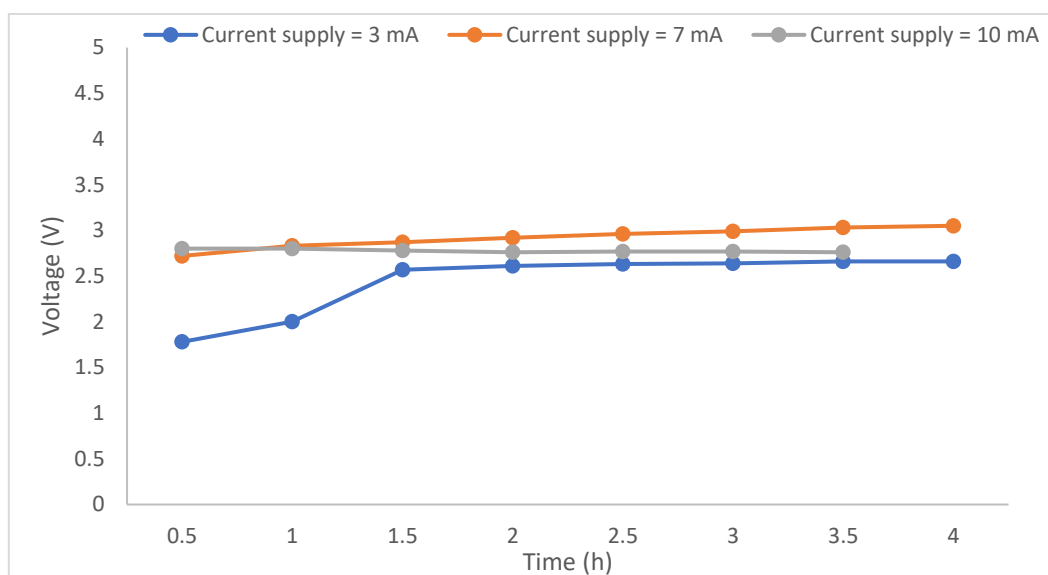
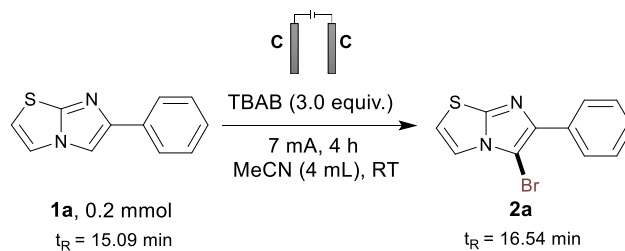


Figure S-04: Voltage vs time plot at several time intervals by applying different amount of constant current.

6. GC analysis to confirm exclusive regioselectivity:



After completion of the above model electrolysis reaction, an aliquot portion (100 μ L) was taken from the reaction mixture using a micro-pipette and the analytical sample solution was prepared in MeCN (1 mL). From the resulting solution, 2 μ L solution was then subjected to GC analysis. The GC analysis (Fig S-05) reveals the formation of mono-brominated product **2a** ($t_R = 16.54$ min) exclusively. No other regioisomeric product was observed. It is also important to mention that there was no peak related to the cyanomethylated imidazothiazole was observed.

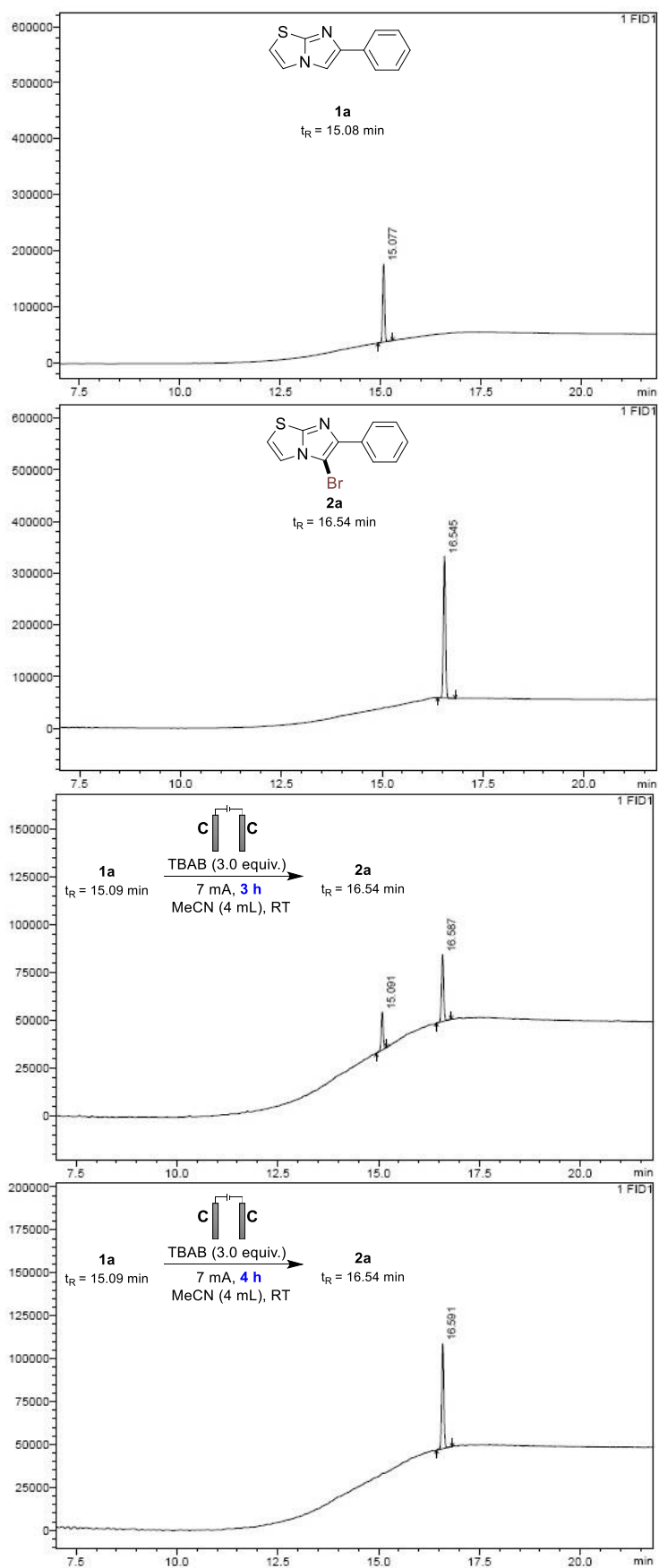


Figure S-05: GC analysis of crude reaction mixture at 3 h and 4 h.

We have also carried out the GC analysis of the crude reaction mixture for the following two reactions using the same sampling process mentioned above and got the desired single mono brominated regioisomer exclusively as mentioned in the Figure S-06 and Figure S-07:

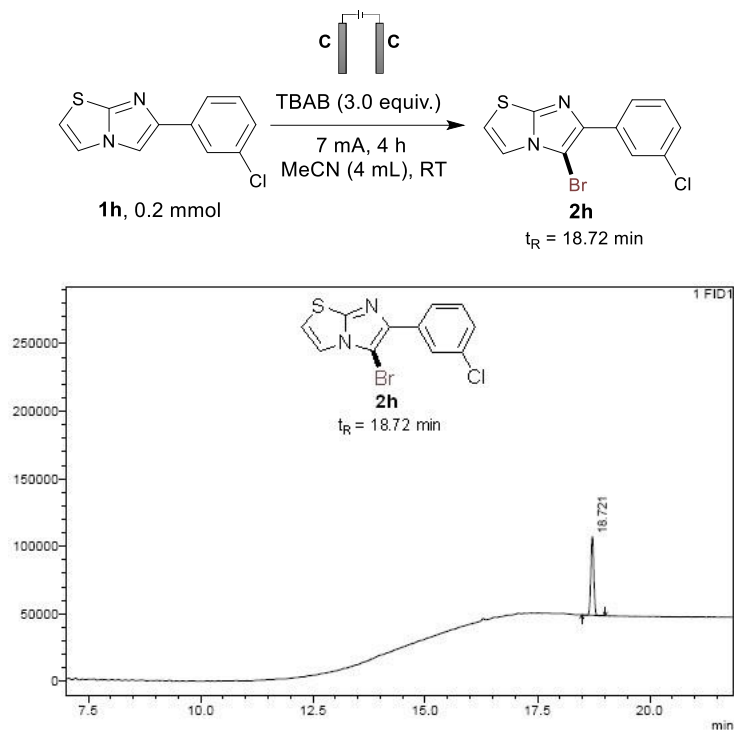


Figure S-06: GC analysis of crude reaction mixture after 4 h.

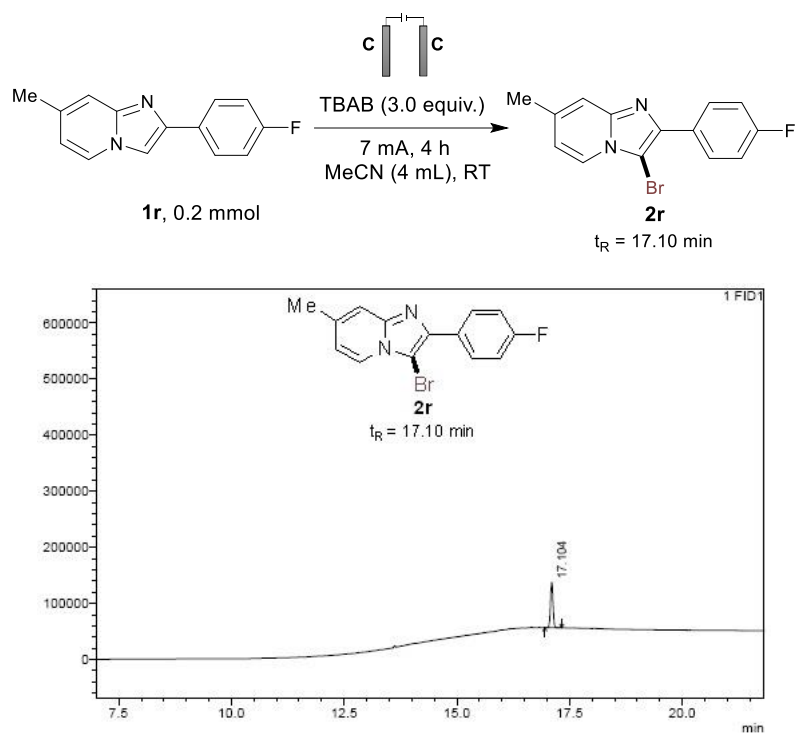


Figure S-07: GC analysis of crude reaction mixture after 4 h.

7. Reusability of the electrodes:

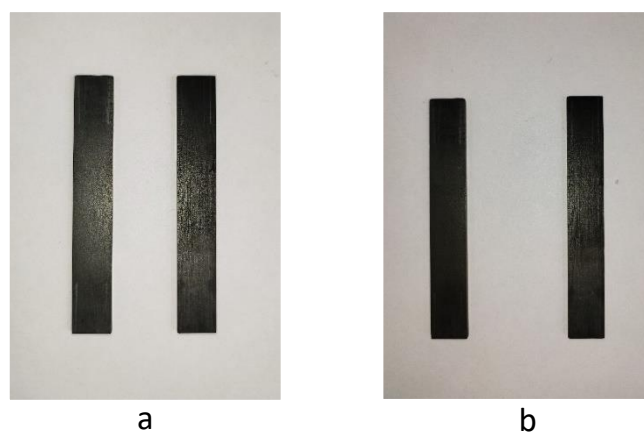


Figure S-08: (a) Electrodes before one pot scale up reaction, (b) Electrodes after one pot scale up reaction.

8. Controlled experiment using sodium thiosulfate:

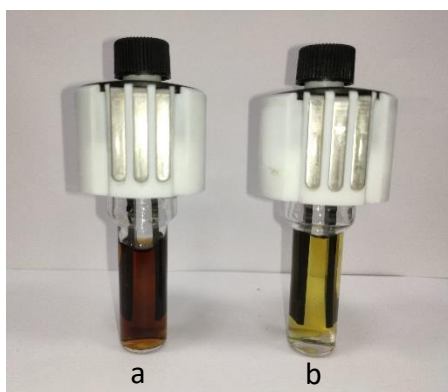
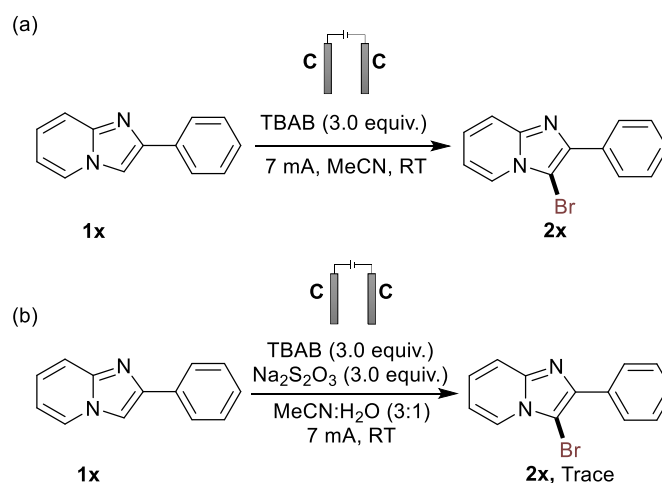


Figure S-09: (a) Reaction mixture after standard electrochemical bromination of **1x**, (b) Reaction mixture after standard electrochemical bromination of **1x** in presence of 3.0 equivalent sodium thiosulfate.

9. Cyclic voltammetry experiments:

All the cyclic voltammetry (CV) experiments were performed using Electrsyn (segment = 3, initial voltage = 0.0 V, final voltage = 2.0 V, sweep = 200 mV/s) in a vial of 5 mL capacity using 0.1 M $n\text{Bu}_4\text{NPF}_6$ (in MeCN).

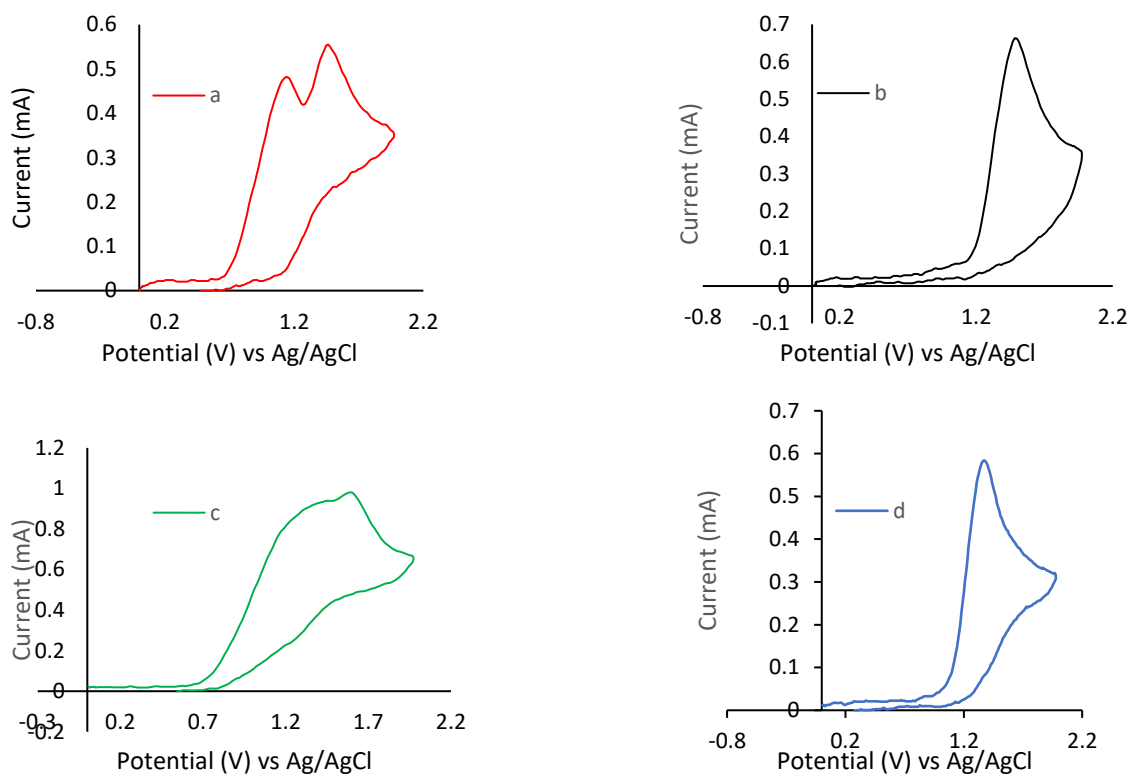


Figure S-10: (a) 0.02 M TBAB, (b) 0.02 M **1a**, (c) 0.02 M **1a** + 0.02 M TBAB, (d) 0.02 M **1x**

10. Characterization data of the synthesized compounds:

5-bromo-6-phenylimidazo[2,1-b]thiazole (2a):² white solid, 51.4 mg, 92% yield, ^1H NMR (500 MHz, CDCl_3) δ 8.02 – 8.00 (m, 1H), 7.44 – 7.41s (m, 1H), 7.34 – 7.30 (m, 1H), 6.81 (d, $J = 4.5$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 148.6, 143.7, 132.9, 128.3, 127.6, 126.7, 117.3, 112.8, 90.0.

5-bromo-6-(*p*-tolyl)imidazo[2,1-b]thiazole (2b):³ yellow solid, 44.0 mg, 75% yield, ^1H NMR (500 MHz, CDCl_3) δ 7.90 (d, $J = 8.0$ Hz, 2H), 7.39 (dd, $J = 4.4, 2.7$ Hz, 1H), 7.25 (d, $J = 7.8$ Hz, 2H), 6.88 (dd, $J = 4.4, 2.7$ Hz, 1H), 2.39 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 148.6, 144.0, 137.6, 130.2, 129.1, 126.8, 117.5, 112.7, 89.7, 21.3.

5-bromo-6-(4-methoxyphenyl)imidazo[2,1-b]thiazole (2c): off-white solid, 48.2 mg, 78% yield, mp = 125 $^\circ\text{C}$, ^1H NMR (400 MHz, CDCl_3) δ 7.94 (d, $J = 8.7$ Hz, 2H), 7.37 (d, $J = 4.5$ Hz, 1H), 6.98 (d, $J = 8.7$ Hz, 2H), 6.86 (d, $J = 4.4$ Hz, 1H), 3.84 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.2, 148.5, 143.8, 128.2, 125.7, 117.4, 113.8, 112.6, 89.1, 55.2. ATR FT-IR ν (cm^{-1}): 652, 816, 950, 1126, 1158, 1306, 1464, 2884, 2929, 2969. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{12}\text{H}_{10}\text{BrN}_2\text{OS}$, 308.9692; found, 308.9702.

5-bromo-6-(4-(trifluoromethyl)phenyl)imidazo[2,1-b]thiazole (2d): white solid, 48.6 mg, 70% yield, mp = 165 $^\circ\text{C}$, ^1H NMR (500 MHz, CDCl_3) δ 8.15 (d, $J = 8.1$ Hz, 2H), 7.68 (d, $J = 8.2$ Hz, 2H), 7.42 (d, $J = 4.5$ Hz, 1H), 6.94 (d, $J = 4.5$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 149.1, 142.4, 136.5,

129.3 (q, J_{C-F} = 32.8 Hz), 126.9, 125.4 (q, J_{C-F} = 3.8 Hz), 124.2 (q, J_{C-F} = 272.2 Hz), 117.5, 113.6, 91.1. ATR FT-IR ν (cm^{-1}): 512, 598, 650, 842, 1062, 1101, 1251, 1322, 1473, 1544, 1612, 3123, 3150. HRMS (ESI⁺) m/z : [M+H]⁺ calcd for C₁₂H₇BrF₃N₂S, 346.9460; found, 346.9468.

4-(5-bromoimidazo[2,1-b]thiazol-6-yl)benzonitrile (2e): white solid, 40.8 mg, 67% yield, mp = 206 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.17 (d, J = 8.6 Hz, 2H), 7.72 – 7.71 (m, 2H), 7.44 (d, J = 4.5 Hz, 1H), 6.98 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 149.4, 141.9, 137.5, 132.3, 127.0, 119.00, 117.5, 114.0, 110.9, 91.6. ATR FT-IR ν (cm^{-1}): 542, 660, 710, 813, 845, 968, 1124, 1245, 1319, 1397, 1467, 1533, 1603, 2219, 2853, 2923, 3109, 3145. HRMS (ESI⁺) m/z : [M+H]⁺ calcd for C₁₂H₇BrN₃S, 303.9539; found, 303.9546.

5-bromo-6-(4-fluorophenyl)imidazo[2,1-b]thiazole (2f):^{1a} white solid, 47.5 mg, 80% yield, ¹H NMR (400 MHz, CDCl₃) δ 8.00 – 7.97 (m, 2H), 7.39 (d, J = 4.5 Hz, 1H), 7.12 (t, J = 8.8 Hz, 2H), 6.90 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 162.4 (d, J_{C-F} = 247.0 Hz), 148.7, 143.1, 129.2 (d, J_{C-F} = 3.8 Hz), 128.6 (d, J_{C-F} = 7.6 Hz), 117.5, 115.3 (d, J_{C-F} = 21.4 Hz), 113.0, 89.8.

5-bromo-6-(4-chlorophenyl)imidazo[2,1-b]thiazole (2g):³ white solid, 52.0 mg, 83% yield, ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, J = 8.6 Hz, 2H), 7.41 – 7.39 (m, 3H), 6.90 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 148.9, 142.8, 133.5, 131.5, 128.6, 128.0, 117.4, 113.2, 90.2.

5-bromo-6-(3-chlorophenyl)imidazo[2,1-b]thiazole (2h): white solid, 53.3 mg, 85% yield, mp = 126 °C, ¹H NMR (500 MHz, CDCl₃) δ 8.03 (t, J = 1.8 Hz, 1H), 7.92 – 7.90 (m, 1H), 7.38 (d, J = 4.5 Hz, 1H), 7.35 (t, J = 7.9 Hz, 1H), 7.29 (ddd, J = 8.0, 2.0, 1.1 Hz, 1H), 6.90 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 148.9, 142.3, 134.8, 134.4, 129.6, 127.6, 126.7, 124.7, 117.4, 113.3, 90.5. ATR FT-IR ν (cm^{-1}): 652, 689, 786, 817, 852, 897, 1078, 1132, 1462, 1593, 2922, 3125. HRMS (ESI⁺) m/z : [M+H]⁺ calcd for C₁₁H₇BrClN₂S, 312.9196; found, 312.9211.

5-bromo-6-(4-bromophenyl)imidazo[2,1-b]thiazole (2i): yellowish solid, 63.0 mg, 88% yield, mp = 136 °C, ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, J = 8.5 Hz, 2H), 7.55 (d, J = 8.5 Hz, 2H), 7.39 (d, J = 4.5 Hz, 1H), 6.91 (d, J = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 148.9, 142.8, 132.0, 131.5, 128.3, 121.8, 117.5, 113.2, 90.2. ATR FT-IR ν (cm^{-1}): 661, 709, 823, 965, 1006, 1099, 1245, 1324, 1393, 1462, 1514, 2854, 2922, 3101, 3127. HRMS (ESI⁺) m/z : [M+H]⁺ calcd for C₁₁H₇Br₂N₂S, 358.8691; found, 358.8681.

5-bromo-6-(4-chlorophenyl)-2-methylimidazo[2,1-b]thiazole (2j): off-white solid, 34.0 mg, 52% yield, mp = 160 °C, ¹H NMR (400 MHz, CDCl₃) δ 7.94 (d, J = 8.6 Hz, 2H), 7.39 (d, J = 8.6 Hz, 2H), 7.12 (d, J = 1.3 Hz, 1H), 2.45 (d, J = 1.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 148.3, 141.5, 133.3, 131.7, 128.5, 127.9, 127.3, 114.0, 89.8, 14.5. ATR FT-IR ν (cm^{-1}): 679, 714, 747, 821, 965, 1090, 1339, 1464, 1515, 2922, 3741. HRMS (ESI⁺) m/z : [M+H]⁺ calcd for C₁₂H₉BrClN₂S, 326.9353; found, 326.9376.

3-bromo-2-(4-chlorophenyl)imidazo[1,2-a]pyridine (2k):⁴ white solid, 46.1 mg, 75% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, J = 6.7 Hz, 1H), 8.08 (d, J = 8.2 Hz, 2H), 7.63 (d, J = 8.9 Hz, 1H), 7.45 (d, J = 8.2 Hz, 2H), 7.27 (t, J = 7.5 Hz, 1H), 6.94 (t, J = 6.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 145.5, 141.5, 134.2, 131.4, 129.0, 128.7, 125.3, 124.0, 117.6, 113.2, 91.7.

3-bromo-2-(4-bromophenyl)imidazo[1,2-a]pyridine (2l):⁵ white solid, 56.32 mg, 80% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, J = 6.8 Hz, 1H), 8.02 (d, J = 8.4 Hz, 2H), 7.63 – 7.59 (m, 3H), 7.27 (t, J = 7.6 Hz, 1H), 6.94 (t, J = 6.6 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 145.5, 141.5, 131.8, 131.6, 129.3, 125.3, 124.0, 122.5, 117.6, 113.2, 91.8.

3-bromo-2-(4-fluorophenyl)imidazo[1,2-a]pyridine (2m):⁴ white solid, 42.5 mg, 73% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.17 (d, J = 6.9 Hz, 1H), 8.13 – 8.10 (m, 2H), 7.63 (d, J = 9.1 Hz, 1H), 7.28 – 7.25 (m, 1H), 7.17 (t, J = 8.7 Hz, 2H), 6.94 (dd, J = 9.9, 3.7 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ

162.8 (d, J_{C-F} = 249.5 Hz), 145.4, 141.8, 129.6 (d, J_{C-F} = 7.6 Hz), 129.0 (d, J_{C-F} = 3.8 Hz), 125.2, 123.9, 117.6, 115.4 (d, J_{C-F} = 21.4 Hz), 113.1, 91.4.

3-bromo-7-methyl-2-(4-(trifluoromethyl)phenyl)imidazo[1,2-a]pyridine (2n): off-white solid, 54.7 mg, 77% yield, mp = 111 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.24 (d, J = 8.1 Hz, 2H), 8.02 (d, J = 7.0 Hz, 1H), 7.71 (d, J = 8.2 Hz, 2H), 7.38 (s, 1H), 6.76 (dd, J = 7.0, 1.5 Hz, 1H), 2.43 (d, J = 0.7 Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 145.9, 140.7, 136.7, 136.6, 129.8 (q, J_{C-F} = 32.8 Hz), 127.80, 125.3 (q, J_{C-F} = 3.8 Hz), 124.5 (q, J_{C-F} = 289 Hz), 123.13, 116.12, 116.00, 91.56, 21.29. ATR FT-IR ν (cm^{-1}): 600, 650, 690, 843, 1063, 1106, 1158, 1321, 1615, 2925. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{15}\text{H}_{11}\text{BrF}_3\text{N}_2$, 355.0052; found, 355.0054.

4-(3-bromo-7-methylimidazo[1,2-a]pyridin-2-yl)benzotrile (2o): white solid, 37.4 mg, 60% yield, mp = 192 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.27 (dd, J = 8.3, 1.5 Hz, 2H), 8.06 (d, J = 7.0 Hz, 1H), 7.74 (dd, J = 8.4, 1.8 Hz, 2H), 7.39 (s, 1H), 6.81 (d, J = 7.0 Hz, 1H), 2.45 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 146.0, 140.1, 137.6, 137.1, 132.2, 128.0, 123.2, 119.0, 116.3, 116.2, 111.3, 92.1, 21.4. ATR FT-IR ν (cm^{-1}): 599, 669, 767, 845, 1020, 1091, 1349, 1469, 1534, 1644, 2218, 2853, 2919, 3060, 3741. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{15}\text{H}_{11}\text{BrN}_3$, 312.0131; found, 312.0136.

3-bromo-2-(4-methoxyphenyl)-7-methylimidazo[1,2-a]pyridine (2p):⁶ white solid, 44.4 mg, 76% yield, ^1H NMR (500 MHz, CDCl_3) δ 8.07 – 8.04 (m, 2H), 8.02 (d, J = 7.0 Hz, 1H), 7.37 (s, 1H), 7.01 (d, J = 8.8 Hz, 2H), 6.74 (dd, J = 7.0, 1.2 Hz, 1H), 3.86 (s, 3H), 2.43 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 159.6, 145.7, 142.2, 135.9, 129.1, 125.6, 123.0, 115.8, 115.4, 113.8, 89.9, 55.3, 21.3.

3-bromo-2-(4-chlorophenyl)-7-methylimidazo[1,2-a]pyridine (2q): white solid, 47.0 mg, 73% yield, mp = 147 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.06 (d, J = 8.5 Hz, 2H), 8.02 (d, J = 7.0 Hz, 1H), 7.43 (d, J = 8.5 Hz, 2H), 7.37 (s, 1H), 6.75 (dd, J = 7.0, 1.0 Hz, 1H), 2.43 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 145.8, 141.2, 136.4, 134.0, 131.5, 128.9, 128.6, 123.1, 116.0, 115.8, 90.8, 21.3. ATR FT-IR ν (cm^{-1}): 598, 664, 719, 771, 827, 1015, 1091, 1159, 1346, 1642, 1727, 2853, 2918, 3062, 3740. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{14}\text{H}_{11}\text{BrClN}_2$, 320.9789; found, 320.9760.

3-bromo-2-(4-fluorophenyl)-7-methylimidazo[1,2-a]pyridine (2r): white solid, 42.7 mg, 70% yield, mp = 110 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.10 – 8.07 (m, 2H), 8.02 (d, J = 7.0 Hz, 1H), 7.37 (s, 1H), 7.17 – 7.14 (m, 2H), 6.75 (dd, J = 7.0, 1.3 Hz, 1H), 2.42 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 162.7 (d, J_{C-F} = 248.2 Hz), 145.8, 141.5, 136.3, 129.5 (d, J_{C-F} = 7.6 Hz), 129.2 (d, J_{C-F} = 3.8 Hz), 123.1, 115.9, 115.7, 115.5 (d, J_{C-F} = 21.4 Hz), 90.5, 21.3. ATR FT-IR ν (cm^{-1}): 605, 662, 764, 826, 988, 1018, 1151, 1216, 1479, 2852, 2922, 3037, 3745. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{14}\text{H}_{11}\text{BrFN}_2$, 305.0084; found, 305.0083.

3-bromo-7-chloro-2-(4-fluorophenyl)imidazo[1,2-a]pyridine (2s): white solid, 46.9 mg, 72% yield, mp = 143 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.09 – 8.06 (m, 3H), 7.61 (d, J = 1.4 Hz, 1H), 7.18 – 7.15 (m, 2H), 6.91 (dd, J = 7.3, 2.0 Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 162.9 (d, J_{C-F} = 249.5 Hz), 145.0, 142.7, 131.8, 129.6 (d, J_{C-F} = 7.6 Hz), 128.6 (d, J_{C-F} = 3.8 Hz), 124.2, 116.3, 115.5 (d, J_{C-F} = 21.4 Hz), 114.7, 91.6. ATR FT-IR ν (cm^{-1}): 597, 654, 730, 770, 800, 830, 932, 985, 1066, 1143, 1216, 1348, 1478, 1625, 3033, 3103. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{13}\text{H}_8\text{BrClFN}_2$, 324.9538; found, 324.9540.

3,6-dibromo-2-(4-chlorophenyl)imidazo[1,2-a]pyridine (2t): white solid, 50.2 mg, 65 % yield, mp = 186 °C, ^1H NMR (400 MHz, CDCl_3) δ 8.32 (d, J = 0.7 Hz, 1H), 8.06 (d, J = 8.5 Hz, 2H), 7.52 (d, J = 9.5 Hz, 1H), 7.45 (d, J = 8.5 Hz, 2H), 7.33 (dd, J = 9.5, 1.7 Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 143.9, 142.4, 134.6, 130.9, 129.0, 128.9, 128.8, 124.2, 118.2, 108.1, 92.0. ATR FT-IR ν (cm^{-1}): 722, 793, 1080, 1322, 1456, 1517, 2922, 3071. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{13}\text{H}_8\text{Br}_2\text{ClN}_2$, 386.8717; found, 386.8718.

5-bromo-6-(tert-butyl)imidazo[2,1-b]thiazole (2u):^{7a} white solid, 41.4 mg, 80% yield, ¹H NMR (500 MHz, CDCl₃) δ 7.32 (d, *J* = 4.5 Hz, 1H), 6.83 (d, *J* = 4.5 Hz, 1H), 1.44 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 153.0, 146.5, 117.3, 112.0, 88.2, 33.4, 29.89.

5-bromo-6-methylimidazo[2,1-b]thiazole (2v):^{7a} off white solid, 32.1 mg, 74% yield, ¹H NMR (500 MHz, CDCl₃) δ 7.29 (d, *J* = 4.5 Hz, 1H), 6.83 (d, *J* = 4.5 Hz, 1H), 2.32 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 147.9, 142.5, 117.3, 111.9, 91.1, 13.6

3-bromo-2-(tert-butyl)imidazo[1,2-a]pyridine (2w):^{8a} brown oil, 39.5 mg, 78% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.12 (d, *J* = 6.9 Hz, 1H), 7.59 (d, *J* = 9.0 Hz, 1H), 7.20-7.17 (m, 1H), 6.88-6.86 (m, 1H), 1.53 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 152.0, 143.8, 124.0, 123.2, 117.3, 112.5, 90.0, 33.1, 29.7.

3-bromo-2-phenylimidazo[1,2-a]pyridine (2x):⁴ white solid, 35.5 mg, 65% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.14 – 8.12 (m, 2H), 7.98 (d, *J* = 6.9 Hz, 1H), 7.55 (d, *J* = 9.0 Hz, 1H), 7.45 (t, *J* = 7.7 Hz, 2H), 7.34 (t, *J* = 7.4 Hz, 1H), 7.12 – 7.09 (m, 1H), 6.74 (dd, *J* = 10.0, 3.6 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 145.0, 142.2, 132.6, 128.1, 127.9, 127.5, 124.7, 123.5, 117.1, 112.6, 91.3.

4-(3-bromoimidazo[1,2-a]pyridin-2-yl)benzotrile (2y):^{7b} white solid, 29.8 mg, 50% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.30 – 8.28 (m, 2H), 8.20 (dt, *J* = 6.9, 1.0 Hz, 1H), 7.77 – 7.75 (m, 2H), 7.65 (dd, *J* = 9.1, 0.9 Hz, 1H), 7.32 (ddd, *J* = 9.1, 6.8, 1.2 Hz, 1H), 6.99 (td, *J* = 6.8, 1.0 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 145.7, 140.5, 137.4, 132.2, 128.1, 125.9, 124.1, 118.9, 117.9, 113.6, 111.5, 92.9.

3-bromo-2-(4-(trifluoromethyl)phenyl)imidazo[1,2-a]pyridine (2z):⁴ white solid, 38.2 mg, 56% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.27 (d, *J* = 8.1 Hz, 2H), 8.19 (d, *J* = 6.9 Hz, 1H), 7.73 (d, *J* = 8.2 Hz, 2H), 7.65 (d, *J* = 9.1 Hz, 1H), 7.30 (ddd, *J* = 9.0, 6.8, 1.1 Hz, 1H), 6.97 (td, *J* = 6.8, 0.8 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 145.6, 141.1, 136.4, 130.0 (q, *J*_{C-F} = 32.8 Hz), 128.0, 125.6, 125.4 (q, *J*_{C-F} = 3.8 Hz), 124.2 (q, *J*_{C-F} = 272.2 Hz), 124.1, 117.8, 113.4, 92.5.

3-bromo-7-methyl-2-phenylimidazo[1,2-a]pyridine (2aa):^{8b} white solid, 47.1 mg, 62% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.12 – 8.11 (m, 2H), 8.03 (d, *J* = 7.0 Hz, 1H), 7.49 – 7.46 (m, 2H), 7.39 – 7.36 (m, 2H), 6.74 (dd, *J* = 7.0, 1.4 Hz, 1H), 2.43 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 145.8, 142.3, 136.1, 133.0, 128.4, 128.1, 127.8, 123.1, 116.0, 115.1, 90.8, 21.3.

3-bromo-7-methyl-2-(*p*-tolyl)imidazo[1,2-a]pyridine (2ab):⁶ white solid, 40.4 mg, 67% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 8.2 Hz, 2H), 7.92 (d, *J* = 0.9, 1H), 7.52 (d, *J* = 9.1 Hz, 1H), 7.28 (d, *J* = 7.9 Hz, 2H), 7.08 (dd, *J* = 9.1, 1.6 Hz, 1H), 2.40 (s, 3H), 2.37 (d, *J* = 0.9 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 144.4, 142.4, 138.0, 130.1, 129.1, 128.1, 127.6, 122.7, 121.6, 116.8, 90.9, 21.3, 18.3.

5-chloro-6-phenylimidazo[2,1-b]thiazole (3a):¹ white solid, 36.6 mg, 77% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.02 – 7.99 (m, 2H), 7.46 – 7.43 (m, 2H), 7.39 (d, *J* = 4.5 Hz, 1H), 7.35 – 7.31 (m, 1H), 6.89 (d, *J* = 4.5 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 147.3, 141.0, 132.8, 128.5, 127.6, 126.5, 116.6, 113.2, 106.1.

5-chloro-6-(4-fluorophenyl)imidazo[2,1-b]thiazole (3b): white solid, 39.9 mg, 79% yield, mp = 112 °C, ¹H NMR (500 MHz, CDCl₃) δ 7.99 – 7.96 (m, 2H), 7.38 (d, *J* = 4.4 Hz, 1H), 7.14 – 7.11 (m, 2H), 6.90 (d, *J* = 4.4 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 162.3 (d, *J*_{C-F} = 249.2 Hz), 147.3, 140.2, 128.9 (d, *J*_{C-F} = 2.5 Hz), 128.3 (d, *J*_{C-F} = 7.6 Hz), 116.6, 115. (d, *J*_{C-F} = 21.4 Hz), 113.3, 105.8. ATR FT-IR ν (cm⁻¹): 600, 661, 710, 773, 826, 980, 1088, 1219, 1328, 1464, 1535, 1608, 2922, 2957, 3046, 3100, 3138. HRMS (ESI⁺) *m/z*: [M+H]⁺ calcd for C₁₁H₇ClFN₂S, 252.9997; found, 252.9996.

3-chloro-2-phenylimidazo[1,2-a]pyridine (3c):⁹ yellow solid, 36.6 mg, 80% yield, ¹H NMR (500 MHz, CDCl₃) δ 8.15 – 8.13 (m, 2H), 8.09 (dt, *J* = 6.9, 1.1 Hz, 1H), 7.63 (dt, *J* = 9.1, 0.9 Hz, 1H), 7.50 – 7.47 (m, 2H), 7.40 – 7.36 (m, 1H), 7.23 (ddd, *J* = 9.0, 6.8, 1.2 Hz, 1H), 6.91 (td, *J* = 6.8, 1.0 Hz, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 143.7, 139.8, 132.5, 128.5, 128.2, 127.4, 124.8, 122.6, 117.6, 112.8, 105.6.

3-chloro-7-methyl-2-(4-(trifluoromethyl)phenyl)imidazo[1,2-a]pyridine (3d): white solid, 46.0 mg, 74% yield, mp = 102 °C, ^1H NMR (500 MHz, CDCl_3) δ 8.24 (d, J = 8.2 Hz, 2H), 7.95 (d, J = 7.0 Hz, 1H), 7.70 (d, J = 8.3 Hz, 2H), 7.37 (s, 1H), 6.76 (dd, J = 7.0, 1.3 Hz, 1H), 2.42 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 144.2, 137.8, 136.5, 136.2, 129.7 (q, $J_{\text{C-F}}$ = 32.8 Hz), 128.7, 127.8, 127.4, 125.35 (q, $J_{\text{C-F}}$ = 3.8 Hz), 124.4 (q, $J_{\text{C-F}}$ = 272.2 Hz), 121.9, 116.1, 115.9, 105.8, 21.3. ATR FT-IR ν (cm^{-1}): 685, 767, 840, 1008, 1064, 1099, 1322, 1164, 1322, 1616, 2923, 2962, 3036. HRMS (ESI⁺) m/z : $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{15}\text{H}_{11}\text{ClF}_3\text{N}_2$, 311.0557; found, 311.0556.

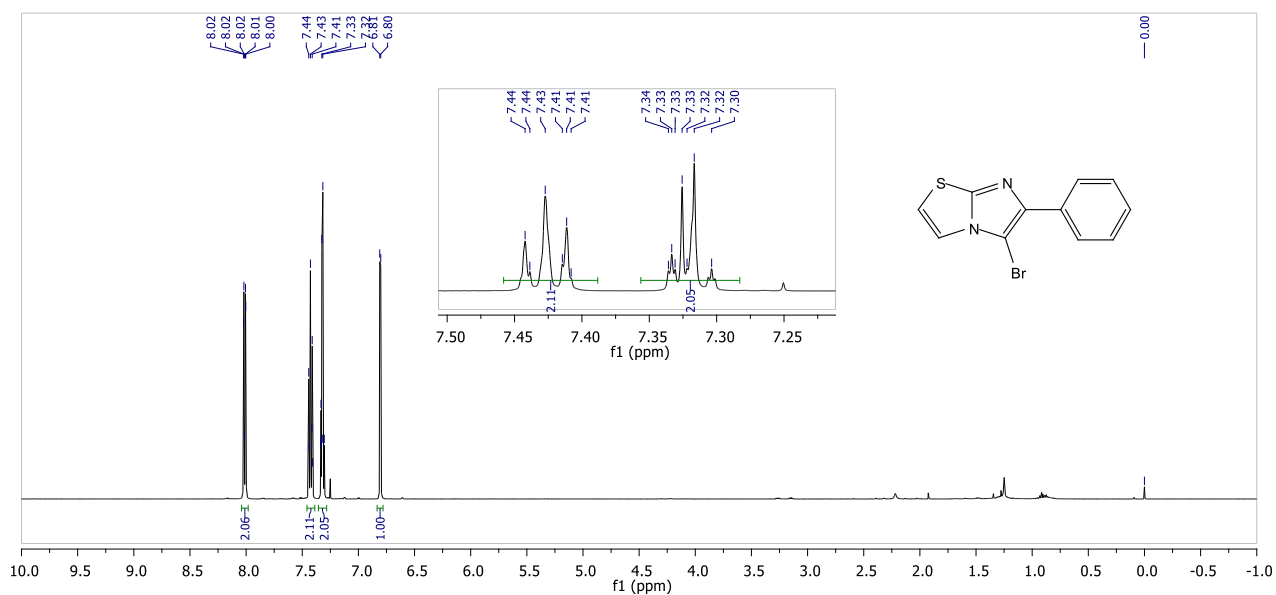
2-(tert-butyl)-3-chloroimidazo[1,2-a]pyridine (3e):^{8a} yellow oil, 27.5 mg, 66% yield, ^1H NMR (500 MHz, CDCl_3) δ 8.04 (d, J = 6.9 Hz, 1H), 7.58 (d, J = 9.1 Hz, 1H), 7.18 – 7.15 (m, 1H), 6.88-6.86 (m, 1H), 1.50 (s, 9H). ^{13}C NMR (126 MHz, CDCl_3) δ 149.5, 142.2, 123.7, 122.0, 117.3, 112.4, 104.4, 32.9, 29.5.

11. References:

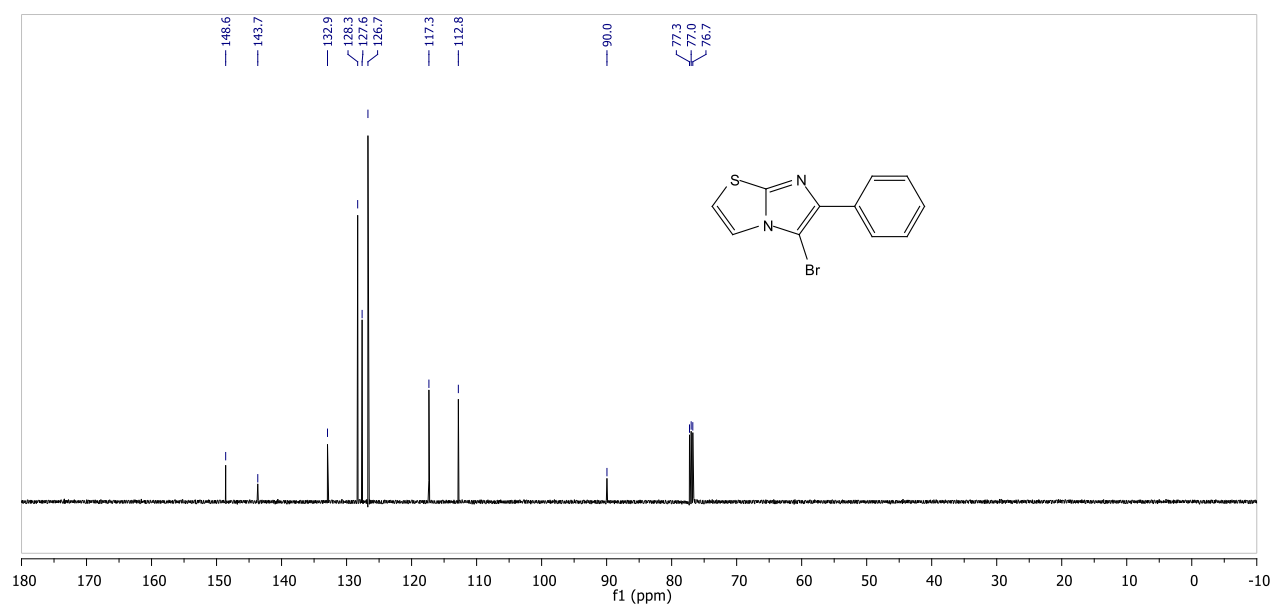
1. (a) M. A. ODaly, C. P. Hopkinson, G. D. Meakins and A. J. Raybould, *J. Chem. SOC., Perkin Trans. 1*, 1991, 855; (b) Y. Rival, G. Grassy and G. Michel, *Chem. Pharm. Bull.*, 1992, **40**, 1170; (c) **For the synthesis of monobromoacetone**: M. Musiejuk, J. Doroszuk, B. Jędrzejewski, G. O. Nieto, M. Navarro and D. Witt, *Adv. Synth. Catal.*, 2020, **362**, 618
2. A. Andreani, M. Rambaldi and A. Locatelli, *Collect. Czech. Chem. Commun.*, 1991, **56**, 2430.
3. R. Semwal, C. Ravi, R. Kumar, R. Meena and S. Adimurthy, *J. Org. Chem.*, 2019, **84**, 792.
4. X. Zhou, H. Yan, C. Ma, Y. He, Y. Li, J. Cao, R. Yan and G. Huang, *J. Org. Chem.*, 2016, **81**, 25.
5. H. Salgado-Zamora, M. Velazquez, D. Mejia; M. E. Campos-Aldrete, R. Jimenez, H. Cervantes, *Heterocycl. Commun.*, 2011, **14**, 27.
6. D. R. Indukuri, G. R. Potuganti and M. Alla, *Synlett*, 2019, **30**, 1573.
7. (a) M. A. ODaly, C. P. Hopkinson, G. D. Meakins and A. J. Raybould, *J. Chem. SOC., Perkin Trans. 1*, 1991, 855; (b) J. Weisner, R. Gontla, L. van der Westhuizen, S. Oeck, J. Ketzer, P. Janning, A. Richters, T. Mühlenberg, Z. Fang, A. Taher, V. Jendrosseck, S. C. Pelly, S. Bauer, W. A. van Otterlo and D. Rauh, *Angew. Chem., Int. Ed.*, 2015, **54**, 10313.
8. (a) J. Li, J. Tang, Y. Wu, Q. He and Y. Yu, *RSC Adv.*, 2018, **8**, 5058; (b) Y. Liu, L. Lu, H. Zhou, F. Xu, C. Ma, Z. Huang, J. Xu and S. Xu, *RSC Adv.*, 2019, b, 34671.
9. X. Xiao, Y. Xie, S. Bai, Y. Deng, H. Jiang and W. Zeng, *Org. Lett.*, 2015, **17**, 3998.

12. ^1H and ^{13}C NMR Spectra:

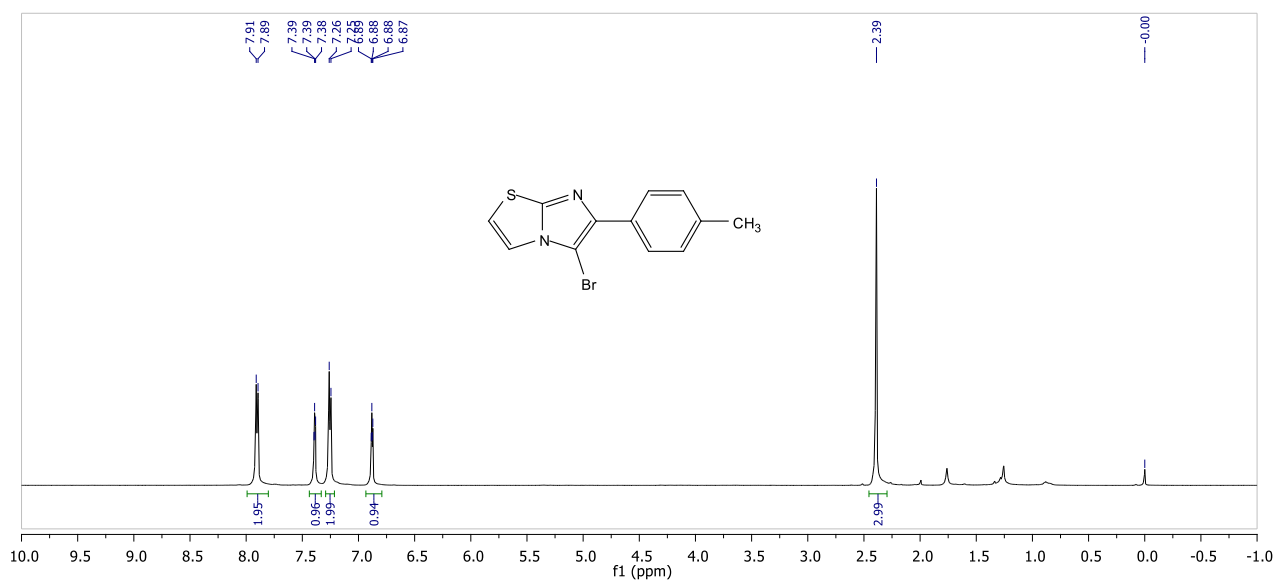
^1H NMR (500 MHz, CDCl_3) of compound **2a**



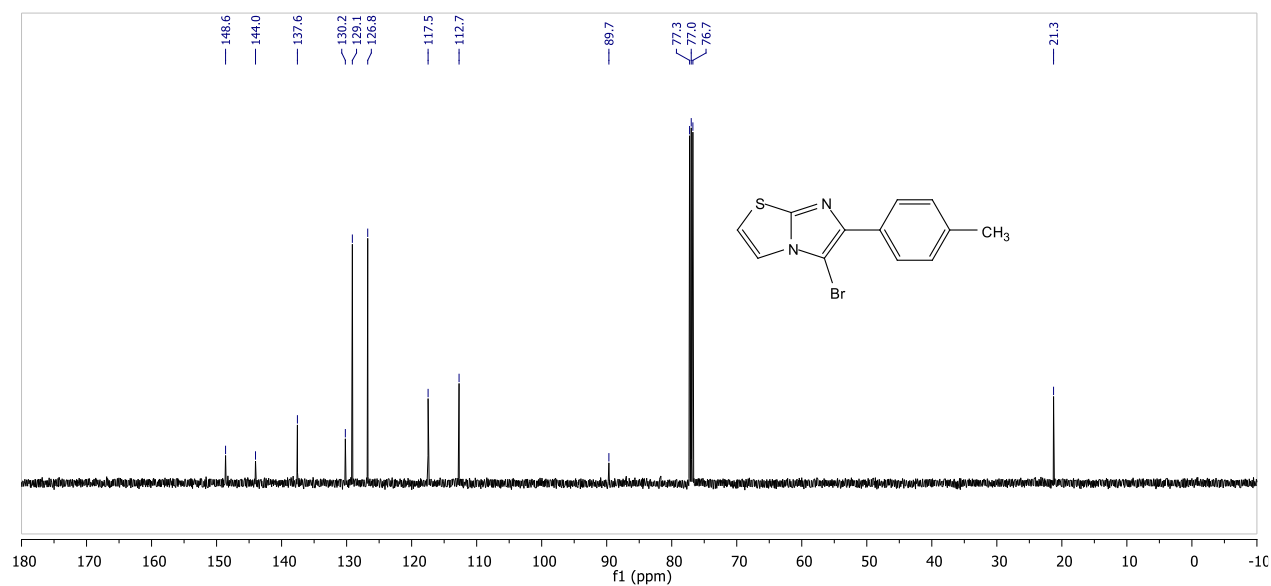
^{13}C NMR (126 MHz, CDCl_3) of compound **2a**



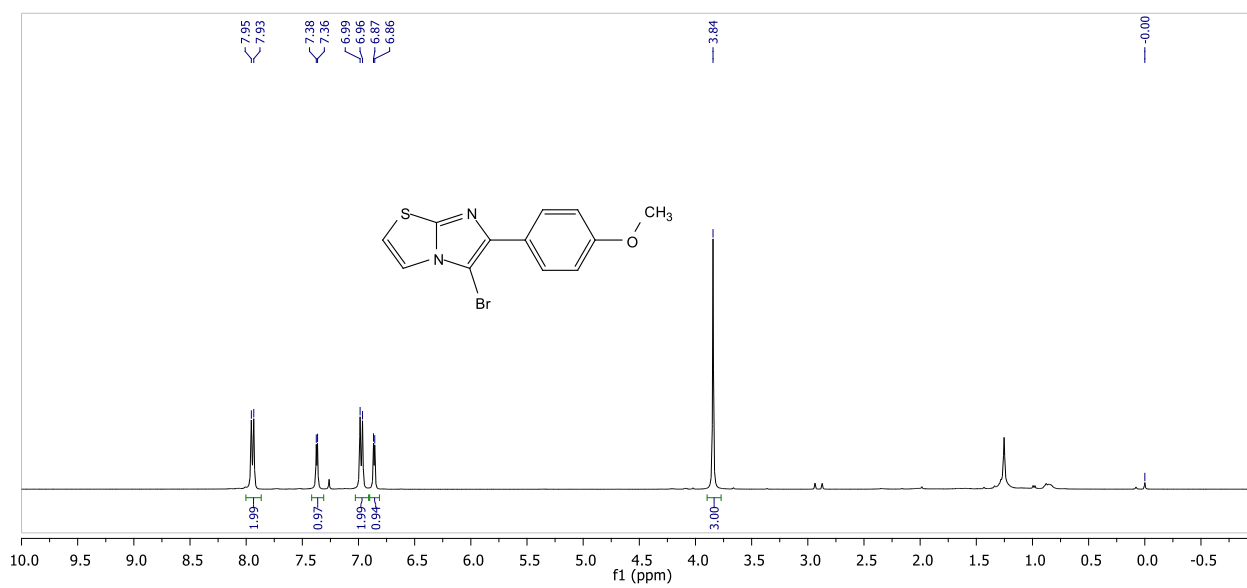
¹H NMR (500 MHz, CDCl₃) of compound **2b**



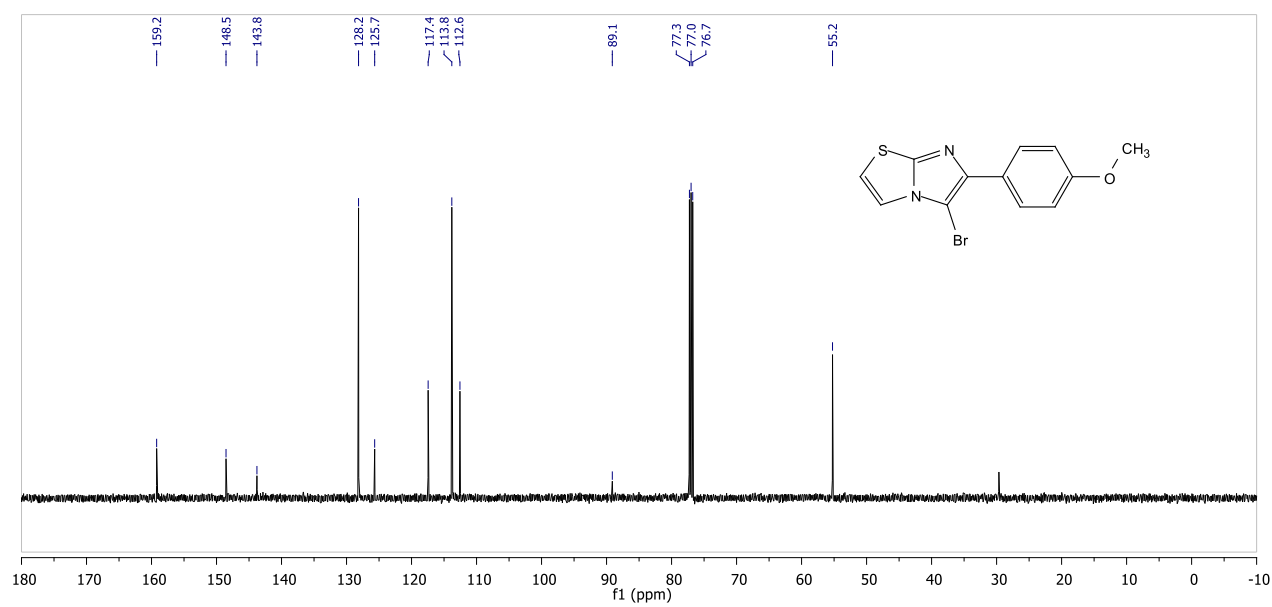
¹³C NMR (126 MHz, CDCl₃) of compound **2b**



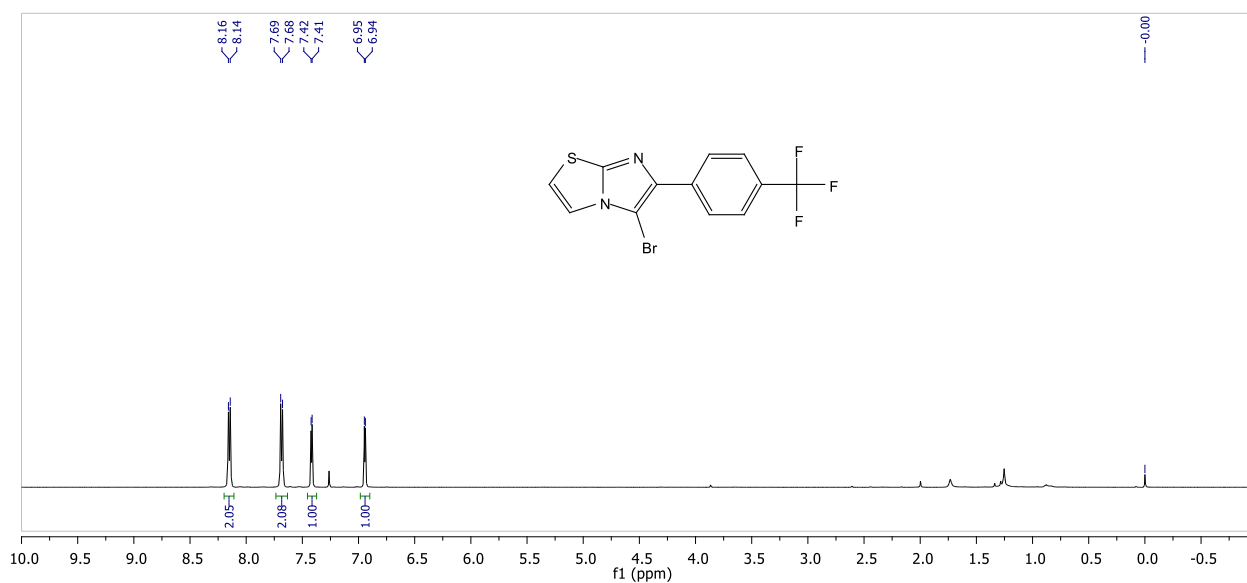
^1H NMR (400 MHz, CDCl_3) of compound **2c**



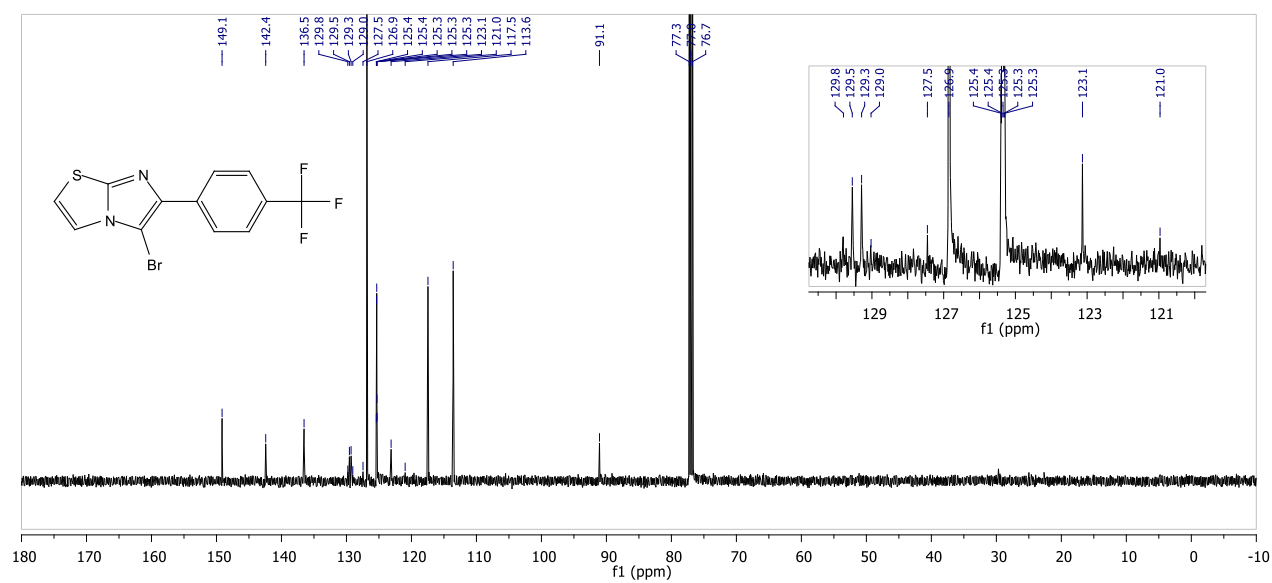
^{13}C NMR (126 MHz, CDCl_3) of compound **2c**



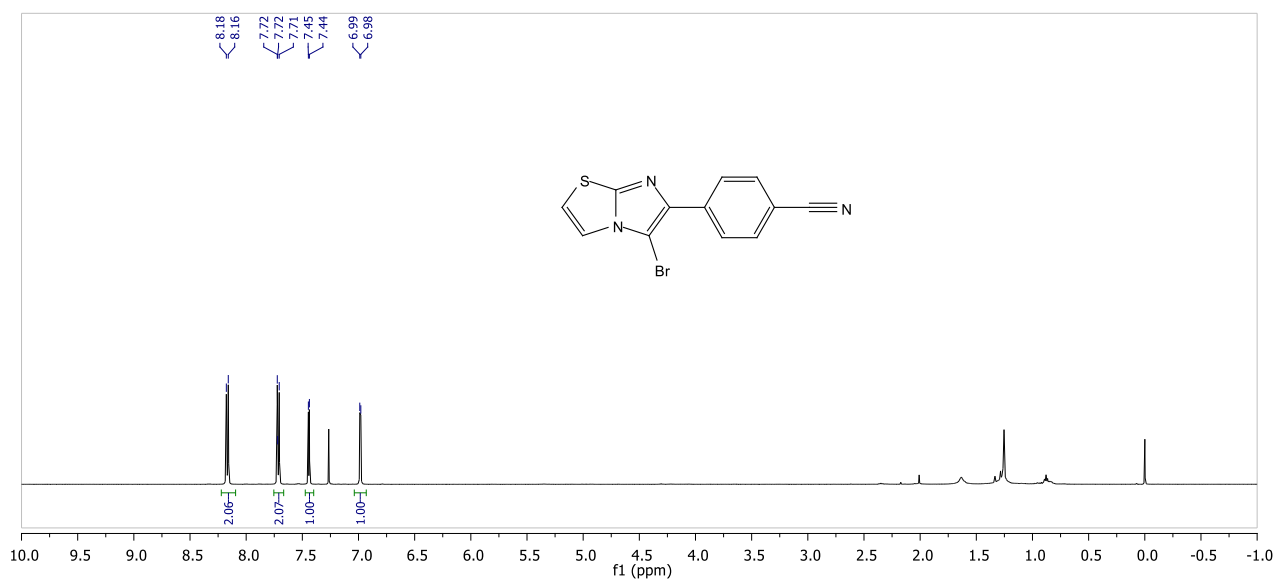
¹H NMR (500 MHz, CDCl₃) of compound **2d**



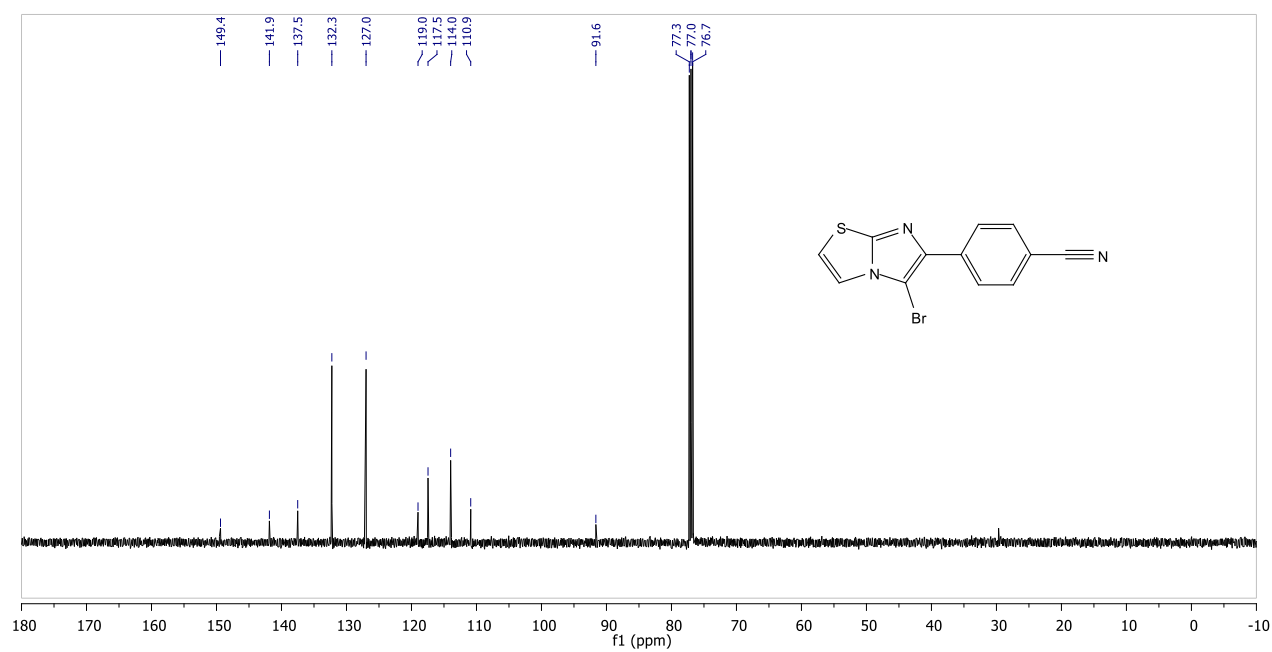
¹³C NMR (126 MHz, CDCl₃) of compound **2d**



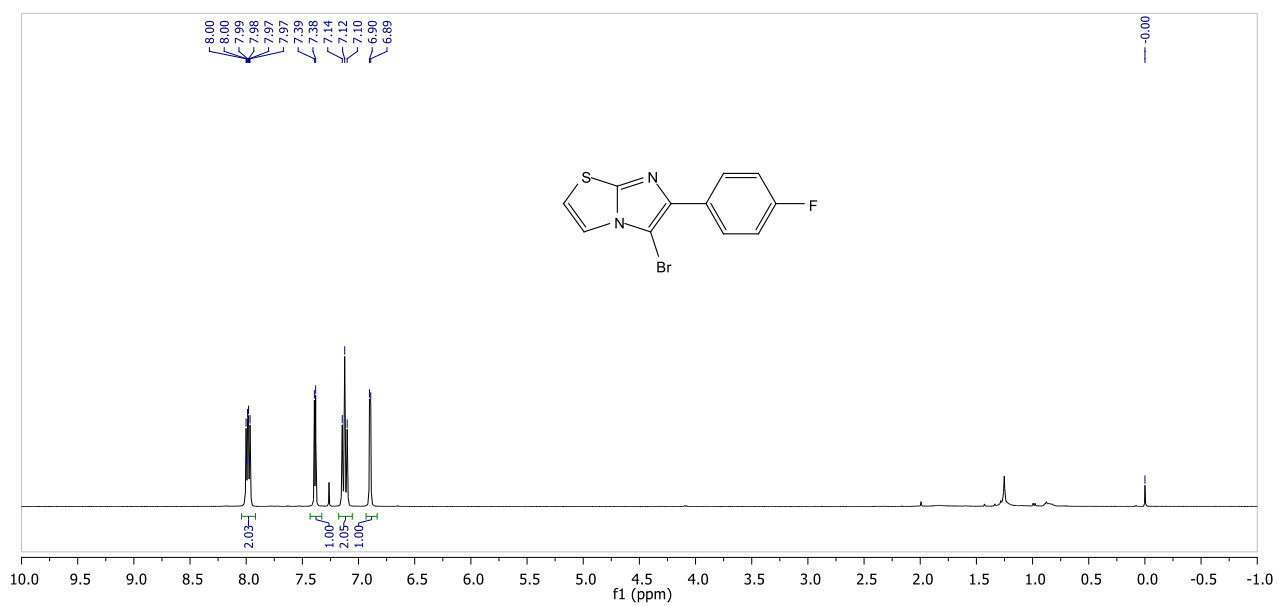
^1H NMR (500 MHz, CDCl_3) of compound **2e**



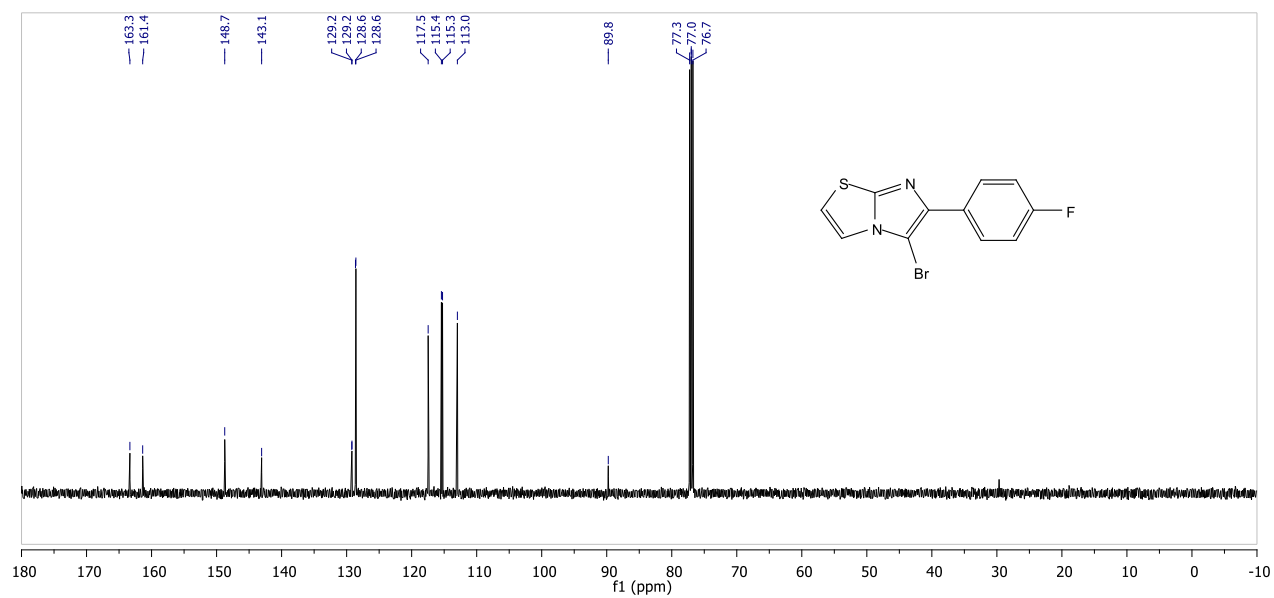
^{13}C NMR (126 MHz, CDCl_3) of compound **2e**



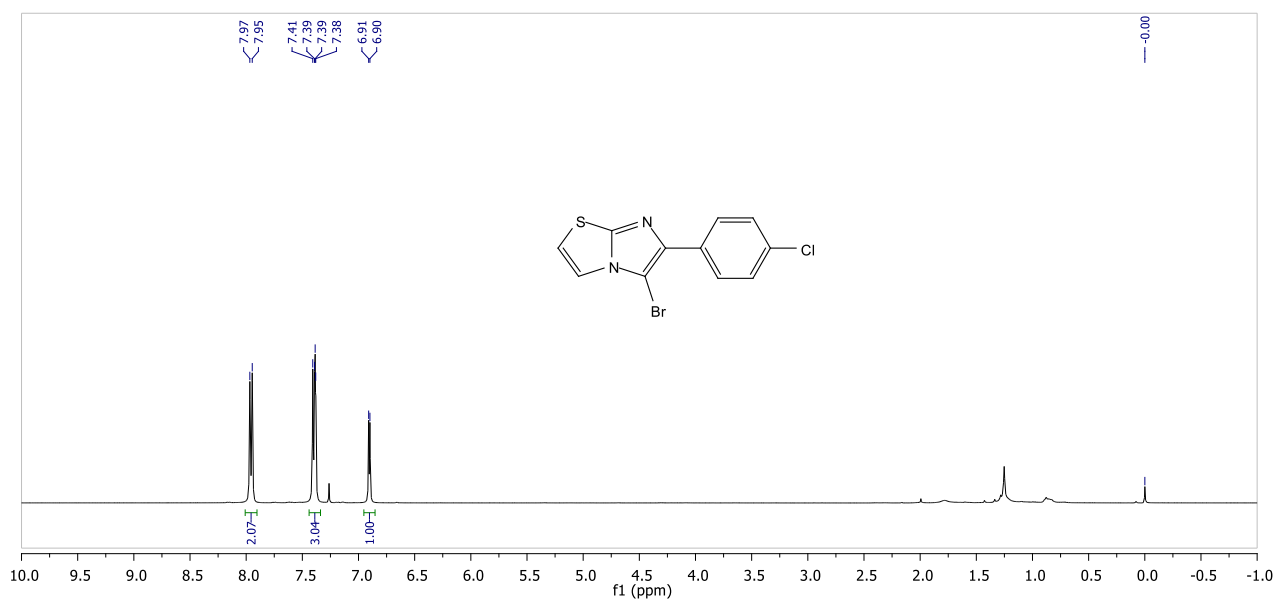
¹H NMR (400 MHz, CDCl₃) of compound **2f**



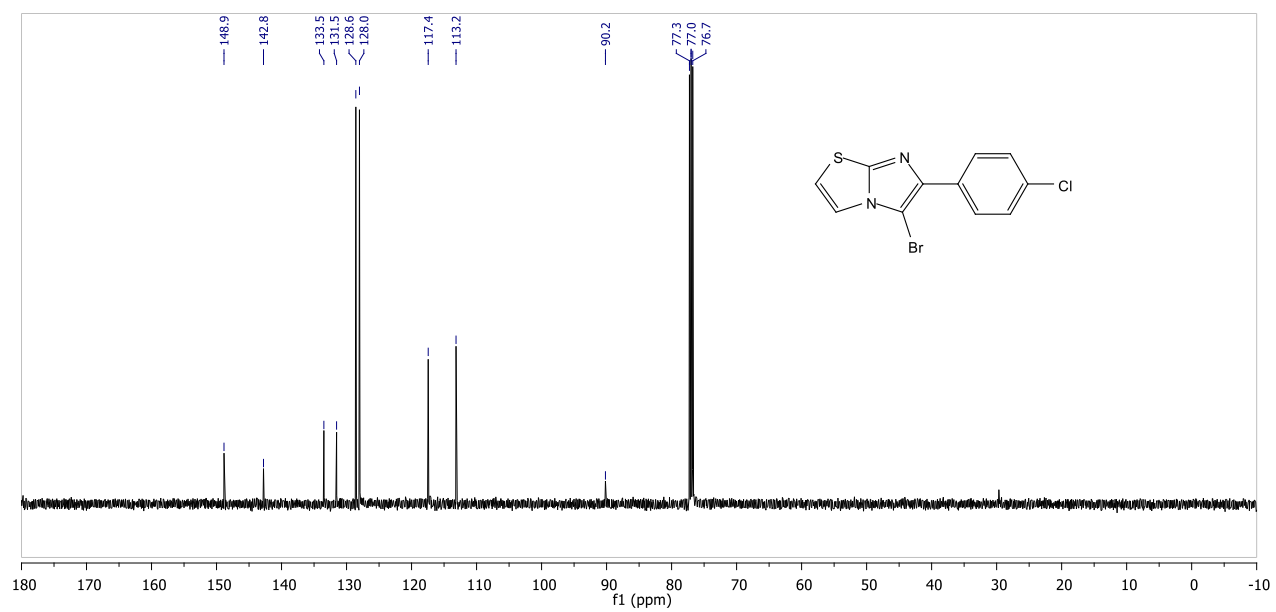
¹³C NMR (126 MHz, CDCl₃) of compound **2f**



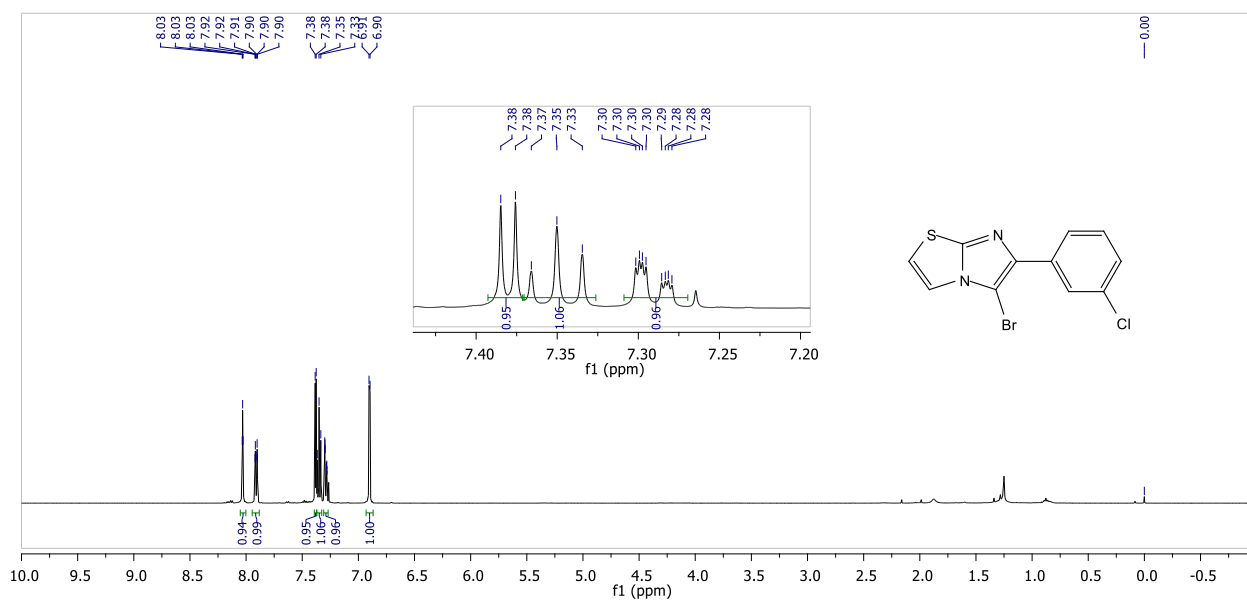
¹H NMR (400 MHz, CDCl₃) of compound **2g**



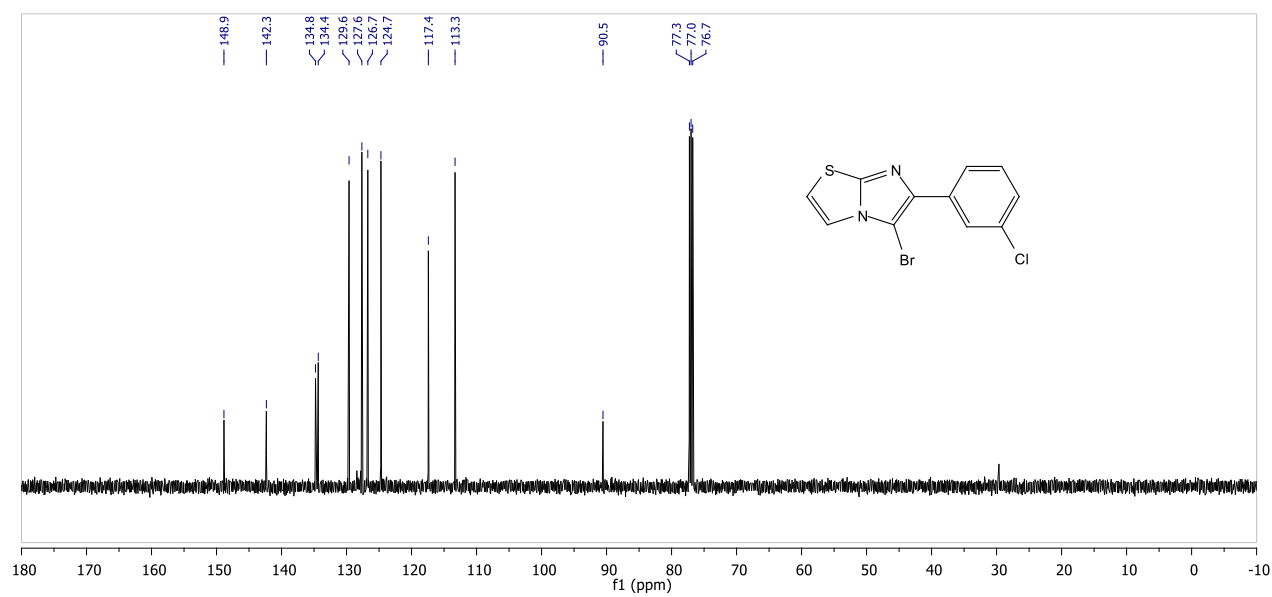
¹³C NMR (126 MHz, CDCl₃) of compound **2g**



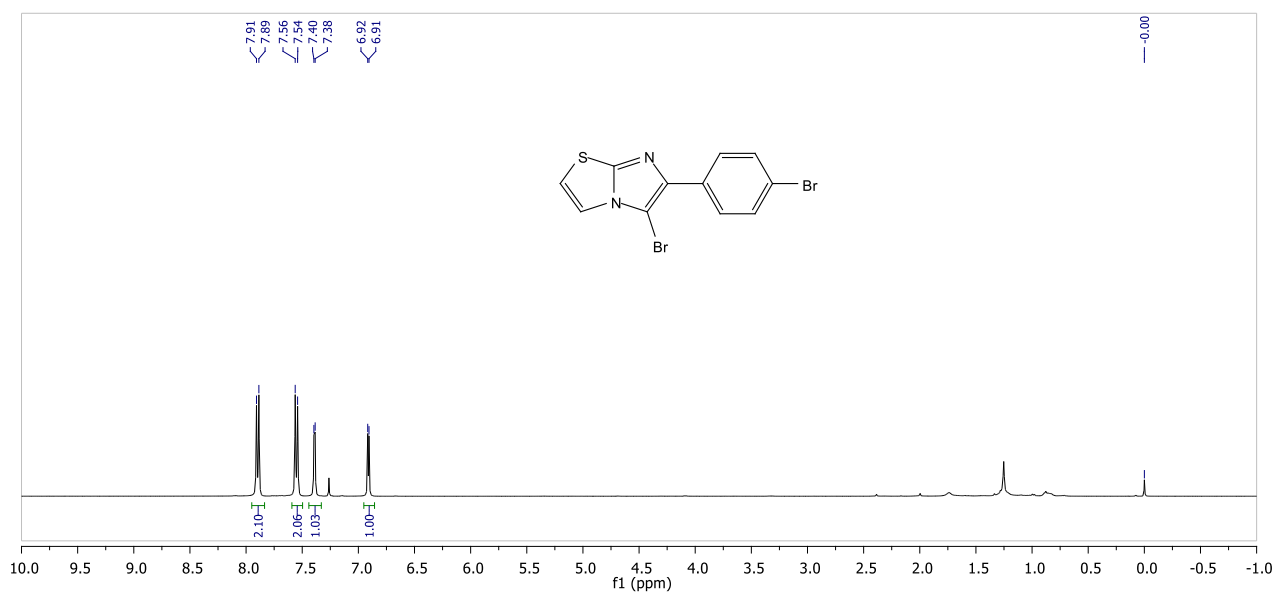
¹H NMR (500 MHz, CDCl₃) of compound **2h**



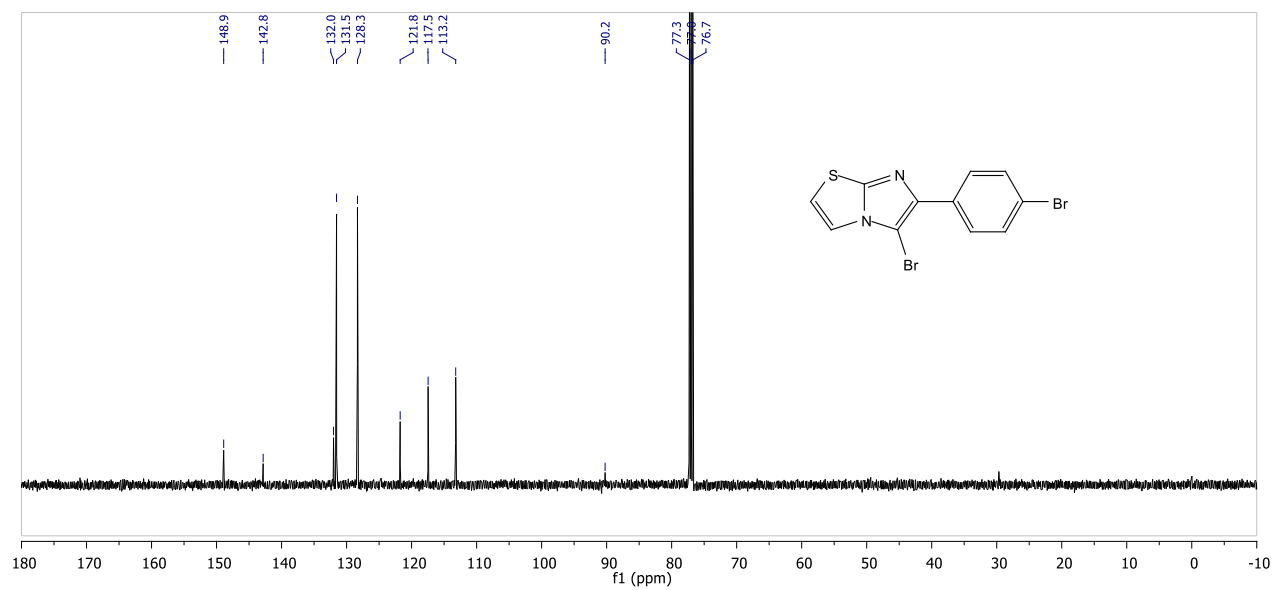
¹³C NMR (126 MHz, CDCl₃) of compound **2h**



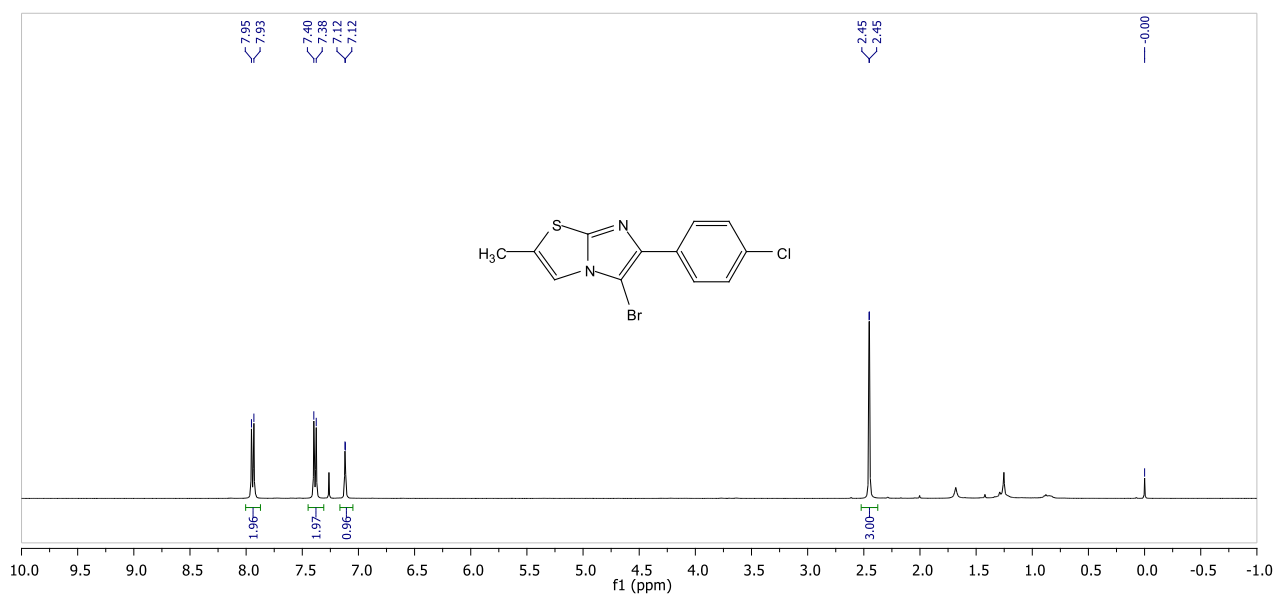
¹H NMR (400 MHz, CDCl₃) of compound **2i**



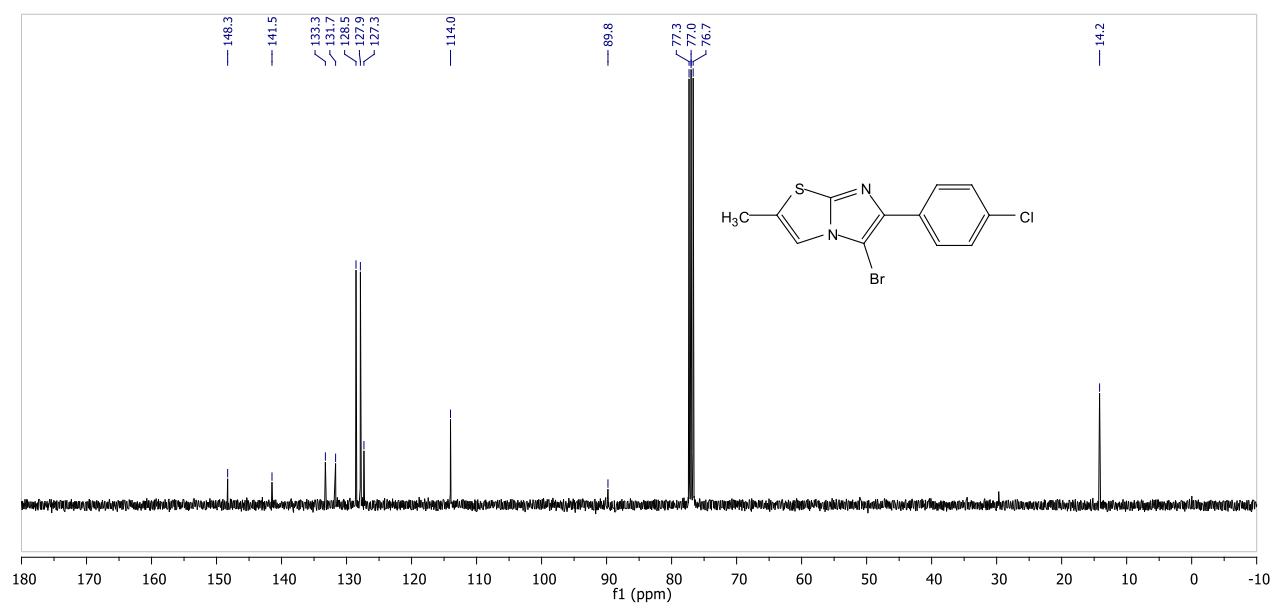
¹³C NMR (126 MHz, CDCl₃) of compound **2i**



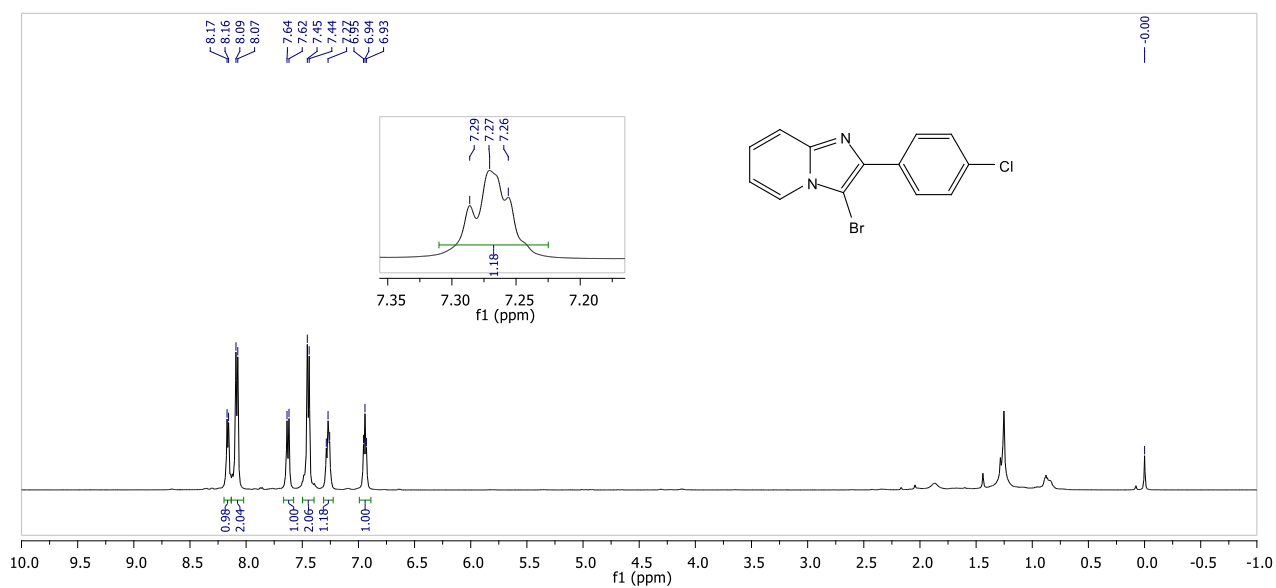
¹H NMR (400 MHz, CDCl₃) of compound **2j**



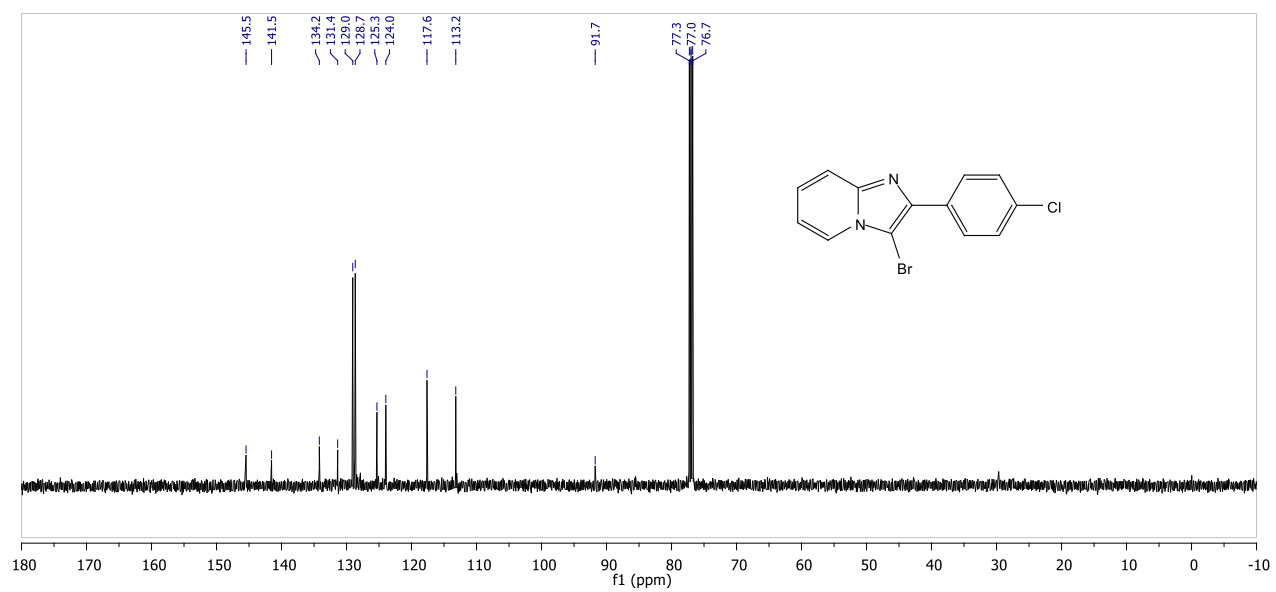
¹³C NMR (101 MHz, CDCl₃) of compound **2j**



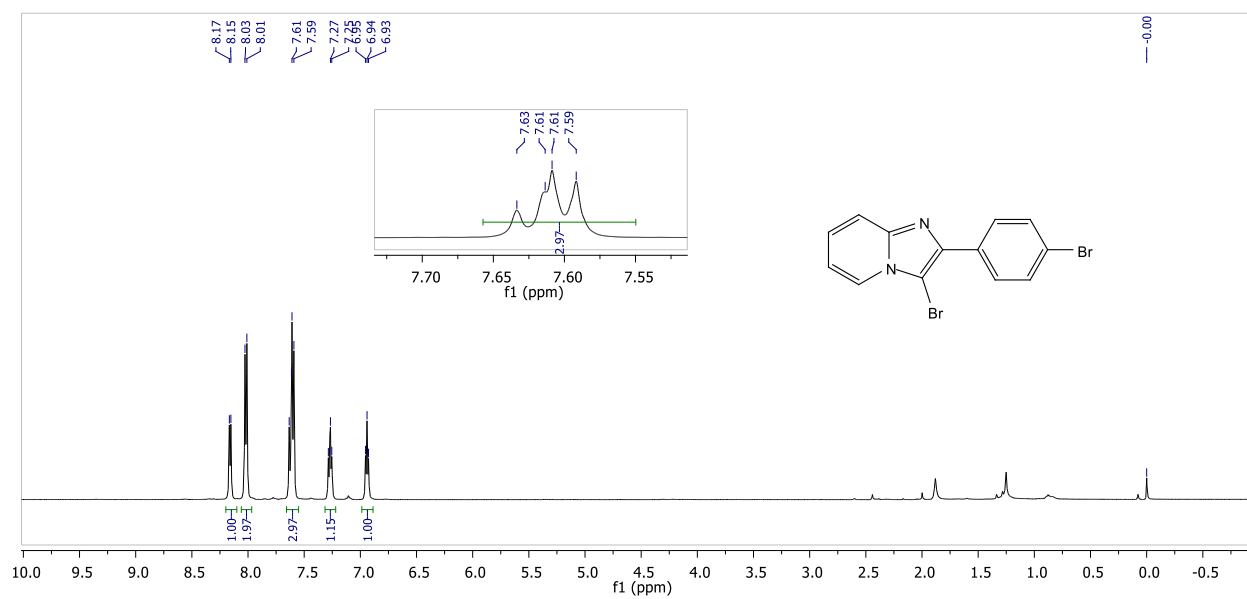
¹H NMR (500 MHz, CDCl₃) of compound **2k**



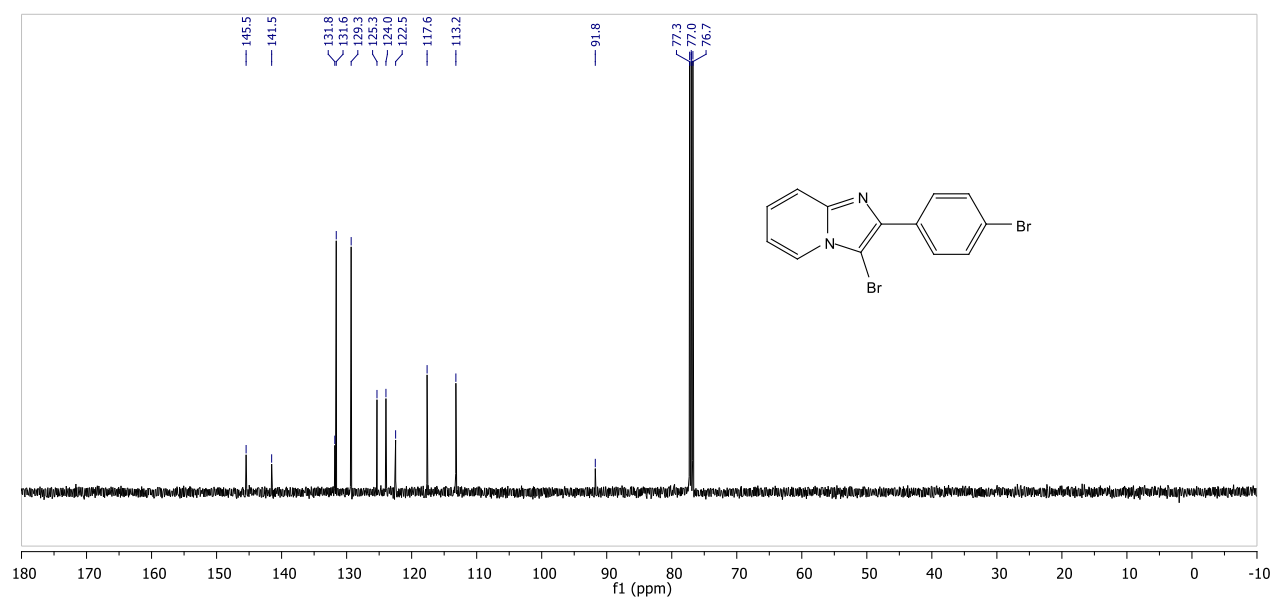
¹³C NMR (126 MHz, CDCl₃) of compound **2k**



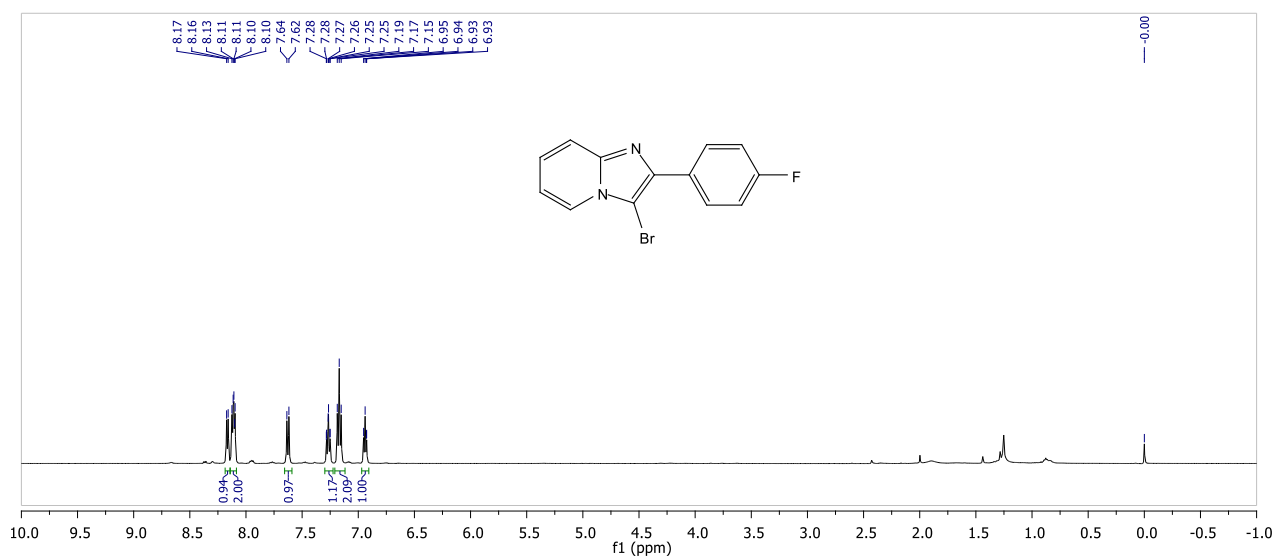
¹H NMR (500 MHz, CDCl₃) of compound **21**



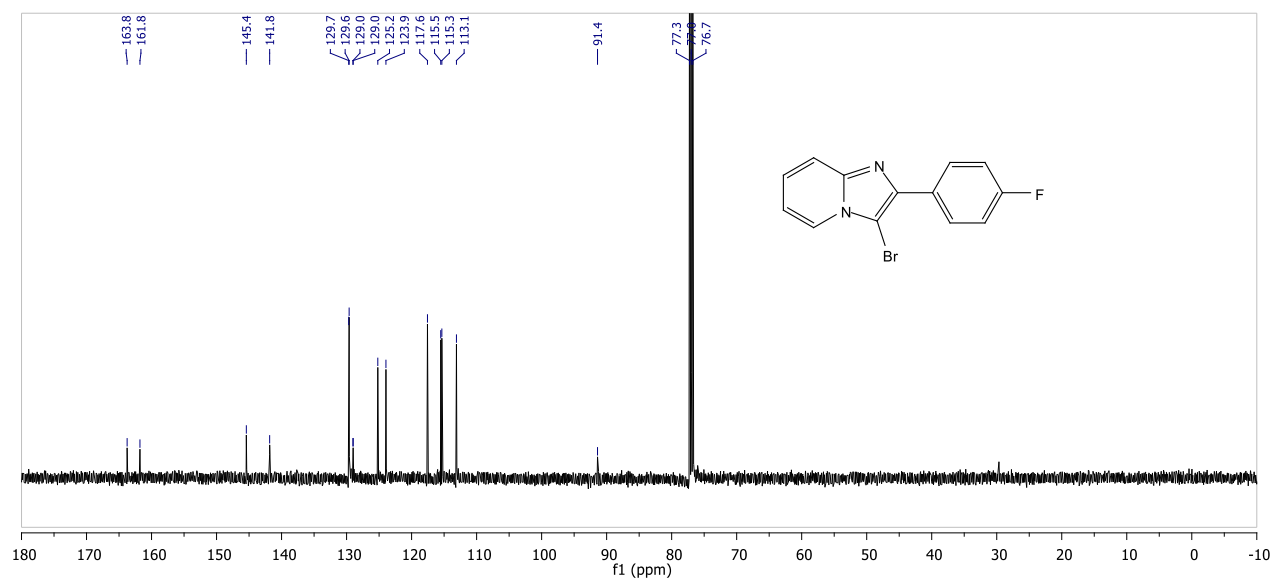
¹³C NMR (126 MHz, CDCl₃) of compound **21**



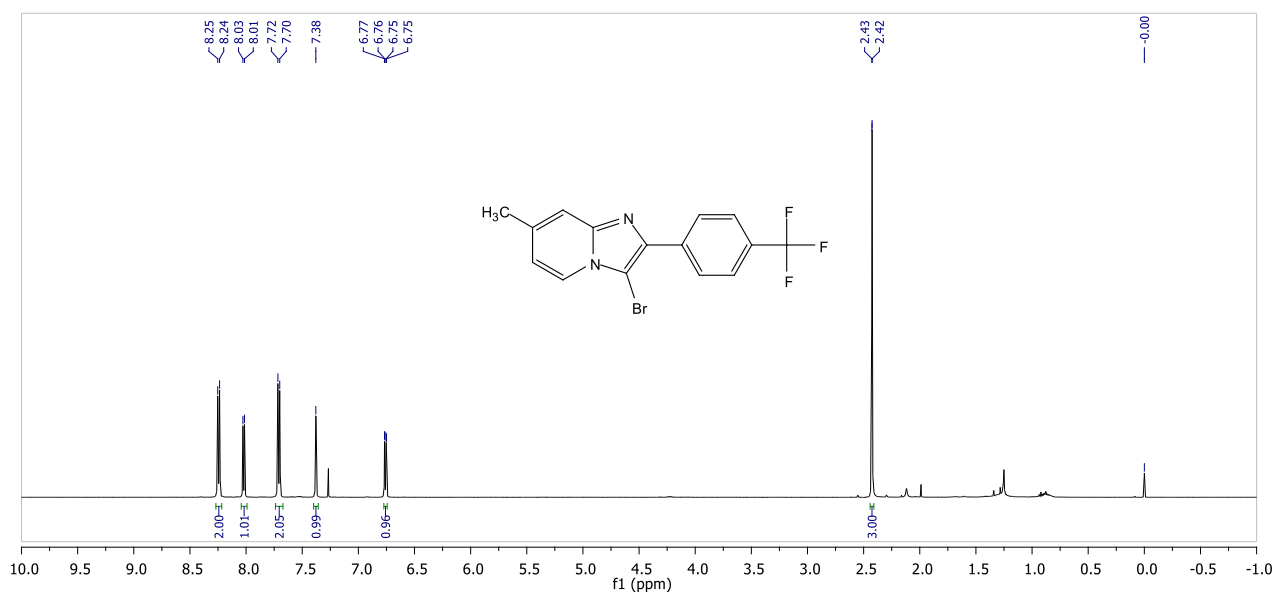
¹H NMR (500 MHz, CDCl₃) of compound **2m**



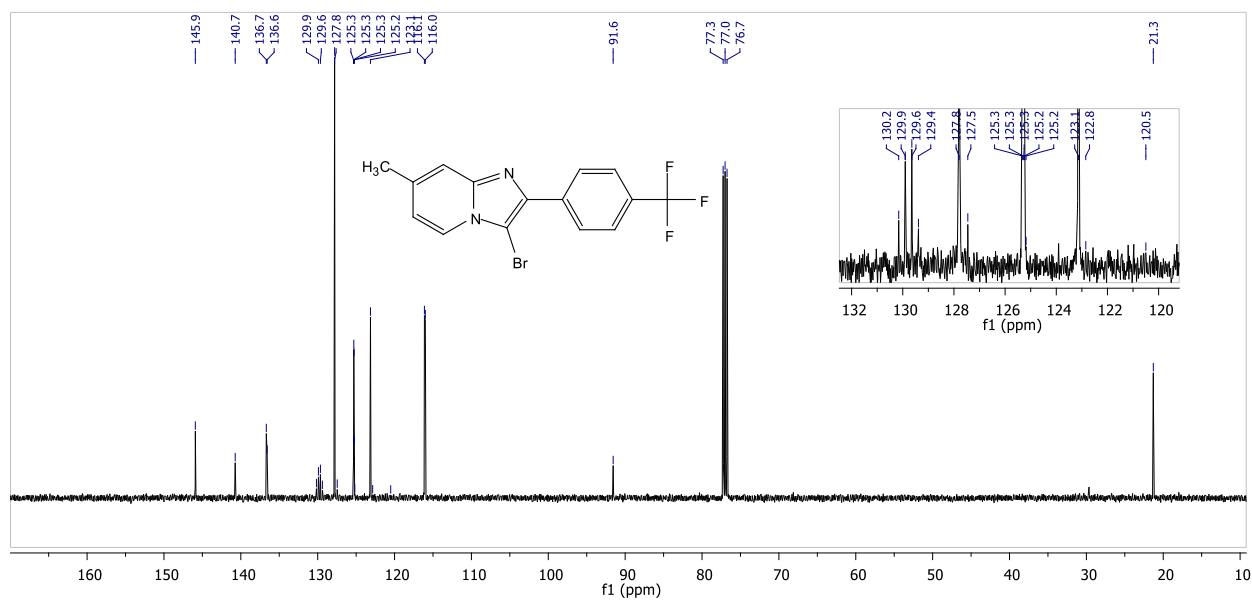
¹³C NMR (126 MHz, CDCl₃) of compound **2m**



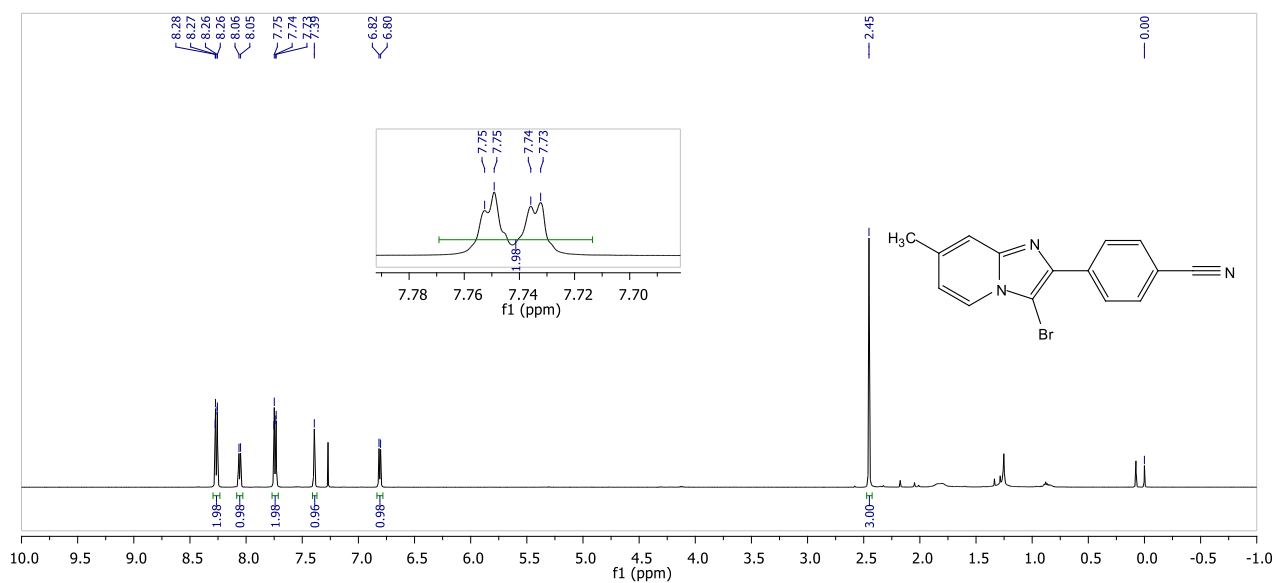
^1H NMR (500 MHz, CDCl_3) of compound **2n**



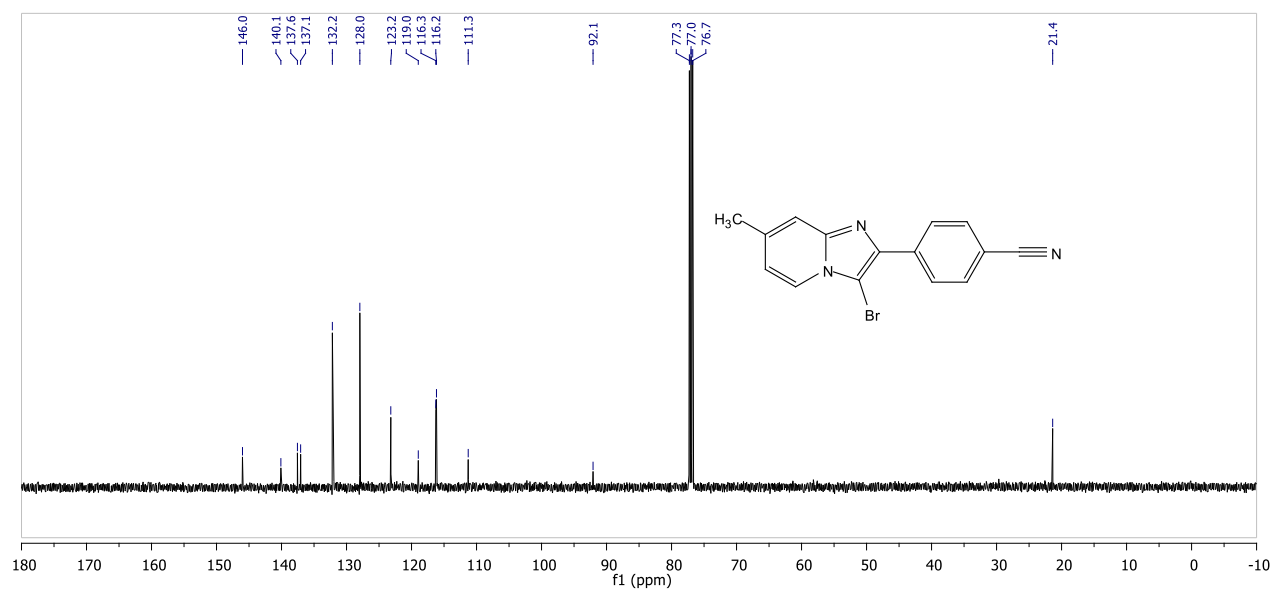
^{13}C NMR (126 MHz, CDCl_3) of compound **2n**



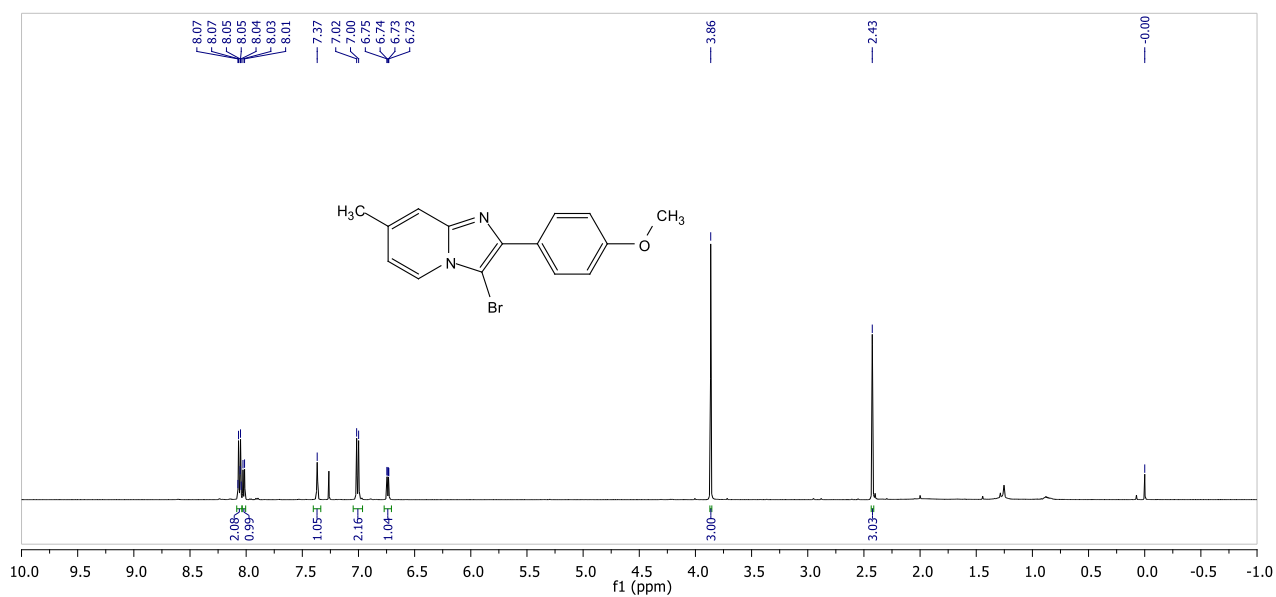
^1H NMR (500 MHz, CDCl_3) of compound **2o**



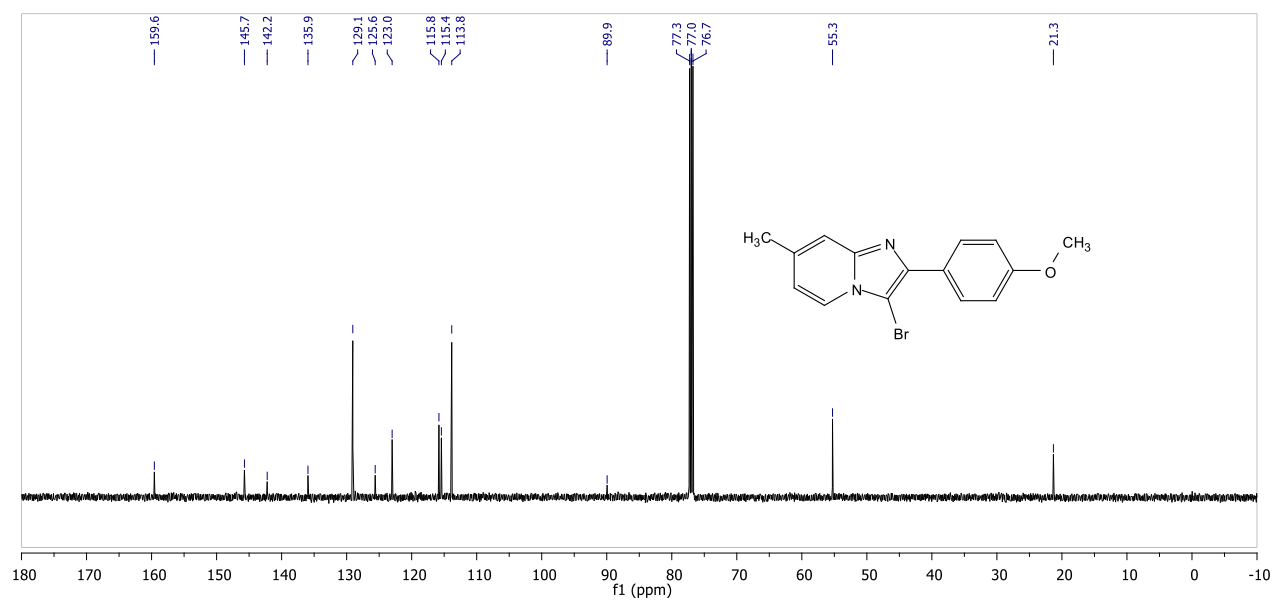
^{13}C NMR (126 MHz, CDCl_3) of compound **2o**



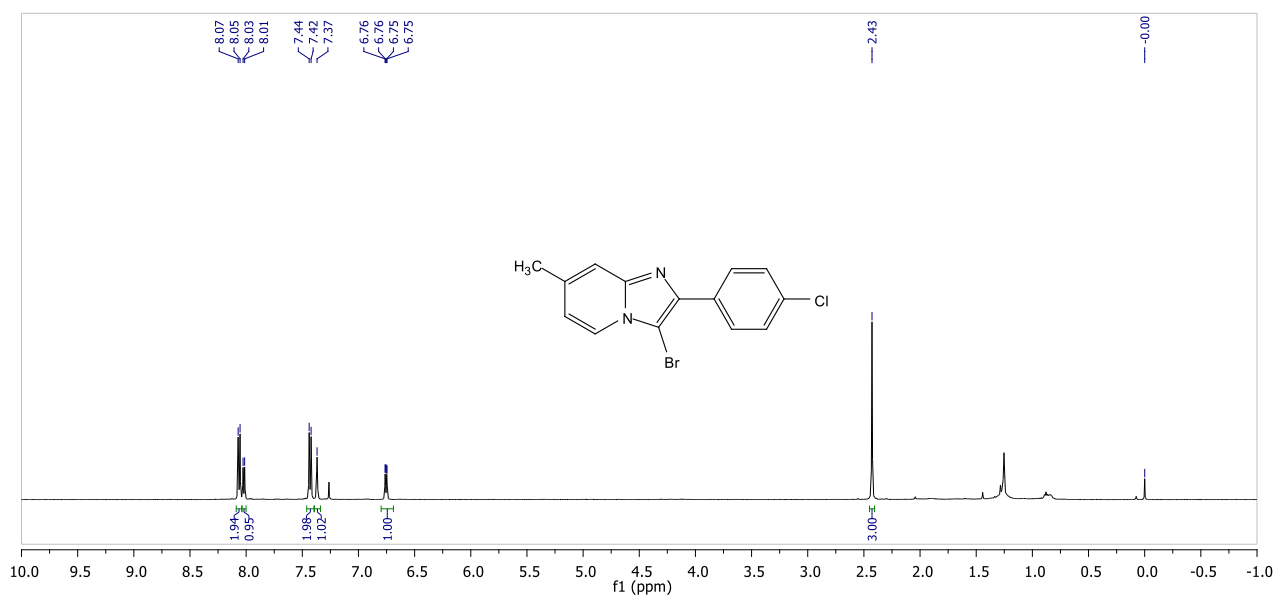
¹H NMR (500 MHz, CDCl₃) of compound **2p**



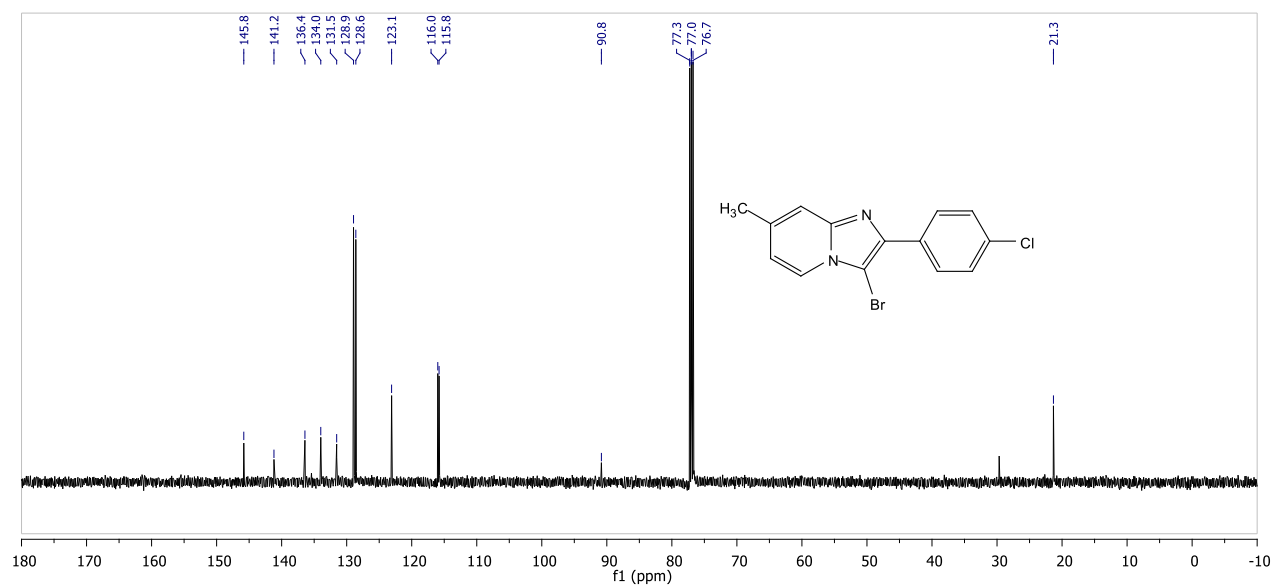
¹³C NMR (126 MHz, CDCl₃) of compound **2p**



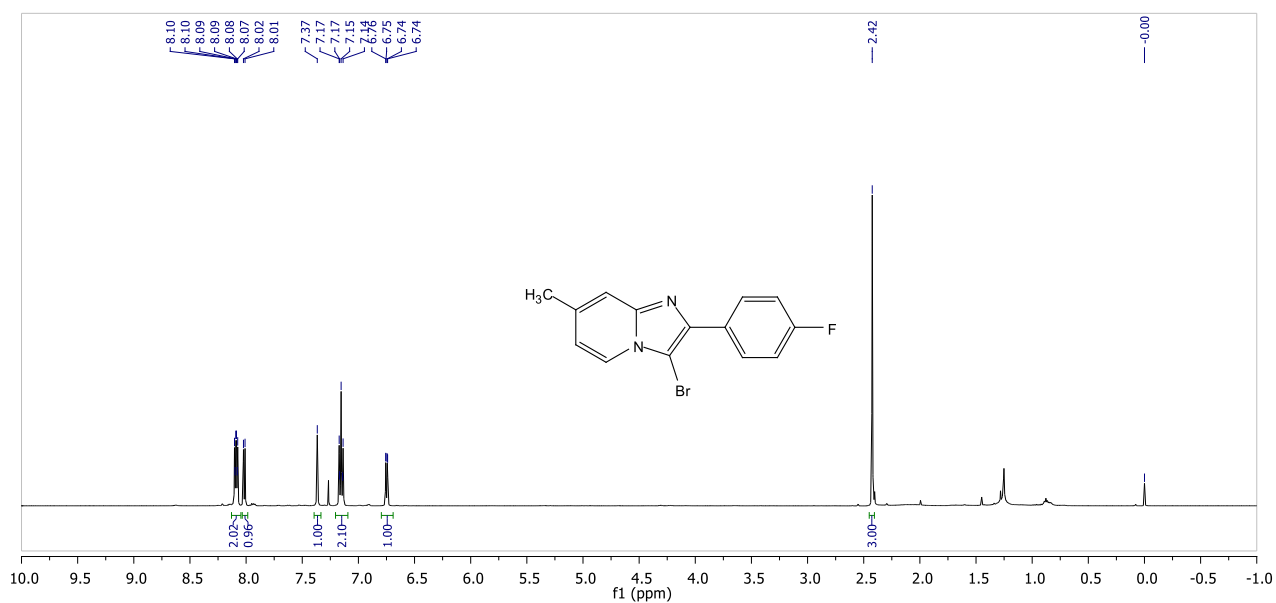
^1H NMR (500 MHz, CDCl_3) of compound **2q**



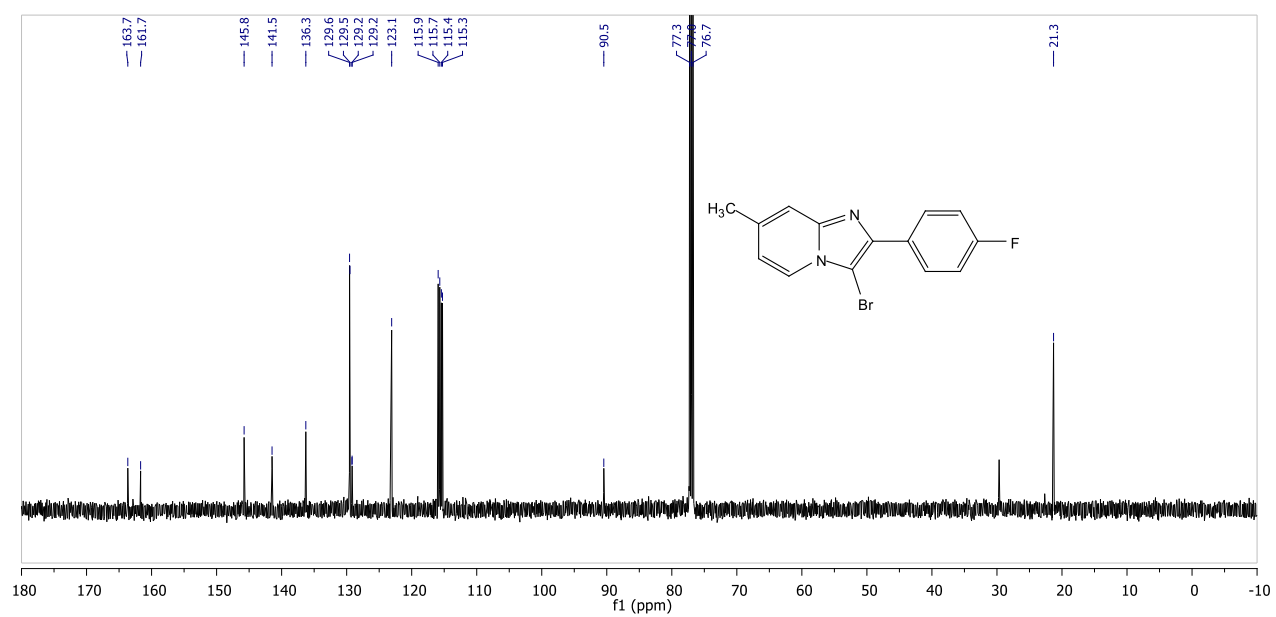
^{13}C NMR (126 MHz, CDCl_3) of compound **2q**



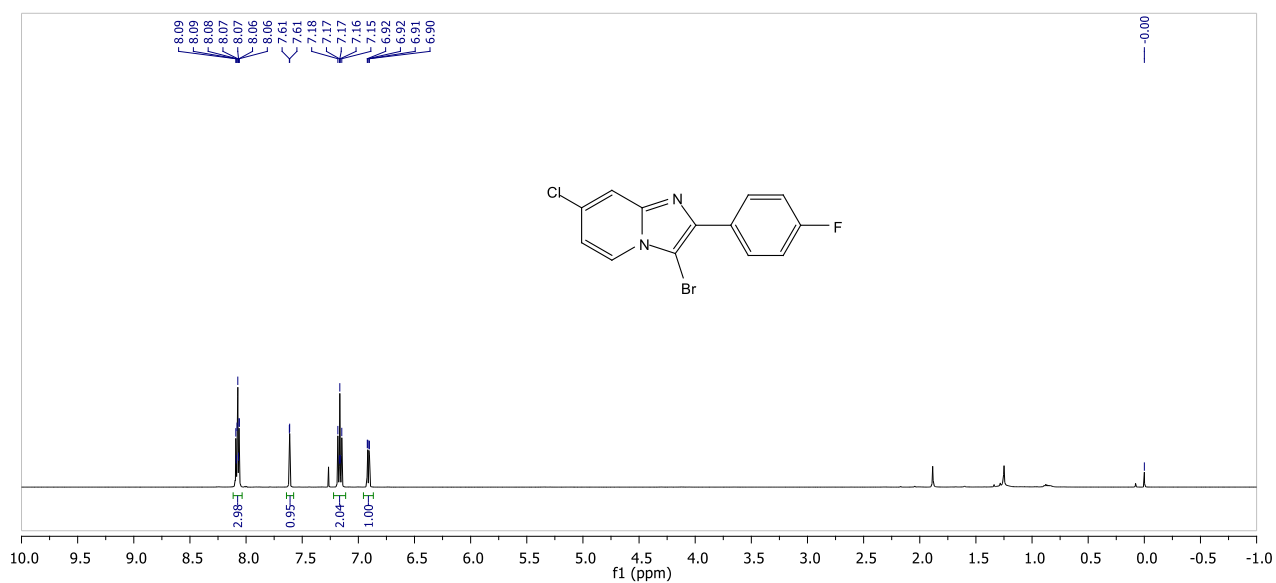
¹H NMR (500 MHz, CDCl₃) of compound **2r**



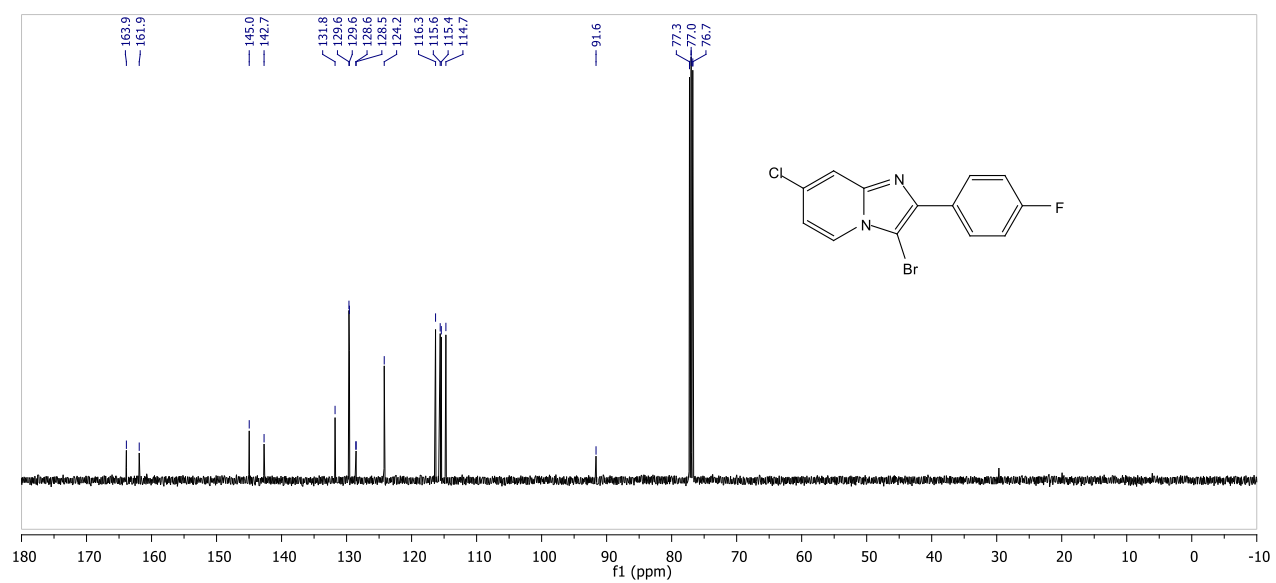
¹³C NMR (126 MHz, CDCl₃) of compound **2r**



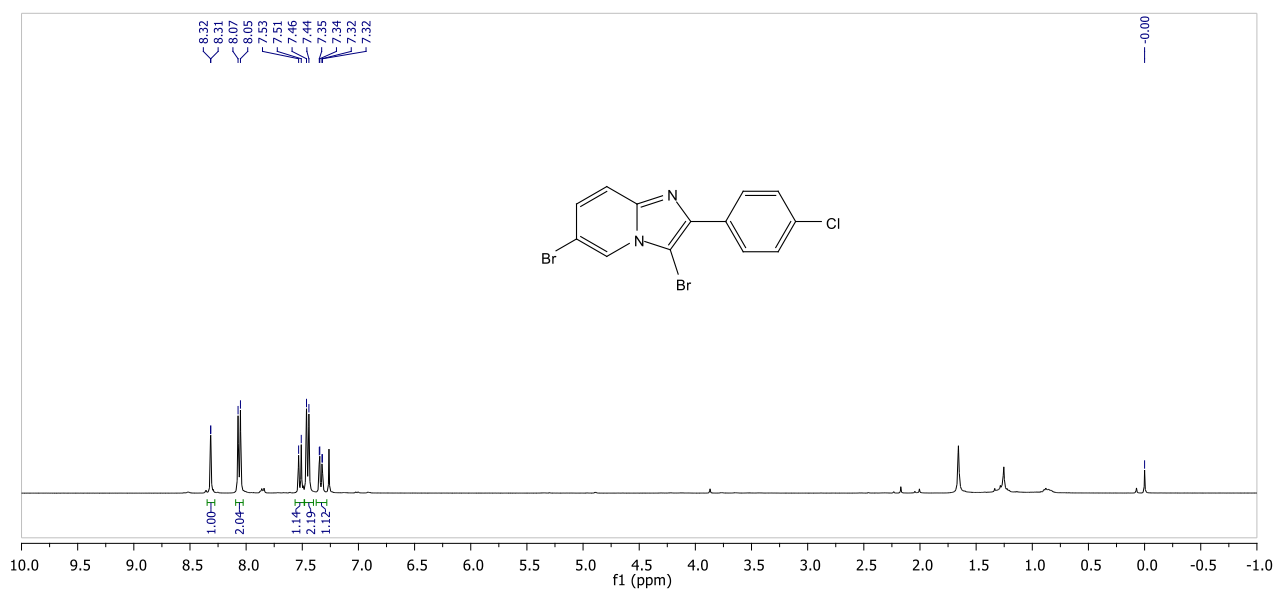
¹H NMR (500 MHz, CDCl₃) of compound **2s**



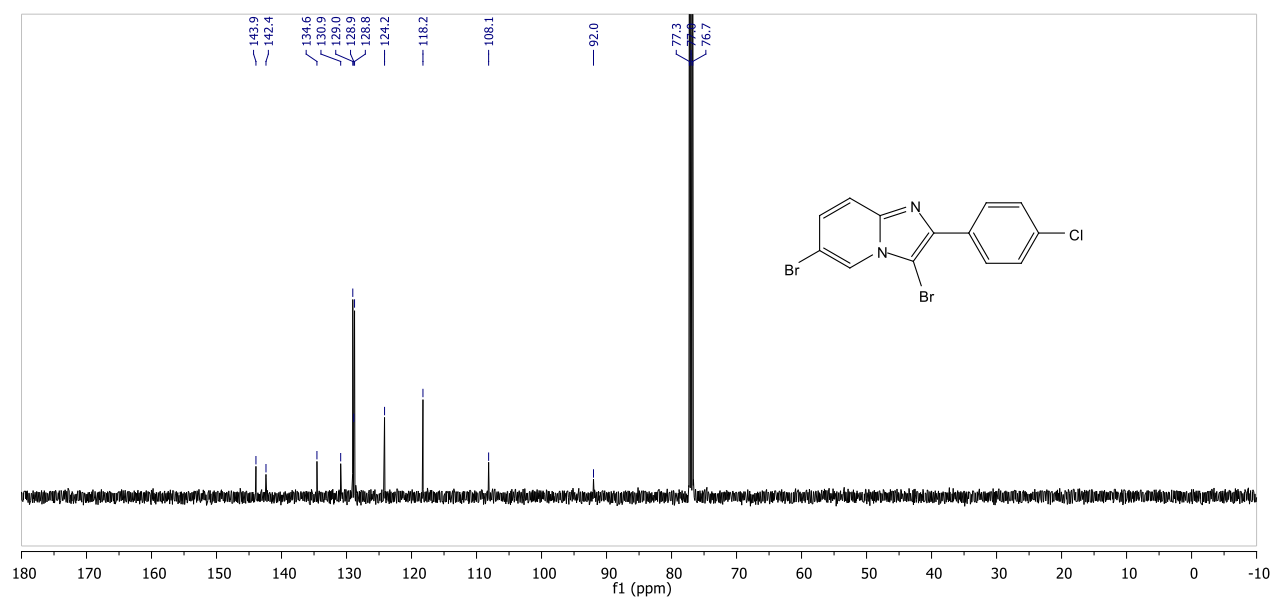
¹³C NMR (126 MHz, CDCl₃) of compound **2s**



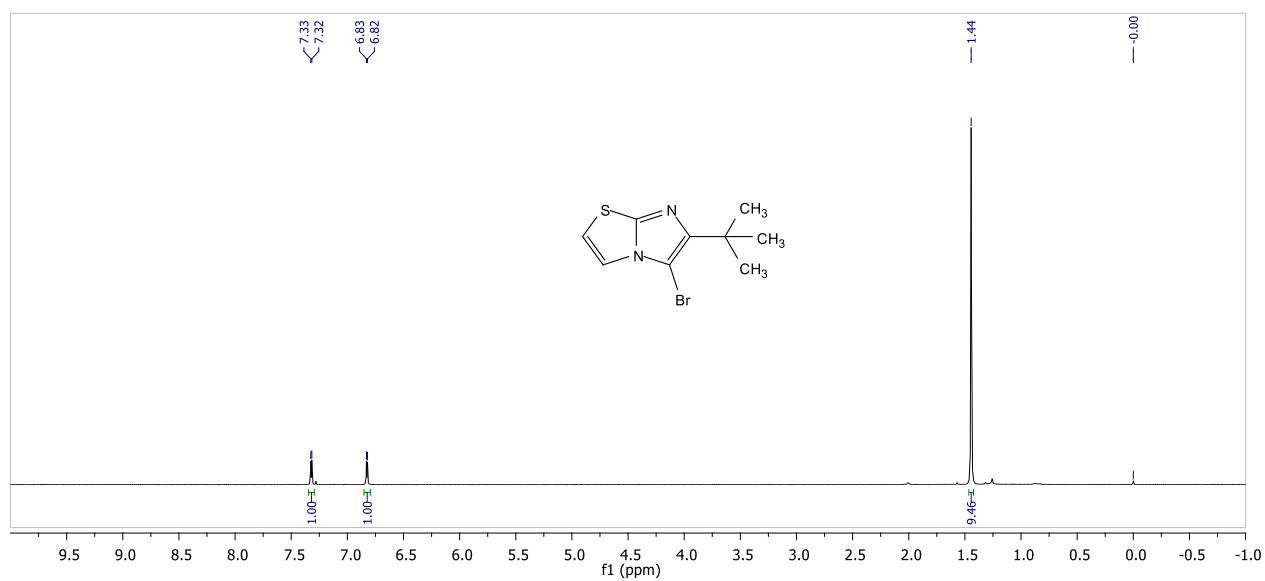
¹H NMR (500 MHz, CDCl₃) of compound **2t**



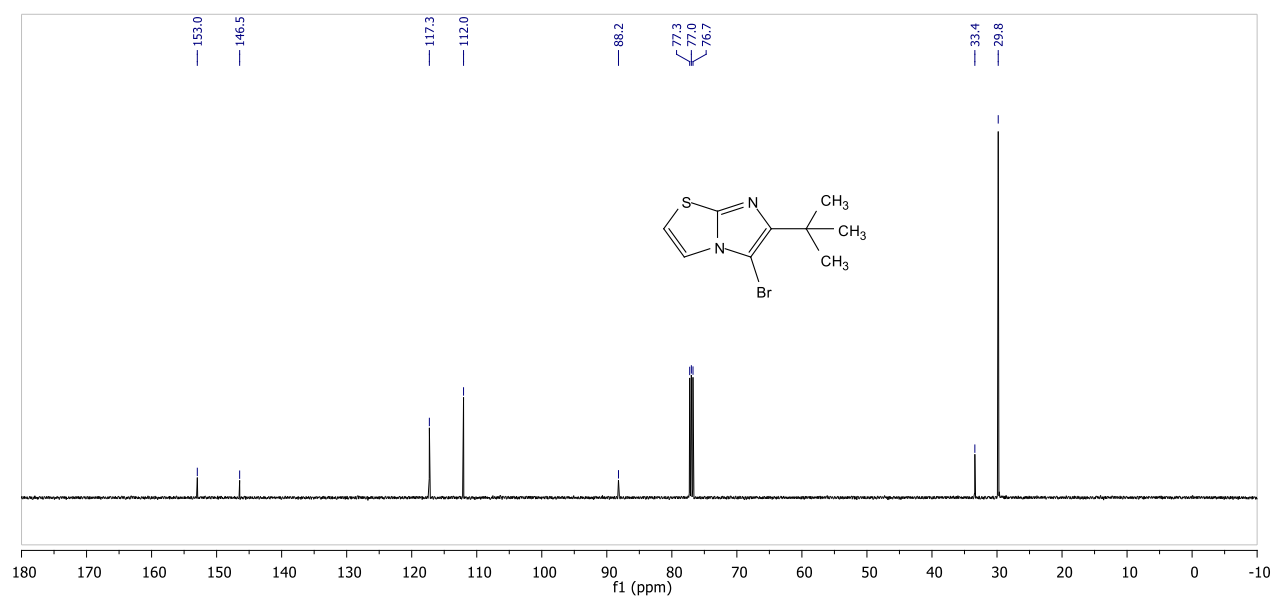
¹³C NMR (126 MHz, CDCl₃) of compound **2t**



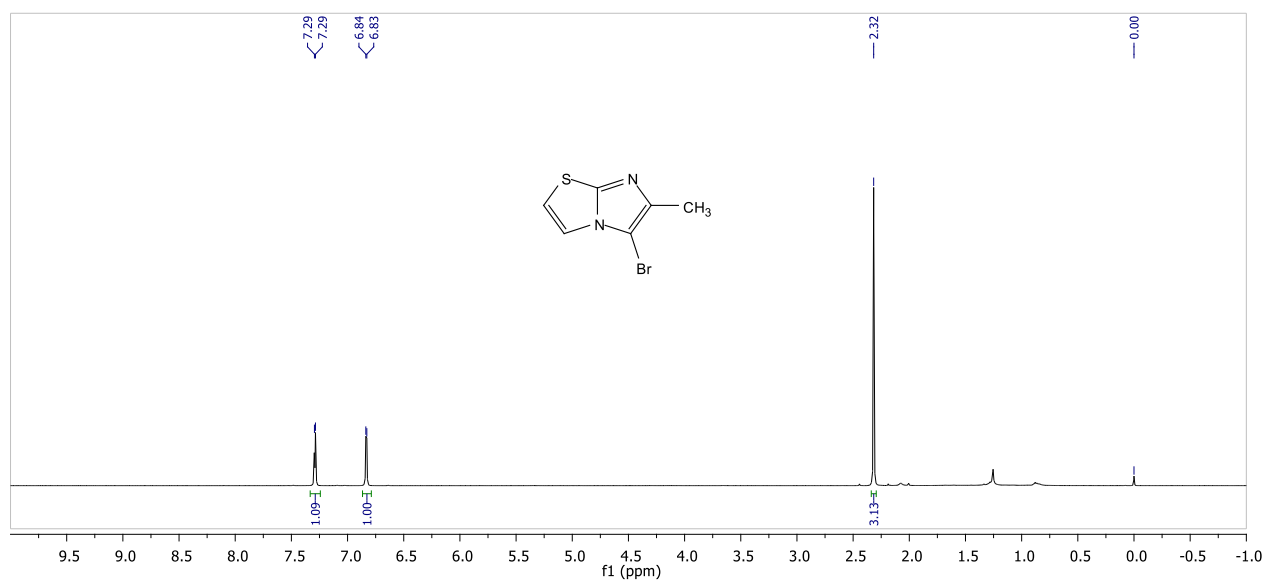
¹H NMR (500 MHz, CDCl₃) of compound **2u**



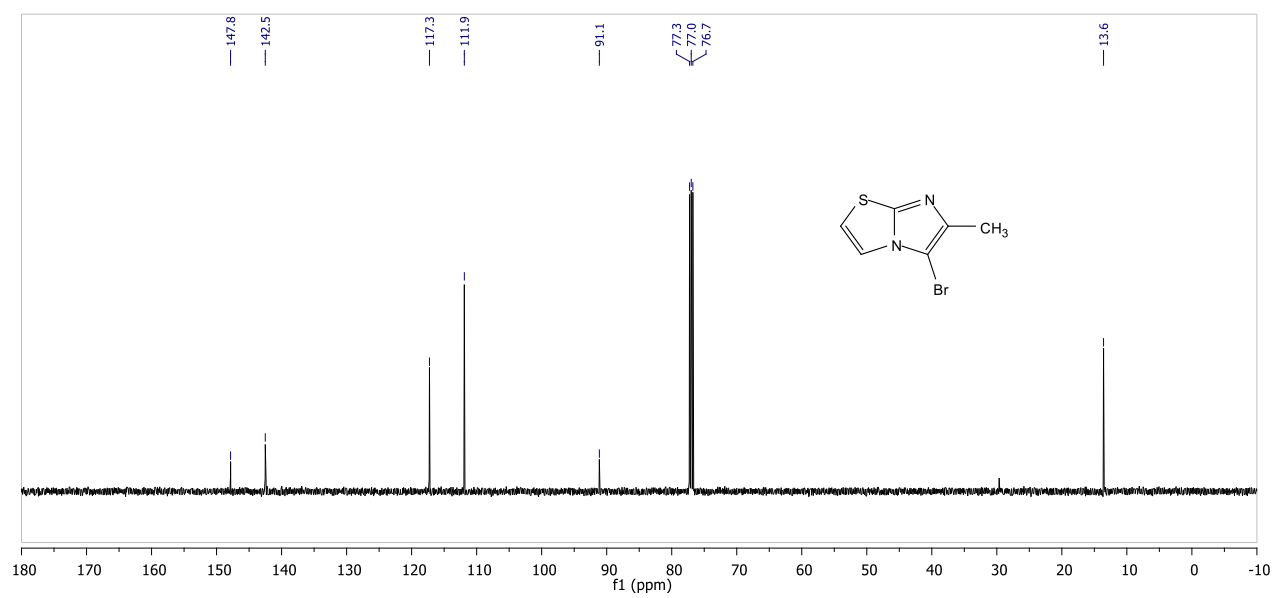
¹³C NMR (126 MHz, CDCl₃) of compound **2u**



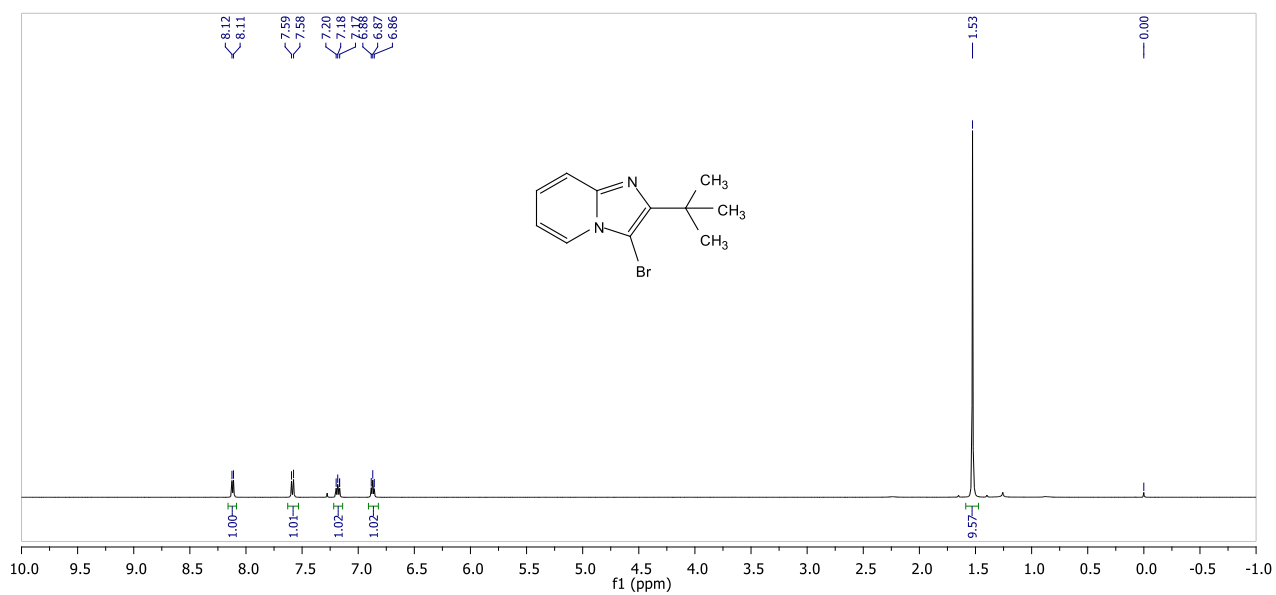
^1H NMR (500 MHz, CDCl_3) of compound **2v**



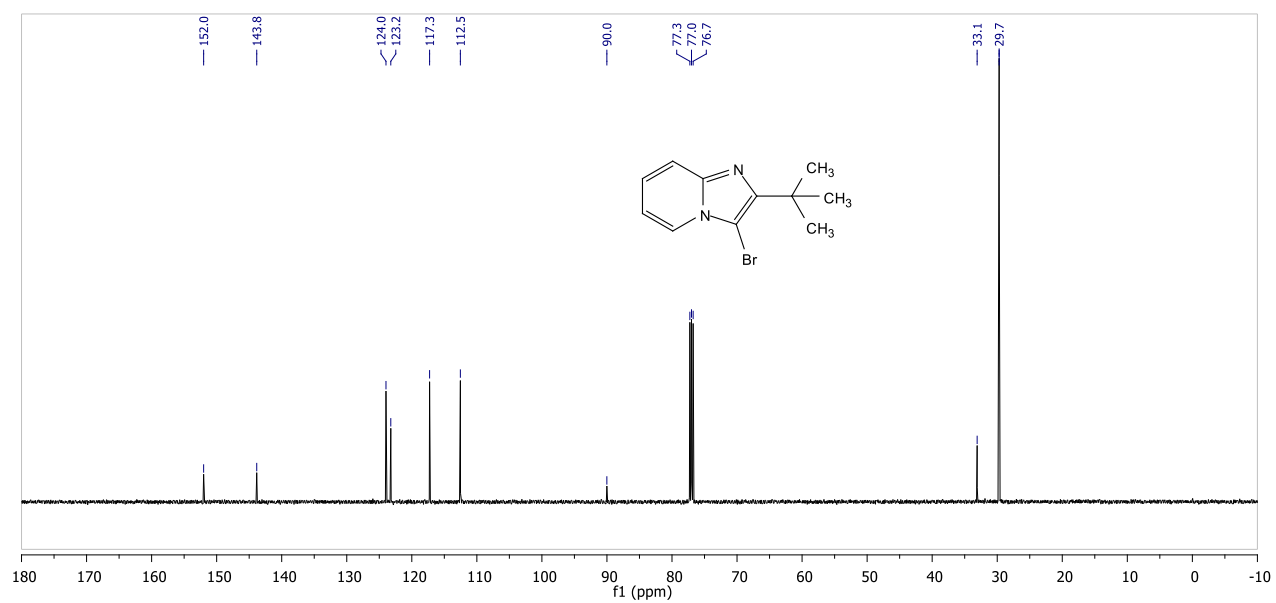
^{13}C NMR (126 MHz, CDCl_3) of compound **2v**



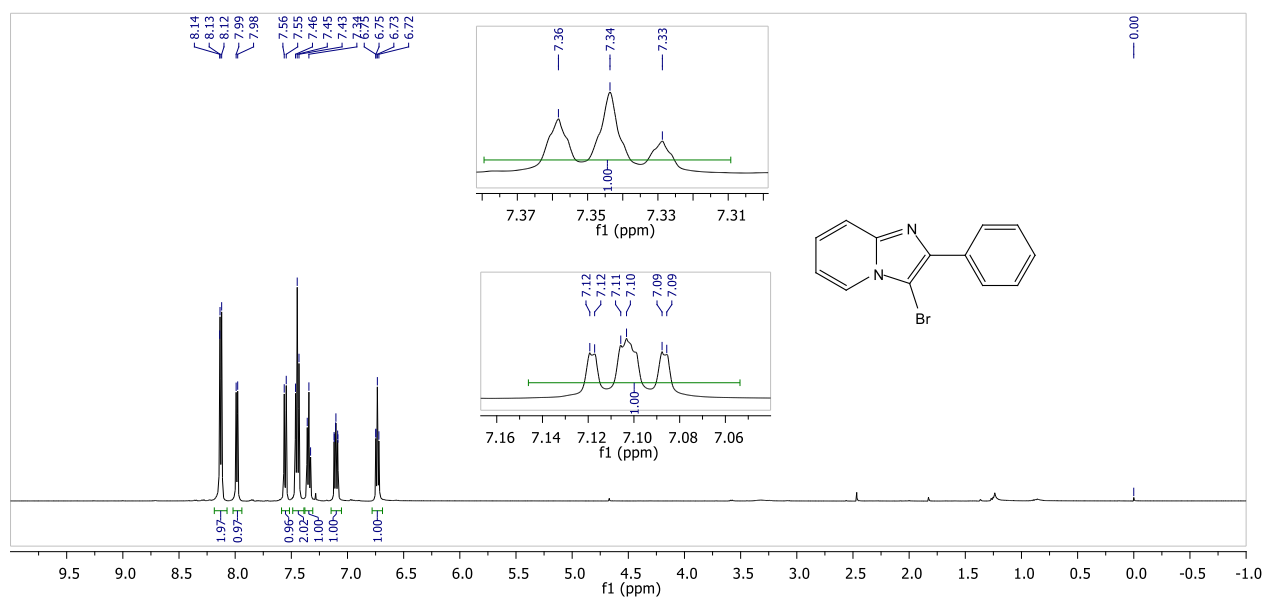
^1H NMR (500 MHz, CDCl_3) of compound **2w**



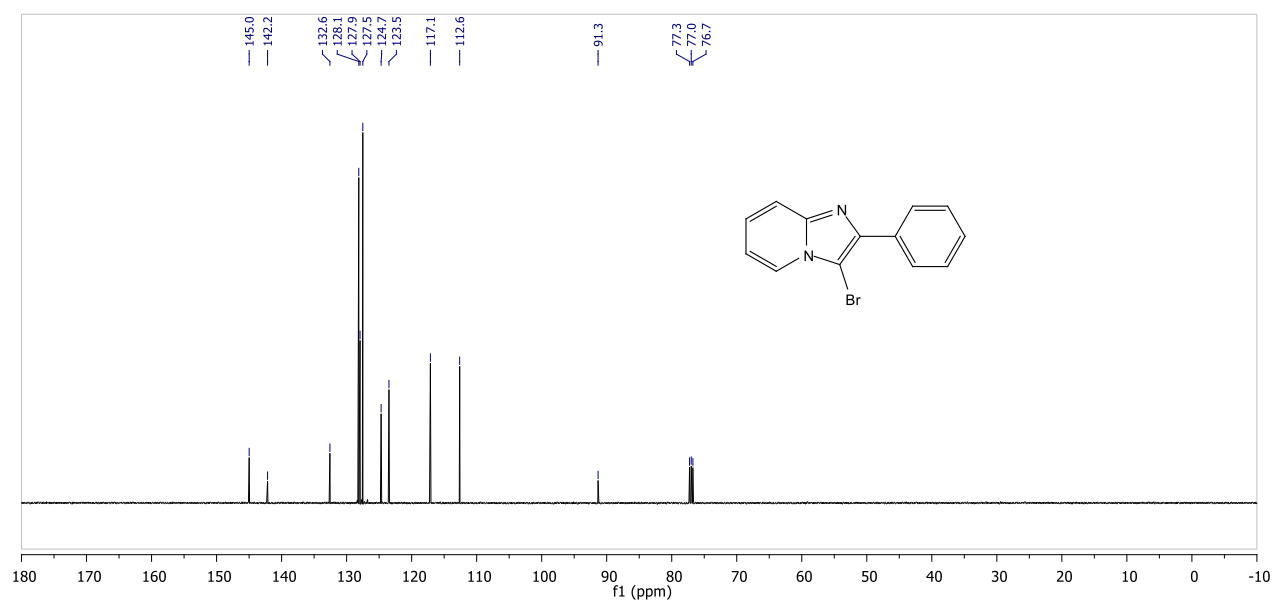
^{13}C NMR (126 MHz, CDCl_3) of compound **2w**



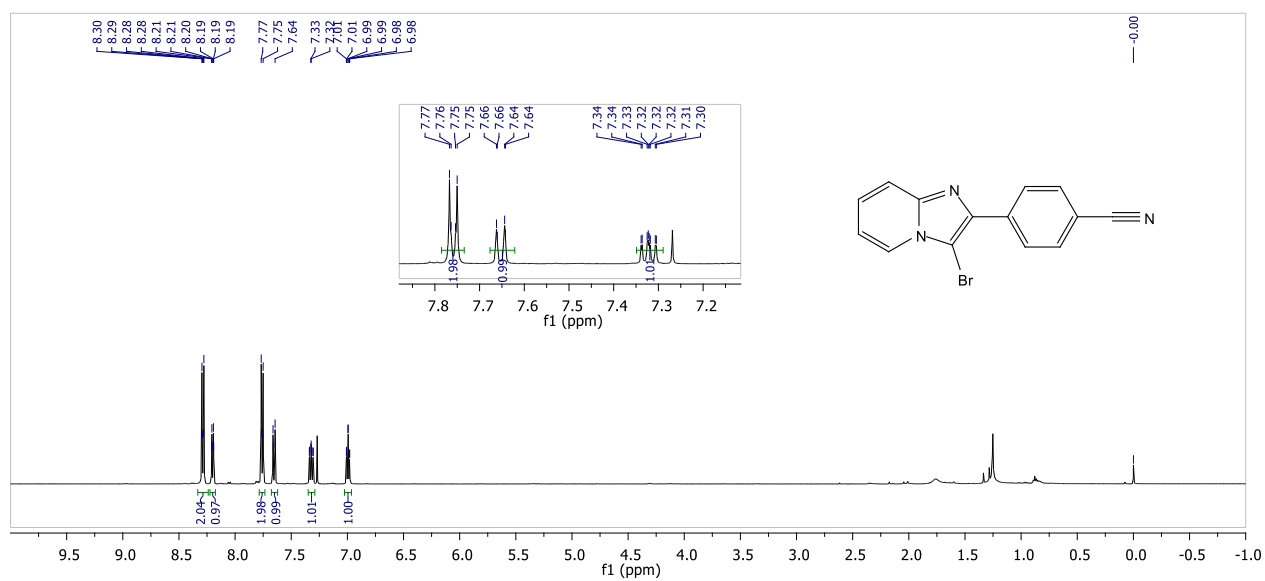
¹H NMR (500 MHz, CDCl₃) of compound **2x**



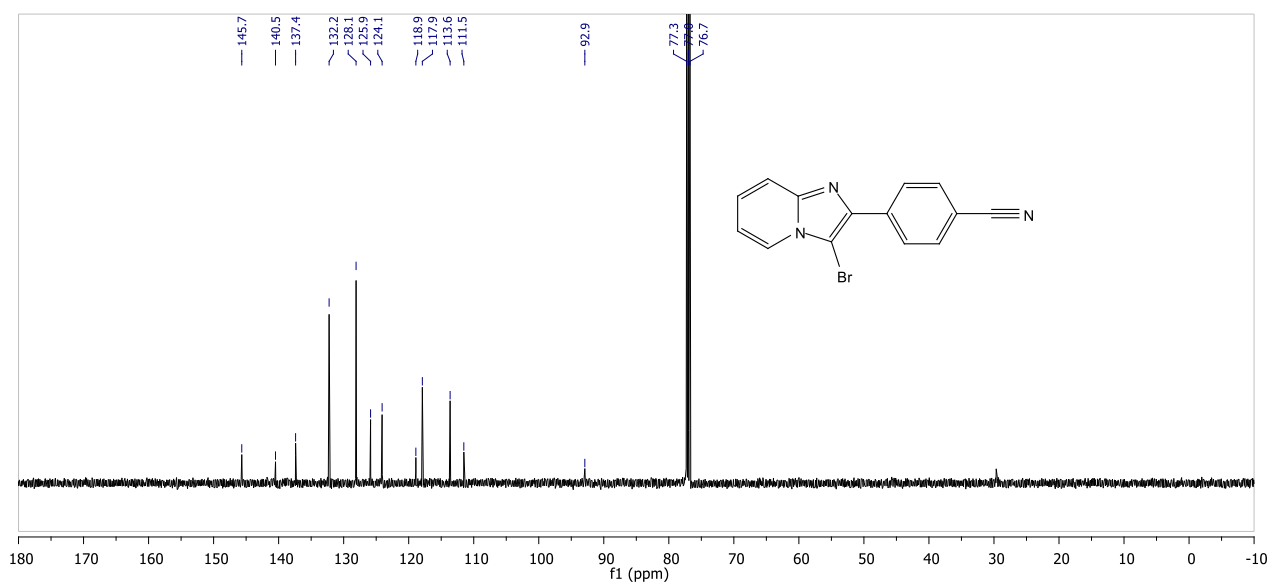
¹³C NMR (126 MHz, CDCl₃) of compound **2x**



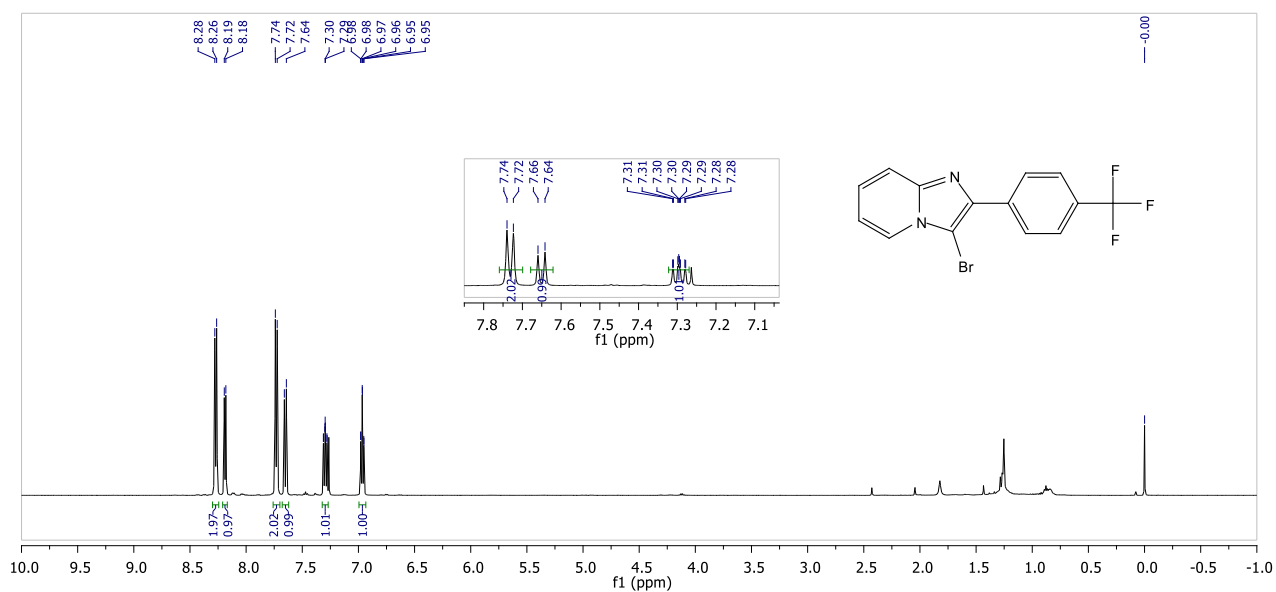
¹H NMR (500 MHz, CDCl₃) of compound **2y**



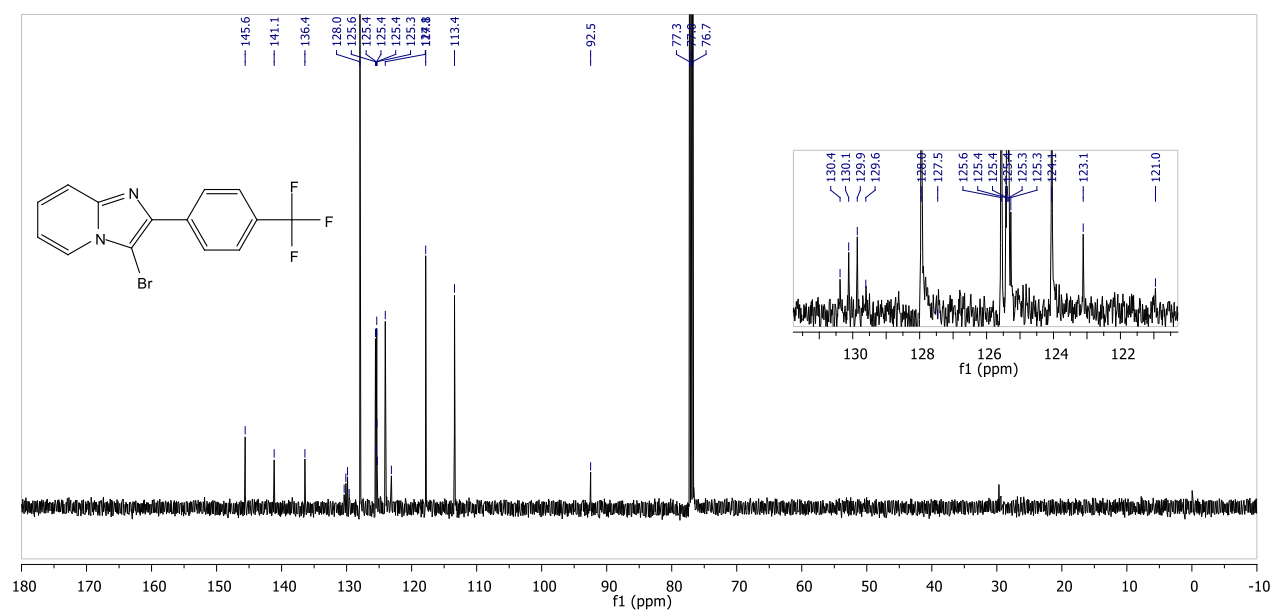
¹³C NMR (126 MHz, CDCl₃) of compound **2y**



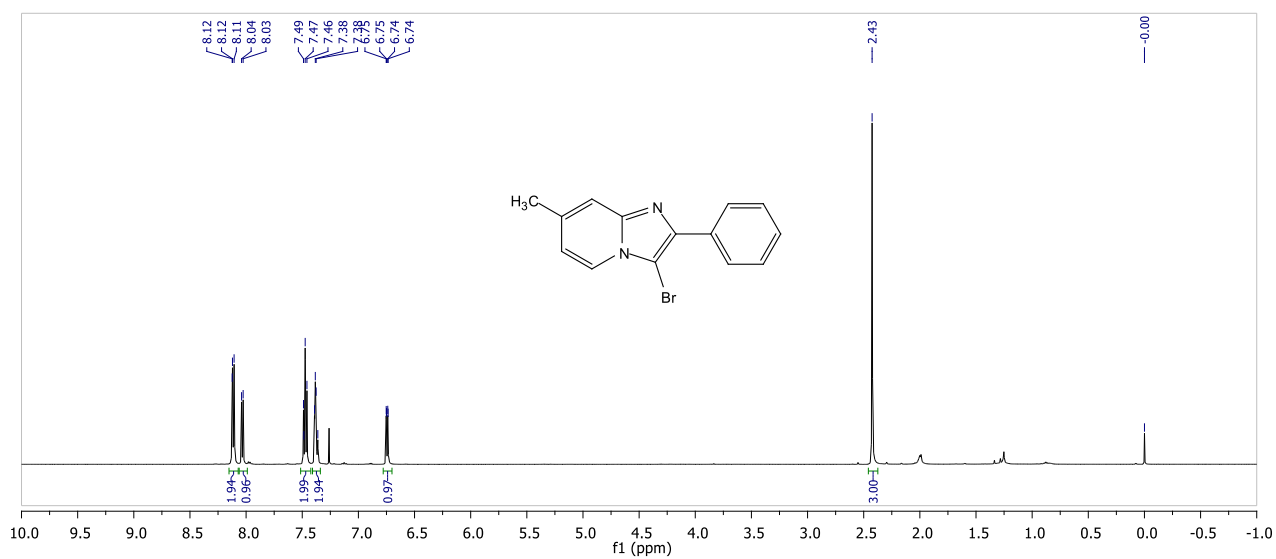
¹H NMR (500 MHz, CDCl₃) of compound **2z**



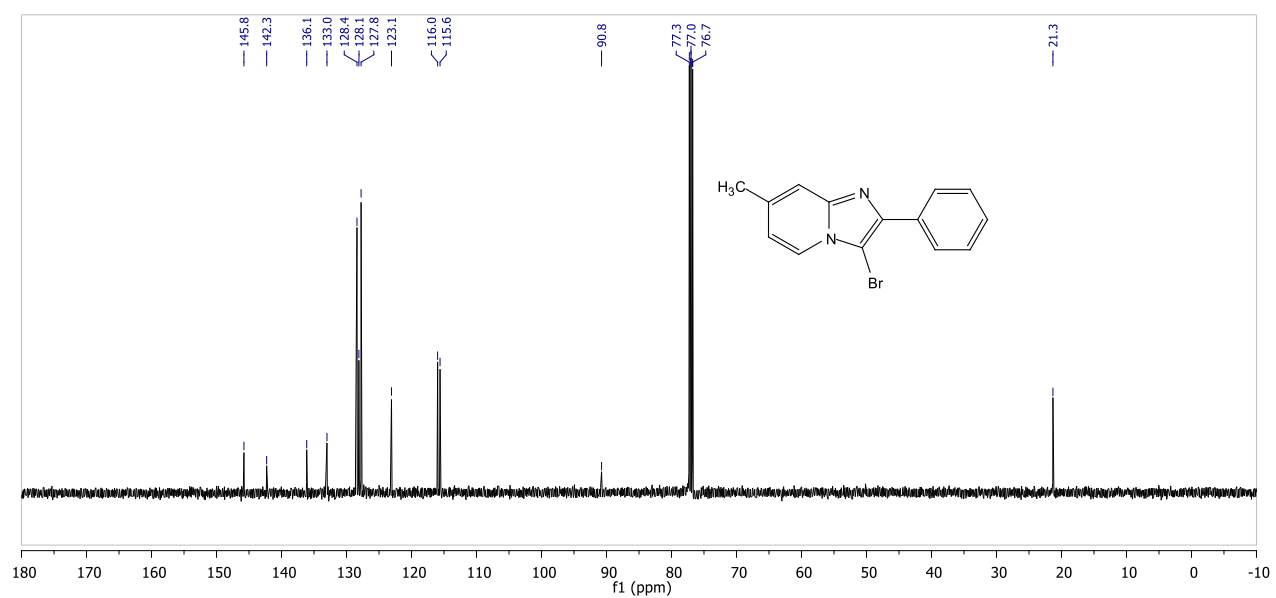
¹³C NMR (126 MHz, CDCl₃) of compound **2z**



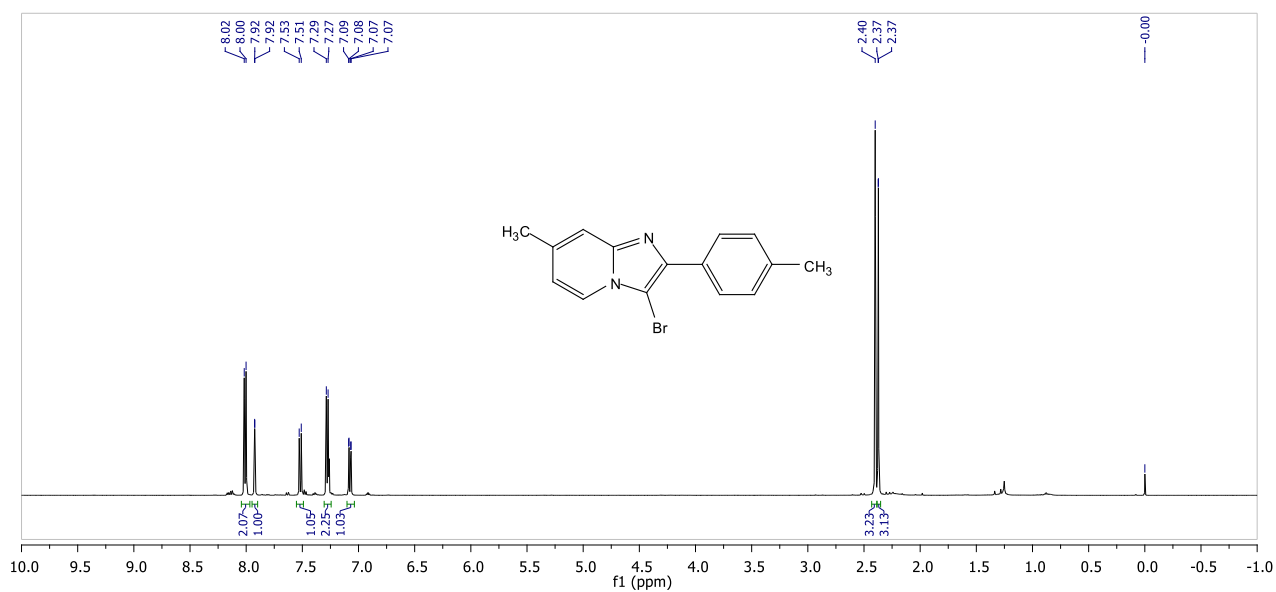
¹H NMR (500 MHz, CDCl₃) of compound **2aa**



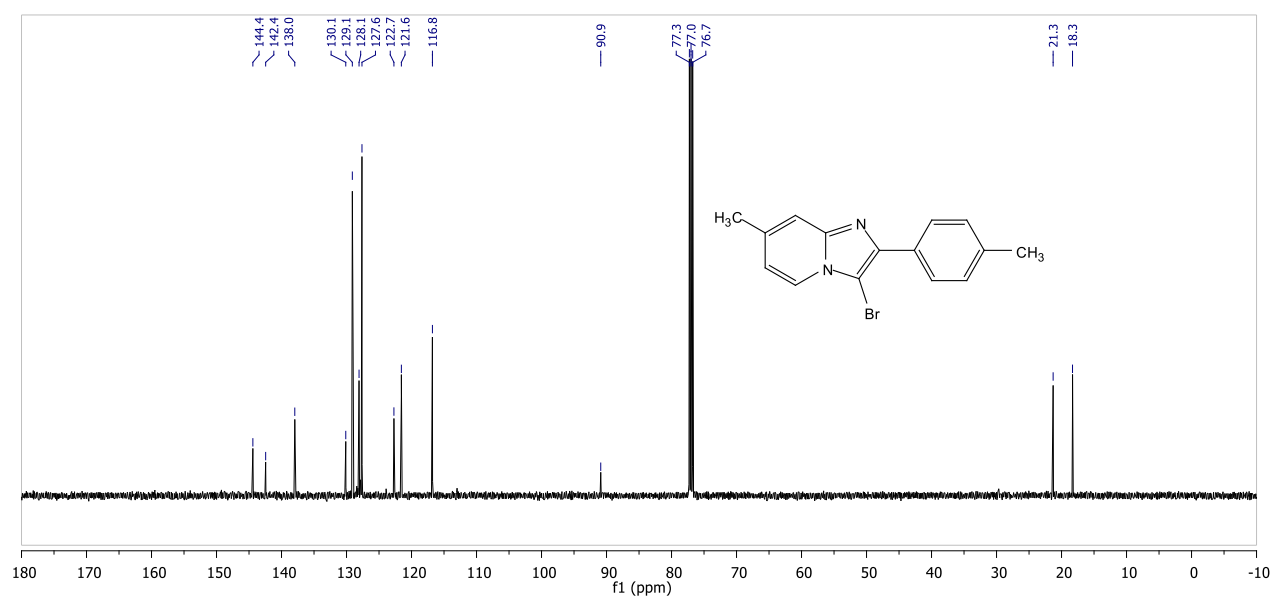
¹³C NMR (126 MHz, CDCl₃) of compound **2aa**



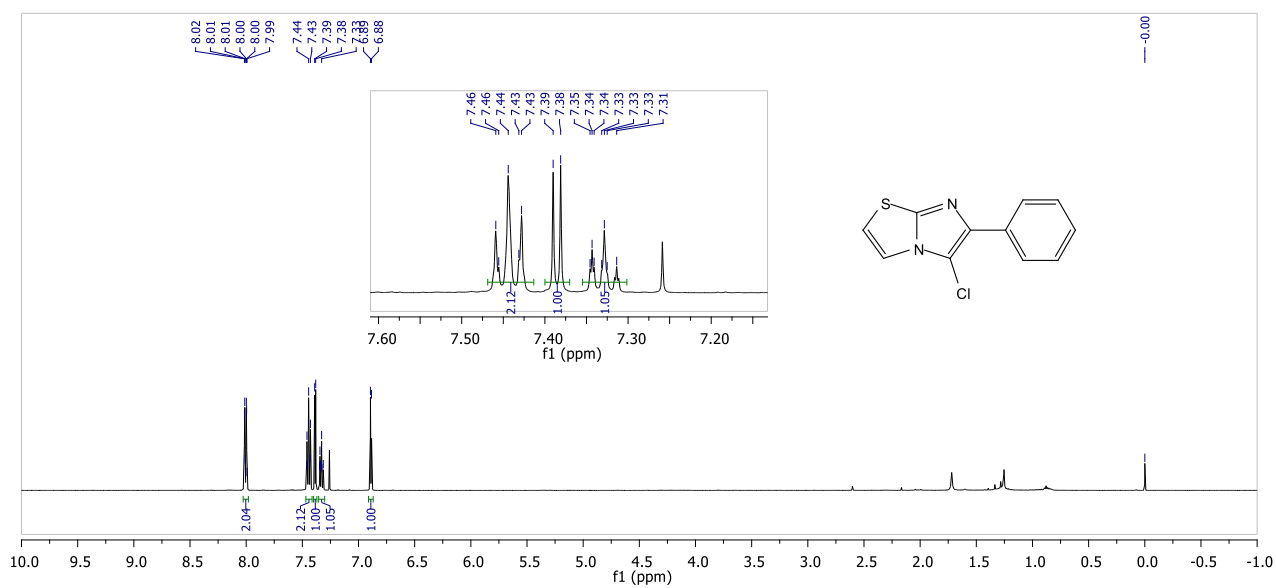
¹H NMR (500 MHz, CDCl₃) of compound **2ab**



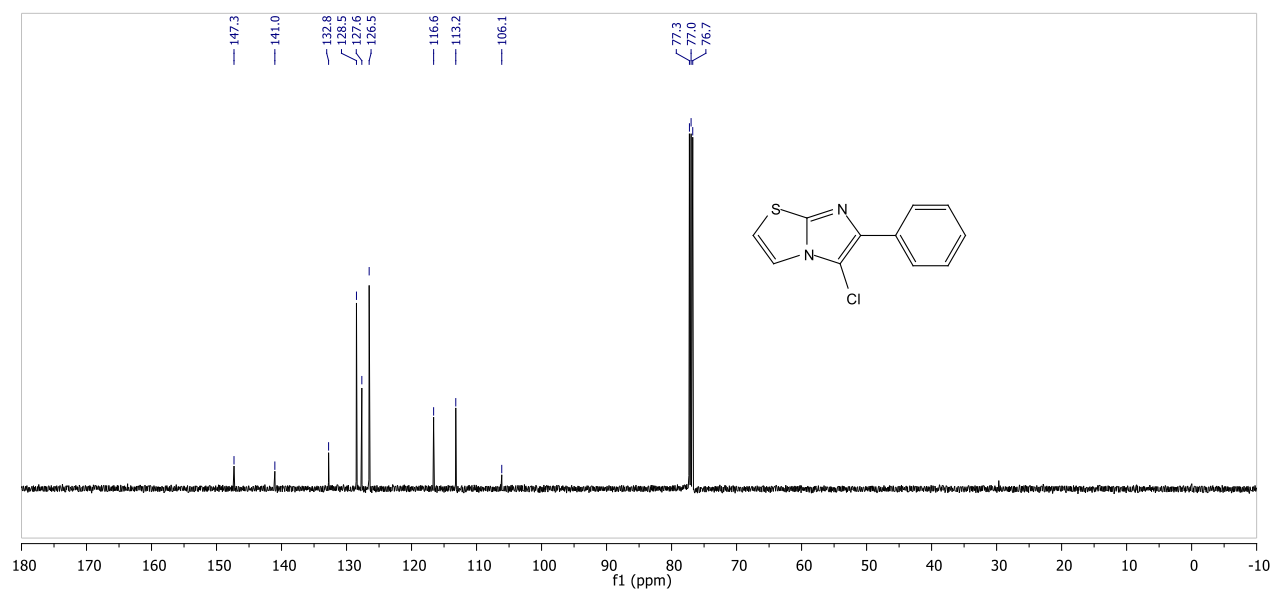
¹³C NMR (126 MHz, CDCl₃) of compound **2ab**



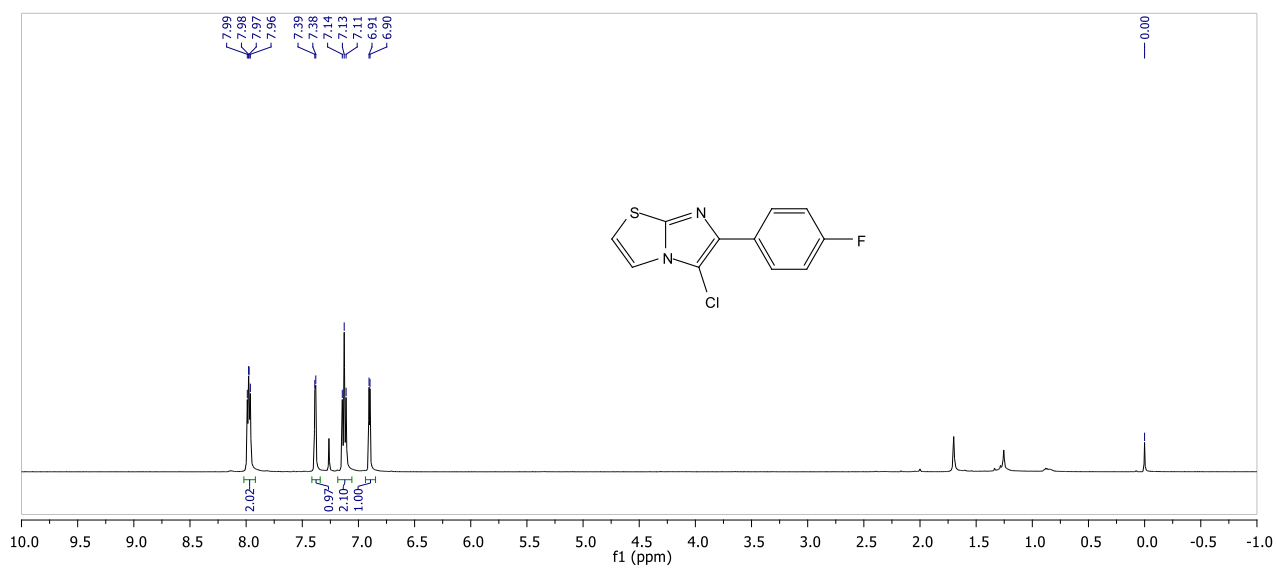
¹H NMR (500 MHz, CDCl₃) of compound **3a**



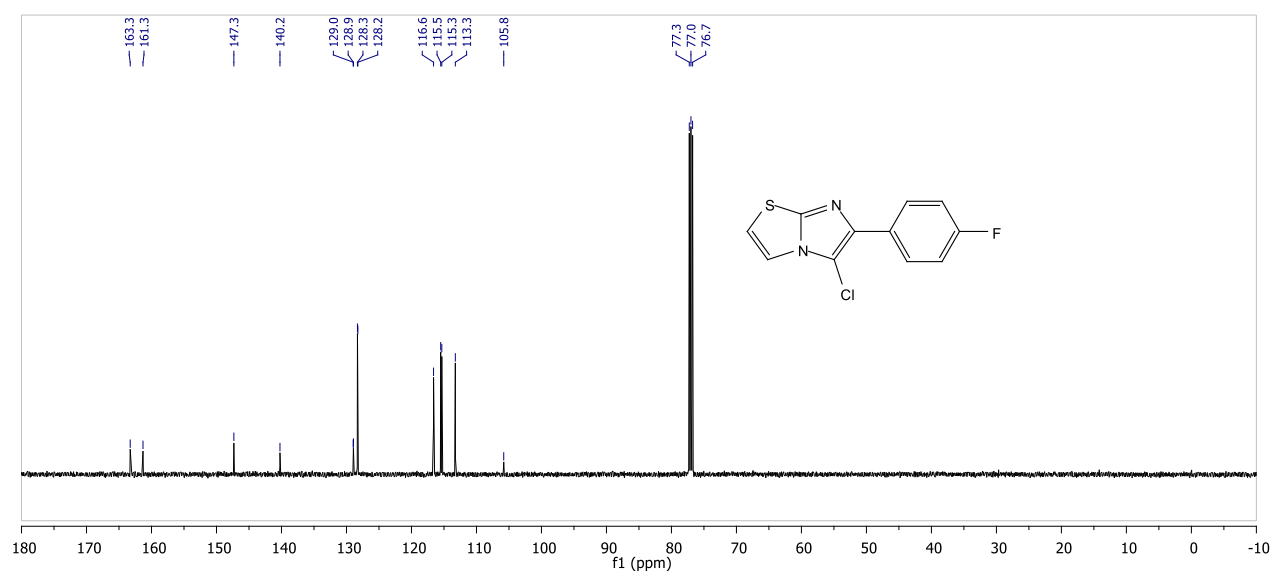
¹³C NMR (126 MHz, CDCl₃) of compound **3a**



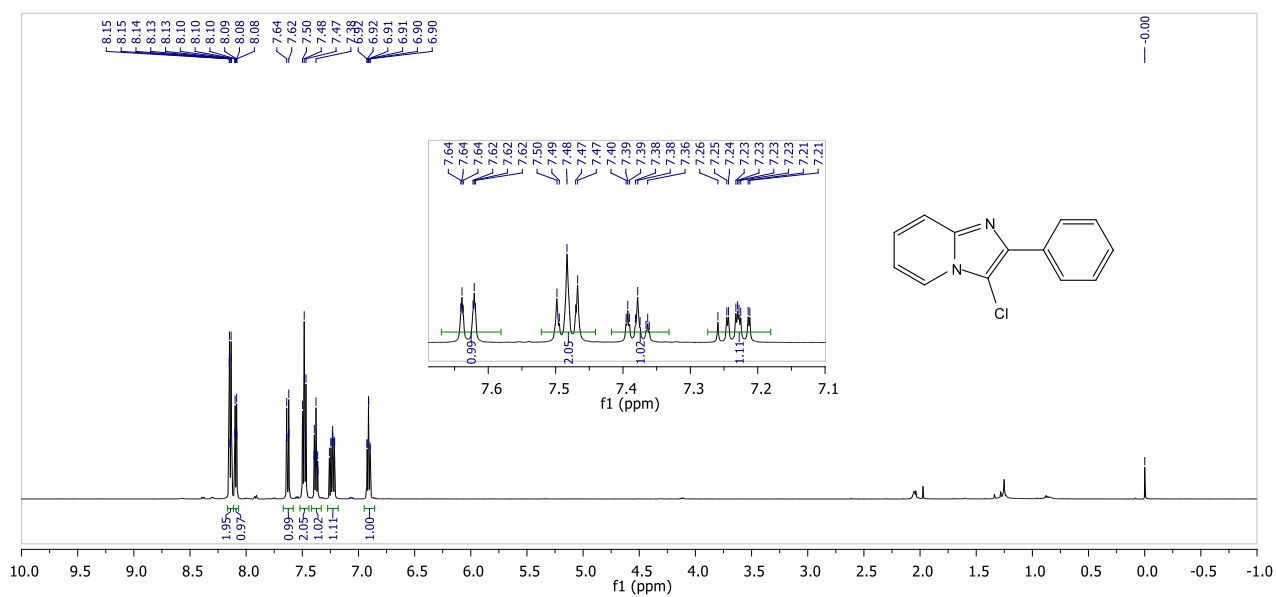
¹H NMR (500 MHz, CDCl₃) of compound **3b**



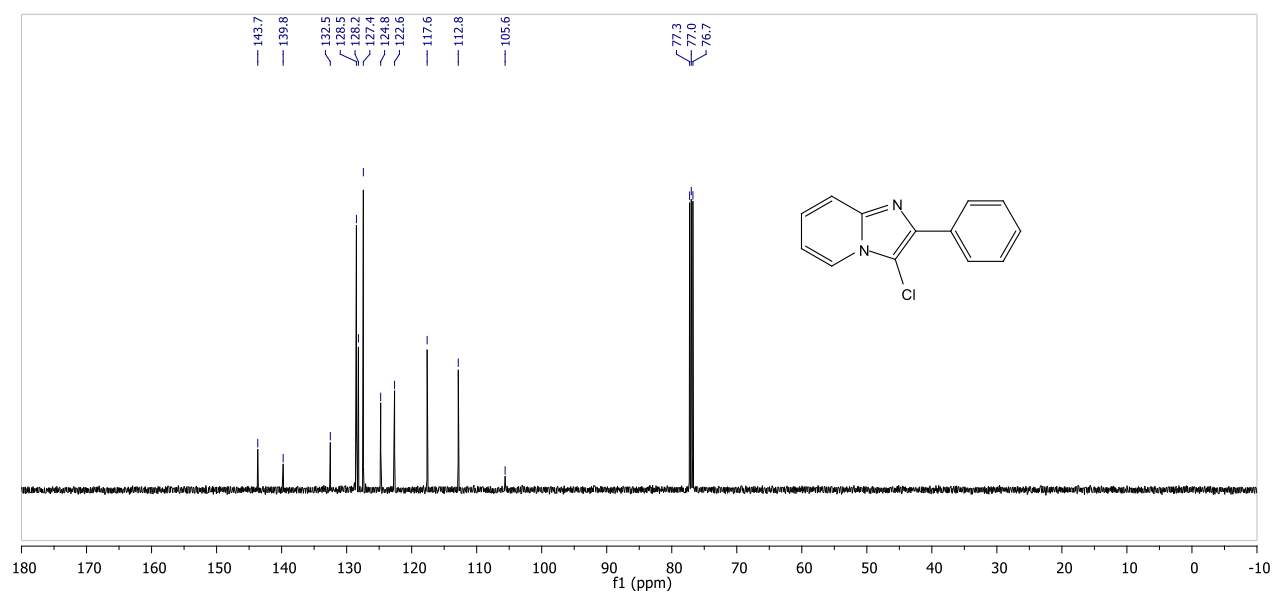
¹³C NMR (126 MHz, CDCl₃) of compound **3b**



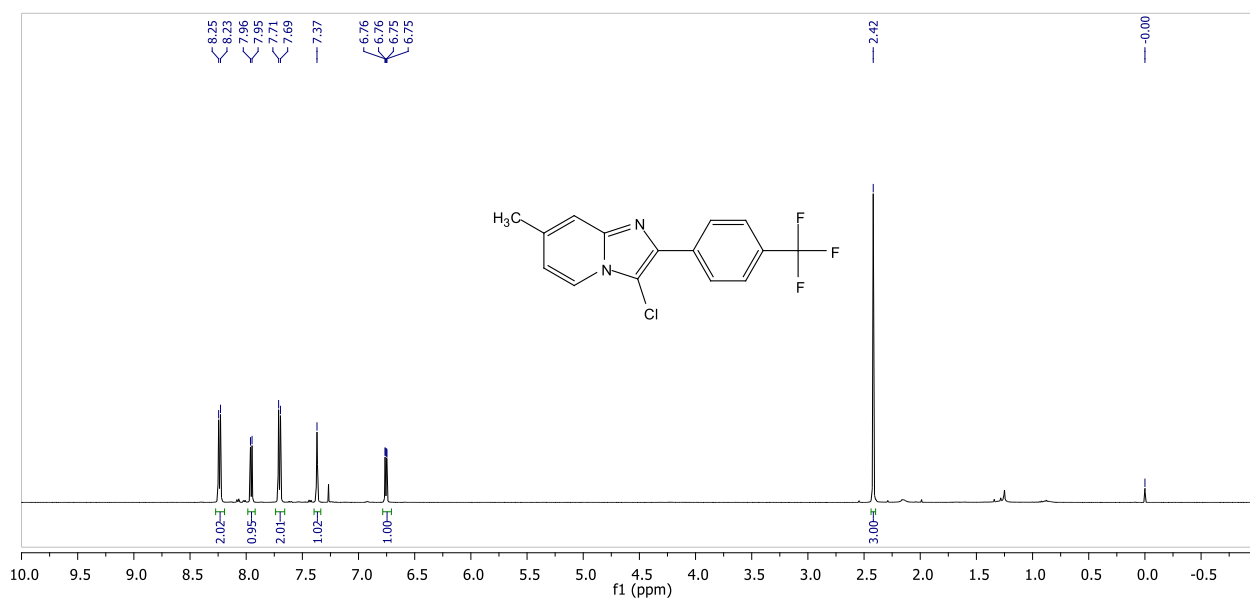
¹H NMR (500 MHz, CDCl₃) of compound **3c**



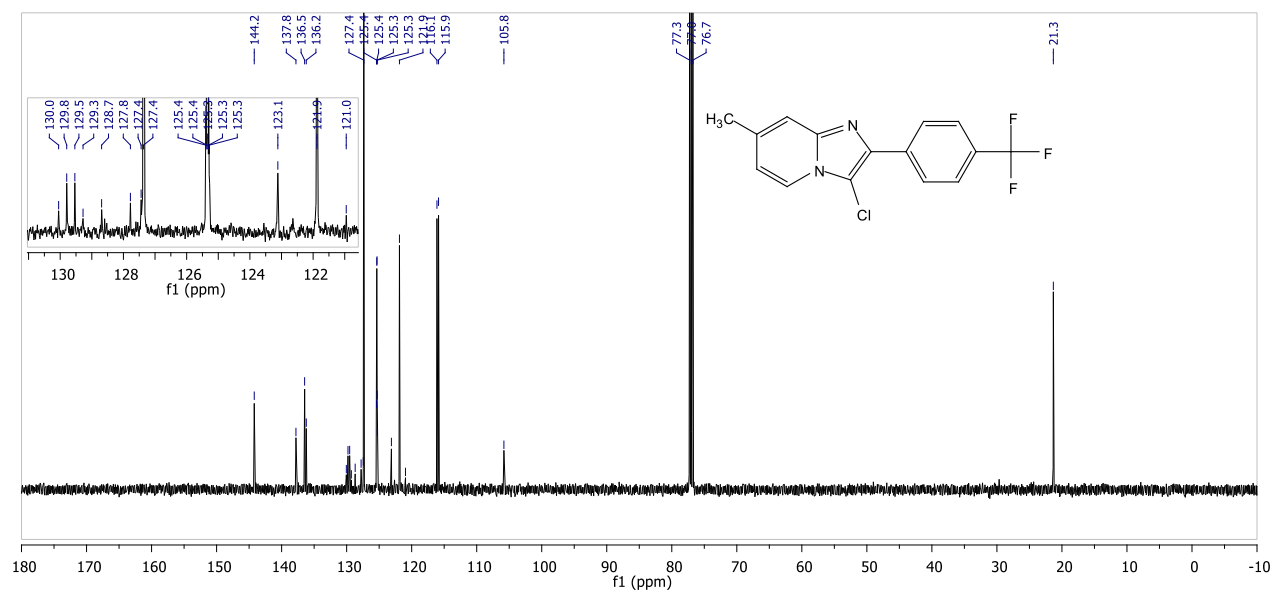
¹³C NMR (126 MHz, CDCl₃) of compound **3c**



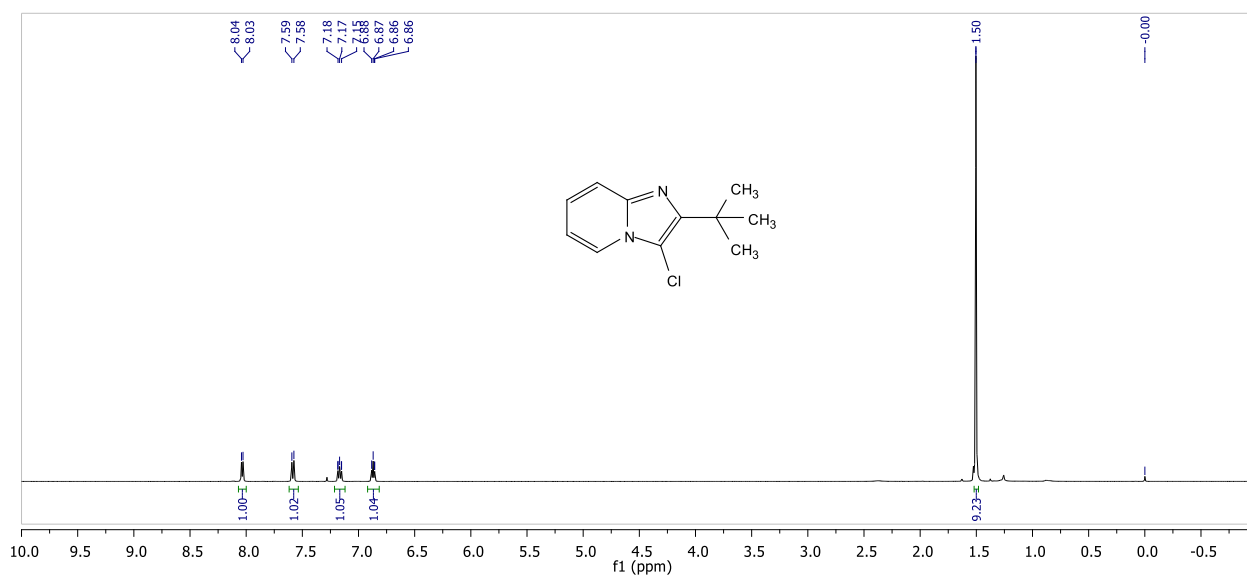
^1H NMR (500 MHz, CDCl_3) of compound **3d**



^{13}C NMR (126 MHz, CDCl_3) of compound **3d**



¹H NMR (500 MHz, CDCl₃) of compound **3e**



¹³C NMR (126 MHz, CDCl₃) of compound **3e**

