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Electronic Supplementary Information

Carbon dot/TiO₂ nanocomposites as photocatalysts for metallaphotocatalytic carbon-heteroatom cross-couplings

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1. General materials and methods

All substrates, reagents, and solvents were obtained from commercial suppliers and were used without further purification unless otherwise noted. All light-emitting diode purchased from Kessil (LED) lamps were Lighting (https://www.kessil.com/science/index.php). Analytical thin layer chromatography (TLC) was performed on pre-coated TLC-sheets, ALUGRAM Xtra SIL G/UV₂₅₄ sheets (Macherey-Nagel) and visualized with 254 nm light or staining solutions followed by heating. Purification of final compounds was carried out by flash chromatography on the Reveleris X2 Flash Chromatography System from GRACE using prepacked columns with 40 µm silica gel. Silica 60 M (0.04-0.063 mm) silica gel (Sigma Aldrich) was used for dry loading of the crude compounds on the flash chromatography system. Carbon dots (CDs) were synthesized using a domestic microwave (SEVERIN). Centrifugation of CDs and CD/titanium dioxide (TiO₂ P25) nanocomposites were carried out using an Eppendorf 5810R centrifuge and an Eppendorf 5430 centrifuge, respectively. NMR spectra were recorded on an AscendTM 400 (400 MHz, Bruker) spectrometer, and are reported in ppm relative to the residual solvent peaks. Peaks are reported as: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet or unresolved, with coupling constants in Hz. Infrared (IR) spectra were recorded on a Perkin-Elmer FT-IR spectrometer (Spectrum 100). The morphologies of CDs were observed using transmission electron microscopy (TEM, Zeiss EM 912Ω). The prepared CDs were diluted with distilled water to suitable concentrations and then deposited onto carboncoated copper grids. After drying at room temperature, the samples were imaged. The morphologies and elemental composition of CD/TiO₂ P25 nanocomposites were measured using scanning electron microscopy and energy dispersive X-ray spectroscopy (SEM-EDX, LEO 1550 system). The freeze-dried CD/TiO₂ P25 nanocomposites were suspended in distilled water and sonicated for 5 min. The resulting suspension was deposited on the silicon substrate and coated with Au. After drying at room temperature, the samples were imaged. Fluorescence spectra of CDs were measured using a microplate reader (SpectraMax M5, Molecular Devices). Absorption spectra of CDs and CD/TiO₂ P25 nanocomposites were collected using a Shimadzu UV-1900 (solutions), or a Shimadzu UV-2600 spectrometer equipped with an integrating sphere (solids). X-ray diffraction (XRD) spectrum was measured with a Bruker D8 Advanced X-ray diffractometer with Cu Kα radiation. Zeta potential was measured with a dynamic light scattering instrument (DLS, Zetasizer Nano, Malvern). Photoluminescence lifetime was measured using the time-correlated single photon counting technique (TCSPC, FluoTime 250, fluorescence lifetime spectrometer). Inductively coupled plasma - optical emission spectrometry (ICP-OES) was carried out using a Horiba Ultra 2 instrument equipped with a photomultiplier tube detection system.

2. Synthesis of CDs

The carbohydrate carbon source was dissolved in ultrapure H₂O (20 mL) in a conical flask (300 mL). The doping agent was added and the mixture was agitated to yield a homogenous solution. The flask was transferred into a domestic microwave in a fume hood and heated at 700 W for the respective time. The crude mixture was cooled for 10 min before ultrapure H₂O (40 mL) was added. The resulting solution was filtered through a filter paper and centrifuged at 8000 rpm for 0.5 h through Amicon® Ultra-15 centrifugal filter units. The filtrate was lyophilized to yield the desired CDs.

Table S1 Summary of the conditions used for the synthesis of the CDs.

Entry	CD	Carbon source (mmol/mL)	Doping agent (mmol/mL)	Time / min
1	CD1	GlcN·HCl (0.12)	β-Ala (0.13)	3
2	CD2 ^a	GlcN·HCl (0.15)	1,3-Diaminobenzene (0.17)	3
3	CD3		1-Cys (0.25)	3
4	CD4	GlcN·HCl (0.23)	PEG (0.25)	9
5	CD5		Gly (0.25)	3
6	CD6	Glc (0.12)		4.5
7	CD7	GlcNAc (0.12)		4.5
8	CD8	Gal (0.12)	β-Ala (0.13)	4.5
9	CD9	Lac (0.12)		4.5
10	CD10	Pullulan (25 mg/mL)		5

^aThe doping agent was first dissolved in MeOH (10 mL) and then added to the aqueous solution (20 mL) of carbon source. GlcN·HCl = glucosamine hydrochloride. Glc = glucose. GlcNAc = *N*-acetylglucosamine. Gal = galactose. Lac = D-lactose. β-Ala = β-alanine. l-Cys = l-cysteine. PEG = poly(ethylene glycol) (average M_n 400). Gly = glycine.

Fig. S1 Chemical structures of carbon sources and doping agents used for CD synthesis.

3. Preparation of CD/TiO₂ P25 nanocomposites

TiO₂ P25 (120 mg) was dispersed in ultrapure H₂O (80 mL) in a round-bottom flask. The respective amount of CDs was added to prepare nanocomposites with different CD-to-TiO₂ P25 mass ratios. The mixture was shielded from light and stirred for 24 h at room temperature. The resulting CD/TiO₂ nanocomposites were centrifuged at 5000 rpm for 15 min and further washed two times with ultrapure H₂O. The nanocomposites were lyophilized to afford a light brown powder. The amount of CDs immobilized on the TiO₂ P25 surface was determined using UV-Vis absorption spectroscopy.

Table S2 Conditions used for the preparation of the CD/TiO₂ nanocomposites.

Entry	CD	CD-to-TiO ₂ P25 mass ratio	CD Immobilization (weight % (CD/nanocomposite))
1		1:20	1.5
2		1:10	1.8
3	CD1	1:4	2.4
4	CD1	1:1	5.0
5		4:1	8.1
6		1:1 (CD-to-SiO ₂)	1.6
7	CD2	1:1	_a
8	CD3	1:1	2.5
9	CD4	1:1	13.6
10	CD5	1:1	3.4
11	CD6	1:1	3.7
12	CD7	1:1	11.9
13	CD8	1:1	3.0
14	CD9	1:1	11.4
15	CD10	1:1	27.2

^aQuantification by UV-Vis was not possible due to the formation of side-products in the solution that affected the measurements.

4. Characterization of CDs

4.1. CD1

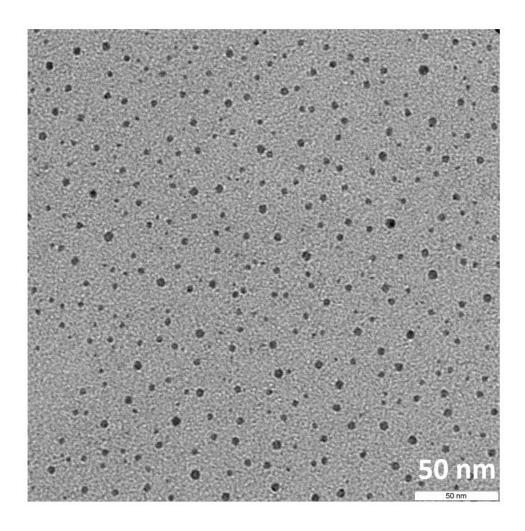


Fig. S2 TEM image of CD1.



Fig. S3 Photographs of an aqueous solution of CD1 in daylight (left) and under UV light irradiation ($\lambda_{ex} = 366$ nm, right).

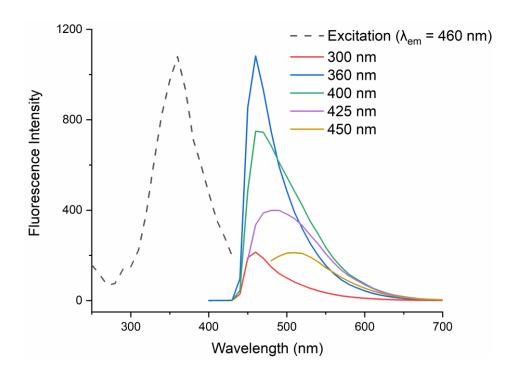


Fig. S4 Excitation ($\lambda_{em} = 460$ nm) and emission spectra of **CD1** recorded upon excitation with different excitation wavelengths (H₂O, 298 K).

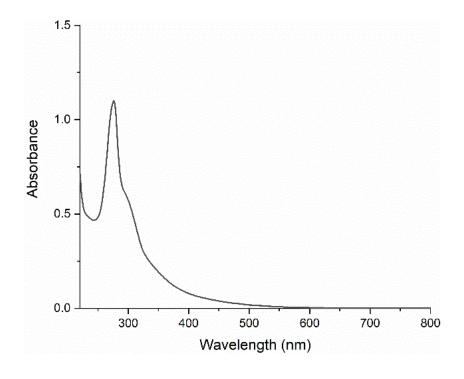


Fig. S5 Absorption spectrum of CD1 (H₂O, 298 K).

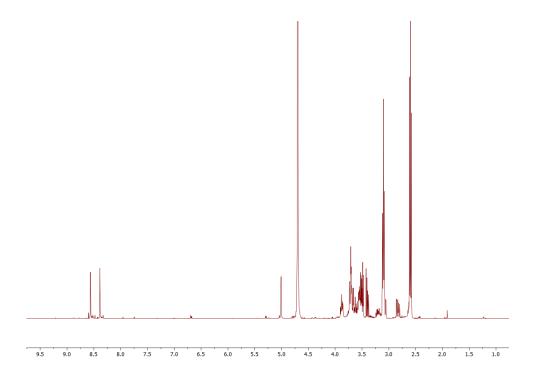


Fig. S6 1 H NMR spectrum of **CD1** in D₂O (400 MHz). The spectrum is in agreement with previously reported data, 1 indicating the presence of the β-alanine on **CD1** surface.

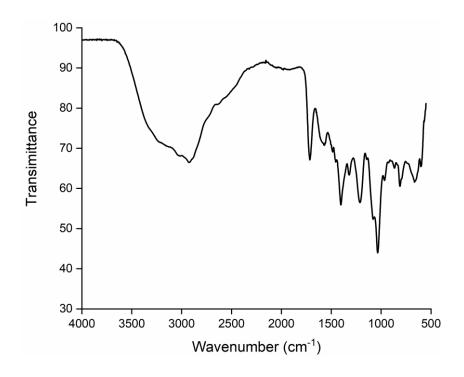


Fig. S7 IR spectrum of **CD1**. Key features: 2927 cm⁻¹ (O-H); 1715 cm⁻¹ (C=O).

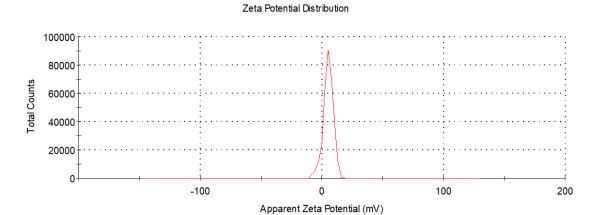


Fig. S8 Zeta potential of **CD1** (-11.1 to +18.7 mV, H₂O, 298 K).

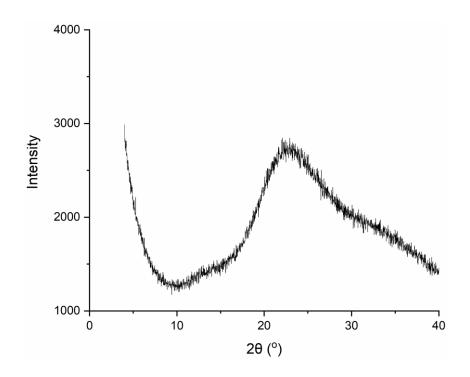
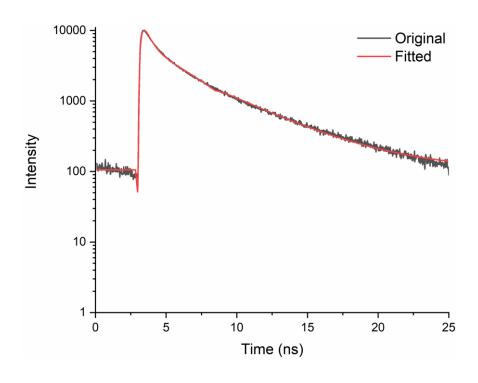


Fig. S9 Powder XRD profile of CD1 confirming its amorphous nature.



A_1	$\mathbf{A_2}$	τ ₁ (ns)	τ_2 (ns)
10.49	6.131	0.451	4.454

Fig. S10 Photoluminescence lifetime of CD1 (H₂O, 298 K).

4.2. Characterization of CD2-CD10

CD2 CD3 CD5 CD4 CD6 CD7

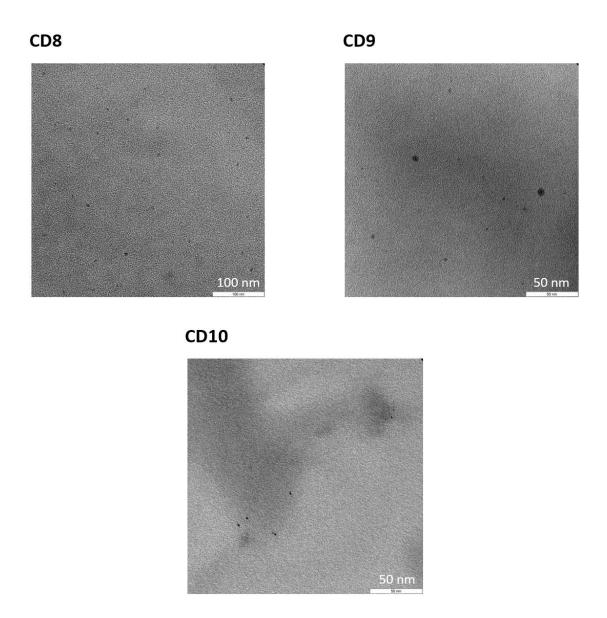


Fig. S11 TEM images of CD2, CD3, CD4, CD5, CD6, CD7, CD8, CD9 and CD10.

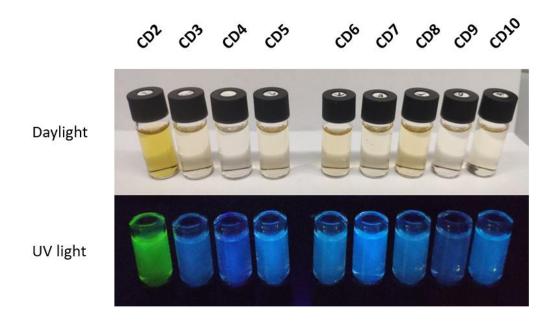
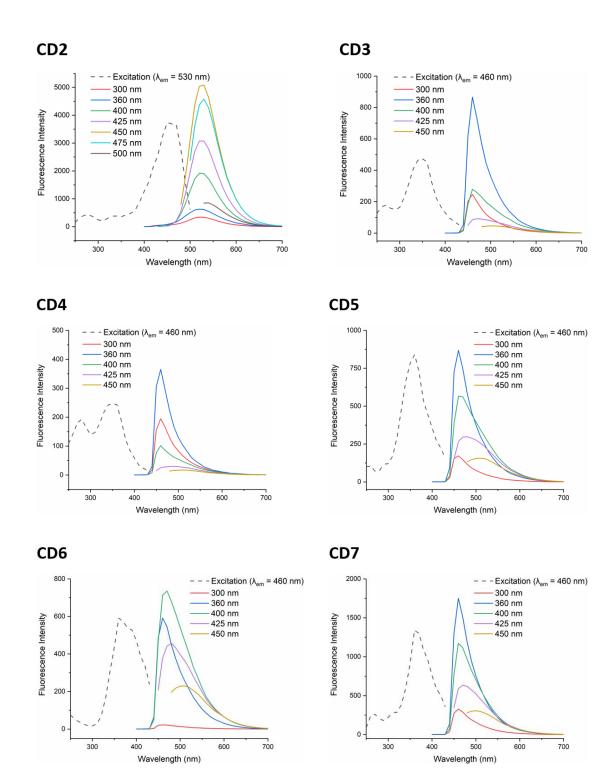


Fig. S12 Photographs of aqueous solutions of CD2, CD3, CD4, CD5, CD6, CD7, CD8, CD9 and CD10 in daylight and under UV light irradiation ($\lambda_{ex} = 366$ nm).



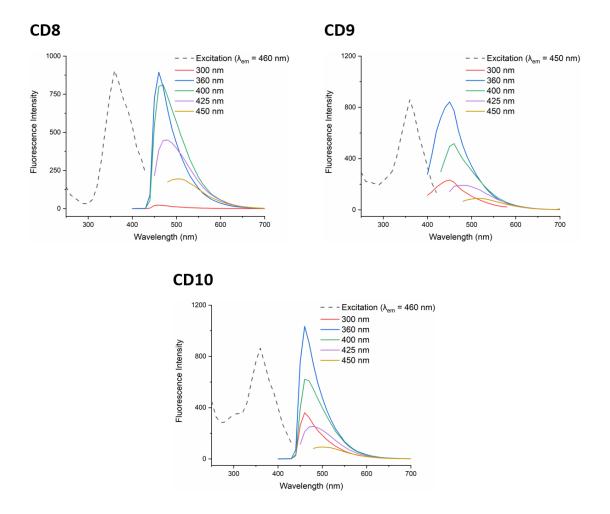
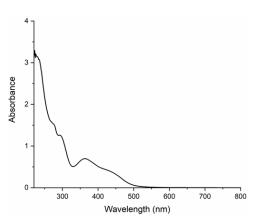
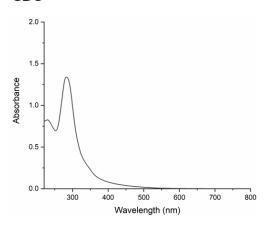


Fig. S13 Excitation and emission spectra of **CD2**, **CD3**, **CD4**, **CD5**, **CD6**, **CD7**, **CD8**, **CD9** and **CD10** recorded upon excitation with different excitation wavelengths (H₂O, 298 K).

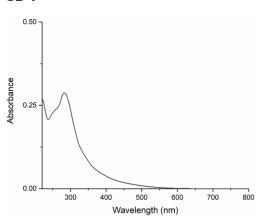
CD2



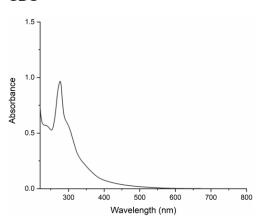
CD3



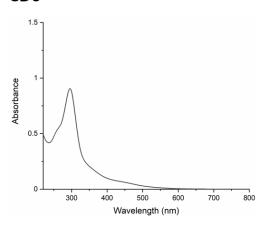
CD4



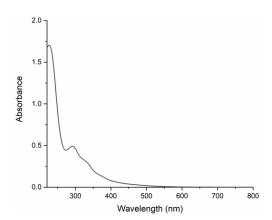
CD5



CD6



CD7



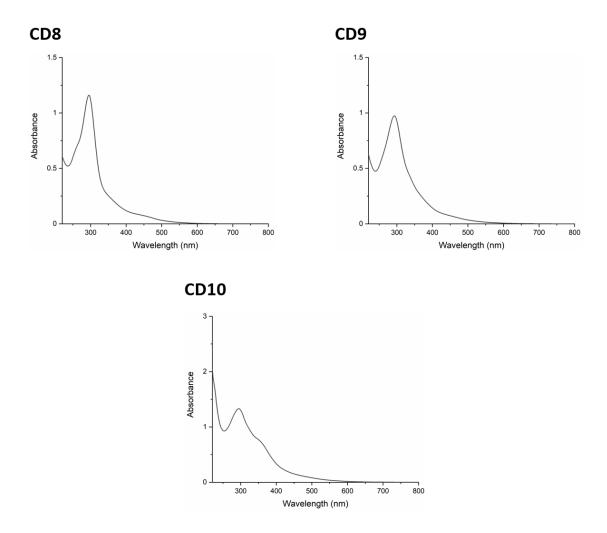


Fig. S14 Absorption spectra of CD2, CD3, CD4, CD5, CD6, CD7, CD8, CD9 and CD10 (H_2O , 298 K).

5. Characterization of CD/TiO2 nanocomposites

5.1. CD1/TiO₂

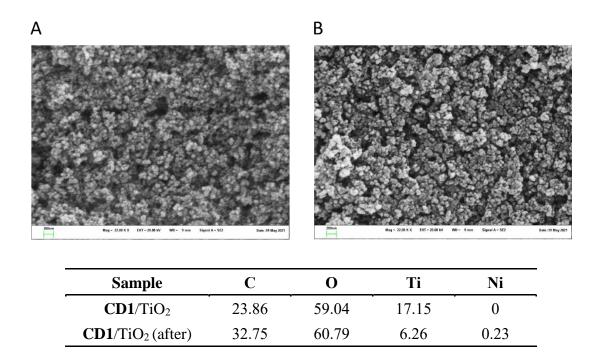


Fig. S15 SEM image and elemental composition of **CD1**/TiO₂ P25 nanocomposite (A) and **CD1**/TiO₂ P25 nanocomposite after the catalytic reaction (B). All values are given as atomic %.

Table S3 ICP-OES measurements of the nickel content of **CD1**/TiO₂ P25 nanocomposite and **CD1**/TiO₂ P25 nanocomposite after the catalytic reaction.

Sample	Ni (mg/g)
CD1/TiO ₂	0.016
CD1/TiO ₂ (after)	24.8

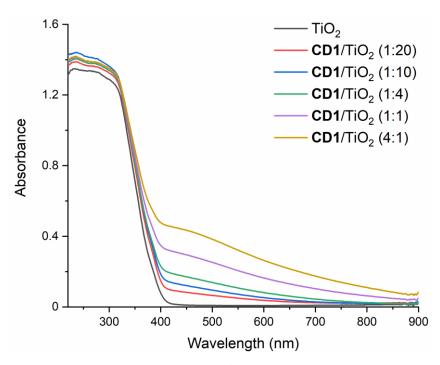


Fig. S16 Absorption spectra (solid state) of **CD1**/TiO₂ nanocomposites prepared using different **CD1**-to-TiO₂ P25 mass ratios.

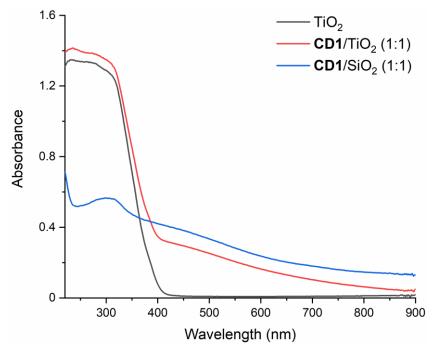
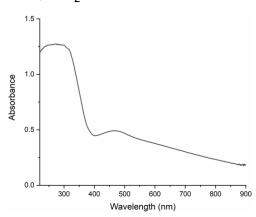


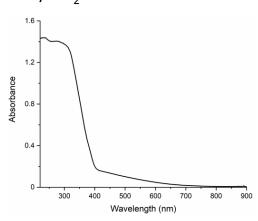
Fig. S17 Absorption spectra (solid state) of **CD1**/TiO₂ and **CD1**/SiO₂ nanocomposites (mass ratio = 1:1).

5.2. Characterization of nanocomposites prepared using CD2-CD10

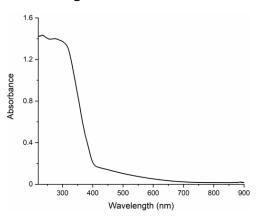
CD2/TiO₂



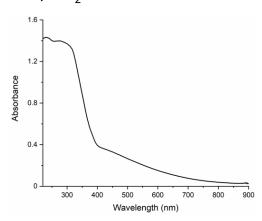
CD3/TiO₂



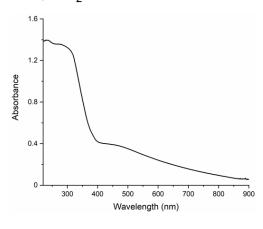
CD4/TiO₂



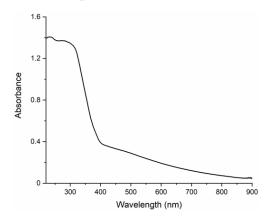
CD5/TiO₂



CD6/TiO₂



CD7/TiO₂



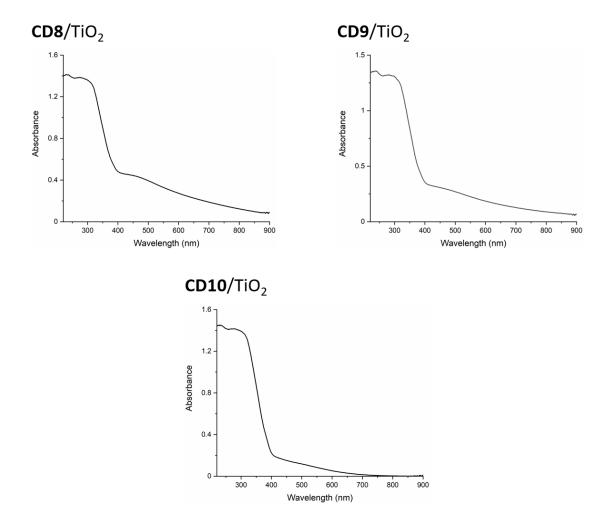


Fig. S18 Absorption spectra (solid state) of $CD2/TiO_2$, $CD3/TiO_2$, $CD4/TiO_2$, $CD5/TiO_2$, $CD6/TiO_2$, $CD7/TiO_2$, $CD8/TiO_2$, $CD9/TiO_2$ and $CD10/TiO_2$ nanocomposites.

6. Cross-coupling reaction

Setup for blue light experiments.¹ Two vials were placed in the middle of the stirring plate (4.5 cm away from single lamp). The reaction was irradiated with a single blue LED lamp (Kessil PR160L-440). The fan was used to avoid possible heating of the reaction mixture.



Fig. S19 Image of the setup using a single blue LED lamp.

Setup for green light experiments.¹ Two vials were placed in the middle of the stirring plate (4.5 cm away from each lamp). The reaction was irradiated with two green LED lamps (Kessil PR160L-525). The fan was used to avoid the possible heating of the reaction mixture.



Fig. S20 Image of the setup using two green LED lamps.

6.1. C-O arylation of Boc-Pro-OH

General experimental procedure. Boc-Pro-OH (*N*-(*tert*-butoxycarbonyl)-L-proline, 41.0 mg, 190.3 μmol), methyl 4-iodobenzoate (74.8 mg, 285.4 μmol) and the respective CD/TiO₂ nanocomposite (*Table S4*) were added to an oven-dried glass vial equipped with a stir bar. Subsequently, a DMSO solution (3 mL) of dcbpy (2,2'-bipyridine-4,4'-dicarboxylic acid, 4.7 mg, 19.0 μmol), NiCl₂·6H₂O (nickel(II) chloride hexahydrate, 4.5 mg, 19.0 μmol), and BIPA (*N*-*tert*-butylisopropylamine, 90.5 μL, 570.8 μmol) were added. The glass vial was sealed with a septum and Parafilm. The reaction mixture was stirred and sonicated for 10 min to obtain a fine dispersion and subsequently degassed with Argon for 10 min. The vial was then irradiated with the respective LED lamps at room temperature for the respective time. 1,3,5-Trimethoxybenzene (32.0 mg, 190.3 μmol) was added as internal standard to determine NMR yields. An aliquot of the resulting mixture (~250 μL) was filtered through a syringe filter, diluted with DMSO-d₆ (~250 μL) and subjected to ¹H-NMR analysis.

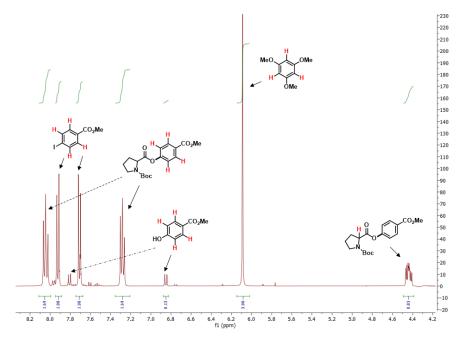


Fig. S21 Representative ¹H-NMR spectrum of the crude reaction mixture to determine yields by ¹H-NMR analysis using 1,3,5-trimethoxybenzene as internal standard (DMSO-d₆, 400 MHz).

Table S4 Summary of the CD/TiO₂ nanocomposites used in the photocatalyzed C-O cross-coupling of Boc-Pro-OH with methyl 4-iodobenzoate.

Entry	Nanocomposite	Initial CD-to-TiO ₂ P25 mass ratio ^[a]	CD Immobilization ^[b] (weight % (CD/nanocomposite))	Amount ^[c] / mg
1	TiO ₂ P25	-	-	30.0
2	CD1	-	-	30.0
3		1:20	1.5	30.5
4		1:10	1.8	30.5
5	CD1/T:O	1:4	2.4	30.8
6	$CD1/TiO_2$	1:1	5.0	31.6
7		4:1	8.1	32.6
8		1:1 (CD-to-SiO ₂)	1.6	30.5
9	CD2/TiO ₂	1:1	-	30.0
10	$CD3/TiO_2$	1:1	2.5	30.8
11	CD4/TiO ₂	1:1	13.6	34.7
12	CD5 /TiO ₂	1:1	3.4	31.1
13	CD6/TiO ₂	1:1	3.7	31.1
14	CD7 /TiO ₂	1:1	11.9	34.1
15	$\mathbf{CD8}/\mathrm{TiO}_2$	1:1	3.0	30.9
16	CD9/TiO ₂	1:1	11.4	33.9
17	CD10/TiO ₂	1:1	27.2	41.2

[[]a] CD-to-TiO₂ P25 mass ratio used for the preparation of the nanocomposite. [b] Weight % of CD immobilized on TiO₂ as calculated by UV-Vis spectroscopy. [c] Amount of nanocomposite used in the C-O arylation of Boc-Pro-OH.

Procedure to obtain the isolated yield. An oven-dried vial (19 x 100 mm) equipped with a stir bar was charged with CD1/TiO₂ (63.2 mg), Boc-Pro-OH (82 mg, 380.6 μmol, 1.0 equiv), methyl 4-iodobenzoate (149.6 mg, 571 μmol, 1.5 equiv), NiCl₂· 6H₂O (9 mg, 38 μmol, 10 mol%) and dcbpy (9.3 mg, 38 μmol, 10 mol%). Subsequently, DMSO (anhydrous, 6 mL) and BIPA (180.9 µL, 1.14 mmol, 3.0 equiv) were added and the vial was sealed with a septum and Parafilm. The reaction mixture was sonicated for 5-10 min followed by stirring for 5 min to obtain a fine dispersion. The mixture was then degassed by bubbling Argon for 10 min. The mixture was irradiated using the 440 nm setup with rapid stirring (1400 rpm). After 40 h, one equivalent of 1,3,5trimethoxybenzene (internal standard, 64 mg, 380 µmol) was added and the mixture was stirred for 5 min. An aliquot of the reaction mixture (~200 µL) was filtered, diluted with DMSO-d₆ and subjected to ¹H-NMR analysis to determine the NMR yield (90%). Thereafter, the NMR sample was combined with the reaction mixture. The reaction mixture was diluted with H₂O (40 mL) and extracted with dichloromethane (3 x 30 mL). The combined organic phases were washed with brine (50 mL), dried over Na₂SO₄ and concentrated. The product was purified by flash column chromatography (SiO₂, Hexane/EtOAc elution gradient of 0-20%) on a Grace Reveleris system using a 12 g cartridge. In some cases, mixed fractions containing small amounts of the phenol byproduct and the desired product were observed. These could be easily purified by a basic extraction (DCM and 0.5 M NaOH), followed by drying the organic phase over Na₂SO₄ and solvent evaporation to maximize the reaction yield. The title compound was isolated as a yellowish solid.

Isolated yield: 84% (111.5 mg, 319 µmol)

1-(tert-butyl) 2-(4-(methoxycarbonyl)phenyl) pyrrolidine-1,2-dicarboxylate 1: 1 H NMR (400 MHz, CDCl₃) rotameric mixture, δ 8.04 (m, 2H), 7.16 (m, 2H), 4.49 (dd, J = 8.6, 4.4 Hz, 0.4H), 4.43 (dd, J = 8.7, 4.3 Hz, 0.6H), 3.87 (m, 3H), 3.66 – 3.39 (m, 2H), 2.42 – 2.26 (m, 1H), 2.19 – 2.09 (m, 1H), 2.07 – 1.86 (m, 2H), 1.44 (m, 9H). 13 C NMR (151 MHz, CDCl₃) rotameric mixture, signals for minor rotamer are enclosed in parenthesis δ (171.21) 171.16, (166.37) 166.23, 154.51 (154.23), 153.68, 131.29 (131.15), 127.87 (127.71), (121.57) 121.20, 80.36, (80.16), 59.23 (59.14), 52.28 (52.21), (46.69) 46.50, 31.06 (30.01), 28.45, (24.60) 23.77.

The data are in full agreement with those previously published in the literature².

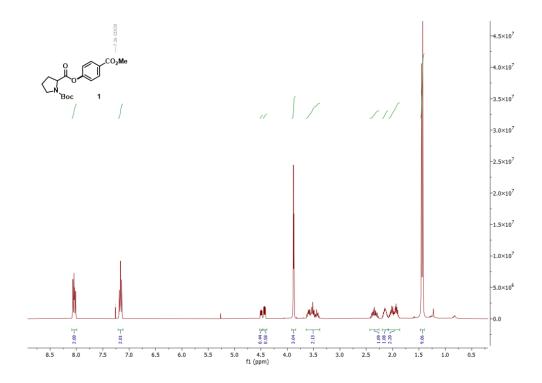


Fig. S22 ¹H-NMR spectrum of compound 1 (CDCl₃, 400 MHz).

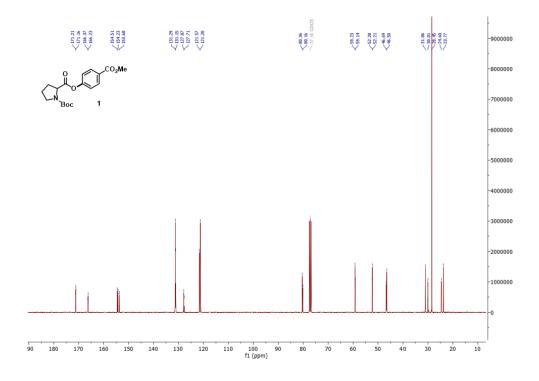


Fig. S23 ¹³C-NMR spectrum of compound **1** (CDCl₃, 151 MHz).

Table S5 Screening of **CD1**/TiO₂ nanocomposites prepared using different **CD1**-to-TiO₂ P25 mass ratios for the metallaphotocatalytic C-O arlyation of Boc-Pro-OH with methyl 4-iodobenzoate.

Entry	CD1-to-TiO ₂ P25 mass ratio ^[a]	1 [%] ^[b]	1 [%] ^[c]
1	1:20	52	8
2	1:10	64	10
3	1:4	80	19
4	1:1	83	22
5	$1:1^{[d]}$	84	-
6	4:1	70	14

[a] CD-to-TiO₂ P25 mass ratio used for the preparation of the nanocomposite. Reaction conditions: methyl 4-iodobenzoate (285.4 μ mol), Boc-Pro-OH (190.3 μ mol), NiCl₂·6H₂O (19.0 μ mol) and dcbpy (19.0 μ mol) in DMSO (anhydrous, 3 mL), BIPA (570.8 μ mol), **CD1**/TiO₂, 24 h. NMR yields were determined by ¹H-NMR using 1,3,5-trimethoxybenzene as internal standard. [b] 440 nm LED lamp (50% power). [c] 525 nm LED lamp (200% power). [d] **CD1**/TiO₂ stored at room temperature for 26 weeks. deg. = degassed.

Table S6 Screening of different amount of **CD1**/TiO₂ nanocomposites for the metallaphotocatalytic C-O arylation of Boc-Pro-OH with methyl 4-iodobenzoate.

Entry	CD1/TiO ₂ / mg	1 [%]
1	20	85
2	10	70
3	5	63
4	1	49
5	0	7

Reaction conditions: methyl 4-iodobenzoate (285.4 µmol), Boc-Pro-OH (190.3 µmol), NiCl₂· $6H_2O$ (19.0 µmol) and dcbpy (19.0 µmol) in DMSO (anhydrous, 3 mL), BIPA (570.8 µmol), CD1/TiO₂, 440 nm LED lamp (50% power), 24 h. NMR yields were determined by 1H -NMR using 1,3,5-trimethoxybenzene as internal standard. deg. = degassed.

6.2. C-O arylation of MeOH

An oven-dried vial (19 x 100 mm) equipped with a stir bar was charged with CD1/TiO2 (15.8 mg), methyl 4-bromobenzoate (20.47 mg, 95.2 μ mol, 1.0 equiv), NiBr₂·3H₂O (nickel(II) bromide trihydrate, 2.6 mg, 9.5 μ mol, 10 mol%) and mcbpy (4'-methyl-2,2'-bipyridine-4-carboxylic acid, 2.1 mg, 9.5 μ mol, 10 mol%). Subsequently, MeOH (anhydrous, 1.5 mL) and BIPA (45 μ L, 285.6 μ mol, 3.0 equiv) were added and the vial was sealed with a septum and Parafilm. The reaction mixture was sonicated for 5-10 min followed by stirring for 5 min to obtain a fine dispersion. The mixture was then degassed by bubbling Argon for 10 min. The mixture was irradiated using the 440 nm LED setup with rapid stirring (1400 rpm). After 14 h, one equivalent of 1,3,5-trimethoxybenzene (internal standard, 16 mg, 95.2 μ mol) was added and the mixture was stirred for 5 min. An aliquot of the reaction mixture (~200 μ L) was filtered, diluted with DMSO-d₆ and subjected to ¹H-NMR analysis. The product was identified by spiking the crude reaction mixture with a pure sample of the desired product.

The data are in full agreement with those previously published in the literature.³

Table S7 C-O arylation of methanol using CD1/TiO₂ and the 440 nm LED setup.

Entry	Conversion [%] ^[a]	2 [%]
1	quant.	93

Reaction conditions: methyl 4-bromobenzoate (95.2 μ mol), NiBr $_2$ ·3H $_2$ O (9.5 μ mol), mcbpy (9.5 μ mol) and BIPA (285.6 μ mol) in MeOH (1.5 mL), CD1/TiO $_2$ (15.8 mg), 440 nm LED lamp (100% power), 14 h. NMR yields determined by 1 H-NMR using 1,3,5-trimethoxybenzene as internal standard. [a] Conversion of methyl 4-bromobenzoate determined by 1 H-NMR using 1,3,5-trimethoxybenzene as internal standard. quant. = quantitative.

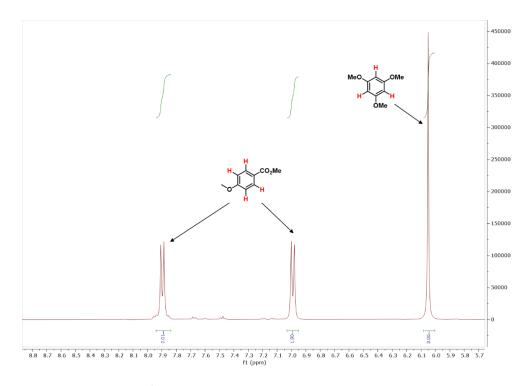


Fig. S24 Representative ¹H-NMR spectrum of a crude reaction mixture for determining NMR yields in the C-O arylation of methanol (DMSO-d₆, 400 MHz).

6.3. C-S arylation of methyl 3-mercaptopropionate

An oven-dried vial (19 x 100 mm) equipped with a stir bar was charged with **CD1**/TiO₂ (31.6 mg), methyl 4-iodobenzoate (49.87 mg, 190.3 μ mol, 1.0 equiv), methyl 3-mercaptopropionate (45.2 μ L, 380.5 μ mol, 2.0 equiv), NiBr₂·3H₂O (5.2 mg, 19 μ mol, 10 mol%) and mcbpy (4.2 mg, 19 μ mol, 10 mol%). Subsequently, MeCN (anhydrous, 3 mL) and BIPA (150.8 μ L, 951.5 μ mol, 5.0 equiv) were added and the vial was sealed with a septum and Parafilm. The reaction mixture was sonicated for 5-10 min followed by stirring for 5 min to obtain a fine dispersion. The mixture was then degassed by bubbling Argon for 10 min. The mixture was irradiated using the 440 nm LED setup with rapid stirring (1400 rpm). After 17 h, one equivalent of 1,3,5-trimethoxybenzene (internal standard, 32 mg, 190.3 μ mol) was added and the mixture was stirred for 5 min. An aliquot of the reaction mixture (~200 μ L) was filtered, diluted with DMSO-d₆ and subjected to ¹H-NMR analysis. The product was identified by spiking the crude reaction mixture with a pure sample of the desired product.

The data are in full agreement with those previously published in the literature.³

Table S8 C-S arylation of methyl 3-mercaptopropionate using **CD1**/TiO₂ and the 440 nm LED setup.

Entry	Conversion [%] ^[a]	3 [%]
1	quant.	98

Reaction conditions: methyl 4-iodobenzoate (190.3 µmol), methyl 3-mercaptopropionate (380.5 µmol), NiBr $_2\cdot 3H_2O$ (19.0 µmol), mcbpy (19.0 µmol) and BIPA (951.5 µmol) in MeCN (3 mL), CD1/TiO $_2$ (31.6 mg), 440 nm LED lamp (50% power), 17 h. NMR yields determined by $^1\text{H-NMR}$ using 1,3,5-trimethoxybenzene as internal standard. [a] Conversion of methyl 4-iodobenzoate determined by $^1\text{H-NMR}$ using 1,3,5-trimethoxybenzene as internal standard. deg. = degassed. quant. = quantitative.

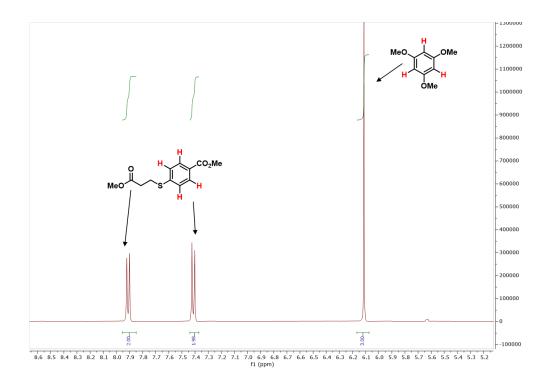


Fig. S25 Representative ¹H-NMR spectrum of a crude reaction mixture for determining NMR yields in the C-S arylation of methyl 3-mercaptopropionate (DMSO-d₆, 400 MHz).

6.4. C-S arylation of sodium *p*-toluensulfinate

An oven-dried vial (19 x 100 mm) equipped with a stir bar was charged with **CD1**/TiO₂ (31.6 mg), 4-iodobenzotrifluoride (14.7 μ L, 100 μ mol, 1.0 equiv), sodium *p*-toluensulfinate (38.8 mg, 200 μ mol, 2.0 equiv), NiCl₂·glyme (nickel(II) chloride ethylene glycol dimethyl ether complex, 2.2 mg, 10 μ mol, 10 mol%) and mcbpy (2.1 mg, 10 μ mol, 10 mol%). Subsequently, DMAc (dimethylacetamide, anhydrous, 2 mL) was added and the vial was sealed with a septum and Parafilm. The reaction mixture was sonicated for 5-10 min followed by stirring for 5 min to obtain a fine dispersion. The mixture was then degassed by bubbling Argon for 10 min. The mixture was irradiated using the 440 nm LED setup with rapid stirring (1400 rpm). After 72 h, one equivalent of 1,3,5-trimethoxybenzene (internal standard, 16.8 mg, 100 μ mol) was added and the mixture was stirred for 5 min. An aliquot of the reaction mixture (~200 μ L) was filtered, diluted with DMSO-d₆ and subjected to ¹H-NMR analysis. The product was identified by spiking the crude reaction mixture with a pure sample of the desired product.

The data are in full agreement with those previously published in the literature.⁴

Table S9 C-S arylation of sodium p-toluensulfinate using **CD1**/TiO₂ and the 440 nm LED setup.

Entry	Conversion [%] ^[a]	4 [%]
1	quant.	55

Reaction conditions: 4-iodobenzotrifluoride (100 µmol), sodium p-toluensulfinate (200 µmol), NiCl $_2$ ·glyme (10 µmol) and mcbpy (10 µmol) in DMAc (2 mL), CD1/TiO $_2$ (31.6 mg), 440 nm blue LED lamp (100% power), 72 h. NMR yields determined by 1 H-NMR using 1,3,5-trimethoxybenzene as internal standard. [a] Conversion of 4-iodobenzotrifluoride determined by 1 H-NMR using 1,3,5-trimethoxybenzene as internal standard. deg. = degassed. quant. = quantitative.

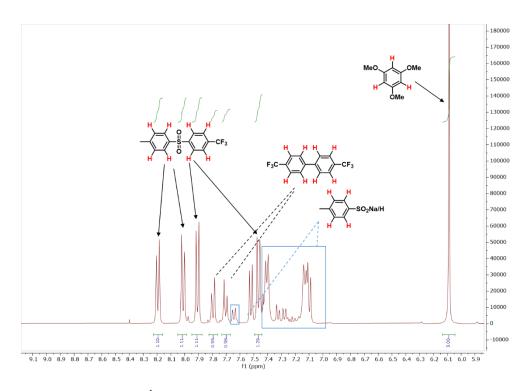


Fig. S26 Representative 1 H-NMR spectrum of a crude reaction mixture for determining NMR yields in the C-S arylation of sodium p-toluensulfinate (DMSO-d₆, 400 MHz).

6.5. C-N arylation of *p*-toluensulfonamide

Entry

An oven-dried vial (19 x 100 mm) equipped with a stir bar was charged with CD1/TiO2 (31.6 mg), 4-iodobenzotrifluoride (14.7 μ L, 100 μ mol, 1.0 equiv), *p*-toluensulfonamide (34.2 mg, 200 μ mol, 2.0 equiv), NiBr₂·3H₂O (2.7 mg, 10 μ mol, 10 mol%), mcbpy (2.1 mg, 10 μ mol, 10 mol%) and DBU (1,8-diazabicyclo[5.4.0]undec-7-en, 22.4 μ L, 150 μ mol, 1.5 equiv). Subsequently, DMSO (anhydrous, 2 mL) was added and the vial was sealed with a septum and Parafilm. The reaction mixture was sonicated for 5-10 min followed by stirring for 5 min to obtain a fine dispersion. The mixture was then degassed by bubbling Argon for 10 min. The mixture was irradiated using the 440 nm LED setup with rapid stirring (1400 rpm). After 14 h, one equivalent of 1,3,5-trimethoxybenzene (internal standard, 16.8 mg, 100 μ mol) was added and the mixture was stirred for 5 min. An aliquot of the reaction mixture (~200 μ L) was filtered, diluted with DMSO-d₆ and subjected to ¹H-NMR analysis.The product was identified by spiking the crude reaction mixture with a pure sample of the desired product.

The data are in full agreement with those previously published in the literature.⁵

Table S10 C-N arylation of p-toluensulfonamide using **CD1**/TiO₂ and the 440 nm LED setup.

1	quant.	90		
Reaction conditions:	4-iodobenzotrifluoride	(100 μmol), p-		
toluensulfonamide (20	0 μmol), NiBr ₂ ·3H ₂ O (10	μmol), mcbpy (10 μmol)		
and DBU (150 µmol) in	DMSO (2 mL), CD1/TiO	O ₂ (31.6 mg), 440 nm blue		
LED lamp (100% power	c), 14 h. NMR yields dete	rmined by ¹ H-NMR using		
1,3,5-trimethoxybenzene as internal standard. [a] Conversion of 4-				
iodobenzotrifluoride determined by ¹ H-NMR using 1,3,5-trimethoxybenzene				
as internal standard. deg. = degassed. quant. = quantitative.				

Conversion [%][a]

5 [%]

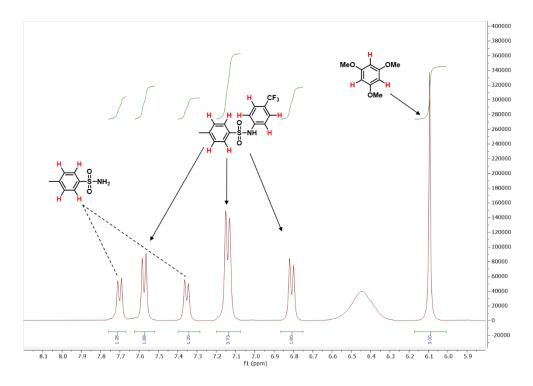


Fig. S27 Representative ¹H-NMR spectrum of a crude reaction mixture for determining NMR yields in the C-N arylation of *p*-toluensulfonamide (DMSO-d₆, 400 MHz).

7. Photobleaching experiments

The photocatalyst (**CD1**/TiO₂, 31.6 mg; Fluo (fluorescein)/TiO₂, 0.54 mg of Fluo and 30 mg of TiO₂) was added to an oven-dried glass vial equipped with a stir bar.² DMSO (1.5 mL) was added and the reaction mixture was stirred and sonicated for 10 min to obtain a fine dispersion. The vial equipped with an air balloon was then transferred into a dark fume hood and irradiated with a blue LED lamp (440 nm LED lamp, 50% power) at room temperature. After the respective time, Boc-Pro-OH (41.0 mg, 190.3 μ mol), methyl 4-iodobenzoate (74.8 mg, 285.4 μ mol), a DMSO solution (1.5 mL) of dcbpy (4.7 mg, 19.0 μ mol), NiCl₂·6H₂O (4.5 mg, 19.0 μ mol), and BIPA (90.5 μ L, 570.8 μ mol) were added. The glass vial was sealed with a septum and Parafilm. The reaction mixture was stirred and sonicated for 10 min to obtain a fine dispersion and subsequently degassed with Argon for 10 min. The vial was then irradiated with a blue LED lamp (440 nm LED lamp, 50% power) at room temperature for 24 h. 1,3,5-Trimethoxybenzene (32.0 mg, 190.3 μ mol) was added as internal standard to determine NMR yields. An aliquot of the resulting mixture (~250 μ L) was filtered through a syringe filter, diluted with DMSO-d₆ (~250 μ L) and subjected to ¹H-NMR analysis.

8. Recycling experiments

Boc-Pro-OH (41.0 mg, 190.3 μ mol), methyl 4-iodobenzoate (74.8 mg, 285.4 μ mol) and CD1/TiO2 nanocomposite (31.6 mg) were added to an oven-dried glass vial equipped with a stir bar. Subsequently, a DMSO solution (3 mL) of dcbpy (4.7 mg, 19.0 μ mol), NiCl2·6H2O (4.5 mg, 19.0 μ mol), and BIPA (90.5 μ L, 570.8 μ mol) were added. The glass vial was sealed with a septum and Parafilm. The reaction mixture was stirred and sonicated for 10 min to obtain a fine dispersion and subsequently degassed with Argon for 10 min. The vial was then irradiated with a blue LED lamp (440 nm LED lamp, 50% power) at room temperature. After 24 h, the reaction mixture was centrifuged and washed twice with DMSO (3 mL). The remaining nanocomposite was lyophilized overnight and reused in the next reaction. For the controlled studies, NiCl2·6H2O (4.5 mg, 19.0 μ mol) or NiCl2·6H2O (4.5 mg, 19.0 μ mol)/dcbpy (4.7 mg, 19.0 μ mol) were added to the new reaction mixture.

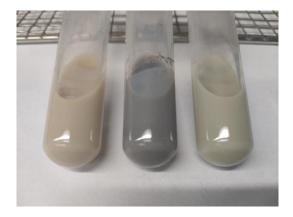


Fig. S28 Photograph of the reaction mixture (+ none group, left; + NiCl₂·6H₂O group, middle; + NiCl₂·6H₂O/dcbpy, right) after C-O cross-coupling reaction (cycle 4).

9. References

- 1. S. A. Hill, D. Benito-Alifonso, S. A. Davis, D. J. Morgan, M. Berry and M. C. Galan, *Sci. Rep.*, 2018, **8**, 12234.
- 2. S. Reischauer, V. Strauss and B. Pieber, *ACS Catal.*, 2020, **10**, 13269.
- 3. C. Cavedon, A. Madani, P. H. Seeberger and B. Pieber, *Org. Lett.*, 2019, **21**, 5331.
- 4. H. Yue, C. Zhu and M. Rueping, *Angew. Chem. Int. Ed.*, 2018, **57**, 1371.
- 5. F. Ye, F. Berger, H. Jia, J. Ford, A. Wortman, J. Börgel, C. Genicot and T. Ritter, *Angew. Chem. Int. Ed.*, 2019, **58**, 14615.