

Supplementary Information

Lipids as Versatile Solvents for Chemical Synthesis

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General considerations

Commercially available starting materials, reagents, catalysts and anhydrous and degassed solvents were used without further purification. Flash column chromatography was performed with Merck silica gel 60 (230-400 mesh). The solvents for column chromatography were distilled before use (in case of technical solvents). Thin layer chromatography was carried out using Merck TLC Silica gel 60 F₂₅₄ and visualized by short-wavelength ultraviolet light or by treatment with potassium permanganate (KMnO₄) stain. ¹H, ¹³C and ¹⁹F NMR spectra were recorded on a Bruker Avance 400 MHz at 20 °C. All ¹H NMR spectra are reported in parts per million (ppm) downfield of TMS and were measured relative to the signal for CHCl₃ (7.26 ppm). All ¹³C NMR spectra were reported in ppm relative to residual CDCl₃ (77.20 ppm) and were obtained with ¹H decoupling. Coupling constants, *J*, are reported in Hertz (Hz). High-resolution mass spectra (HRMS) were recorded from methanol solutions on an LTQ Orbitrap XL (Thermo Scientific) either in negative or in positive electrospray ionization (ESI) mode.

Triglycerides, phospholipids and related lipids possess a varying number of internal double bonds. The fact that we could develop highly efficient methodologies for a range of different cross-coupling reactions in vegetable oils, avoiding side processes involving the solvent (including the Heck coupling), is intriguing. The Heck cross-coupling reaction is known to be quite sensitive to steric hindrance, which can be one of the reasons for the efficiency of vegetable oils and related systems as solvents. Thus, control experiments with internal olefins or substituted styrenes, using the best conditions for Heck coupling, provided partial recovery of starting materials and reductive homocoupling of the aryl halide (Scheme 4, **11i,j**).

The solvent was not affected in a series of control experiments, where the best conditions for cross-couplings described above were run in rapeseed oil in the presence of only 3,5-bis(trifluoromethyl)bromobenzene (**2a**). In all cases, we observed partial recovery of the starting aryl halide alongside the biaryl formed from the reductive homocoupling of the aryl halide. ¹H, ¹³C and ¹⁹F NMR spectra of the solvent before and after the control experiments were identical. As lipids/oils consist of esters, decomposition via saponification is another possible side reaction. In our work saponification was observed only in the presence of water or strong bases such as alkoxides. Considerable amount of saponification is easily detectable, as the liquid phase turns into a wax-like solid when a reaction mixture is cooled. This phenomenon was never seen for anhydrous conditions or bases like carbonates, fluorides, acetates and amines, which are commonly used for cross-coupling reactions, including the present work.

The following oils were bought from Sigma Aldrich and are available worldwide: Rapeseed oil from *Brassica rapa* (CAS: 8002-13-9); Sunflower seed oil from *Helianthus annuus* (CAS: 8001-21-6); Soybean oil from *Glycine max* (CAS: 8001-22-7); Fish oil from menhaden (CAS: 8002-50-4).

The rest of vegetable oils and butter were bought from local groceries (Norway) and are available in Nordic countries (for detailed descriptions see Figures S1-S16). Before use, all oils were treated with anhydrous CaCl₂ powder for 24h (butter and coconut oil were melted during the treatment), which was followed by hot filtration to remove CaCl₂ (along with NaCl in case of butter). The resulting oils were

degassed and stored over activated molecular sieves (4Å) under Ar atmosphere (for detailed descriptions see Figures S17-S19).

Natural waxes are commercially available and were bought from the following international suppliers: Carnauba wax No. 1 yellow (Sigma Aldrich, CAS: 8015-86-9); Beeswax (Sigma Aldrich, CAS: 8012-89-3); Lanolin (Alfa Aesar, CAS: 8006-54-0). After the delivery, these waxes were stored under Ar atmosphere.

For the production of waste rapeseed oil, we fried potatoes in virgin rapeseed oil from the brand Coop. Frying was performed under air at 130 °C. After every 2 hours, a portion of waste rapeseed oil was separated and filtered through a short plug of silica gel. Accordingly, we generated four fractions of waste rapeseed oil, which were used for frying potatoes for 2, 4, 6 and 8 hours respectively. Obtained waste rapeseed oils were stored under air and used for Suzuki-Miyaura and Heck cross-couplings without additional treatment. Products obtained in waste rapeseed oils can be isolated using column chromatography. For more details on the production and processing of waste rapeseed oils, see Figures S30-S32.

Monoterpenes used for preparative column chromatography are commercially available and were bought from the following international suppliers: (-)- α -Pinene (Sigma Aldrich, CAS: 7785-26-4, Cat No: W290203-8KG-K); 3-Carene (Sigma Aldrich, CAS: 13466-78-9, Cat No: W382108-4KG); Dipentene (Sigma Aldrich, CAS: 138-86-3, Cat No: 334111-4L); (R)-(+)-Limonene (Sigma Aldrich, CAS: 5989-27-5, Cat No: W263303-1KG-K); γ -Terpinene (Sigma Aldrich, CAS: 99-85-4, Cat No: W355909-1KG-K); α -Terpinene (Sigma Aldrich, CAS: 99-86-5, Cat No: W355801-4KG-K); Terpinolene (Sigma Aldrich, CAS: 586-62-9, Cat No: W304603-4KG-K); Sabinene (Sigma Aldrich, CAS: 3387-41-5, Cat No: W530597-1KG-K); Myrcene (Sigma Aldrich, CAS: 123-35-3, Cat No: W276200-1KG-K); α -Phellandrene (Sigma Aldrich, CAS: 99-83-2, Cat No: W285609-1KG-K). Monoterpenes were evaporated before use.

Description of oils from grocery



Figure S1. Rapeseed oils and rapeseed oil from the brand Askim.

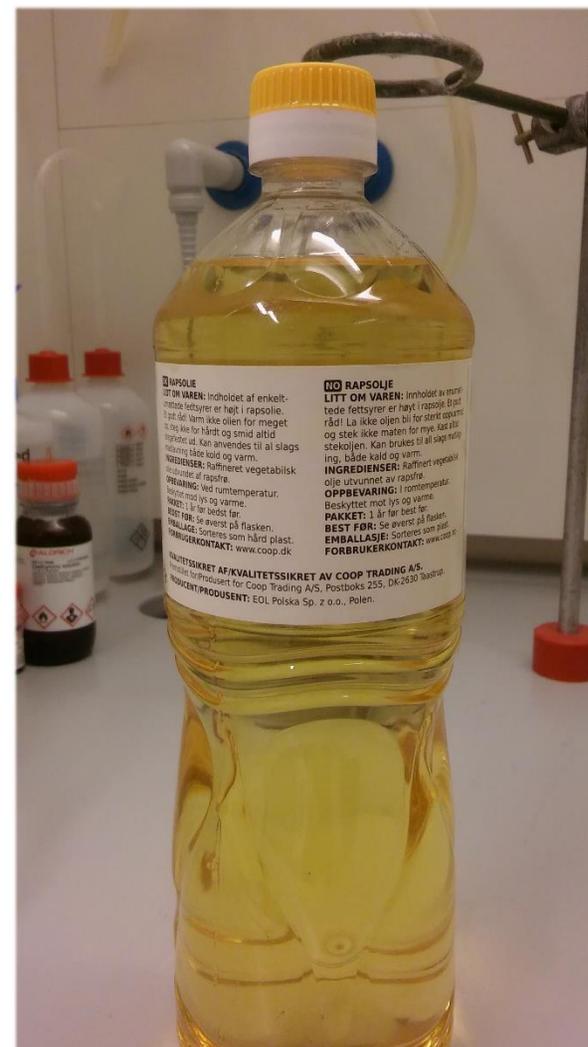
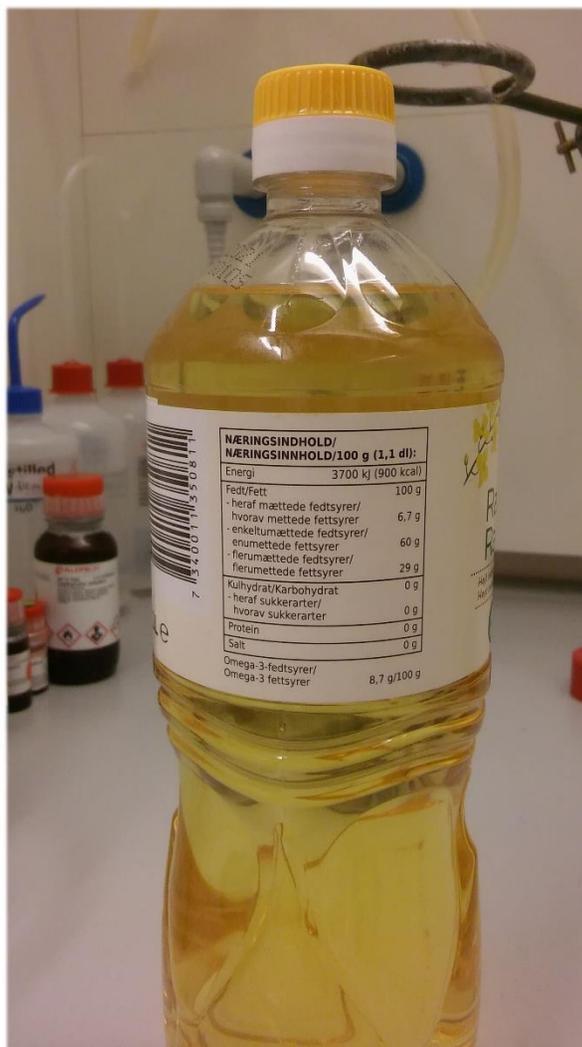


Figure S2. Rapeseed oil from the brand Coop.



Figure S3. Rapeseed oil from the brand Odelia.



Figure S4. Rapeseed oil from the brand Rema.



Figure S6. Sunflower oil.



Figure S7. Olive oil.



Figure S8. Soybean oil.



Figure S9. Corn oil.



Figure S10. Avocado oil.



Næringsinnhold		pr. 100ml
Energi		3400kj / (827kcal)
Fett		91,9g
Hvorav mettet		14g
Karbohydrater		0g
Hvorav sukkerarter		0g
Protein		0g
Salt		0g

INGREDIENSER: Sesamolje
Allergener: Se uthøvet skrift

Oppbevares tørt og kjølig. Unngå direkte kontakt med sollys

250ml e

Best før: Se dato over etiketten



Tappet i England av: AAK (UK) Ltd.
www.internationalcollectionoils.com

Importert av:
Oluf Lorentzen A/S Vestby Norge

Figure S11. Sesame oil.



Figure S12. Rice bran oil.



Figure S13. Mixture of oils consisting of rapeseed oil (60%), sunflower oil (25%) and olive oil (15%).

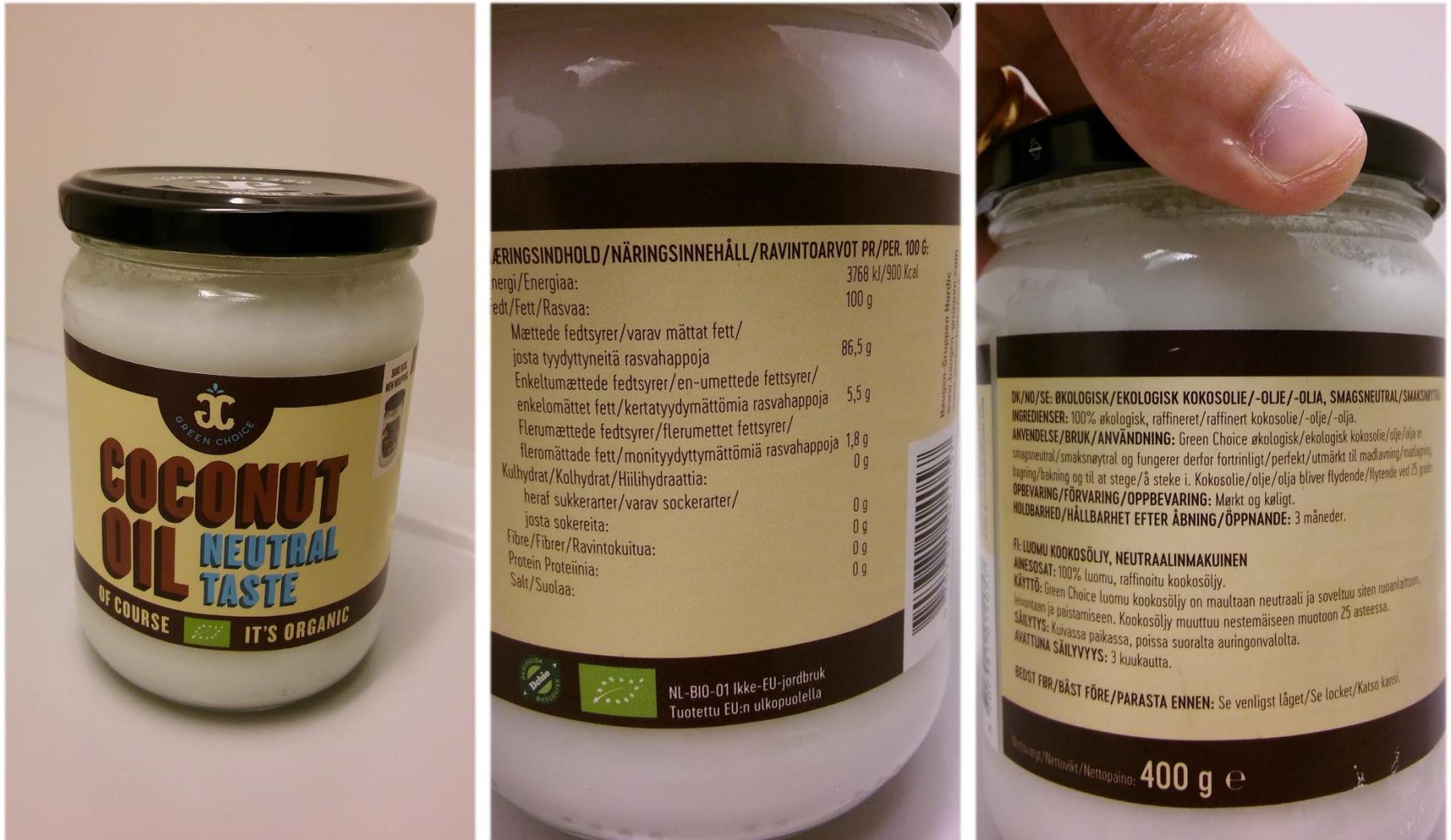


Figure S14. Coconut oil.

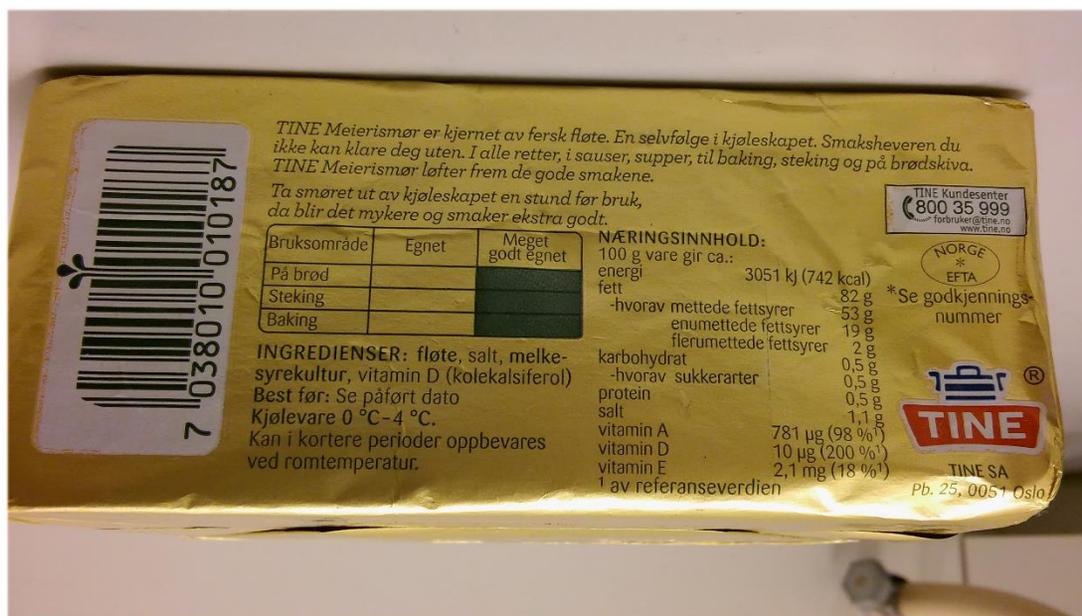


Figure S15. Butter.



Figure S16. Fish oil.

Drying and storage of vegetable oils



Figure S17. Treatment of oils with anhydrous CaCl₂ powder.

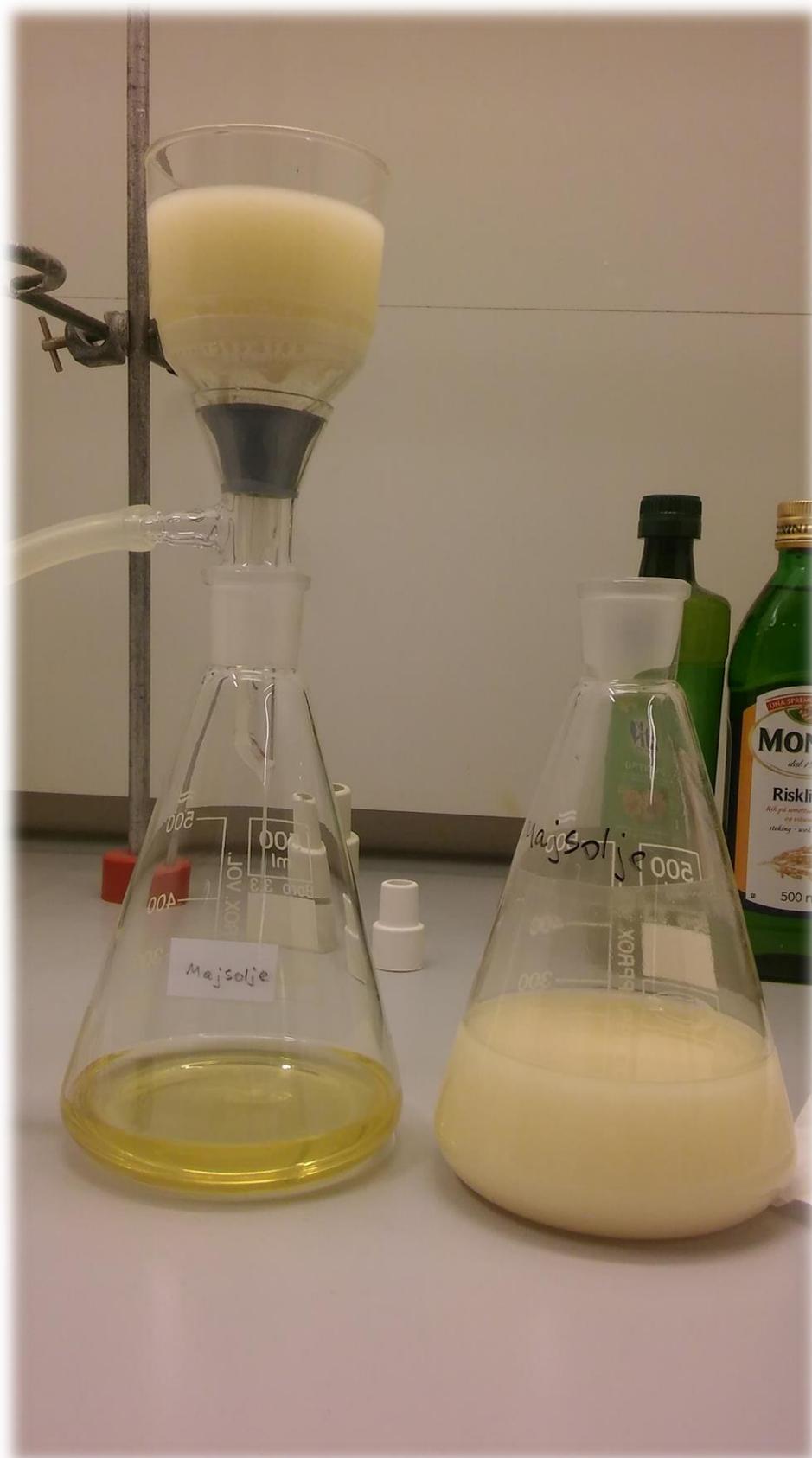


Figure S18. Hot filtration of corn oil from CaCl_2 .

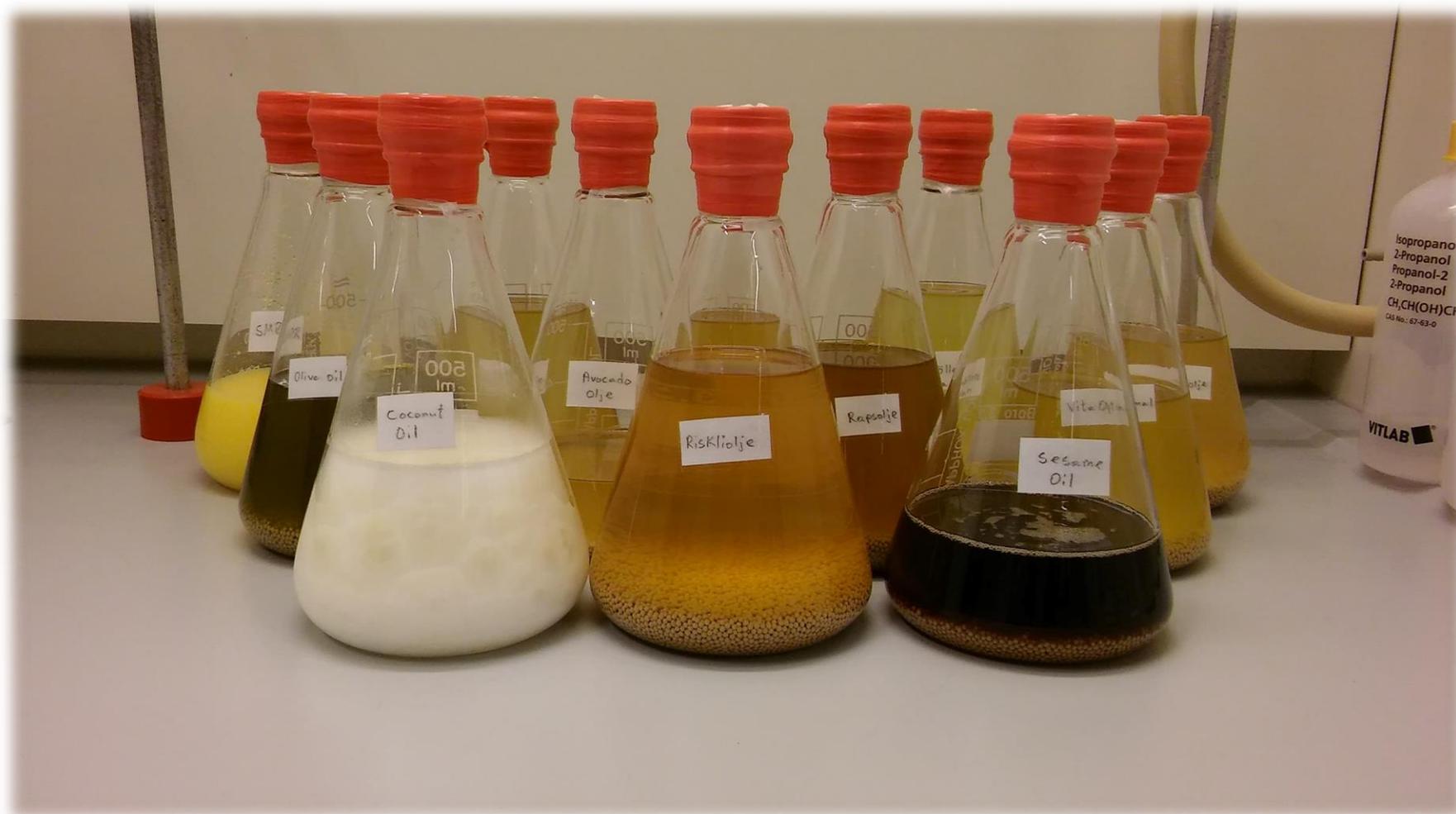
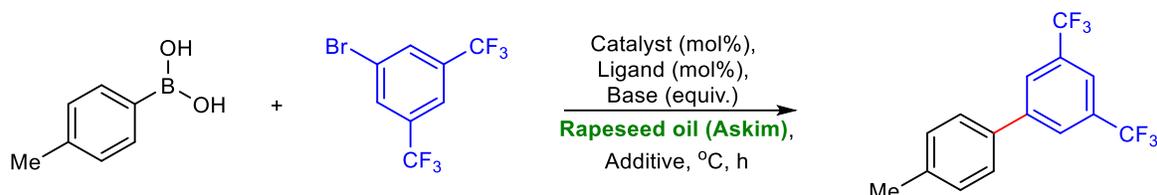


Figure S19. Degassed oils over activated molecular sieves (4Å) and under Ar atmosphere.

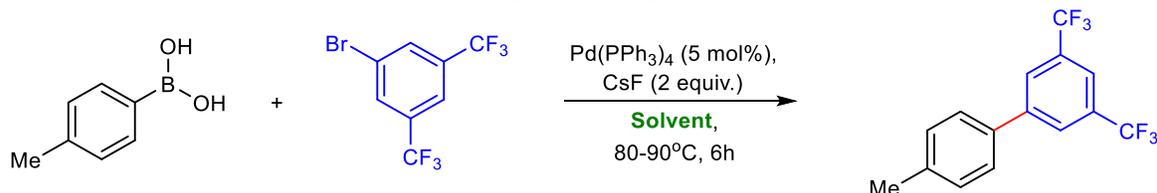
General experimental procedures for optimization of cross-couplings

Optimization of Suzuki-Miyaura coupling in rapeseed oil from Askim. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with 4-tolylboronic acid (1 equiv., 0.368 mmol), corresponding Pd-complex (0.4-5 mol%), ligand (0-12 mol%) and the base (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL), corresponding additive (0-0.3 mL) and 3,5-bis(trifluoromethyl)bromobenzene (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 20-110 °C for 6-24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of solvents for Suzuki-Miyaura coupling. For general setup, see Figures S20-S27.

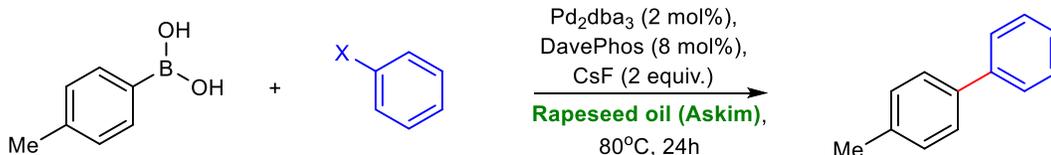


Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with 4-tolylboronic acid (1 equiv., 0.368 mmol), Pd(PPh₃)₄ (5 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added corresponding solvent (2 mL)¹ and 3,5-bis(trifluoromethyl)bromobenzene (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80-90 °C for 6 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL² of CDCl₃ (or CCl₄). In case of 2MeTHF, acetal, dioxane, toluene or DMF, the solvent was evaporated before addition of the internal standard and chloroform. The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

¹ Beeswax and Carnauba wax were weighed inside of glove box like other solids (2g). Lanolin, Butter and coconut oil were melted before addition (80 °C). In general, vegetable oils are viscous liquids. The transfer of vegetable oils from the main container into the reaction vessel with the syringe can be greatly facilitated by heating the oil to 50-80 °C.

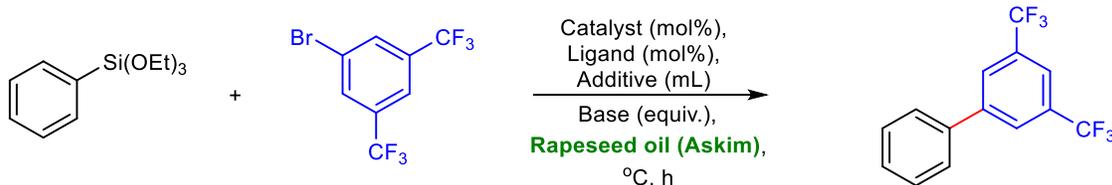
² In case of Beeswax and Carnauba wax, the mixture was diluted with 6 mL of CDCl₃ (CCl₄).

Screening of electrophiles for Suzuki-Miyaura coupling. For general setup, see Figures S20-S27.



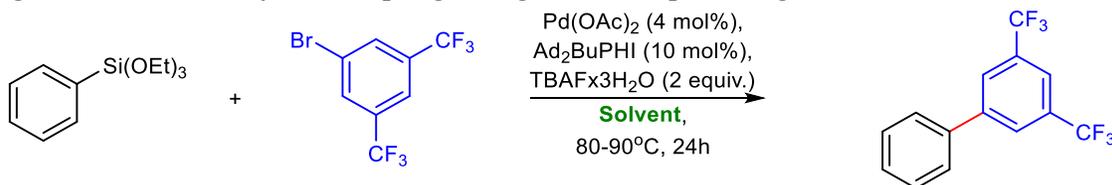
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with 4-tolylboronic acid (1 equiv., 0.368 mmol), Pd₂dba₃ (2 mol%), DavePhos (8 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL) and corresponding aryl halide/sulfonate ester³ (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Optimization of Hiyama coupling in rapeseed oil from Askim. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with corresponding Pd-complex (0.4-4 mol%), ligand (0-10 mol%) and the base (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL), corresponding additive (0-0.2 mL), phenyltriethoxysilane (1 equiv., 0.333 mmol) and 3,5-bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 20-110 °C for 6-24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of solvents for Hiyama coupling. For general setup, see Figures S20-S27.

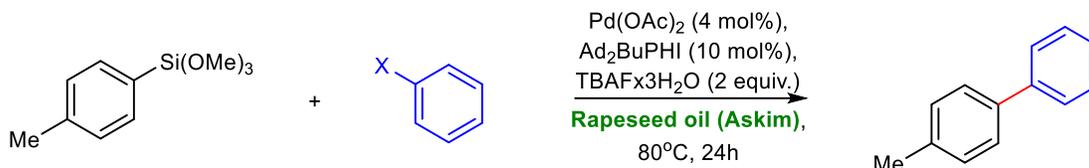


Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd(OAc)₂ (4 mol%), Ad₂BuPHI (10 mol%) and TBAF·3H₂O (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added corresponding solvent (2 mL),¹ phenyltriethoxysilane (1 equiv., 0.333 mmol) and 3,5-

³ Solid electrophiles were weighed in the glove box.

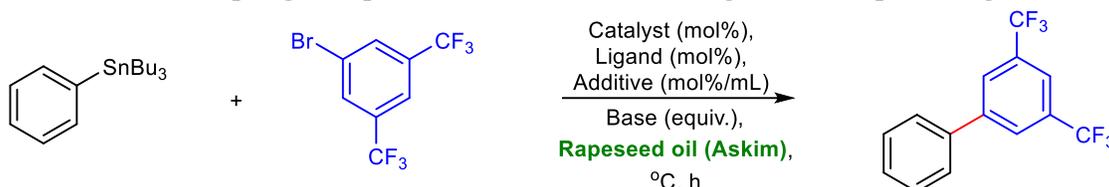
bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80-90 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL² of CDCl₃ (or CCl₄). In case of 2MeTHF, acetal, dioxane, toluene or DMF, the solvent was evaporated before addition of the internal standard and chloroform. The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of electrophiles for Hiyama coupling. For general setup, see Figures S20-S27.



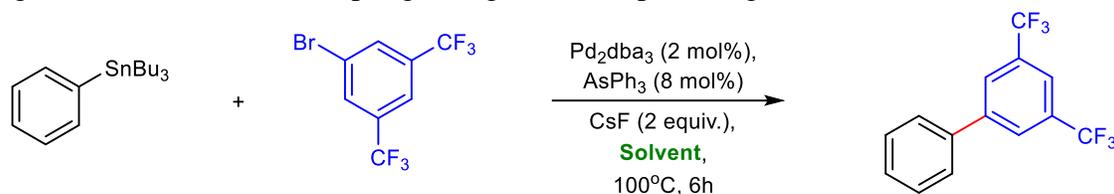
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd(OAc)₂ (4 mol%), Ad₂BuPHI (10 mol%) and TBAFx3H₂O (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL), trimethoxy(*p*-tolyl)silane (1 equiv., 0.471 mmol) and corresponding aryl halide/sulfonate ester³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Optimization of Stille coupling in rapeseed oil from Askim. For general setup, see Figures S20-S27.



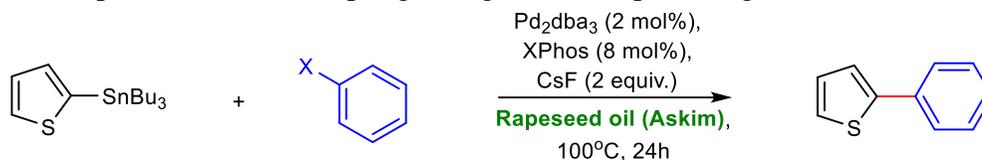
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with corresponding Pd-complex (0.5-2 mol%), ligand (2-8 mol%), CuI (0-4 mol%) and the base (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL), absolute ethanol (0-0.3 mL), tributylphenylstannane (1 equiv., 0.545 mmol) and 3,5-bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 20-100 °C for 6-24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,2,4,5-tetramethylbenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of solvents for Stille coupling. For general setup, see Figures S20-S27.



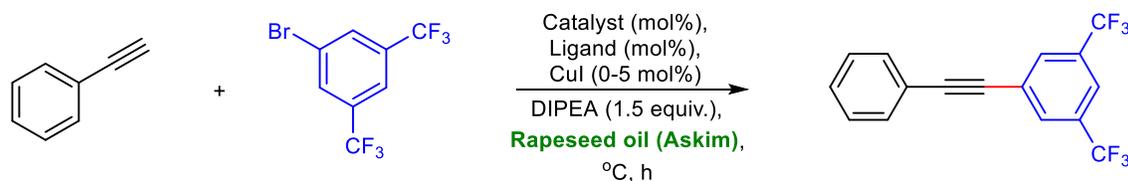
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), AsPh₃ (8 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added corresponding solvent (2 mL),¹ tributylphenylstannane (1 equiv., 0.545 mmol) and 3,5-bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 100 °C for 6 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,2,4,5-tetramethylbenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL² of CDCl₃ (or CCl₄). In case of 2MeTHF, acetal, dioxane, toluene or DMF, the solvent was evaporated before addition of the internal standard and chloroform. The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of electrophiles for Stille coupling. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), XPhos (8 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (2 mL), 2-(tributylstannyl)thiophene (1 equiv., 0.536 mmol) and corresponding aryl halide/sulfonate ester³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 100 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,2,4,5-tetramethylbenzene (1 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

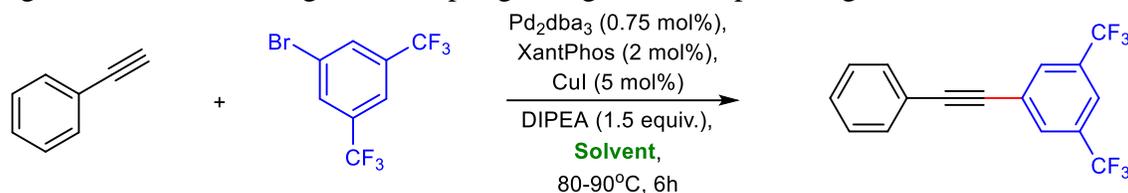
Optimization of Sonogashira coupling in rapeseed oil from Askim. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with corresponding Pd-complex (0.75-3 mol%), ligand (0-6 mol%), CuI (0-5 mol%), phenylacetylene (1 equiv., 0.783 mmol, degassed), 3,5-bis(trifluoromethyl)bromobenzene (1.5 equiv., degassed), rapeseed oil from Askim (2 mL, degassed) and DIPEA (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed

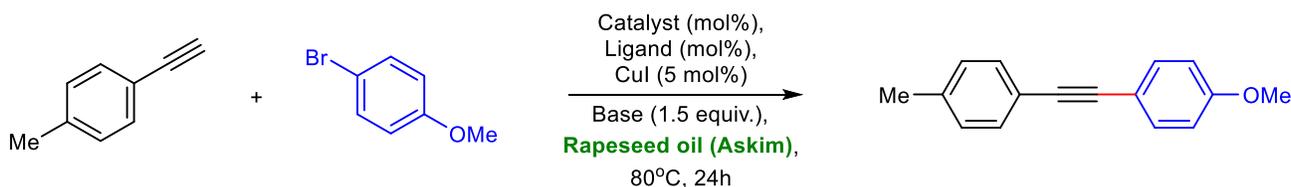
from the glove box and stirred at 20-80 °C for 6-24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (0.5 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of solvents for Sonogashira coupling. For general setup, see Figures S20-S27.



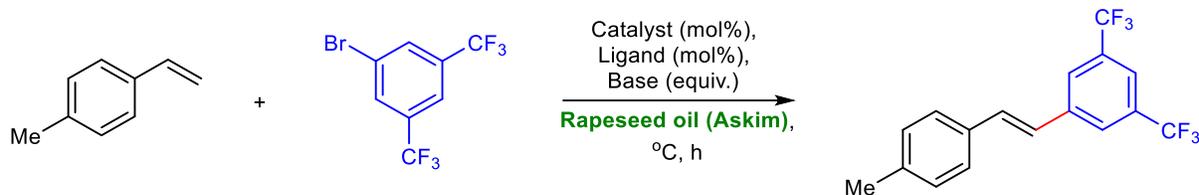
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (0.75 mol%), XantPhos (2 mol%), CuI (5 mol%), phenylacetylene (1 equiv., 0.783 mmol, degassed), 3,5-bis(trifluoromethyl)bromobenzene (1.5 equiv., degassed), corresponding solvent (2 mL, degassed)¹ and DIPEA (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed from the glove box and stirred at 80-90 °C for 6 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (0.5 equiv., internal standard), 20 drops of CHCl₃ and 2 mL² of CDCl₃ (or CCl₄). In case of 2MeTHF, acetal, dioxane, toluene or DMF, the solvent was evaporated before addition of the internal standard and chloroform. The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Optimization of Sonogashira coupling for unactivated aryl halides. For general setup, see Figures S20-S27.



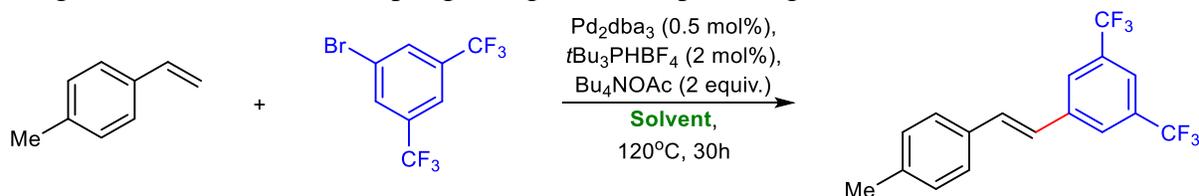
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with corresponding Pd-complex (2-5 mol%), ligand (0-8 mol%), CuI (5 mol%), 4-ethynyltoluene (1 equiv., 0.689 mmol, degassed), 4-bromoanisole (1.5 equiv., degassed), rapeseed oil from Askim (2 mL, degassed) and appropriate base (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed from the glove box and stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of 1,3,5-trimethoxybenzene (0.5 equiv., internal standard), 20 drops of CHCl₃ and 2 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Optimization of Heck coupling in rapeseed oil from Askim. For general setup, see Figures S20-S27.



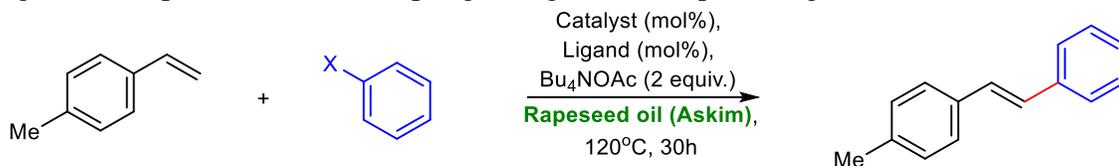
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with corresponding Pd-complex (0.5-5 mol%), ligand (0-12 mol%) and the base (2 equiv.).⁴ The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), 4-methylstyrene (1 equiv., 0.846 mmol) and 3,5-bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80-140 °C for 24-48 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of methyl 3,5-dinitrobenzoate (0.5 equiv., internal standard), 20 drops of CHCl₃ and 3 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of solvents for Heck coupling. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (0.5 mol%), *t*Bu₃PHBF₄ (2 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added corresponding solvent (3 mL),¹ 4-methylstyrene (1 equiv., 0.846 mmol) and 3,5-bis(trifluoromethyl)bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of methyl 3,5-dinitrobenzoate (0.5 equiv., internal standard), 20 drops of CHCl₃ and 3 mL² of CDCl₃ (or CCl₄). In case of 2MeTHF, acetal, dioxane, toluene or DMF, the solvent was evaporated before addition of the internal standard and chloroform. The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Screening of electrophiles for Heck coupling. For general setup, see Figures S20-S27.



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (0.5-2 mol%), corresponding ligand (2-8 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber

⁴ Liquid bases were added outside of the glove box following the addition of rapeseed oil.

septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), 4-methylstyrene (1 equiv., 0.846 mmol) and corresponding aryl halide/sulfonate ester³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was cooled down, which was followed by addition of methyl 3,5-dinitrobenzoate (0.5 equiv., internal standard), 20 drops of CHCl₃ and 3 mL of CDCl₃ (or CCl₄). The resulting mixture was thoroughly shaken for 2 minutes than centrifuged. This was followed by NMR analysis of an aliquot taken from the resulting mixture.

Setup of high-throughput screening

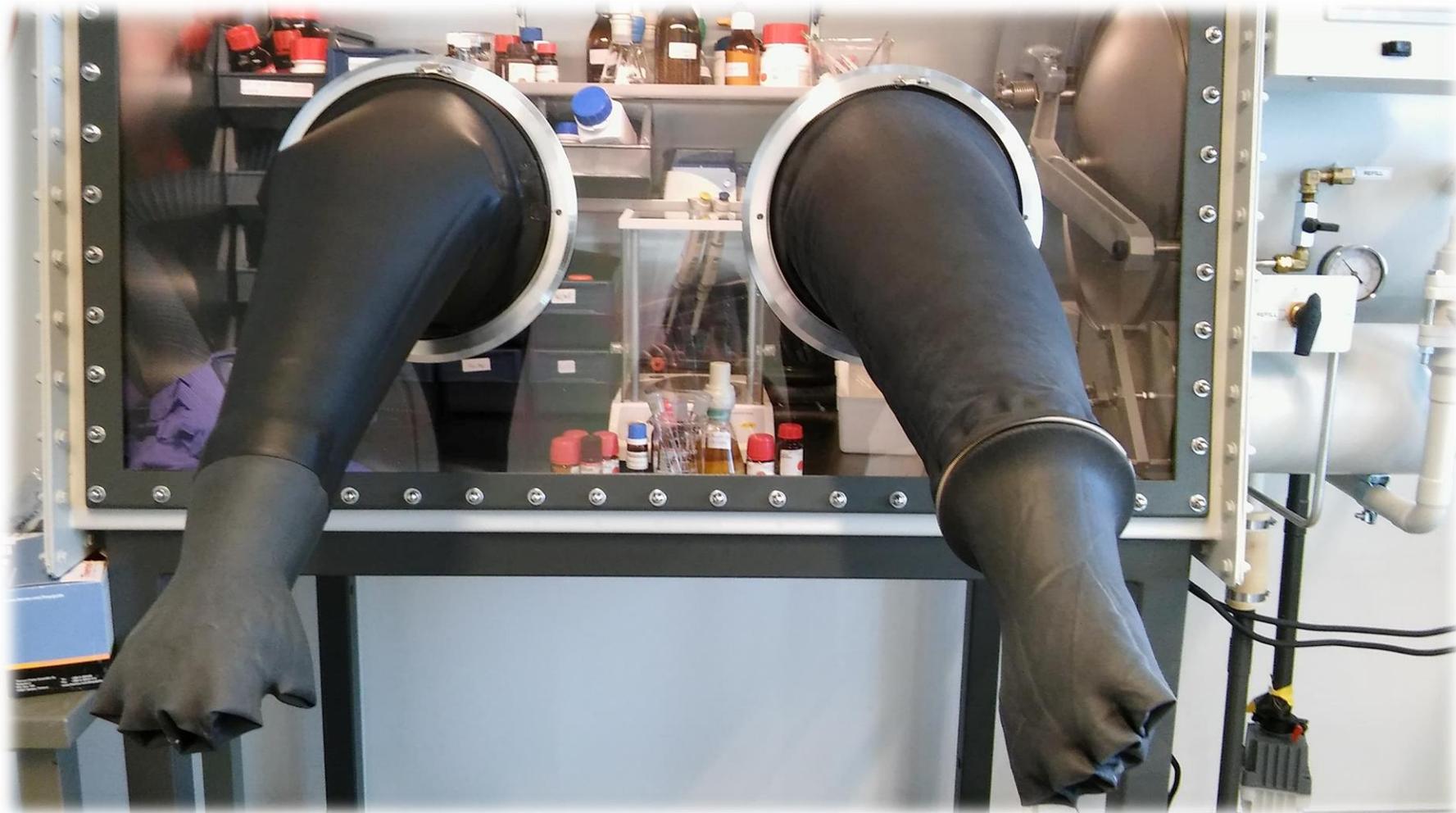


Figure S20. Charging of reaction vials with solid reactants and catalyst in the glove box.

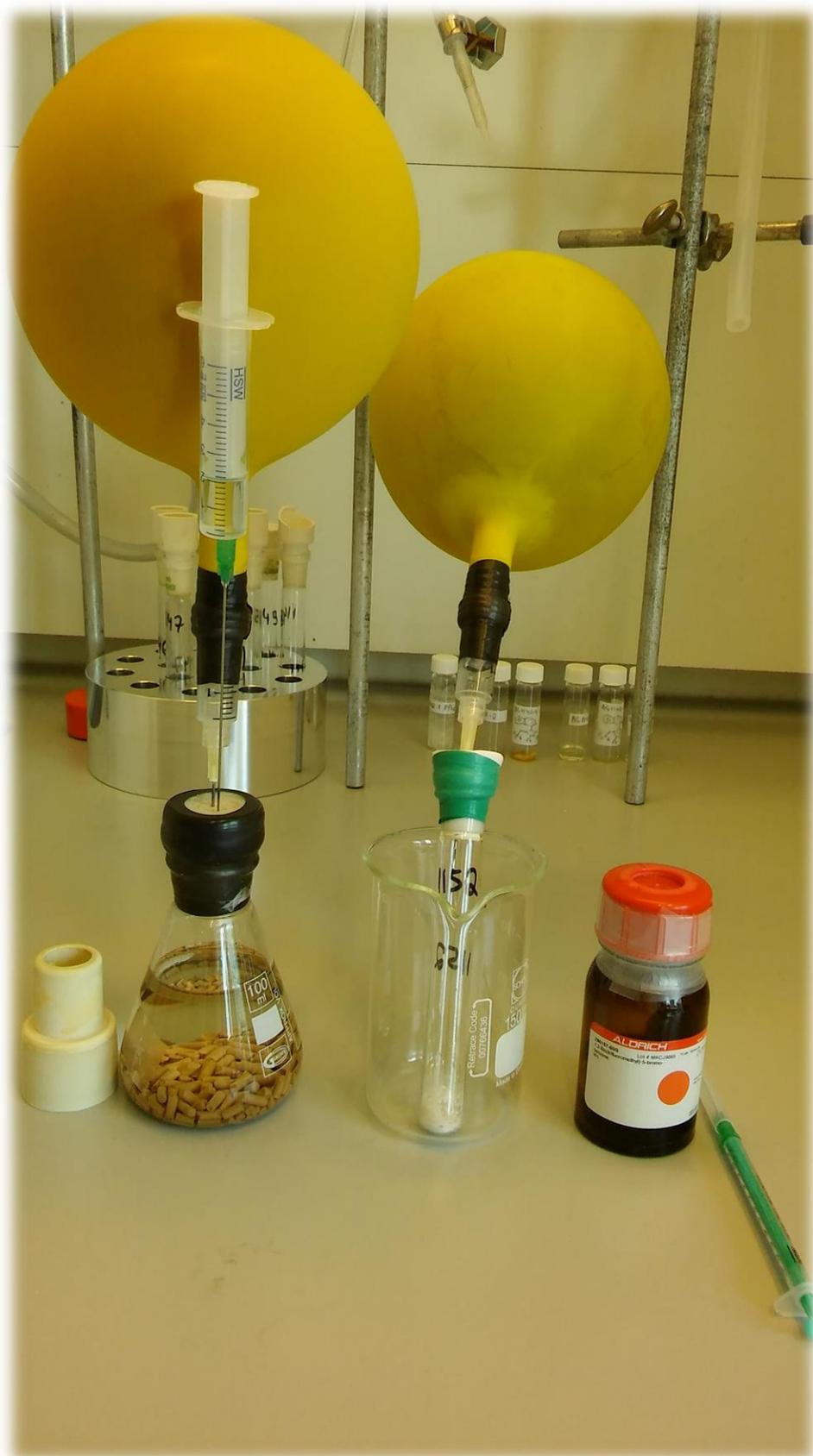


Figure S21. Addition of vegetable oil and liquid reaction components.



Figure S22. Stirring of the reaction mixture at appropriate temperature.



Figure S23. Addition of an internal standard to the reaction mixture at room temperature.



Figure S24. Addition of 20 drops of CHCl_3 and CDCl_3 (or CCl_4).

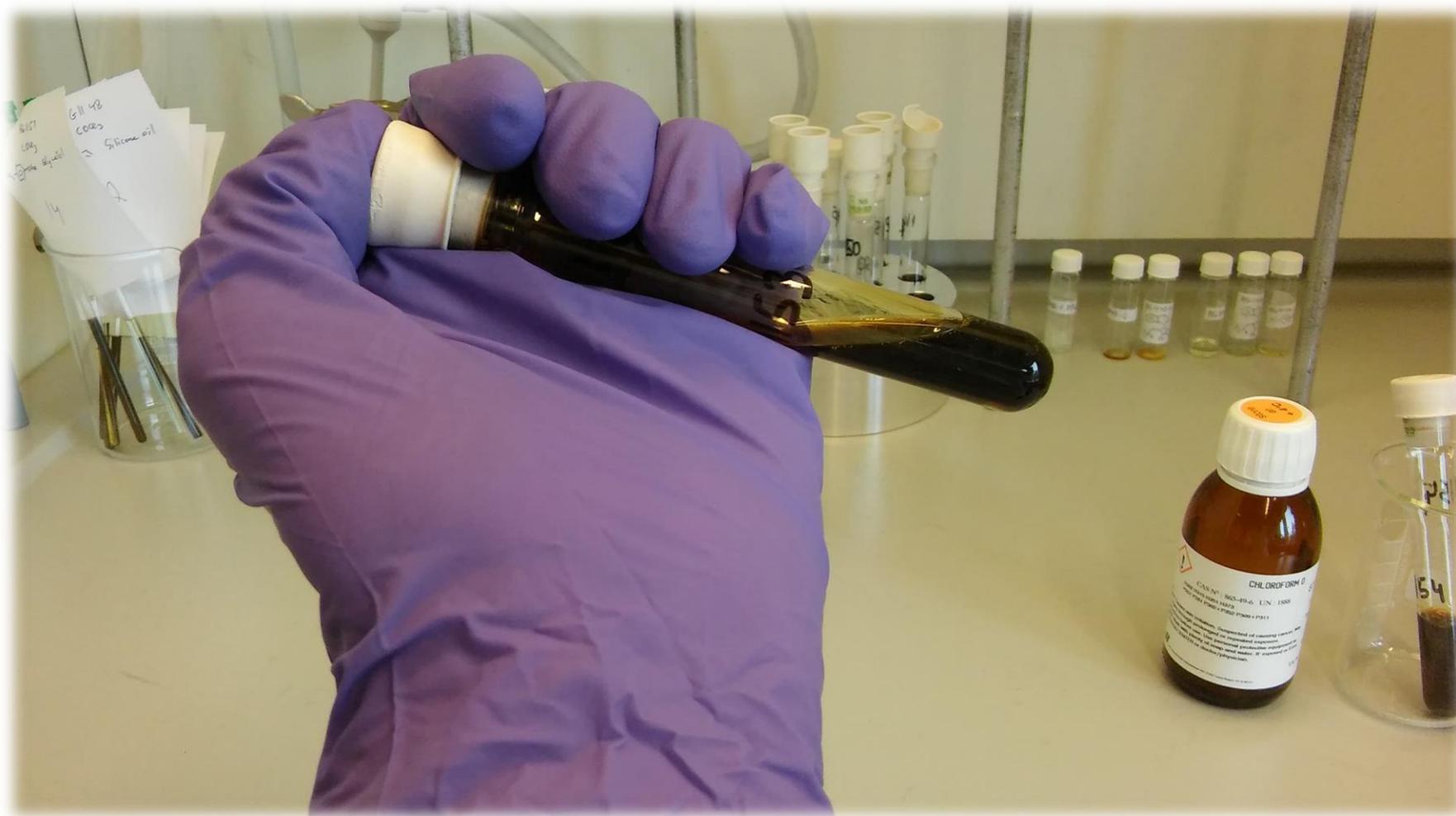


Figure S25. Shaking of the reaction mixture to dissolve all organic matter.



Figure S26. Centrifugation to precipitate inorganic matter.

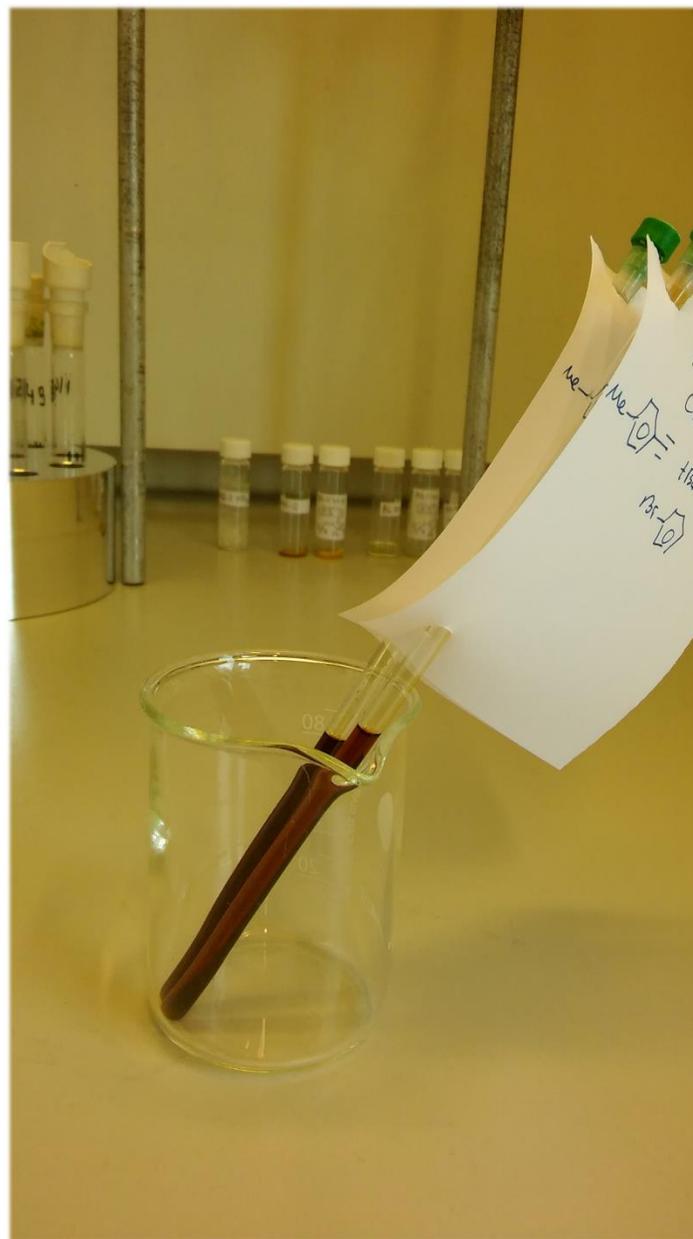


Figure S27. Taking an aliquot for crude ^1H NMR analysis.

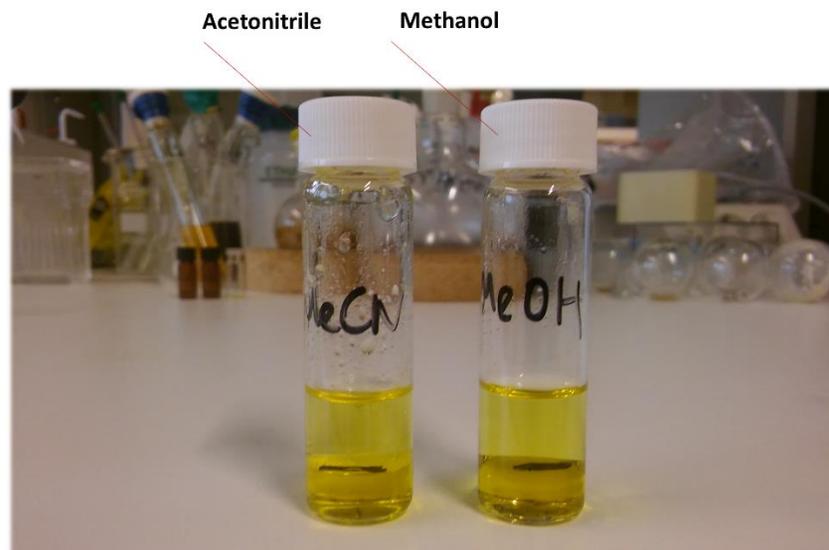
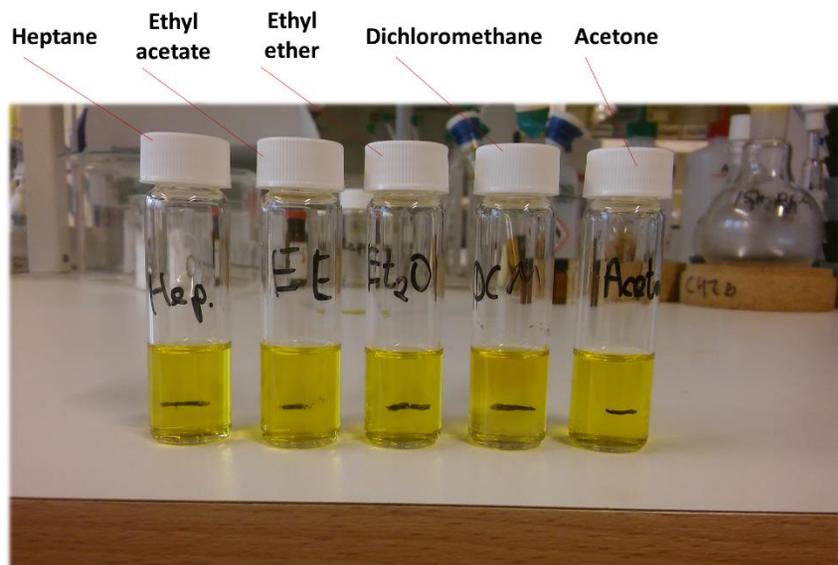
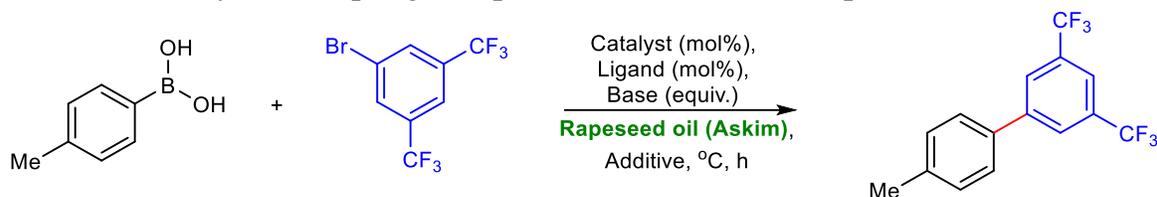
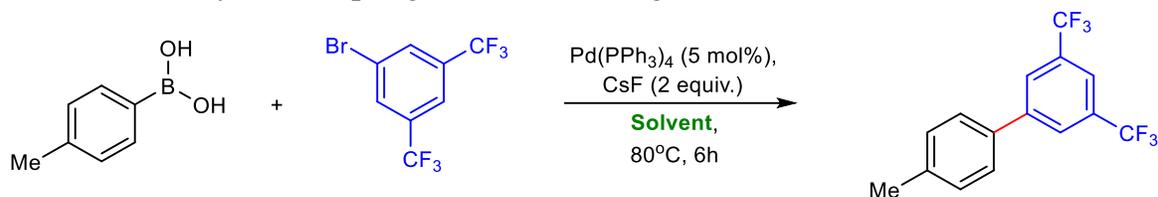


Figure S28. Miscibility of rapeseed oil with common organic solvents.

Table S1. Suzuki-Miyaura coupling in rapeseed oil from Askim: optimization

Entry	Catalyst (mol%)	Ligand (mol%)	Base (equiv.)	Additive (mL)	°C, h	Yield % ^a
1	Pd ₂ dba ₃ (2)	XPhos (8)	K ₂ CO ₃ (2)	-	80, 24	60
2	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (8)	K ₂ CO ₃ (2)	-	80, 24	33
3	Pd ₂ dba ₃ (2)	DavePhos (8)	K ₂ CO ₃ (2)	-	80, 24	82
4	Pd ₂ dba ₃ (2)	DavePhos (8)	CsF (2)	-	80, 24	100
5	Pd ₂ dba ₃ (2)	SPhos (8)	K ₂ CO ₃ (2)	-	80, 24	77
6	Pd ₂ dba ₃ (2)	BrettPhos (8)	K ₂ CO ₃ (2)	-	80, 24	80
7	Pd ₂ dba ₃ (2)	RuPhos (8)	K ₂ CO ₃ (2)	-	80, 24	63
8	XPhos Pd G3 (0.4)	XPhos (0.4)	K ₂ CO ₃ (2)	-	80, 24	63
9	Pd ₂ dba ₃ (2)	XantPhos (4)	K ₂ CO ₃ (2)	-	80, 24	58
10	Pd ₂ dba ₃ (2)	Ad ₂ BuPHI (8)	K ₂ CO ₃ (2)	-	80, 24	37
11	Pd(OAc) ₂ (3)	<i>t</i> Bu ₃ PHBF ₄ (6)	K ₂ CO ₃ (2)	-	80, 24	57
12	[PdCl(allyl)] ₂ (2)	IPrHCl (5)	K ₂ CO ₃ (2)	-	80, 24	8
13	Pd ₂ dba ₃ (2)	AsPh ₃ (8)	K ₂ CO ₃ (2)	-	80, 24	67
14	Pd(OAc) ₂ (3)	P(2-furyl) ₃ (12)	K ₂ CO ₃ (2)	-	80, 24	82
15	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	-	80, 24	90
16	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	-	110, 24	89
17	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	-	20, 24	0
18	Pd(PPh ₃) ₄ (2.5)	-	K ₂ CO ₃ (2)	-	80, 24	78
19	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	EtOH (0.3)	80, 24	100
20	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	EtOH (0.3)	20, 24	100
21	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	EtOH (0.3)	80, 6	100
22	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	EtOH (0.1)	80, 6	89
23	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	EtOH (0.3)	20, 6	34
24	Pd(PPh ₃) ₄ (2.5)	-	K ₂ CO ₃ (2)	EtOH (0.3)	20, 24	48
25	Pd(PPh ₃) ₄ (5)	-	K ₂ CO ₃ (2)	Glycerol (0.3)	80, 24	60
26	Pd(PPh ₃) ₄ (5)	-	K ₃ PO ₄ (2)	-	80, 24	67
27	Pd(PPh ₃) ₄ (5)	-	Bu ₄ NOAc (2)	-	80, 24	0
28	Pd(PPh ₃) ₄ (5)	-	Cs ₂ CO ₃ (2)	-	80, 24	92
29	Pd(PPh ₃) ₄ (5)	-	CsOAc (2)	-	80, 24	83
30	Pd(PPh ₃) ₄ (5)	-	CsF (2)	-	80, 24	100
31	Pd(PPh ₃) ₄ (5)	-	CsF (2)	-	80, 6	100/92 ^b

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield.

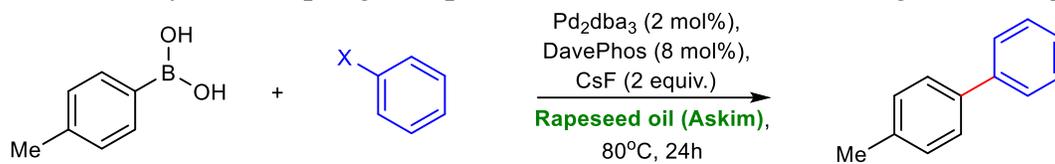
Table S2. Suzuki-Miyaura coupling in oils: screening of solvents

Entry	Solvent (2 mL)	Yield % ^a
1	Triacetin	88
2	Tributylin	100
3	Rapeseed oil (Askim)	100/92 ^b
4	Rapeseed oil (Coop)	87
5	Rapeseed oil (Odelia)	100
6	Rapeseed oil (Rema)	80/100 ^c
7	Rapeseed oil (Anglamark)	100
8	Rapeseed oil (Sigma)	97
9	Sunflower oil	95
10	Sunflower oil (Sigma)	78/100 ^c
11	Olive oil	100
12	Soybean oil	94
13	Soybean oil (Sigma)	88
14	Corn oil	100
15	Avocado oil	89
16	Sesame oil	100
17	Rice bran oil	95
18	Mixture of oils	98
19	Coconut oil	98
20	Butter	100
21	Fish oil	83/100 ^c
22	Fish oil (Sigma)	100
23	Carnauba wax No. 1 yellow	97 ^d
24	Beeswax, refined	22
25	Lanolin	96
26	2MeTHF	91
27	Acetal	98
28	Dioxane	100
29	Toluene	100
30	DMF	86

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield. ^c The reaction was performed in the presence of DavePhos/Pd₂dba₃-based catalytic system.

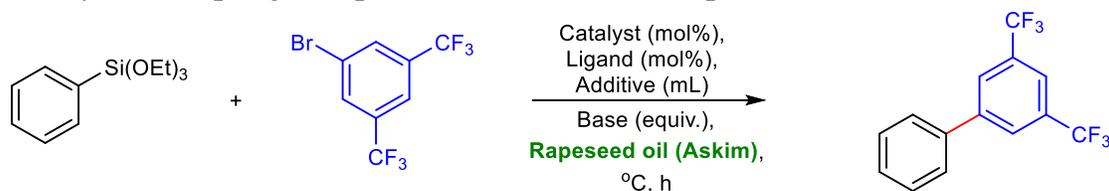
^d The reaction was performed at 90 °C.

Table S3. Suzuki-Miyaura coupling in rapeseed oil from Askim: screening of electrophiles



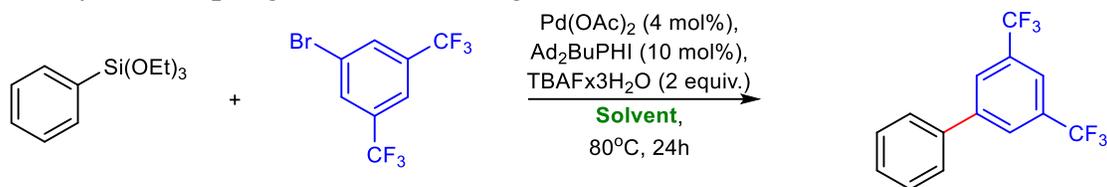
Entry	X	Yield % ^a
1	X = Br	100/99 ^b
2	X = Cl	59/49 ^b
3	X = OTf	100/99 ^b
4	X = OTs	0
5	X = OMs	0

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield.

Table S4. Hiyama coupling in rapeseed oil from Askim: optimization

Entry	Catalyst (mol%)	Ligand (mol%)	Base (2 equiv.)	Additive (mL)	°C, h	Yield % ^a
1	Pd ₂ dba ₃ (2)	XPhos (10)	TBAF _x 3H ₂ O	-	80, 24	72
2	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	-	80, 24	60
3	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	-	80, 24	58 ^b
4	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	MS 4 Å ^c	80, 24	32
5	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (10)	TBAF _x 3H ₂ O	-	80, 24	69
6	Pd ₂ dba ₃ (2)	DavePhos (10)	TBAF _x 3H ₂ O	-	80, 24	65
7	Pd ₂ dba ₃ (2)	SPhos (10)	TBAF _x 3H ₂ O	-	80, 24	80
8	Pd ₂ dba ₃ (2)	BrettPhos (10)	TBAF _x 3H ₂ O	-	80, 24	51
9	Pd ₂ dba ₃ (2)	RuPhos (10)	TBAF _x 3H ₂ O	-	80, 24	60
10	<i>t</i> BuXPhos Pd G3 (0.4)	<i>t</i> BuXPhos (0.4)	TBAF _x 3H ₂ O	-	80, 24	20
11	Pd ₂ dba ₃ (2)	XantPhos (5)	TBAF _x 3H ₂ O	-	80, 24	44
12	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (10)	TBAF _x 3H ₂ O	-	80, 24	57
13	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₂ MePHBF ₄ (10)	TBAF _x 3H ₂ O	-	80, 24	46
14	Pd ₂ dba ₃ (2)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	-	80, 24	93
15	Pd(OAc) ₂ (4)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	-	80, 24	96/79 ^d
16	Pd(OAc) ₂ (4)	DABCO (8)	TBAF _x 3H ₂ O	-	80, 24	62
17	Pd(OAc) ₂ (4)	IPrHCl (4)	TBAF _x 3H ₂ O	-	80, 24	60
18	PdCl ₂ [P(<i>o</i> -Tol) ₃] ₂ (4)	-	TBAF _x 3H ₂ O	-	80, 24	36
19	Pd(OAc) ₂ (4)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	-	110, 24	60
20	Pd(OAc) ₂ (2)	Ad ₂ BuPHI (5)	TBAF _x 3H ₂ O	-	80, 24	89
21	Pd(OAc) ₂ (4)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	EtOH (0.2)	80, 24	97
22	Pd(OAc) ₂ (4)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	EtOH (0.2)	20, 24	0
23	Pd(OAc) ₂ (4)	Ad ₂ BuPHI (10)	TBAF _x 3H ₂ O	EtOH (0.2)	80, 6	90
24	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	EtOH (0.2)	80, 24	99/90 ^d
25	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	EtOH (0.1)	80, 24	89
26	Pd(OAc) ₂ (4)	XPhos (10)	TBAF _x 3H ₂ O	Glycerol (0.2)	80, 24	20
27	Pd(OAc) ₂ (4)	XPhos (10)	CsF	-	80, 24	0
28	Pd(OAc) ₂ (4)	XPhos (10)	CsF/TBAB (0.5)	-	80, 24	9

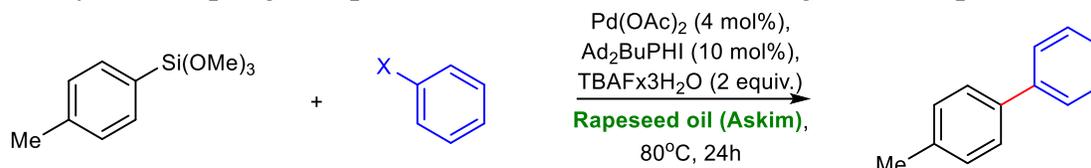
^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Silane used in 2 equiv.. ^c 80 mg. ^d Isolated yield.

Table S5. Hiyama coupling in oils: screening of solvents

Entry	Solvent (2 mL)	Yield % ^a
1	Triacetin	93
2	Tributylin	96
3	Rapeseed oil (Askim)	96/79 ^b
4	Rapeseed oil (Coop)	86
5	Rapeseed oil (Odelia)	93
6	Rapeseed oil (Rema)	86
7	Rapeseed oil (Anglamark)	92
8	Rapeseed oil (Sigma)	86
9	Sunflower oil	86
10	Sunflower oil (Sigma)	72/88 ^c
11	Olive oil	84
12	Soybean oil	93
13	Soybean oil (Sigma)	82
14	Corn oil	93
15	Avocado oil	91
16	Sesame oil	74/92 ^c
17	Rice bran oil	79
18	Mixture of oils	85
19	Coconut oil	78
20	Butter	88
21	Fish oil	88
22	Fish oil (Sigma)	92
23	Carnauba wax No. 1 yellow	100 ^d
24	Beeswax, refined	86
25	Lanolin	79
26	2MeTHF	98
27	Acetal	85
28	Dioxane	88
29	Toluene	91
30	DMF	67

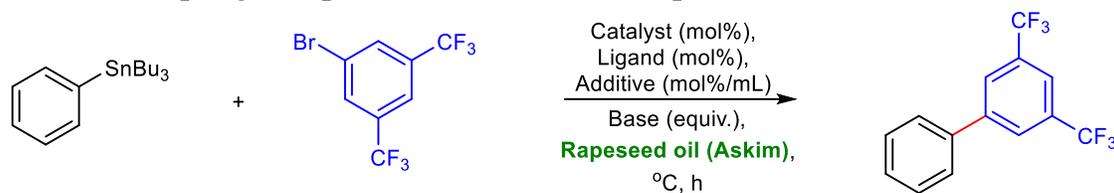
^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield. ^c The reaction was performed in the presence of ethanol (0.2 mL). ^d The reaction was performed at 90 °C.

Table S6. Hiyama coupling in rapeseed oil from Askim: screening of electrophiles



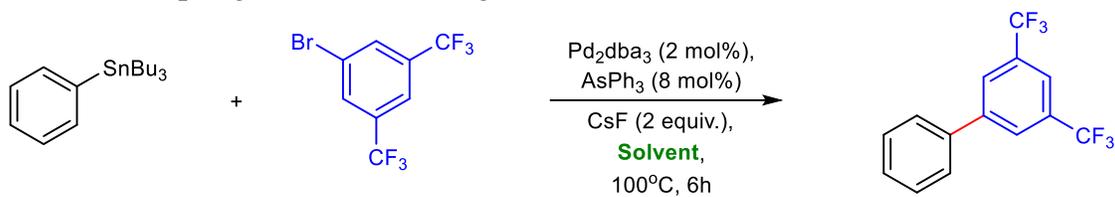
Entry	X	Yield % ^a
1	X = Br	98
2	X = Cl	24
3	X = OTf	51
4	X = OTs	0
5	X = OMs	0

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard.

Table S7. Stille coupling in rapeseed oil from Askim: optimization

Entry	Catalyst (mol%)	Ligand (mol%)	Base (2 equiv.)	Additive	°C, h	Yield % ^a
1	Pd ₂ dba ₃ (2)	XPhos (8)	KF	-	100, 24	98
2	Pd ₂ dba ₃ (2)	XPhos (8)	TBAF·3H ₂ O	-	100, 24	89
3	Pd ₂ dba ₃ (2)	XPhos (8)	CsF	-	100, 24	99
4	Pd ₂ dba ₃ (2)	XPhos (8)	K ₂ CO ₃	-	100, 24	95
5	Pd ₂ dba ₃ (2)	XPhos (8)	Cs ₂ CO ₃	-	100, 24	84
6	Pd(OAc) ₂ (2)	XPhos (3)	CsF	-	100, 24	93
7	Pd(OAc) ₂ (2)	XPhos (3)	CsF	EtOH (0.3 mL)	100, 24	97
8	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (8)	CsF	-	100, 24	4
9	Pd ₂ dba ₃ (2)	DavePhos (8)	CsF	-	100, 24	100
10	Pd ₂ dba ₃ (2)	SPhos (8)	CsF	-	100, 24	91
11	Pd ₂ dba ₃ (2)	BrettPhos (8)	CsF	-	100, 24	95
12	Pd ₂ dba ₃ (2)	RuPhos (8)	CsF	-	100, 24	95
13	Pd ₂ dba ₃ (2)	XantPhos (5)	CsF	-	100, 24	100
14	Pd ₂ dba ₃ (1.5)	Ad ₂ BuPHI (8)	CsF	-	100, 24	90
15	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	CsF	-	100, 24	37
16	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	CsF	CuI (4 mol%)	100, 24	1
17	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	CsF	CuI (4 mol%)/EtOH (0.3 mL)	100, 24	68
18	Pd(OAc) ₂ (2)	IPrHCl (3)	CsF	-	100, 24	98
19	Pd ₂ dba ₃ (2)	P(2-furyl) ₃ (8)	CsF	-	100, 24	98
20	Pd ₂ dba ₃ (2)	AsPh ₃ (8)	CsF	-	100, 24	99
21	Pd ₂ dba ₃ (2)	AsPh ₃ (8)	CsF	-	20, 24	16
22	Pd ₂ dba ₃ (2)	AsPh ₃ (8)	CsF	-	100, 6	100/99 ^b
23	Pd ₂ dba ₃ (0.5)	AsPh ₃ (2)	CsF	-	100, 6	92

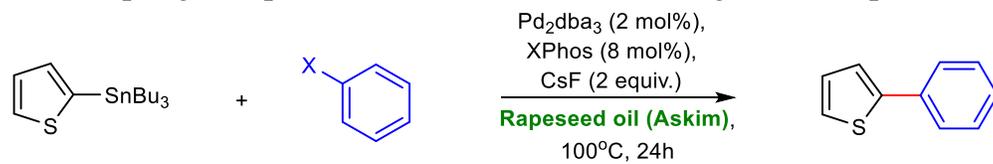
^a Yields determined by ¹H NMR using 1,2,4,5-tetramethylbenzene as internal standard. ^b Isolated yield.

Table S8. Stille coupling in oils: screening of solvents

Entry	Solvent (2 mL)	Yield % ^a
1	Triacetin	95
2	Tributylin	96
3	Rapeseed oil (Askim)	100/99 ^b
4	Rapeseed oil (Coop)	100
5	Rapeseed oil (Odelia)	98
6	Rapeseed oil (Rema)	95
7	Rapeseed oil (Anglamark)	97
8	Rapeseed oil (Sigma)	94
9	Sunflower oil	100
10	Sunflower oil (Sigma)	100
11	Olive oil	95
12	Soybean oil	98
13	Soybean oil (Sigma)	98
14	Corn oil	100
15	Avocado oil	94
16	Sesame oil	96
17	Rice bran oil	100
18	Mixture of oils	97
19	Coconut oil	98
20	Butter	95
21	Fish oil	98
22	Fish oil (Sigma)	97
23	Carnauba wax No. 1 yellow	100
24	Beeswax, refined	94
25	Lanolin	100
26	2MeTHF	100
27	Acetal	100
28	Dioxane	100
29	Toluene	100
30	DMF	75

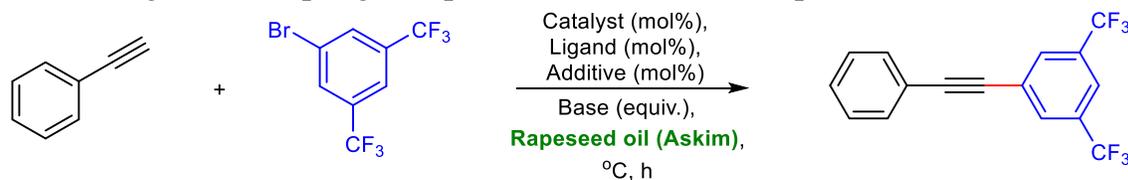
^a Yields determined by ^1H NMR using 1,2,4,5-tetramethylbenzene as internal standard. ^b Isolated yield.

Table S9. Stille coupling in rapeseed oil from Askim: screening of electrophiles



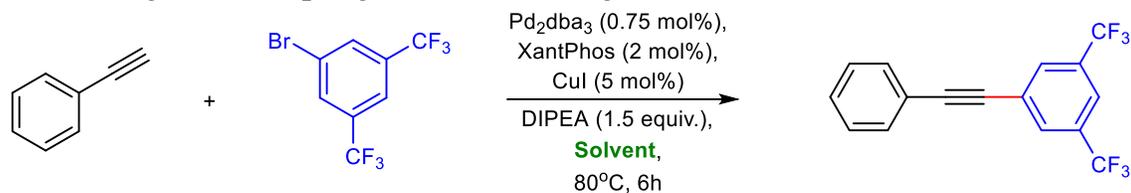
Entry	X	Yield % ^a
1	X = Br	92
2	X = Cl	100/88 ^b
3	X = OTf	74/71 ^b
4	X = OTs	47
5	X = OMs	45

^a Yields determined by ¹H NMR using 1,2,4,5-tetramethylbenzene as internal standard. ^b Isolated yields.

Table S10. Sonogashira coupling in rapeseed oil from Askim: optimization

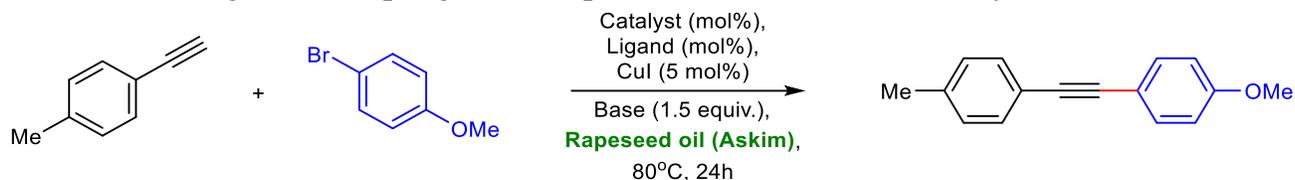
Entry	Catalyst (mol%)	Ligand (mol%)	Additive (mol%)	Base (equiv.)	°C, h	Yield % ^a
1	Pd ₂ dba ₃ (1.5)	XPhos (6)	CuI (5)	DIPEA (1.5)	80, 24	57
2	Pd ₂ dba ₃ (1.5)	<i>t</i> BuXPhos (6)	CuI (5)	DIPEA (1.5)	80, 24	73
3	Pd ₂ dba ₃ (1.5)	DavePhos (6)	CuI (5)	DIPEA (1.5)	80, 24	27
4	Pd ₂ dba ₃ (1.5)	SPhos (6)	CuI (5)	DIPEA (1.5)	80, 24	8
5	Pd ₂ dba ₃ (1.5)	BrettPhos (6)	CuI (5)	DIPEA (1.5)	80, 24	73
6	Pd ₂ dba ₃ (1.5)	RuPhos (6)	CuI (5)	DIPEA (1.5)	80, 24	11
7	Pd ₂ dba ₃ (1.5)	<i>t</i> Bu ₃ PHBF ₄ (6)	CuI (5)	DIPEA (1.5)	80, 24	21
8	Pd ₂ dba ₃ (1.5)	Ad ₂ BuP (6)	CuI (5)	DIPEA (1.5)	80, 24	23
9	Pd ₂ dba ₃ (1.5)	XantPhos (4)	CuI (5)	DIPEA (1.5)	80, 24	100
10	Pd ₂ dba ₃ (0.75)	XantPhos (2)	CuI (5)	DIPEA (1.5)	80, 6	100/86 ^b
11	Pd ₂ dba ₃ (1.5)	XantPhos (4)	CuI (5)	DIPEA (1.5)	20, 24	100
12	PdCl ₂ (PPh ₃) ₂ (3)	-	CuI (5)	DIPEA (1.5)	80, 24	99/96 ^b
13	PdCl ₂ (PPh ₃) ₂ (3)	-	CuI (5)	DIPEA (1.5)	20, 24	98
14	PdCl ₂ (PPh ₃) ₂ (3)	-	CuI (5)	DIPEA (1.5)	20, 6	36
15	PdCl ₂ (PPh ₃) ₂ (3)	-	CuI (5)	DIPEA (1.5)	80, 6	98
16	PdCl ₂ (PPh ₃) ₂ (1.5)	-	CuI (5)	DIPEA (1.5)	20, 24	89
17	PdCl ₂ (PPh ₃) ₂ (1.5)	-	CuI (5)	DIPEA (1.5)	80, 6	96
18	PdCl ₂ (PPh ₃) ₂ (3)	-	-	DIPEA (1.5)	80, 24	91
19	Pd ₂ dba ₃ (1.5)	Ad ₂ BuP (6)	-	DIPEA (1.5)	80, 24	24
20	Pd ₂ dba ₃ (1.5)	<i>t</i> Bu ₃ PHBF ₄ (6)	-	DIPEA (1.5)	80, 24	44
21	Pd ₂ dba ₃ (1.5)	XPhos (6)	-	DIPEA (1.5)	80, 24	63
22	Pd ₂ dba ₃ (1.5)	XantPhos (4)	-	DIPEA (1.5)	80, 24	45

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield.

Table S11. Sonogashira coupling in oils: screening of solvents

Entry	Solvent (2 mL)	Yield % ^a
1	Triacetin	100
2	Tributyryn	100
3	Rapeseed oil (Askim)	100/86 ^b
4	Rapeseed oil (Coop)	100
5	Rapeseed oil (Odelia)	100
6	Rapeseed oil (Rema)	100
7	Rapeseed oil (Anglamark)	100
8	Rapeseed oil (Sigma)	100
9	Sunflower oil	100
10	Sunflower oil (Sigma)	100
11	Olive oil	100
12	Soybean oil	100
13	Soybean oil (Sigma)	100
14	Corn oil	100
15	Avocado oil	99
16	Sesame oil	100
17	Rice bran oil	100
18	Mixture of oils	100
19	Coconut oil	100
20	Butter	100
21	Fish oil	100
22	Fish oil (Sigma)	100
23	Carnauba wax No. 1 yellow	100 ^c
24	Beeswax, refined	99
25	Lanolin	99
26	2MeTHF	100
27	Acetal	97
28	Dioxane	100
29	Toluene	96
30	DMF	95

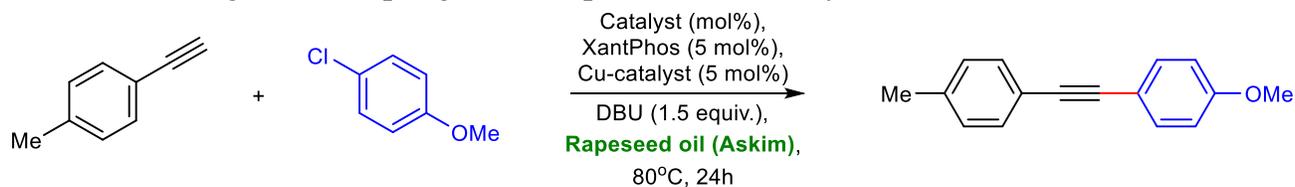
^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^b Isolated yield. ^c The reaction was performed at 90 °C.

Table S12. Sonogashira coupling in oils: optimization for unactivated aryl bromides

Entry	Catalyst (mol%)	Ligand (mol%)	Base	Yield % ^a
1	Pd ₂ dba ₃ (2)	XantPhos (5)	DIPEA	28
2	Pd ₂ dba ₃ (2)	XantPhos (5)	TEA	59
3	Pd ₂ dba ₃ (2)	XantPhos (5)	Cy ₂ NMe	67
4	Pd ₂ dba ₃ (2)	XantPhos (5)	Bu ₄ NOAc	51
5	Pd ₂ dba ₃ (2)	XantPhos (5)	Cs ₂ CO ₃	61
6	Pd ₂ dba ₃ (2)	<u>DPEPhos</u> (5)	Cy ₂ NMe	29
7	Pd ₂ dba ₃ (2)	<u><i>t</i>Bu-Xantphos</u> (5)	Cy ₂ NMe	0
8	Pd ₂ dba ₃ (2)	<u><i>N</i>-XantPhos</u> (5)	Cy ₂ NMe	25
9	Pd ₂ dba ₃ (2)	<u>dppBz</u> (5)	Cy ₂ NMe	0
10	Pd ₂ dba ₃ (2)	<u>dppf</u> (5)	Cy ₂ NMe	27
11	Pd ₂ dba ₃ (2)	<u>dippf</u> (5)	Cy ₂ NMe	21
12	Pd ₂ dba ₃ (2)	<u>1-Diphenylphosphino-1'-(di-<i>tert</i>-butylphosphino)ferrocene</u> (5)	Cy ₂ NMe	18
13	Pd ₂ dba ₃ (2)	<u>dppb</u> (5)	Cy ₂ NMe	0
14	XantPhos Pd G3 (4)	-	Cy ₂ NMe	47
15	<u>PdCl₂(dtbpf)</u> (4)	-	Cy ₂ NMe	13
16	Pd ₂ dba ₃ (2)	XPhos (8)	Cy ₂ NMe	31
17	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (8)	Cy ₂ NMe	46
18	Pd ₂ dba ₃ (2)	BrettPhos (8)	Cy ₂ NMe	36
19	Pd ₂ dba ₃ (2)	Ad ₂ BuPHI (8)	Cy ₂ NMe	14
20	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	Cy ₂ NMe	0
21	PdCl ₂ (PPh ₃) ₂ (5)	-	Cy ₂ NMe	34
22	Pd ₂ dba ₃ (2)	XantPhos (5)	<u><i>i</i>PrN(Me)<i>t</i>Bu</u>	53
23	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>2-(Diethylamino)ethanol</u>	58
24	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>Triethanolamine</u>	61
25	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>Hexamethyldisilazane</u>	8
26	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>TMEDA</u>	88
27	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>2,6-Lutidine</u>	0
28	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>DMAP</u>	57
29	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>2,2,6,6-Tetramethylpiperidine</u>	87
30	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>Pempidine</u>	77
31	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>1,1,3,3-Tetramethylguanidine</u>	66
32	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>DBU</u>	100

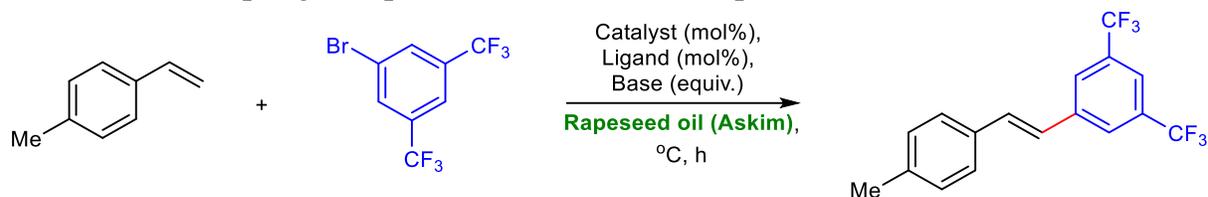
33	Pd ₂ dba ₃ (2)	XantPhos (5)	<u>TBD</u>	61
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^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard.

Table S13. Sonogashira coupling in oils: optimization for aryl chlorides

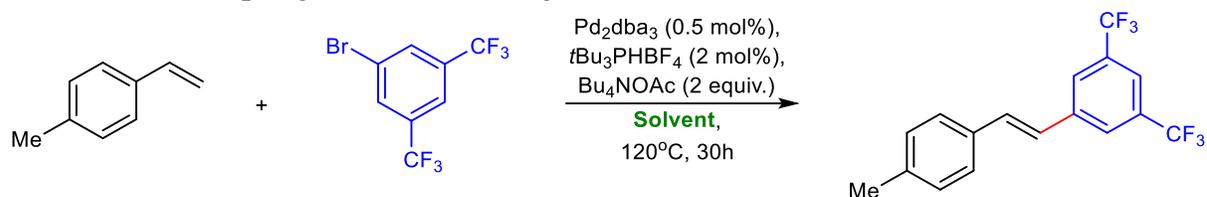
Entry	Catalyst (mol%)	Cu-catalyst (5 mol%)	Yield % ^a
1	Pd ₂ dba ₃ (2)	CuI	12
2	Pd ₂ dba ₃ (2)	<u>Cu(TC)</u>	6
3	Pd ₂ dba ₃ (2)	[(MeCN) ₄ Cu]PF ₆	2
4	PdCl ₂ (4)	CuI	0
5	Pd(OAc) ₂ (4)	CuI	0
6	[PdCl(allyl)] ₂ (2)	CuI	0
7	<u>[(η³-1-<i>tert</i>-Butylindenyl)(μ-Cl)Pd]₂</u> (2)	CuI	0
8	<u>[(Cinnamyl)PdCl]₂</u> (2)	CuI	0
9	<u>Di-μ-chlorobis[2-[(dimethylamino)methyl]phenyl-C,N]dipalladium(II)</u> (2)	CuI	0
10	<u>Di-μ-mesylobis[2'-(amino-N)[1,1'-biphenyl]-2-yl-C]dipalladium(II)</u> (2)	CuI	0
11	<u>cataCXium[®] C</u> (2)	CuI	20

^a Yields determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard.

Table S14. Heck coupling in rapeseed oil from Askim: optimization

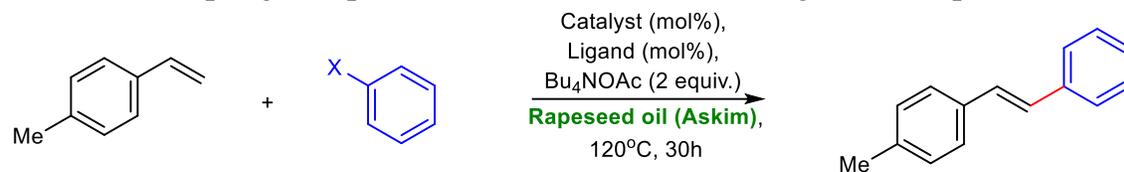
Entry	Catalyst (mol%)	Ligand (mol%)	Base (2 equiv.)	°C, h	Yield % ^a
1	Pd ₂ dba ₃ (3)	XPhos (12)	TEA	120°C, 48h	12
2	Pd ₂ dba ₃ (3)	<i>t</i> BuXPhos (12)	TEA	120°C, 48h	71
3	Pd ₂ dba ₃ (3)	<i>t</i> BuXPhos (12)	Bu ₄ NOAc	120°C, 30h	100
4	Pd ₂ dba ₃ (3)	DavePhos (12)	TEA	120°C, 48h	64
5	Pd ₂ dba ₃ (3)	SPhos (12)	TEA	120°C, 48h	20
6	Pd ₂ dba ₃ (3)	BrettPhos (12)	TEA	120°C, 48h	0
7	Pd ₂ dba ₃ (3)	RuPhos (12)	TEA	120°C, 48h	16
8	Pd ₂ dba ₃ (3)	JohnPhos (12)	TEA	120°C, 48h	56
9	XPhos Pd G3 (1)	XPhos (1)	TEA	120°C, 48h	21
10	<i>t</i> BuXPhos Pd G3 (1)	<i>t</i> BuXPhos (1)	TEA	120°C, 48h	50
11	Pd ₂ dba ₃ (3)	XantPhos (7)	TEA	120°C, 48h	0
12	Pd ₂ dba ₃ (3)	Ad ₂ BuPHI (12)	TEA	120°C, 48h	7
13	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	TEA	120°C, 48h	82
14	[PdCl(allyl)] ₂ (3)	IPrHCl (7)	TEA	120°C, 48h	17
15	PdCl ₂ (PPh ₃) ₂ (5)	-	TEA	120°C, 48h	13
16	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	TEA	80°C, 48h	8
17	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	TEA	140°C, 30h	83
18	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	TEA	120°C, 24h	82
19	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	TEA	120°C, 30h	82
20	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	Cs ₂ CO ₃	120°C, 30h	14
21	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	DIPEA	120°C, 30h	90
22	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	DIPEA	120°C, 30h	63 ^b
23	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	Cy ₂ NMe	120°C, 30h	93
24	Pd ₂ dba ₃ (3)	<i>t</i> Bu ₃ PHBF ₄ (12)	Bu ₄ NOAc	120°C, 30h	100
25	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	Bu ₄ NOAc	120°C, 30h	100
26	Pd ₂ dba ₃ (1.5)	<i>t</i> Bu ₃ PHBF ₄ (6)	Bu ₄ NOAc	120°C, 30h	100
27	Pd ₂ dba ₃ (0.5)	<i>t</i> Bu ₃ PHBF ₄ (2)	Bu ₄ NOAc	120°C, 30h	100/94 ^c

^a Yields determined by ¹H NMR using methyl 3,5-dinitrobenzoate as internal standard. ^b 4-Methylstyrene used in 2 equiv.. ^c Isolated yield.

Table S15. Heck coupling in oils: screening of solvents

Entry	Solvent (3 mL)	Yield % ^a
1	Triacetin	68/86 ^b
2	Tributylin	65/82 ^b
3	Rapeseed oil (Askim)	100/94 ^c
4	Rapeseed oil (Coop)	100
5	Rapeseed oil (Odelia)	100
6	Rapeseed oil (Rema)	100
7	Rapeseed oil (Anglamark)	100
8	Rapeseed oil (Sigma)	100
9	Sunflower oil	100
10	Sunflower oil (Sigma)	98
11	Olive oil	100
12	Soybean oil	100
13	Soybean oil (Sigma)	100
14	Corn oil	100
15	Avocado oil	95
16	Sesame oil	98
17	Rice bran oil	96
18	Mixture of oils	100
19	Coconut oil	85/96 ^b
20	Butter	97
21	Fish oil	100
22	Fish oil (Sigma)	92
23	Carnauba wax No. 1 yellow ^d	100
24	Beeswax, refined ^d	85
25	Lanolin	100
26	2MeTHF	35
27	Acetal	76
28	Dioxane	89
29	Toluene	65
30	DMF	87

^a Yields determined by ¹H NMR using methyl 3,5-dinitrobenzoate as internal standard. ^b The catalyst loading was Pd₂dba₃ (3 mol%), *t*Bu₃PHBF₄ (12 mol%). ^c Isolated yield. ^d We have used 2 g of corresponding solvent.

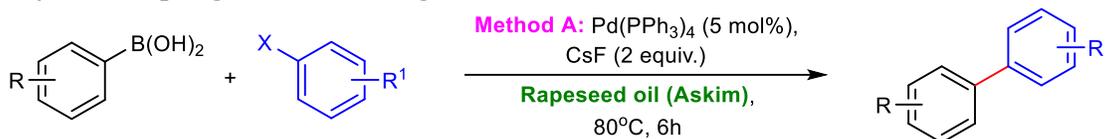
Table S16. Heck coupling in rapeseed oil from Askim: screening of electrophiles

Entry	Catalyst (mol%)	Ligand (mol%)	X	Yield % ^a
1	Pd ₂ dba ₃ (0.5)	<i>t</i> Bu ₃ PHBF ₄ (2)	X = Br	67
2	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	X = Br	100/99 ^b
3	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	X = Cl	41
4	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (8)	X = Cl	0
5	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	X = OTf	37
6	Pd ₂ dba ₃ (2)	<i>t</i> BuXPhos (8)	X = OTf	0
7	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	X = OTs	0
8	Pd ₂ dba ₃ (2)	<i>t</i> Bu ₃ PHBF ₄ (8)	X = OMs	0

^a Yields determined by ¹H NMR using methyl 3,5-dinitrobenzoate as internal standard. ^b Isolated yield.

General experimental procedures for cross-couplings

Suzuki-Miyaura coupling, Method A (Figure S29).



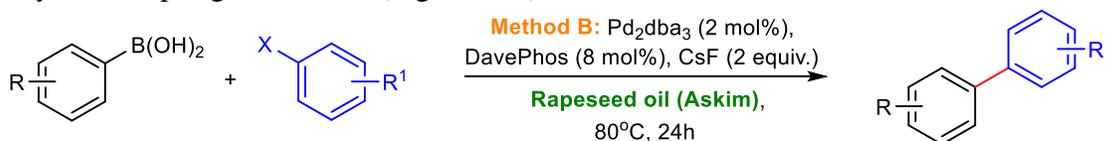
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with appropriate boronic acid (1 equiv., 0.724-0.786 mmol), Pd(PPh₃)₄ (5 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim or waste rapeseed oil (3 mL) and corresponding aryl halide³ (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 6 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Suzuki-Miyaura coupling, Method B (Figure S29).



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with appropriate boronic acid (1 equiv., 1.011-1.103 mmol), Pd₂dba₃ (2 mol%), DavePhos (8 mol%) and CsF (2 equiv.).

⁵ DCM can be substituted with 2MeTHF, Et₂O or EtOAc.

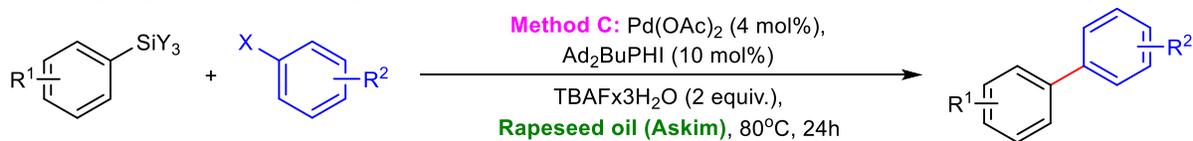
The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL) and corresponding aryl halide/sulfonate ester³ (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Hiyama coupling, Method C (Figure S29).



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd(OAc)₂ (4 mol%), Ad₂BuPHI (10 mol%) and TBAFx3H₂O (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), appropriate alkoxy-silane (1 equiv., 0.725-0.848 mmol) and corresponding aryl halide/triflate³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

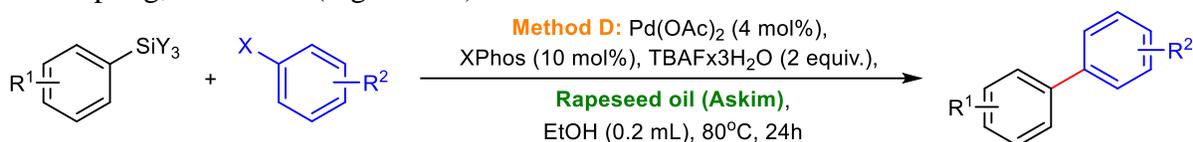
(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate,

dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Hiyama coupling, Method D (Figure S29).



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd(OAc)₂ (4 mol%), XPhos (10 mol%) and TBAFx₃H₂O (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), absolute ethanol (0.2 mL), appropriate alkoxy-silane (1 equiv., 0.725-0.848 mmol) and corresponding aryl halide³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

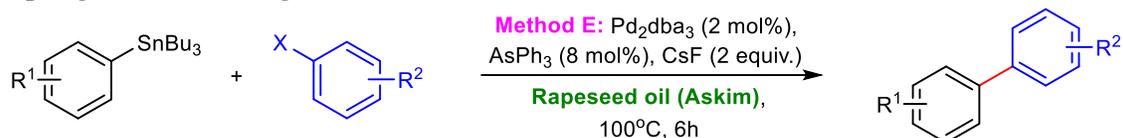
(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture.

Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Stille coupling, Method E (Figure S29).



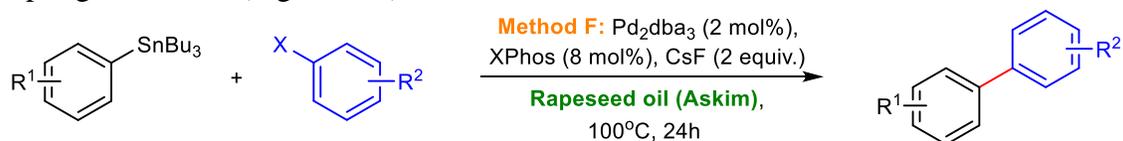
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), AsPh₃ (8 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), appropriate organotin reagent (1 equiv., 0.634-0.716 mmol) and corresponding aryl halide³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 100 °C for 6 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Stille coupling, Method F (Figure S29).



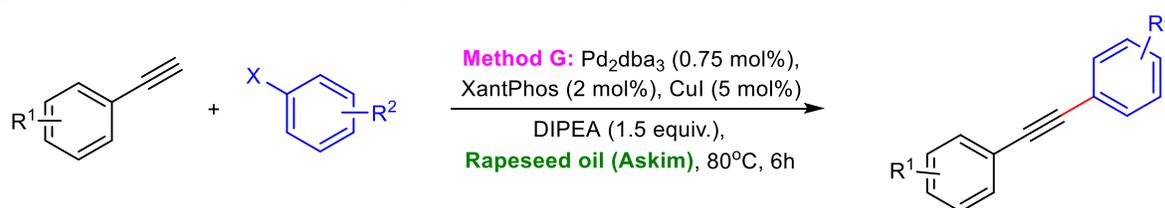
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), XPhos (8 mol%) and CsF (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), appropriate organotin reagent (1 equiv., 0.634-0.716 mmol) and corresponding aryl halide/triflate³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 100 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Sonogashira coupling, Method G (Figure S29).



Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (0.75 mol%), XantPhos (2 mol%), CuI (5 mol%), suitable acetylene (1 equiv., 0.947-0.999 mmol, degassed), corresponding aryl halide (1.5 equiv., degassed), rapeseed oil from Askim (3 mL, degassed) and DIPEA (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed from the glove box and stirred at 60-80 °C for 6-24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

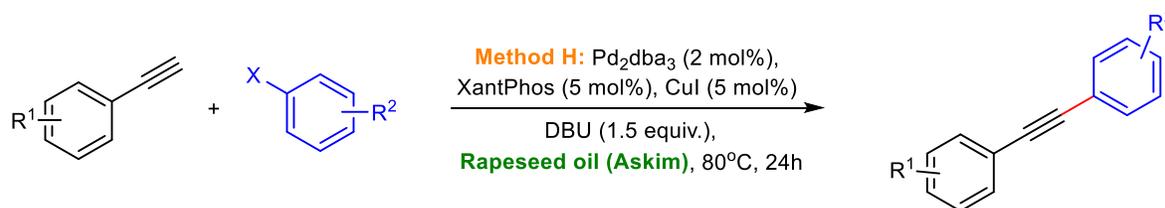
(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation

using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 250 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Sonogashira coupling, Method H (Figure S29).



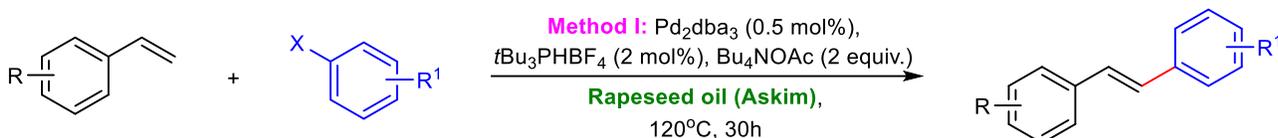
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), XantPhos (5 mol%), CuI (5 mol%), suitable acetylene (1 equiv., 0.979 mmol, degassed), corresponding aryl halide/sulfonate ester (1.5 equiv., degassed), rapeseed oil from Askim (3 mL, degassed) and DBU (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed from the glove box and stirred at 80 °C for 24 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 250 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Heck coupling, Method I (Figure S29).



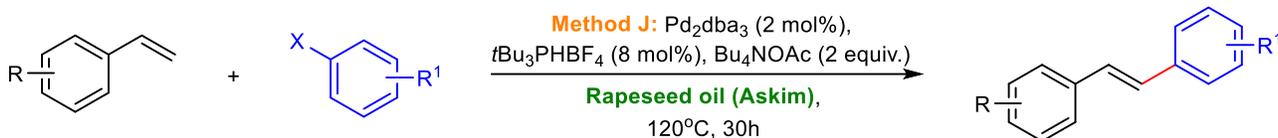
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (0.5 mol%), tBu₃PHBF₄ (2 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), appropriate olefin (1 equiv., 0.819-0.894 mmol) and corresponding aryl halide³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 250 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask. The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Heck coupling, Method J (Figure S29).



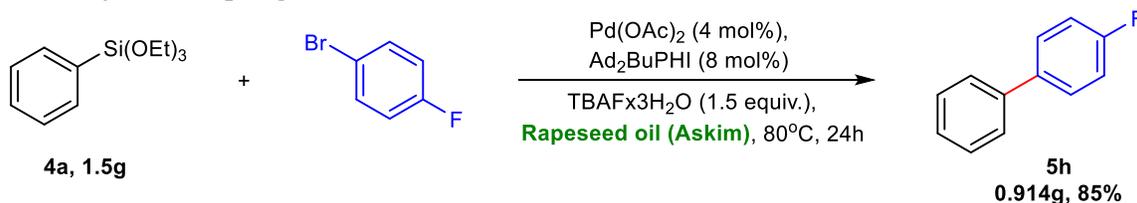
Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), *t*Bu₃PHBF₄ (8 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), appropriate olefin (1 equiv., 0.819-0.894 mmol) and corresponding aryl halide³ (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was cooled down, which was followed by isolation of the product according to one of the following methods:

(I) The reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent. Rapeseed oil can be washed out from the column using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(II) The flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 250 °C under reduced pressure. The product is being condensed in the receiving bulb within 40-60 minutes. If necessary, the condensed product can be further purified by column chromatography. Rapeseed oil from the flask containing the reaction mixture can be filtered through a short pad of silica gel using ethyl acetate, dried and applied for another experiment. NMR spectra of rapeseed oil before and after the reaction are identical.

(III) The reaction mixture was transferred into a 250 mL round bottom flask.⁶ The reaction vial was washed using EtOAc or THF and transferred into the 250 mL flask containing the reaction mixture. Afterwards, volatiles were removed using rotary evaporator that was followed by addition of 40 mL of 5M NaOH solution. The flask was equipped with a condenser and stirred under reflux for 4 h. The resulting mixture was treated with 100 mL of saturated NaCl solution, transferred into a 500 mL separating funnel and extracted with DCM (3 x 50 mL).⁵ Organic fractions were collected, evaporated to dryness and further purified using a column chromatography.

Gram-scale Hiyama coupling.

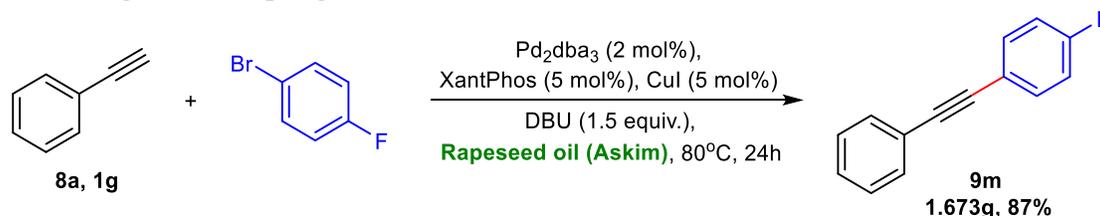


Inside of an Ar filled glove box an oven dried 50 mL round bottom flask was sequentially charged with Pd(OAc)₂ (4 mol%), Ad₂BuPHI (8 mol%) and TBAFx₃H₂O (1.5 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were

⁶ This method of separation is not applicable for the product **11h**.

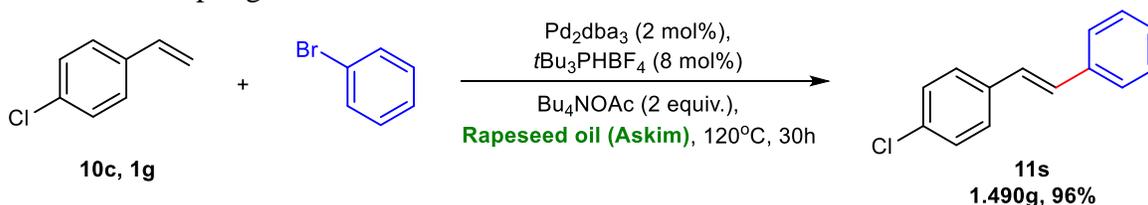
added rapeseed oil from Askim (15 mL), triethoxyphenylsilane (1 equiv., 6.240 mmol) and 1-bromo-4-fluorobenzene (1.5 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 80 °C for 24 h. Afterwards, the flask containing the reaction mixture was attached to the short-path vacuum distillation apparatus (Kugelrohr) and heated to 200 °C under reduced pressure. The product is being condensed in the receiving bulb within 60 minutes. The condensed product was further purified by column chromatography.

Gram-scale Sonogashira coupling.



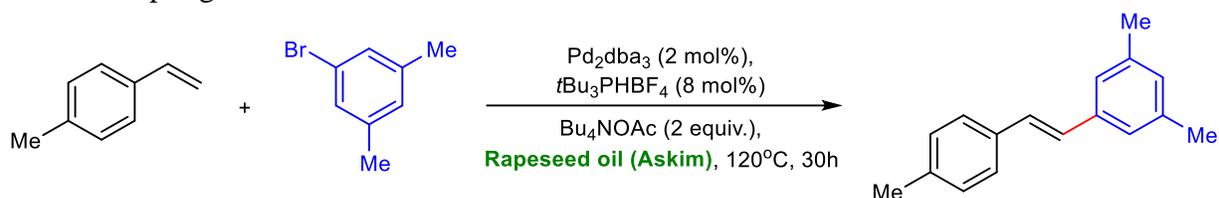
Inside of an Ar filled glove box an oven dried 50 mL round bottom flask was sequentially charged with Pd₂dba₃ (2 mol%), XantPhos (5 mol%), CuI (5 mol%), phenylacetylene (1 equiv., 9.790 mmol, degassed), 1-bromo-4-fluorobenzene (1.5 equiv., degassed), rapeseed oil from Askim (20 mL, degassed) and DBU (1.5 equiv., degassed). The flask was sealed with a rubber septa, removed from the glove box and stirred at 80 °C for 24 h. Afterwards, the reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 5 mL of DCM that was diluted with 5 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent.

Gram-scale Heck coupling.



Inside of an Ar filled glove box an oven dried 50 mL round bottom flask was sequentially charged with Pd₂dba₃ (2 mol%), *t*Bu₃PHBF₄ (8 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (20 mL), 4-chlorostyrene (1 equiv., 7.216 mmol) and bromobenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was transferred onto the top of a column, filled with silica gel, using a disposable Pasteur pipette. The reaction vial was washed with 5 mL of DCM that was diluted with 5 mL of heptane and transferred onto the top of the column. This was followed by a classical column separation using mixtures of heptane as eluent.

Substitution of heptane with other renewable solvents for column chromatography on the instance of Heck cross-coupling.

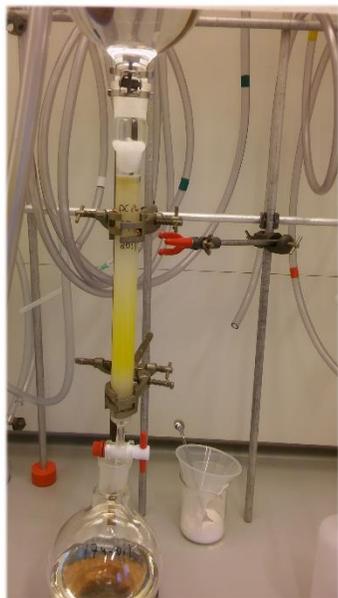


Inside of an Ar filled glove box an oven dried 10 mL flask was sequentially charged with Pd₂dba₃ (2 mol%), *t*Bu₃PHBF₄ (8 mol%) and Bu₄NOAc (2 equiv.). The flask was sealed with a rubber septa, removed from the glove box and equipped with an Ar balloon. Next, sequentially were added rapeseed oil from Askim (3 mL), 4-methylstyrene (1 equiv., 0.846 mmol) and 1-bromo-3,5-dimethylbenzene (2 equiv.). The Ar balloon was removed and the resulting mixture was stirred at 120 °C for 30 h. Afterwards, the reaction mixture was transferred onto the top of a column, filled with silica gel (suspended in suitable monoterpene), using a disposable Pasteur pipette. The reaction vial was washed with 1 mL of DCM that was diluted with 1 mL of appropriate monoterpene and transferred onto the top of the column. This was followed by a classical column separation using mixtures of corresponding monoterpene as eluent.

(A) Separation using a Kugelrohr



(B) Separation with column chromatography



(C) Separation based on hydrolysis and extraction



Figure S29. Overview of separation methods.

Production and processing of waste rapeseed oil

(A) Frying potatoes in rapeseed oil



(B) Filtration of waste rapeseed oil



(C) Waste rapeseed oil after filtration

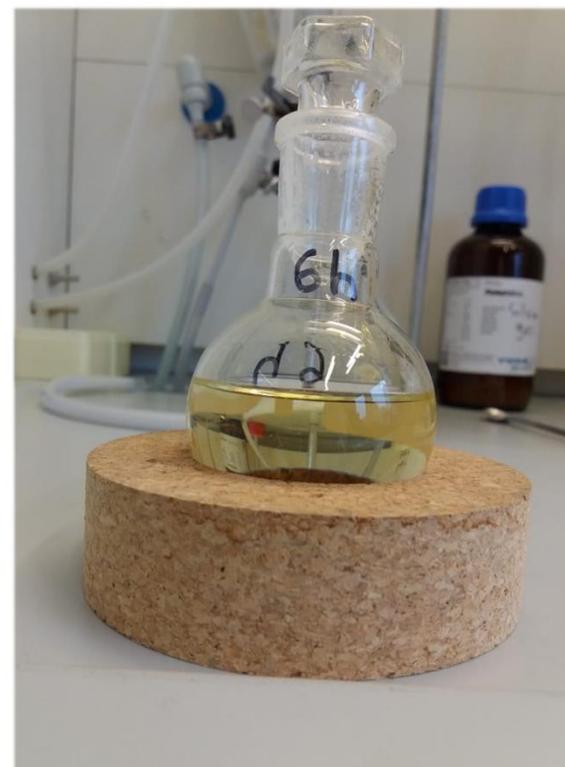


Figure S30. Production and processing of waste rapeseed oil.

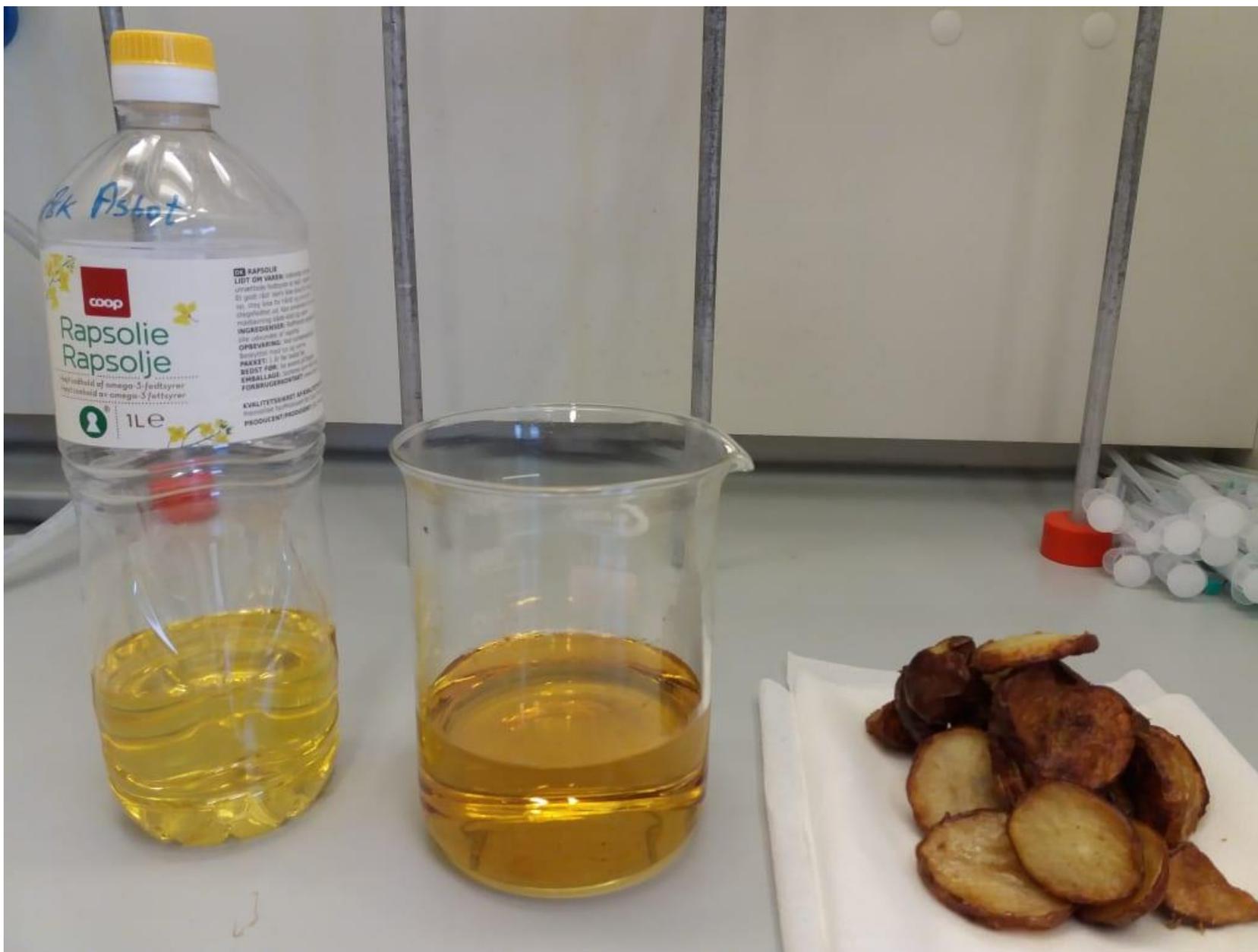
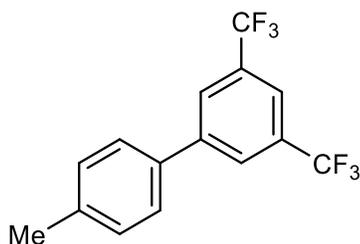


Figure S31. Rapeseed oil from Coop before and after frying potatoes at 130 °C for 8h.

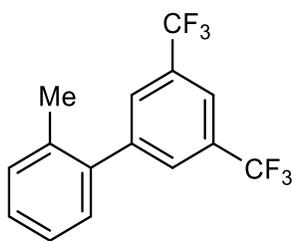


Figure S32. Waste rapeseed oils used for frying potatoes for 2, 4, 6 and 8 hours respectively.

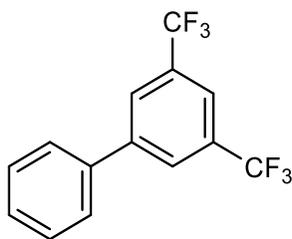
Characterization of products



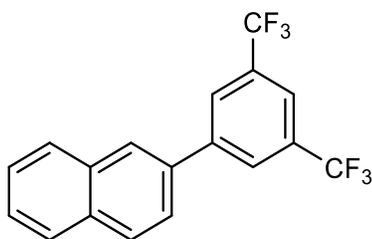
4'-Methyl-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 3a.⁷ Starting from 0.736 mmol of corresponding boronic acid the product was obtained as a white solid, yield 92% (0.205 g, method A). For the reaction performed in waste rapeseed oil, used for frying at 130 °C for 8h, starting from 0.736 mmol of corresponding boronic acid the product was obtained as a white solid, yield 93% (0.207 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 2.46 (s, 3H, Me), 7.34 (d, *J* = 7.9 Hz, 2H, Ar), 7.53-7.55 (m, 2H, Ar), 7.88 (s, 1H, Ar), 8.03 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.3, 120.8 (p, *J* = 3.9 Hz), 123.7 (q, *J* = 271 Hz), 127.0-127.1 (m), 127.2, 130.2, 132.3 (q, *J* = 33.2 Hz), 135.5, 139.2, 143.5.



2-Methyl-3',5'-bis(trifluoromethyl)-1,1'-biphenyl, 3b.⁸ Starting from 0.736 mmol of corresponding boronic acid the product was obtained as a colourless oil, yield 91% (0.203 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 2.31 (s, 3H, Me), 7.25-7.27 (m, 1H, Ar), 7.31-7.40 (m, 3H, Ar), 7.84 (s, 2H, Ar), 7.92 (s, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 20.4, 121.0 (hept, *J* = 3.9 Hz), 123.6 (q, *J* = 272.6 Hz), 126.5, 128.9, 129.6 (q, *J* = 3.8 Hz), 129.8, 131.0, 131.8 (q, *J* = 33.2 Hz), 135.4, 139.1, 144.2.



3,5-Bis(trifluoromethyl)-1,1'-biphenyl, 3c.⁹ Starting from 0.738 mmol of corresponding boronic acid the product was obtained as a white solid, yield 61% (0.131 g, method A). Starting from 1.066 mmol of corresponding boronic acid the product was obtained as a white solid, 94% (0.291 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.44-7.49 (m, 1H, Ar), 7.50-7.54 (m, 2H, Ar), 7.60-7.63 (m, 2H, Ar), 7.88 (s, 1H, Ar), 8.03 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.1 (p, *J* = 3.8 Hz), 123.6 (q, *J* = 271 Hz), 127.4, 129.1, 129.5, 132.3 (q, *J* = 33.2 Hz), 138.4, 143.5.



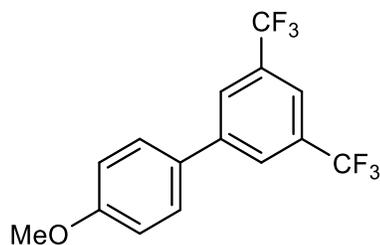
2-(3,5-Bis(trifluoromethyl)phenyl)naphthalene, 3d.¹⁰ Starting from 0.756 mmol of corresponding boronic acid the product was obtained as a white solid, yield 97% (0.249 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 7.59 (td, *J* = 5.7, 4.8, 3.3 Hz, 2H, Ar), 7.71 (dd, *J* = 8.5, 1.9 Hz, 1H, Ar), 7.90-7.98 (m, 4H, Ar), 8.05 (s, 1H, Ar), 8.17 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.1 (hept, *J* = 3.9 Hz), 123.7 (q, *J* = 271 Hz), 124.8, 126.7, 127.0, 127.1, 127.5 (q, *J* = 3.5 Hz), 127.9, 128.6, 129.3, 132.4 (q, *J* = 33.2 Hz), 133.4, 133.7, 135.5, 143.4.

⁷ F. D'Accriscio, A. Ohleier, E. Nicolas, M. Demange, O. T. D. Boullay, N. Saffon-Merceron, M. Fustier-Boutignon, E. Rezabal, G. Frison, N. Nebra and N. Mezailles, *Organometallics*, 2020, **39**, 1688-1699.

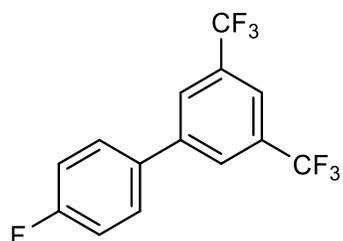
⁸ Y. Uozumi and Y. Nakai, *Org. Lett.*, 2002, **4**, 2997-3000.

⁹ S. Ichii, G. Hamasaka and Y. Uozumi, *Chem. Asian J.*, 2019, **14**, 3850-3854.

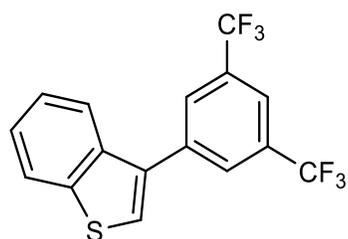
¹⁰ Y.-N. Wang, X.-Q. Guo, X.-H. Zhu, R. Zhong, L.-H. Cai and X.-F. Hou, *Chem. Commun.*, 2012, **48**, 10437-10439.



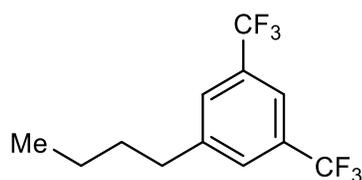
4'-Methoxy-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 3e.¹¹ Starting from 0.724 mmol of corresponding boronic acid the product was obtained as a white solid, yield 95% (0.219 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 3.88 (s, 3H, OMe), 7.02-7.05 (m, 2H, Ar), 7.54-7.58 (m, 2H, Ar), 7.82 (s, 1H, Ar), 7.98 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 55.6, 114.9, 120.4 (p, *J* = 3.8 Hz), 123.7 (q, *J* = 271 Hz), 126.8 (q, *J* = 3.8 Hz), 128.6, 130.8, 132.2 (q, *J* = 33.1 Hz), 143.1, 160.6.



4'-Fluoro-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 3f.¹² Starting from 0.786 mmol of corresponding boronic acid the product was obtained as a white solid, yield 42% (0.102 g, method A). Starting from 1.072 mmol of corresponding boronic acid the product was obtained as a white solid, 94% (0.311 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.18-7.23 (m, 2H, Ar), 7.57-7.62 (m, 2H, Ar), 7.88 (s, 1H, Ar), 7.98 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 116.5 (d, *J* = 21.8 Hz), 121.2 (hept, *J* = 3.9 Hz), 123.6 (q, *J* = 271 Hz), 127.3 (q, *J* = 3.8 Hz), 129.2 (d, *J* = 8.4 Hz), 132.5 (q, *J* = 33.3 Hz), 134.6 (d, *J* = 3.3 Hz), 142.5, 163.6 (d, *J* = 249.2 Hz).



3-(3,5-Bis(trifluoromethyl)phenyl)benzo[b]thiophene, 3g.¹³ Starting from 1.011 mmol of corresponding boronic acid the product was obtained as a white solid, yield 99% (0.345 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.45-7.52 (m, 2H, Ar), 7.55 (s, 1H, Ar), 7.83-7.87 (m, 1H, Ar), 7.96-8.00 (m, 2H, Ar), 8.09 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.5 (p, *J* = 3.8 Hz), 122.2, 123.4, 123.6 (q, *J* = 271 Hz), 125.2, 125.3, 125.9, 128.9 (q, *J* = 3.6 Hz), 132.4 (q, *J* = 33.4 Hz), 135.1, 137.2, 138.3, 141.0.



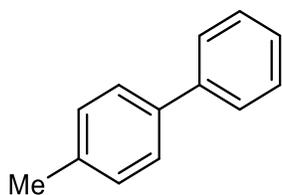
1-Butyl-3,5-bis(trifluoromethyl)benzene, 3h.¹⁴ Starting from 1.079 mmol of corresponding boronic acid the product was obtained as a colourless oil, yield 18% (0.052 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 0.96 (t, *J* = 7.3 Hz, 3H, Me), 1.39 (dq, *J* = 14.7, 7.3 Hz, 2H, CH₂), 1.61-1.69 (m, 2H, CH₂), 2.72-2.76 (m, 2H, CH₂), 7.63 (s, 2H, Ar), 7.70 (s, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 14.0, 22.5, 33.4, 35.6, 120.0 (hept, *J* = 4.0 Hz), 123.7 (q, *J* = 270 Hz), 128.7 (dd, *J* = 5.6, 2.7 Hz), 131.7 (q, *J* = 33.0 Hz), 145.4.

¹¹ A. D. Zotto, F. Amoroso, W. Baratta and P. Rigo, *Eur. J. Org. Chem.*, 2009, 110-116.

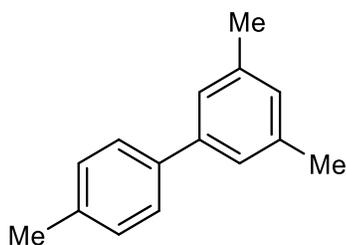
¹² M. Butters, J. N. Harvey, J. Jover, A. J. J. Lennox, G. C. Lloyd-Jones and P. M. Murray, *Angew. Chem. Int. Ed.*, 2010, **49**, 5156-5160.

¹³ C. Colletto, J. Bures and I. Larrosa, *Chem. Commun.*, 2017, **53**, 12890-12893.

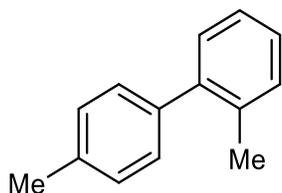
¹⁴ T. Agrawal and S. P. Cook, *Org. Lett.*, 2013, **15**, 96-99.



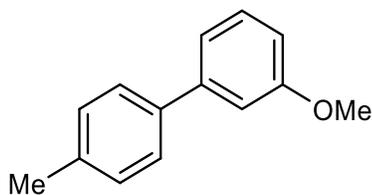
4-Methyl-1,1'-biphenyl, 3i.¹⁵ Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a white solid, yield 99% (0.184 g, X = Br, method B), 49% (0.091 g, X = Cl, method B), 99% (0.185 g, X = OTf, method B). ¹H NMR (400 MHz, CDCl₃): δ = 2.53 (s, 3H, Me), 7.38 (d, *J* = 7.9 Hz, 2H, Ar), 7.44-7.48 (m, 1H, Ar), 7.54-7.58 (m, 2H, Ar), 7.63-7.65 (m, 2H, Ar), 7.71-7.74 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.2, 127.1, 127.2, 128.9, 129.6, 137.1, 138.5, 141.3.



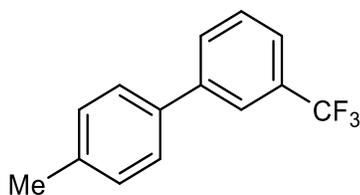
3,4,5-Trimethyl-1,1'-biphenyl, 3j.¹⁶ Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a white solid, yield 99% (0.216 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 2.47-2.48 (m, 9H, 3xMe), 7.07 (s, 1H, Ar), 7.30-7.33 (m, 4H, Ar), 7.56-7.59 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.3, 21.6, 125.1, 127.2, 128.8, 129.5, 136.9, 138.3, 138.7, 141.4.



2,4'-Dimethyl-1,1'-biphenyl, 3k.¹⁷ Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a colourless oil, yield 99% (0.201 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 2.43-2.45 (m, 3H, Me), 2.55-2.56 (m, 3H, Me), 7.37-7.42 (m, 8H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 20.7, 21.3, 125.9, 127.2, 128.9, 129.2, 130.0, 130.4, 135.5, 136.5, 139.2, 142.1.



3-Methoxy-4'-methyl-1,1'-biphenyl, 3l.¹⁸ Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a white solid, yield 98% (0.215 g, X = Br, method B), 80% (0.175 g, X = Cl, method B). ¹H NMR (400 MHz, CDCl₃): δ = 2.48-2.49 (m, 3H, Me), 3.93-3.94 (m, 3H, OMe), 6.95-6.99 (m, 1H, Ar), 7.21-7.24 (m, 1H, Ar), 7.25-7.28 (m, 1H, Ar), 7.32-7.35 (m, 2H, Ar), 7.40-7.45 (m, 1H, Ar), 7.57-7.61 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.2, 55.4, 112.5, 112.9, 119.6, 127.2, 129.6, 129.8, 137.3, 138.4, 142.8, 160.1.



4'-Methyl-3-(trifluoromethyl)-1,1'-biphenyl, 3m.¹⁹ Starting from 0.736 mmol of corresponding boronic acid the product was obtained as a white solid, yield 80% (0.139 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 2.48 (s, 3H, Me), 7.34 (d, *J* = 7.8 Hz, 2H, Ar), 7.55-7.60 (m, 3H, Ar), 7.64-7.66 (m, 1H, Ar), 7.80 (d, *J* = 7.7 Hz, 1H, Ar), 7.91 (s, 1H, Ar). ¹³C

¹⁵ M. M. Heravi, S. Asadi, S. M. H. Chopani and E. Jaderi, *Appl. Organomet. Chem.*, 2020, **34**, 5805.

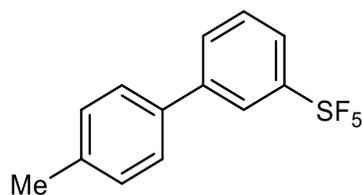
¹⁶ B. R. Barnett, L. A. Labios, J. M. Stauber, C. E. Moore, A. L. Rheingold and J. S. Figueroa, *Organometallics*, 2017, **36**, 944-954.

¹⁷ K. L. Wilson, J. Murray, C. Jamieson and A. J. B. Watson, *Synlett*, 2018, **29**, 650-654.

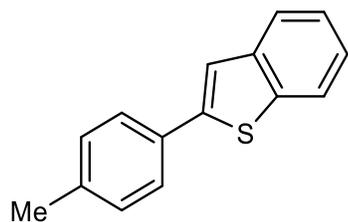
¹⁸ X. Zhou, X. Guo, F. Jian and G. Wei, *ACS Omega*, 2018, **3**, 4418-4422.

¹⁹ A. F. Asachenko, K. R. Sorochkina, P. B. Dzhevakov, M. A. Topchiy and M. S. Nechaev, *Adv. Synth. Catal.*, 2013, **355**, 3553-3557.

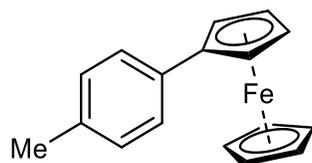
NMR (101 MHz, CDCl₃): δ = 21.1, 123.7 (dq, J = 7.6, 3.8 Hz), 124.4 (q, J = 271 Hz), 127.1, 129.2, 129.8, 130.2-130.3 (m), 131.2 (q, J = 32.1 Hz), 136.9, 138.0, 142.0.



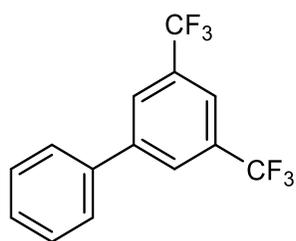
Pentafluoro(4'-methyl-[1,1'-biphenyl]-3-yl)- λ^6 -sulfane, 3n. Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a white solid, yield 99% (0.324 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 2.48 (s, 3H, Me), 7.33-7.35 (m, 2H, Ar), 7.52-7.56 (m, 3H, Ar), 7.73-7.78 (m, 2H, Ar), 8.03 (p, J = 2.0 Hz, 1H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 21.2, 124.6 (dp, J = 20.5, 4.6 Hz), 127.2, 129.2, 130.0, 130.2, 136.7, 138.4, 142.5, 154.7 (p, J = 16.8 Hz). **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₃H₁₂F₅S 295.0574 found 295.0570.



2-(p-Tolyl)benzo[b]thiophene, 3o.²⁰ Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a white solid, yield 62% (0.154 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 2.41 (s, 3H, Me), 7.24-7.27 (m, 2H, Ar), 7.30-7.39 (m, 2H, Ar), 7.50-7.53 (m, 1H, Ar), 7.62-7.65 (m, 2H, Ar), 7.76-7.79 (m, 1H, Ar), 7.83-7.85 (m, 1H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 21.4, 119.0, 122.4, 123.6, 124.3, 124.6, 126.5, 129.8, 131.7, 138.4, 139.5, 141.0, 144.6.



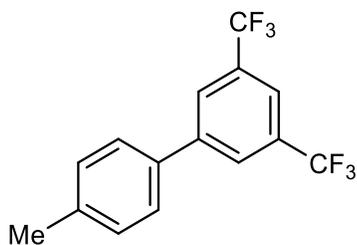
(4-Methylphenyl)ferrocene, 3p. Starting from 1.103 mmol of corresponding boronic acid the product was obtained as a dark red solid, yield 51% (0.154 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 2.40 (s, 3H, Me), 4.11 (s, 5H, ferrocene), 4.35 (t, J = 1.9 Hz, 2H, ferrocene), 4.68 (t, J = 1.9 Hz, 2H, ferrocene), 7.17 (d, J = 7.8 Hz, 2H, Ar), 7.45 (d, J = 7.9 Hz, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 21.3, 66.5, 68.8, 69.7, 85.9, 126.2, 129.2, 135.6, 136.2. **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₇H₁₇Fe 277.0674 found 277.0674.



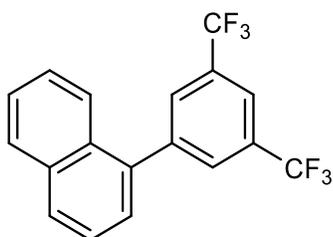
3,5-Bis(trifluoromethyl)-1,1'-biphenyl, 5a.²¹ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless solid, yield 79% (0.190 g, method A), 90% (0.218 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 7.46-7.55 (m, 3H, Ar), 7.62-7.64 (m, 2H, Ar), 7.90 (s, 1H, Ar), 8.05 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 121.1 (hept, J = 3.9 Hz), 123.6 (q, J = 271 Hz), 127.4, 129.1, 129.5, 132.4 (q, J = 33.2 Hz), 138.5, 143.6.

²⁰ A. E. Purta, S. Ichiia, A. Tazawa and Y. Uozumi, *Synlett*, 2020, **31**, 1634-1638.

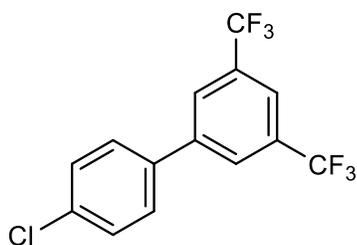
²¹ S. Ichii, G. Hamasaka and Y. Uozumi, *Chem. Asian J.*, 2019, **14**, 3850-3854.



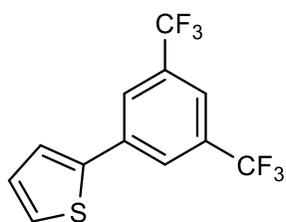
4'-Methyl-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 5b.²² Starting from 0.848 mmol of corresponding silane the product was obtained as a colourless solid, yield 82% (0.211 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 2.47 (s, 3H, Me), 7.34-7.36 (m, 2H, Ar), 7.52-7.55 (m, 2H, Ar), 7.89 (s, 1H, Ar), 8.04 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.3, 120.8 (hept, *J* = 3.9 Hz), 123.7 (q, *J* = 271 Hz), 127.1-127.2 (m), 127.2, 130.2, 132.3 (q, *J* = 33.2 Hz), 135.5, 139.2, 143.5.



1-(3,5-Bis(trifluoromethyl)phenyl)naphthalene, 5c.²³ Starting from 0.725 mmol of corresponding silane the product was obtained as a colourless solid, yield 75% (0.185 g, method A), 78% (0.192 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.44-7.46 (m, 1H, Ar), 7.49-7.60 (m, 3H, Ar), 7.72-7.76 (m, 1H, Ar), 7.96-8.06 (m, 5H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 119.5, 121.4 (p, *J* = 3.8 Hz), 122.3, 125.0, 125.5, 126.0, 126.5, 127.2, 127.6, 127.7, 128.1, 128.9, 129.3, 130.4 (d, *J* = 3.9 Hz), 131.2, 132.0 (q, *J* = 33.2, 32.7 Hz), 134.0, 137.1, 143.1.



4'-Chloro-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 5d.²⁴ Starting from 0.728 mmol of corresponding silane the product was obtained as a colourless solid, yield 95% (0.224 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 7.46-7.50 (m, 2H, Ar), 7.53-7.57 (m, 2H, Ar), 7.89 (s, 1H, Ar), 7.99 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.4 (hept, *J* = 3.8 Hz), 123.5 (q, *J* = 271 Hz), 127.2 (q, *J* = 3.9 Hz), 128.7, 129.7, 132.5 (q, *J* = 33.3 Hz), 135.5, 136.8, 142.3.



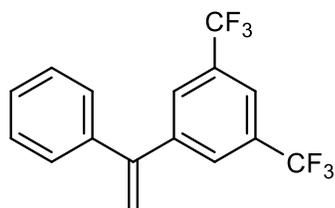
2-(3,5-Bis(trifluoromethyl)phenyl)thiophene, 5e.²⁵ Starting from 0.812 mmol of corresponding silane the product was obtained as a colourless solid, yield 18% (0.043 g, method A), 60% (0.143 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.14-7.16 (m, 1H, thiophene), 7.41-7.45 (m, 2H, thiophene), 7.78 (s, 1H, Ar), 8.00 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 120.9 (dt, *J* = 7.7, 3.9 Hz), 123.4 (q, *J* = 271 Hz), 125.5, 125.8-125.9 (m), 127.2, 128.8, 132.5 (q, *J* = 33.4 Hz), 136.7, 140.6, 141.1.

²² F. D'Accriscio, A. Ohleier, E. Nicolas, M. Demange, O. T. D. Boullay, N. Saffon-Merceron, M. Fustier-Boutignon, E. Rezabal, G. Frison, N. Nebra and N. Mezailles, *Organometallics*, 2020, **39**, 1688-1699.

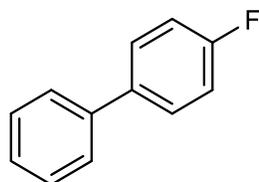
²³ Z. Zhou, H. Liang, W. Xia, H. Chen, Y. Zhang, X. He, S. Yu, R. Cao and L. Qiu, *New J. Chem.*, 2018, **42**, 5967-5971.

²⁴ Y. Uozumi and Y. Nakai, *Org. Lett.*, 2002, **4**, 2997-3000.

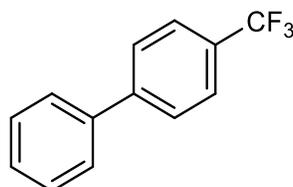
²⁵ J. Yang, S. Liu, J.-F. Zheng and J. Zhou, *Eur. J. Org. Chem.*, 2012, 6248-6259.



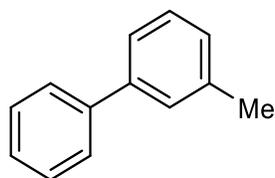
1-(1-Phenylvinyl)-3,5-bis(trifluoromethyl)benzene, 5f.²⁶ Starting from 0.751 mmol of corresponding silane the product was obtained as a colourless oil, yield 31% (0.073 g, method A), 73% (0.173 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 5.58 (s, 1H, olefin), 5.67 (s, 1H, olefin), 7.29-7.35 (m, 2H, Ar), 7.38-7.43 (m, 3H, Ar), 7.80 (s, 2H, Ar), 7.86 (s, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 117.3, 121.7 (dt, *J* = 7.7, 3.9 Hz), 123.5 (q, *J* = 271 Hz), 128.2, 128.5 (q, *J* = 4.0 Hz), 128.8, 128.9, 131.9 (q, *J* = 33.3 Hz), 139.9, 143.9, 148.0.



4-Fluoro-1,1'-biphenyl, 5h.²⁷ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless oil, yield 87% (0.125 g, X = Br, method A), 57% (0.081 g, X = Cl, method A). For gram-scale experiment starting from 6.240 mmol (1.5 g) of corresponding silane the product was obtained as a colourless oil, yield 85% (0.914 g). ¹H NMR (400 MHz, CDCl₃): δ = 7.14-7.19 (m, 2H, Ar), 7.36-7.42 (m, 1H, Ar), 7.46-7.51 (m, 2H, Ar), 7.56-7.61 (m, 4H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 115.8 (d, *J* = 21.4 Hz), 127.2, 127.4, 128.9 (d, *J* = 8.1 Hz), 129.0, 137.5 (d, *J* = 3.3 Hz), 140.4, 162.6 (d, *J* = 246.3 Hz).



4-(Trifluoromethyl)-1,1'-biphenyl, 5i.²⁸ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless oil, yield 81% (0.150 g, X = Br, method A), 87% (0.161 g, X = Cl, method A). ¹H NMR (400 MHz, CDCl₃): δ = 7.45-7.49 (m, 1H, Ar), 7.51-7.56 (m, 2H, Ar), 7.64-7.67 (m, 2H, Ar), 7.72-7.77 (m, 4H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 124.6 (q, *J* = 270 Hz), 125.9 (q, *J* = 3.8 Hz), 127.4, 127.6, 128.4, 129.2, 129.5 (q, *J* = 32.4 Hz), 139.9, 144.9 (q, *J* = 1.4 Hz).



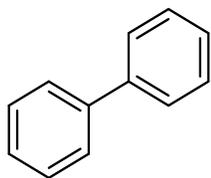
3-Methyl-1,1'-biphenyl, 5j.²⁹ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless solid, yield 86% (0.120 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 2.54 (s, 3H, Me), 7.28-7.30 (m, 1H, Ar), 7.43-7.48 (m, 2H, Ar), 7.52-7.57 (m, 4H, Ar), 7.70-7.73 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.7, 124.4, 127.3, 127.4, 128.1, 128.2, 128.8, 128.9, 138.5, 141.4, 141.5.

²⁶ F. Berthiol, H. Doucet and M. Santelli, *Eur. J. Org. Chem.*, 2003, 1091-1096.

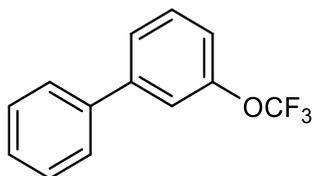
²⁷ R. N. Dhital, A. Sen, T. Sato, H. Hu, R. Ishii, D. Hashizume, H. Takaya, Y. Uozumi and Y. M. A. Yamada, *Org. Lett.*, 2020, **22**, 4797-4801.

²⁸ A. Ohno, T. Sato, T. Mase, Y. Uozumi and Y. M. A. Yamada, *Adv. Synth. Catal.*, 2020, **362**, 4687-4698.

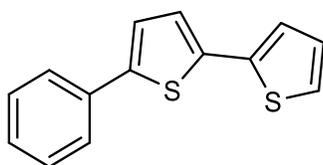
²⁹ S. Yang and S. H. Hong, *Asian J. Org. Chem.*, 2020, **9**, 1846-1851.



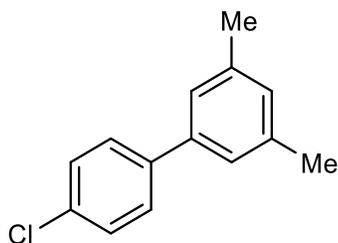
1,1'-Biphenyl, 5k.³⁰ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless solid, yield 86% (0.110 g, X = Br, method A), 58% (0.074 g, X = OTf, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 7.42-7.46 (m, 2H, Ar), 7.51-7.56 (m, 4H, Ar), 7.68-7.71 (m, 4H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 127.3, 127.4, 128.9, 141.4.



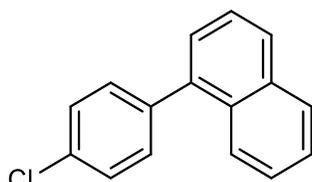
3-(Trifluoromethoxy)-1,1'-biphenyl, 5l.³¹ Starting from 0.832 mmol of corresponding silane the product was obtained as a colourless oil, yield 85% (0.169 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 7.25-7.29 (m, 1H, Ar), 7.42-7.46 (m, 1H, Ar), 7.47-7.53 (m, 4H, Ar), 7.56-7.59 (m, 1H, Ar), 7.61-7.65 (m, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 119.7, 119.9, 120.8 (q, *J* = 255 Hz), 125.7, 127.3, 128.2, 129.1, 130.2, 139.9, 143.6, 149.9 (q, *J* = 1.8 Hz).



5-Phenyl-2,2'-bithiophene, 5m.³² Starting from 0.832 mmol of corresponding silane the product was obtained as a yellow solid, yield 50% (0.100 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 7.04-7.07 (m, 1H, Ar), 7.17-7.18 (m, 1H, Ar), 7.22-7.25 (m, 3H, Ar), 7.29-7.34 (m, 1H, Ar), 7.39-7.43 (m, 2H, Ar), 7.62-7.65 (m, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 123.8, 123.9, 124.5, 124.8, 125.8, 127.7, 128.0, 129.1, 134.2, 136.9, 137.6, 143.3.



4'-Chloro-3,5-dimethyl-1,1'-biphenyl, 5n.³³ Starting from 0.728 mmol of corresponding silane the product was obtained as a colourless solid, yield 95% (0.150 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 2.44 (s, 6H, 2xMe), 7.07 (s, 1H, Ar), 7.23 (s, 2H, Ar), 7.42-7.45 (m, 2H, Ar), 7.54-7.56 (m, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 21.5, 125.1, 128.5, 128.9, 129.2, 129.4, 133.3, 138.6, 140.0, 140.1.



1-(4-Chlorophenyl)naphthalene, 5o.³⁴ Starting from 0.728 mmol of corresponding silane the product was obtained as a colourless solid, yield 84% (0.145 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 7.39-7.54 (m, 7H, Ar), 7.64-7.66 (m, 1H, Ar), 7.83-7.93 (m, 3H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 125.4, 125.5, 125.9, 126.1, 126.3, 126.4, 126.7, 127.1, 127.9, 128.2, 128.4, 128.6, 128.7, 128.8, 129.2, 131.5, 131.6, 132.9, 133.5, 133.7, 133.8, 134.0, 137.5, 139.1, 139.4, 139.8.

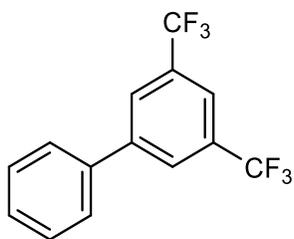
³⁰ M. Xiaojing, L. Gao, Z. Weng, H. Yang and X. Sun, *New J. Chem.*, 2020, **44**, 20525-20529.

³¹ M. Zhou, C. Ni, Z. He and J. Hu, *Org. Lett.*, 2016, **18**, 3754-3757.

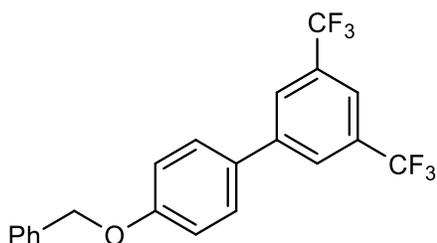
³² M. M. Martinez, M. Pena-Lopez, J. P. Sestelo and L. A. Sarandeses, *Org. Biomol. Chem.*, 2012, **10**, 3892-3898.

³³ P. Pattanayak, D. Patra, P. Brandao, D. Mal and V. Felix, *Inorg. Chem. Commun.*, 2015, **53**, 68-71.

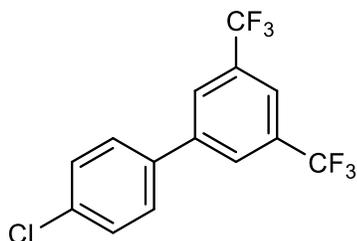
³⁴ P.-F. Li, C.-B. Yi, S.-J. Ren and J. Qu, *Adv. Synth. Catal.*, 2016, **358**, 2088-2092.



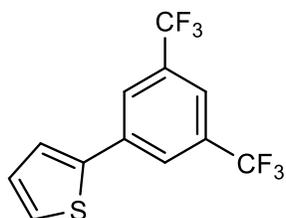
3,5-Bis(trifluoromethyl)-1,1'-biphenyl, 7a.²¹ Starting from 0.681 mmol of corresponding organotin reagent the product was obtained as a colourless solid, yield 99% (0.197 g, method C). **¹H NMR** (400 MHz, CDCl₃): δ = 7.45-7.55 (m, 3H, Ar), 7.62-7.66 (m, 2H, Ar), 7.89 (s, 1H, Ar), 8.04 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 121.1 (p, *J* = 3.9 Hz), 123.6 (q, *J* = 271 Hz), 127.4-127.5 (m), 129.1, 129.5, 132.4 (q, *J* = 33.3 Hz), 138.5, 143.6.



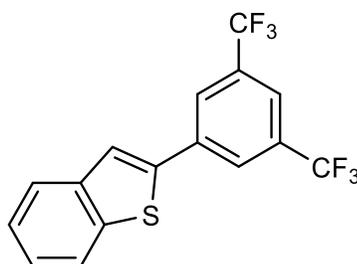
4'-(Benzyloxy)-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 7b. Starting from 0.634 mmol of corresponding organotin reagent the product was obtained as a colourless solid, yield 85% (0.213 g, method C). **¹H NMR** (400 MHz, CDCl₃): δ = 5.17 (s, 2H, CH₂), 7.13-7.16 (m, 2H, Ar), 7.38-7.42 (m, 1H, Ar), 7.43-7.48 (m, 2H, Ar), 7.50-7.53 (m, 2H, Ar), 7.56-7.59 (m, 2H, Ar), 7.87 (s, 1H, Ar), 8.01 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 70.3, 120.4 (hept, *J* = 3.9 Hz), 123.7 (q, *J* = 271 Hz), 126.8 (q, *J* = 3.7 Hz), 127.7, 128.3, 128.6, 128.9, 131.0, 132.2 (q, *J* = 33.1 Hz), 136.8, 143.0, 159.7. **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₂₁H₁₅F₆O 397.1022 found 397.1028.



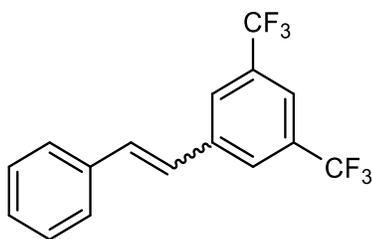
4'-Chloro-3,5-bis(trifluoromethyl)-1,1'-biphenyl, 7c.²⁴ Starting from 0.697 mmol of corresponding organotin reagent the product was obtained as a colourless solid, yield 93% (0.210 g, method C). **¹H NMR** (400 MHz, CDCl₃): δ = 7.47-7.50 (m, 2H, Ar), 7.54-7.57 (m, 2H, Ar), 7.89 (s, 1H, Ar), 7.99 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 121.4 (hept, *J* = 3.9 Hz), 123.5 (q, *J* = 271 Hz), 127.2 (q, *J* = 3.8 Hz), 128.7, 129.7, 132.5 (q, *J* = 33.4 Hz), 135.5, 136.8, 142.3.



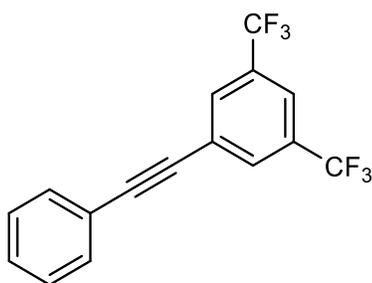
2-(3,5-Bis(trifluoromethyl)phenyl)thiophene, 7d.²⁵ Starting from 0.670 mmol of corresponding organotin reagent the product was obtained as a colourless solid, yield 99% (0.198 g, method D). **¹H NMR** (400 MHz, CDCl₃): δ = 7.15 (dd, *J* = 5.1, 3.7 Hz, 1H, thiophene), 7.41-7.44 (m, 2H, thiophene), 7.79 (s, 1H, Ar), 8.01 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 120.9 (hept, *J* = 3.8 Hz), 123.4 (q, *J* = 271 Hz), 125.4, 125.8 (q, *J* = 3.9 Hz), 127.2, 128.7, 132.5 (q, *J* = 33.4 Hz), 136.7, 141.1.



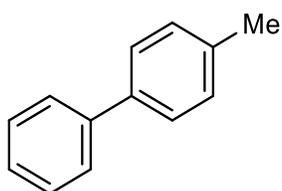
2-(3,5-Bis(trifluoromethyl)phenyl)benzo[b]thiophene, 7e.²⁴ Starting from 0.709 mmol of corresponding organotin reagent the product was obtained as a colourless solid, yield 99% (0.245 g, method D). **¹H NMR** (400 MHz, CDCl₃): δ = 7.36-7.43 (m, 2H, Ar), 7.62 (s, 1H, Ar), 7.78-7.85 (m, 3H, Ar), 8.08 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 121.6 (hept, *J* = 3.8 Hz), 122.0, 122.6, 123.4 (q, *J* = 271 Hz), 124.4, 125.2, 125.6, 126.2 (q, *J* = 3.8 Hz), 132.6 (q, *J* = 33.4 Hz), 136.6, 140.0, 140.4, 140.5.



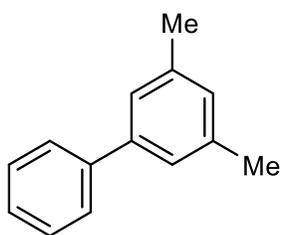
1-Styryl-3,5-bis(trifluoromethyl)benzene, 7f.³⁵ Starting from 0.712 mmol of corresponding organothin reagent the product was obtained as a colourless oil, yield 82%, E/Z = 10/1.2 (0.185 g, method C). ¹H NMR (400 MHz, CDCl₃): δ = 6.60 (d, *J* = 12.1 Hz, 0.12H, olefin, minor isomer), 6.85 (d, *J* = 12.2 Hz, 0.12H, olefin, minor isomer), 7.13 (d, *J* = 16.4 Hz, 1H, olefin, major isomer), 7.18-7.30 (m, 1.84H, Ar, minor isomer/olefin, major isomer), 7.33-7.38 (m, 1H, Ar, major isomer), 7.40-7.44 (m, 2H, Ar, major isomer), 7.50-7.52 (m, 0.14H, Ar, minor isomer), 7.54-7.57 (m, 2H, Ar, major isomer), 7.67-7.70 (m, 0.40H, Ar, minor isomer), 7.77 (s, 1H, Ar, major isomer), 7.92 (s, 2H, Ar, major isomer), 8.02 (s, 0.17H, Ar, minor isomer), 8.04 (s, 0.34H, Ar, minor isomer). ¹³C NMR (101 MHz, CDCl₃): δ = 120.7-120.8 (m), 120.9 (p, *J* = 3.9 Hz), 123.2 (q, *J* = 271 Hz), 123.6 (q, *J* = 271 Hz), 125.7, 126.3 (q, *J* = 3.7 Hz), 127.1, 127.2, 127.3, 127.6, 127.7, 127.9, 128.1, 128.3, 128.8, 129.0, 129.1, 132.2 (q, *J* = 33.0 Hz), 132.7, 133.1 (q, *J* = 33.0 Hz), 134.1, 135.9, 136.2, 138.1, 139.4, 139.6, 140.6, 142.6.



1-(Phenylethynyl)-3,5-bis(trifluoromethyl)benzene, 7g.³⁶ Starting from 0.716 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 77% (0.173 g, method C). ¹H NMR (400 MHz, CDCl₃): δ = 7.38-7.43 (m, 3H, Ar), 7.56-7.61 (m, 2H, Ar), 7.83 (s, 1H, Ar), 7.97 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 86.5, 93.0, 121.7 (p, *J* = 3.8 Hz), 122.1, 123.2 (q, *J* = 271 Hz), 125.9, 128.8, 129.5, 131.6 (q, *J* = 3.9 Hz), 132.0, 132.2 (q, *J* = 33.0 Hz).



4-Methyl-1,1'-biphenyl, 7i.³⁷ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 83% (0.095 g, X = Br, method D), 94% (0.108 g, X = Cl, method D). ¹H NMR (400 MHz, CDCl₃): δ = 2.49 (s, 3H, Me), 7.33-7.36 (m, 2H, Ar), 7.40-7.44 (m, 1H, Ar), 7.50-7.54 (m, 2H, Ar), 7.58-7.62 (m, 2H, Ar), 7.67-7.70 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.3, 127.1, 127.2, 128.9, 129.7, 137.2, 138.5, 141.3.



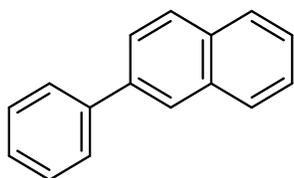
3,5-Dimethyl-1,1'-biphenyl, 7j.³⁸ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 99% (0.124 g, method D). ¹H NMR (400 MHz, CDCl₃): δ = 2.49 (s, 6H, 2xMe), 7.10 (s, 1H, Ar), 7.33 (s, 2H, Ar), 7.41-7.45 (m, 1H, Ar), 7.50-7.55 (m, 2H, Ar), 7.68-7.71 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.6, 125.3, 127.2, 127.4, 128.8, 129.1, 138.4, 141.4, 141.6.

³⁵ A. N. Baumann, A. Music, J. Dechent, N. Meller, T. C. Jagau and D. Didier, *Chem. Eur. J.*, 2020, **26**, 8382-8387.

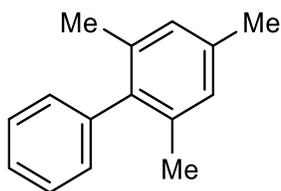
³⁶ R. Zhou, W. Wang, Z.-j. Jiang, H.-y. Fu, X.-l. Zheng, C.-c. Zhang, H. Chen and R.-x. Li, *Catal. Sci. Technol.*, 2014, **4**, 746-751.

³⁷ J. Ishida, M. Nakatsuji, T. Nagata, H. Kawasaki, T. Suzuki and Y. Obora, *ACS Omega*, 2020, **5**, 9598-9604.

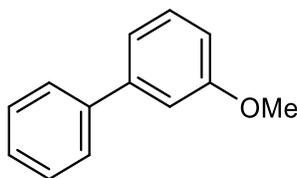
³⁸ D. Kim, G. Choi, W. Kim, D. Kim, Y. K. Kang and S. H. Hong, *Chem. Sci.*, 2021, **12**, 363-373.



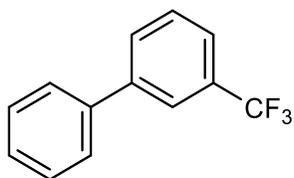
2-Phenylnaphthalene, 7k.³⁹ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 99% (0.138 g, method D). ¹H NMR (400 MHz, CDCl₃): δ = 7.44-7.48 (m, 1H, Ar), 7.53-7.60 (m, 4H, Ar), 7.79-7.85 (m, 3H, Ar), 7.92-7.99 (m, 3H, Ar), 8.13 (d, *J* = 1.9 Hz, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 125.8, 126.0, 126.1, 126.4, 127.5, 127.6, 127.8, 128.4, 128.6, 129.0, 132.8, 133.9, 138.7, 141.3.



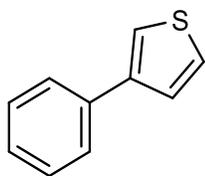
2,4,6-Trimethyl-1,1'-biphenyl, 7l.⁴⁰ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless oil, yield 38% (0.051 g, method D). ¹H NMR (400 MHz, CDCl₃): δ = 2.08 (s, 6H, 2xMe), 2.40 (s, 3H, Me), 7.01 (s, 2H, Ar), 7.19-7.22 (m, 2H, Ar), 7.36-7.42 (m, 1H, Ar), 7.45-7.51 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 20.9, 21.2, 126.7, 127.3, 127.4, 128.2, 128.5, 128.9, 129.5, 136.1, 136.7, 139.2, 141.3.



3-Methoxy-1,1'-biphenyl, 7m.⁴¹ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 85% (0.106 g, X = Br, method D), 99% (0.125 g, X = Cl, method D). ¹H NMR (400 MHz, CDCl₃): δ = 3.90 (m, 3H, OMe), 6.93-6.97 (m, 1H, Ar), 7.18-7.20 (m, 1H, Ar), 7.22-7.25 (m, 1H, Ar), 7.37-7.43 (m, 2H, Ar), 7.46-7.50 (m, 2H, Ar), 7.63-7.66 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 55.4, 112.8, 113.1, 119.8, 127.4, 127.6, 128.9, 129.9, 141.3, 142.9, 160.1.



3-(Trifluoromethyl)-1,1'-biphenyl, 7n.⁴² Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless oil, yield 64% (0.097 g, method C). ¹H NMR (400 MHz, CDCl₃): δ = 7.41-7.46 (m, 1H, Ar), 7.48-7.53 (m, 2H, Ar), 7.56-7.65 (m, 4H, Ar), 7.78-7.81 (m, 1H, Ar), 7.88 (s, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 124.1 (qd, *J* = 3.8, 1.9 Hz), 124.4 (q, *J* = 271 Hz), 127.4, 128.1, 128.2, 129.2, 129.4, 130.6 (d, *J* = 1.5 Hz), 131.4 (q, *J* = 32.2 Hz), 136.7, 140.0, 142.2.



3-Phenylthiophene, 7o.⁴³ Starting from 0.681 mmol of corresponding organothin reagent the product was obtained as a colourless oil, yield 90% (0.098 g, method D). ¹H NMR (400 MHz, CDCl₃): δ = 7.35-7.40 (m, 1H, thiophene), 7.42-7.52 (m, 5H, Ar/thiophene), 7.66-7.70 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 119.9, 120.4, 126.3, 126.5, 126.6, 127.3, 129.0, 136.0, 142.5.

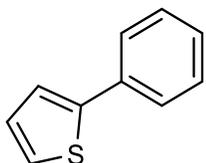
³⁹ M. M. Talukder, J. M. O. Cue, J. T. Miller, P. L. Gamage, A. Aslam, G. T. McCandless, M. C. Biewer and M. C. Stefan, *ACS Omega*, 2020, **5**, 24018-24032.

⁴⁰ G. Pandey and B. Torok, *Green Chem.*, 2017, **19**, 5390-5395.

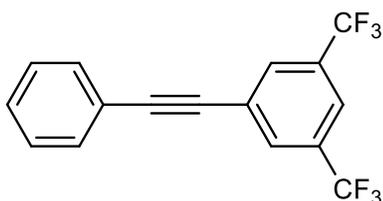
⁴¹ B. Liu, T. Xu, C. Li and J. Bai, *New J. Chem.*, 2020, **44**, 3794-3801.

⁴² P. P. Mpungose, N. I. Sehloko, G. E. M. Maguire and H. B. Friedrich, *New J. Chem.*, 2017, **41**, 13560-13566.

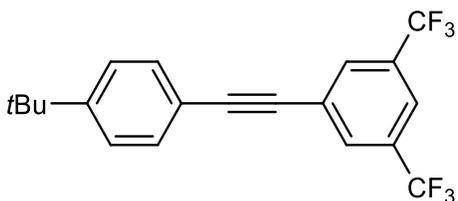
⁴³ J. Duczynski, A. N. Sobolev, S. A. Moggach, R. Dorta and S. G. Stewart, *Organometallics*, 2020, **39**, 105-115.



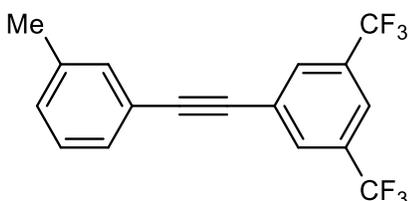
2-Phenylthiophene, 7p.⁴⁴ Starting from 0.670 mmol of corresponding organothin reagent the product was obtained as a colourless solid, yield 88% (0.094 g, X = Cl, method D), 71% (0.076 g, X = OTf, method D). **¹H NMR** (400 MHz, CDCl₃): δ = 7.12-7.15 (m, 1H, thiophene), 7.32-7.38 (m, 3H, Ar/thiophene), 7.41-7.46 (m, 2H, Ar), 7.67-7.70 (m, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 123.2, 124.9, 126.1, 127.6, 128.2, 129.0, 134.6, 144.6.



1-(Phenylethynyl)-3,5-bis(trifluoromethyl)benzene, 9a.⁴⁵ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a colourless oil, yield 86% (0.264 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 7.39-7.44 (m, 3H, Ar), 7.59-7.62 (m, 2H, Ar), 7.85 (s, 1H, Ar), 7.97 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 86.5, 93.0, 121.7 (hept, *J* = 3.8 Hz), 122.2, 123.2 (q, *J* = 271 Hz), 125.9, 128.7, 129.5, 131.6 (q, *J* = 3.9 Hz), 132.1, 132.2 (q, *J* = 34 Hz).



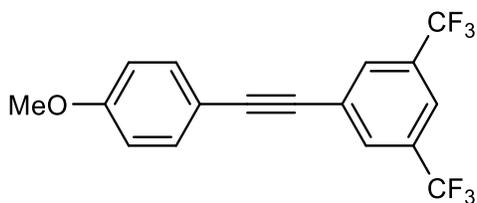
1-((4-(tert-Butyl)phenyl)ethynyl)-3,5-bis(trifluoromethyl)benzene, 9b. Starting from 0.948 mmol of corresponding acetylene the product was obtained as a white solid, yield 93% (0.325 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 1.37 (s, 9H, *t*Bu), 7.44 (d, *J* = 8.8 Hz, 2H, Ar), 7.53 (d, *J* = 8.8 Hz, 2H, Ar), 7.83 (s, 1H, Ar), 7.97 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 31.3, 35.1, 86.0, 93.4, 119.1, 121.5 (p, *J* = 3.8 Hz), 123.3 (q, *J* = 272 Hz), 125.8, 126.2, 131.6 (q, *J* = 4.0 Hz), 131.8, 132.1 (q, *J* = 34 Hz), 153.0. **HRMS-EI** (*m/z*) [M+H]⁺ calcd. for C₂₀H₁₇F₆ 371.1229 found 371.1231.



1-(m-Tolyethynyl)-3,5-bis(trifluoromethyl)benzene, 9c. Starting from 0.947 mmol of corresponding acetylene the product was obtained as a colourless oil, yield 92% (0.285 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 2.40 (s, 3H, Me), 7.23 (d, *J* = 7.7 Hz, 1H, Ar), 7.30 (t, *J* = 7.6 Hz, 1H, Ar), 7.39-7.42 (m, 2H, Ar), 7.83 (s, 1H, Ar), 7.96 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 21.4, 86.2, 93.3, 121.6 (hept, *J* = 3.9 Hz), 121.9, 123.2 (q, *J* = 271 Hz), 126.0, 128.6, 129.1, 130.4, 131.6 (q, *J* = 4.0 Hz), 132.1 (q, *J* = 33 Hz), 132.6, 138.5. **HRMS-EI** (*m/z*) [M+H]⁺ calcd. for C₁₇H₁₁F₆ 329.0759 found 329.0755.

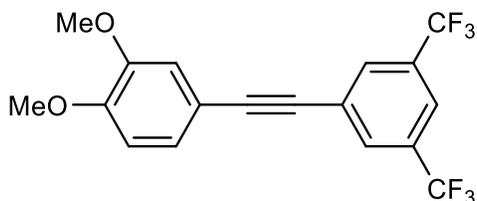
⁴⁴ B. Karimi, M. Tavakolian, F. Mansouri and H. Vali, *ACS Sustainable Chem. Eng.*, 2019, **7**, 3811-3823.

⁴⁵ R. Zhou, W. Wang, Z.-j. Jiang, H.-y. Fu, X.-l. Zheng, C.-c. Zhang, H. Chen and R.-x. Li, *Catal. Sci. Technol.*, 2014, **4**, 746-751.



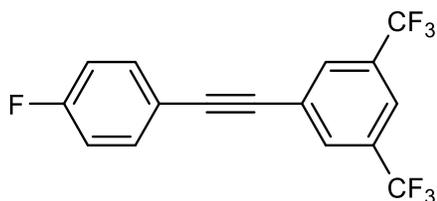
1-((4-Methoxyphenyl)ethynyl)-3,5-bis(trifluoromethyl)benzene, 9d.⁴⁶ Starting from 0.984 mmol of corresponding acetylene the product was obtained as a white solid, yield 87% (0.295 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 3.84 (s, 3H, OMe), 6.92 (d, *J* = 8.4 Hz, 2H, Ar), 7.50-7.52 (m, 2H, Ar), 7.80 (s, 1H, Ar), 7.93 (s, 2H, Ar). ¹³C

NMR (101 MHz, CDCl₃): δ = 55.4, 85.5, 93.3, 114.1, 114.4, 121.3 (p, *J* = 3.9 Hz), 123.2 (q, *J* = 271 Hz), 126.3, 131.4 (q, *J* = 3.8 Hz), 132.1 (q, *J* = 33.6 Hz), 133.6, 160.6.



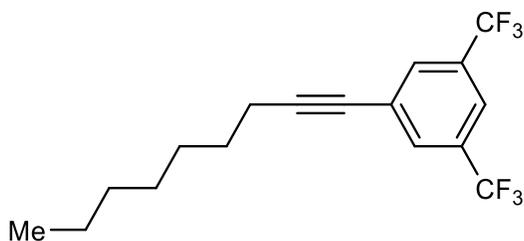
4-((3,5-bis(Trifluoromethyl)phenyl)ethynyl)-1,2-dimethoxybenzene, 9e. Starting from 0.987 mmol of corresponding acetylene the product was obtained as a white solid, yield 97% (0.358 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 3.88 (d, *J* = 2.5 Hz, 6H, 2xOMe), 6.83 (d, *J* = 8.3 Hz, 1H, Ar), 7.03 (d, *J* = 1.8 Hz, 1H, Ar), 7.14 (dd, *J* = 8.3, 1.9

Hz, 1H, Ar), 7.76 (s, 1H, Ar), 7.90 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 55.9, 56.0, 85.2, 93.3, 111.2, 114.1, 114.4, 121.2 (hept, *J* = 3.9 Hz), 123.1 (q, *J* = 272 Hz), 125.5, 126.1, 131.3 (q, *J* = 3.9 Hz), 132.0 (q, *J* = 33.6 Hz), 148.9, 150.5. **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₈H₁₃F₆O₂ 375.0814 found 375.0807.



1-((4-Fluorophenyl)ethynyl)-3,5-bis(trifluoromethyl)benzene, 9f. Starting from 0.999 mmol of corresponding acetylene the product was obtained as a white solid, yield 93% (0.307 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 7.06-7.11 (m, 2H, Ar), 7.55 (dd, *J* = 8.4, 5.3 Hz, 2H, Ar), 7.83 (s, 1H, Ar), 7.94 (s, 2H, Ar). ¹³C

NMR (101 MHz, CDCl₃): δ = 86.3, 91.9, 116.1 (d, *J* = 22.2 Hz), 118.3 (d, *J* = 3.5 Hz), 121.7-121.9 (m), 123.2 (q, *J* = 271 Hz), 125.8, 131.6 (q, *J* = 3.7 Hz), 132.2 (q, *J* = 33.7 Hz), 134.0 (d, *J* = 8.5 Hz), 163.4 (d, *J* = 251.4 Hz). **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₆H₈F₇ 333.0509 found 333.0511.

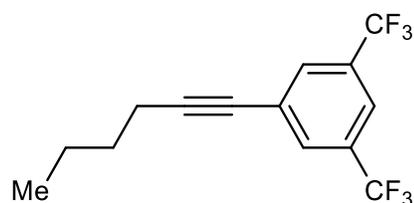


1-(Non-1-yn-1-yl)-3,5-bis(trifluoromethyl)benzene, 9g.

Starting from 0.966 mmol of corresponding acetylene the product was obtained as a colourless oil, yield 72% (0.235 g, method A). ¹H NMR (400 MHz, CDCl₃): δ = 0.87-0.92 (m, 3H, Me), 1.28-1.39 (m, 6H, 3xCH₂), 1.46 (p, *J* = 6.7 Hz, 2H, CH₂), 1.63 (p, *J* = 7.2 Hz, 2H, CH₂), 2.44 (t, *J* = 7.1 Hz,

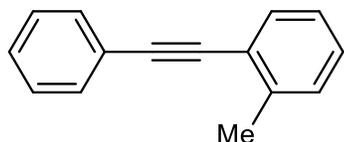
2H, CH₂), 7.75 (s, 1H, Ar), 7.81 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 14.2, 19.6, 22.9, 28.6, 29.0, 29.1, 31.9, 78.3, 94.9, 121.0 (p, *J* = 3.7 Hz), 123.3 (q, *J* = 271 Hz), 126.7, 131.7 (q, *J* = 3.9 Hz), 132.0 (q, *J* = 33 Hz). **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₇H₁₉F₆ 337.1385 found 337.1383.

⁴⁶ J. He, K. Yang, J. Zhao and S. Cao, *Org. Lett.*, 2019, **21**, 9714-9718.



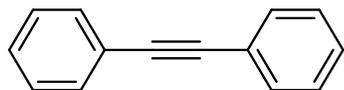
1-(Hex-1-yn-1-yl)-3,5-bis(trifluoromethyl)benzene, 9h.⁴⁷ Starting from 0.974 mmol of corresponding acetylene the product was obtained as a colourless oil, yield 99% (0.286 g, method A). **¹H NMR** (400 MHz, CDCl₃): δ = 0.97 (t, *J* = 7.3 Hz, 3H, Me), 1.45-1.54 (m, 2H, CH₂), 1.58-1.66 (m, 2H, CH₂), 2.44 (t, *J* = 7.0 Hz, 2H, CH₂), 7.74 (s, 1H, Ar), 7.81 (s, 2H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ =

13.7, 19.2, 22.3, 30.7, 78.3, 94.8, 121.0 (hept, *J* = 3.9 Hz), 123.3 (q, *J* = 271 Hz), 126.7, 131.7 (q, *J* = 3.8 Hz), 132.0 (q, *J* = 33 Hz).



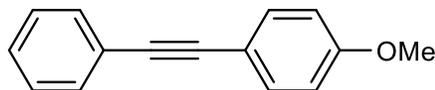
1-Methyl-2-(phenylethynyl)benzene, 9i.⁴⁸ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 58% (0.109 g, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 2.58-2.59 (m, 3H, Me), 7.20-7.25 (m, 1H, Ar), 7.28-7.29 (m, 2H, Ar), 7.35-7.42 (m, 3H, Ar), 7.55-7.62 (m, 3H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 88.5, 93.5, 123.2, 123.7, 125.8, 128.3,

128.4, 128.5, 129.6, 131.7, 132.0, 140.3.



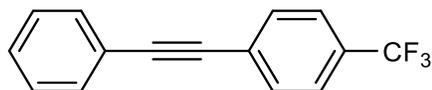
1,2-Diphenylethyne, 9j.⁴⁹ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 99% (0.174 g, X = Br, method B), 90% (0.157 g, X = OTf, method B). **¹H NMR** (400

MHz, CDCl₃): δ = 7.36-7.43 (m, 6H, Ar), 7.59-7.63 (m, 4H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 89.6, 123.4, 128.4, 128.5, 131.8.



1-Methoxy-4-(phenylethynyl)benzene, 9k.⁵⁰ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 97% (0.197 g, method B). **¹H NMR** (400 MHz,

CDCl₃): δ = 3.83 (s, 3H, OMe), 6.90-6.94 (m, 2H, Ar), 7.33-7.41 (m, 3H, Ar), 7.52-7.61 (m, 4H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 55.3, 88.2, 89.6, 114.1, 115.5, 123.7, 128.1, 128.4, 131.6, 133.2, 159.7.



1-(Phenylethynyl)-4-(trifluoromethyl)benzene, 9l.⁵¹ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 91% (0.220 g, X = Br, method A), 7% (0.018

g, X = Cl, method B). **¹H NMR** (400 MHz, CDCl₃): δ = 7.39-7.42 (m, 3H, Ar), 7.59-7.68 (m, 6H, Ar). **¹³C NMR** (101 MHz, CDCl₃): δ = 88.2, 92.0, 122.8, 124.2 (q, *J* = 270 Hz), 125.4 (q, *J* = 3.8 Hz), 127.3 (q, *J* = 1.5 Hz), 128.6, 129.0, 130.1 (q, *J* = 32.6 Hz), 131.9, 132.0.

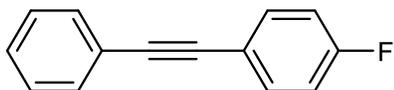
⁴⁷ S. R. Chidipudi, I. Khan and H. W. Lam, *Angew. Chem. Int. Ed.*, 2012, **51**, 12115-12119.

⁴⁸ F. Messa, G. Dilauro, F. M. Perna, P. Vitale, V. Capriati and A. Salomone, *ChemCatChem*, 2020, **12**, 1979-1984.

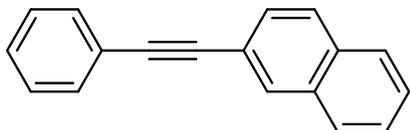
⁴⁹ S. Alapour, M. D. Farahani, D. Ramjugernath, N. A. Koobanally and H. B. Friedrich, *ACS Sustainable Chem. Eng.*, 2019, **7**, 12697-12706.

⁵⁰ G. Hamasaka, D. Roy, A. Tazawa and Y. Uozumi, *ACS Catal.*, 2019, **9**, 11640-11646.

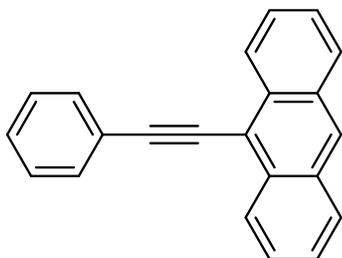
⁵¹ T. Tani, Y. Sawatsugawa, Y. Sano, Y. Hirataka, N. Takahashi, S. Hashimoto, T. Sugiura and T. Tsuchimoto, *Adv. Synth. Catal.*, 2019, **361**, 1815-1834.



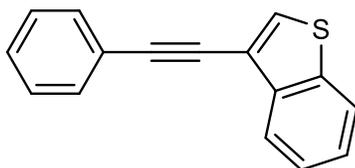
1-Fluoro-4-(phenylethynyl)benzene, 9m.⁵² Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 70% (0.134 g, method A), 99% (0.191 g, method B). For gram-scale experiment starting from 9.790 mmol (1 g) of corresponding acetylene the product was obtained as a white solid, yield 87% (1.673 g). ¹H NMR (400 MHz, CDCl₃): δ = 7.04-7.10 (m, 2H, Ar), 7.36-7.41 (m, 3H, Ar), 7.52-7.59 (m, 4H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 88.5, 89.2 (d, *J* = 1.5 Hz), 115.8 (d, *J* = 22.0 Hz), 119.5 (d, *J* = 3.4 Hz), 123.3, 128.4, 128.5, 131.7, 133.6 (d, *J* = 8.3 Hz), 162.6 (d, *J* = 249.5 Hz).



2-(Phenylethynyl)naphthalene, 9n.⁵³ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a yellow solid, yield 99% (0.223 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.40-7.48 (m, 3H, Ar), 7.54-7.59 (m, 2H, Ar), 7.68-7.72 (m, 3H, Ar), 7.87-7.90 (m, 3H, Ar), 8.16 (s, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 89.9, 90.0, 120.7, 123.5, 126.7, 126.8, 127.9, 127.9, 128.2, 128.4, 128.5, 128.6, 131.6, 131.8, 132.9, 133.2.



9-(Phenylethynyl)anthracene, 9o.⁵⁴ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a yellow solid, yield 98% (0.268 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.49-7.61 (m, 5H, Ar), 7.72 (ddd, *J* = 8.5, 6.6, 1.3 Hz, 2H, Ar), 7.92-7.95 (m, 2H, Ar), 8.05 (d, *J* = 8.5 Hz, 2H, Ar), 8.41 (s, 1H, Ar), 8.83 (dd, *J* = 8.7, 1.2 Hz, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 86.6, 100.9, 117.3, 123.8, 125.7, 126.7, 126.8, 127.8, 128.5, 128.6, 128.8, 131.2, 131.8, 132.7.



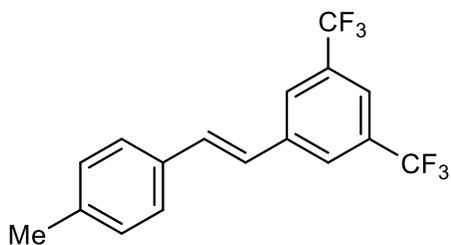
3-(Phenylethynyl)benzo[b]thiophene, 9p.⁵⁵ Starting from 0.979 mmol of corresponding acetylene the product was obtained as a white solid, yield 97% (0.223 g, method B). ¹H NMR (400 MHz, CDCl₃): δ = 7.37-7.50 (m, 4H, Ar), 7.51-7.56 (m, 1H, Ar), 7.66-7.70 (m, 2H, Ar), 7.73 (s, 1H, Ar), 7.90-7.92 (m, 1H, Ar), 8.11-8.14 (m, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 83.2, 92.0, 118.5, 122.8, 123.2, 123.3, 124.9, 125.2, 128.5, 128.6, 129.9, 131.8, 139.0, 139.3.

⁵² S. Ruengsangtongkul, N. Chaisan, C. Thongsornkleeb, J. Tummatorn and S. Ruchirawat, *Org. Lett.*, 2019, **21**, 2514-2517.

⁵³ A. Baralle, H. Yorimitsu and A. Osuka, *Chem. Eur. J.*, 2016, **22**, 10768-10772.

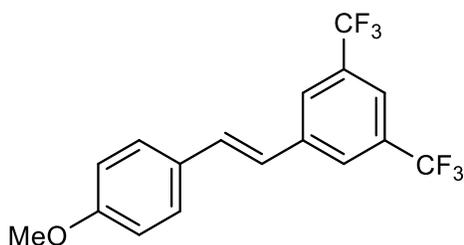
⁵⁴ Y. Thummala, A. K. Morri, G. V. Karunakar and V. R. Doddi, *Eur. J. Org. Chem.*, 2018, 6280-6285.

⁵⁵ S. Prateptongkum, K. M. Driller, R. Jackstell, A. Spannenberg and M. Beller, *Chem. Eur. J.*, 2010, **16**, 9606-9615.



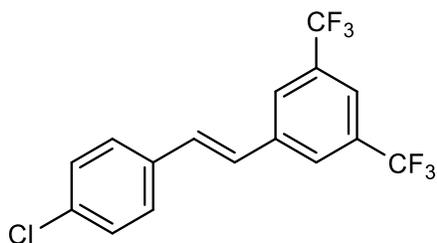
(E)-1-(4-Methylstyryl)-3,5-bis(trifluoromethyl)benzene, 11a.⁵⁶

Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 94% (0.262 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 2.53 (s, 3H, Me), 7.20 (d, *J* = 16.5 Hz, 1H, olefin), 7.32-7.36 (m, 3H, Ar/olefin), 7.58 (d, *J* = 8.5 Hz, 2H, Ar), 7.88 (s, 1H, Ar), 8.03 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 120.7 (hept, *J* = 3.9 Hz), 123.6 (q, *J* = 271 Hz), 124.7, 126.2 (q, *J* = 3.7 Hz), 127.1, 127.7, 127.9, 129.5, 129.8, 132.2 (q, *J* = 33 Hz), 132.6, 133.4, 139.1, 139.8.



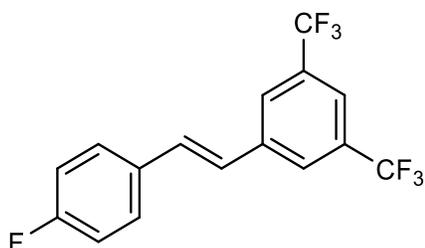
(E)-1-(4-Methoxystyryl)-3,5-bis(trifluoromethyl)benzene, 11b.

Starting from 0.894 mmol of corresponding olefin the product was obtained as a colourless solid, yield 94% (0.291 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 3.85 (s, 3H, OMe), 6.94-6.99 (m, 3H, Ar/olefin), 7.18 (d, *J* = 16.3 Hz, 1H, olefin), 7.47-7.51 (m, 2H, Ar), 7.73 (s, 1H, Ar), 7.87 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 55.4, 114.5, 120.4 (hept, *J* = 3.8 Hz), 123.4, 123.6 (q, *J* = 271 Hz), 126.0 (q, *J* = 4.0 Hz), 128.5, 128.9, 132.1 (q, *J* = 33 Hz), 132.2, 140.0, 160.4. **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₇H₁₃F₆O 347.0865 found 347.0862.



(E)-1-(4-Chlorostyryl)-3,5-bis(trifluoromethyl)benzene, 11c.

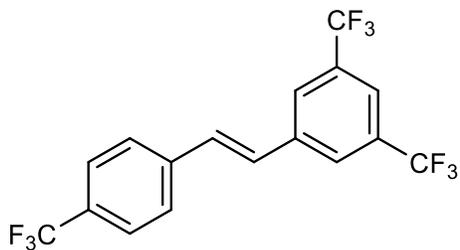
Starting from 0.866 mmol of corresponding olefin the product was obtained as a colourless solid, yield 97% (0.295 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 7.07 (d, *J* = 16.6 Hz, 1H, olefin), 7.17 (d, *J* = 16.4 Hz, 1H, olefin), 7.35-7.38 (m, 2H, Ar), 7.45-7.47 (m, 2H, Ar), 7.77 (s, 1H, Ar), 7.90 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.2 (p, *J* = 3.9 Hz), 123.5 (q, *J* = 271 Hz), 126.2, 126.3 (q, *J* = 3.7 Hz), 128.2, 129.1, 129.2, 129.3, 131.3, 132.3 (q, *J* = 33.2 Hz), 134.7, 139.3. **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₆H₁₀ClF₆ 351.0370 found 351.0377.



(E)-1-(4-Fluorostyryl)-3,5-bis(trifluoromethyl)benzene, 11d.

Starting from 0.819 mmol of corresponding olefin the product was obtained as a colourless oil, yield 82% (0.225 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 7.02 (d, *J* = 16.3 Hz, 1H, olefin), 7.07-7.12 (m, 2H, Ar), 7.19 (d, *J* = 16.3 Hz, 1H, olefin), 7.49-7.53 (m, 2H, Ar), 7.76 (s, 1H, Ar), 7.89 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 116.1 (d, *J* = 21.9 Hz), 121.0 (hept, *J* = 3.8 Hz), 123.6 (q, *J* = 271 Hz), 125.5 (d, *J* = 2.5 Hz), 126.2 (q, *J* = 3.8 Hz), 128.7 (d, *J* = 8.1 Hz), 131.4, 132.3 (q, *J* = 33 Hz), 132.4, 139.5, 163.2 (d, *J* = 249.1 Hz). **HRMS-EI** (m/z) [M+H]⁺ calcd. for C₁₆H₁₀F₇ 335.0665 found 335.0670.

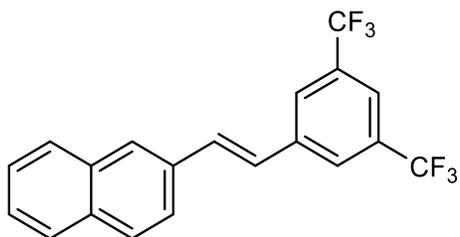
⁵⁶ M. S. M. Pearson and D. R. Carbery, *J. Org. Chem.*, 2009, **74**, 5320-5325.



(E)-1,3-Bis(trifluoromethyl)-5-(4-(trifluoromethyl)styryl)benzene, 11e.⁵⁷

Starting from 0.871 mmol of corresponding olefin the product was obtained as a colourless oil, yield 80% (0.269 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 7.19 (d, *J* = 16.3 Hz, 1H, olefin), 7.25 (d, *J* = 16.2 Hz, 1H, olefin), 7.61-7.67 (m, 4H, Ar), 7.79 (s, 1H, Ar), 7.94 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 121.6 (p, *J* = 3.8

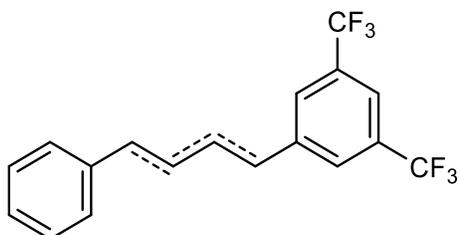
Hz), 123.9 (q, *J* = 271 Hz), 124.3 (q, *J* = 271 Hz), 126.1 (q, *J* = 3.8 Hz), 126.6 (q, *J* = 3.6 Hz), 127.2, 128.1, 130.7 (q, *J* = 33.4 Hz), 131.1, 132.5 (q, *J* = 33.4 Hz), 139.0, 139.6-139.7 (m).



(E)-2-(3,5-Bis(trifluoromethyl)styryl)naphthalene, 11f.

Starting from 0.843 mmol of corresponding olefin the product was obtained as a colourless solid, yield 93% (0.286 g, method E). ¹H NMR (400 MHz, CDCl₃): δ = 7.13 (d, *J* = 16.2 Hz, 1H, olefin), 7.32 (d, *J* = 16.4 Hz, 1H, olefin), 7.54-7.60 (m, 2H, Ar), 7.72 (d, *J* = 8.6 Hz, 1H, Ar), 7.87-7.90 (m, 5H, Ar), 7.94 (s, 2H, Ar). ¹³C

NMR (101 MHz, CDCl₃): δ = 120.8 (p, *J* = 3.9 Hz), 123.3, 123.6 (q, *J* = 271 Hz), 125.6, 126.2 (q, *J* = 3.9 Hz), 126.7, 126.8, 127.9, 128.0, 128.4, 128.7, 132.1 (q, *J* = 33.4 Hz), 132.5, 133.6, 133.7, 133.7, 139.5. HRMS-EI (m/z) [M+H]⁺ calcd. for C₂₀H₁₃F₆ 367.0916 found 367.0910.



Mixture of 1-(4-phenylbut-1-en-1-yl)-3,5-bis(trifluoromethyl)benzene, 1-(4-phenylbut-2-en-1-yl)-3,5-bis(trifluoromethyl)benzene and 1-(4-phenylbut-3-en-1-yl)-3,5-bis(trifluoromethyl)benzene, 11g.⁵⁸

Starting from 0.832 mmol of corresponding olefin the product was obtained as a colourless oil, yield 82% (0.235 g, method E).

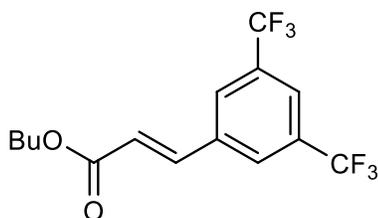
Fraction 1: ¹H NMR (400 MHz, CDCl₃): δ = 2.57-2.62 (m, 1.81H, CH₂), 2.78-2.87 (m, 2.45H, CH₂), 3.45 (dd, *J* = 27.6, 6.8 Hz, 0.27H, CH₂), 3.61 (d, *J* = 7.6 Hz, 0.31H, CH₂), 6.43-6.49 (m, 1.67H, olefin), 7.15-7.19 (m, 0.61H, olefin), 7.22-7.27 (m, 3.25H, Ar), 7.29-7.34 (m, 2.59H, Ar), 7.65-7.73 (m, 3H, Ar), 7.79-7.82 (m, 0.73H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 16.0, 21.2, 34.7, 35.0, 35.3, 35.6, 37.0, 116.1, 120.5 (hept, *J* = 4.0 Hz), 121.2-121.3 (m), 122.3, 123.6 (q, *J* = 271 Hz), 125.0, 126.0 (q, *J* = 3.5 Hz), 126.3, 126.4, 126.5, 127.7, 127.8, 128.2, 128.3, 128.4, 128.6, 128.7, 128.8, 128.9, 130.5, 130.6, 131.5, 131.7, 132.0 (q, *J* = 33.4 Hz), 132.1, 132.4, 132.9, 133.0, 133.6, 134.5, 135.4, 140.0, 140.2, 141.2, 141.4, 143.6, 145.8.

Fraction 2: ¹H NMR (400 MHz, CDCl₃): δ = 3.03-3.10 (m, 2.77H, CH₂), 3.19 (t, *J* = 7.5 Hz, 0.28H, CH₂), 3.32 (t, *J* = 7.8 Hz, 1H, CH₂), 3.41 (t, *J* = 7.8 Hz, 1.78H, CH₂), 3.49 (t, *J* = 7.6 Hz, 0.29H, CH₂), 3.94 (dd, *J* = 28.2, 6.8 Hz, 0.44H, CH₂), 6.65-6.73 (m, 0.89H, olefin), 6.90-6.94 (m, 1.81H, olefin), 7.67-7.72 (m, 3.28H, Ar), 7.77-7.83 (m, 4.91H, Ar), 8.13-8.23 (m, 4.57H, Ar), 8.38 (s, 0.44H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 32.3, 34.3, 34.5, 35.0, 35.6, 35.7, 38.6, 39.1, 120.3 (p, *J* = 4.0 Hz), 120.5 (p, *J* =

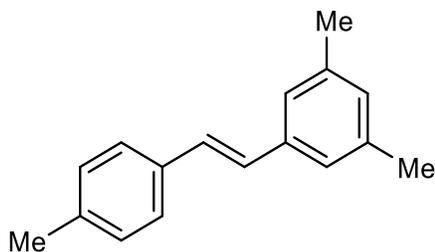
⁵⁷ P. d. Medina, R. Casper, J.-F. Savouret and M. Poirot, *J. Med. Chem.*, 2005, **48**, 287-291.

⁵⁸ C.-C. Tseng, M. Li, B. Mo, S. A. Warren and A. C. Spivey, *Chem. Lett.*, 2011, **40**, 995-997.

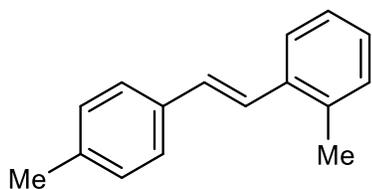
3.8 Hz), 123.5 (q, $J = 272$ Hz), 123.6 (q, $J = 271$ Hz), 126.0 (q, $J = 4.1$ Hz), 126.2, 126.3, 126.4, 126.8, 127.5, 128.2, 128.3, 128.4, 128.5, 128.6, 128.7, 128.8, 128.8-128.9 (m), 129.3, 131.7, 131.8 (q, $J = 32$ Hz), 132.0 (q, $J = 33.4$ Hz), 133.0, 134.5, 137.4, 139.0, 139.9, 140.0, 140.3, 141.4, 143.1, 143.4, 144.0, 144.2.



Butyl (E)-3-(3,5-bis(trifluoromethyl)phenyl)acrylate, 11h.⁵⁹ Starting from 0.858 mmol of corresponding olefin the product was obtained as a colourless solid, yield 90% (0.262 g, method F). ¹H NMR (400 MHz, CDCl₃): $\delta = 0.95$ (t, $J = 7.4$ Hz, 3H, Me), 1.44 (h, $J = 7.4$ Hz, 2H, CH₂), 1.69 (p, $J = 6.4$ Hz, 2H, CH₂), 4.23 (t, $J = 7.4$ Hz, 2H, CH₂), 6.58 (d, $J = 16.0$ Hz, 1H, olefin), 7.71 (d, $J = 16.1$ Hz, 1H, olefin), 7.85 (s, 1H, Ar), 7.94 (s, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): $\delta = 13.8$, 19.3, 30.9, 65.1, 122.5, 123.2 (q, $J = 272$ Hz), 123.4 (hept, $J = 3.8$ Hz), 127.8 (q, $J = 3.9$ Hz), 132.6 (q, $J = 33.6$ Hz), 136.8, 141.0, 166.1.



(E)-1,3-Dimethyl-5-(4-methylstyryl)benzene, 11k.⁶⁰ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 86% (0.161 g, method E), 86% (0.162 g, method F). For isolations based on the use of renewable monoterpenes starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 90% (0.170 g, (-)- α -pinene), 91% (0.172 g, 3-carene), 86% (0.161 g, (R)-(+)-limonene), 89% (0.167 g, γ -terpinene), 87% (0.163 g, sabinene). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.65$ -2.67 (m, 9H, 3xMe), 7.21 (s, 1H, Ar), 7.32 (d, $J = 16.4$ Hz, 1H, olefin), 7.39 (d, $J = 16.2$ Hz, 1H, olefin), 7.45-7.48 (m, 4H, Ar), 7.72 (d, $J = 7.8$ Hz, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): $\delta = 21.4$, 21.5, 124.5, 126.5, 128.0, 128.3, 129.4, 129.5, 134.9, 137.4, 137.6, 138.1.

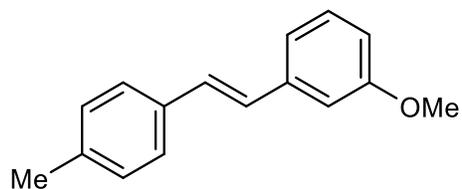


(E)-1-Methyl-2-(4-methylstyryl)benzene, 11l.⁶¹ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless oil, yield 58% (0.102 g, method E), 95% (0.167 g, method F). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.48$ (s, 3H, Me), 2.55 (s, 3H, Me), 7.10 (d, $J = 16.1$ Hz, 1H, olefin), 7.28-7.35 (m, 5H, Ar), 7.42 (d, $J = 16.1$ Hz, 1H, olefin), 7.54 (d, $J = 8.4$ Hz, 2H, Ar), 7.71 (d, $J = 7.9$ Hz, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): $\delta = 20.1$, 21.4, 125.4, 125.7, 126.3, 126.6, 127.5, 129.5, 130.1, 130.5, 135.1, 135.8, 136.7, 137.6.

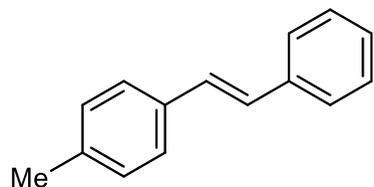
⁵⁹ M. Feuerstein, H. Doucet and M. Santelli, *J. Org. Chem.*, 2001, **66**, 5923-5925.

⁶⁰ K. Song, P. Liu, J. Wang, L. Pang, J. Chen, I. Hussain, B. Tan and T. Li, *Dalton Trans.*, 2015, **44**, 13906-13913.

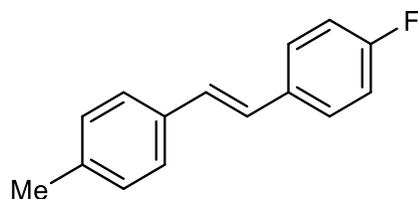
⁶¹ E. Shirakawa, X. Zhang and T. Hayashi, *Angew. Chem. Int. Ed.*, 2011, **50**, 4671-4674.



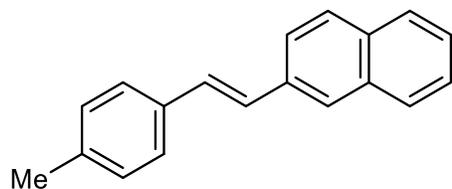
(E)-1-Methoxy-3-(4-methylstyryl)benzene, 11m.⁶² Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 87% (0.165 g, X = Br, method F), 71% (0.134 g, X = Cl, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.61 (s, 3H, Me), 4.08 (s, 3H, OMe), 7.06 (d, *J* = 8.2 Hz, 1H, olefin), 7.30-7.36 (m, 4H, Ar/olefin), 7.42 (d, *J* = 7.8 Hz, 2H, Ar), 7.52 (t, *J* = 8.0 Hz, 1H, Ar), 7.66 (d, *J* = 7.1 Hz, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 55.3, 111.8, 113.2, 119.3, 126.6, 127.7, 129.1, 129.5, 129.7, 134.6, 137.7, 139.1, 160.0.



(E)-1-Methyl-4-styrylbenzene, 11n.⁶³ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 99% (0.164 g, X = Br, method F), 0% (0 g, X = OTf, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.66 (s, 3H, Me), 7.34-7.43 (m, 2H, olefin), 7.47 (d, *J* = 7.8 Hz, 2H, Ar), 7.54-7.57 (m, 1H, Ar), 7.63-7.67 (m, 2H, Ar), 7.72 (d, *J* = 8.5 Hz, 2H, Ar), 7.81 (d, *J* = 7.7 Hz, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 126.5, 126.6, 127.5, 127.8, 128.7, 128.8, 129.5, 134.7, 137.5, 137.6.



(E)-1-Fluoro-4-(4-methylstyryl)benzene, 11o.⁶⁴ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 73% (0.131 g, X = Br, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.40 (s, 3H, Me), 7.00-7.10 (m, 4H, Ar/olefin), 7.20 (d, *J* = 7.8 Hz, 2H, Ar), 7.43 (d, *J* = 7.8 Hz, 2H, Ar), 7.49 (dd, *J* = 8.4, 5.4 Hz, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 115.6, 115.8, 126.5, 126.6, 128.0 (d, *J* = 7.9 Hz), 128.6 (d, *J* = 2.5 Hz), 129.6, 133.9 (d, *J* = 3.4 Hz), 134.6, 137.7, 162.4 (d, *J* = 246.8 Hz).



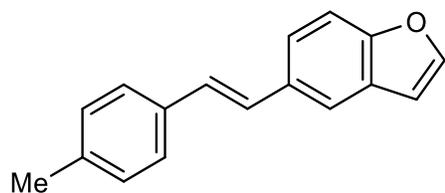
(E)-2-(4-Methylstyryl)naphthalene, 11p.⁶⁵ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless solid, yield 64% (0.132 g, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.39 (s, 3H, Me), 7.18-7.23 (m, 4H, Ar/olefin), 7.42-7.51 (m, 4H, Ar), 7.75 (dd, *J* = 8.6, 1.8 Hz, 1H, Ar), 7.80-7.86 (m, 4H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.5, 123.7, 126.0, 126.5, 126.6, 126.7, 127.9, 128.0, 128.1, 128.4, 129.2, 129.6, 133.1, 133.9, 134.8, 135.2, 137.8.

⁶² S. Kumar, A. K. Pandey, R. Singh and K. N. Singh, *Eur. J. Org. Chem.*, 2018, 5942-5946.

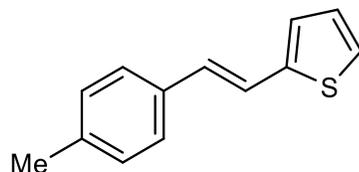
⁶³ D. Gong, B. Hu, W. Yang, D. Kong, H. Xia and D. Chen, *Organometallics*, 2020, **39**, 862-869.

⁶⁴ A. L. Isfahani, I. Mohammadpoor-Baltork, V. Mirkhani, A. R. Khosropour, M. Moghadam, S. Tangestaninejad and R. Kia, *Adv. Synth. Catal.*, 2013, **355**, 957-972.

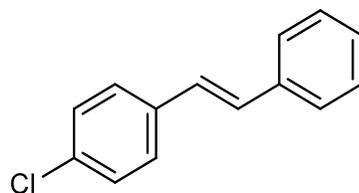
⁶⁵ S. M. Sutar, H. M. Savanur, S. S. Malunavar, P. Prabhala, R. G. Kalkhambkar and K. K. Laali, *Eur. J. Org. Chem.*, 2019, 6088-6093.



(E)-5-(4-Methylstyryl)benzofuran, 11q.⁶⁶ Starting from 0.846 mmol of corresponding olefin the product was obtained as a yellow solid, yield 69% (0.136 g, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.43 (s, 3H, Me), 6.80 (d, *J* = 2.2 Hz, 1H, Ar), 7.13 (d, *J* = 16.3 Hz, 1H, olefin), 7.21-7.25 (m, 3H, Ar/olefin), 7.48-7.50 (m, 2H, Ar), 7.54 (d, *J* = 1.3 Hz, 2H, Ar), 7.66 (d, *J* = 2.2 Hz, 1H, Ar), 7.75-7.76 (m, 1H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 106.8, 111.6, 119.2, 123.1, 126.4, 127.8, 128.0, 128.1, 129.5, 132.8, 134.9, 137.4, 145.6, 154.8.



(E)-2-(4-Methylstyryl)thiophene, 11r.⁶⁷ Starting from 0.846 mmol of corresponding olefin the product was obtained as a colourless oil, yield 37% (0.062 g, method F). ¹H NMR (400 MHz, CDCl₃): δ = 2.39 (s, 3H, Me), 6.95 (d, *J* = 16.1 Hz, 1H, olefin), 7.03 (dd, *J* = 5.1, 3.5 Hz, 1H, thiophene), 7.07-7.08 (m, 1H, thiophene), 7.18-7.24 (m, 4H, Ar/thiophene/olefin), 7.39-7.41 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 21.4, 121.0, 124.2, 125.9, 126.4, 127.7, 128.5, 129.6, 134.3, 137.7, 143.3.



(E)-1-Chloro-4-styrylbenzene, 11s.⁶⁸ Starting from 7.216 mmol (1 g) of corresponding olefin the product was obtained as a colourless solid, yield 96% (1.496 g). ¹H NMR (400 MHz, CDCl₃): δ = 7.03-7.12 (m, 2H, olefin), 7.27-7.40 (m, 5H, Ar), 7.43-7.47 (m, 2H, Ar), 7.51-7.53 (m, 2H, Ar). ¹³C NMR (101 MHz, CDCl₃): δ = 126.7, 127.5, 127.8, 128.0, 128.9, 129.0, 129.5, 133.3, 136.0, 137.2.

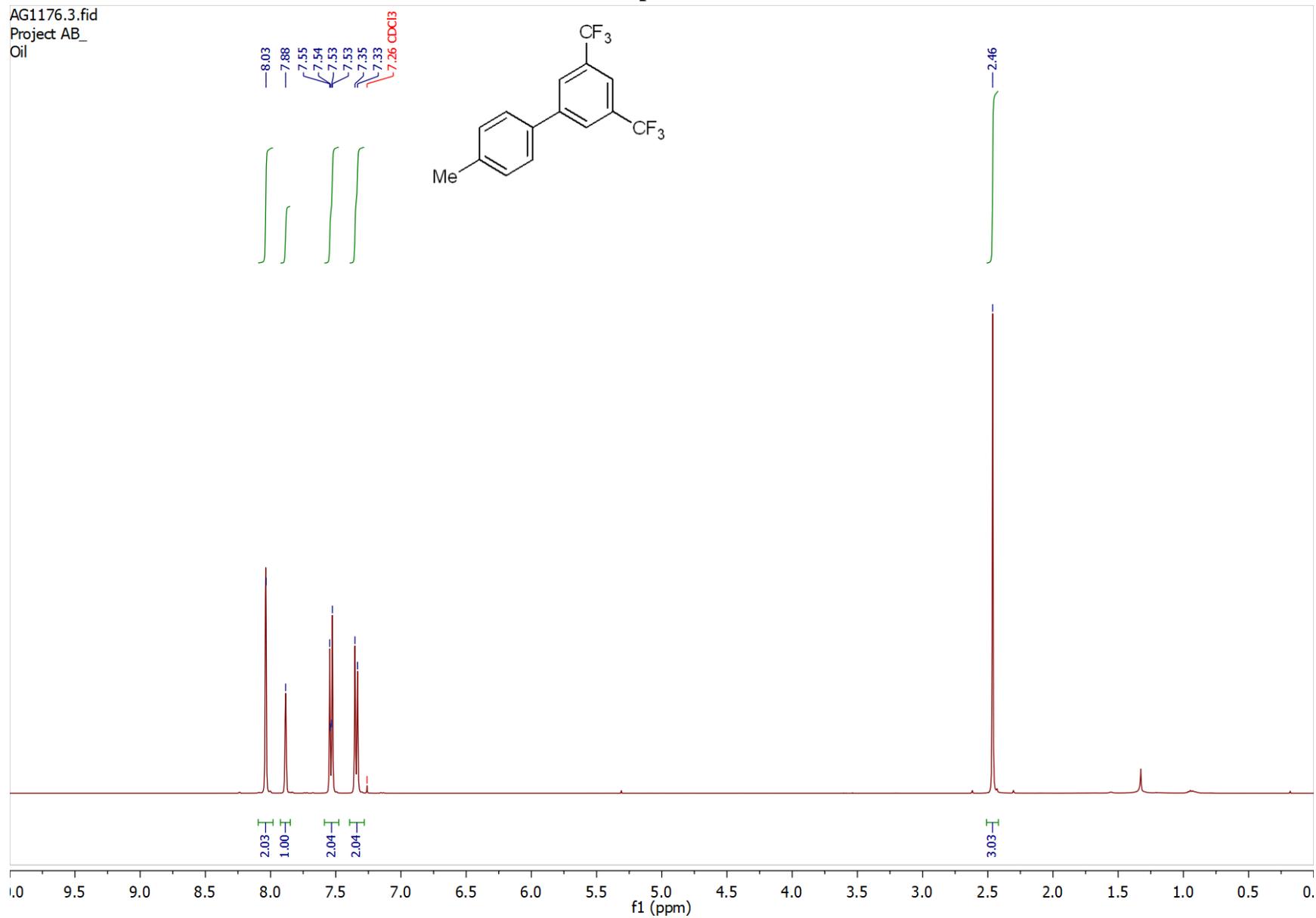
⁶⁶ K. T. Neumann, S. Klimczyk, M. N. Burhardt, B. Bang-Andersen, T. Skrydstrup and A. T. Lindhardt, *ACS Catal.*, 2016, **6**, 4710-4714.

⁶⁷ K. Itami, T. Nokami, Y. Ishimura, K. Mitsudo, T. Kamei and J.-i. Yoshida, *J. Am. Chem. Soc.*, 2001, **123**, 11577-11585.

⁶⁸ A. Ekebergh, R. Begon and N. Kann, *J. Org. Chem.*, 2020, **85**, 2966-2975.

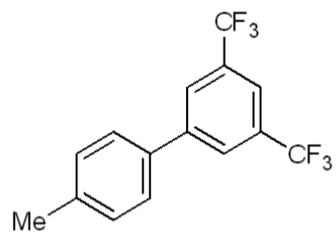
Copies of spectra

Compound 3a

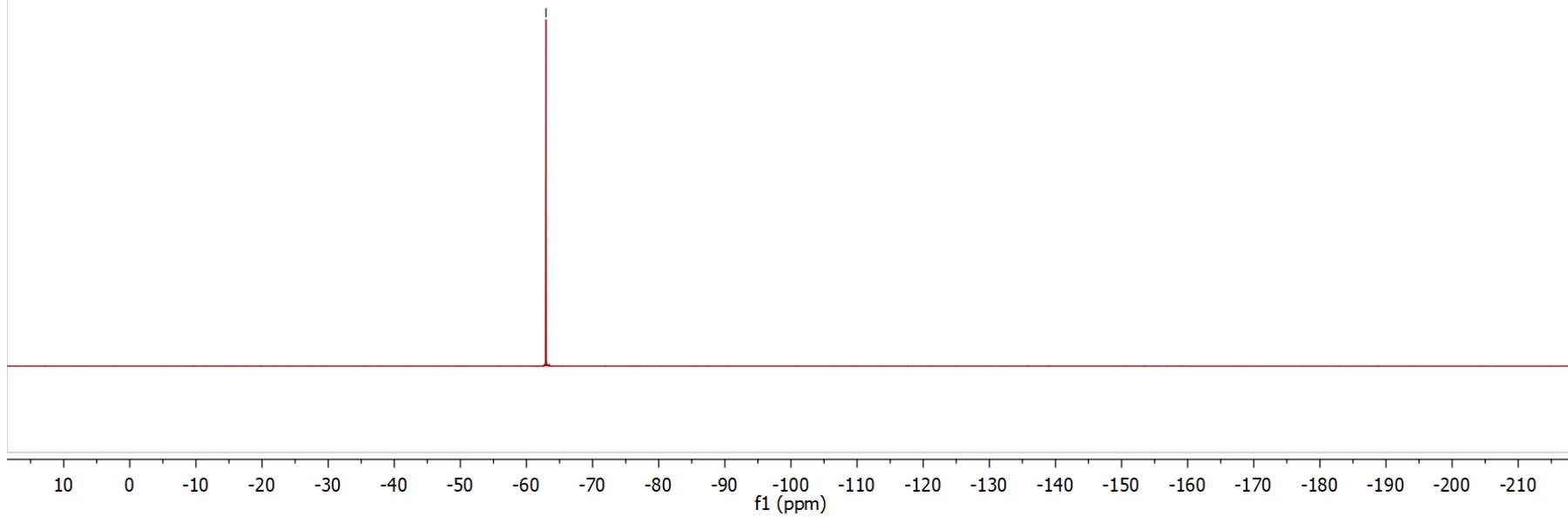


Compound 3a

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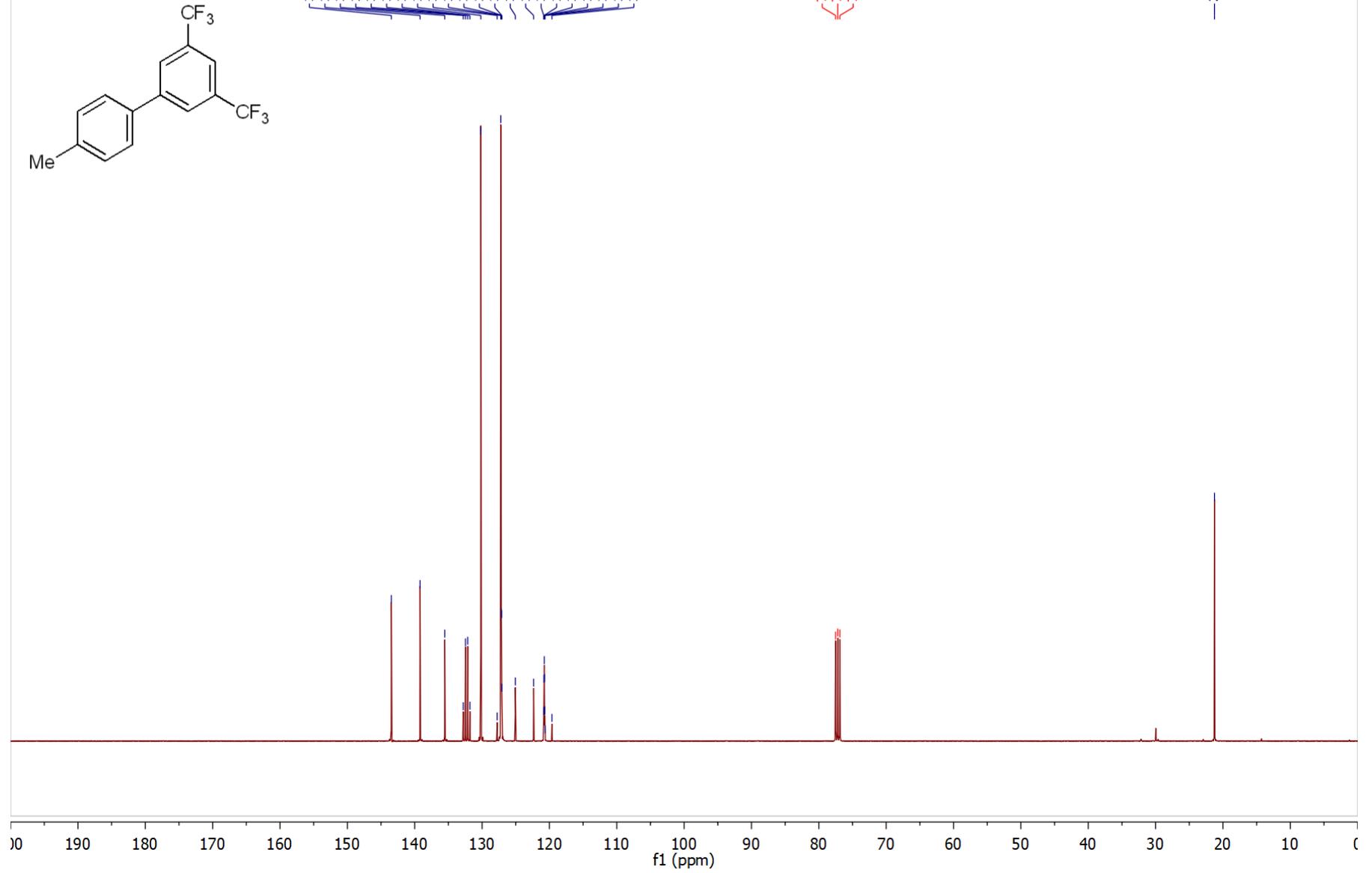


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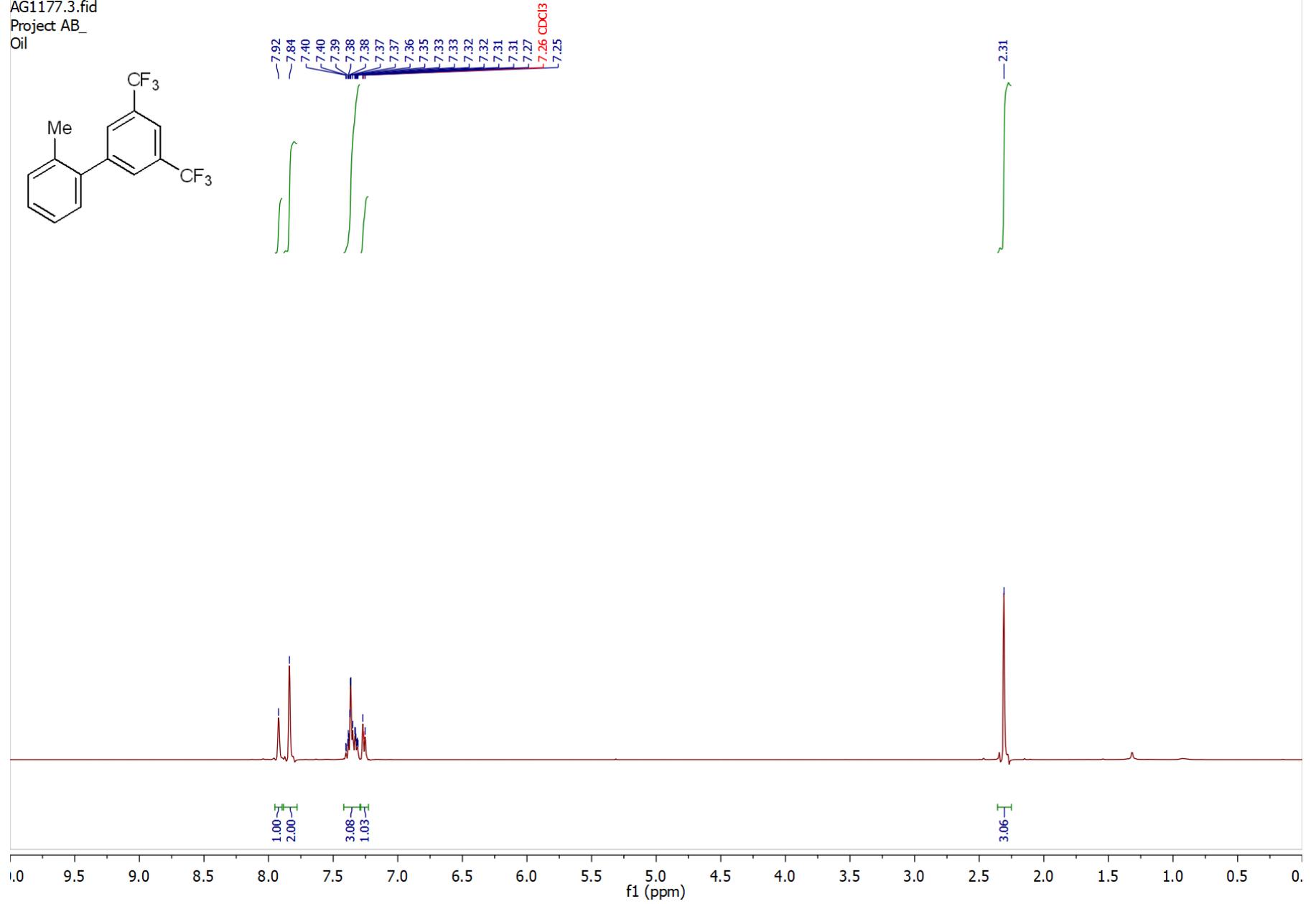
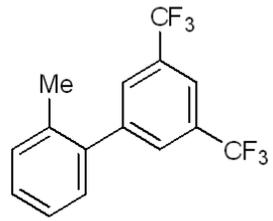
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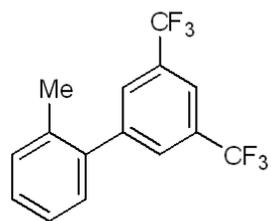
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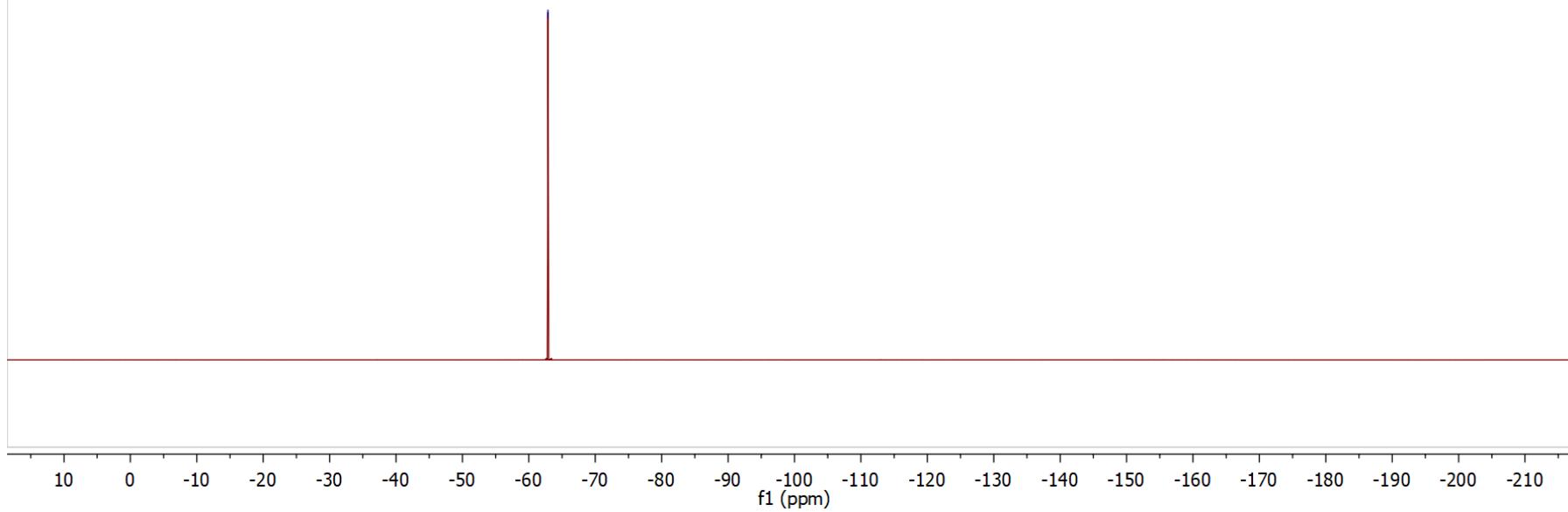


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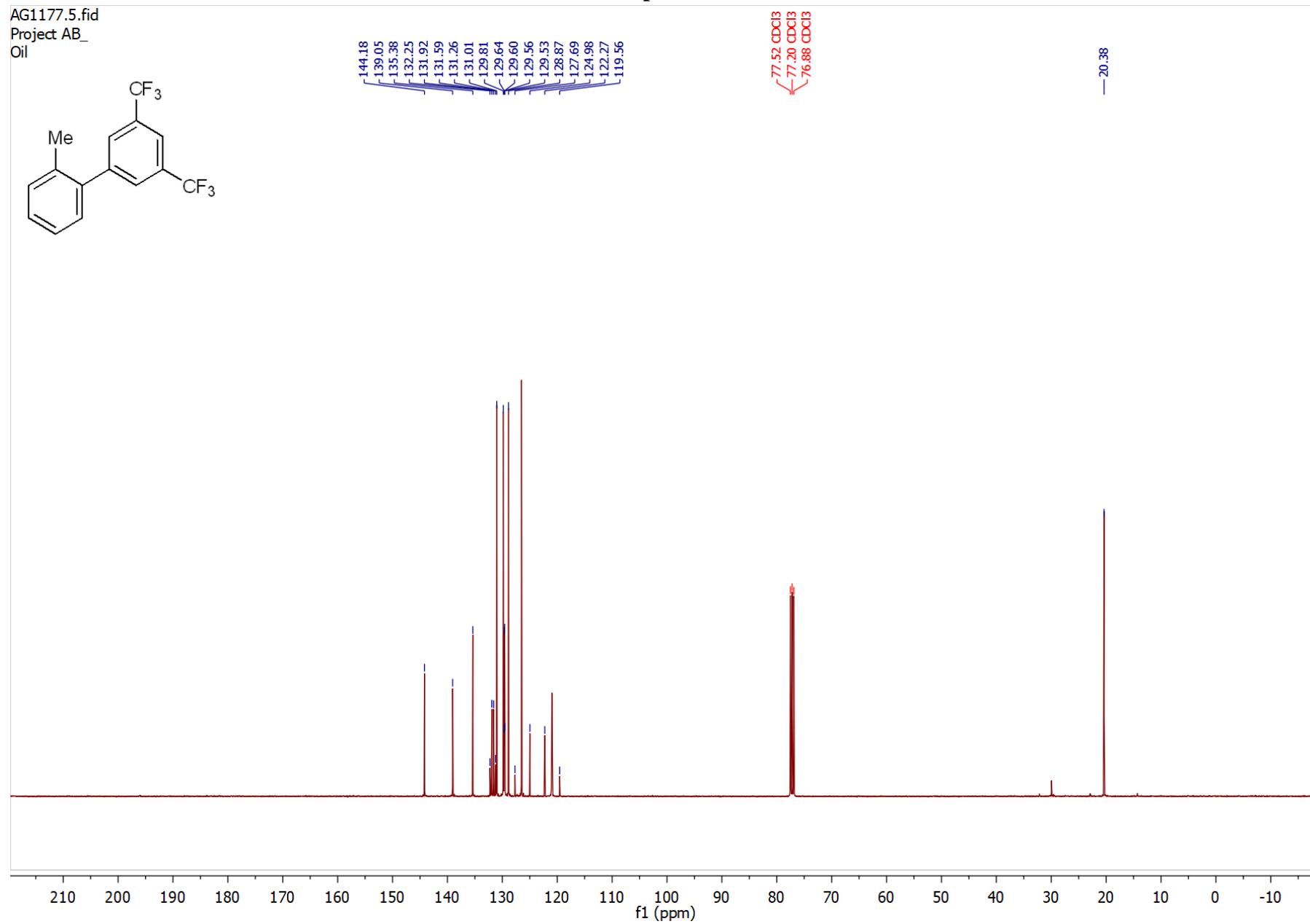
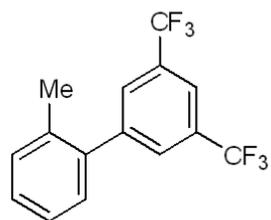


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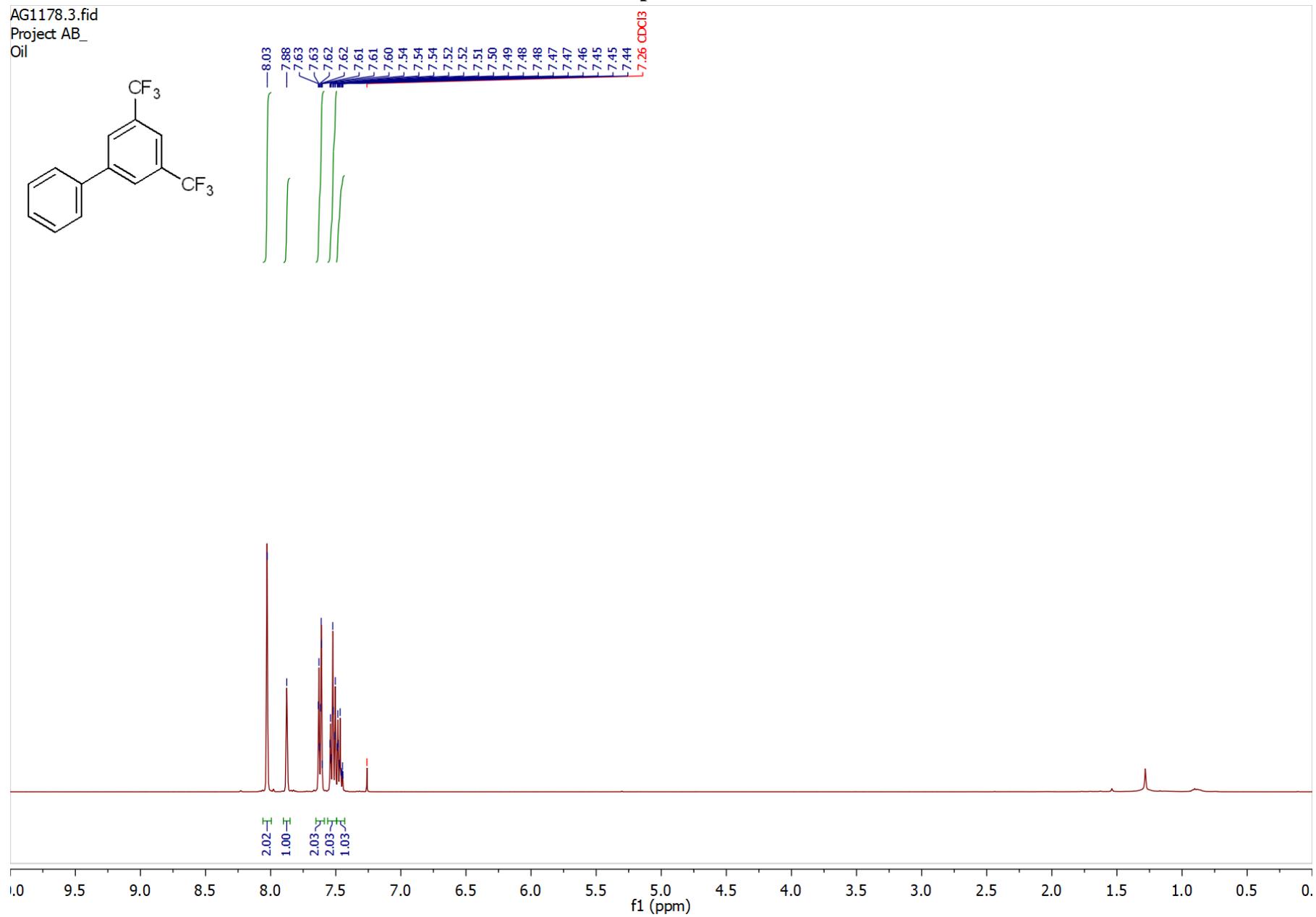
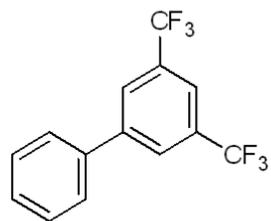
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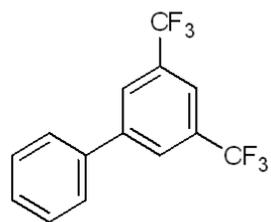
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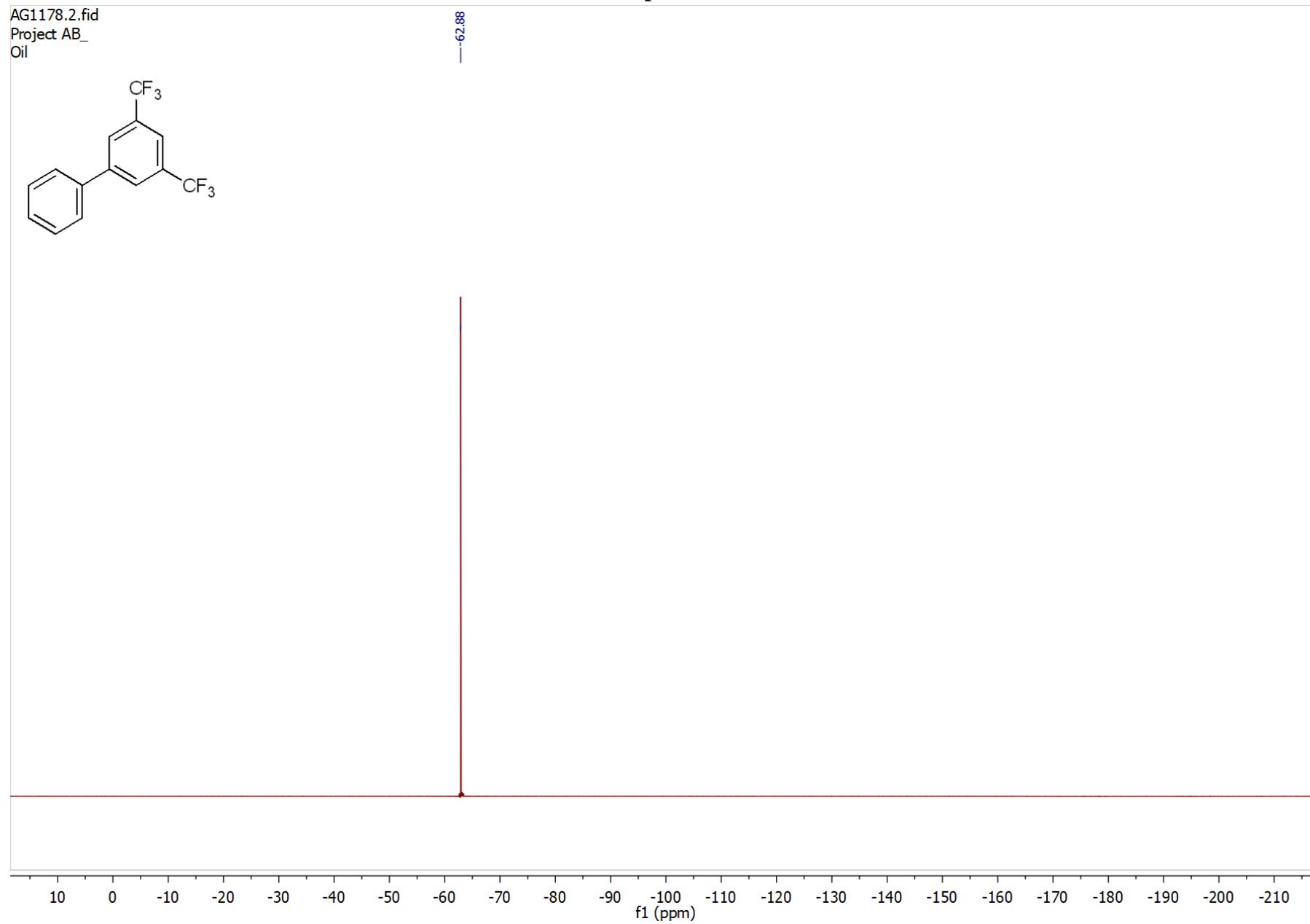


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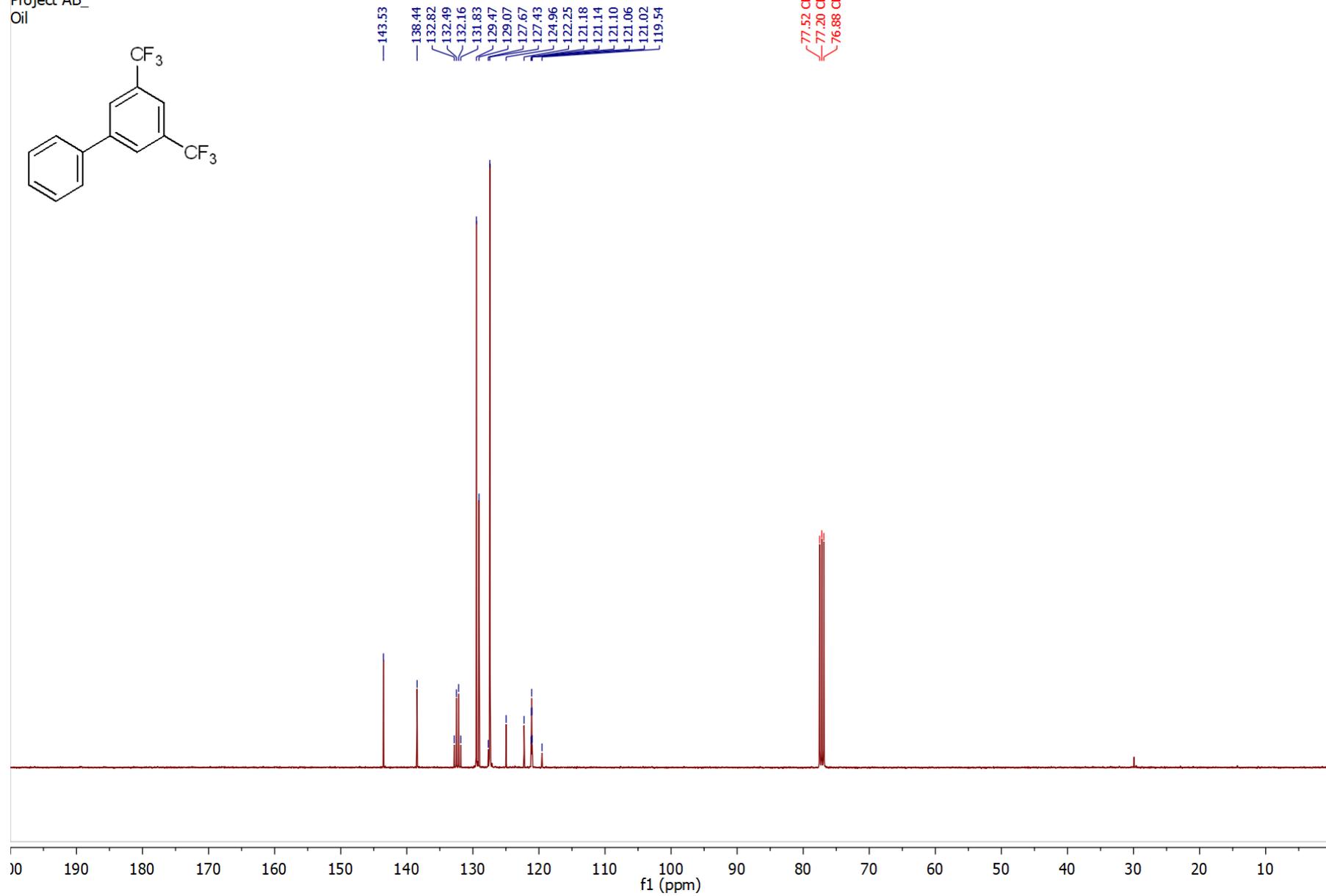
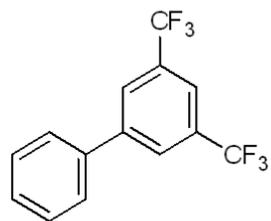


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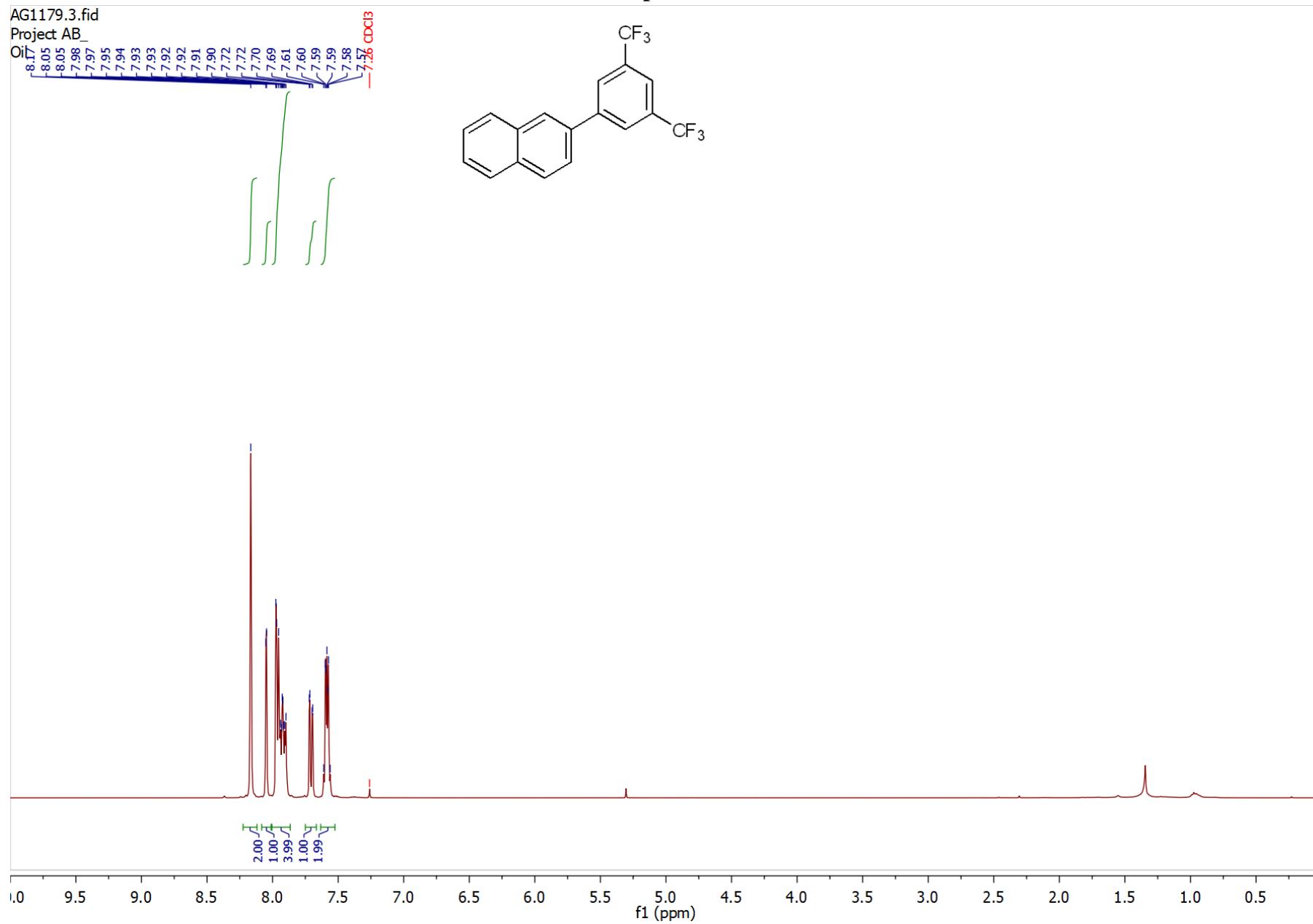


Compound 3c

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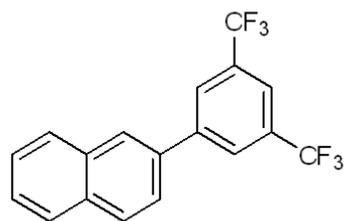


Compound 3d

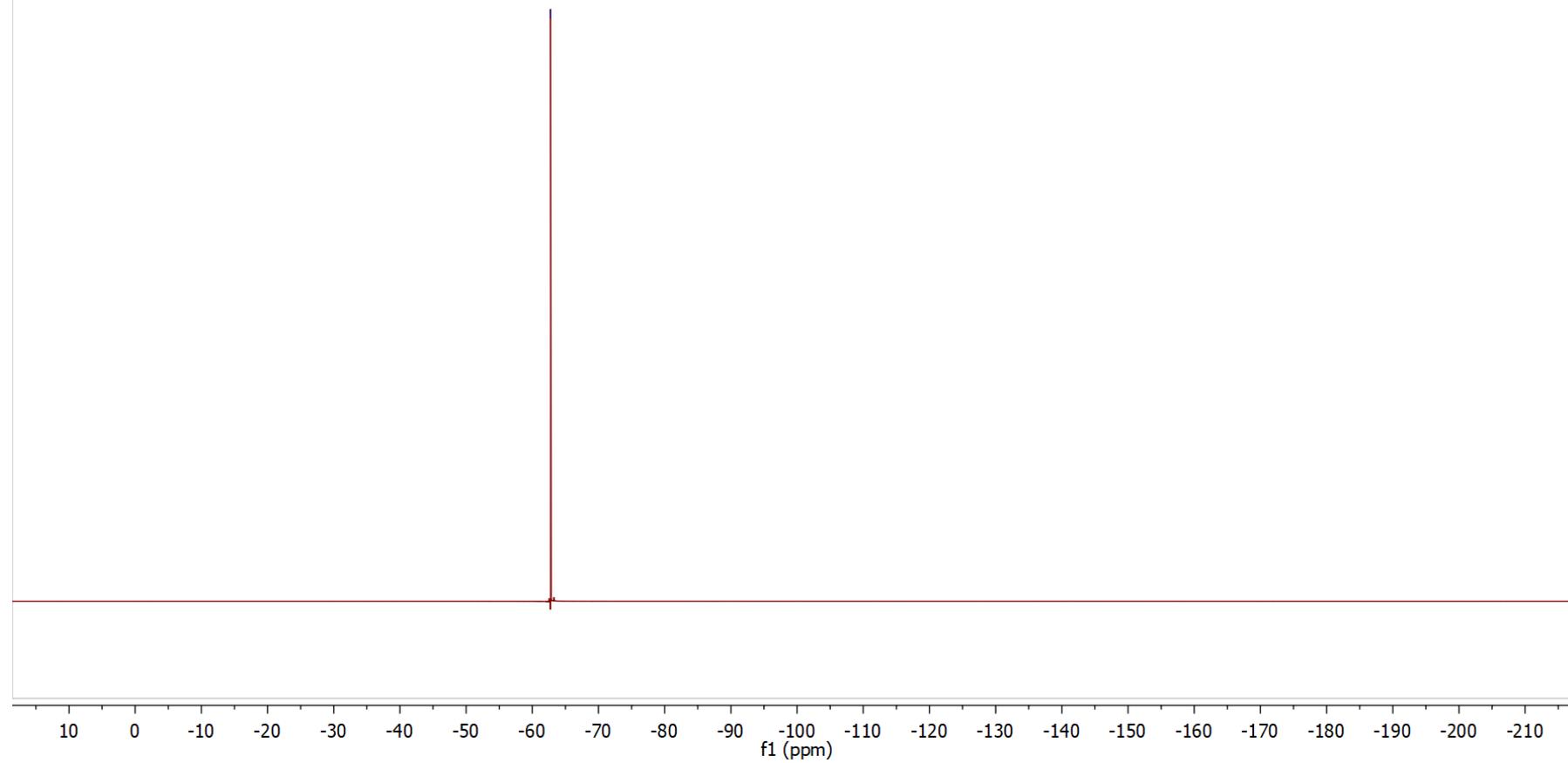


Compound 3d

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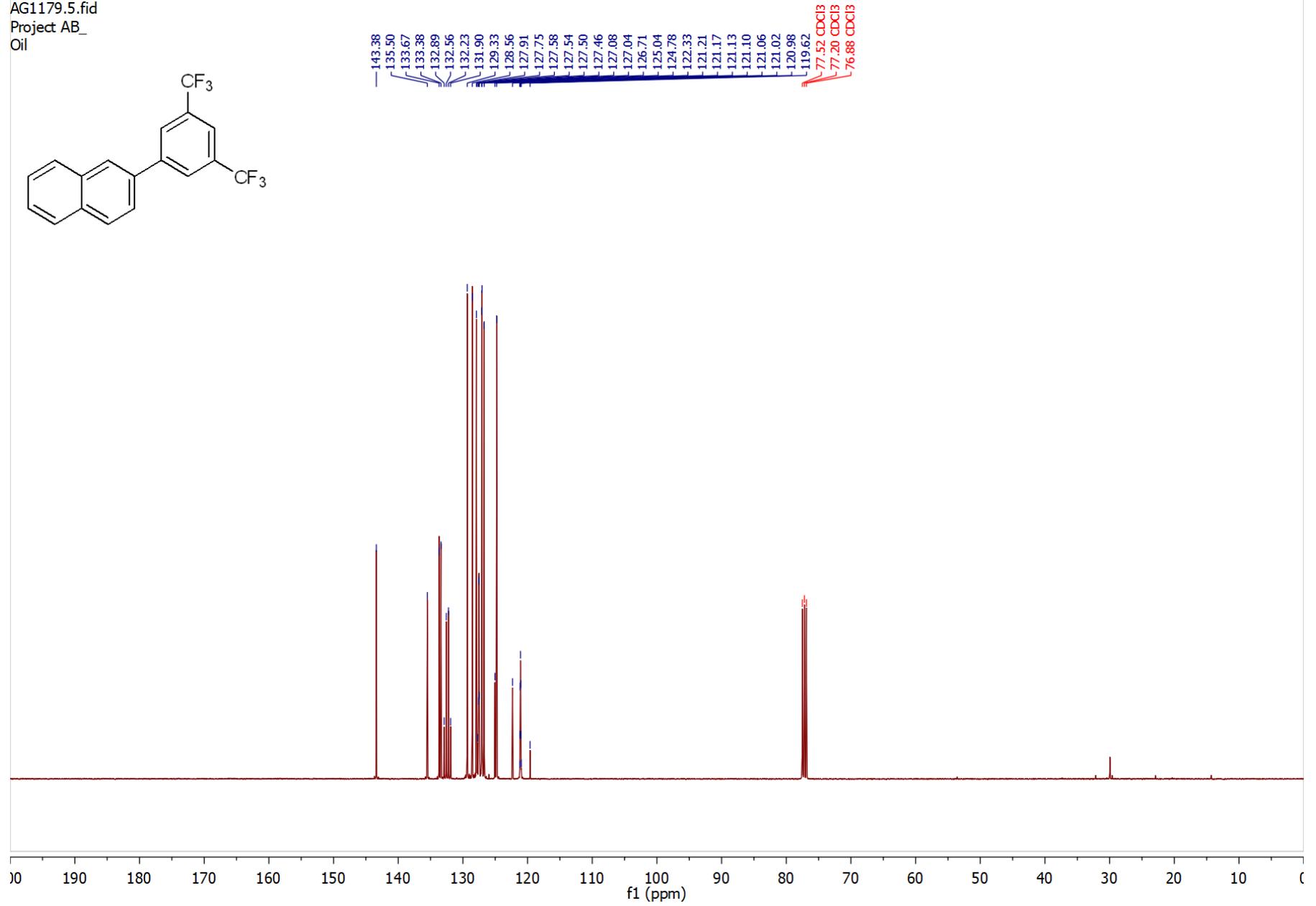
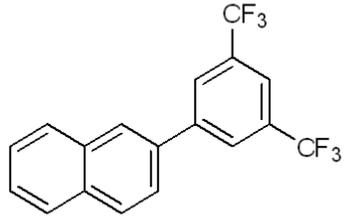


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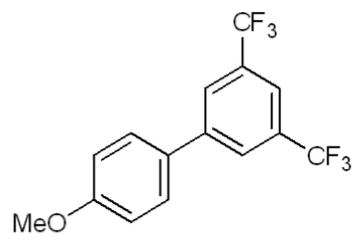
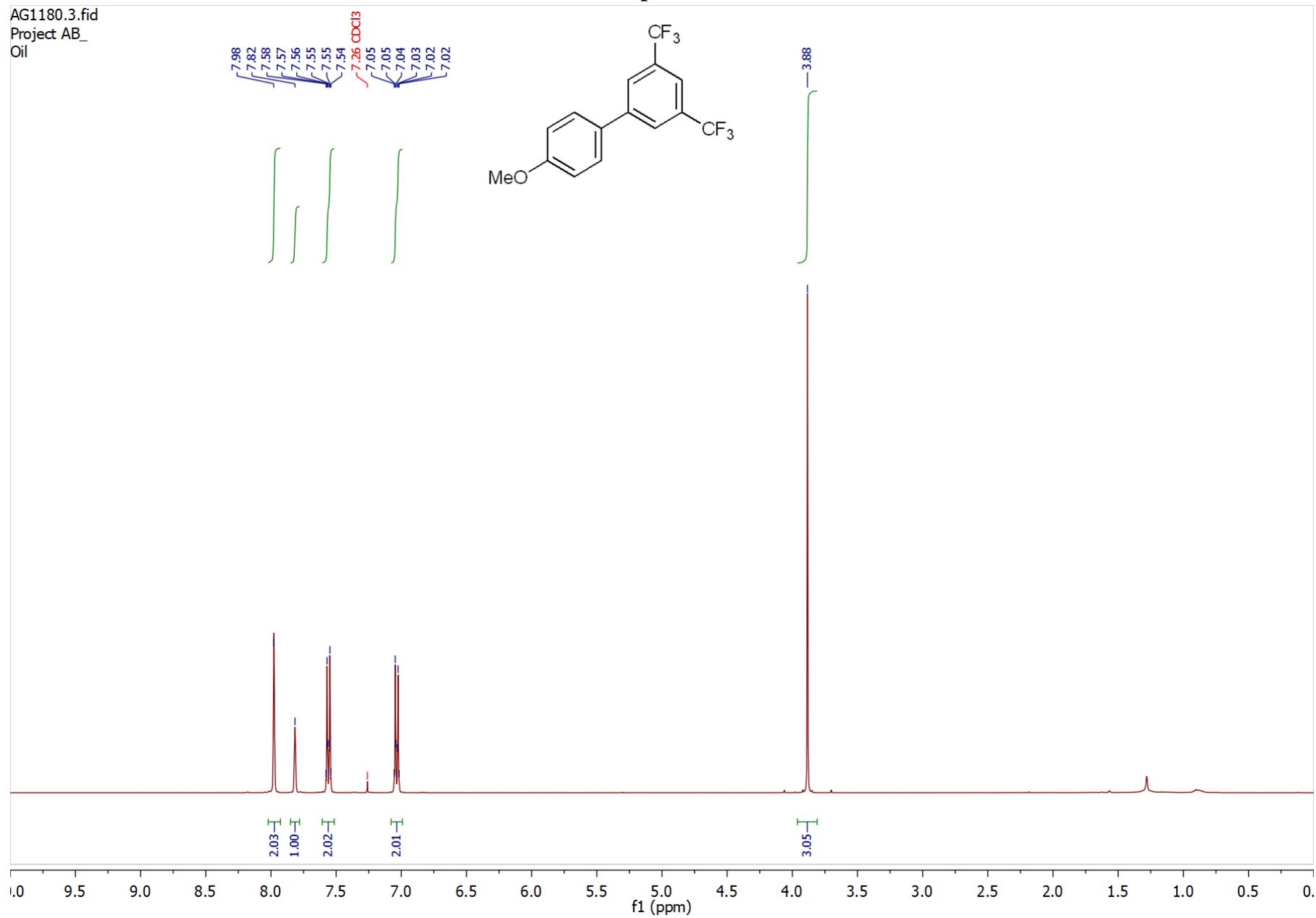
Compound 3d

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Compound 3e

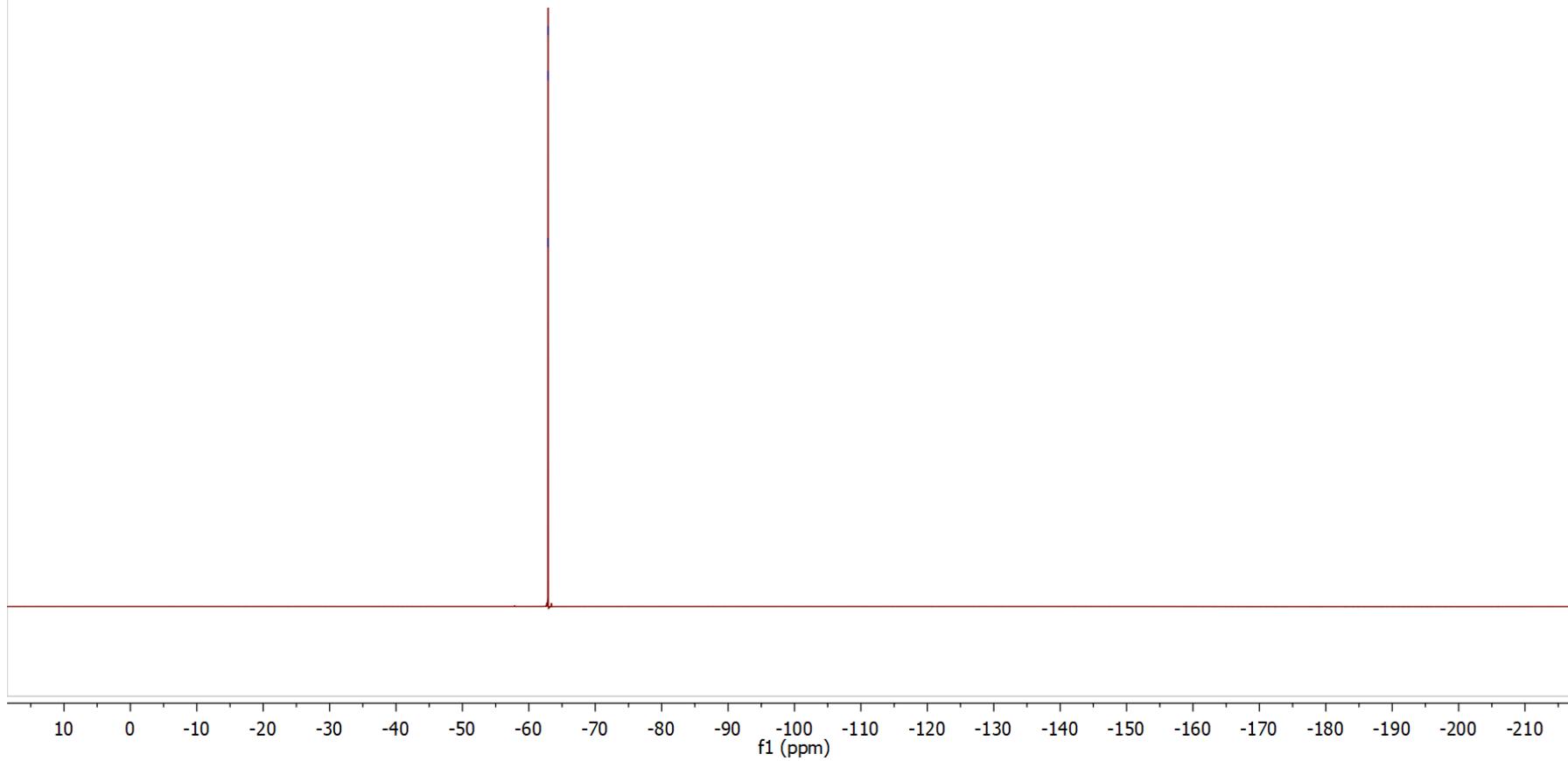
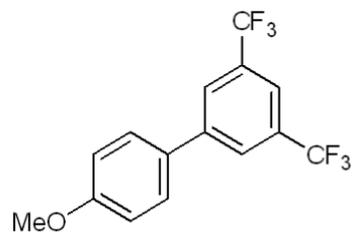
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Compound 3e

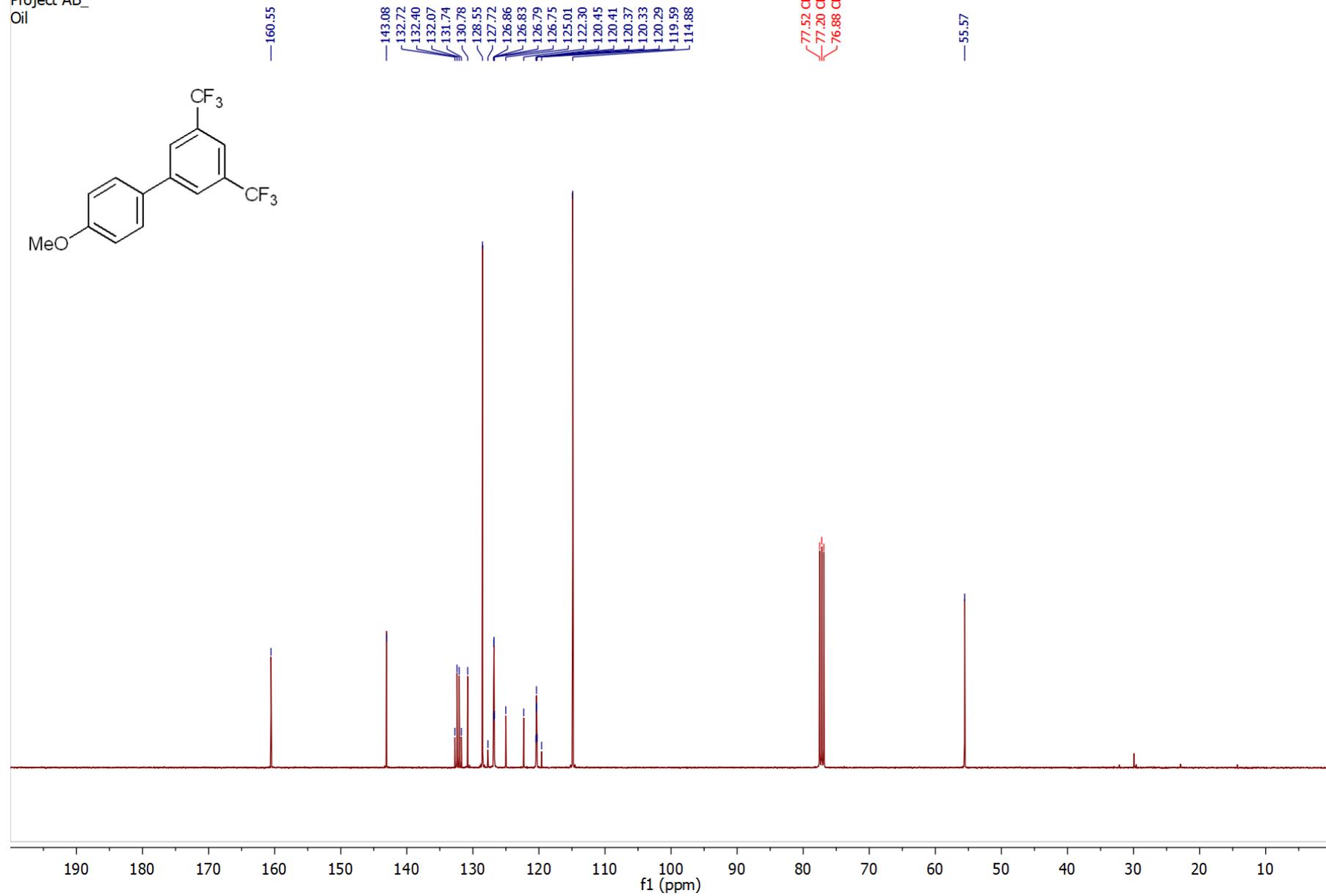
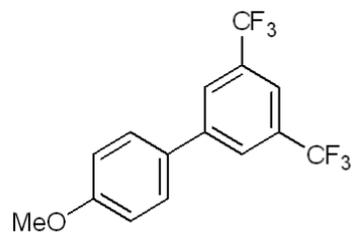
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62.90
62.91
62.92



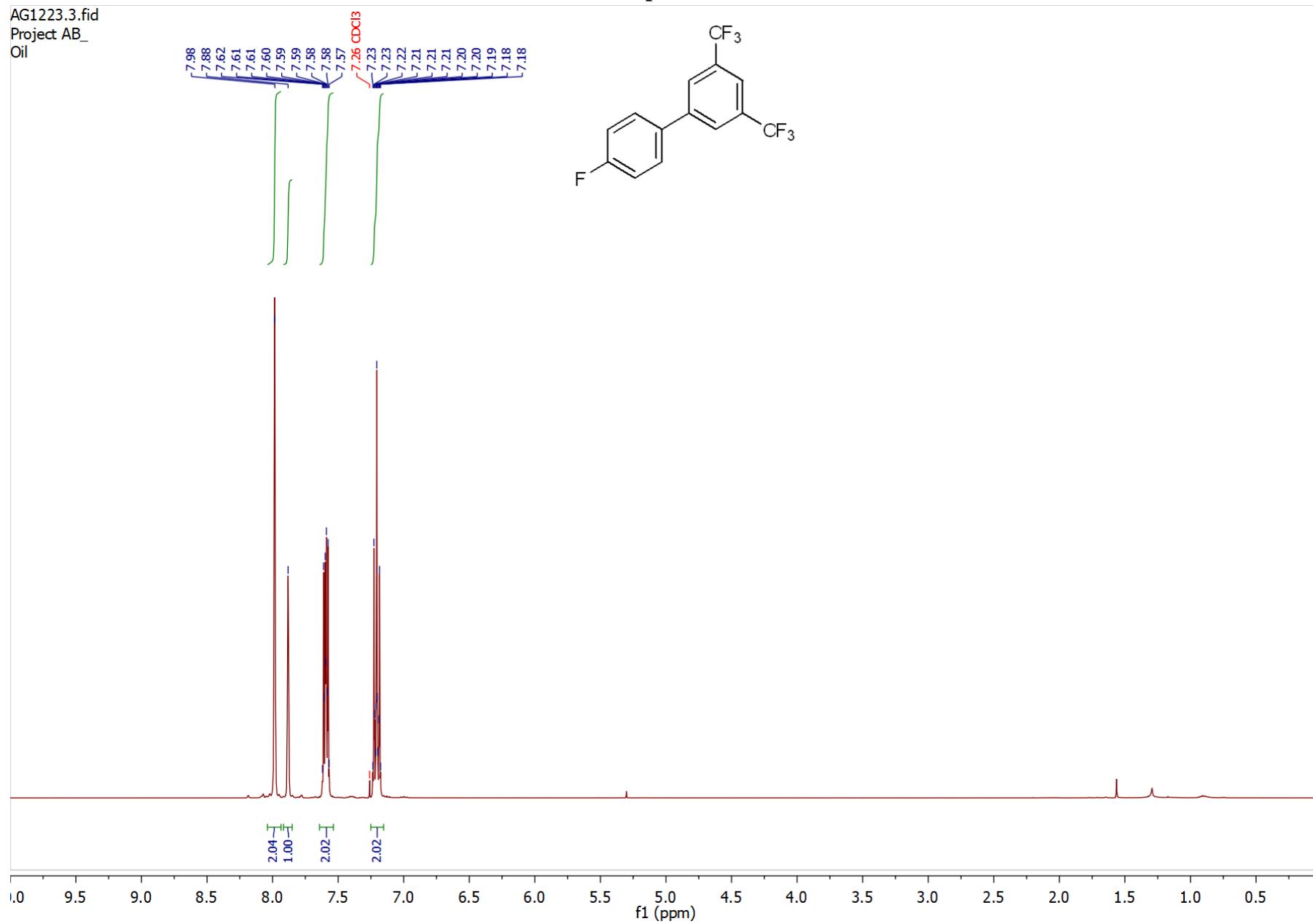
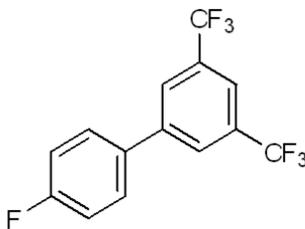
Compound 3e

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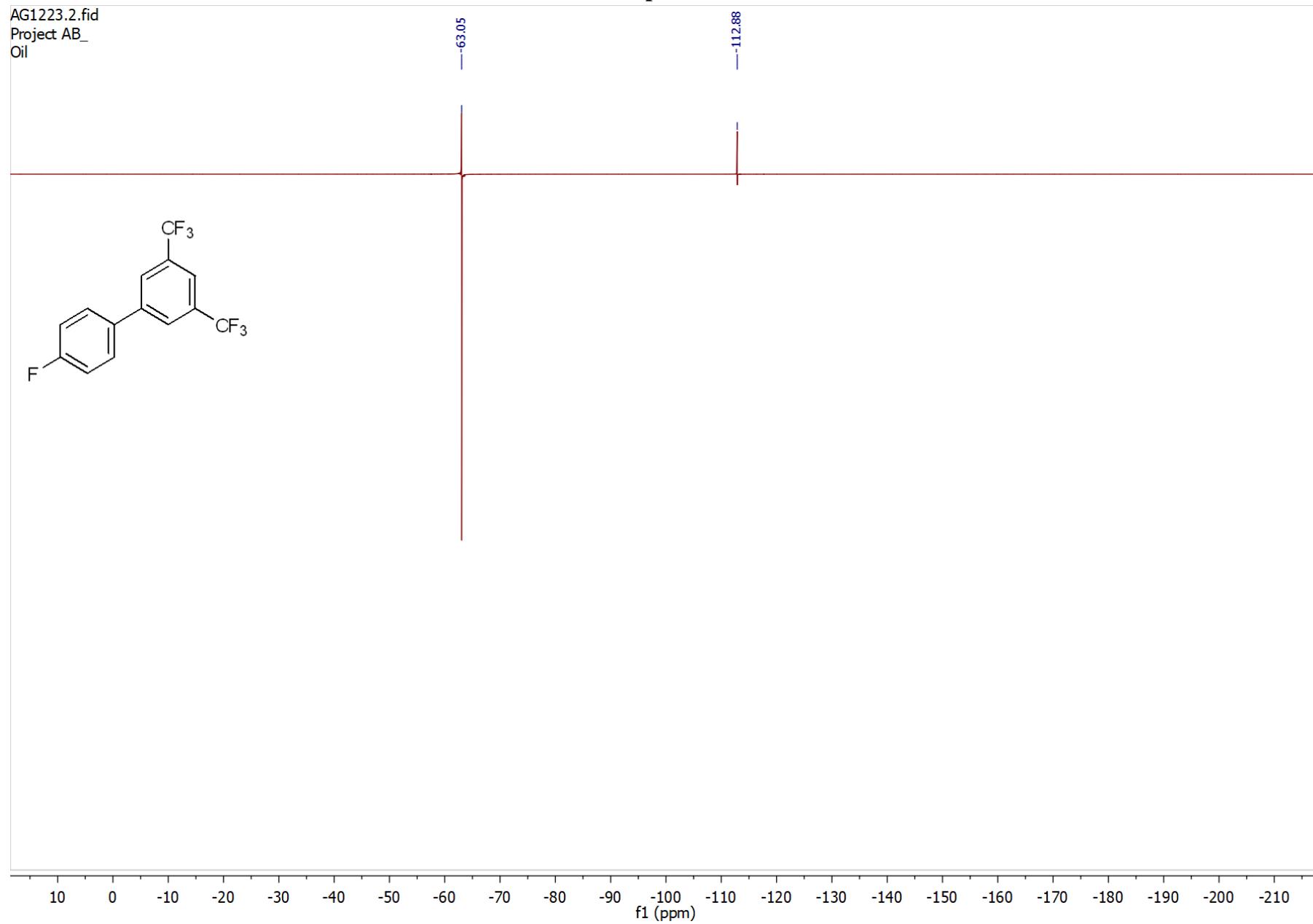
Compound 3f

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Project AB_
Oil



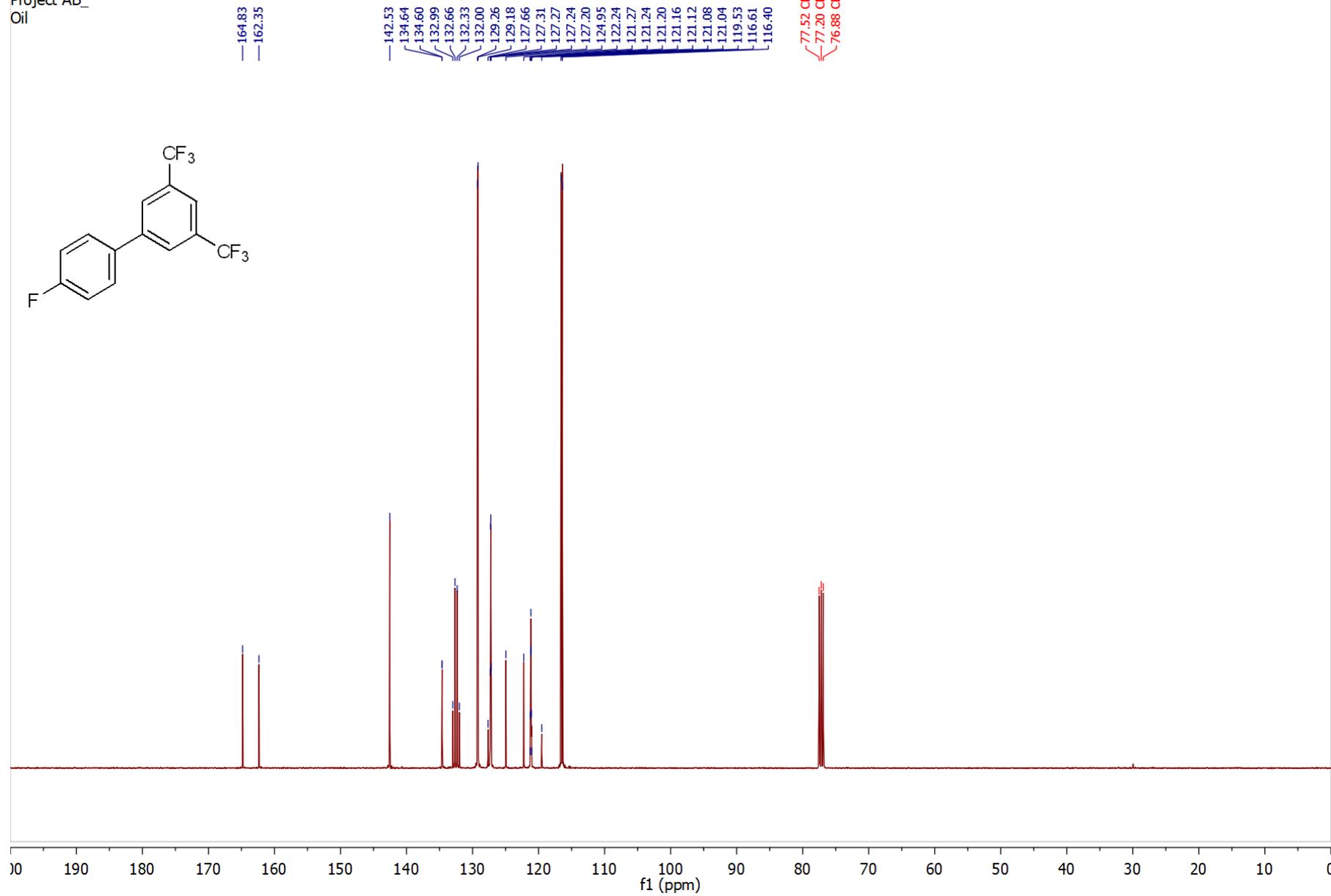
Compound 3f

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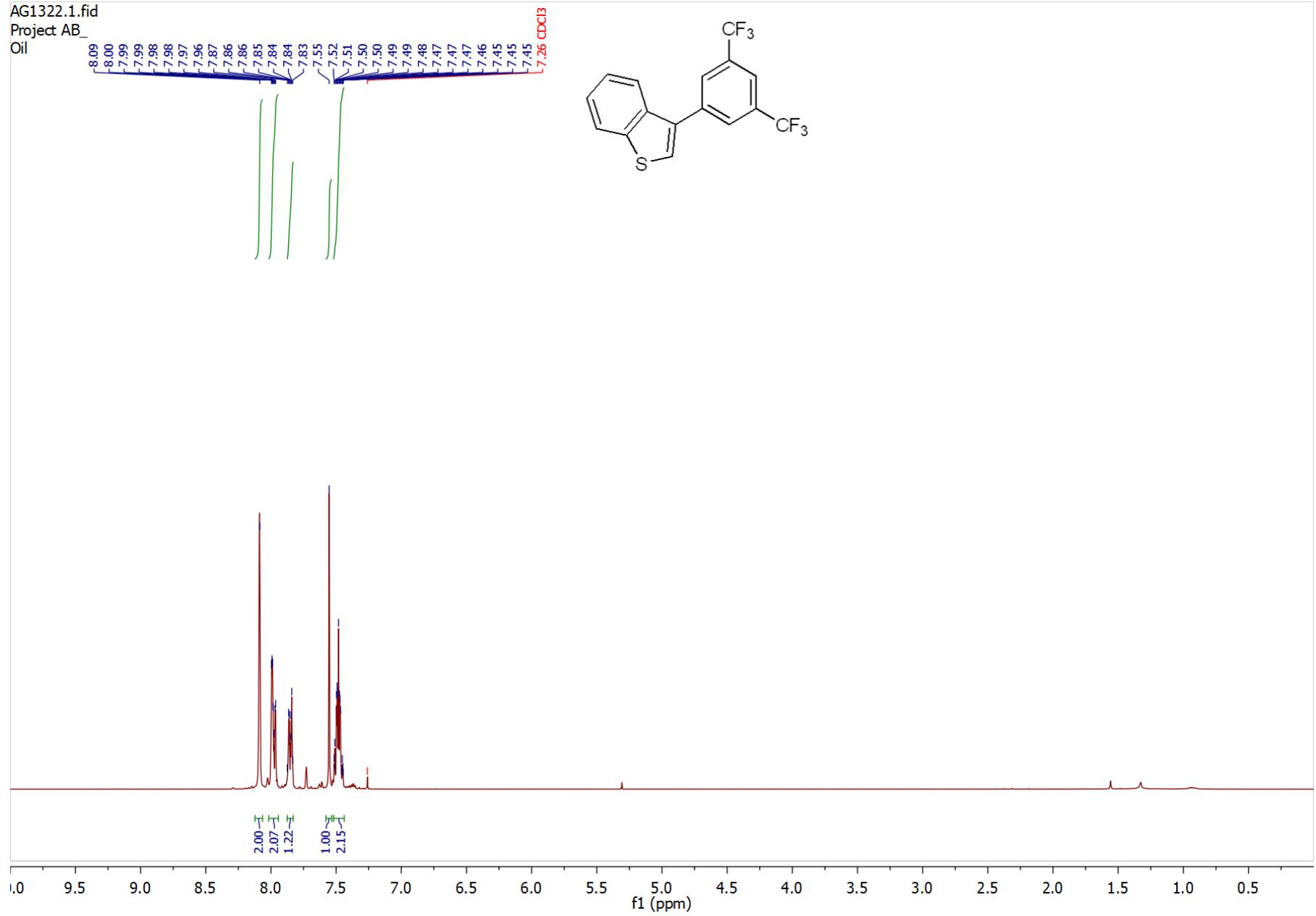
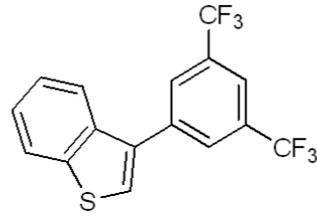
Compound 3f

AG1223.5.fid
Project AB_
Oil



Compound 3g

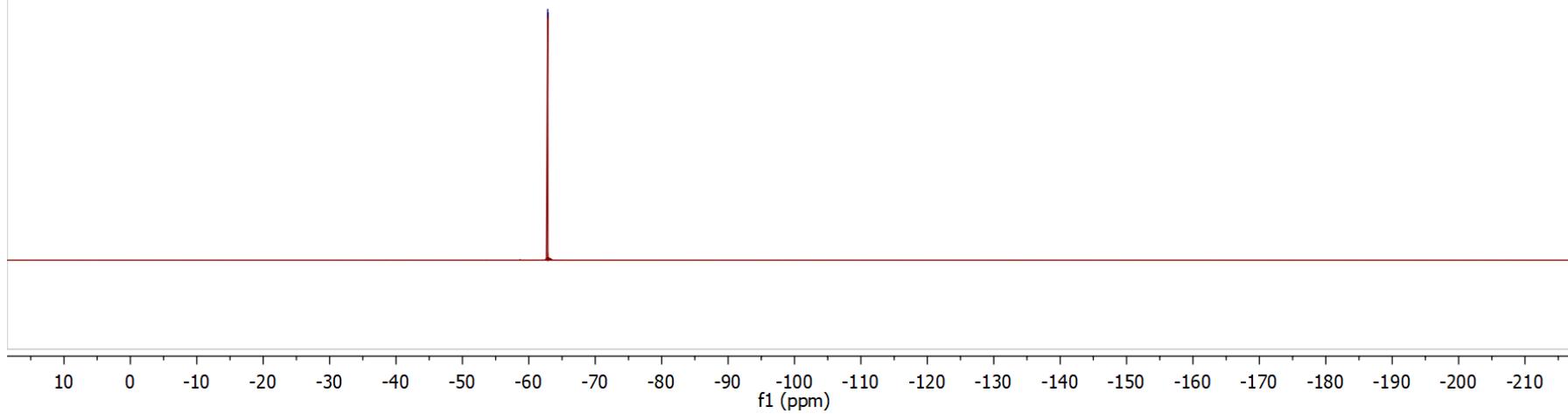
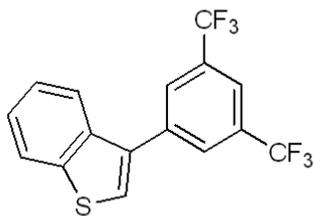
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Project AB_
Oil



Compound 3g

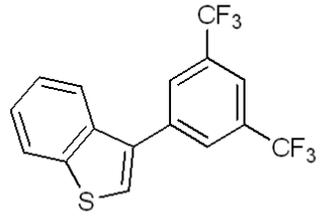
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-62.84



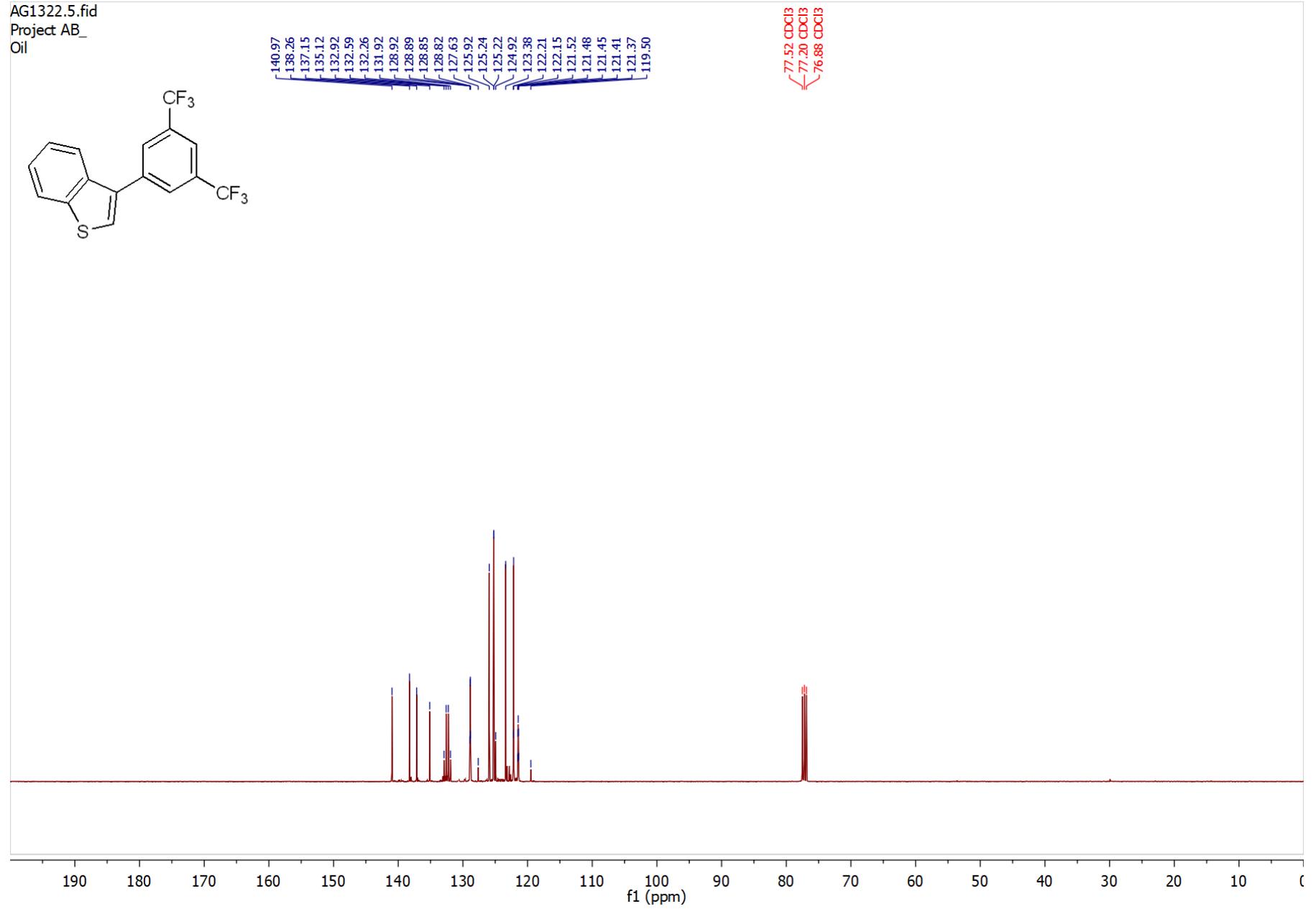
Compound 3g

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Project AB_
Oil



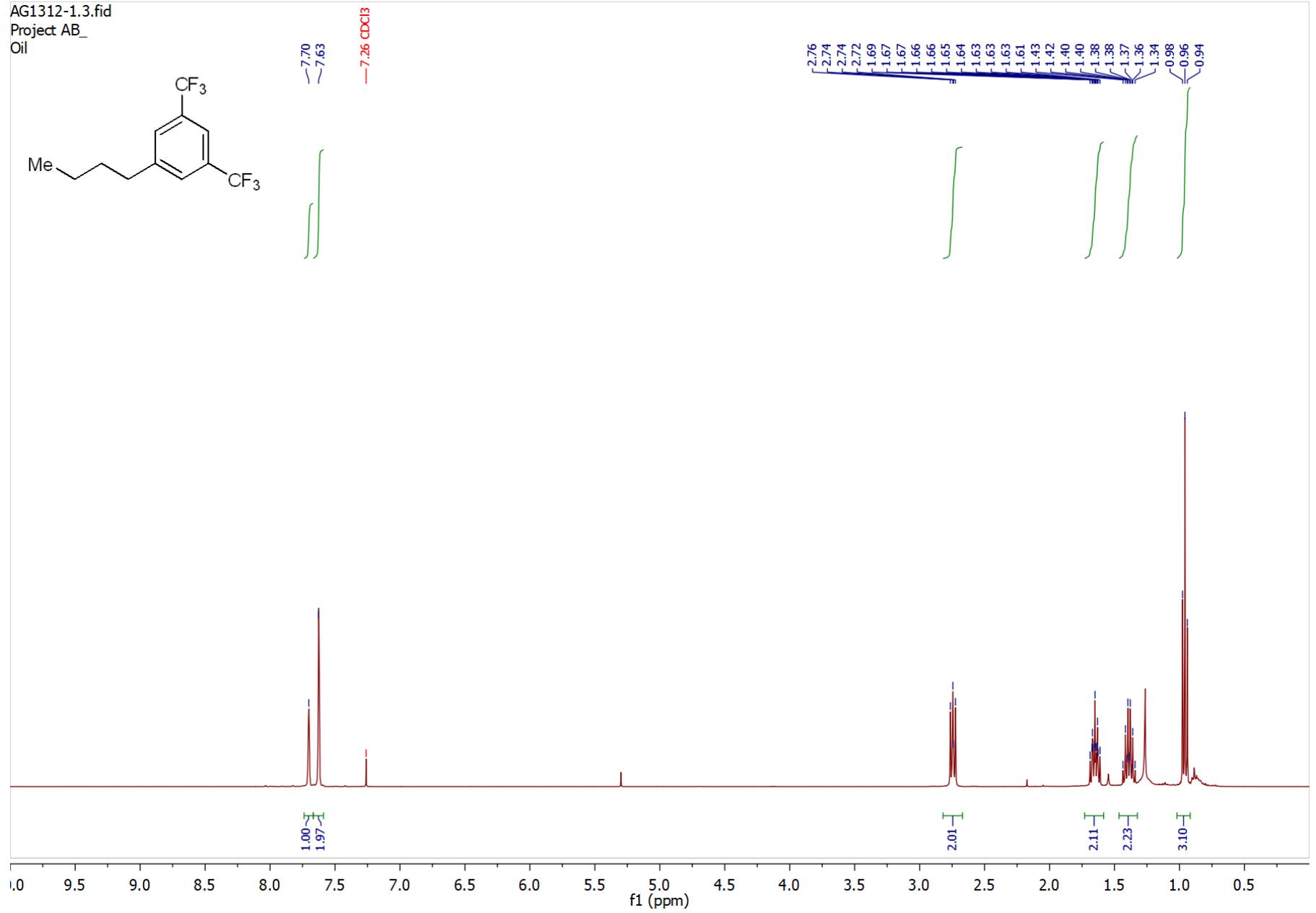
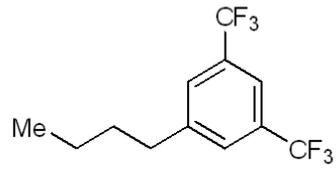
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124.92
123.38
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121.37
119.50

77.52 CDCl3
77.20 CDCl3
76.88 CDCl3



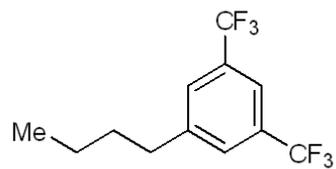
Compound 3h

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Project AB_
Oil

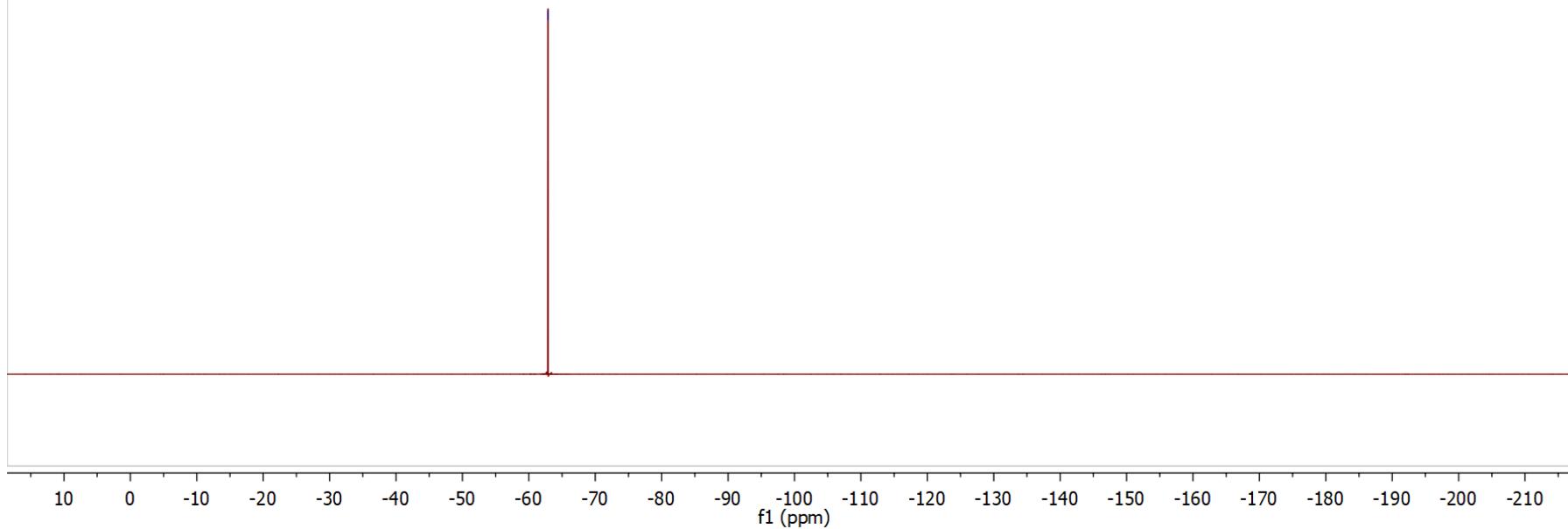


Compound 3h

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Project AB_
Oil

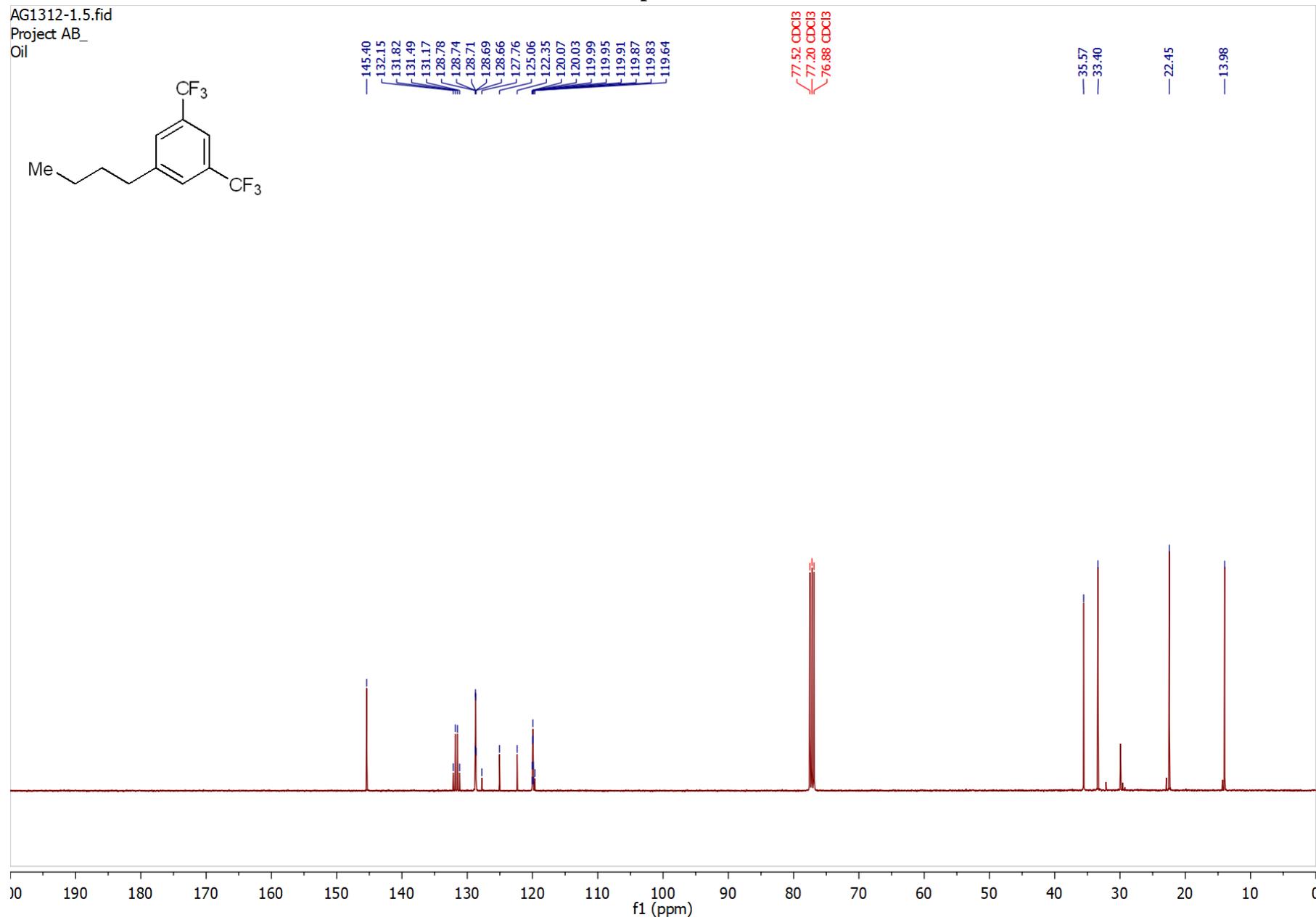
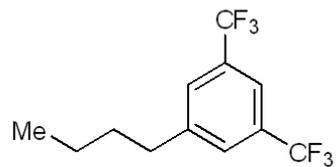


-62.90



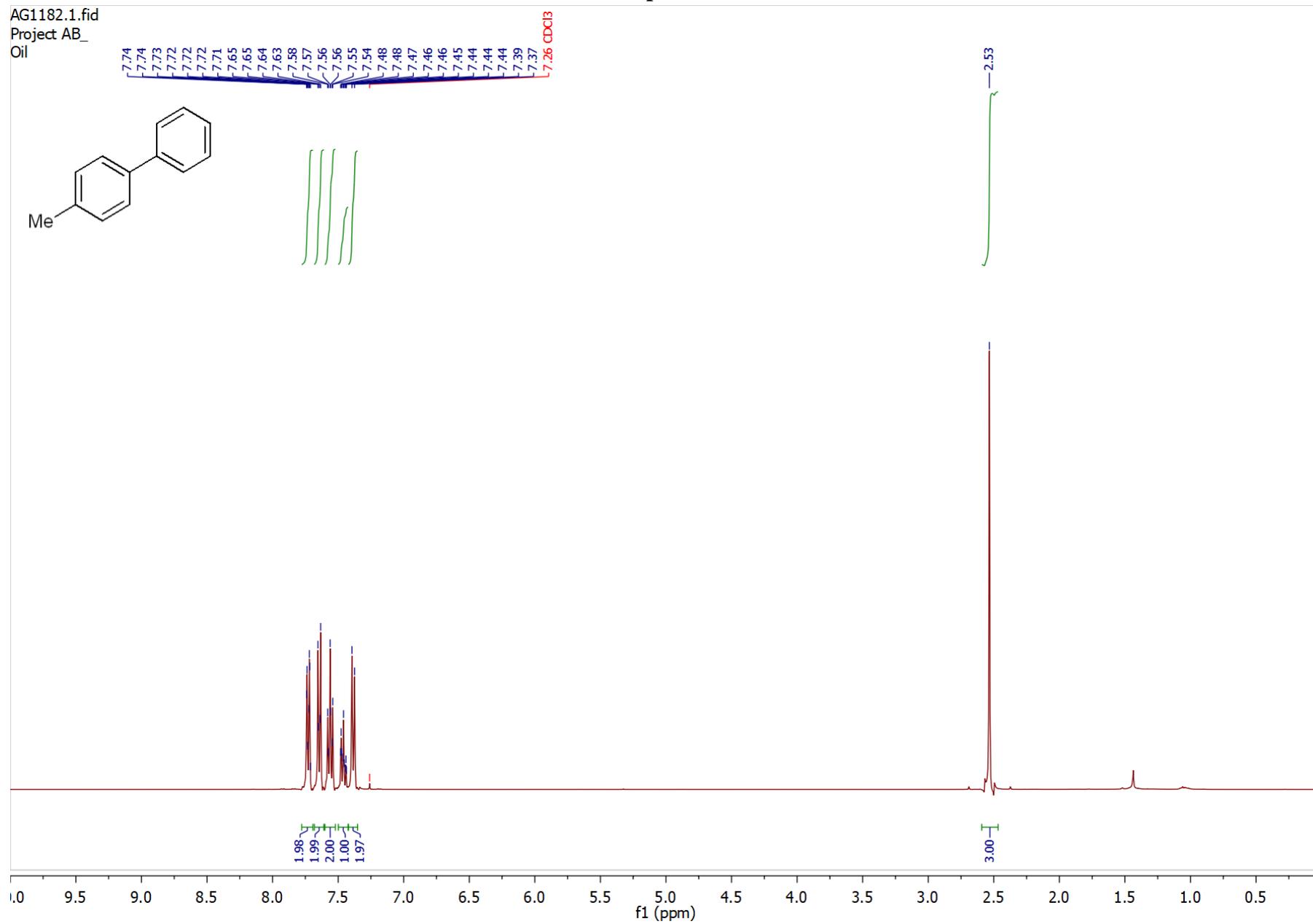
Compound 3h

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Project AB_
Oil



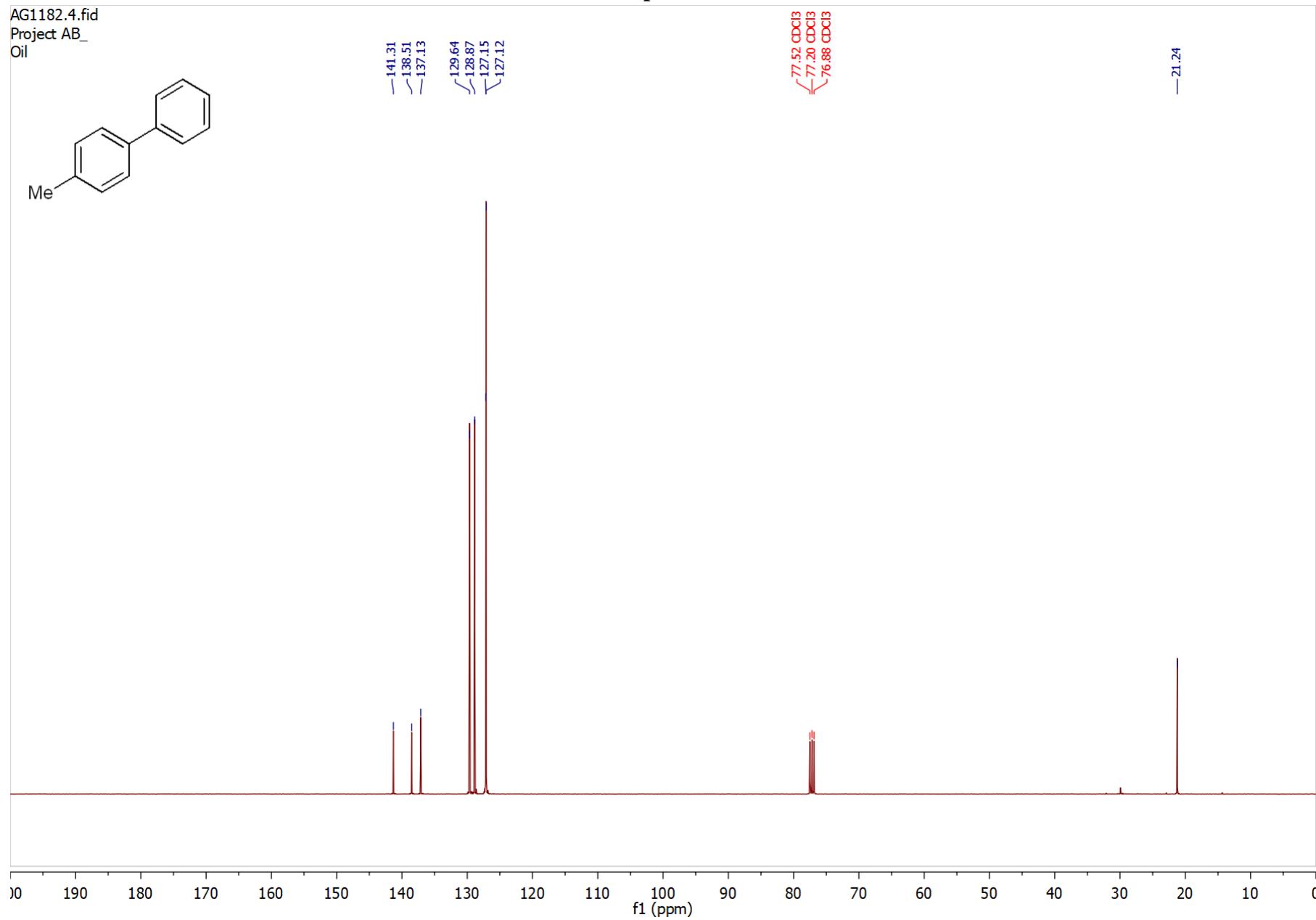
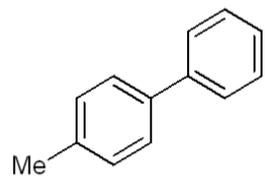
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AG1182.1.fid
Project AB_
Oil



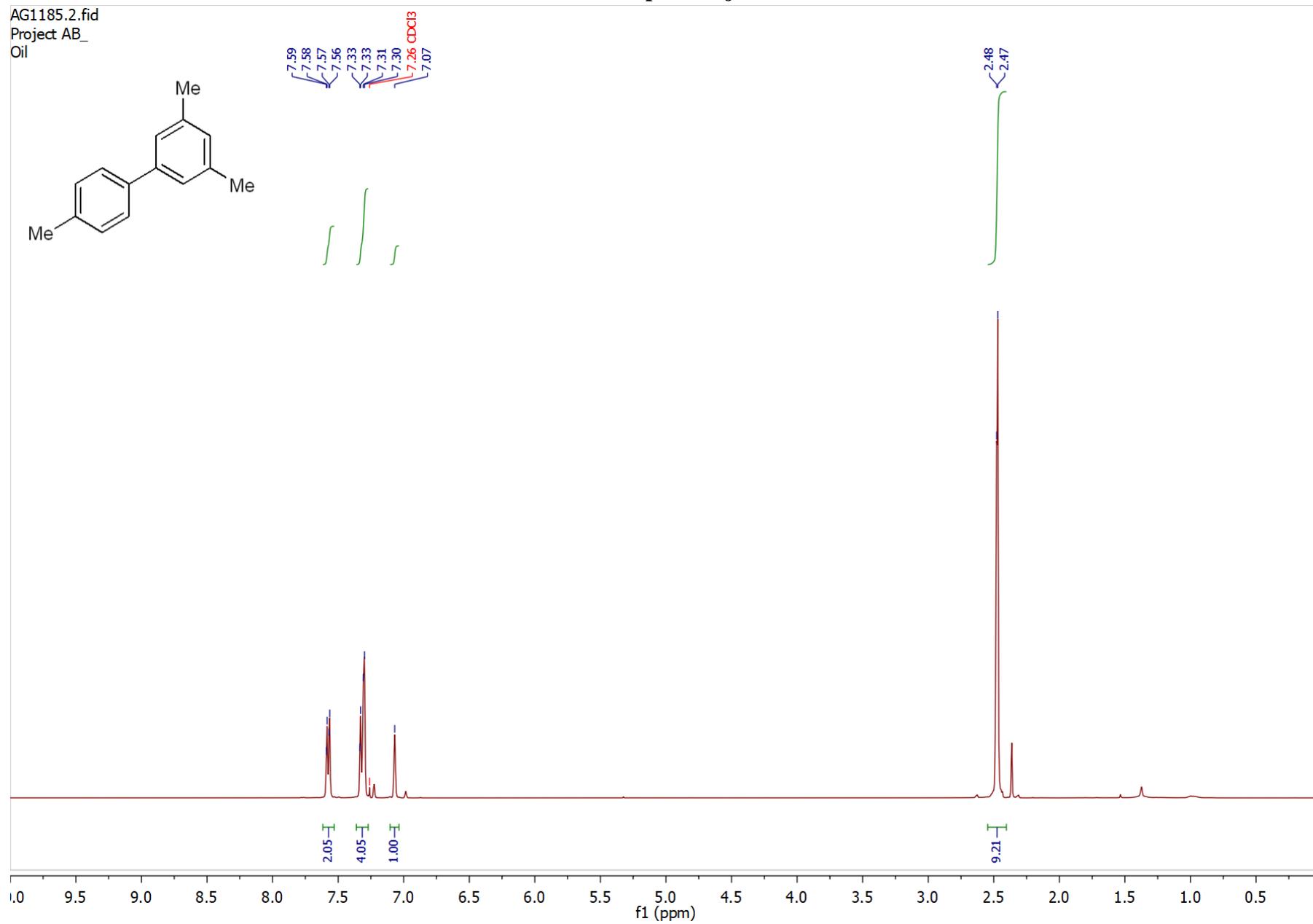
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Project AB_
Oil



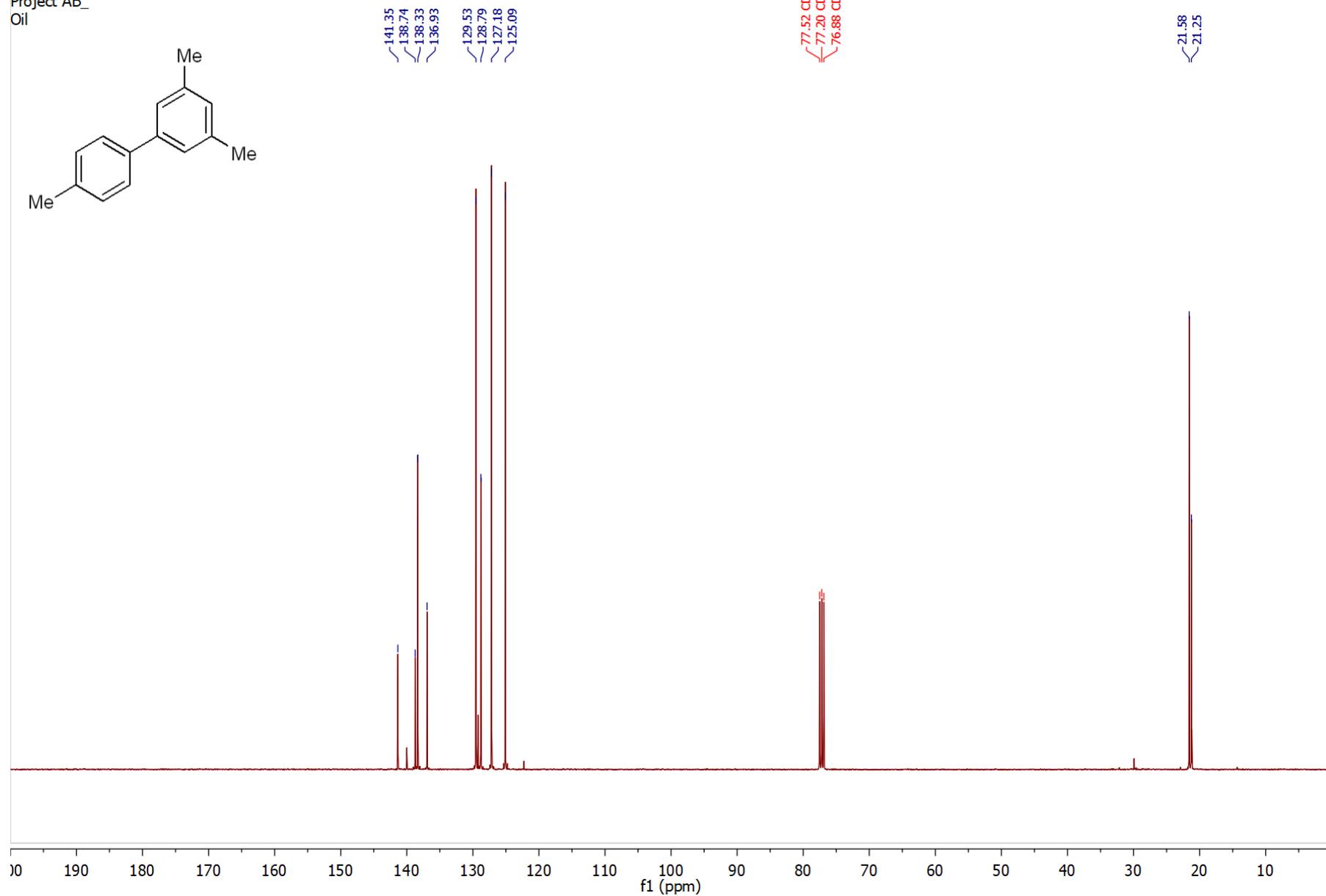
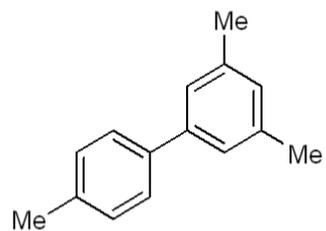
Compound 3j

AG1185.2.fid
Project AB_
Oil



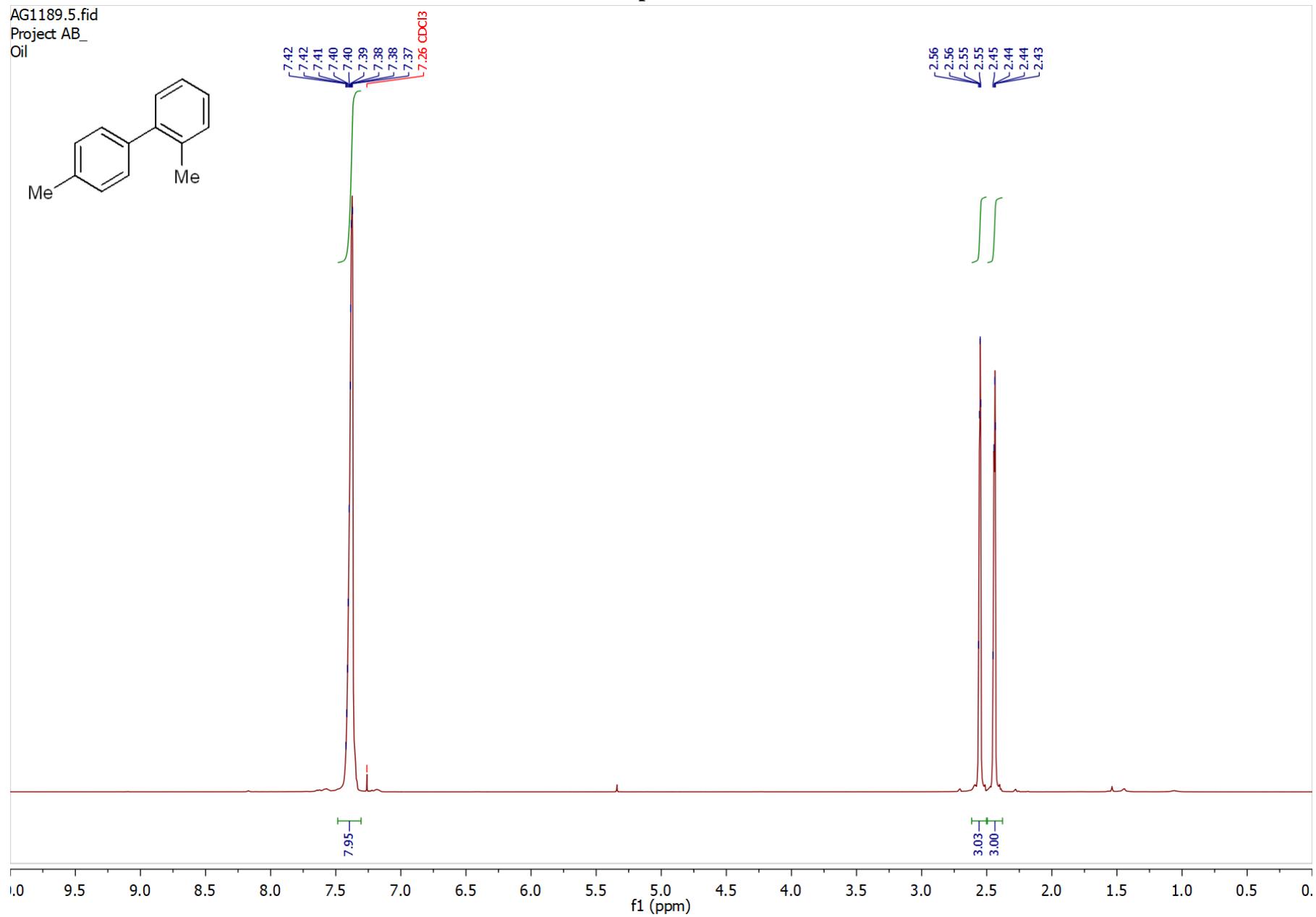
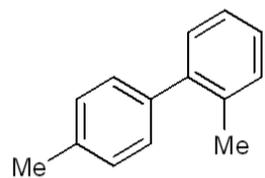
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Project AB_
Oil



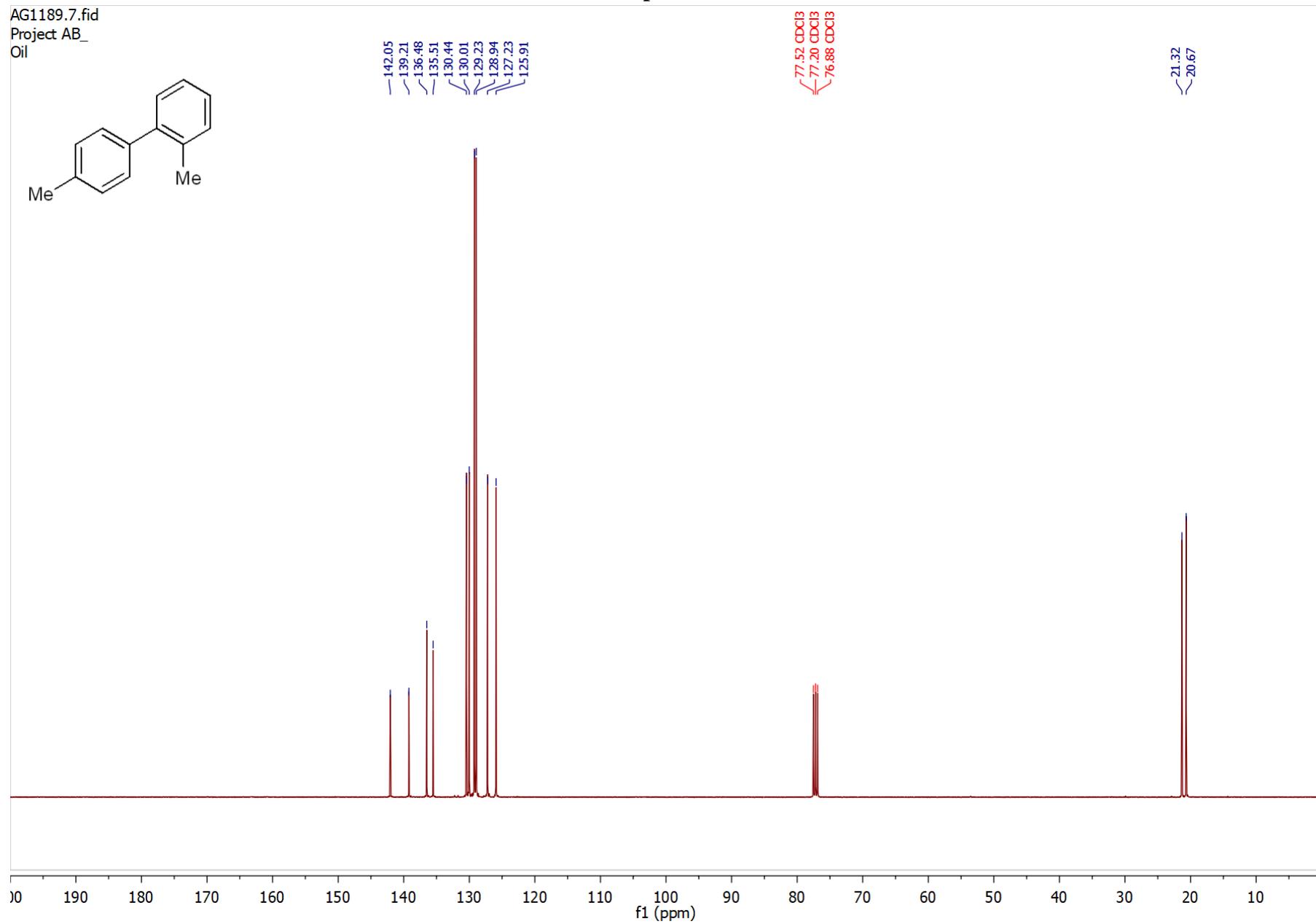
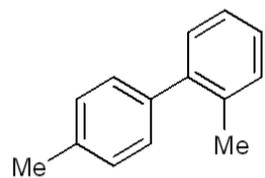
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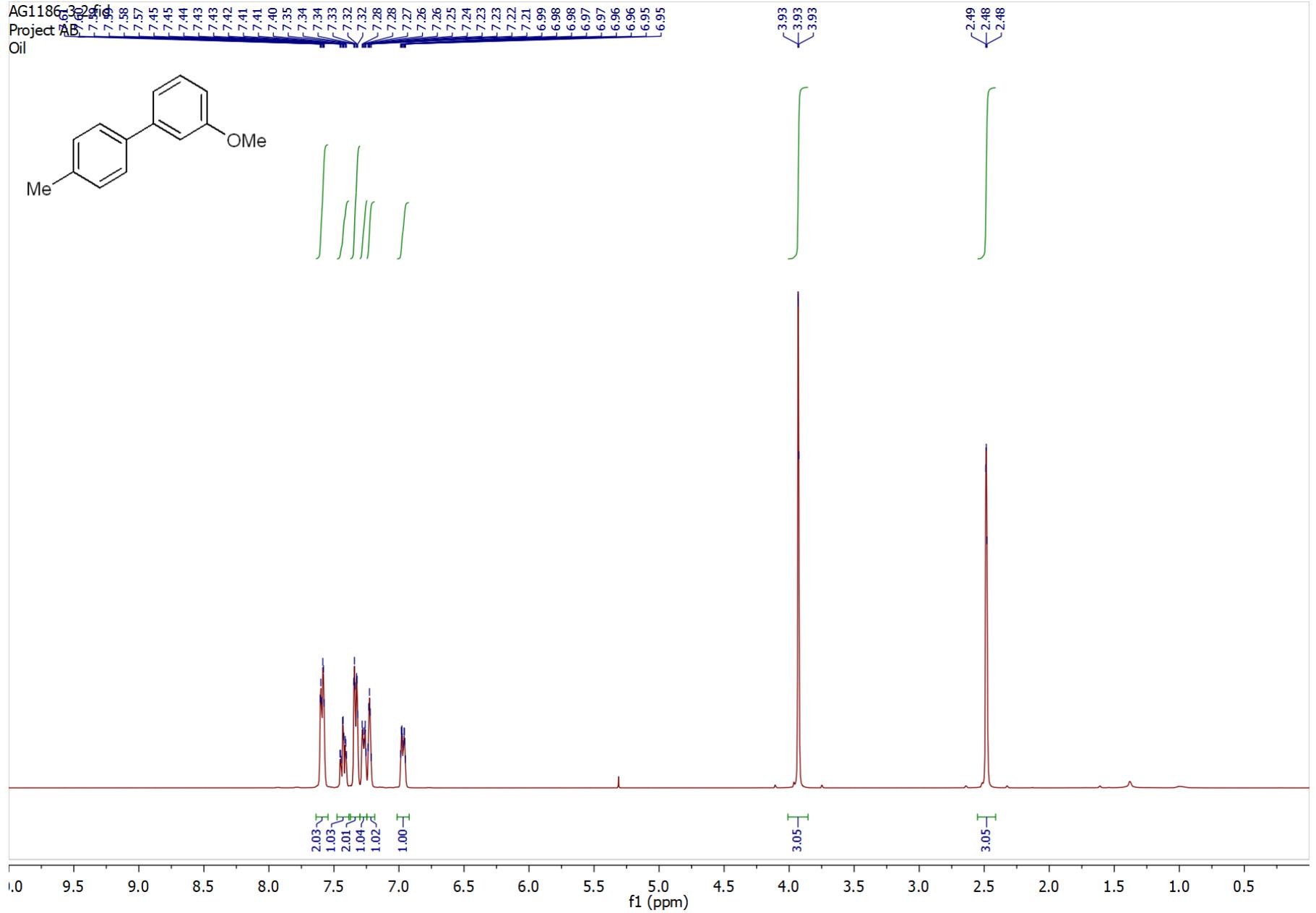


Compound 3k

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Project AB_
Oil

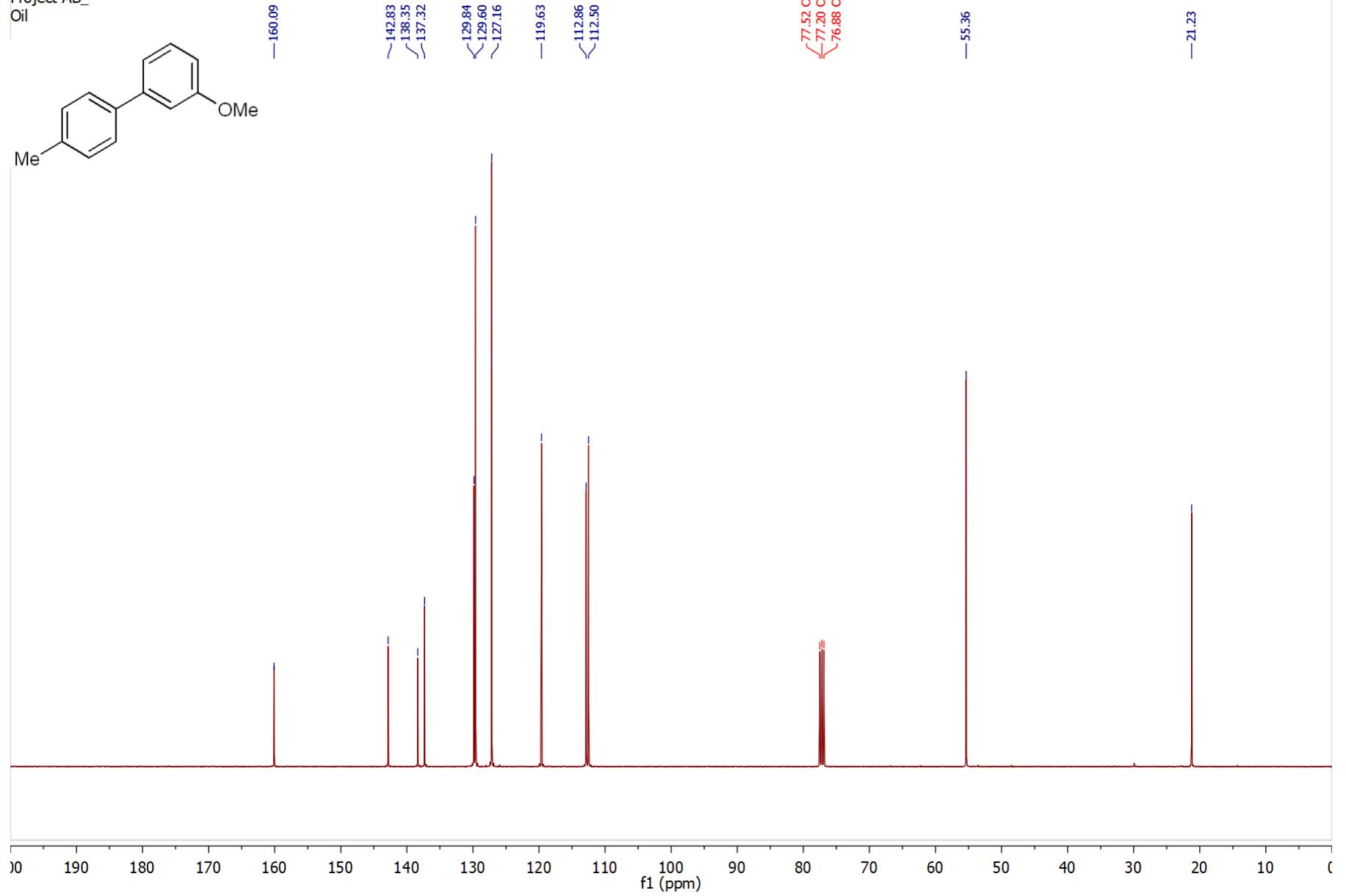
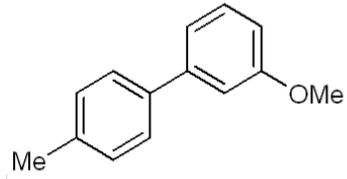


Compound 3I



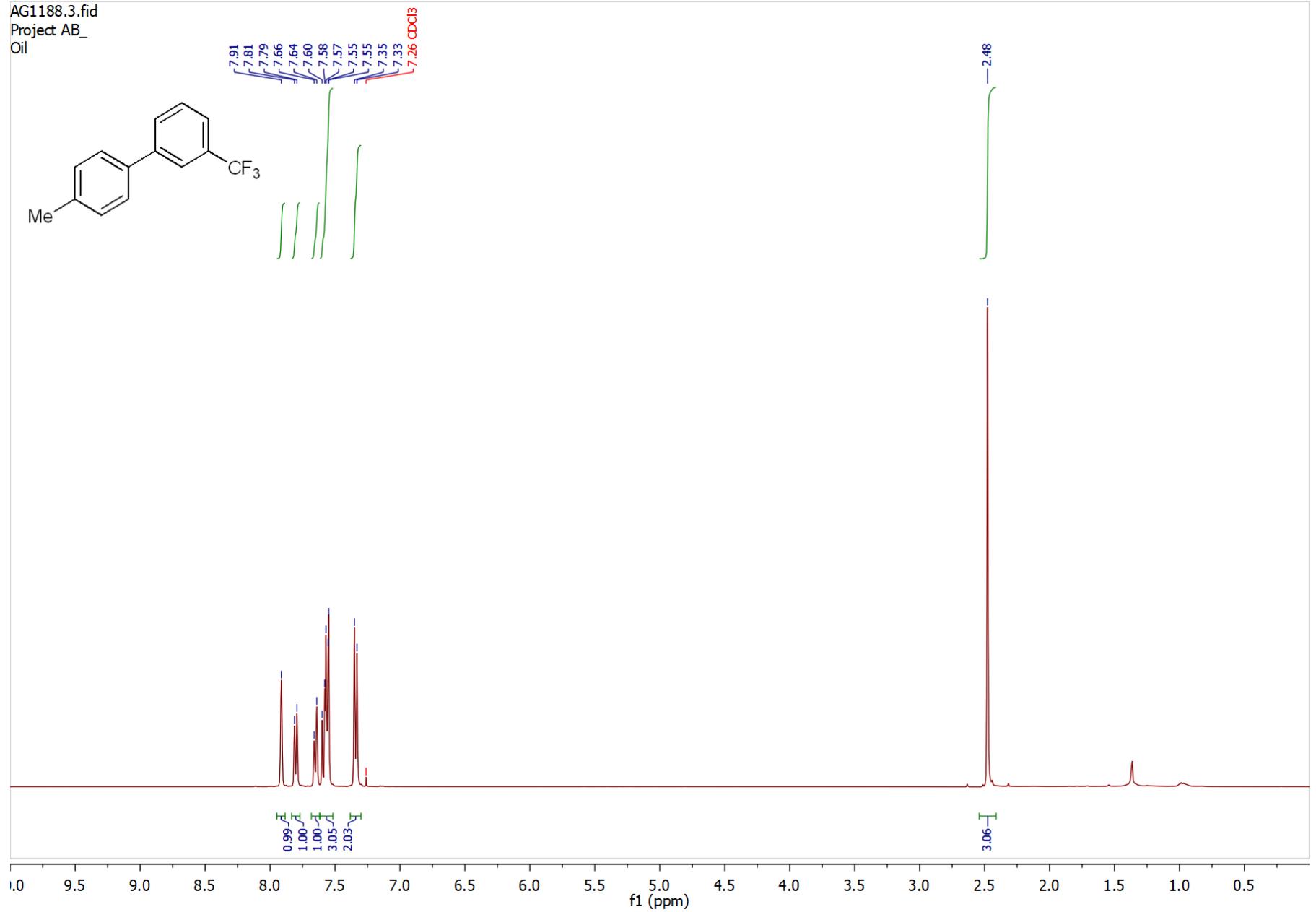
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Project AB_
Oil



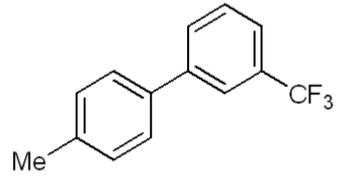
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Project AB_
Oil

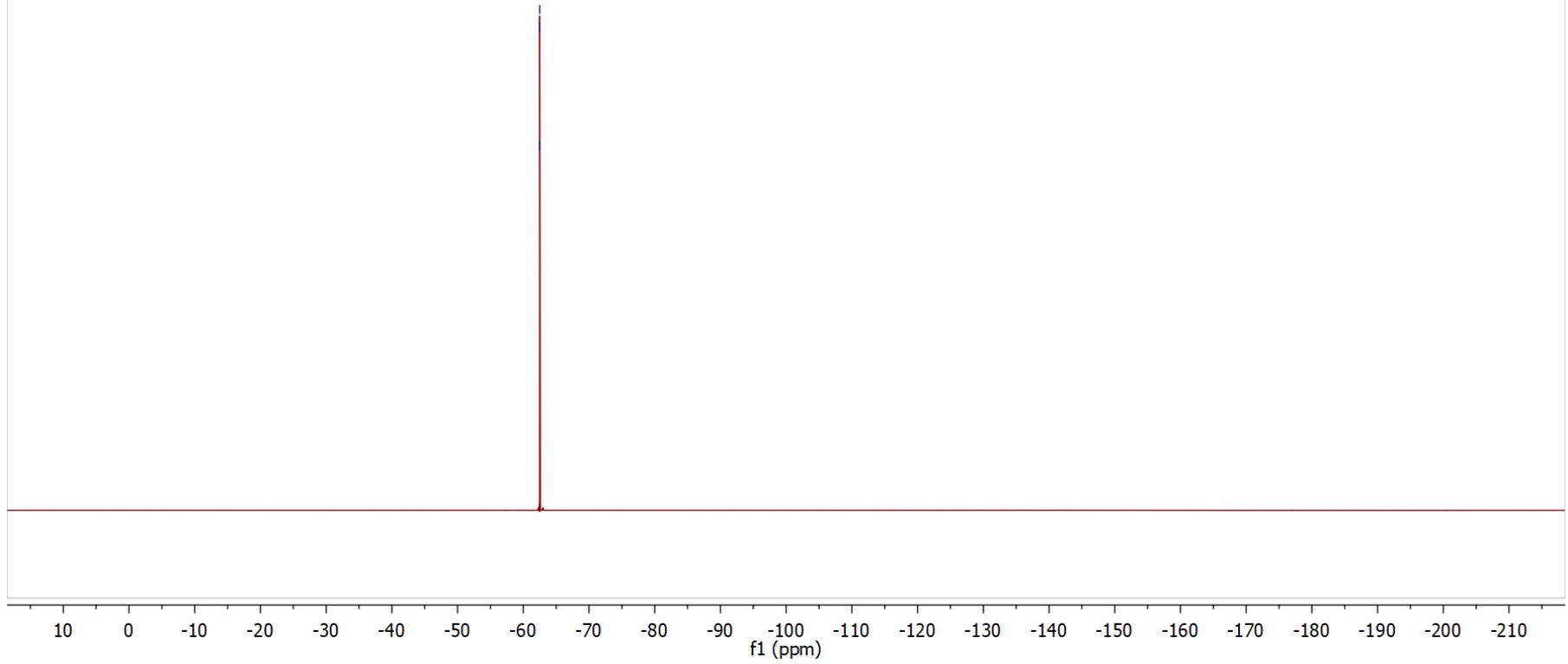


Compound 3m

AG1188.2.fid
Project AB_
Oil

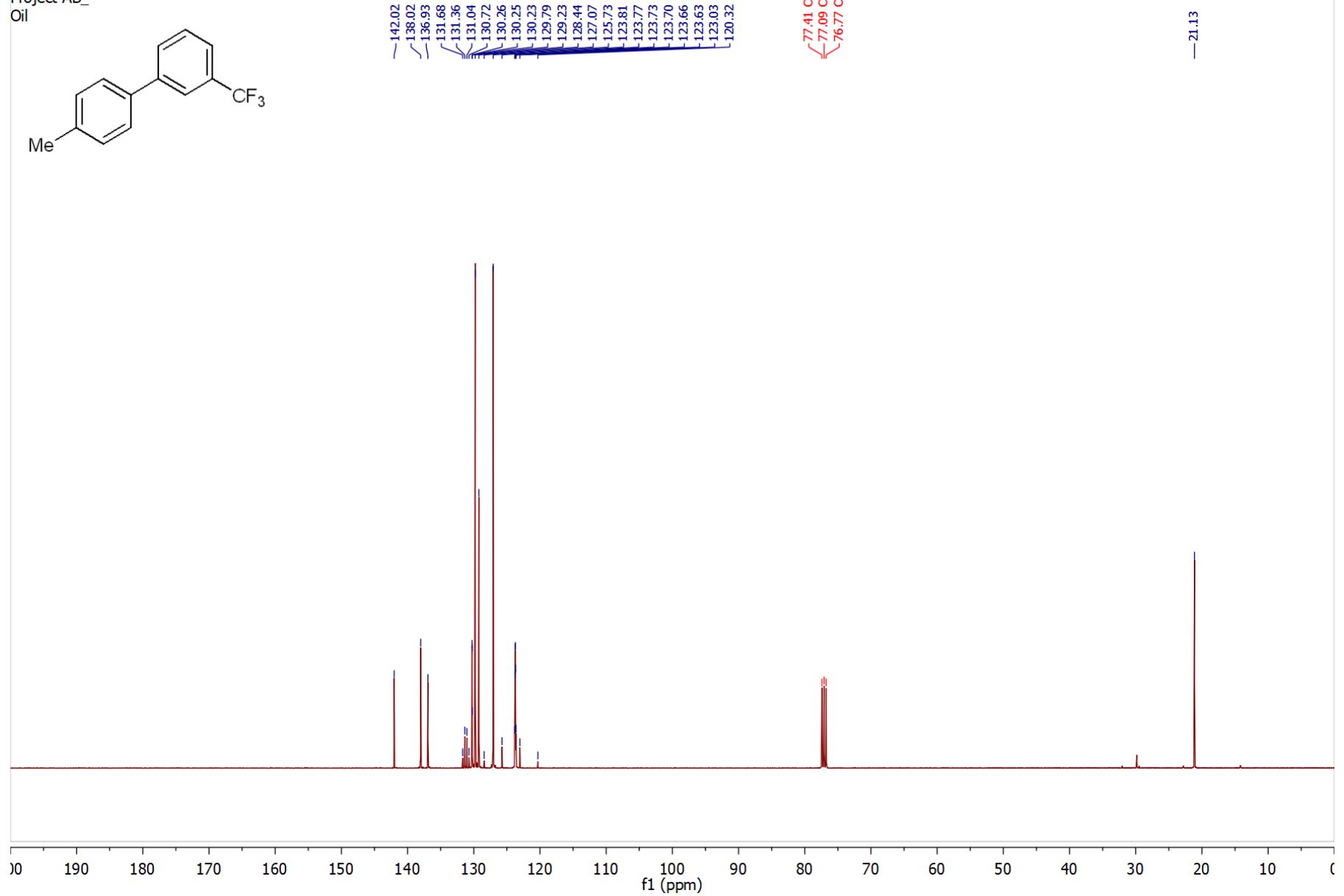
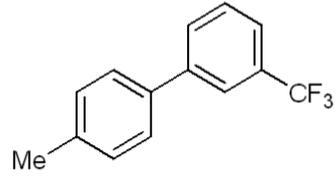


-62.50
-62.51
-62.51



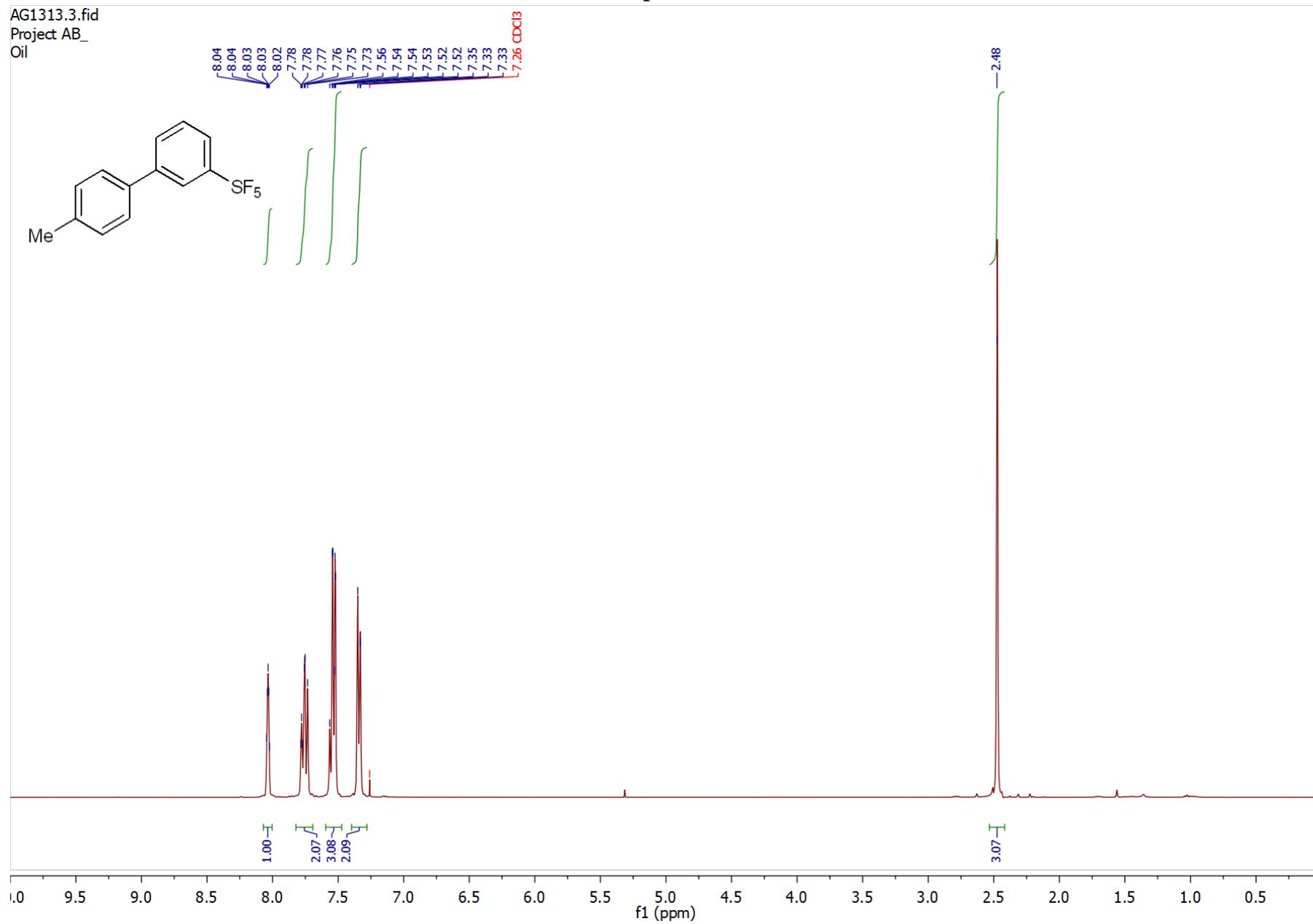
Compound 3m

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Project AB_
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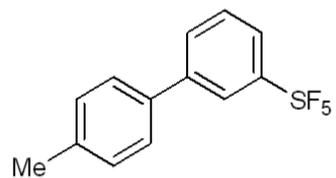
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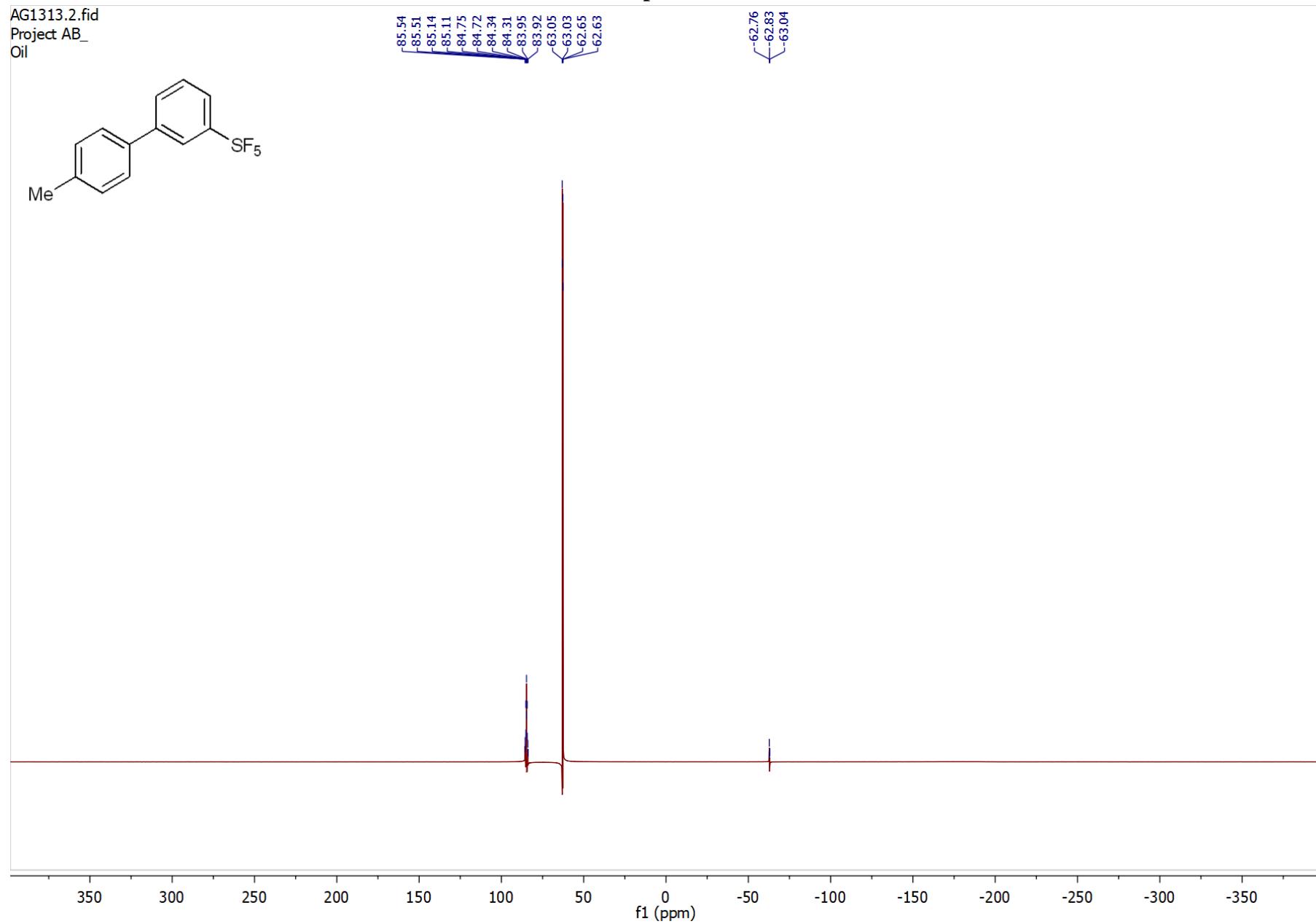


Compound 3n

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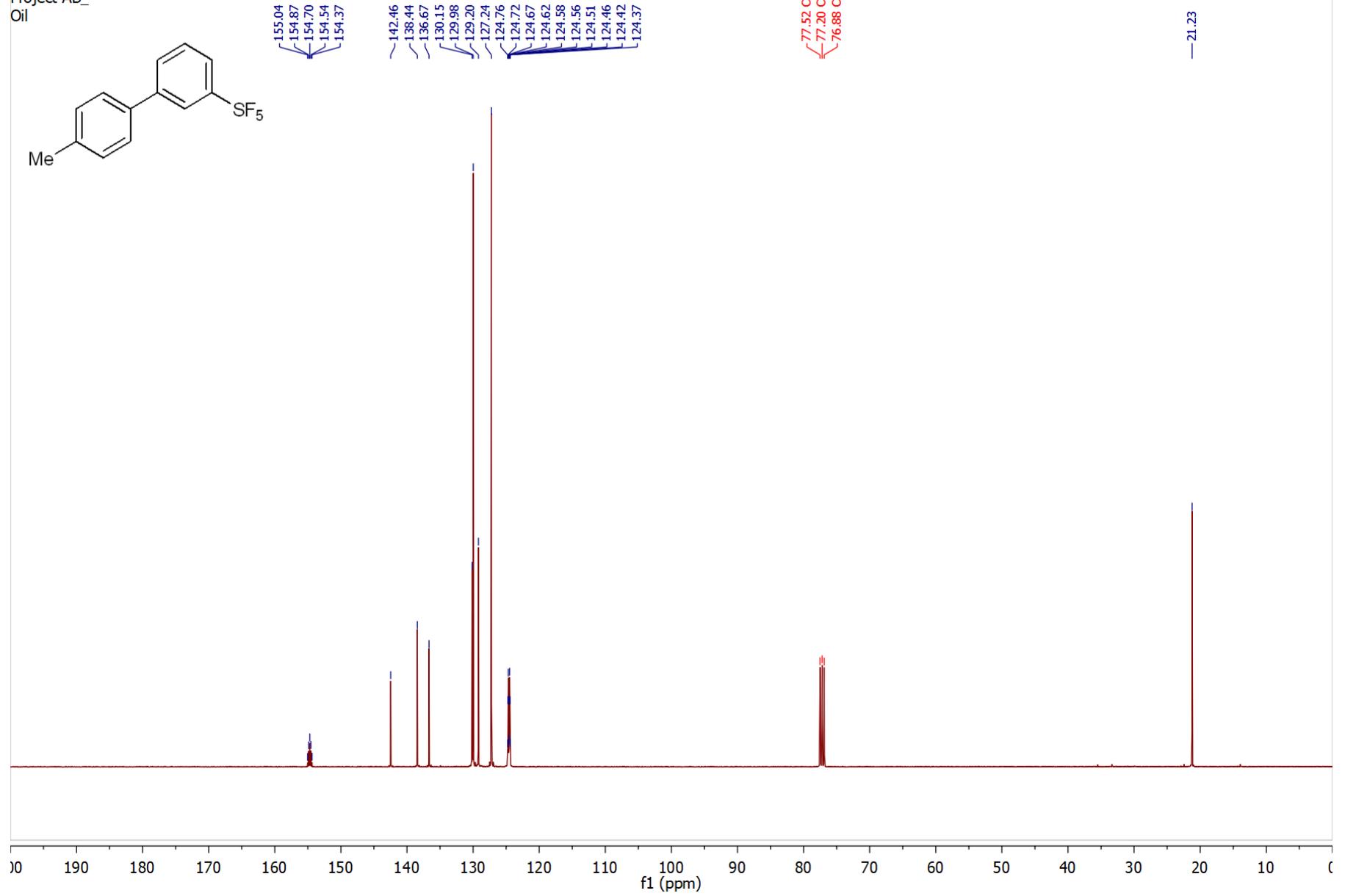
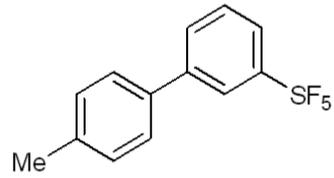


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85.11
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84.72
84.34
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83.95
83.92
63.05
63.03
62.65
62.63
-62.76
-62.83
-63.04



Compound 3n

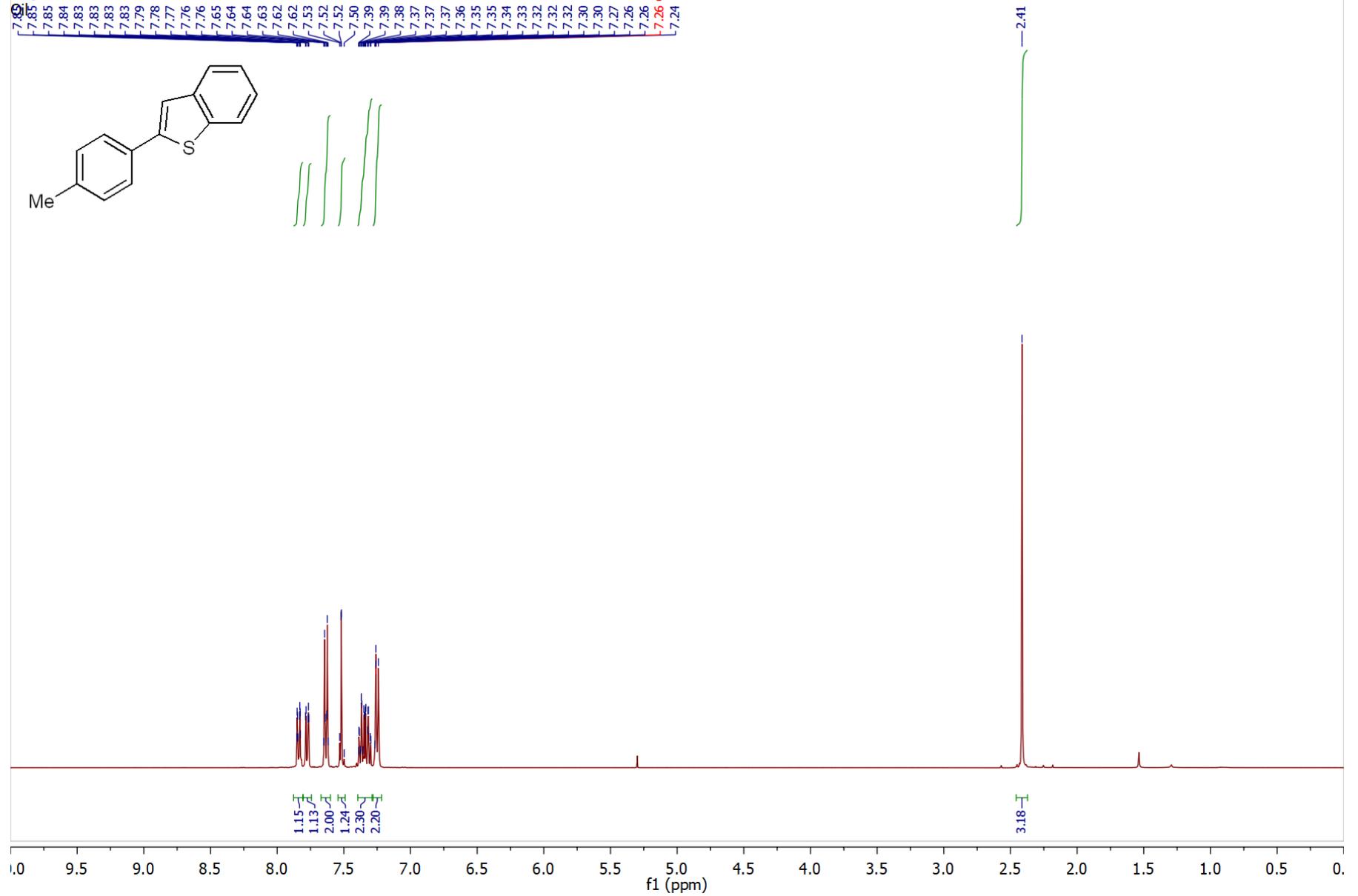
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Project AB_
Oil



Compound 3o

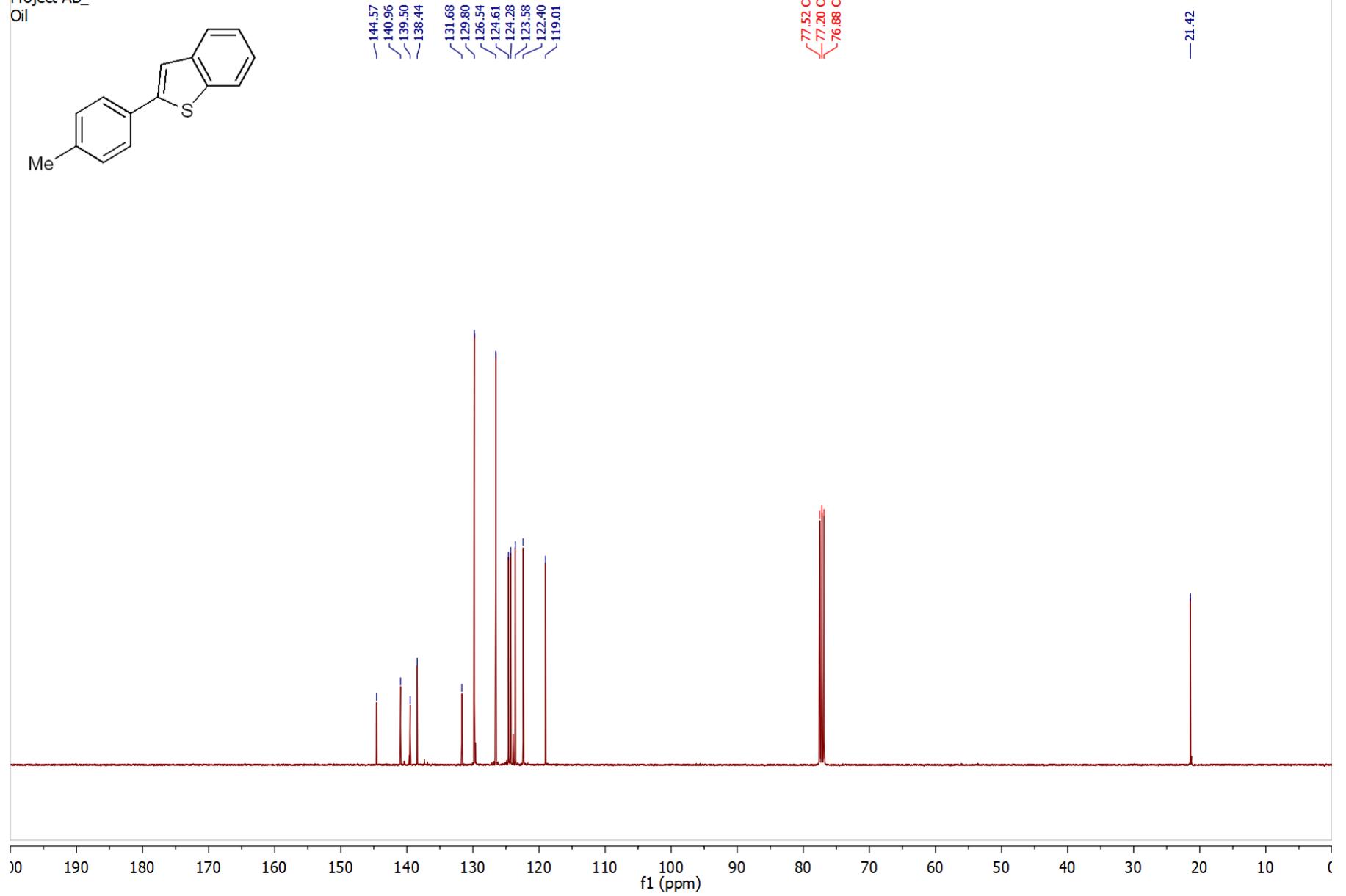
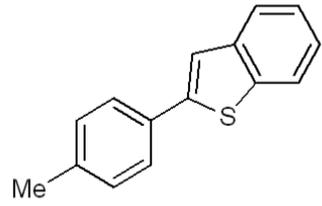
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Project AB



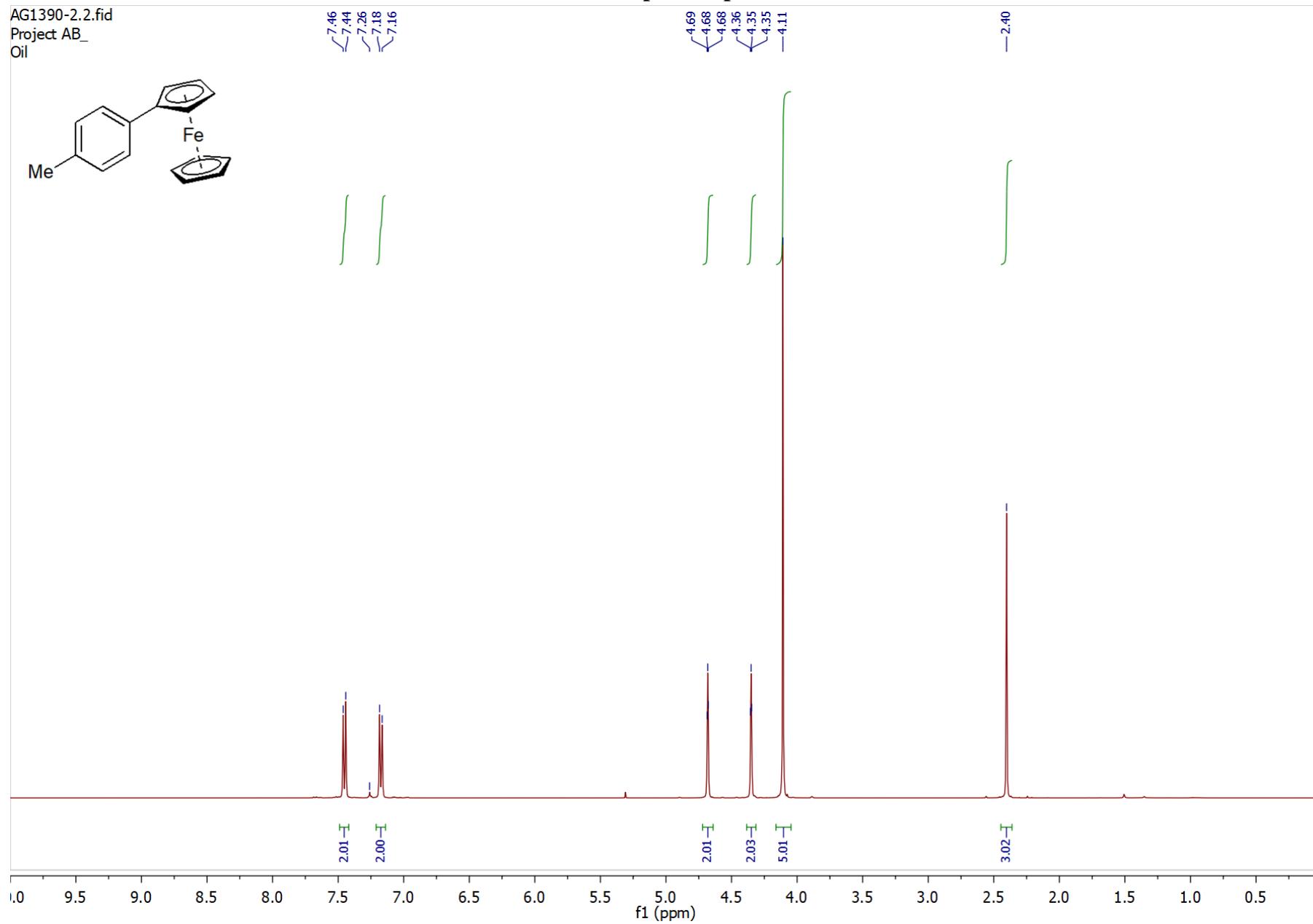
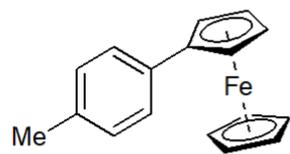
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AG1190.4.fid
Project AB_
Oil



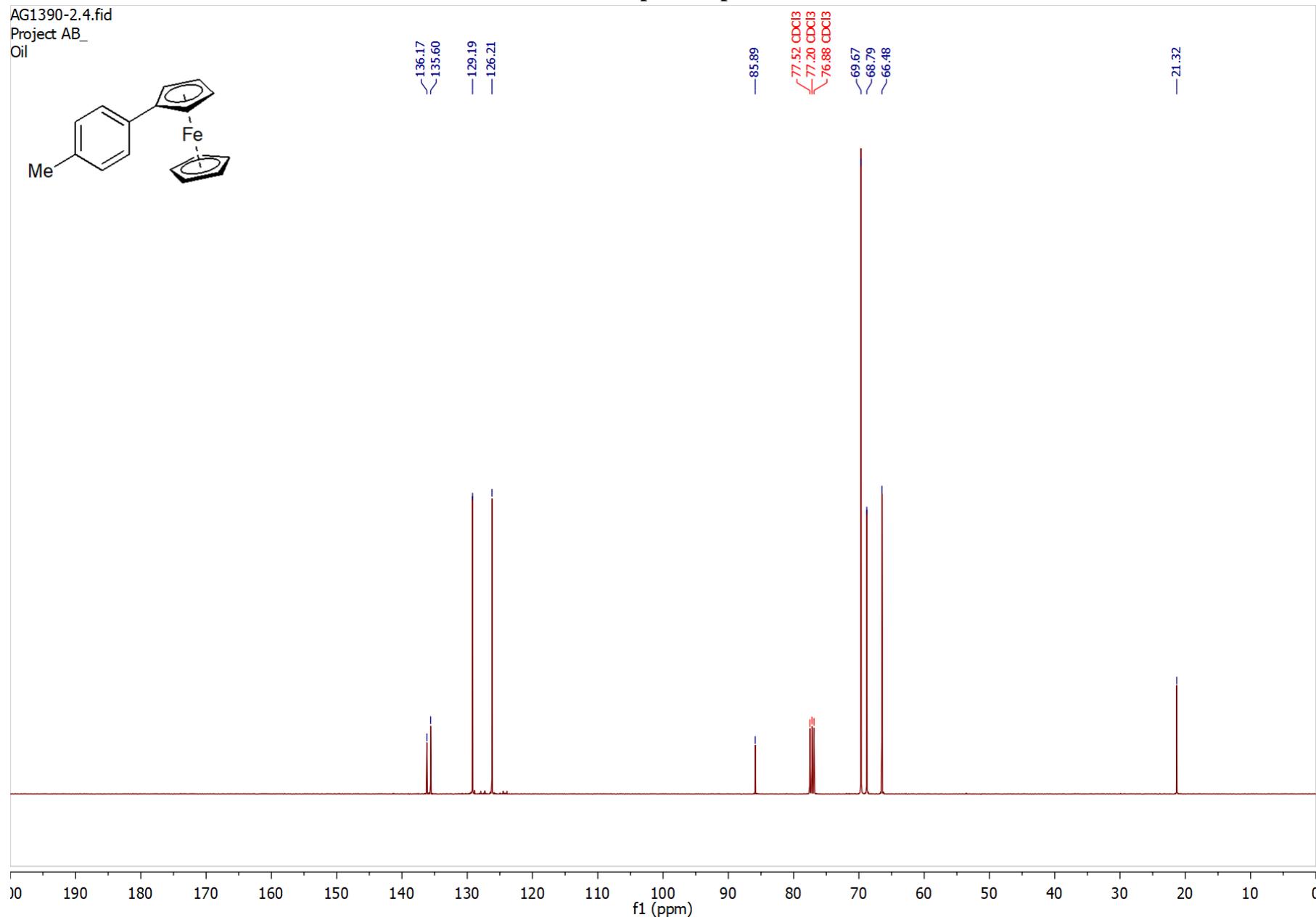
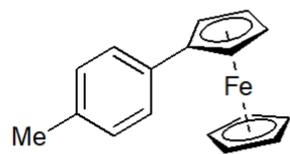
Compound 3p

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Project AB_
Oil



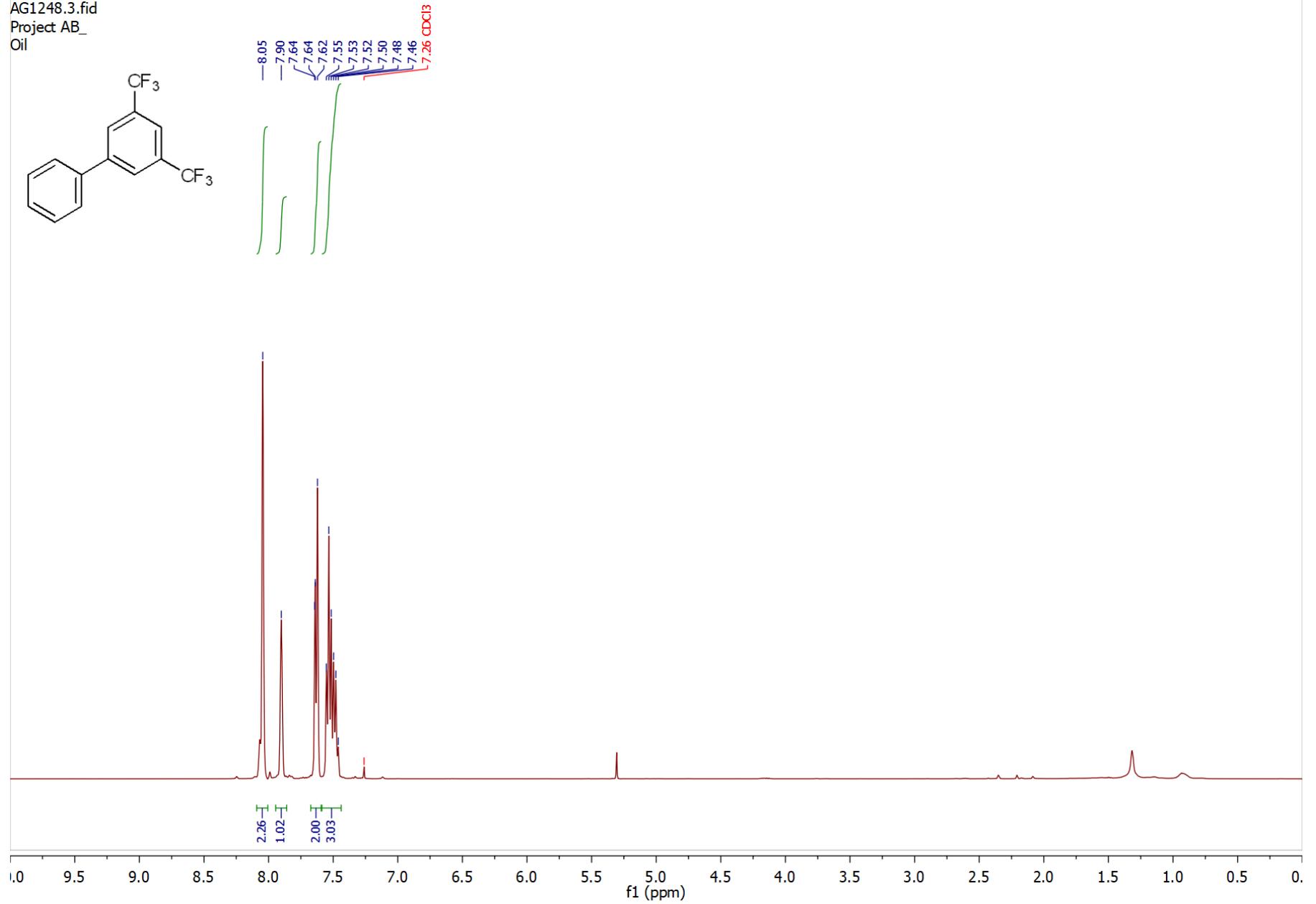
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AG1390-2.4.fid
Project AB_
Oil



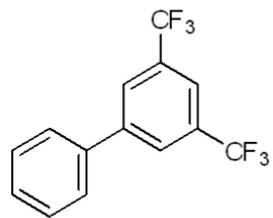
Compound 5a

AG1248.3.fid
Project AB_
Oil

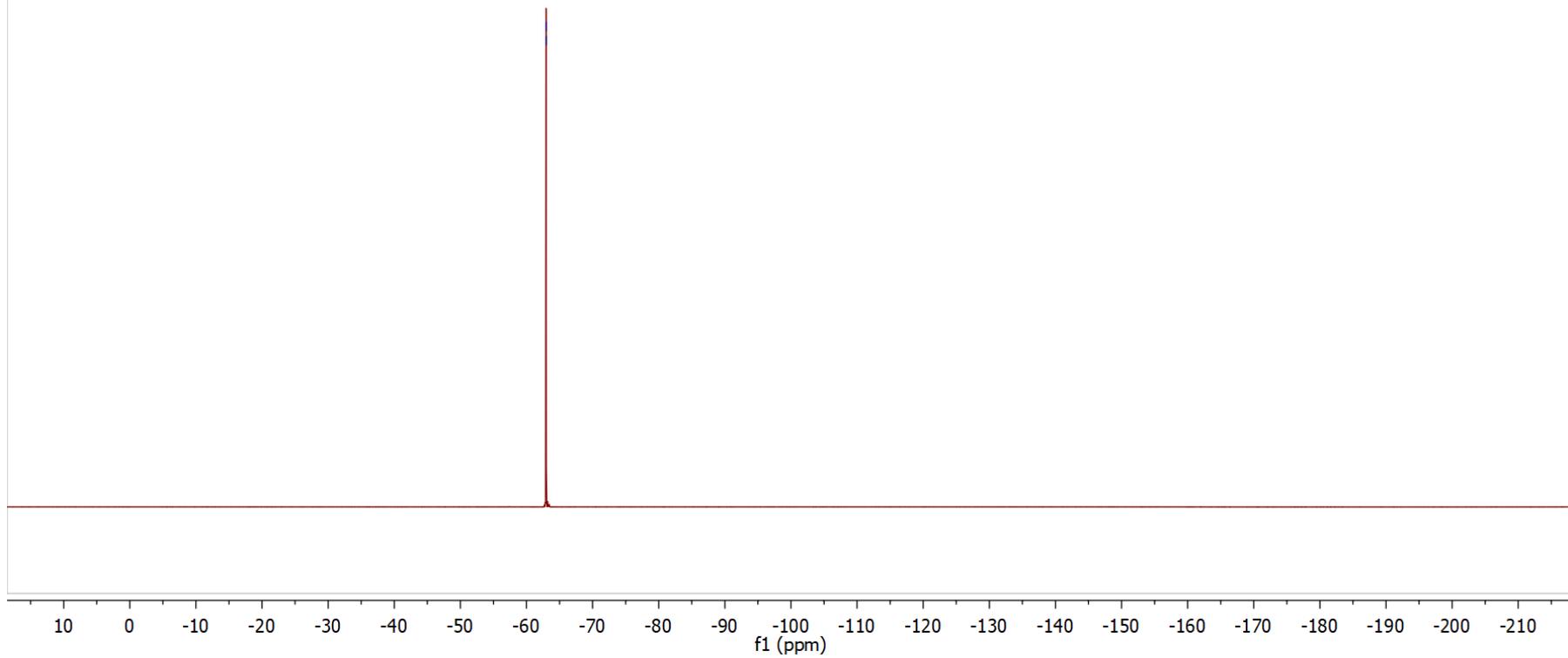


Compound 5a

AG1248.2.fid
Project AB_
Oil

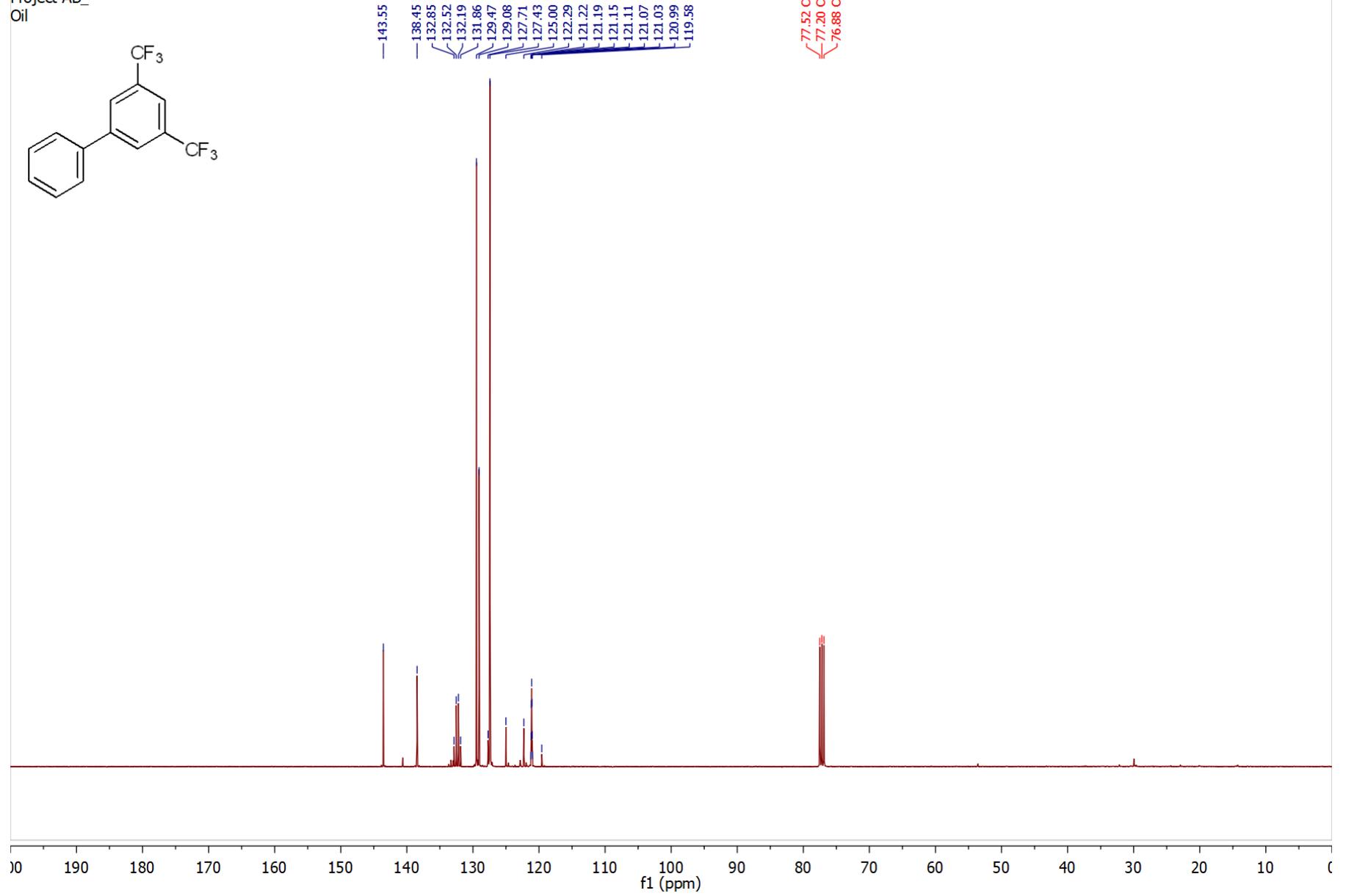
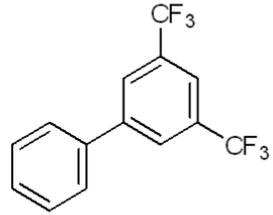


-62.95
-62.96



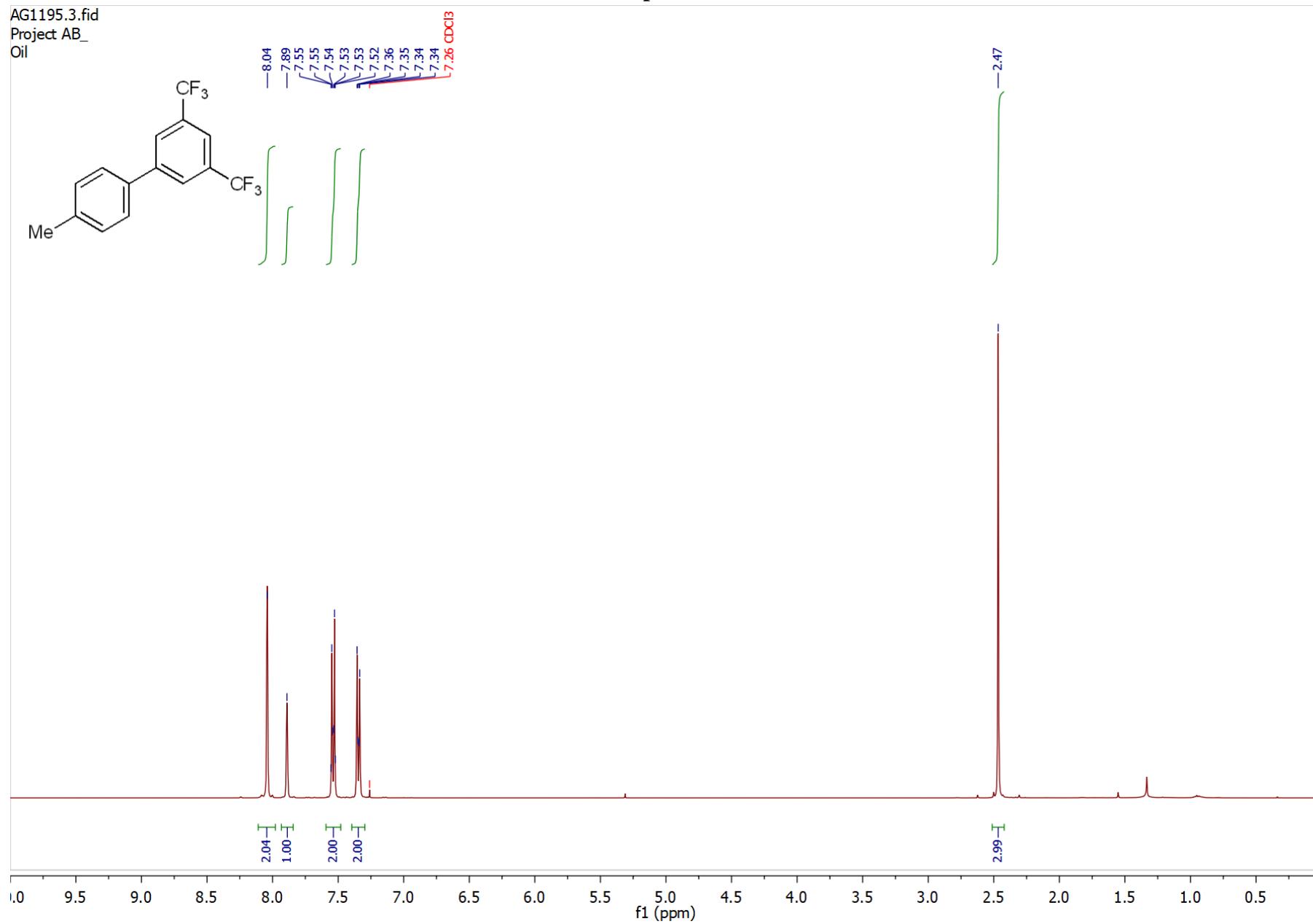
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Project AB_
Oil



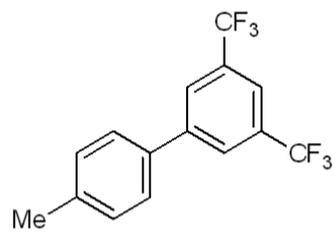
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Project AB_
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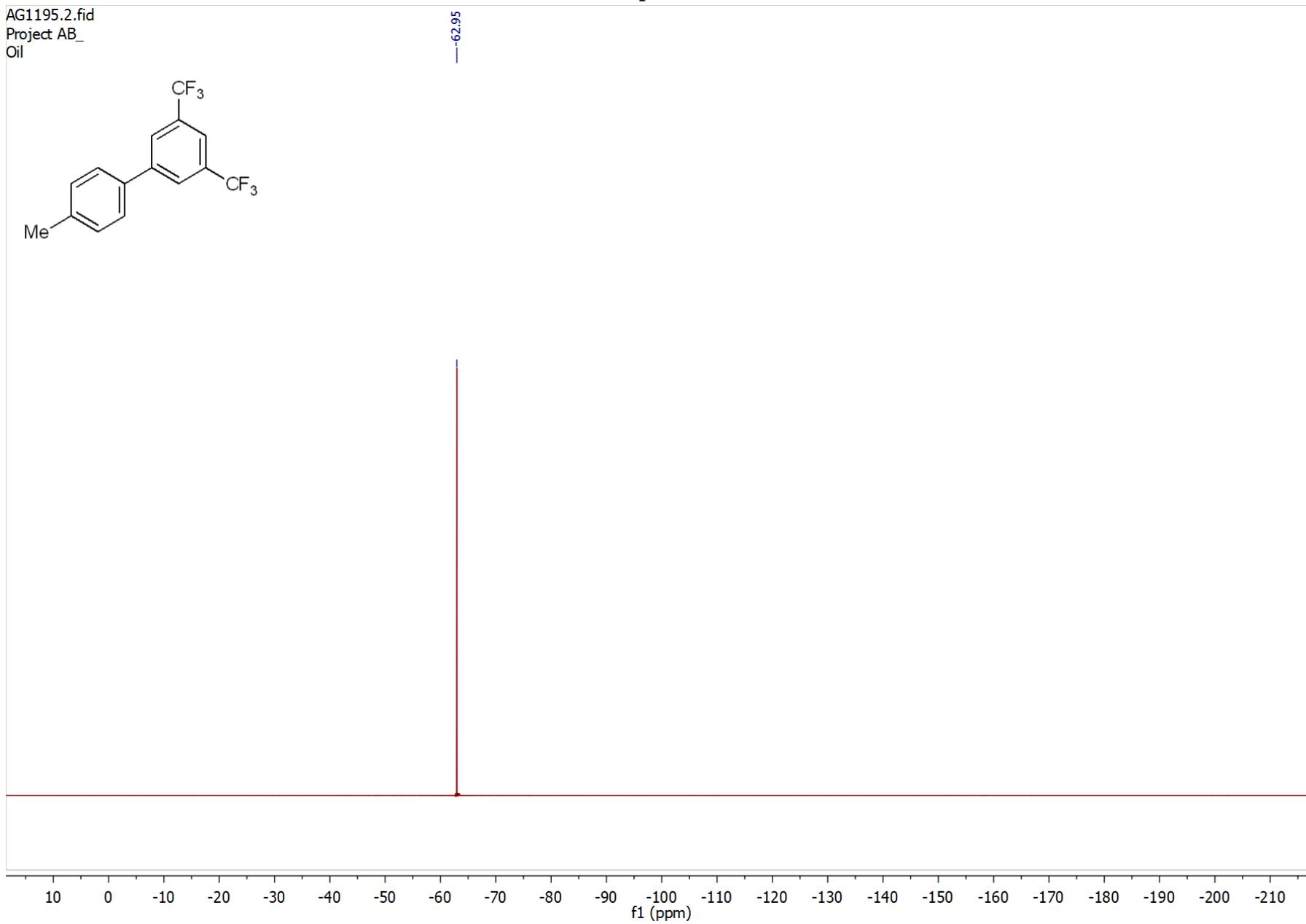


Compound 5b

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Project AB_
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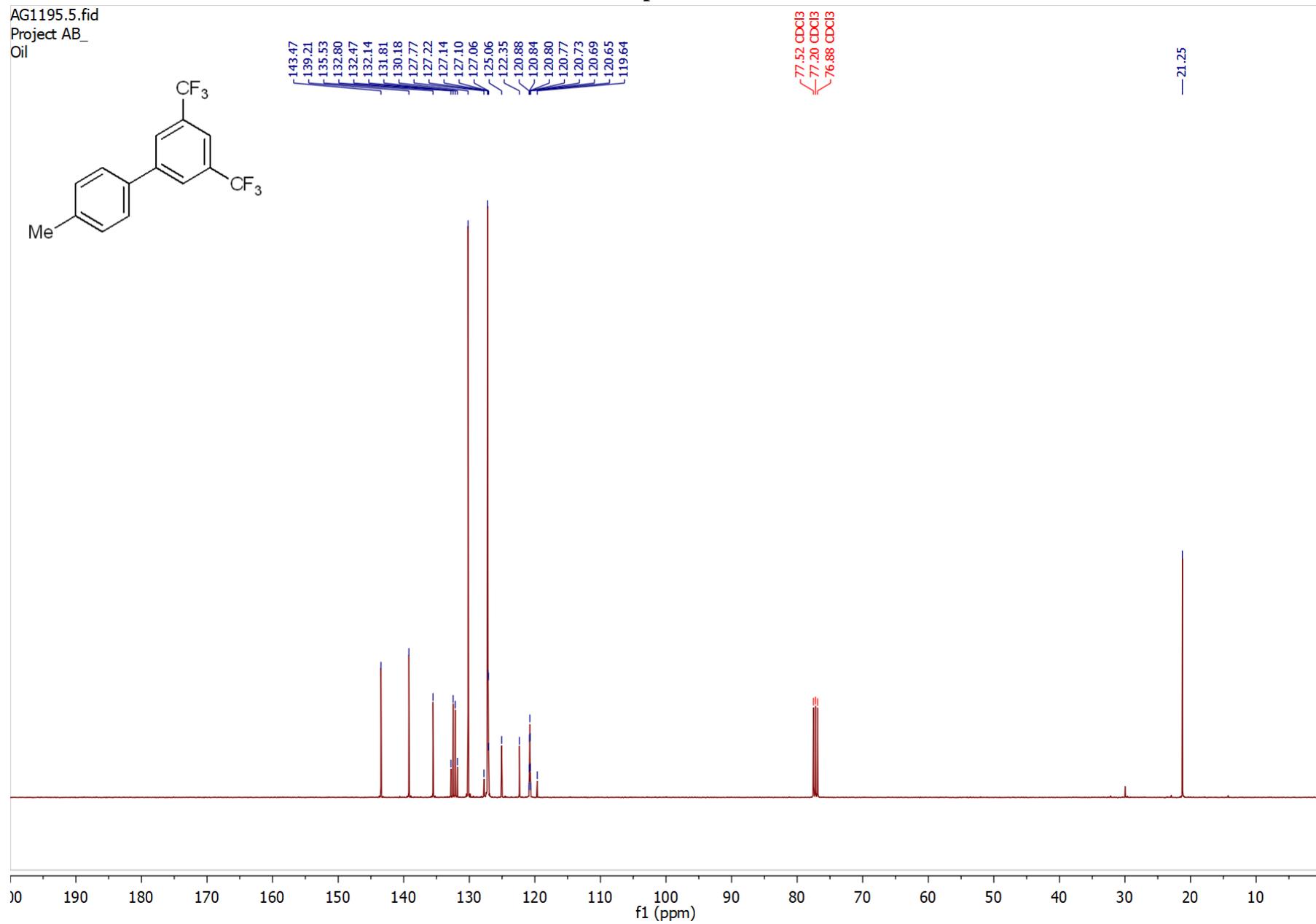


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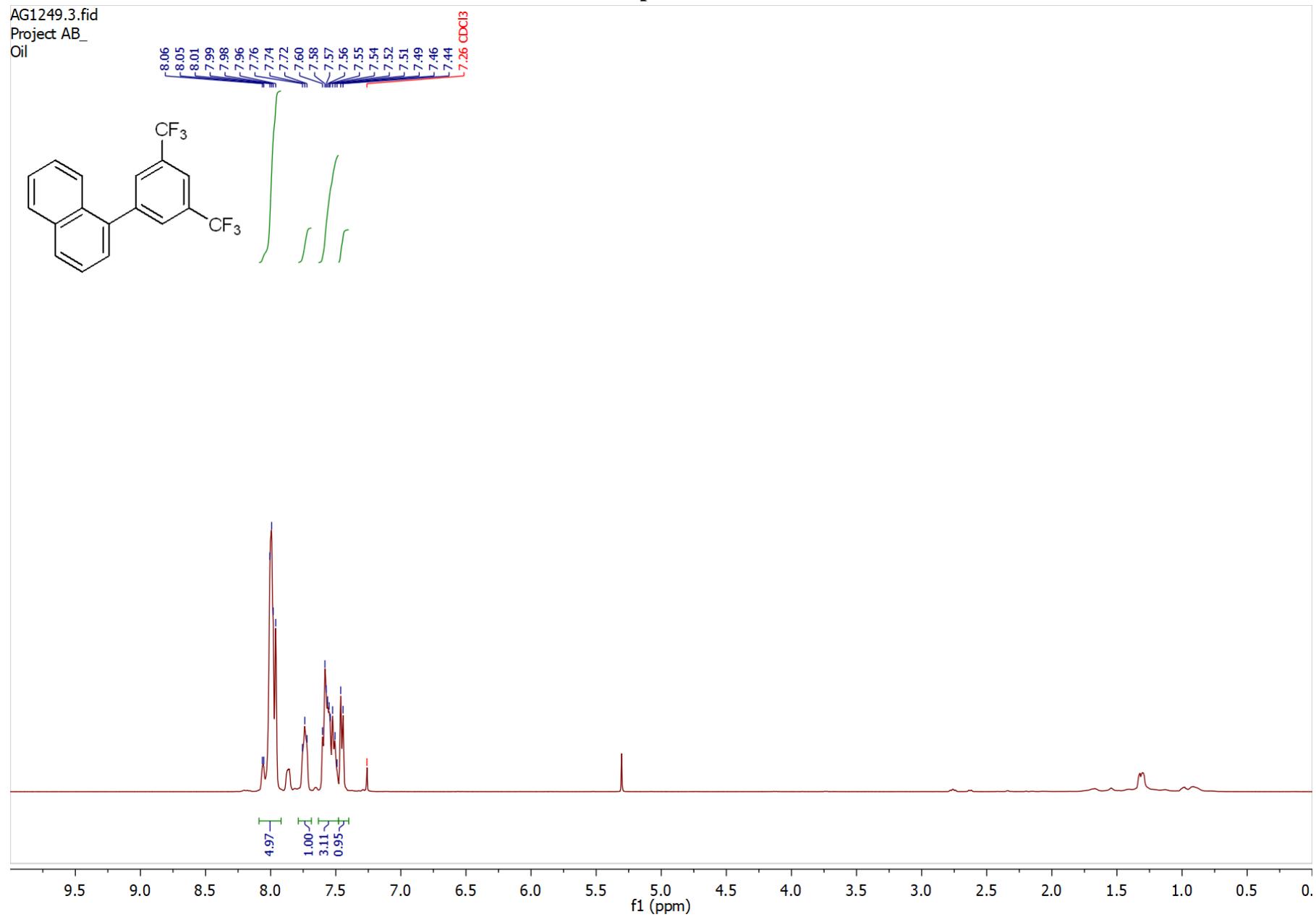
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Project AB_
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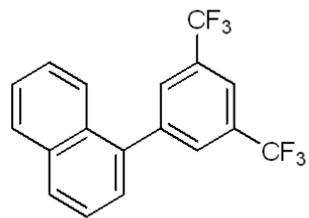
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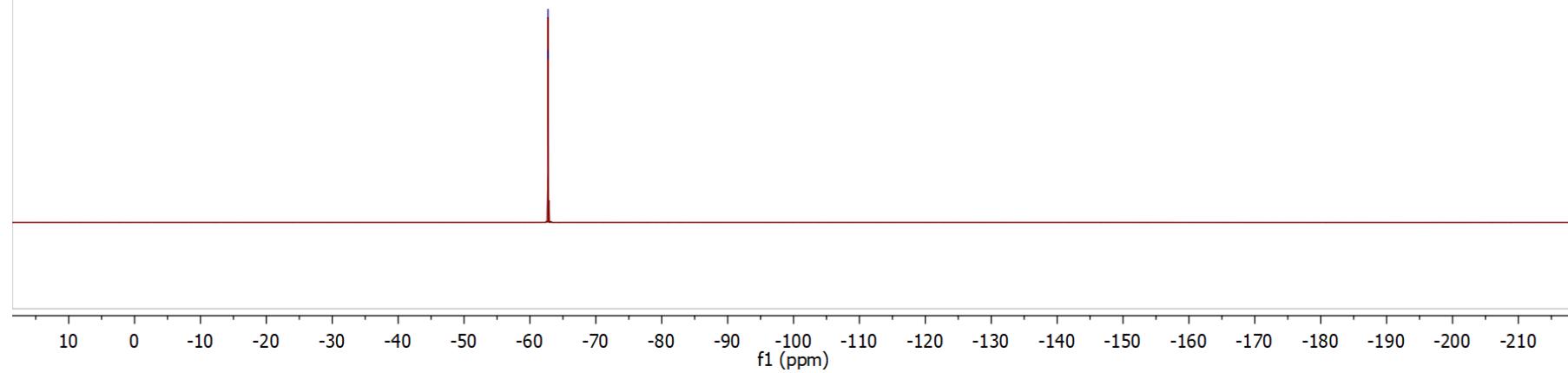


Compound 5c

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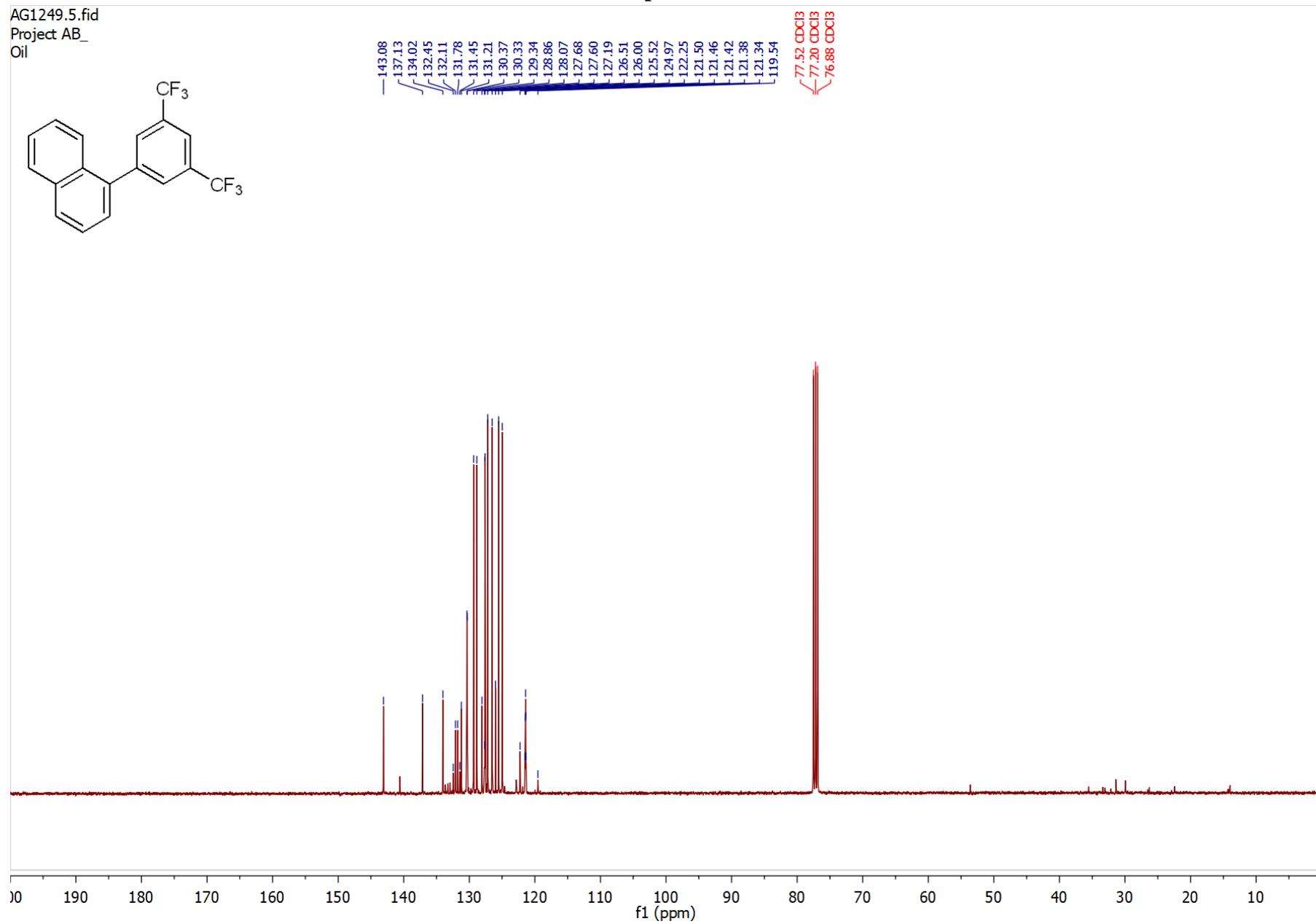
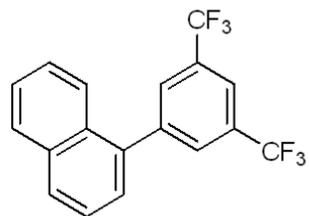


-62.73
-62.74



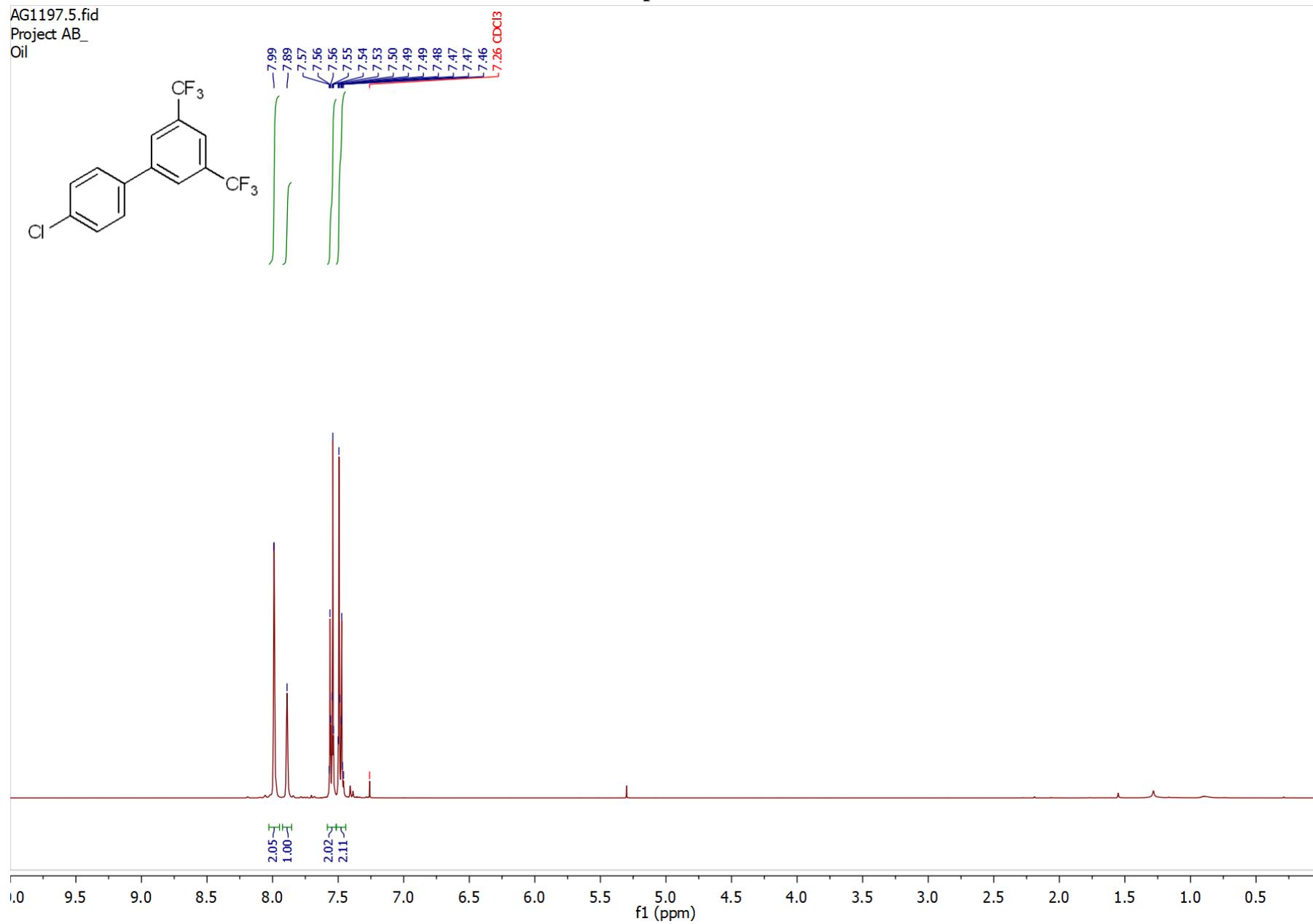
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Project AB_
Oil



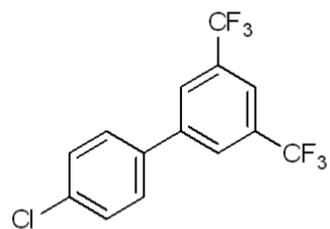
Compound 5d

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Project AB_
Oil

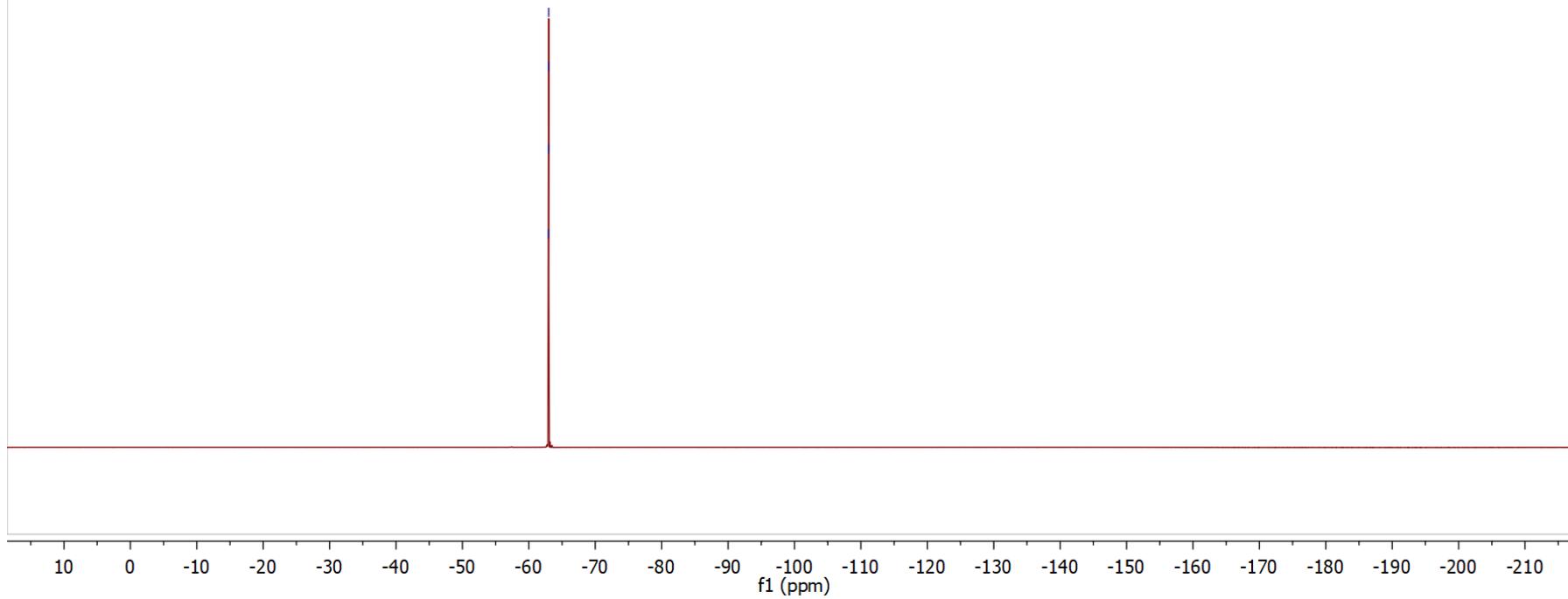


Compound 5d

AG1197.2.fid
Project AB_
Oil

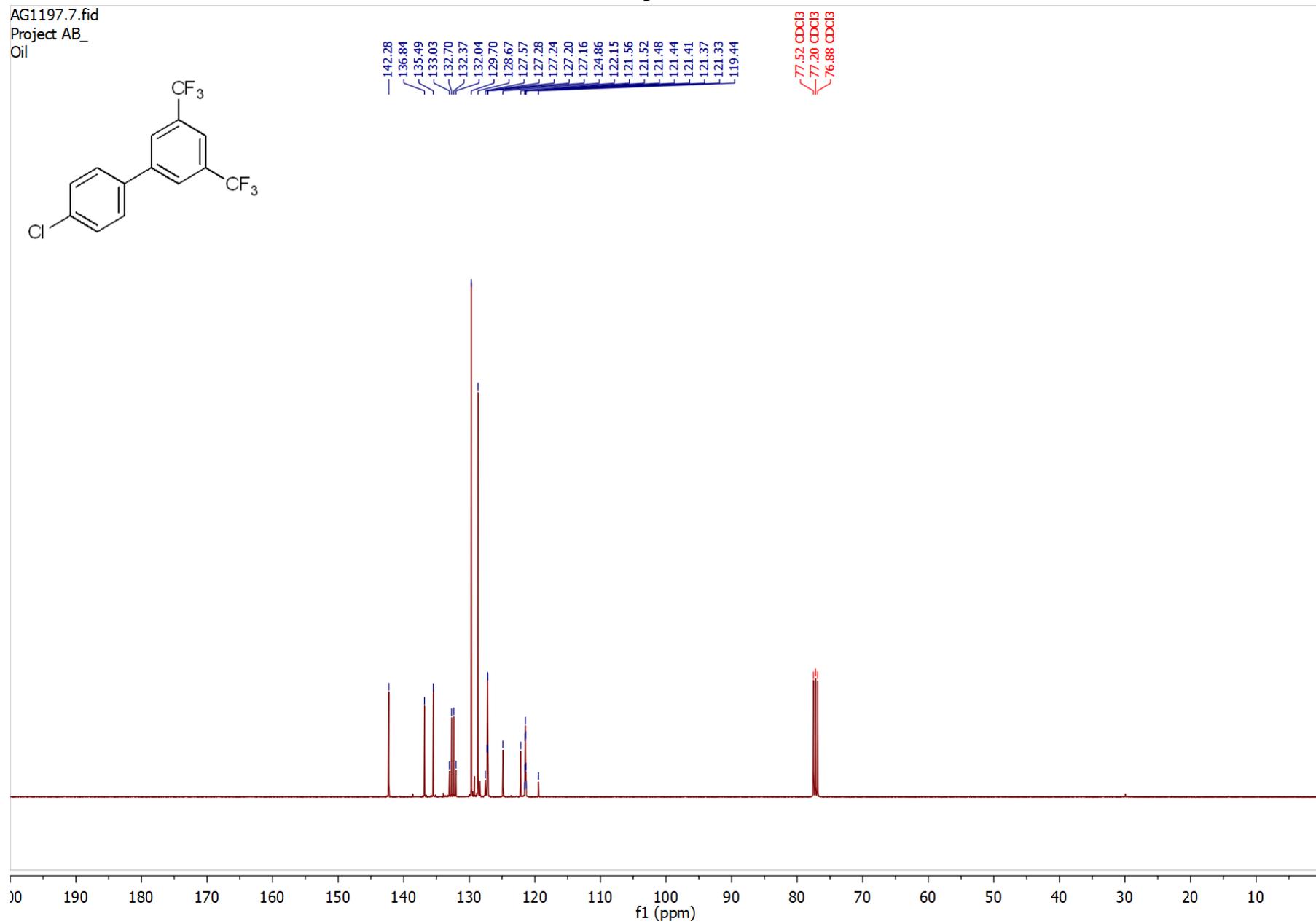
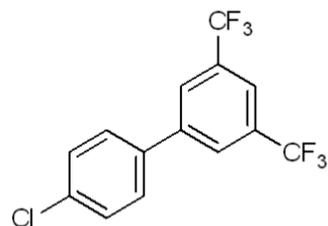


-62.94
-62.95
-62.97
-62.98



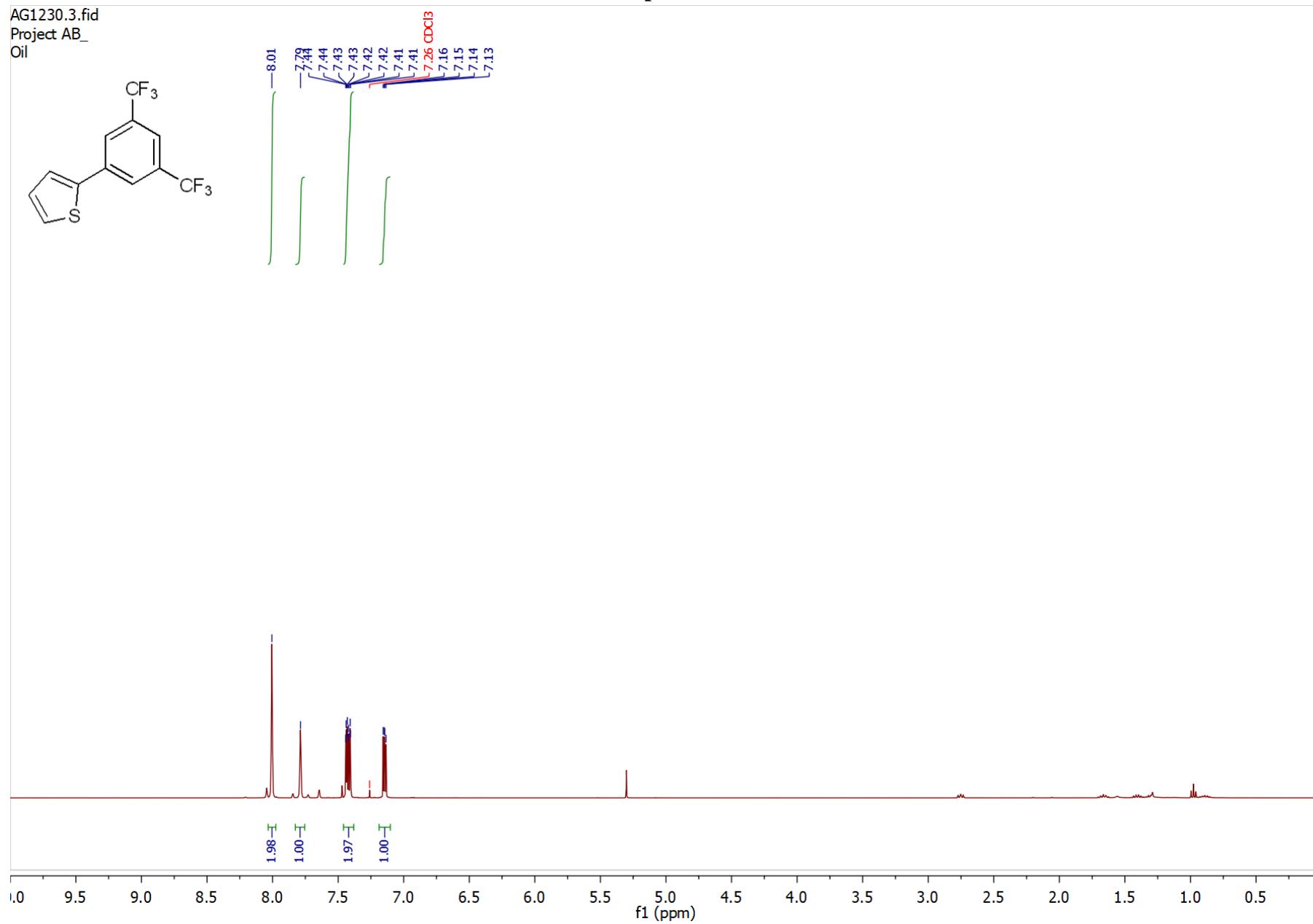
Compound 5d

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Project AB_
Oil



Compound 5e

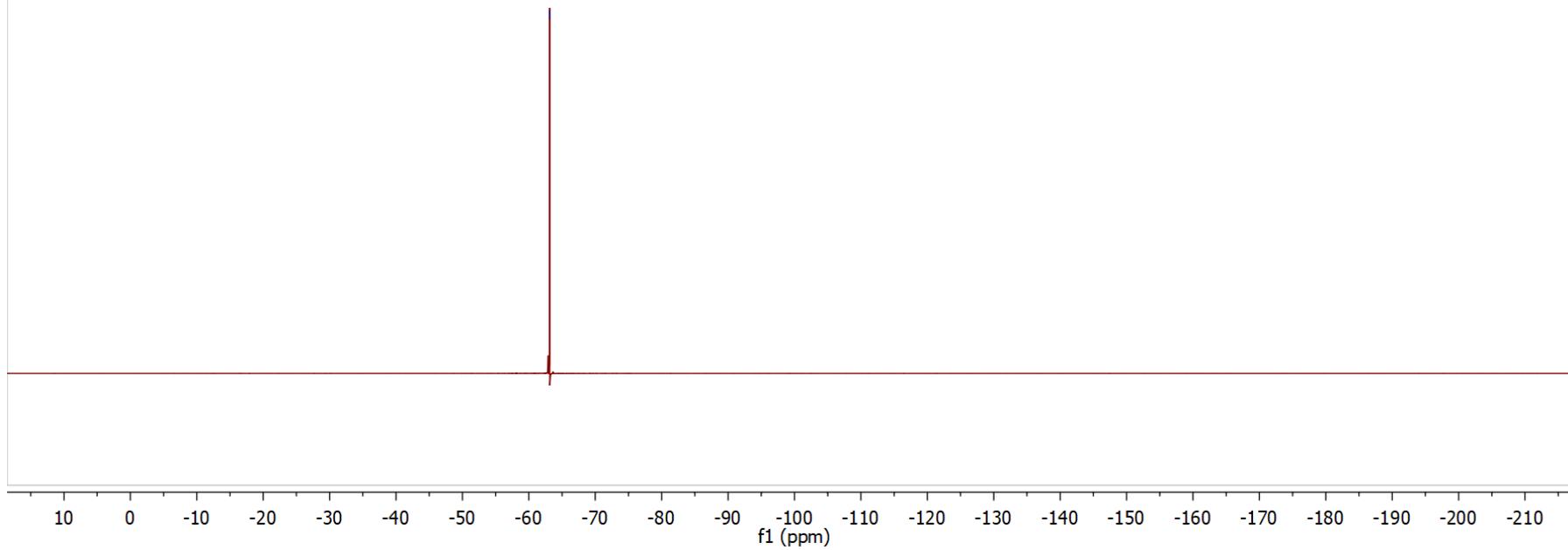
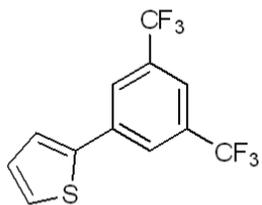
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Project AB_
Oil



Compound 5e

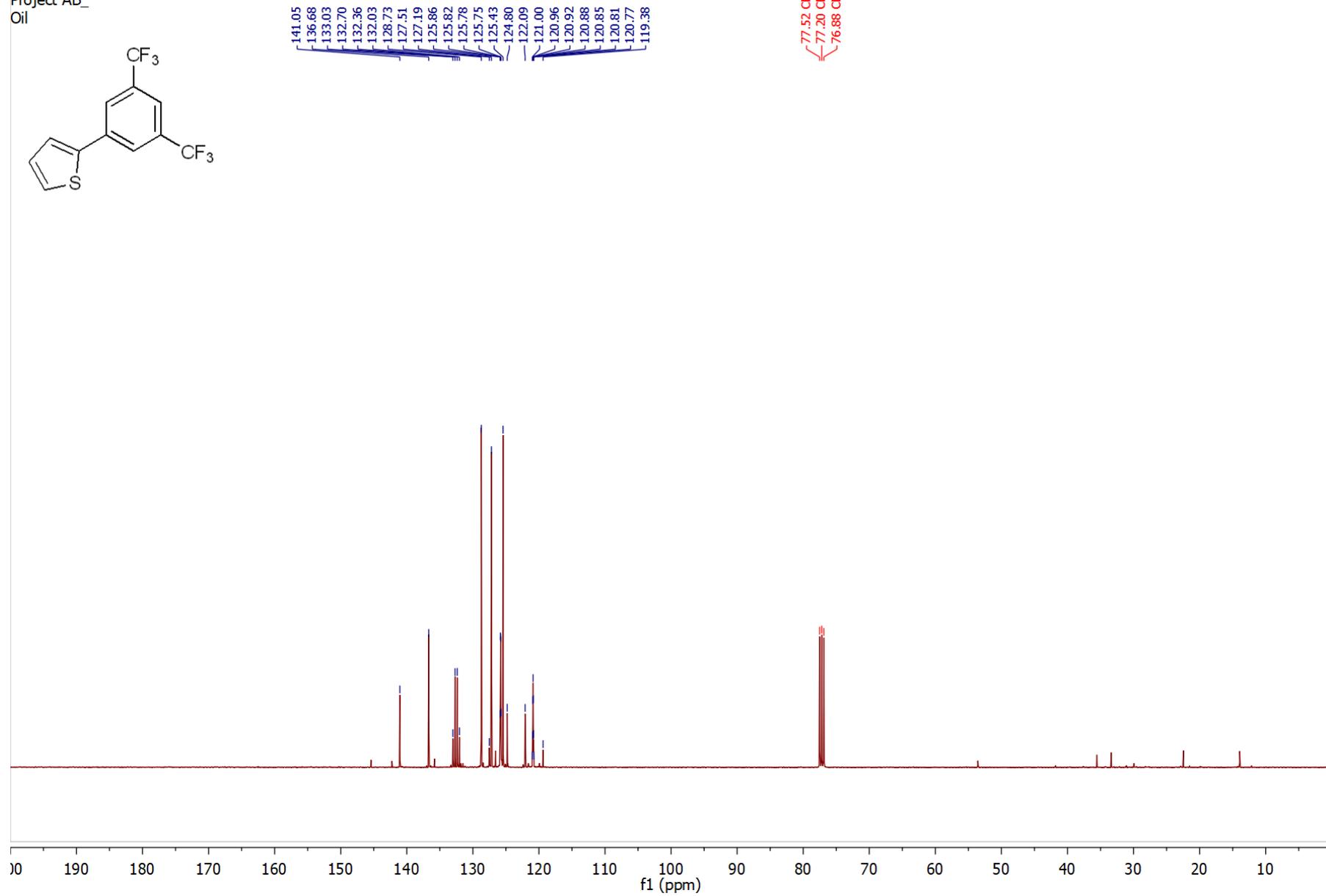
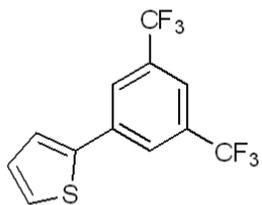
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Project AB_
Oil

63.14



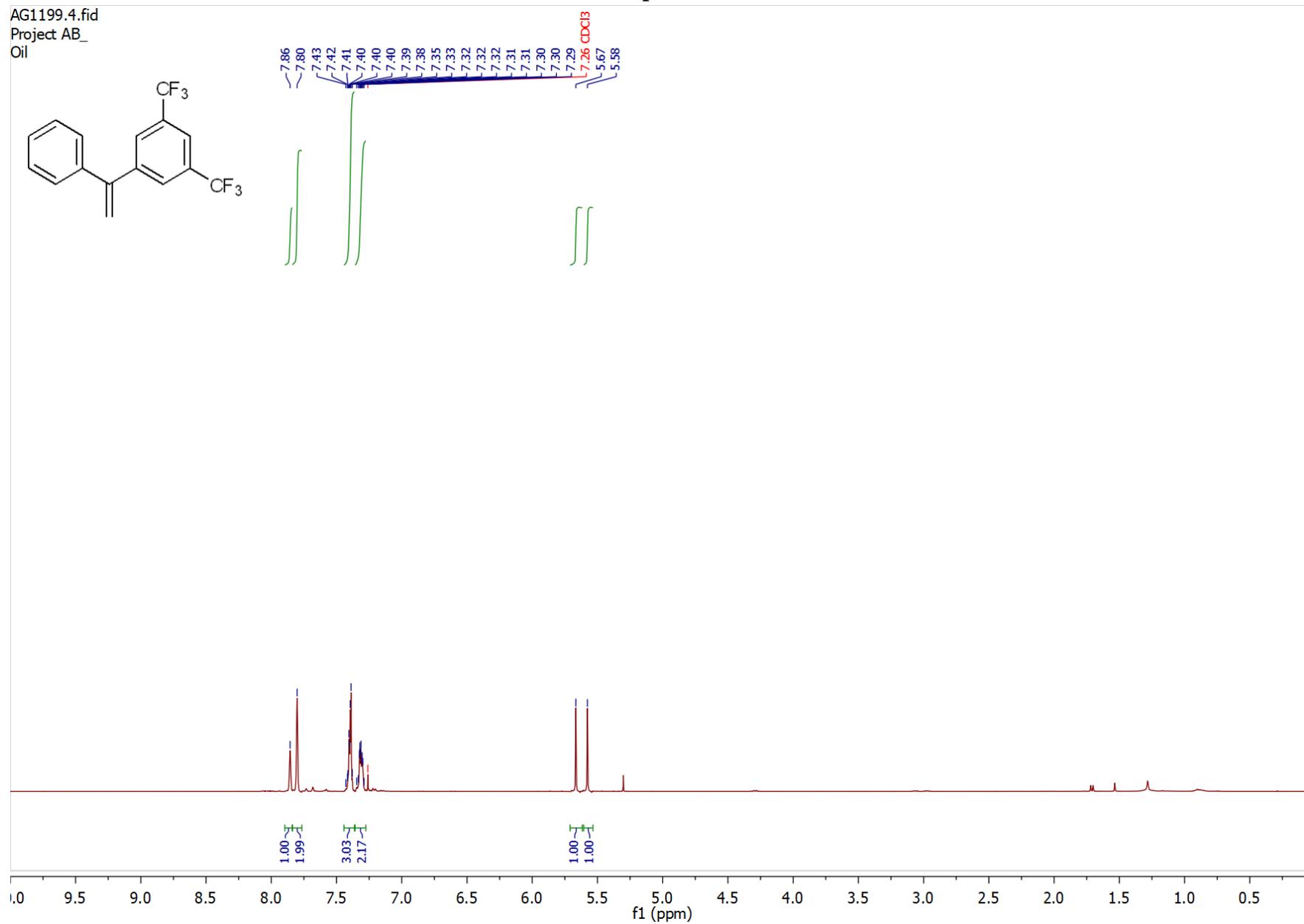
Compound 5e

AG1230.5.fid
Project AB_
Oil



Compound 5f

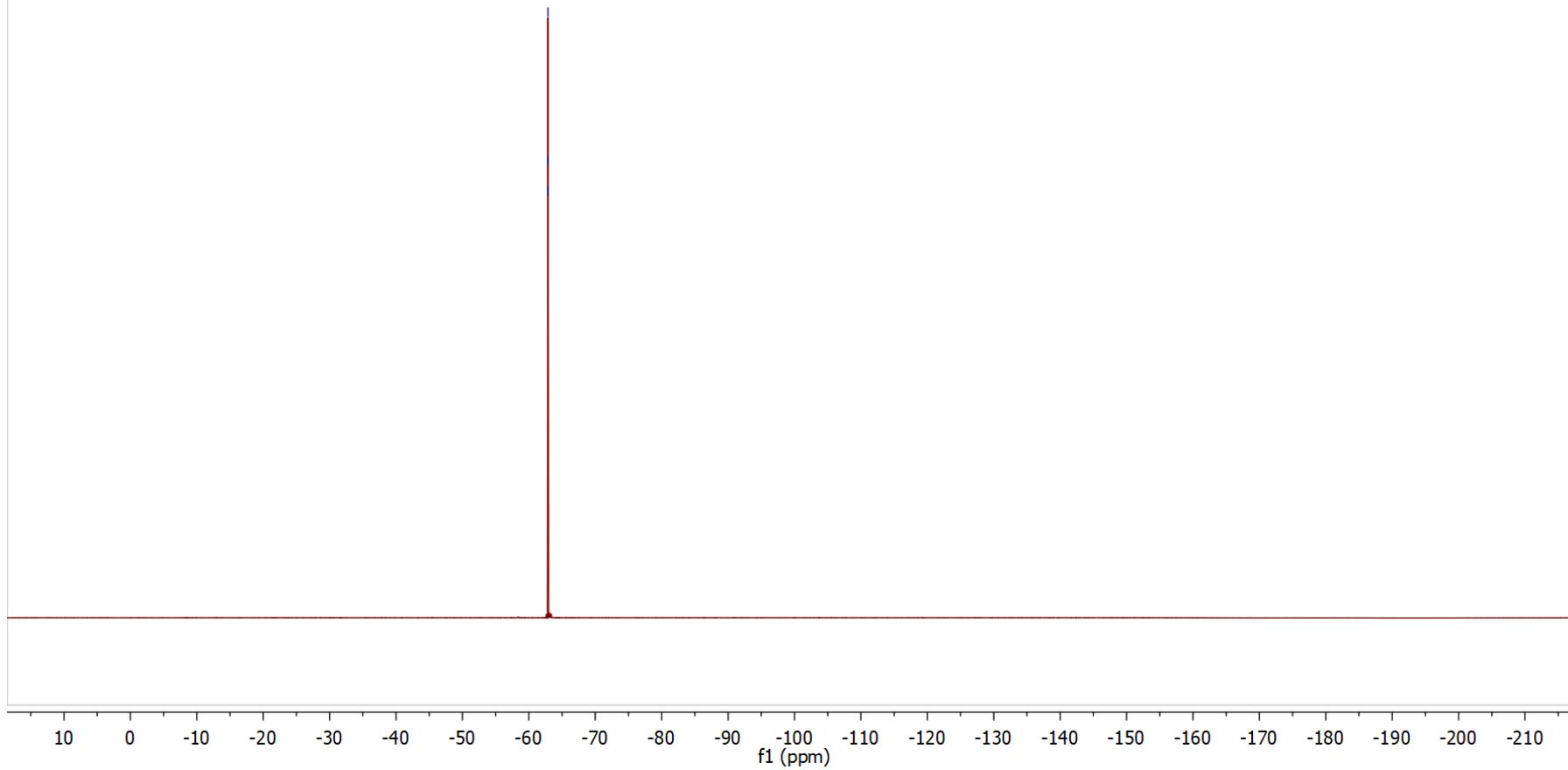
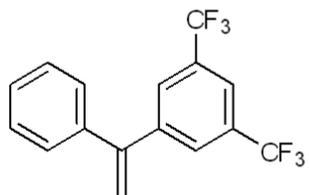
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Project AB_
Oil



Compound 5f

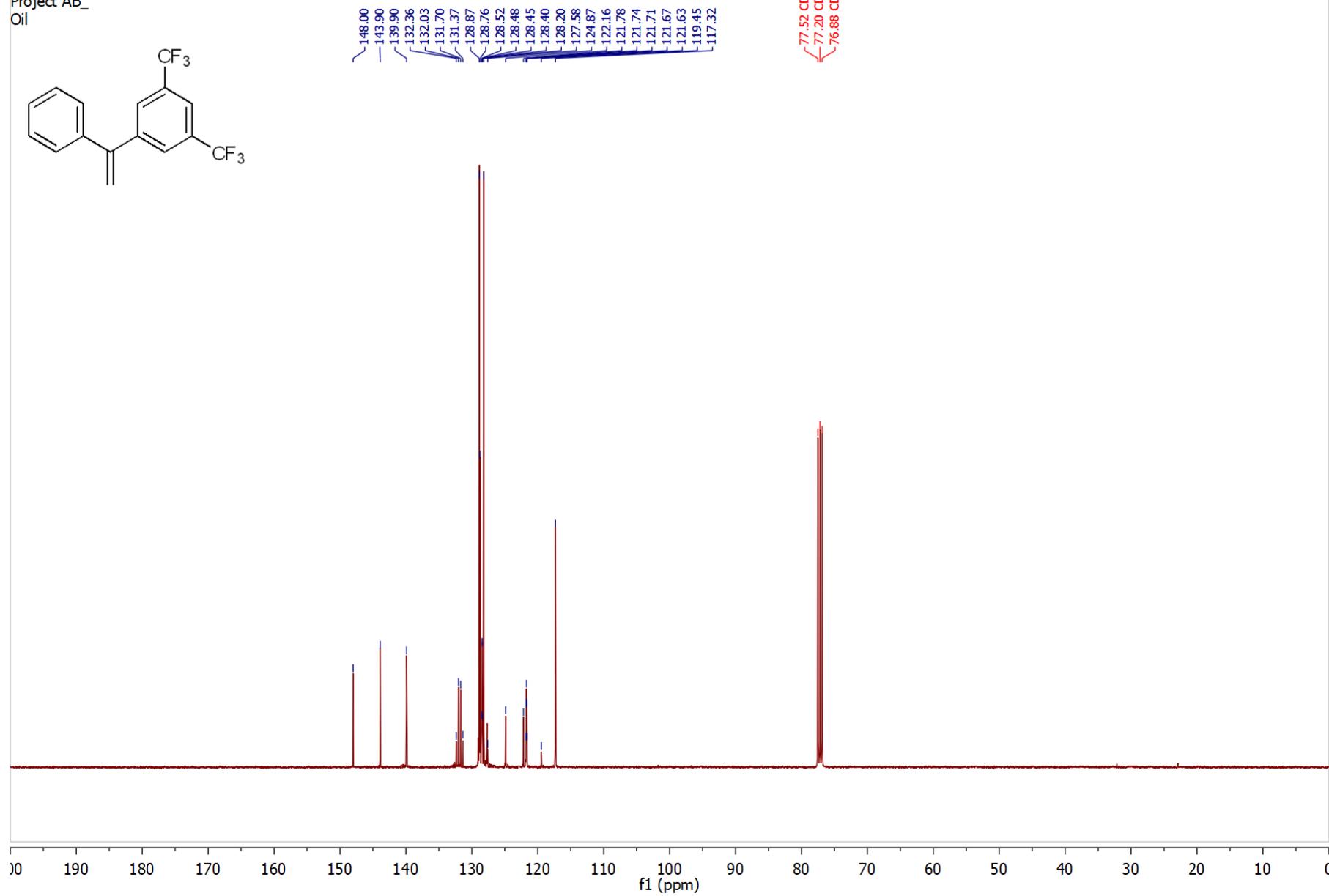
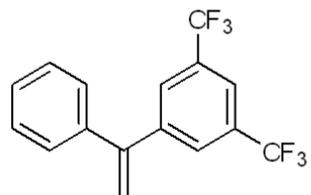
AG1199.2.fid
Project AB_
Oil

-62.86
-62.87
-62.88



Compound 5f

AG1199.6.fid
Project AB_
Oil

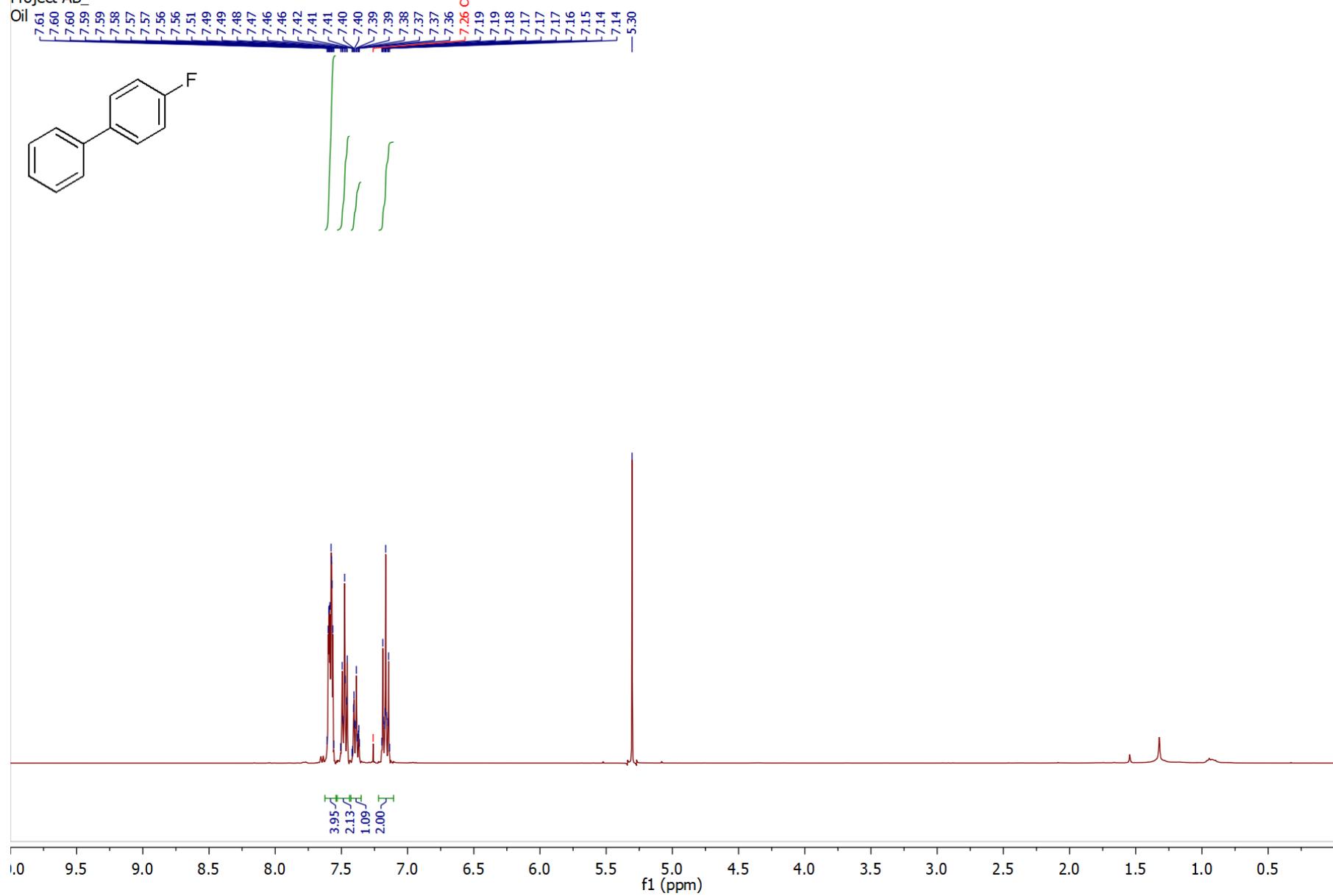


Compound 5h

AG1201.5.fid

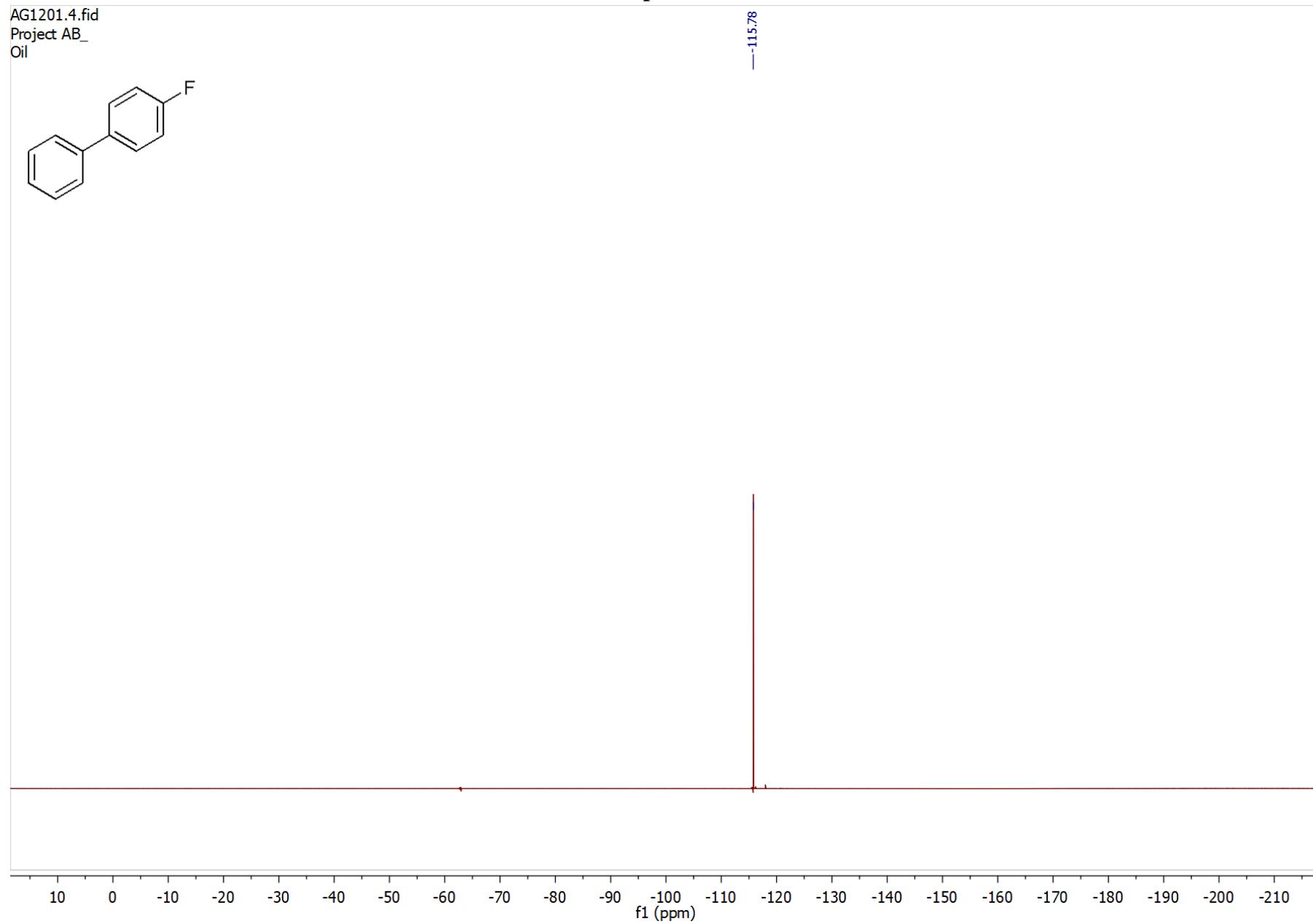
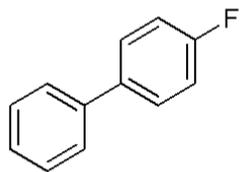
Project AB

Oil



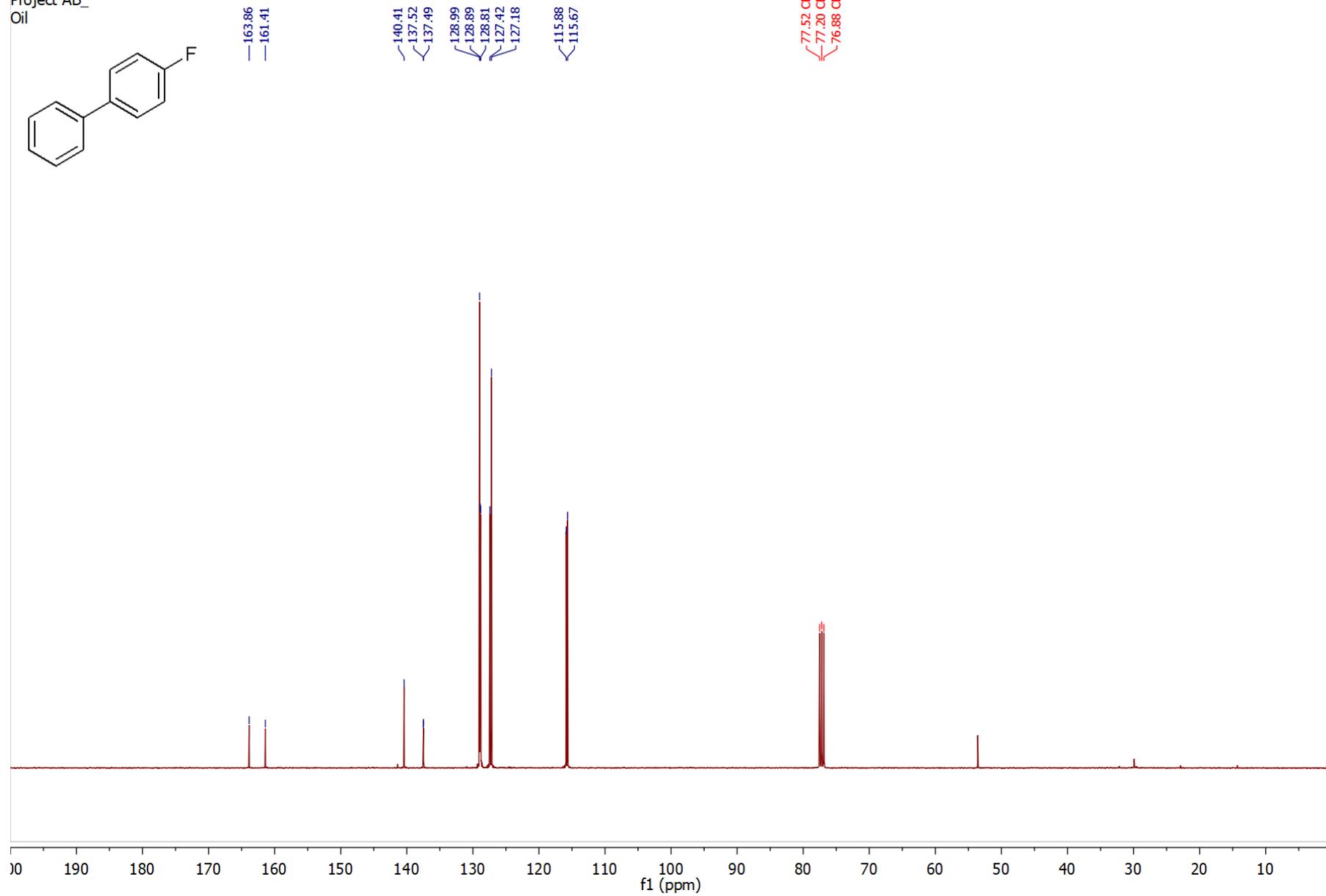
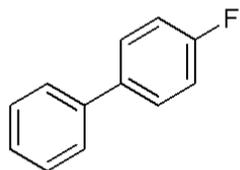
Compound 5h

AG1201.4.fid
Project AB_
Oil



Compound 5h

AG1201.7.fid
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Oil

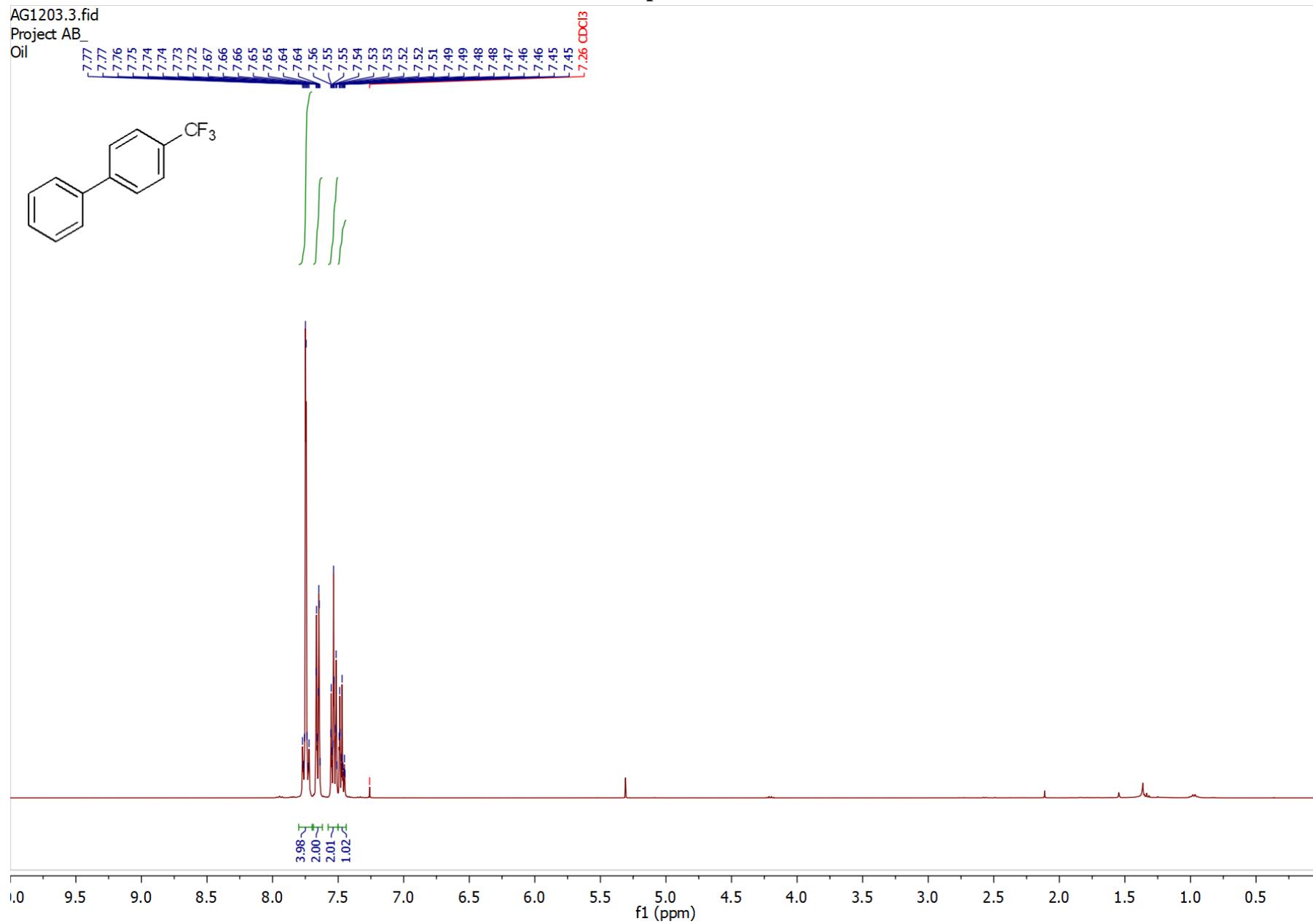


Compound 5i

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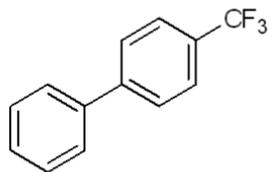
Project AB_

Oil

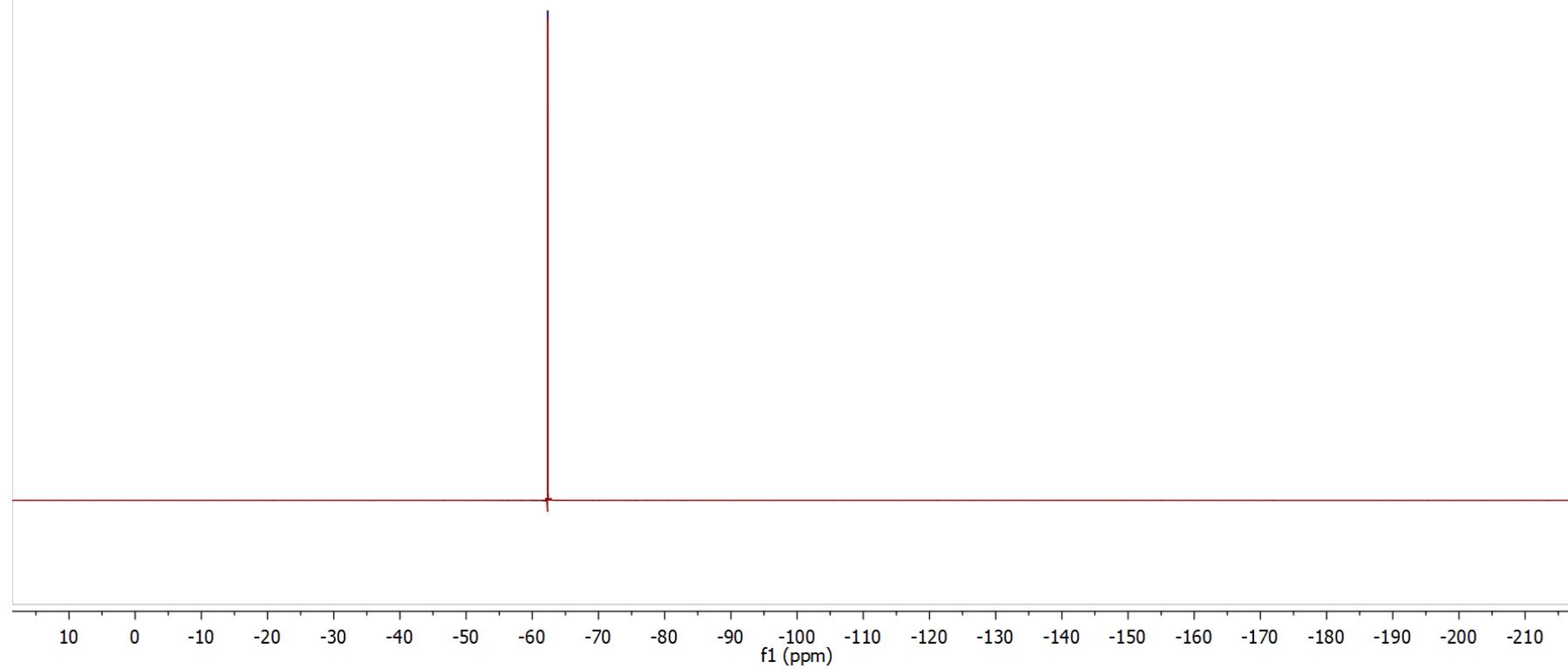


Compound 5i

AG1203.2.fid
Project AB_
Oil

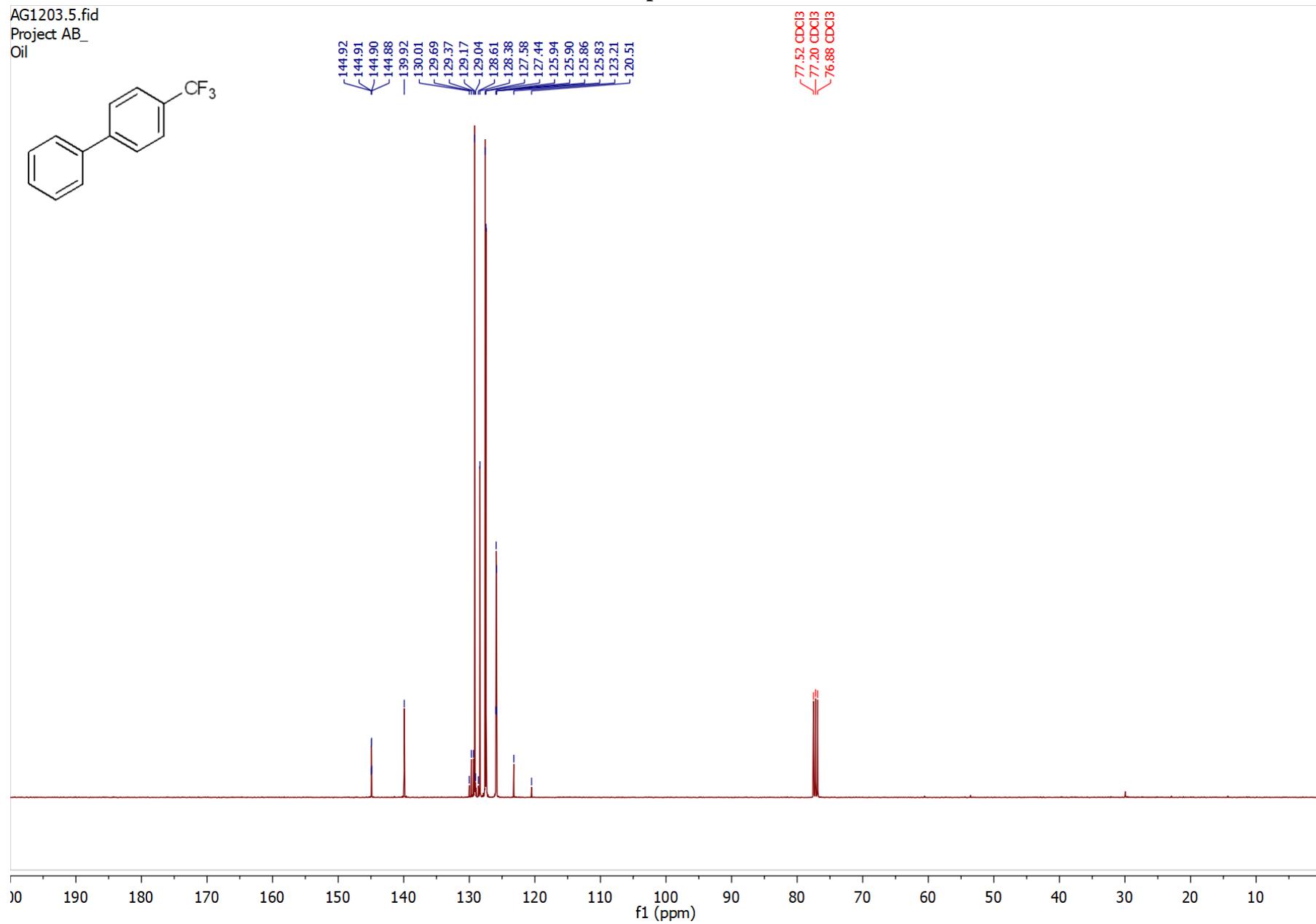
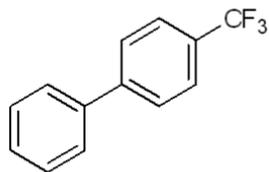


—62.31



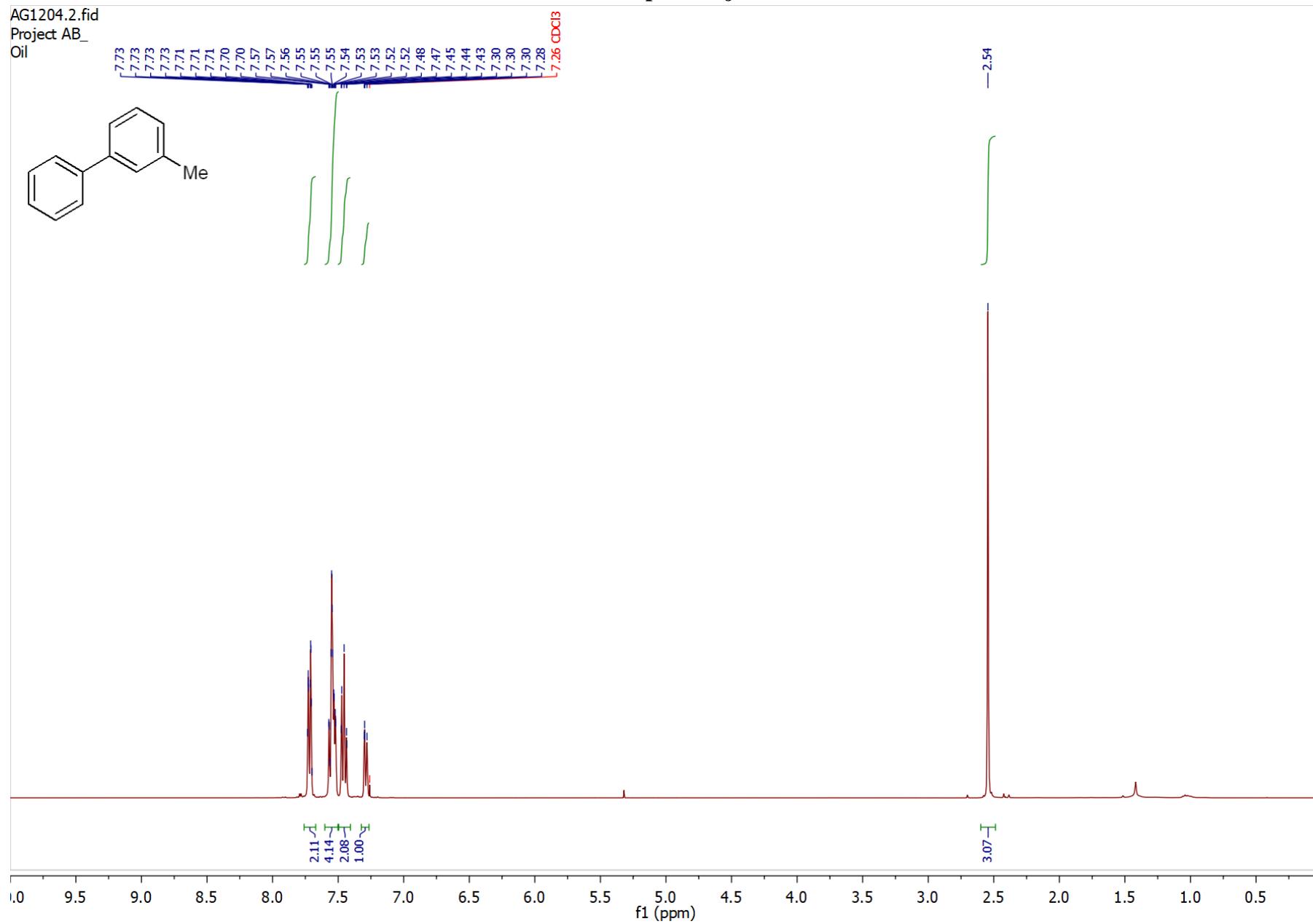
Compound 5i

AG1203.5.fid
Project AB_
Oil



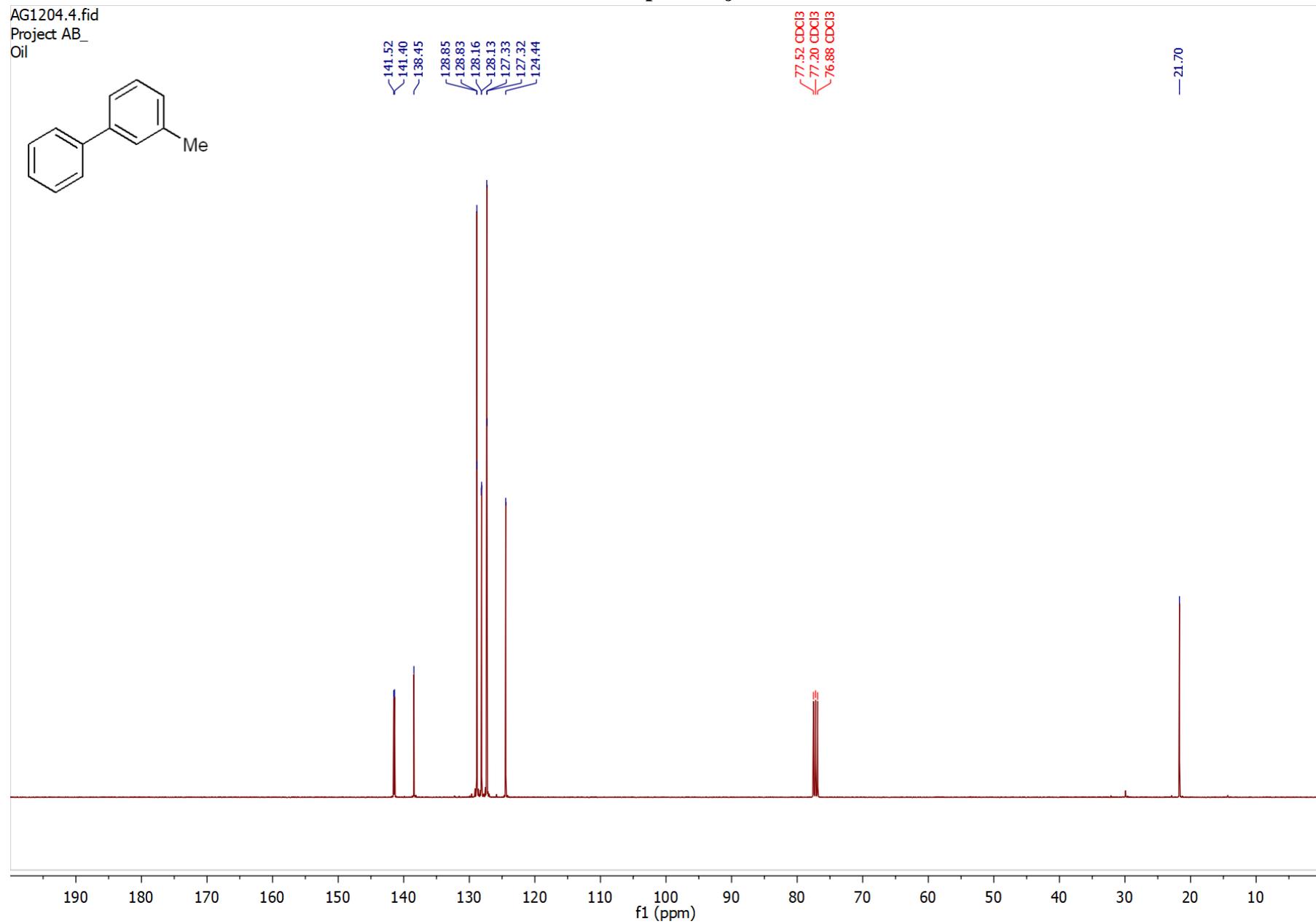
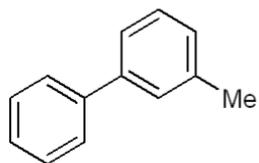
Compound 5j

AG1204.2.fid
Project AB_
Oil



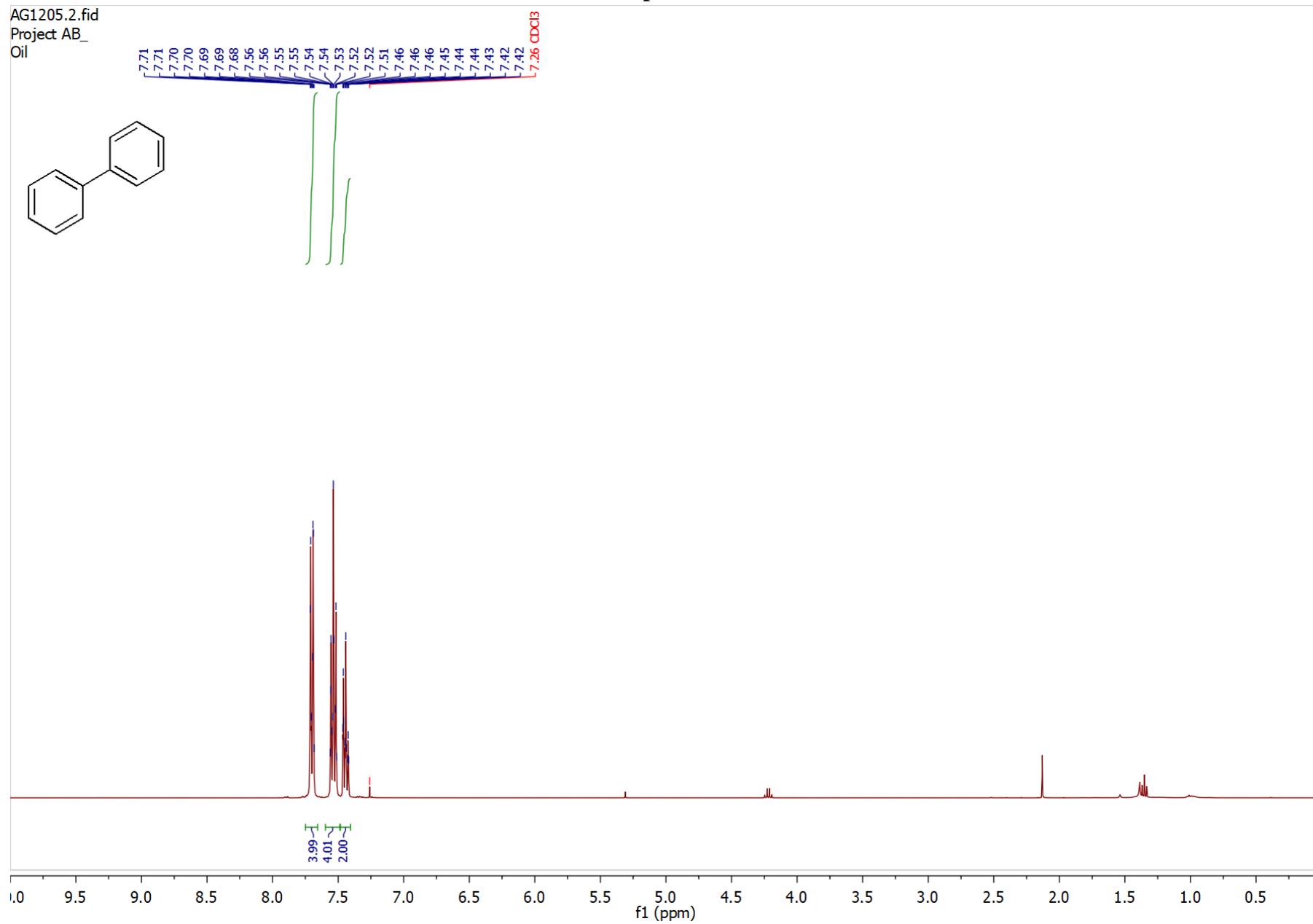
Compound 5j

AG1204.4.fid
Project AB_
Oil



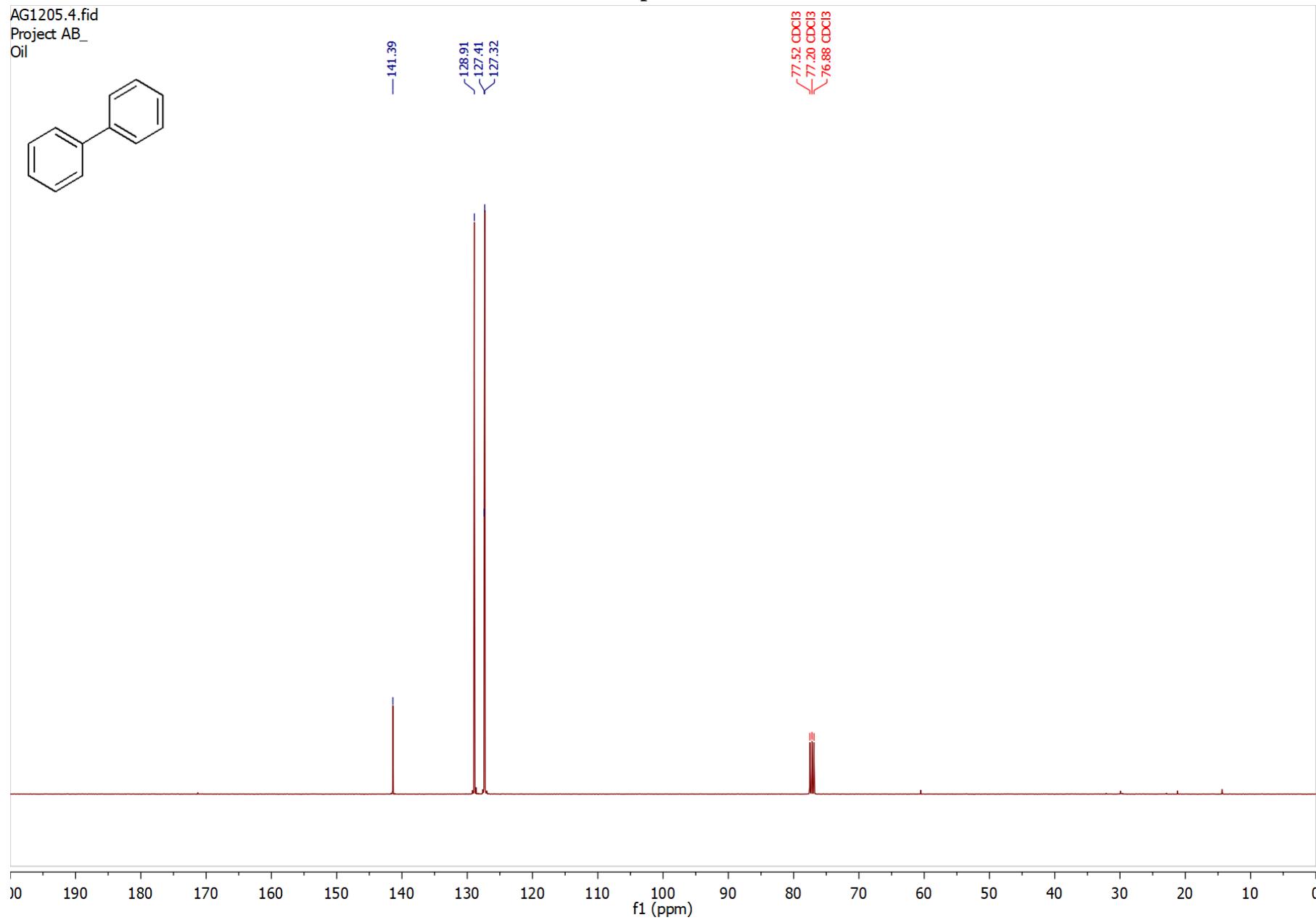
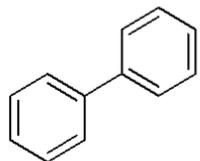
Compound 5k

AG1205.2.fid
Project AB_
Oil

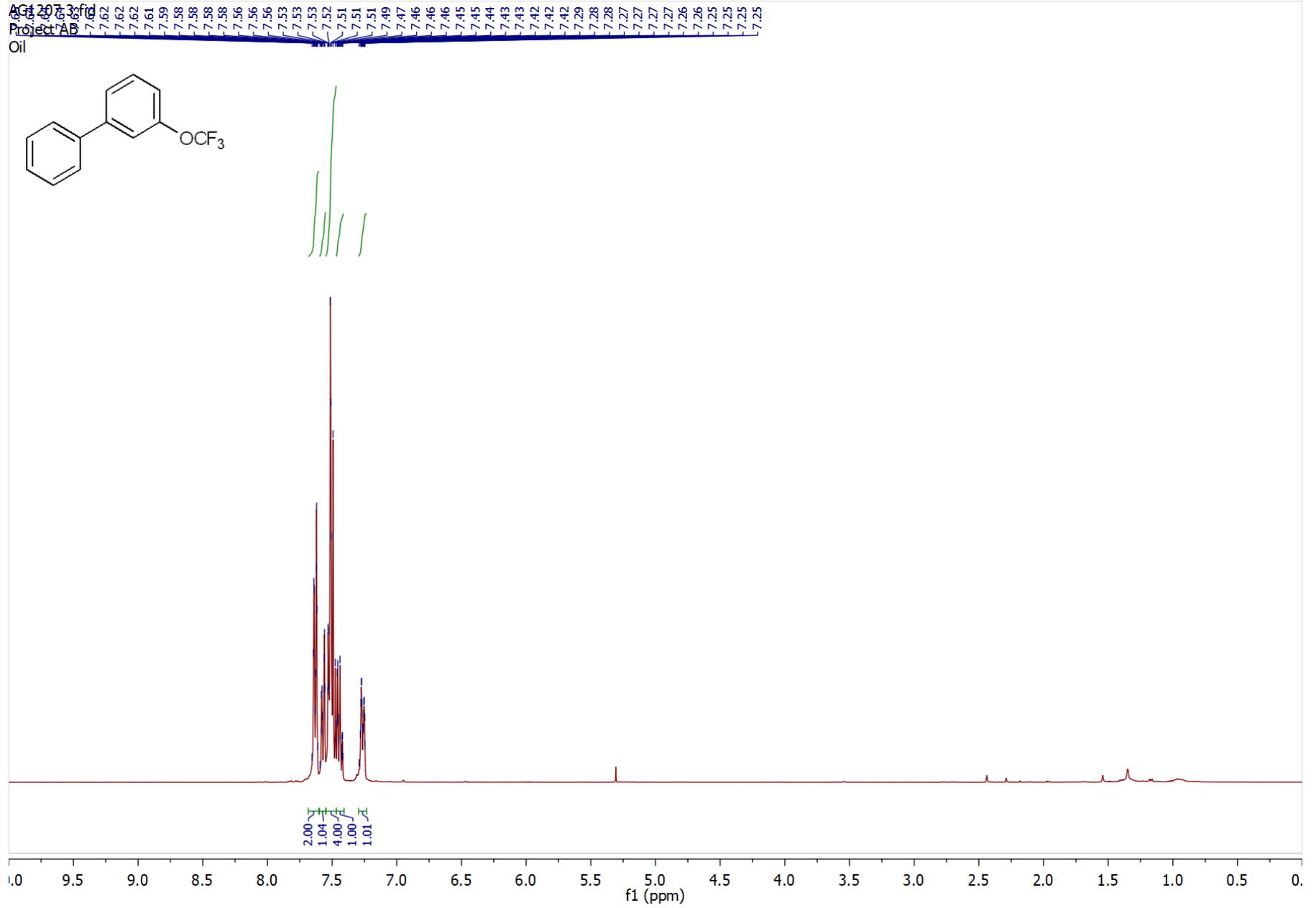


Compound 5k

AG1205.4.fid
Project AB_
Oil

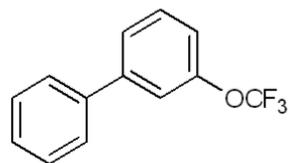


Compound 51

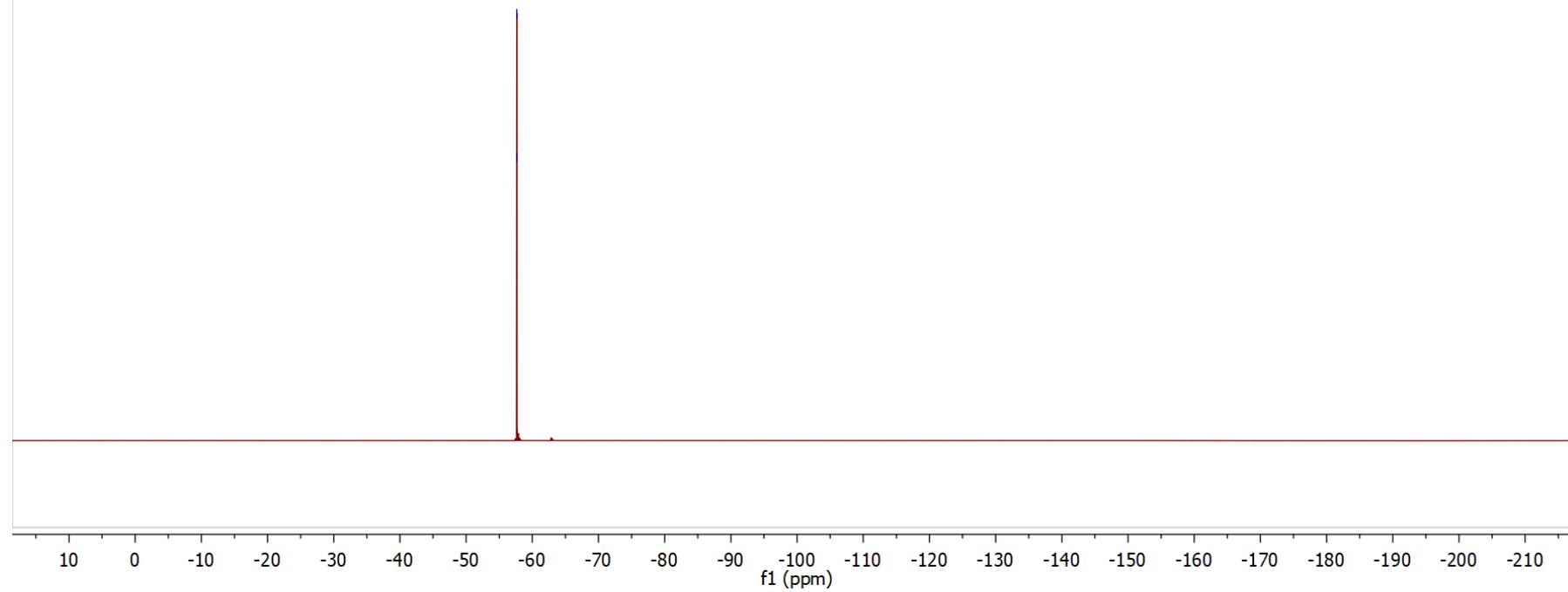


Compound 51

AG1207.2.fid
Project AB_
Oil

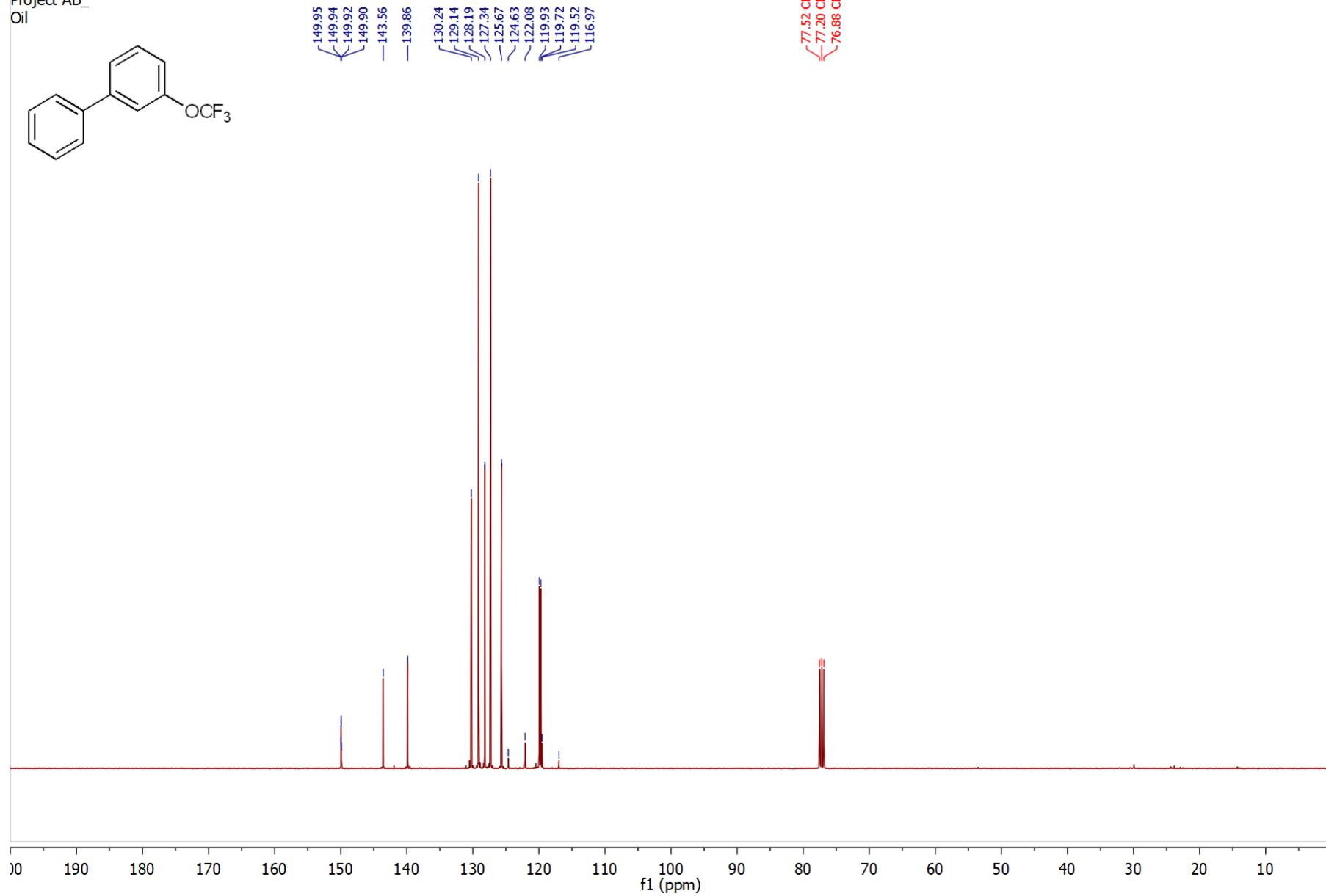
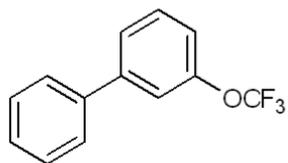


57.65
57.66



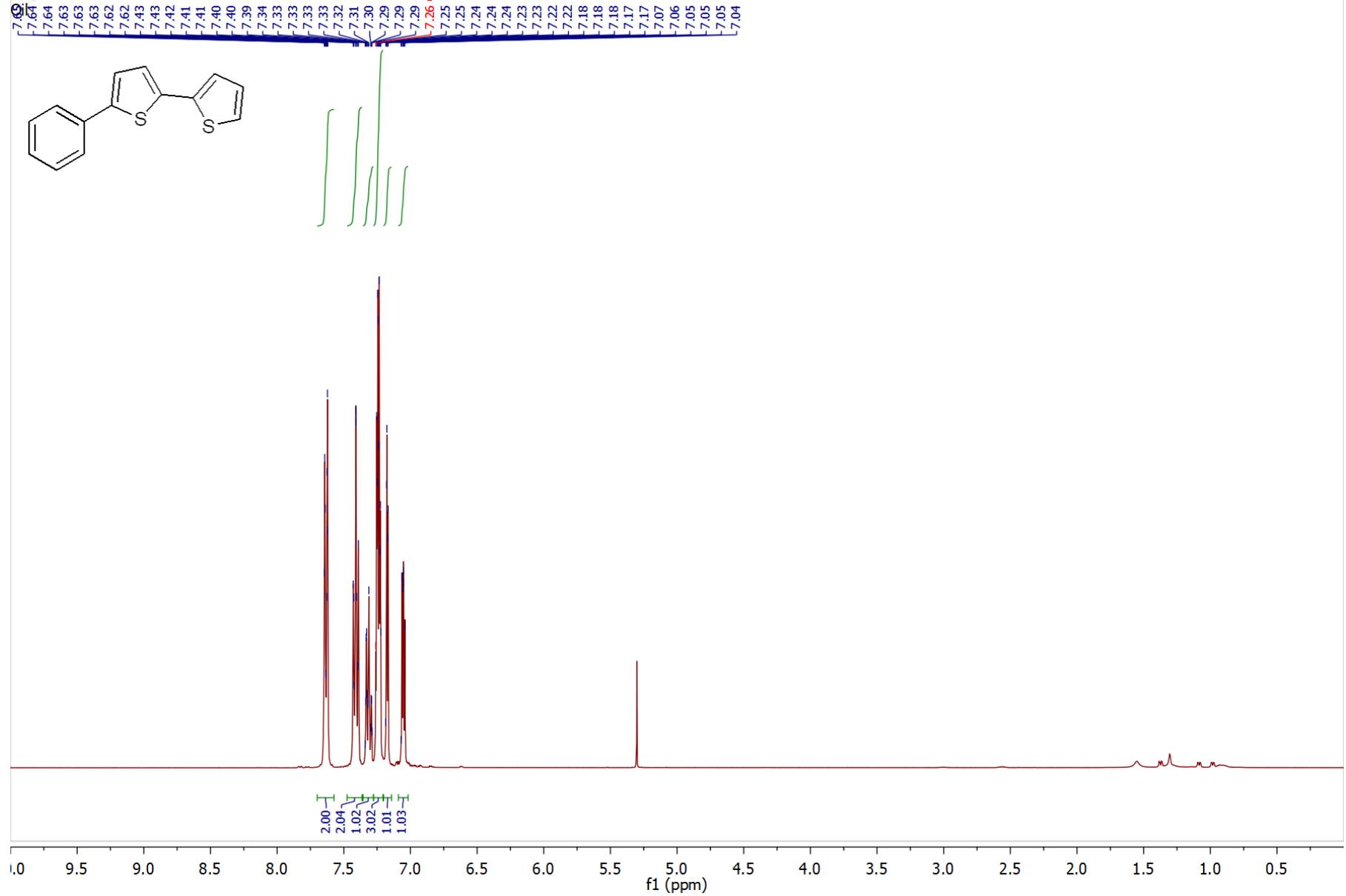
Compound 51

AG1207.5.fid
Project AB_
Oil



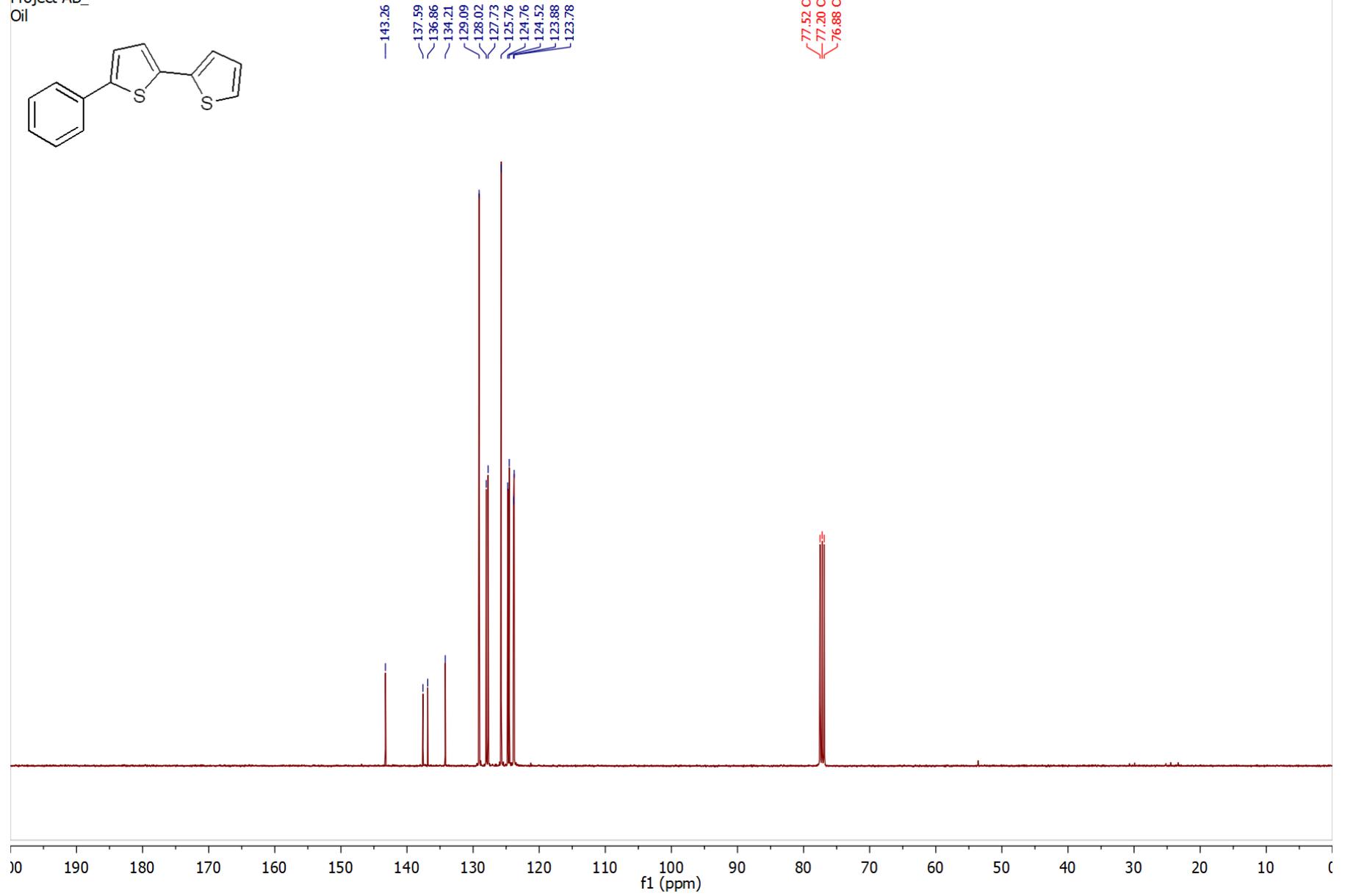
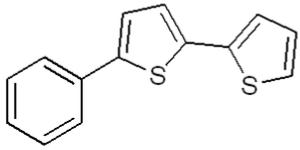
Compound 5m

AG1208.2.fid
Project AB



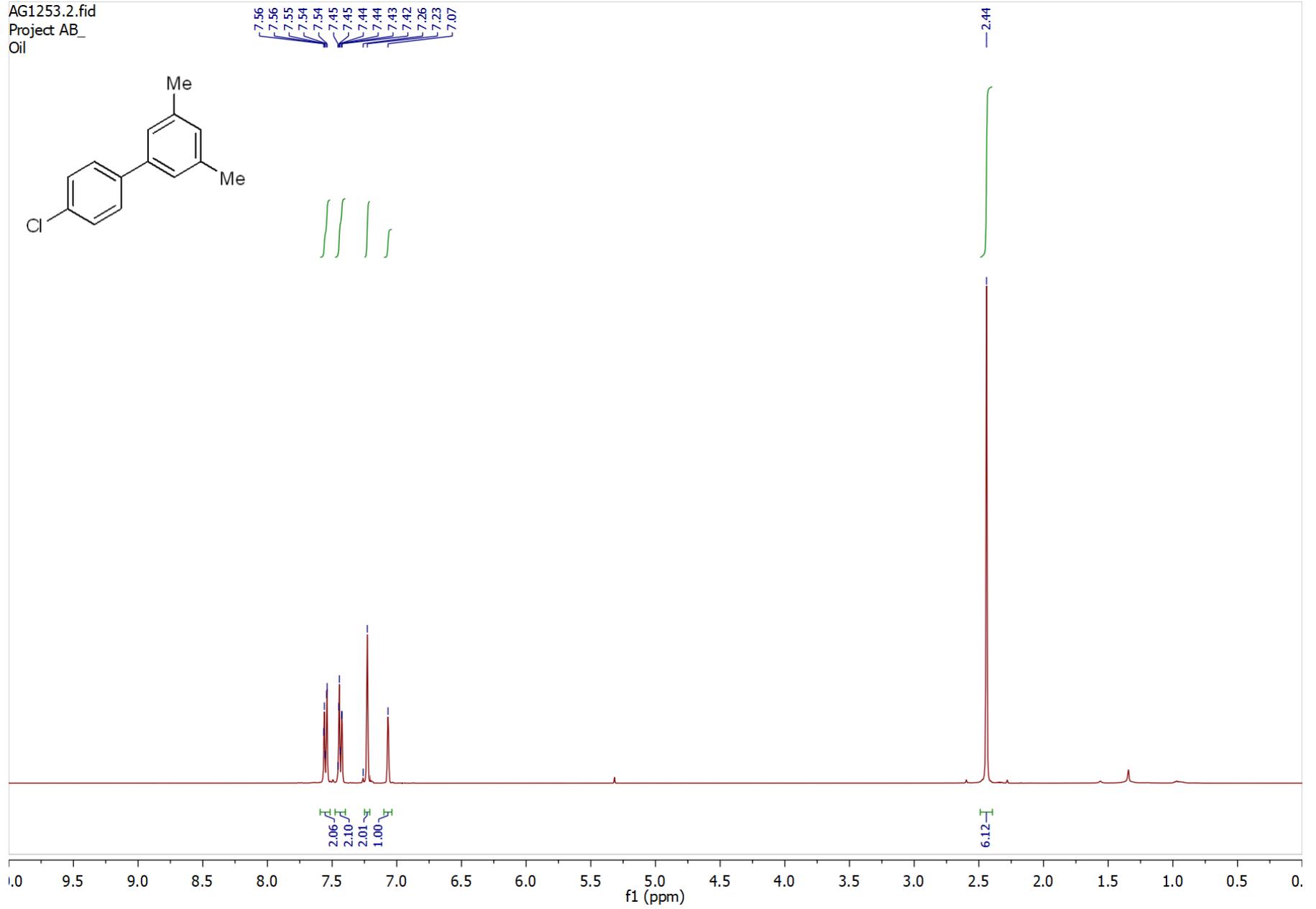
Compound 5m

AG1208.4.fid
Project AB_
Oil



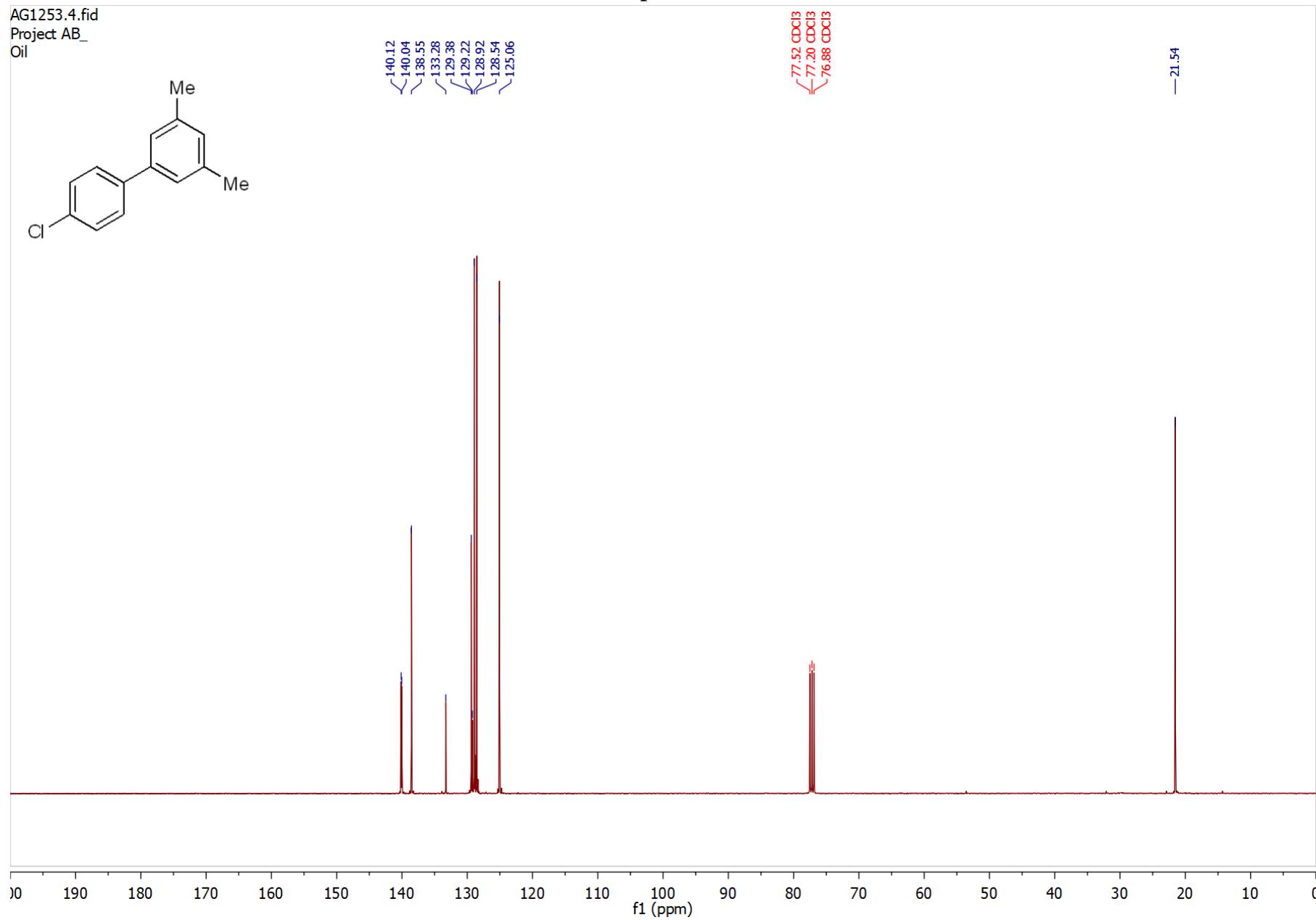
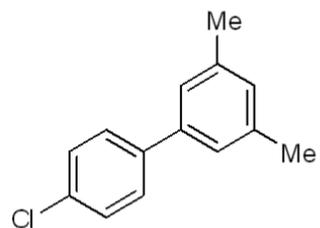
Compound 5n

AG1253.2.fid
Project AB_
Oil



Compound 5n

AG1253.4.fid
Project AB_
Oil

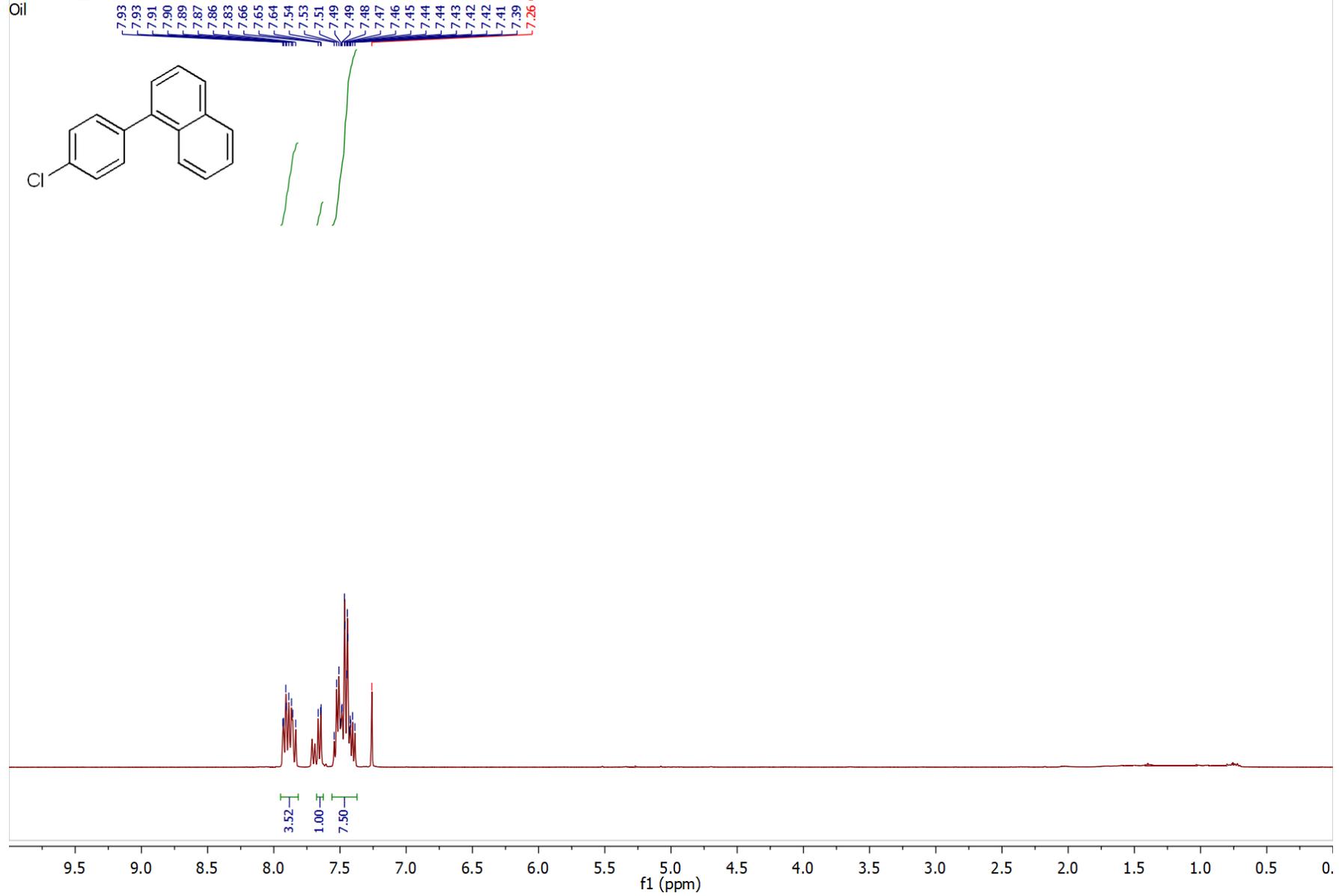


Compound 5o

AG1254-2.2.fid

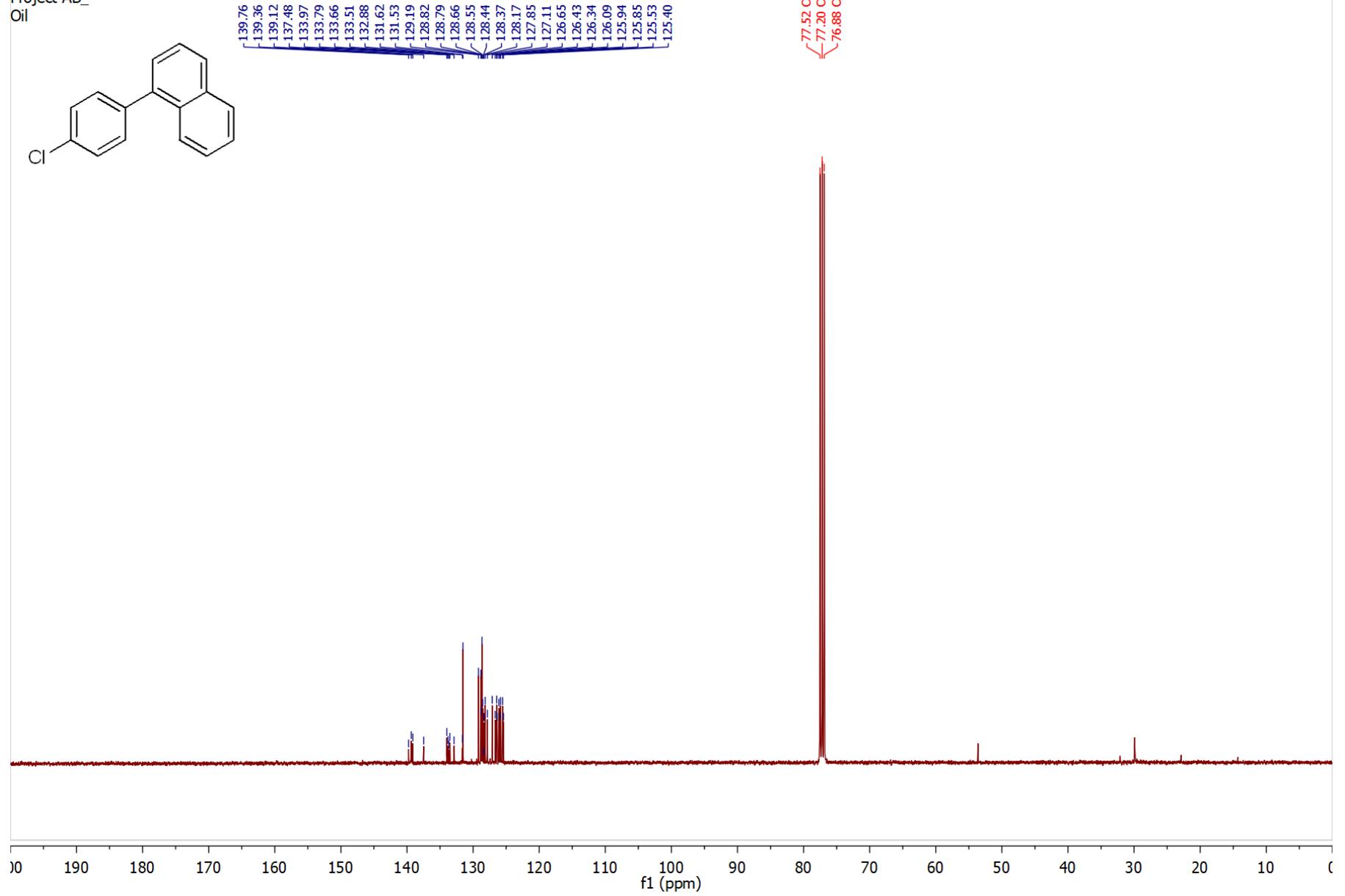
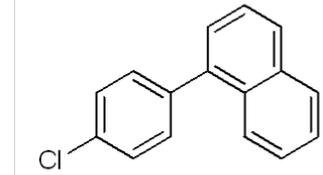
Project AB_

Oil



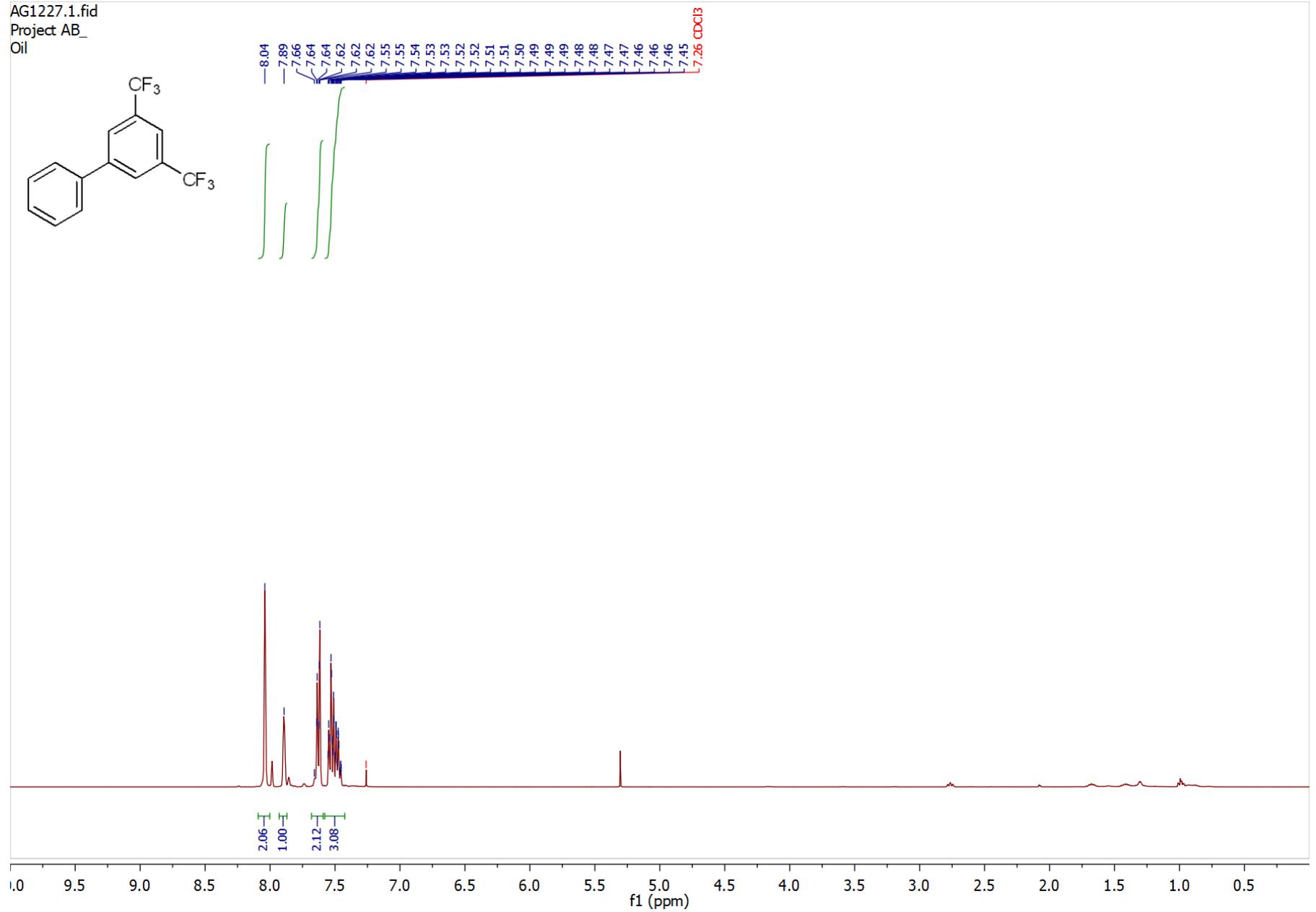
Compound 5o

AG1254-2.4.fid
Project AB_
Oil



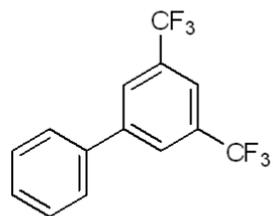
Compound 7a

AG1227.1.fid
Project AB_
Oil

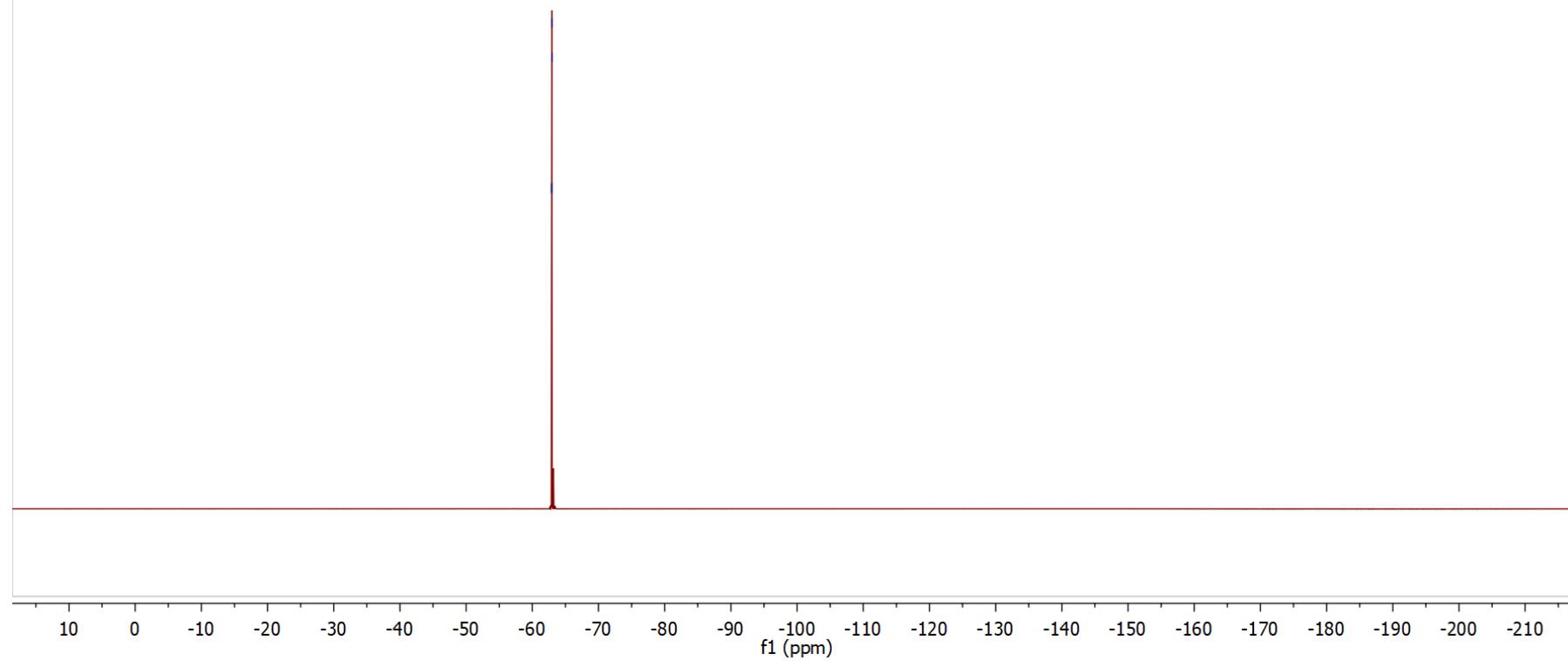


Compound 7a

AG1227.2.fid
Project AB_
Oil

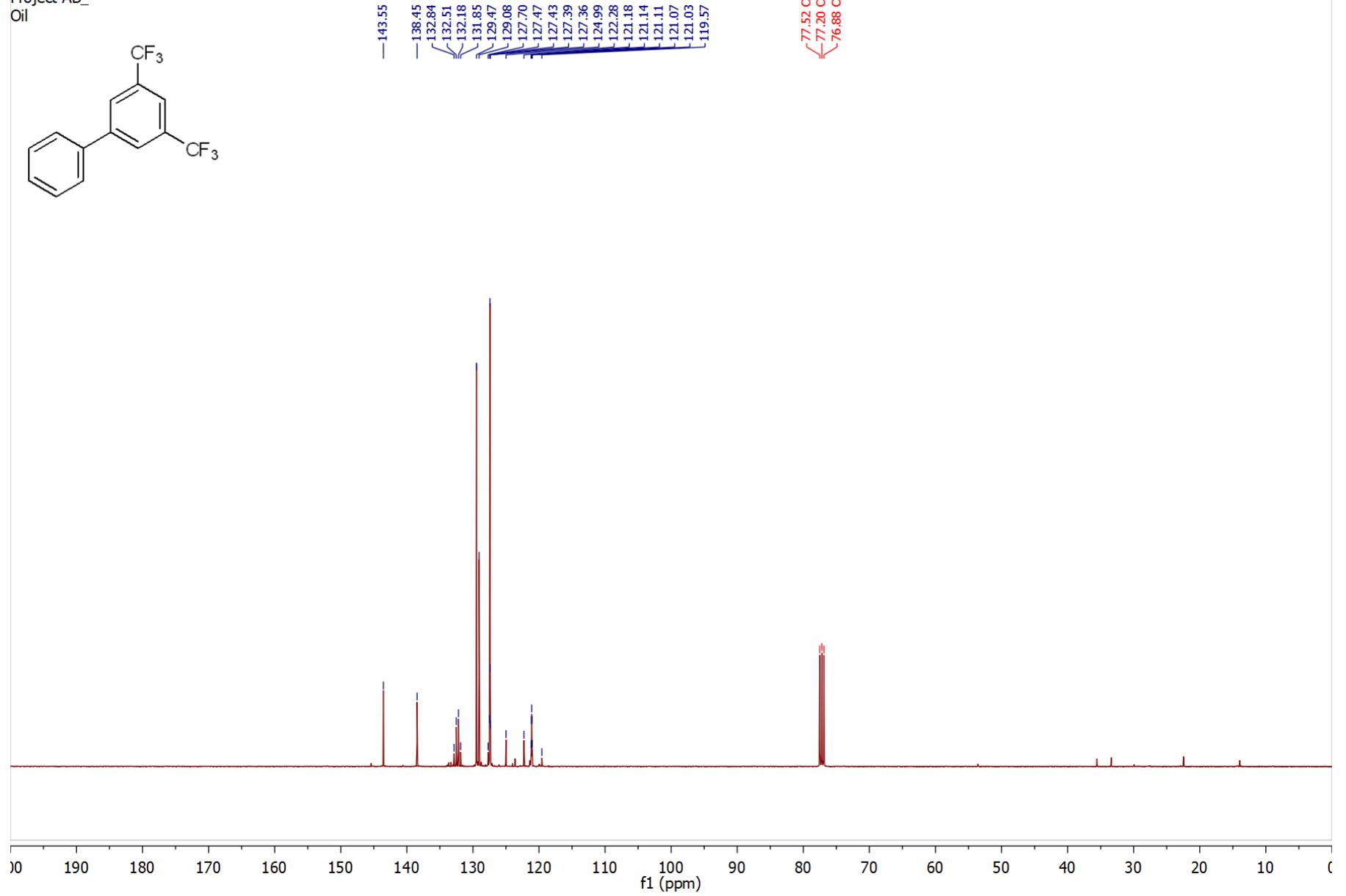
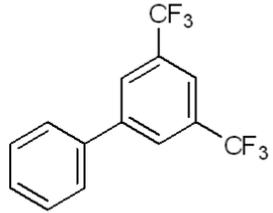


62.92
62.93
62.94
62.95

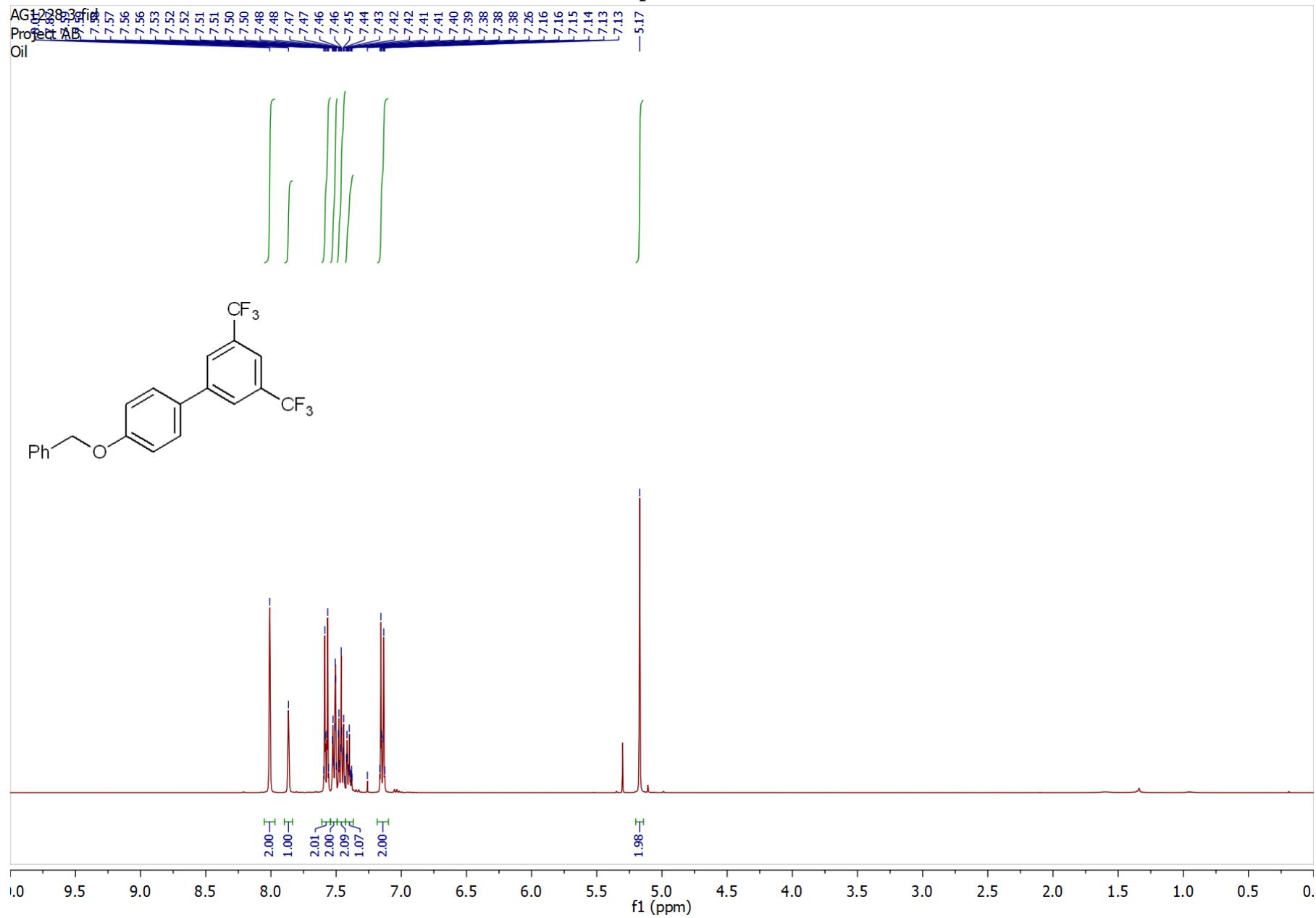


Compound 7a

AG1227.5.fid
Project AB_
Oil

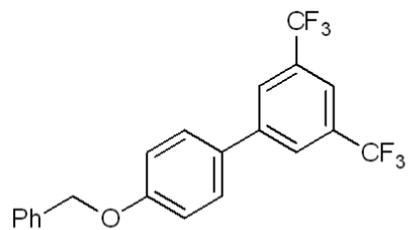


Compound 7b

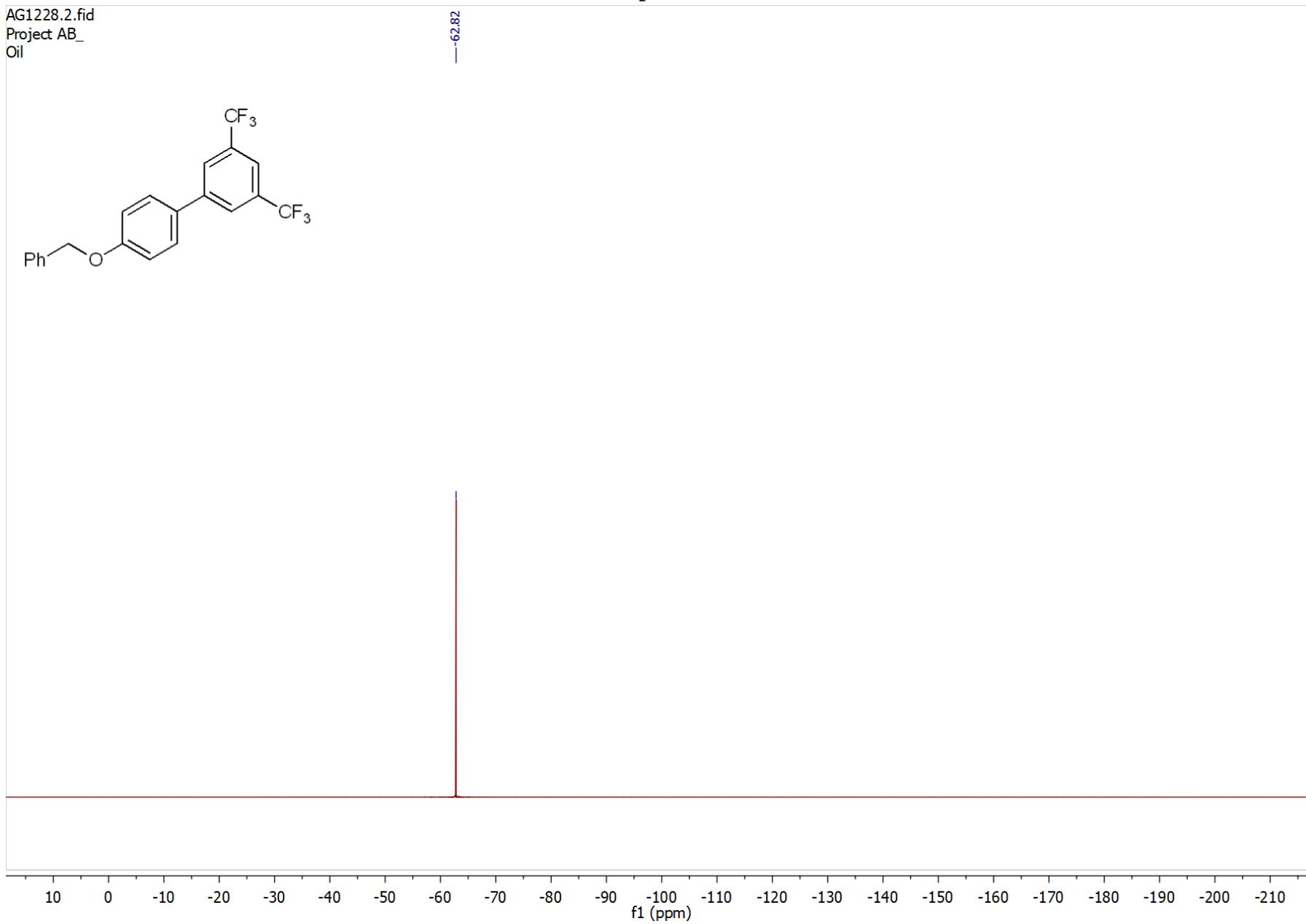


Compound 7b

AG1228.2.fid
Project AB_
Oil

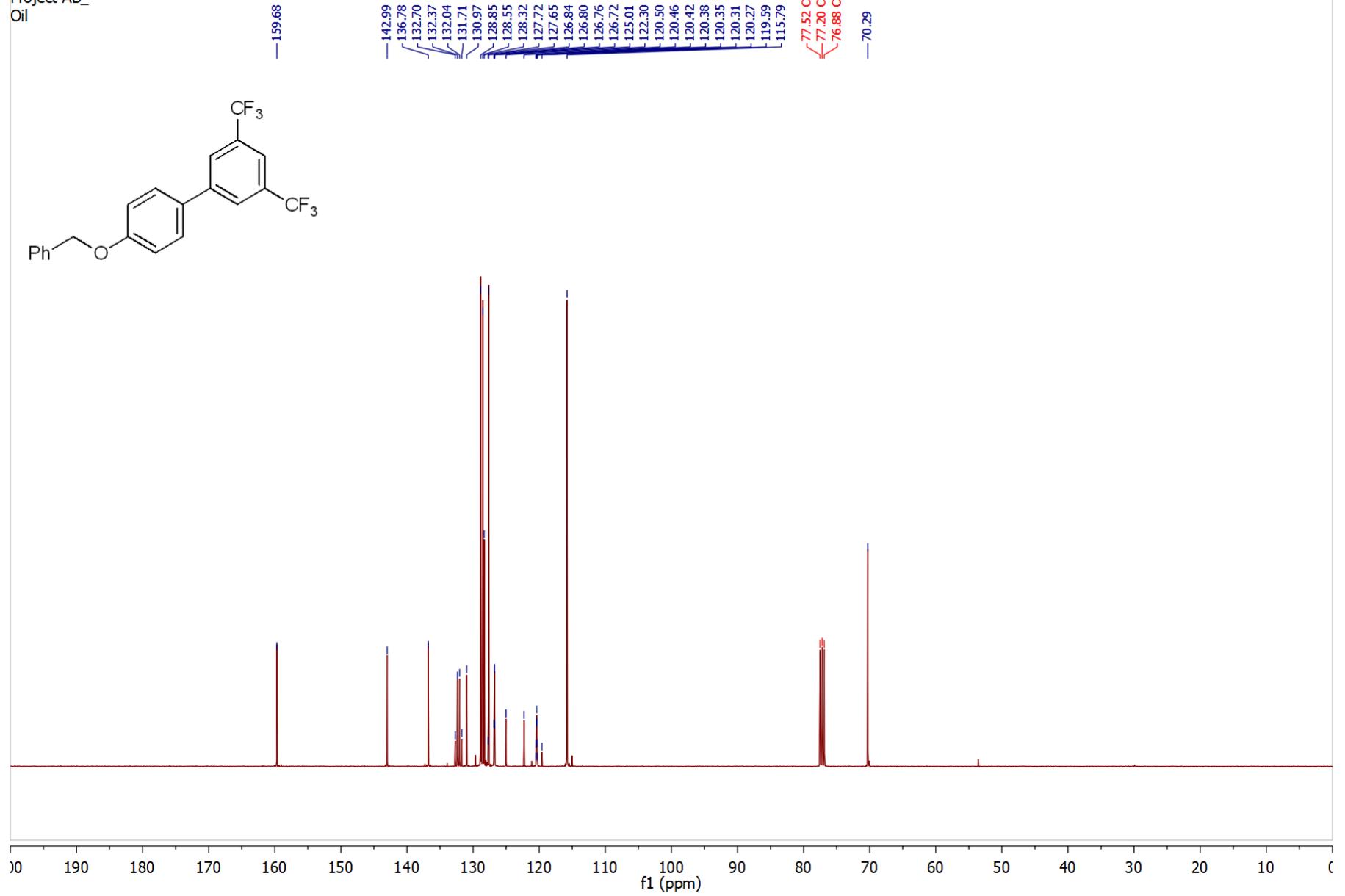


-62.82



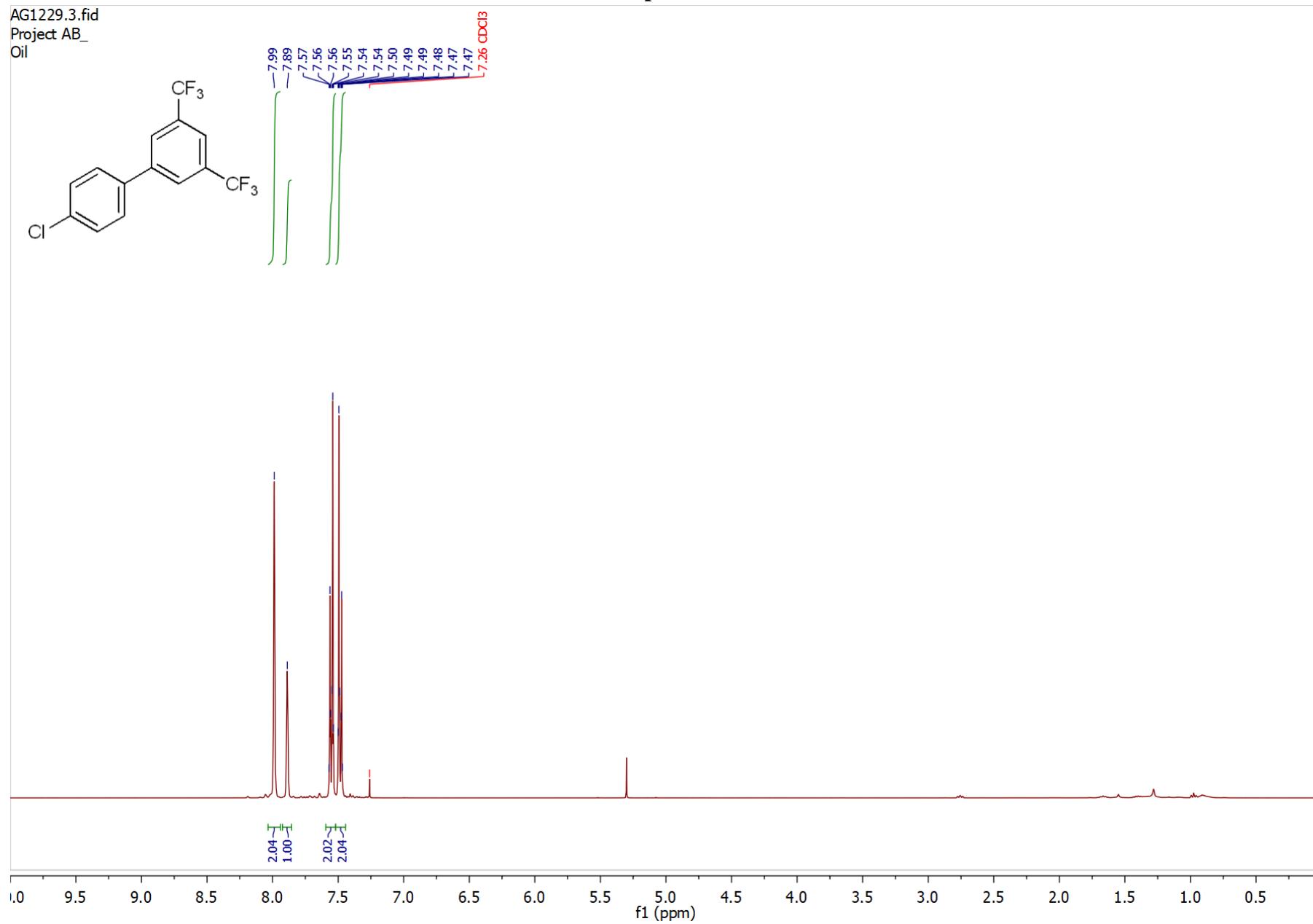
Compound 7b

AG1228.5.fid
Project AB_
Oil



Compound 7c

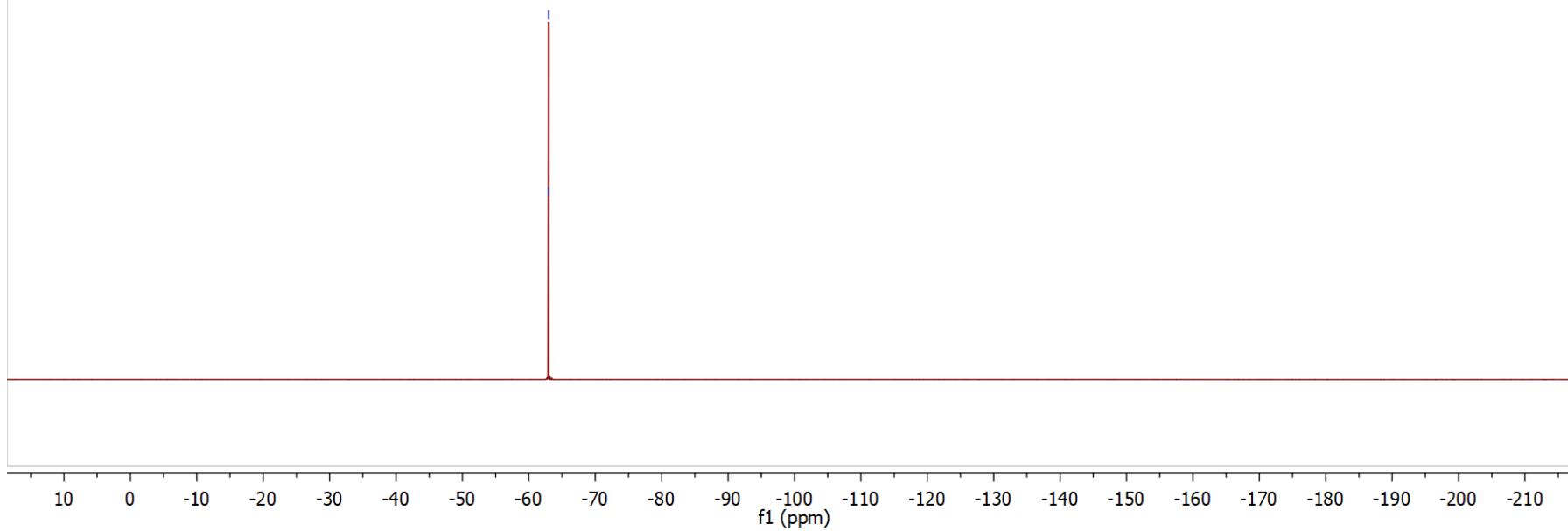
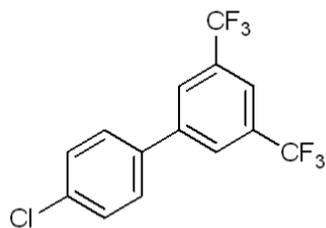
AG1229.3.fid
Project AB_
Oil



Compound 7c

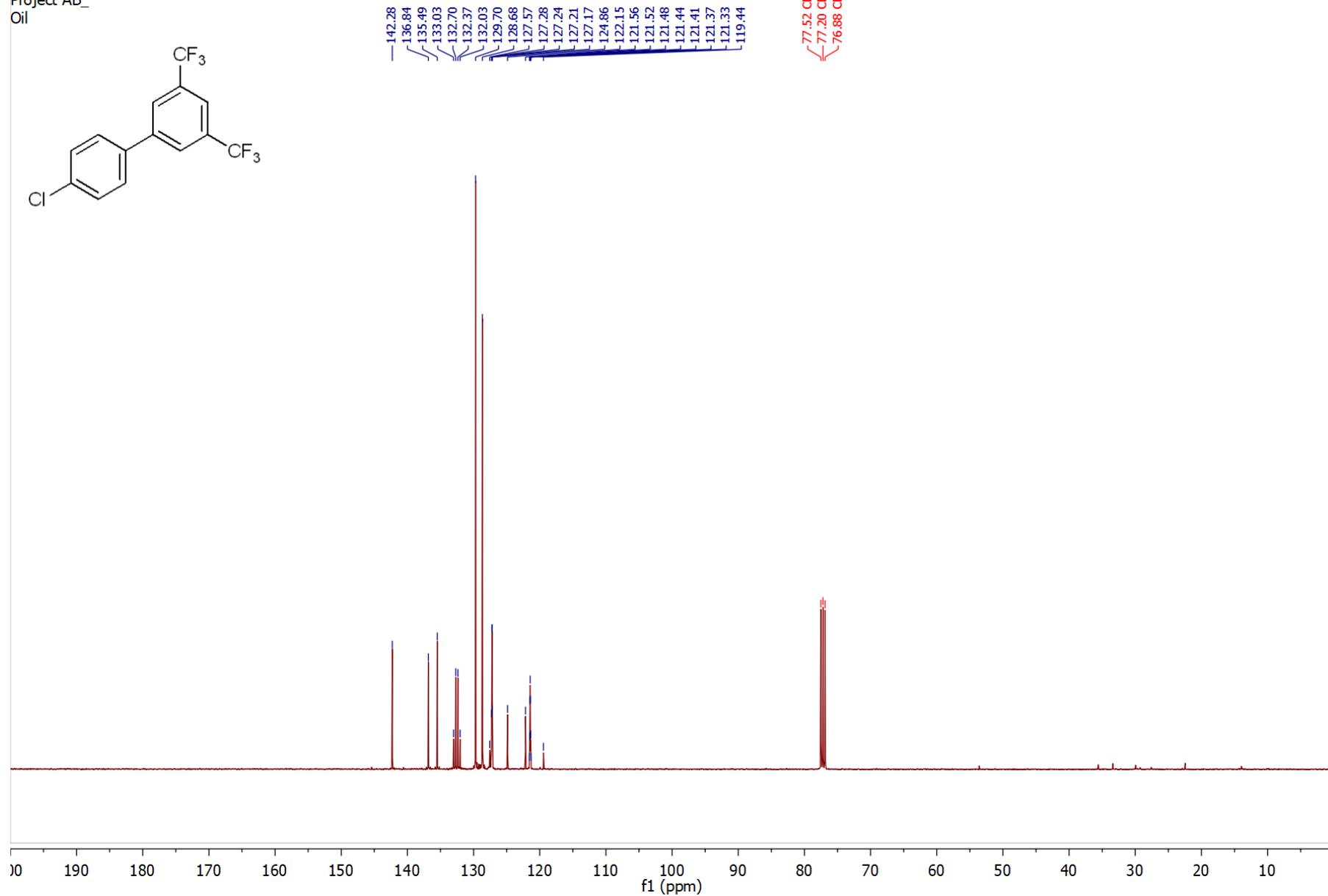
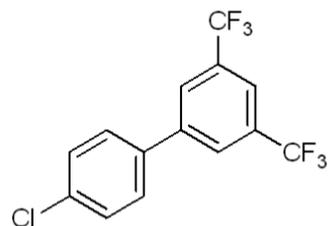
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Project AB_
Oil

-62.97
-62.98



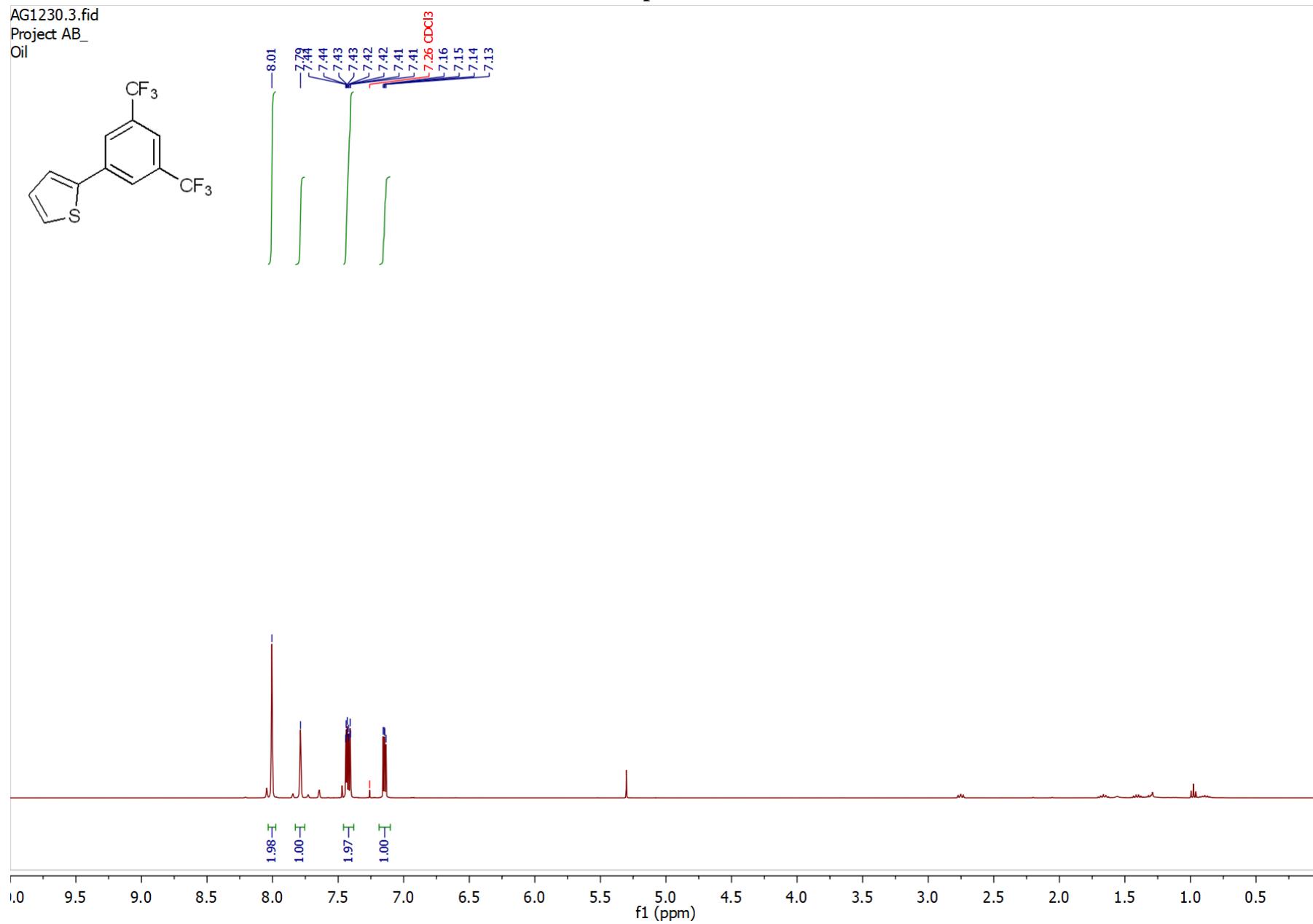
Compound 7c

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Project AB_
Oil



Compound 7d

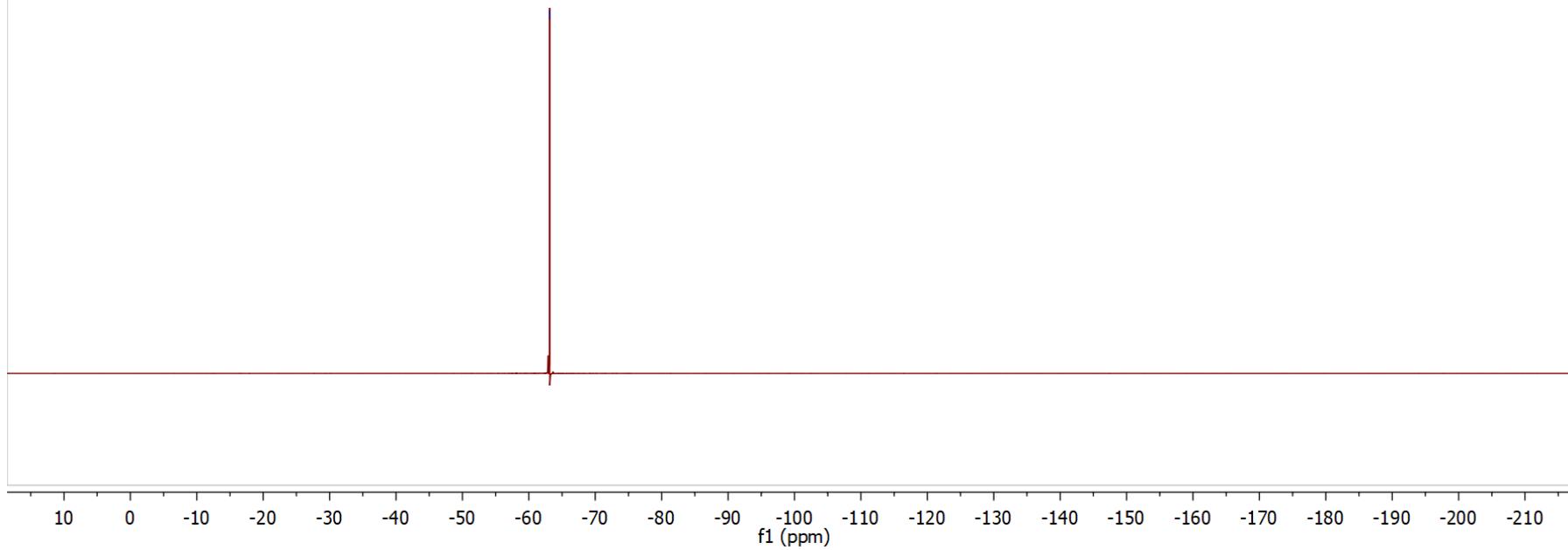
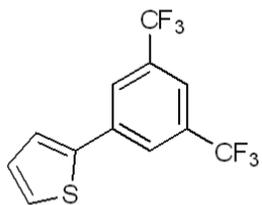
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Project AB_
Oil



Compound 7d

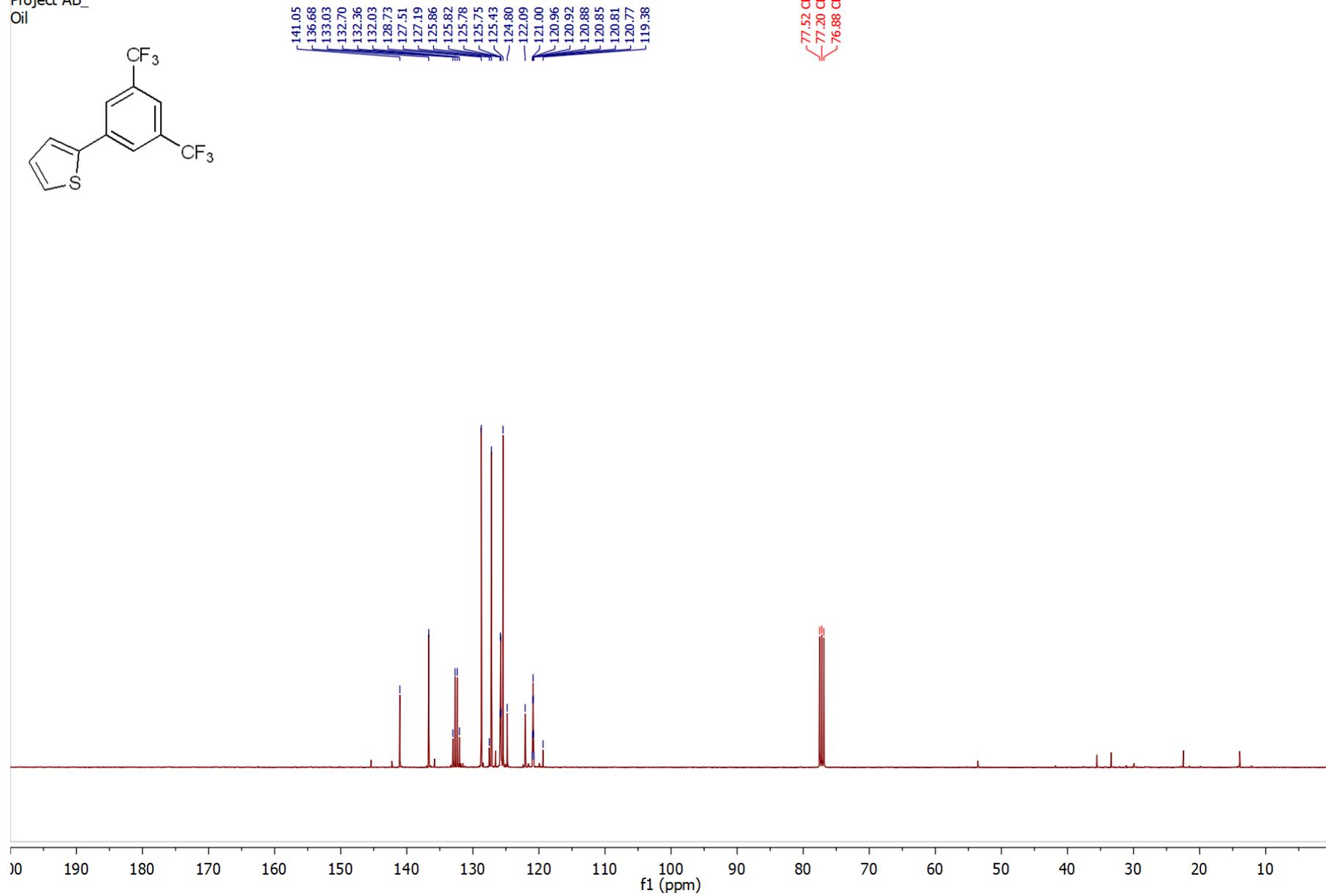
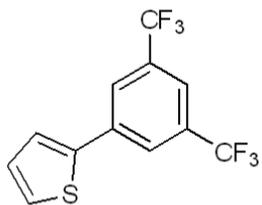
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Project AB_
Oil

—63.14



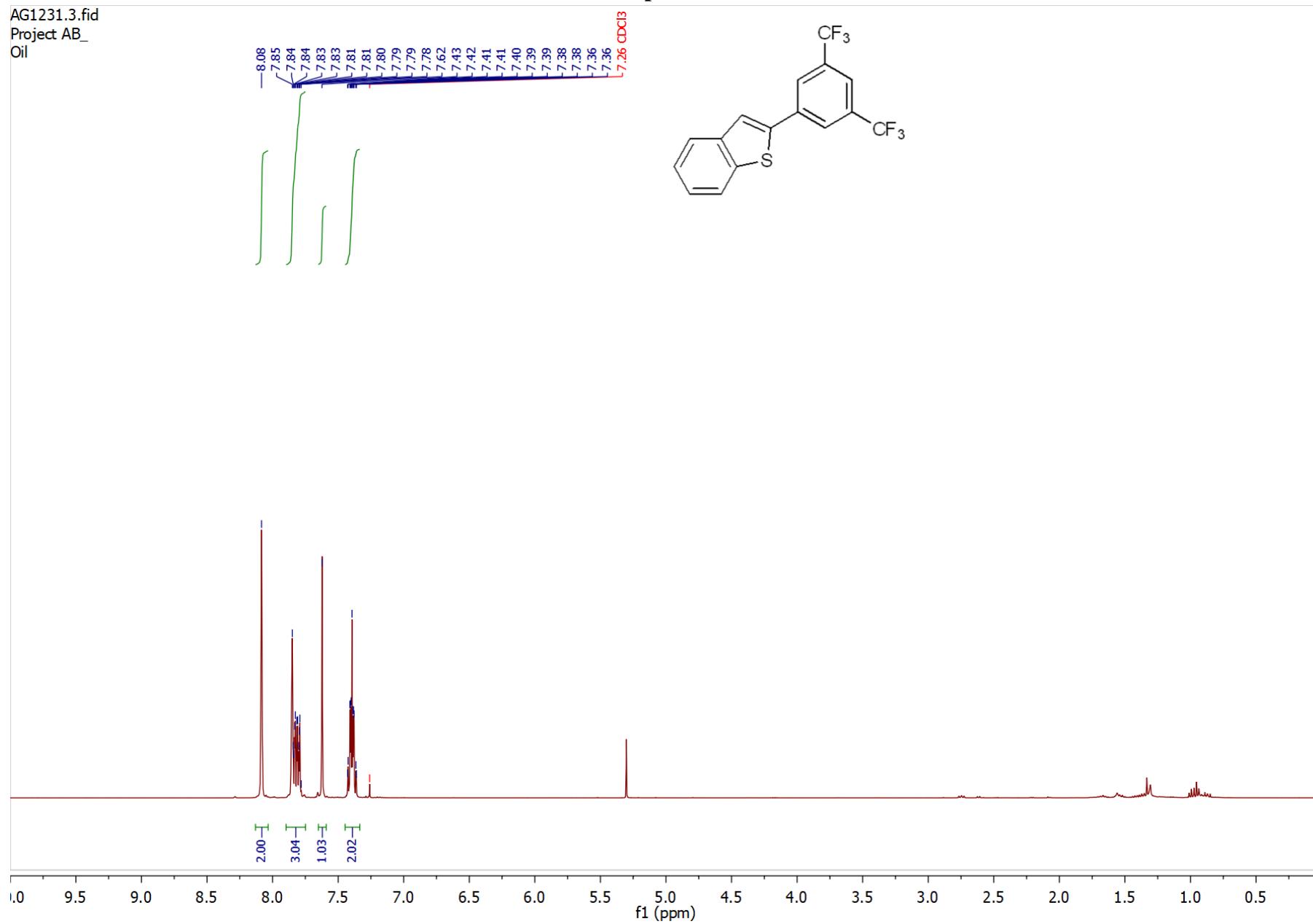
Compound 7d

AG1230.5.fid
Project AB_
Oil



Compound 7e

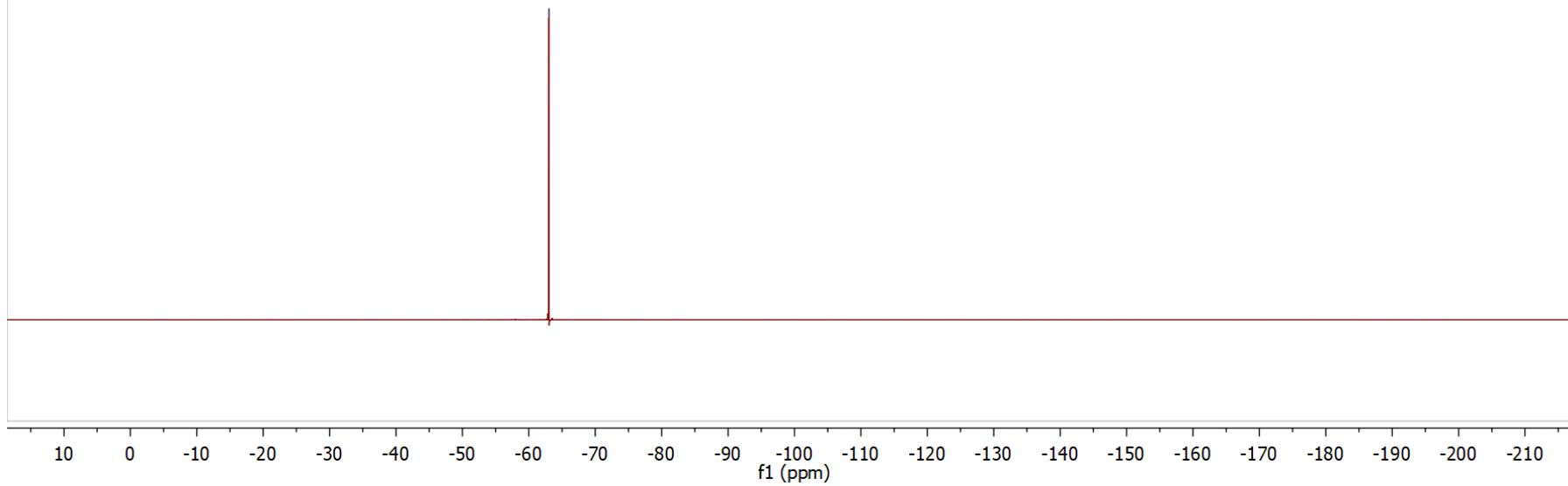
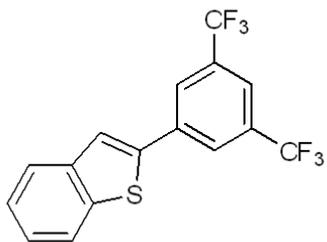
AG1231.3.fid
Project AB_
Oil



Compound 7e

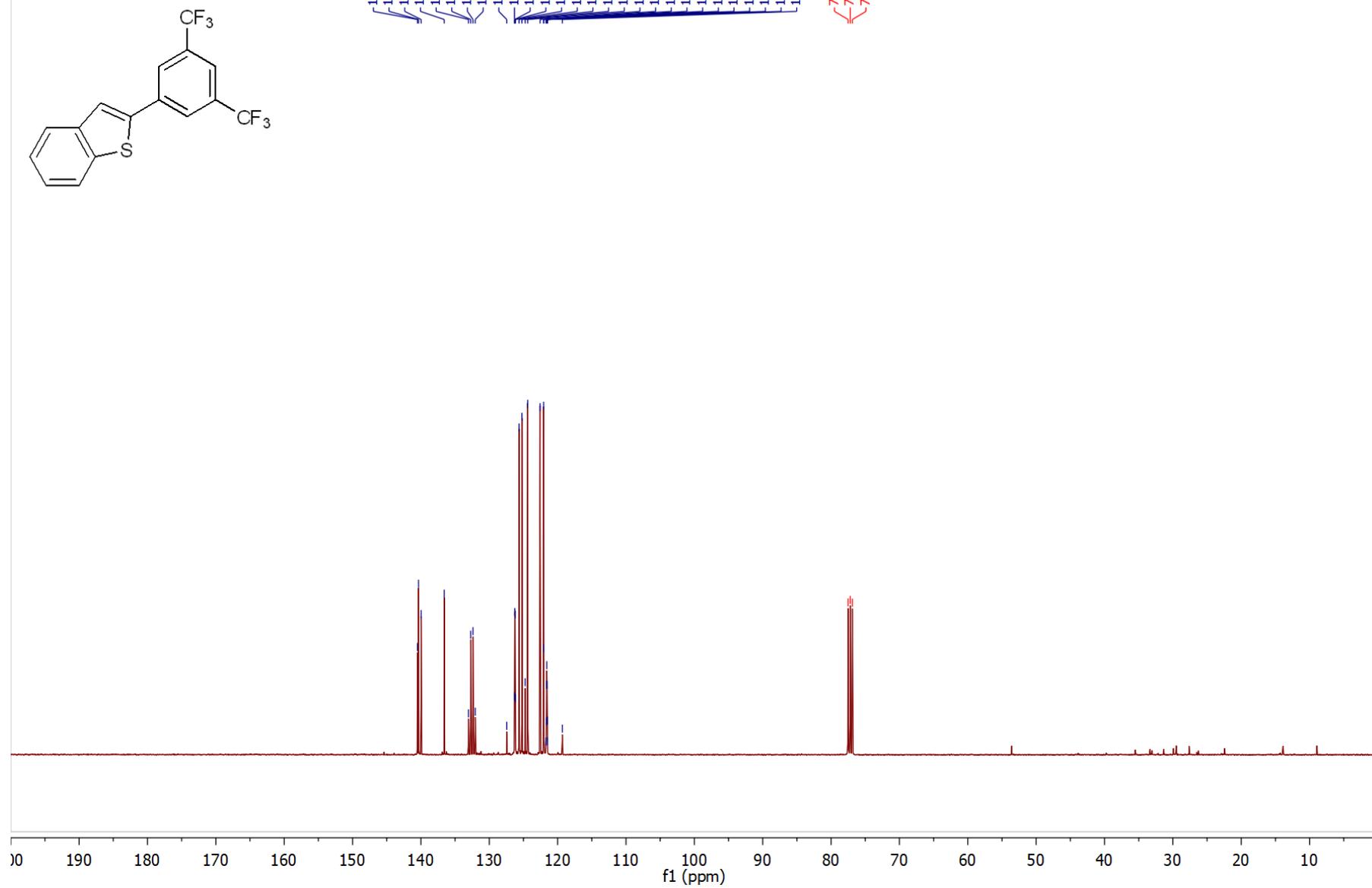
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Project AB_
Oil

63.03

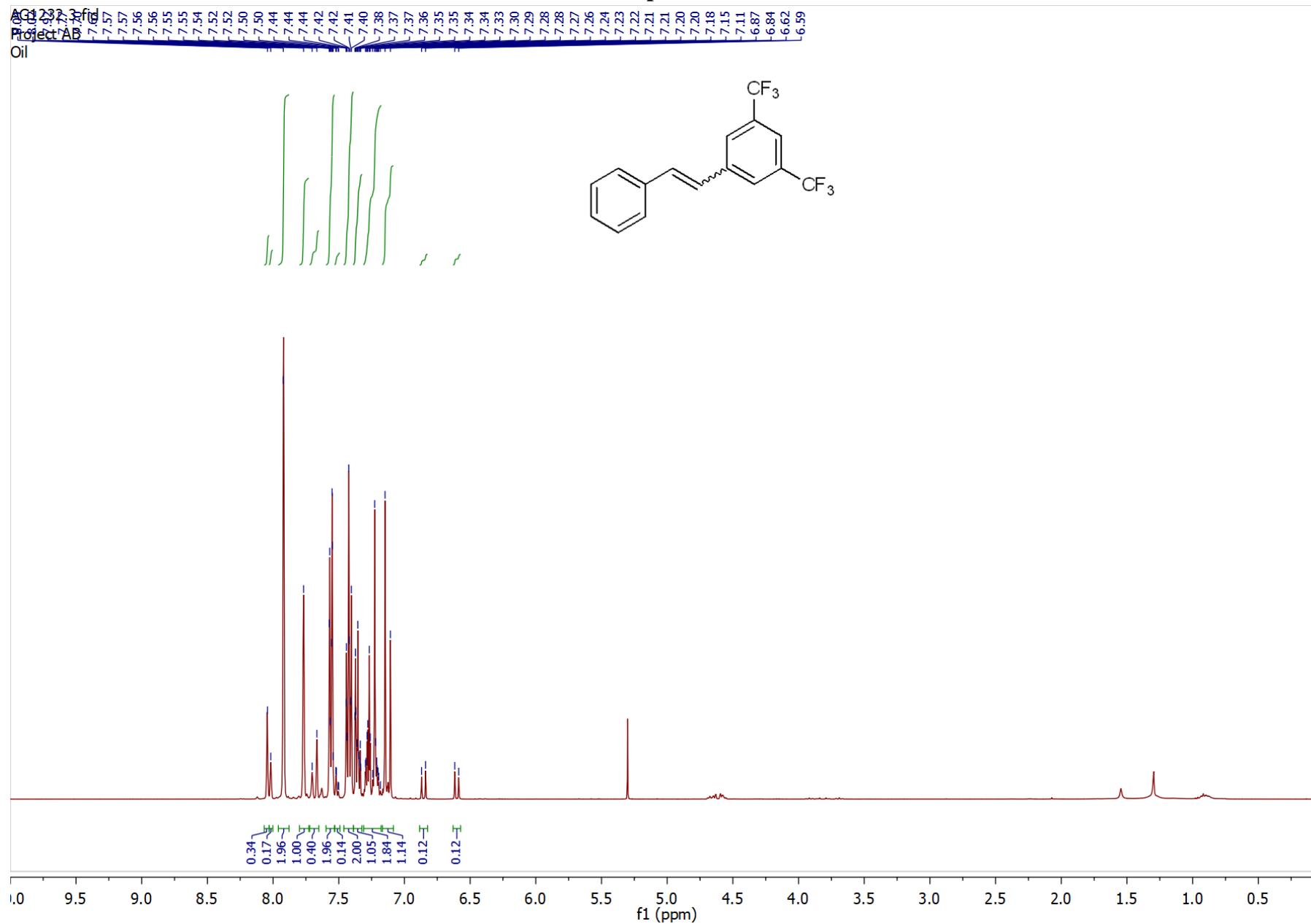


Compound 7e

AG1231.5.fid
Project AB_
Oil



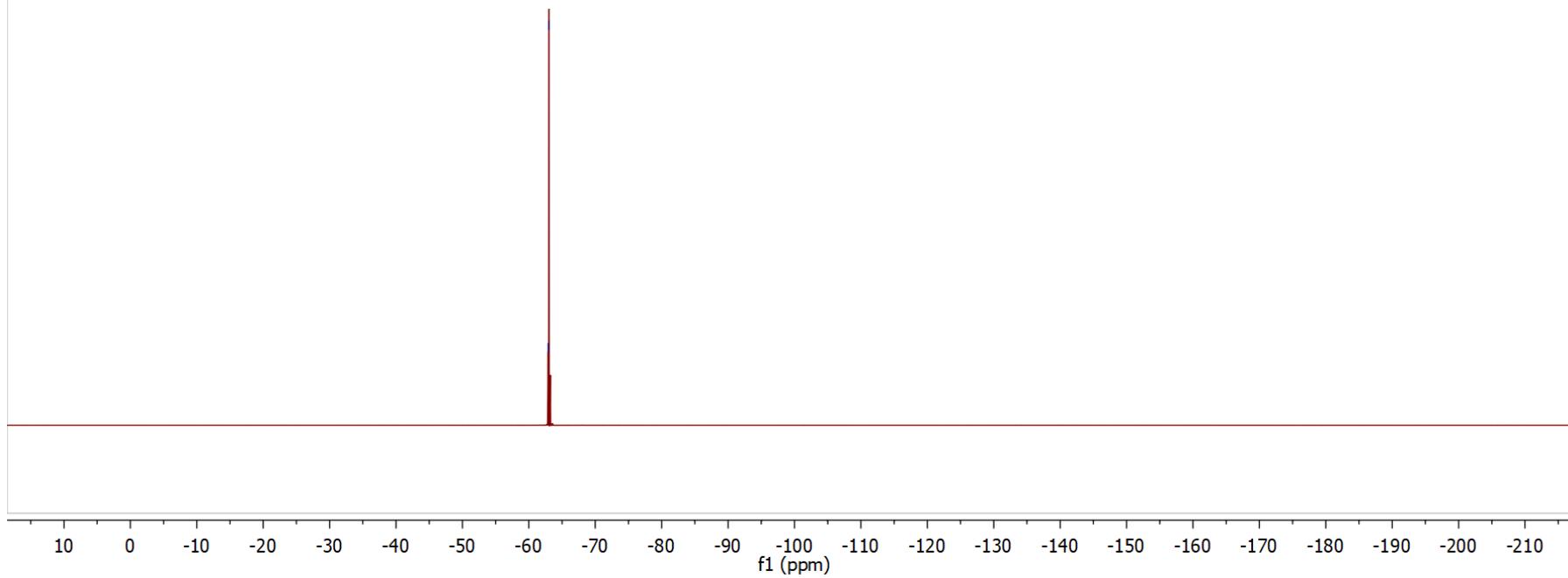
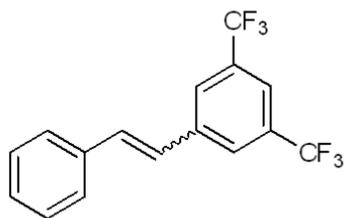
Compound 7f



Compound 7f

AG1232.2.fid
Project AB_
Oil

-62.91
-63.04

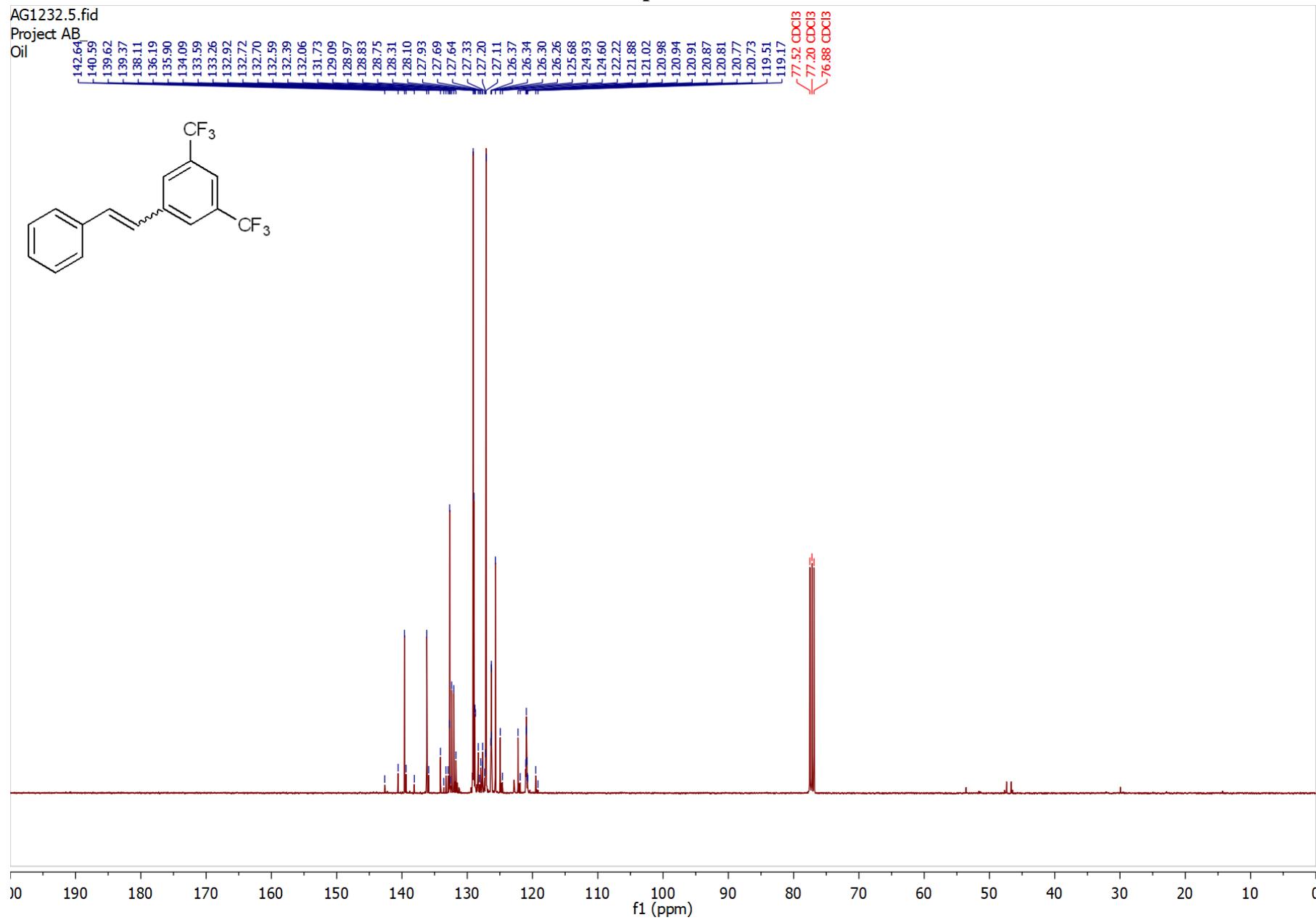


Compound 7f

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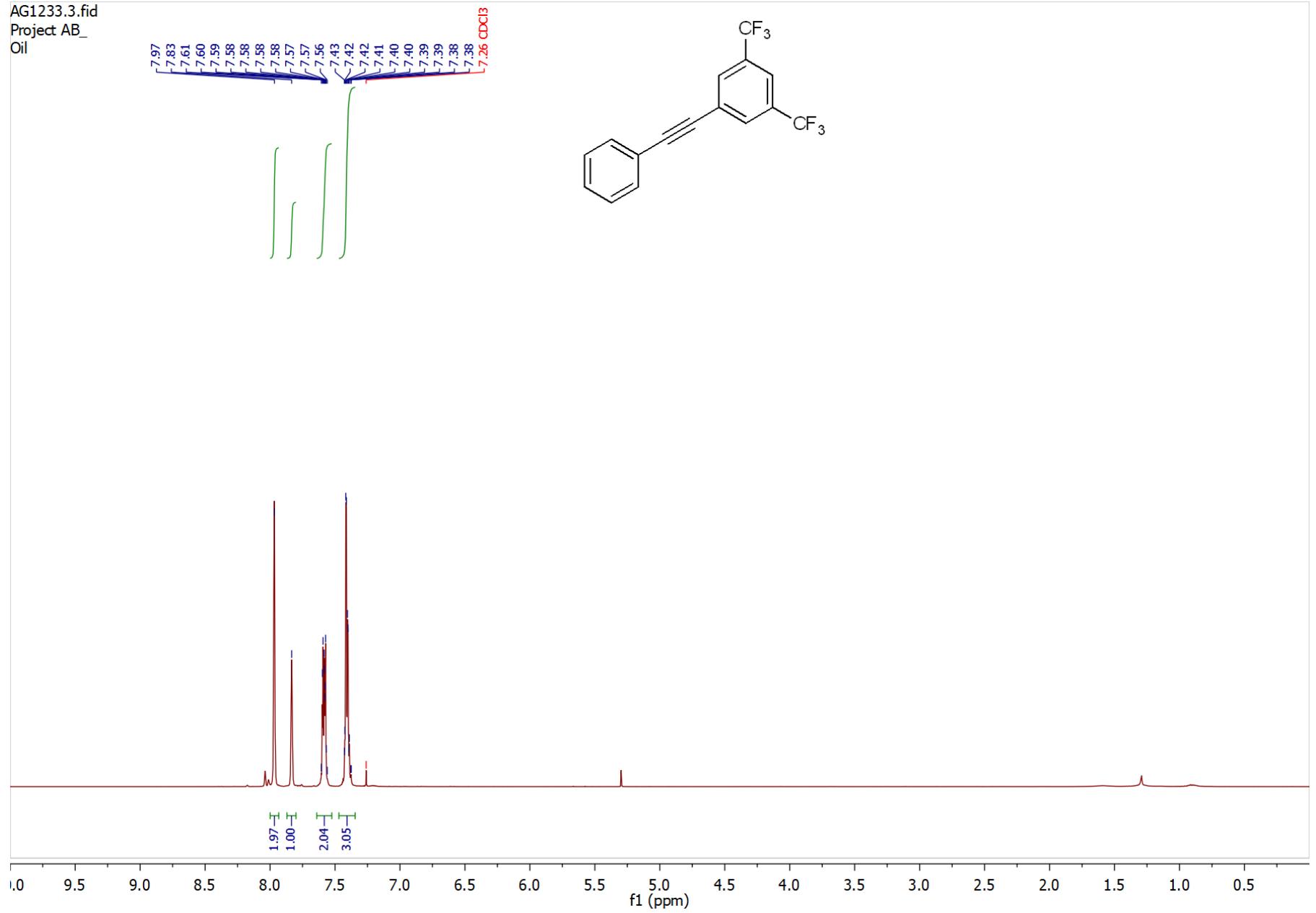
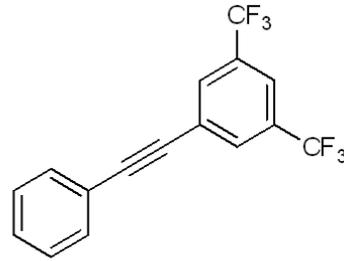
Project AB

Oil



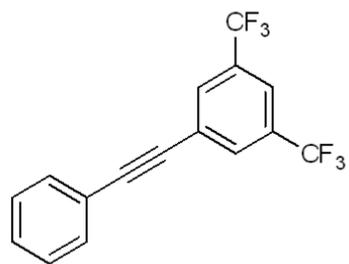
Compound 7g

AG1233.3.fid
Project AB_
Oil

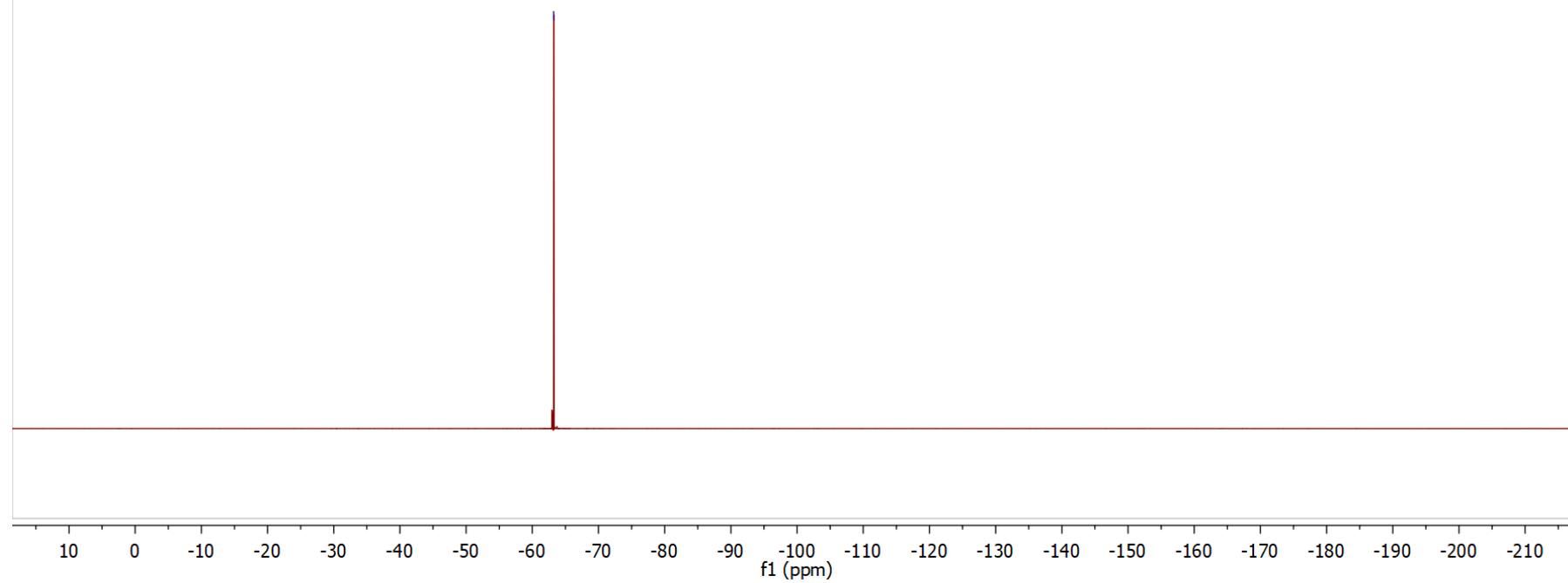


Compound 7g

AG1233.2.fid
Project AB_
Oil

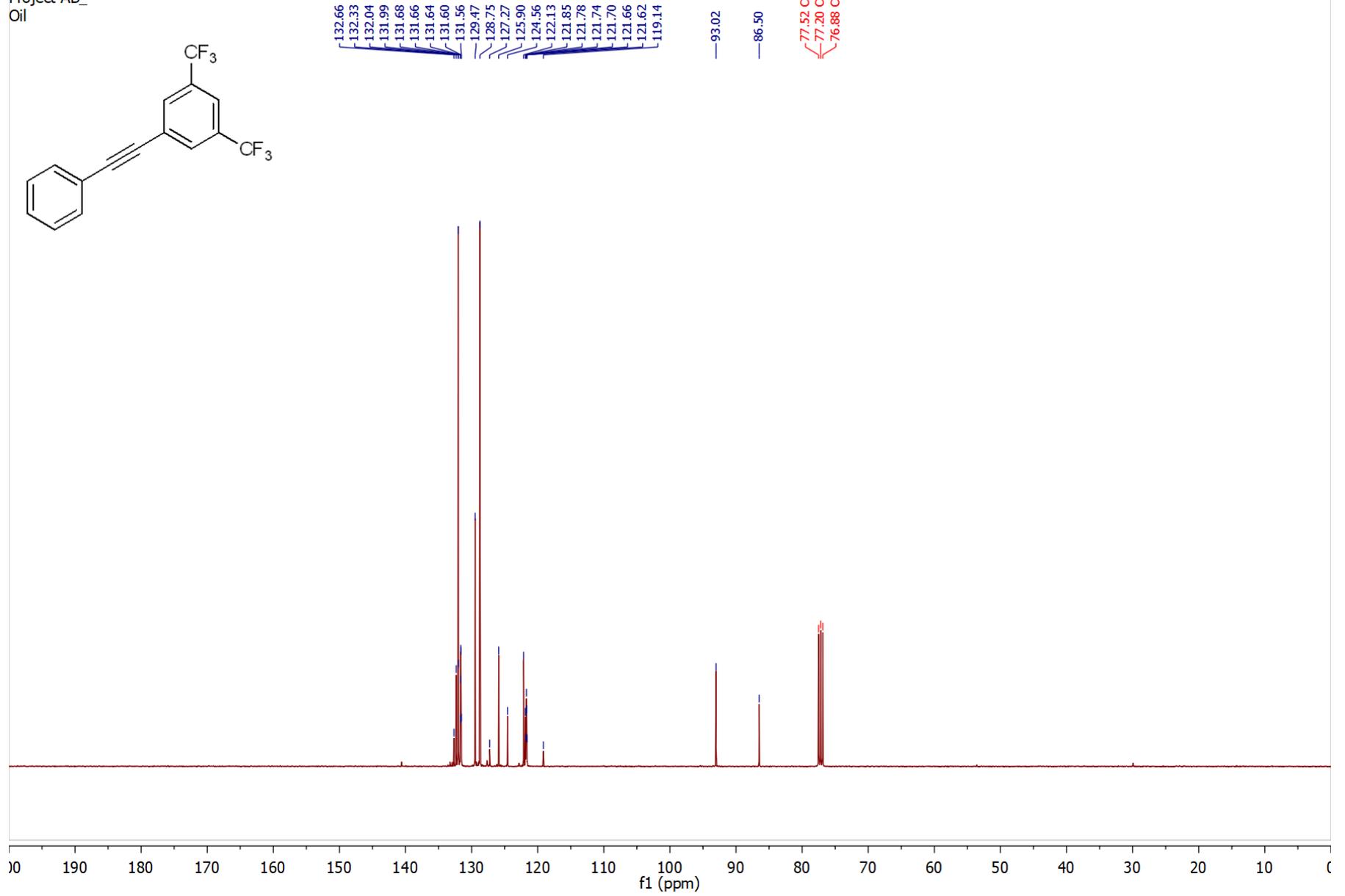
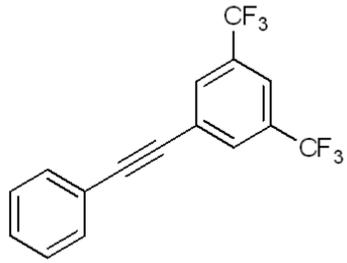


—63.21



Compound 7g

AG1233.5.fid
Project AB_
Oil

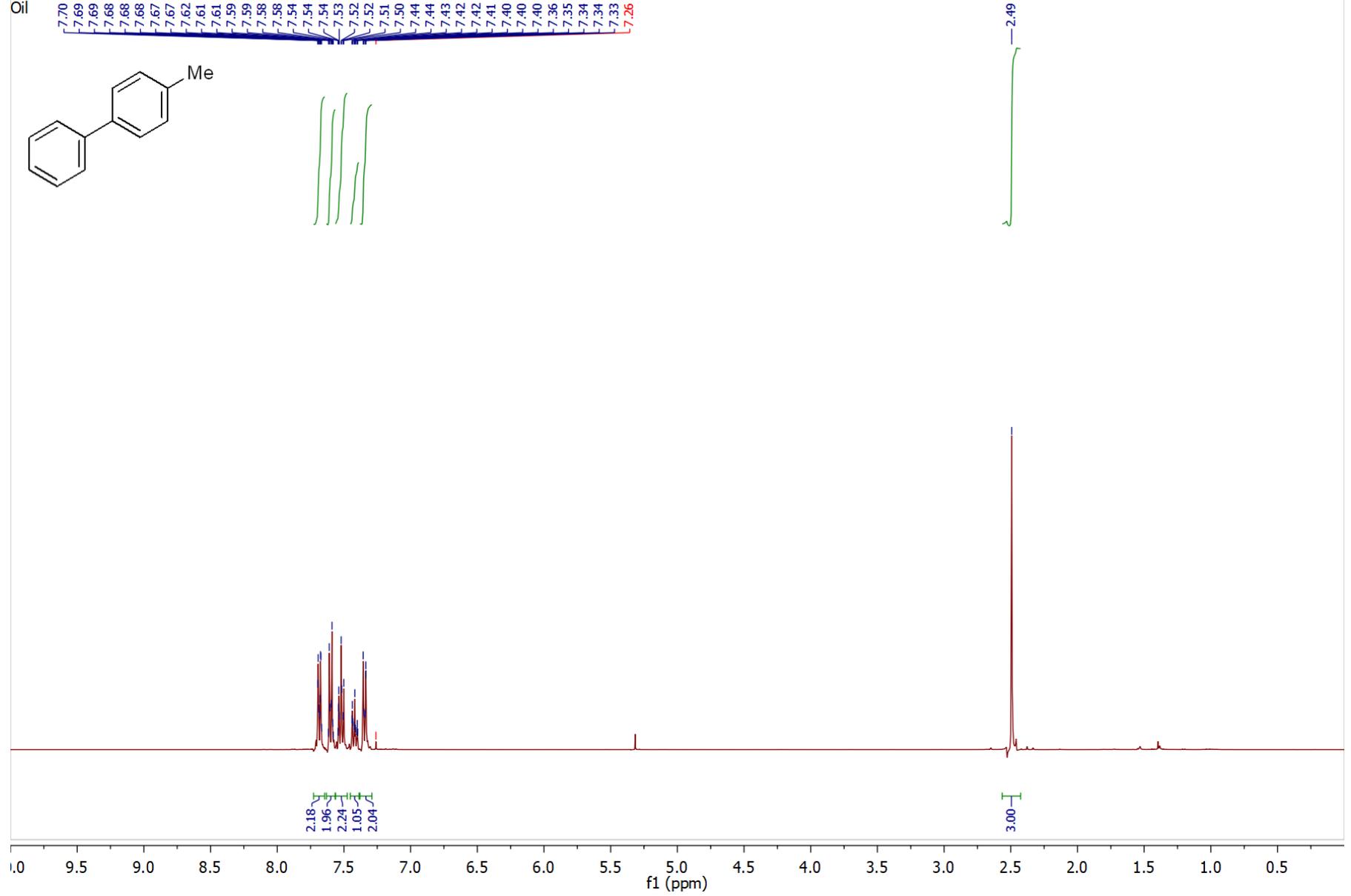


Compound 7i

AG1234.2.fid

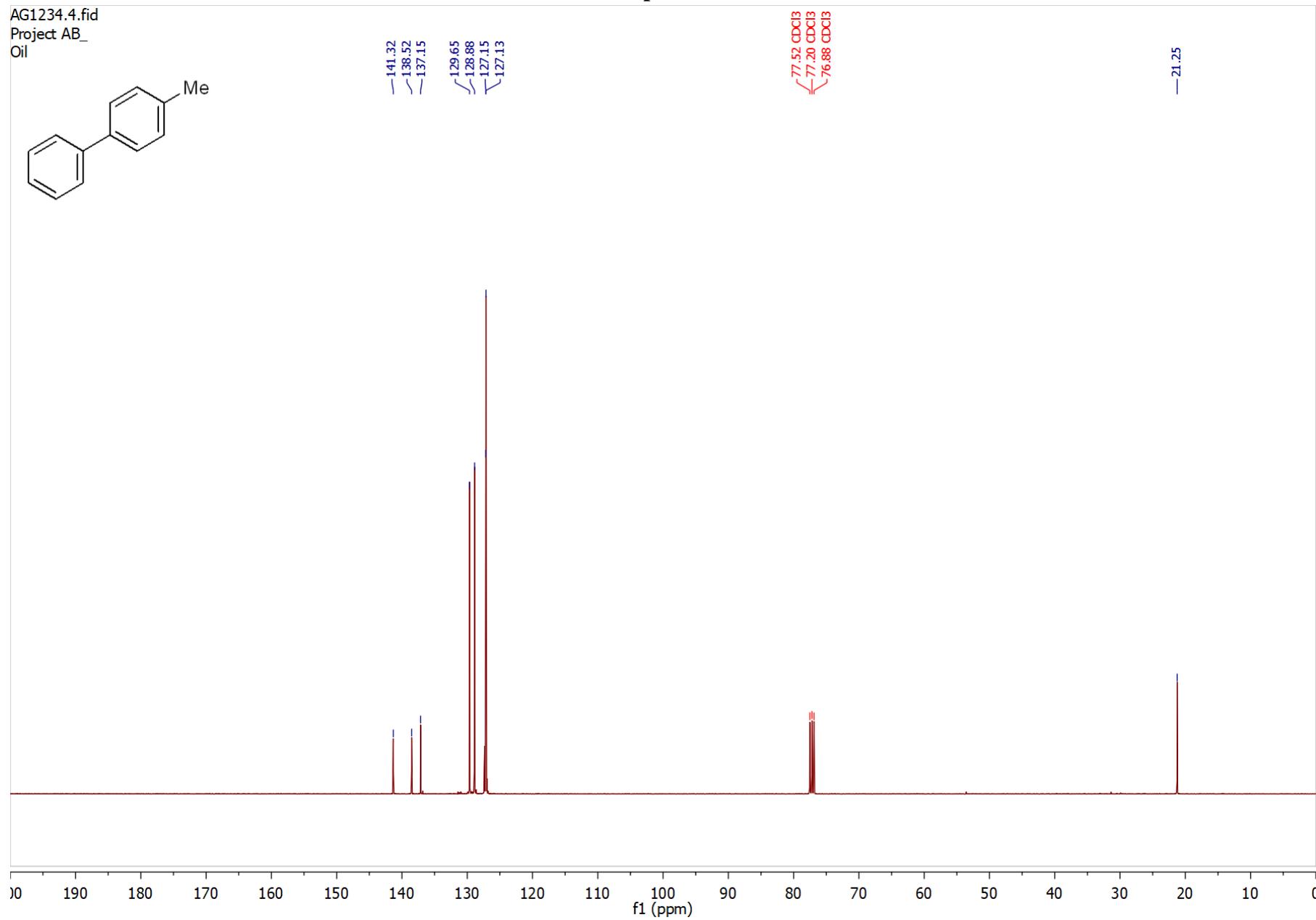
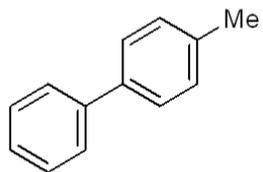
Project AB

Oil



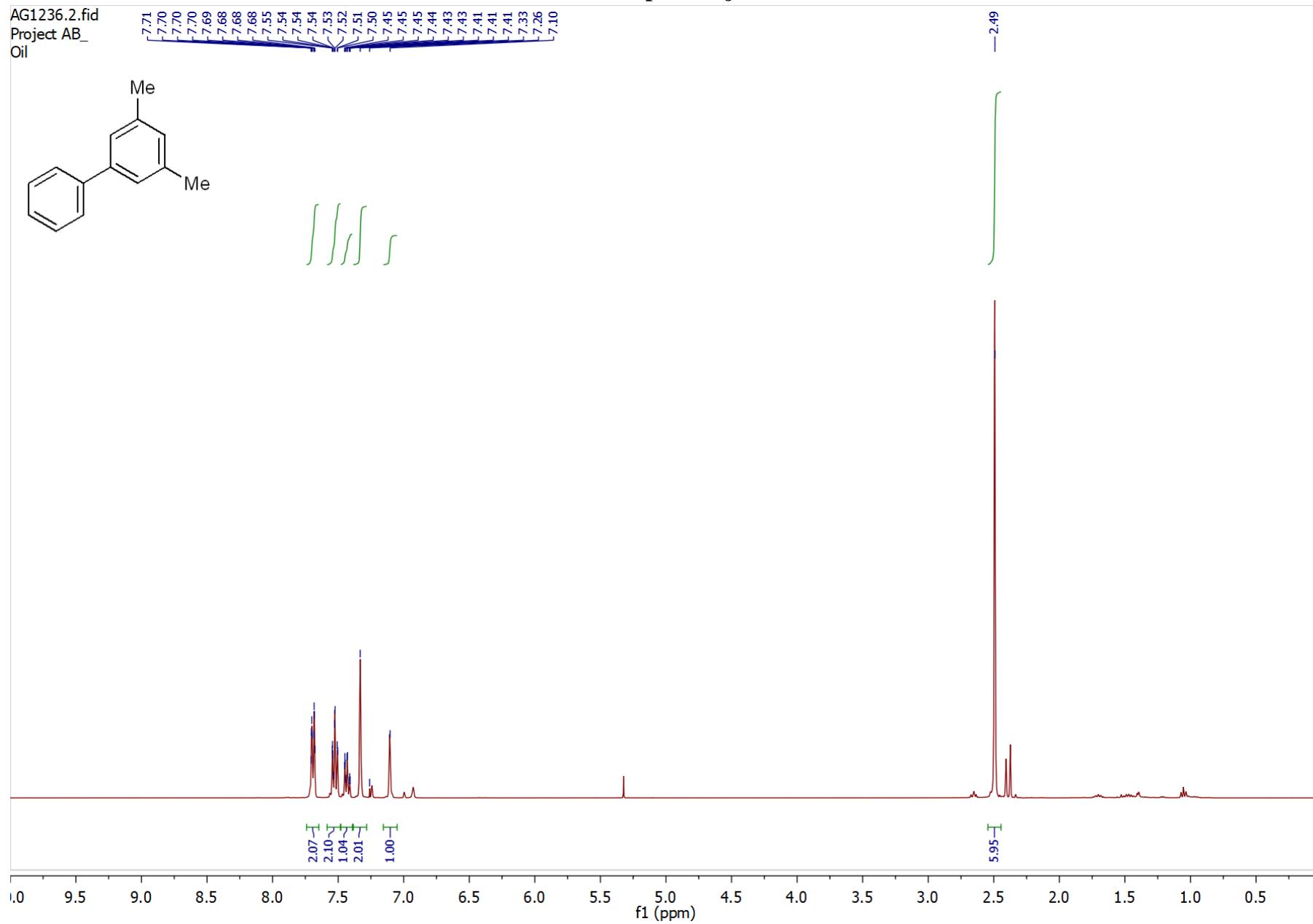
Compound 7i

AG1234.4.fid
Project AB_
Oil



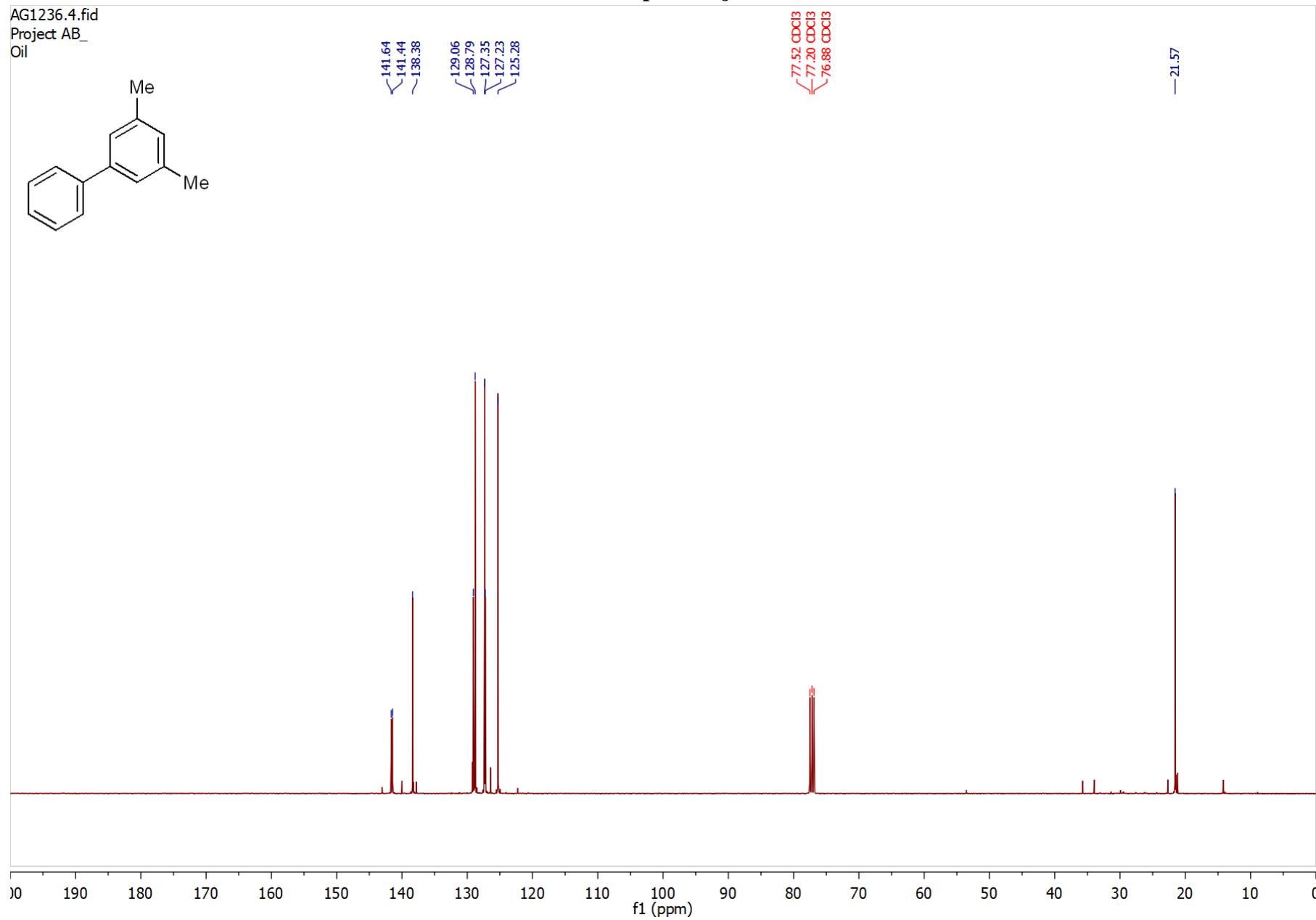
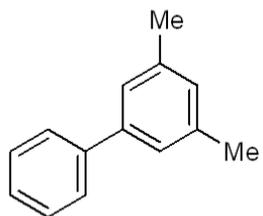
Compound 7j

AG1236.2.fid
Project AB_
Oil



Compound 7j

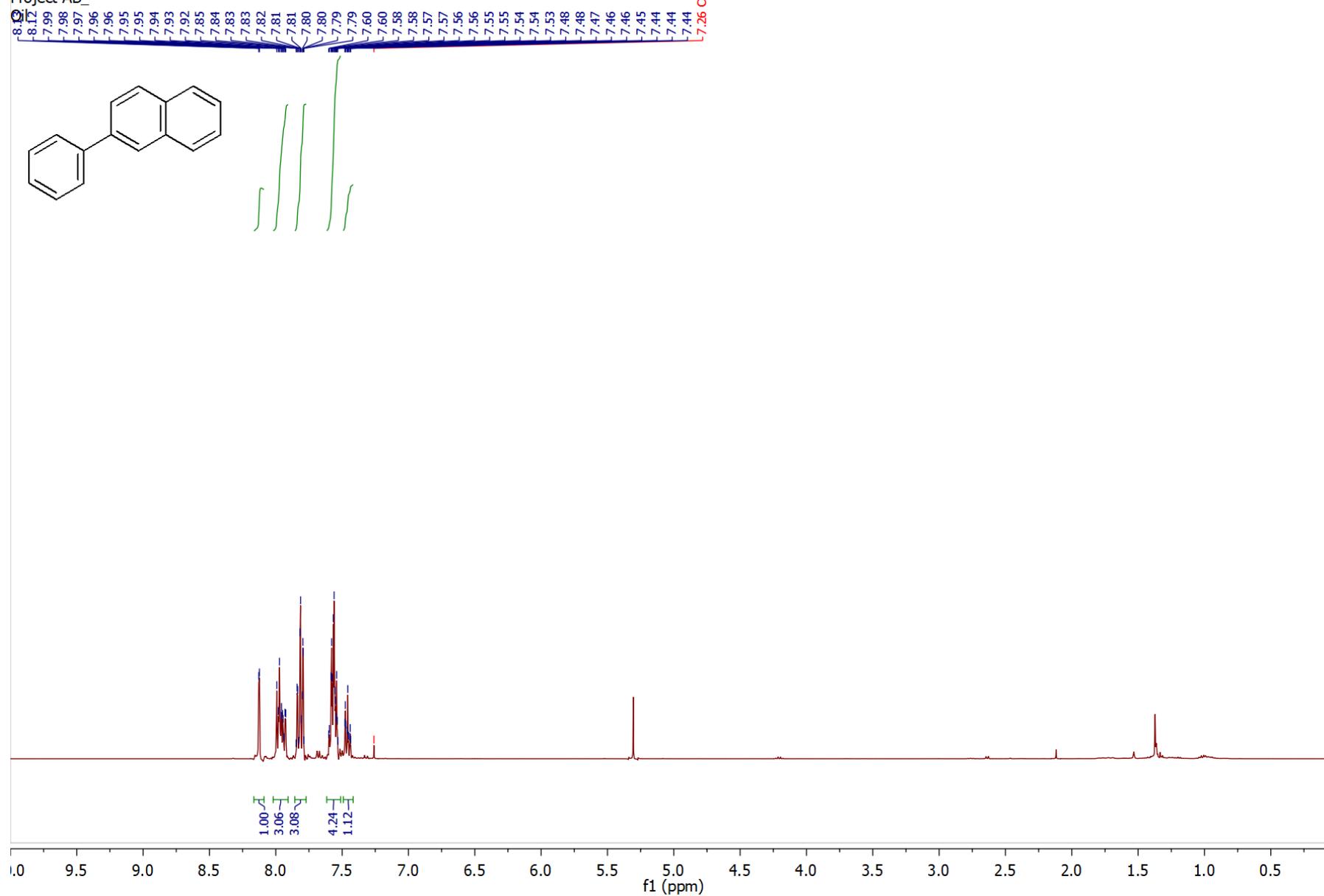
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Project AB_
Oil



Compound 7k

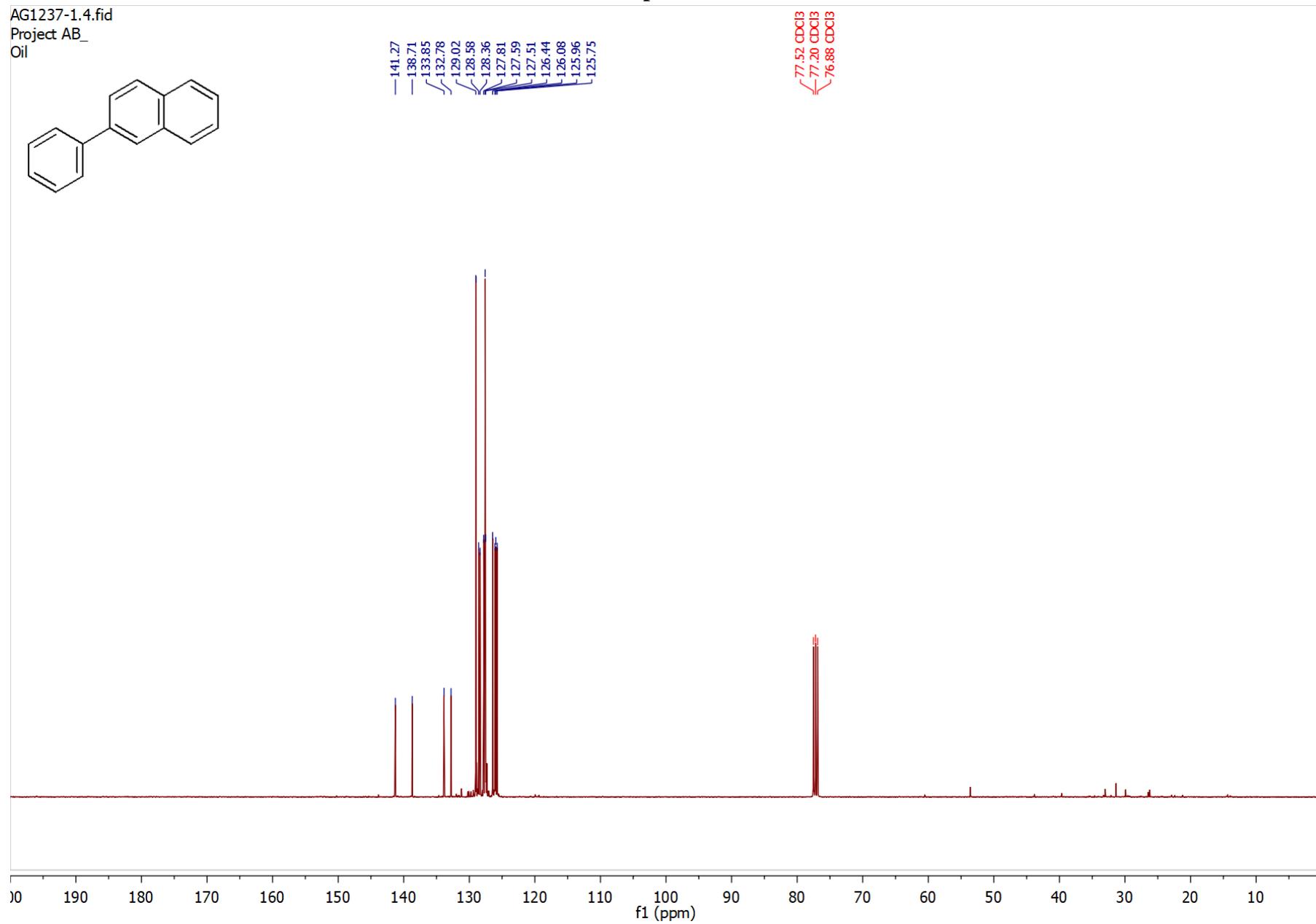
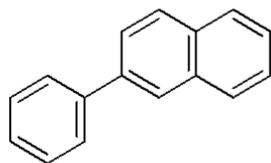
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Project AB



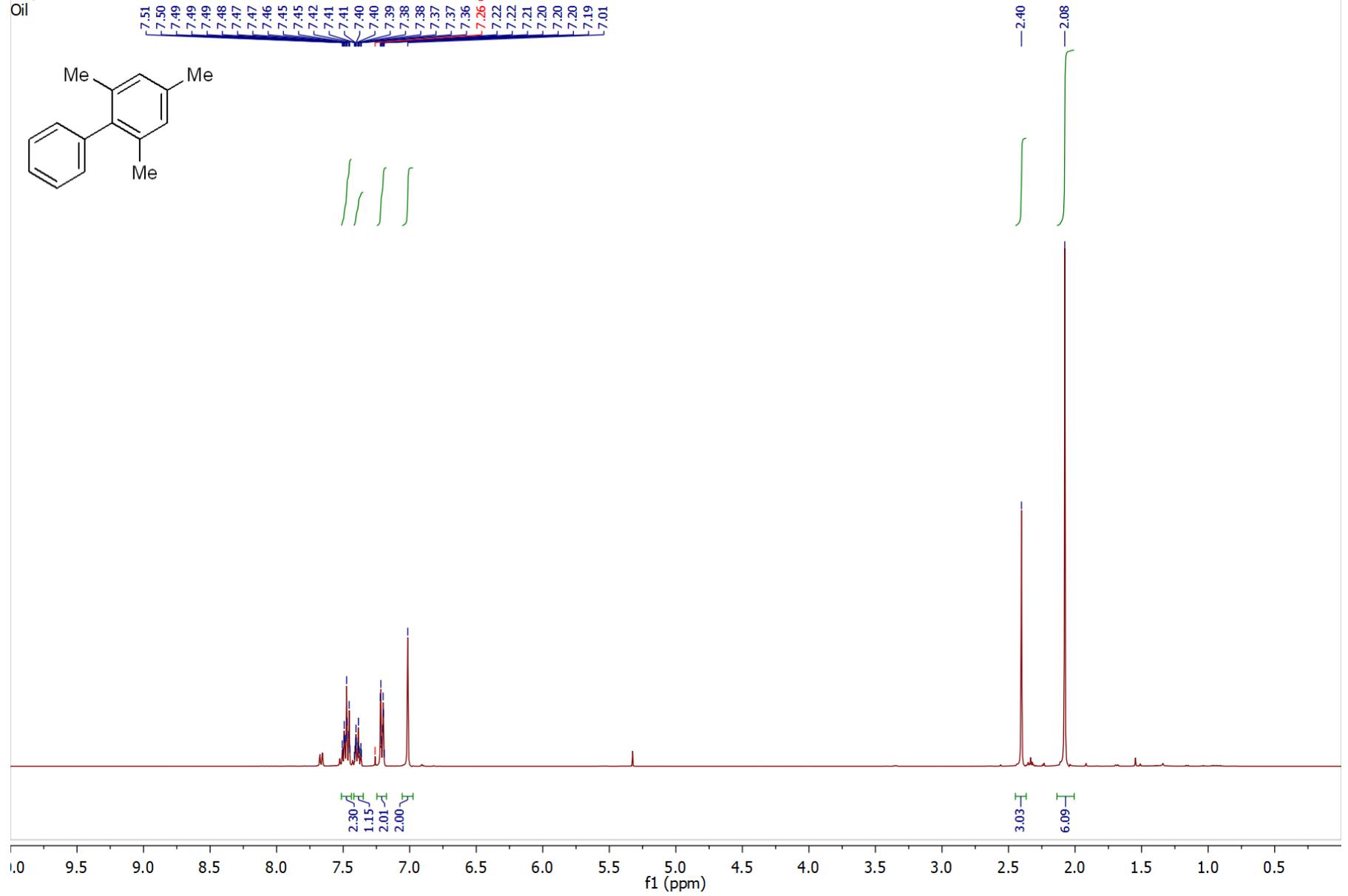
Compound 7k

AG1237-1.4.fid
Project AB_
Oil



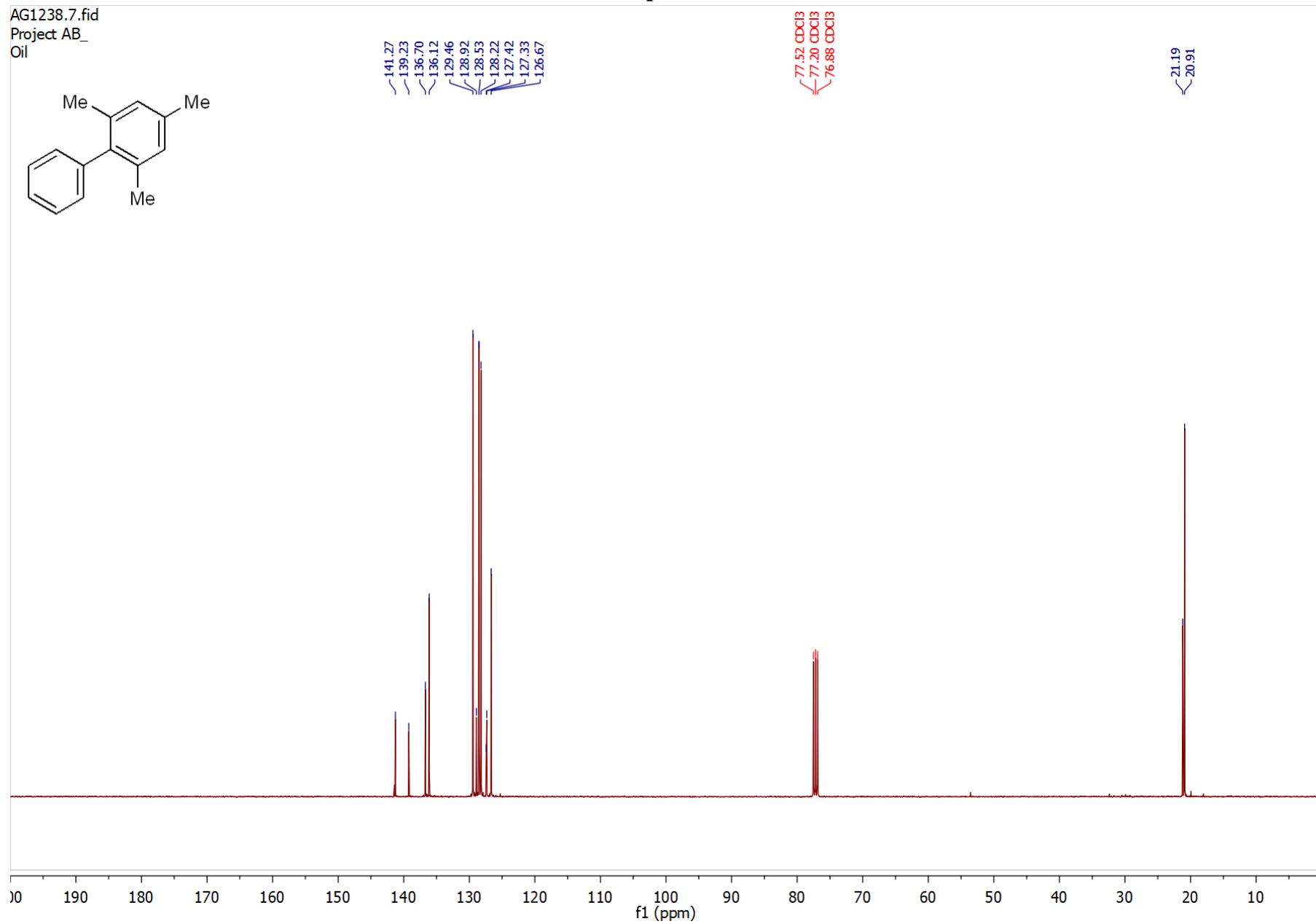
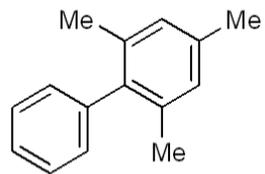
Compound 71

AG1238.3.fid
Project AB_
Oil



Compound 71

AG1238.7.fid
Project AB_
Oil

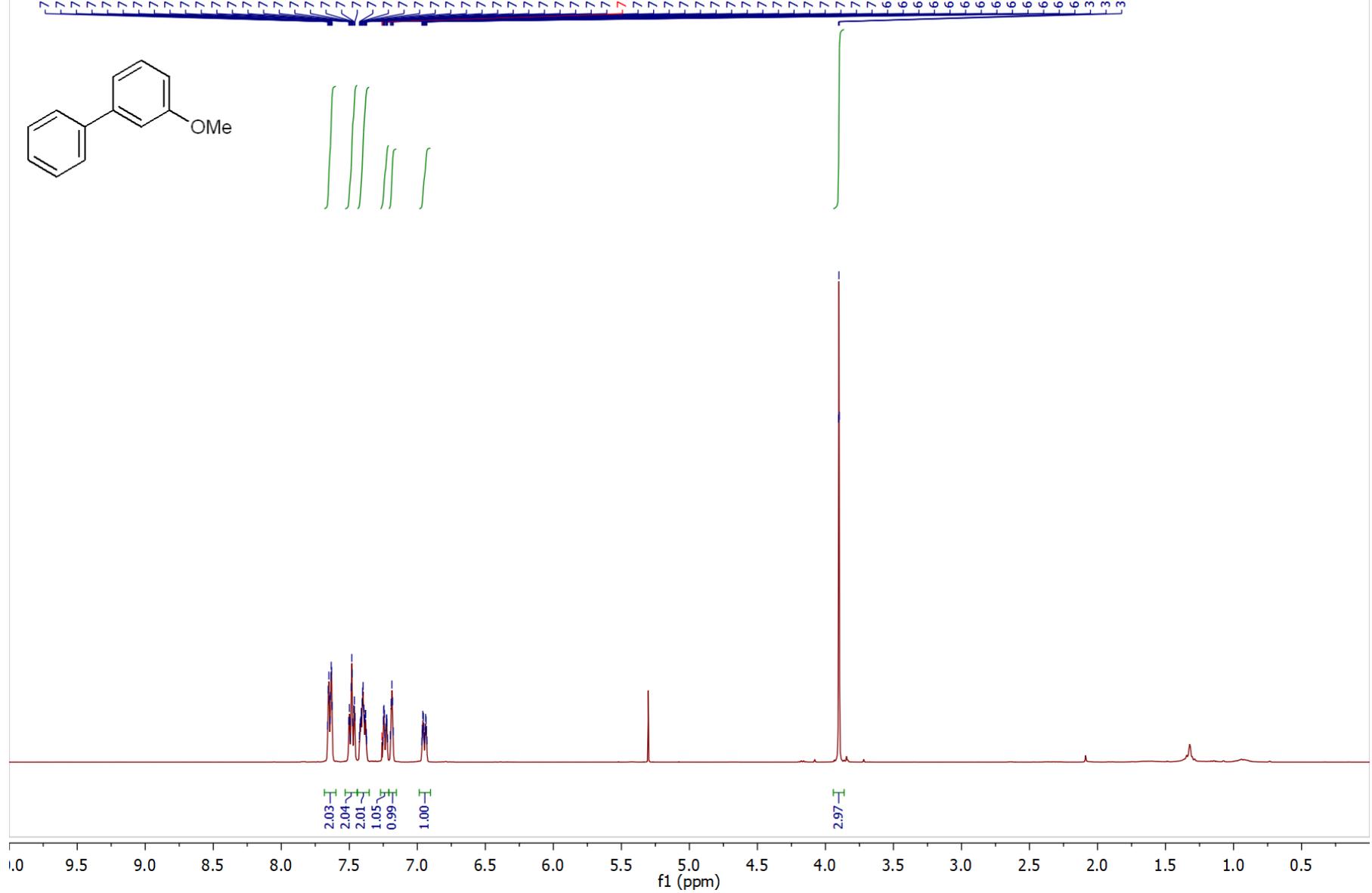


Compound 7m

AG1239.2.fid

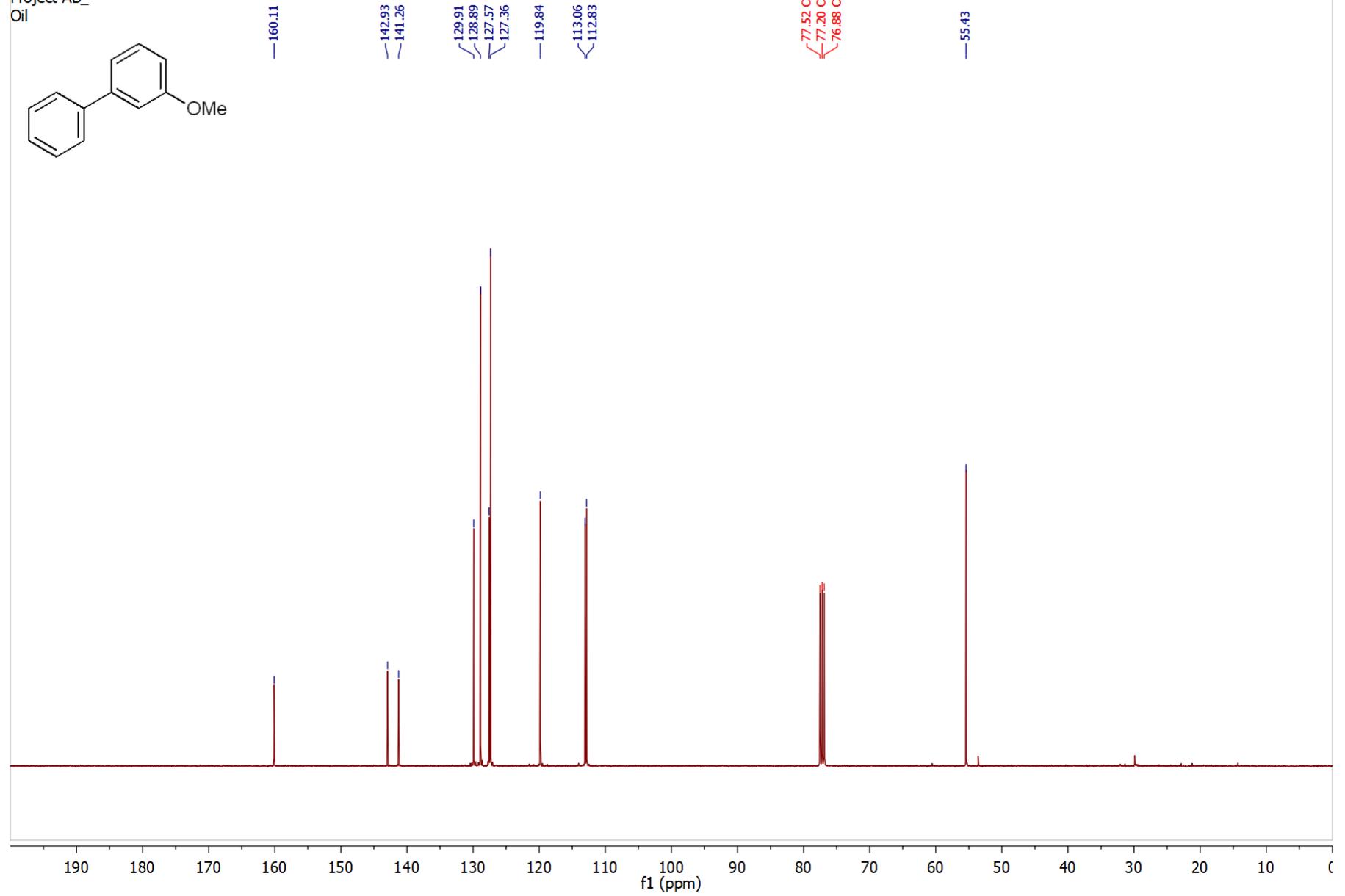
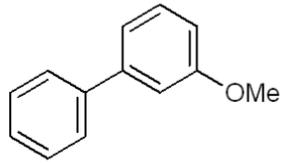
Project AB

Oil



Compound 7m

AG1239.4.fid
Project AB_
Oil

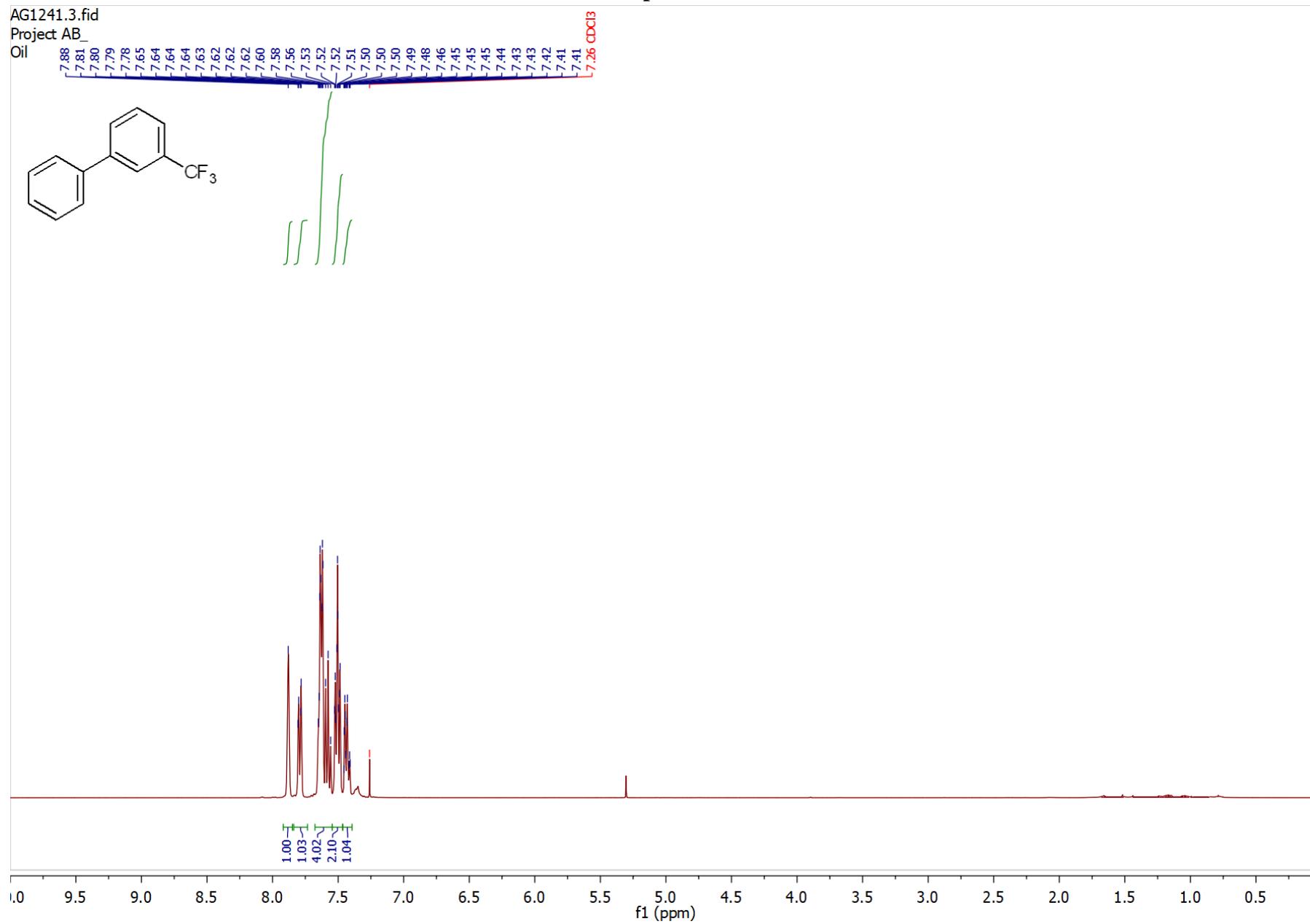


Compound 7n

AG1241.3.fid

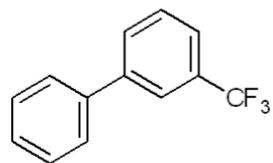
Project AB

Oil

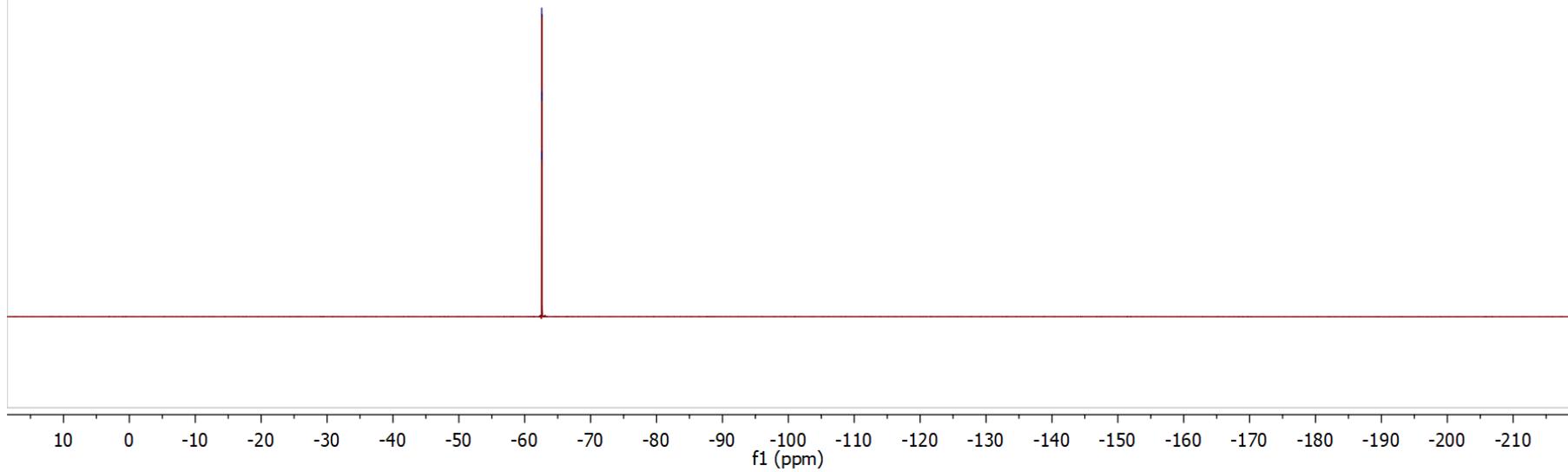


Compound 7n

AG1241.2.fid
Project AB_
Oil

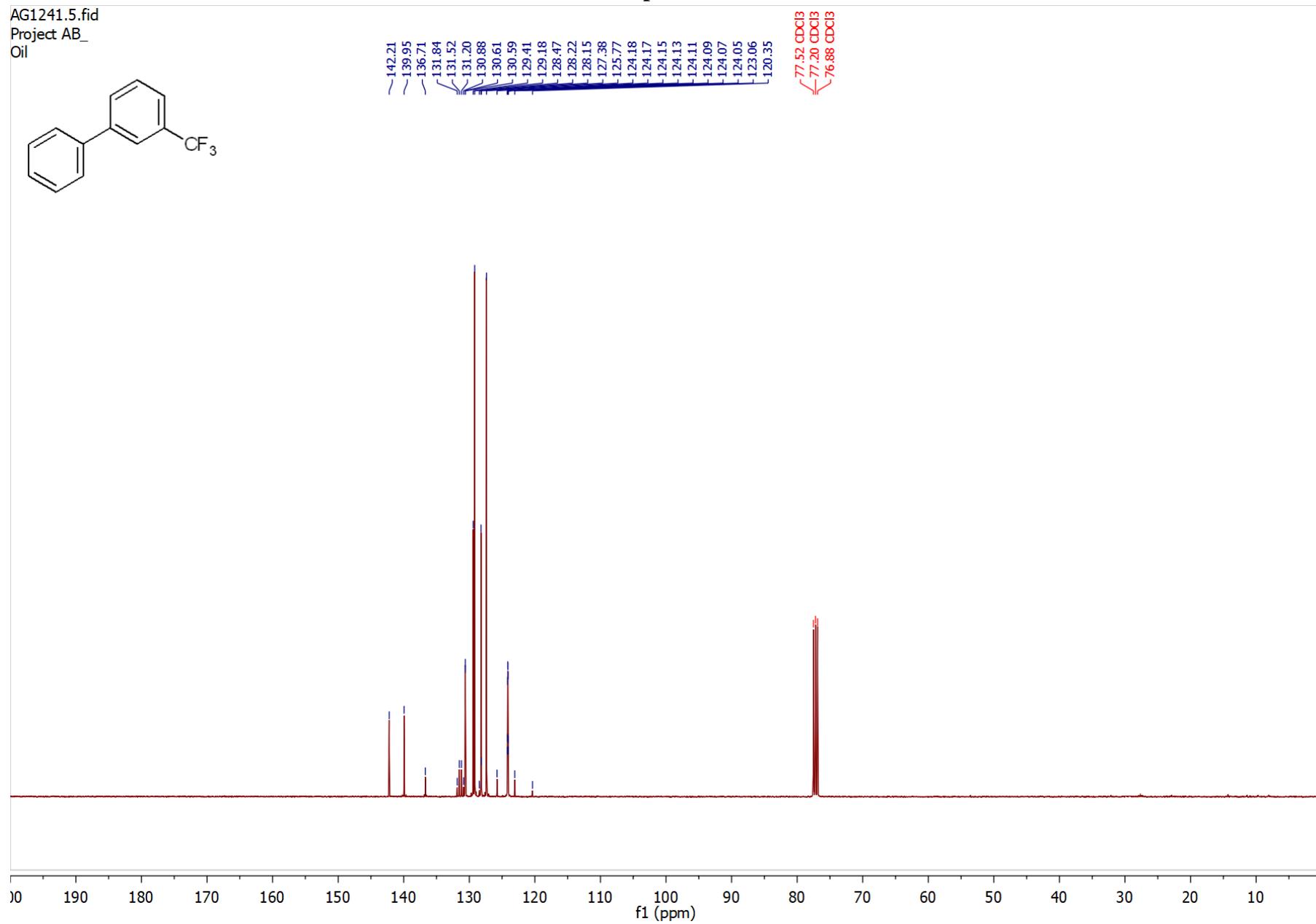
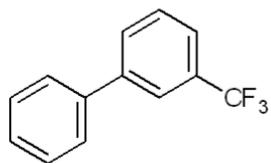


-62.56
-62.56
-62.57



Compound 7n

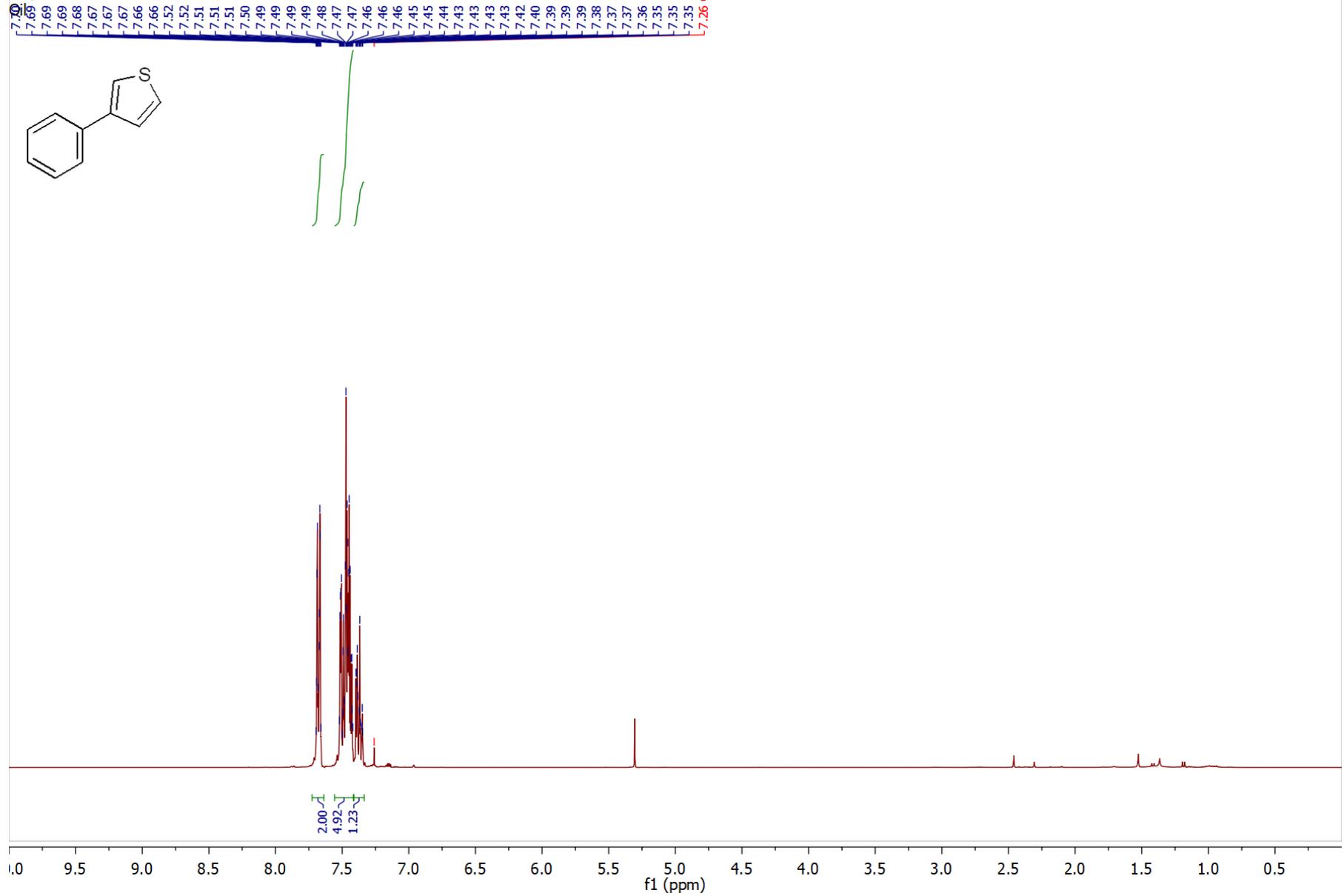
AG1241.5.fid
Project AB_
Oil



Compound 7o

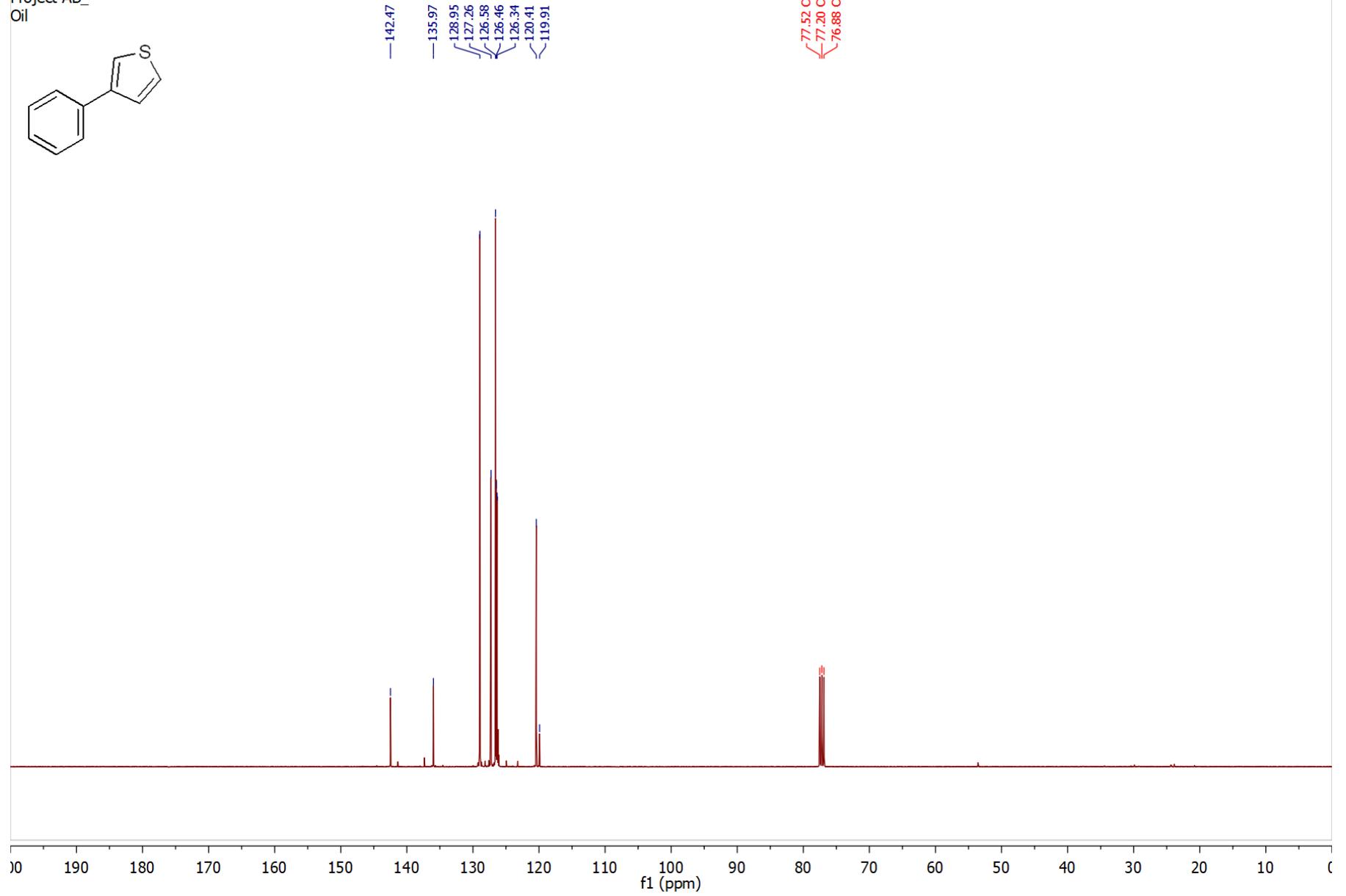
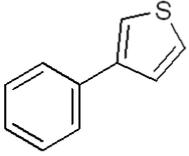
AG1242.1.fid

Project AB



Compound 7o

AG1242.4.fid
Project AB_
Oil

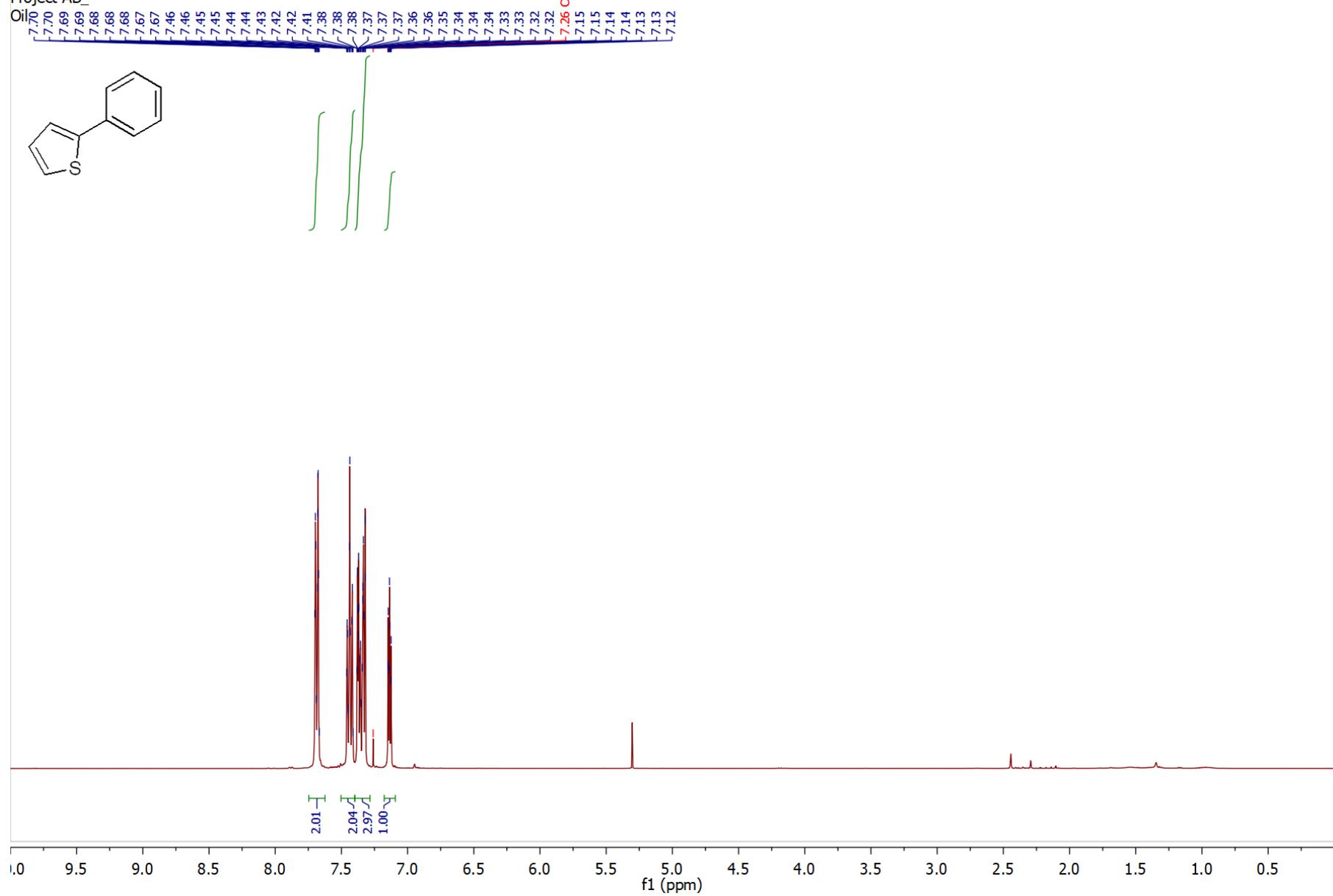


Compound 7p

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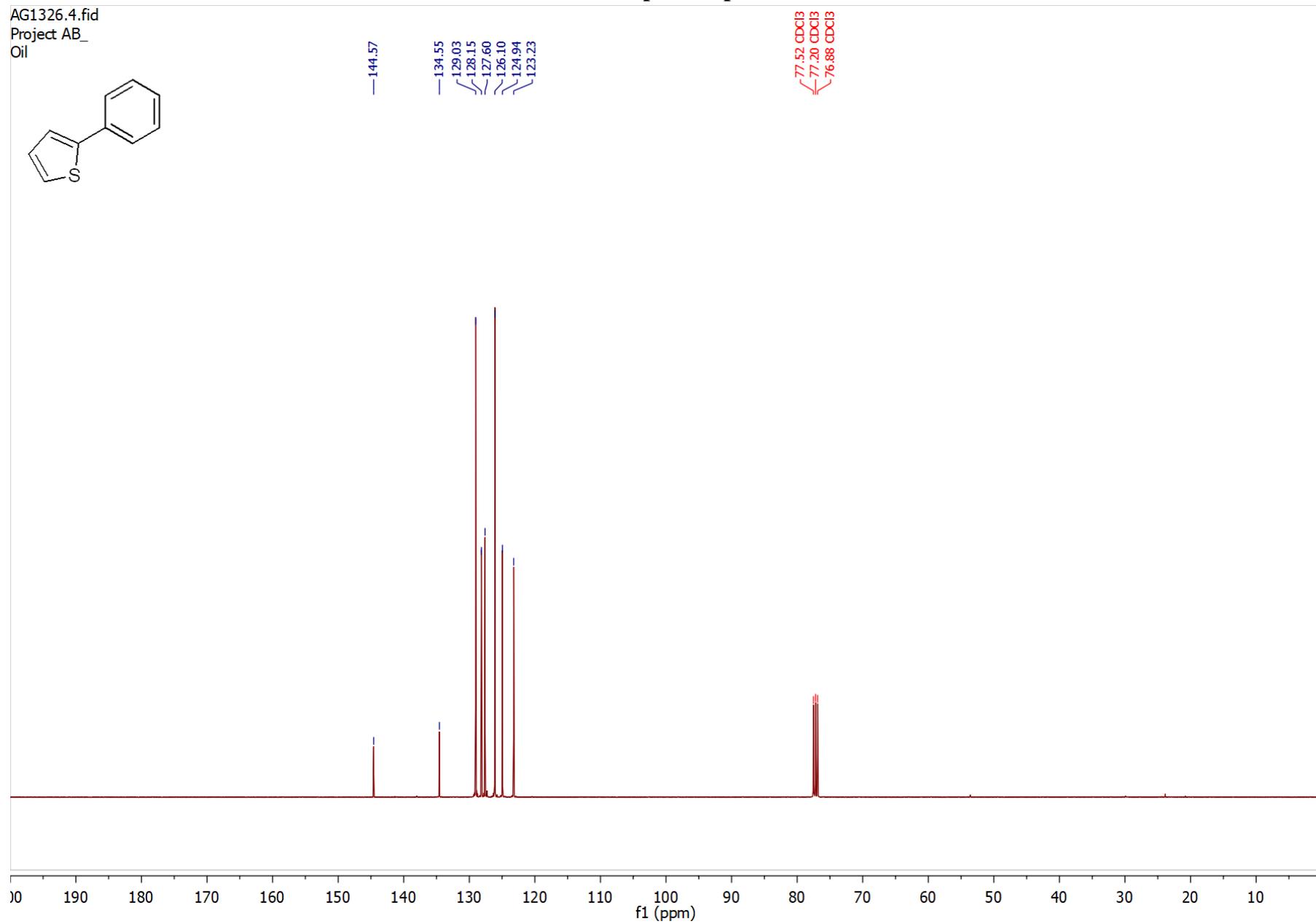
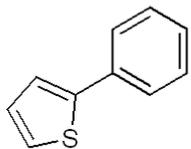
Project AB

Oil



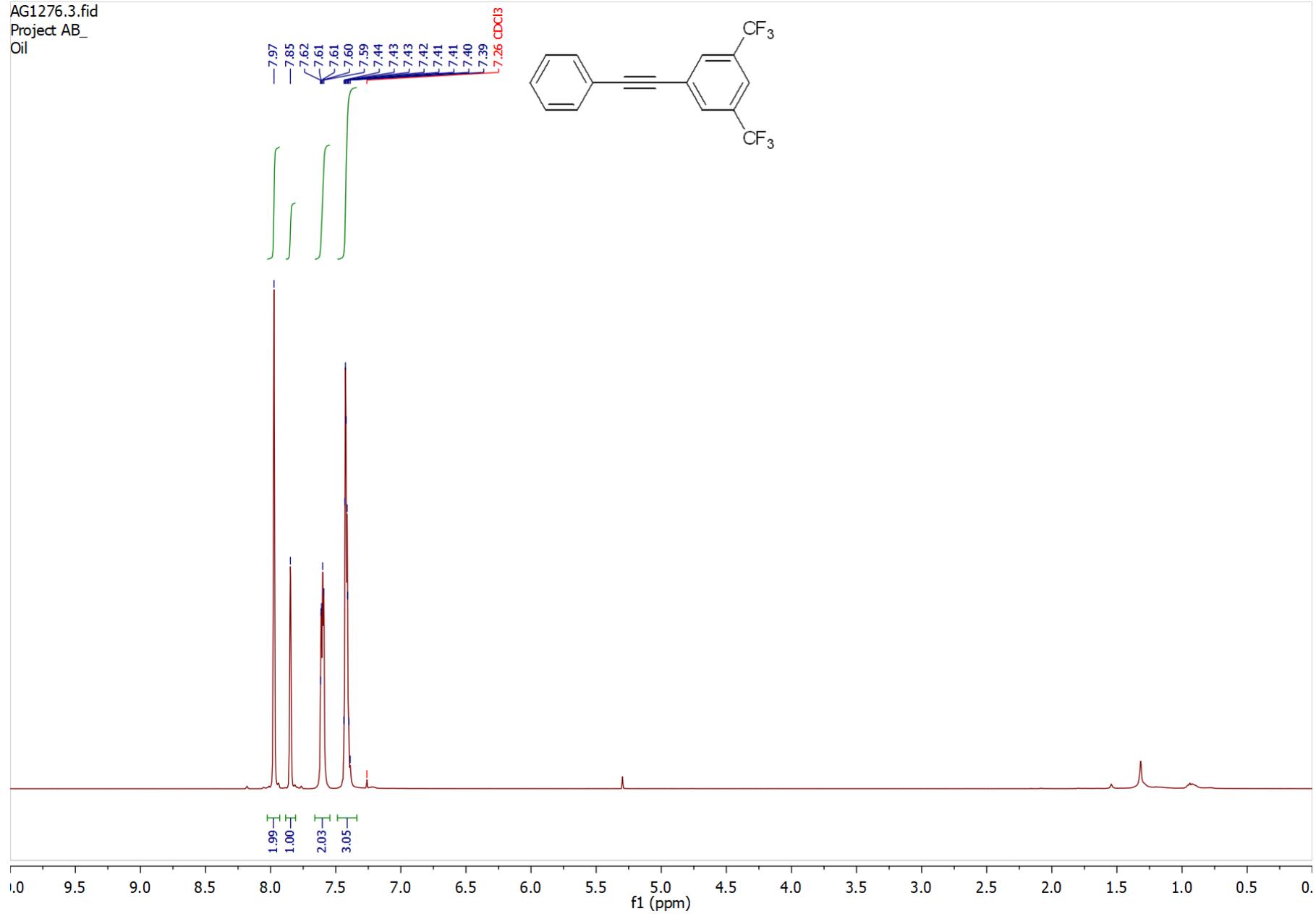
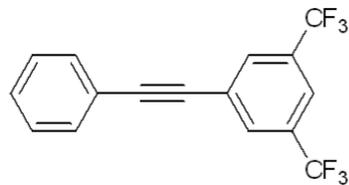
Compound 7p

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Project AB_
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Compound 9a

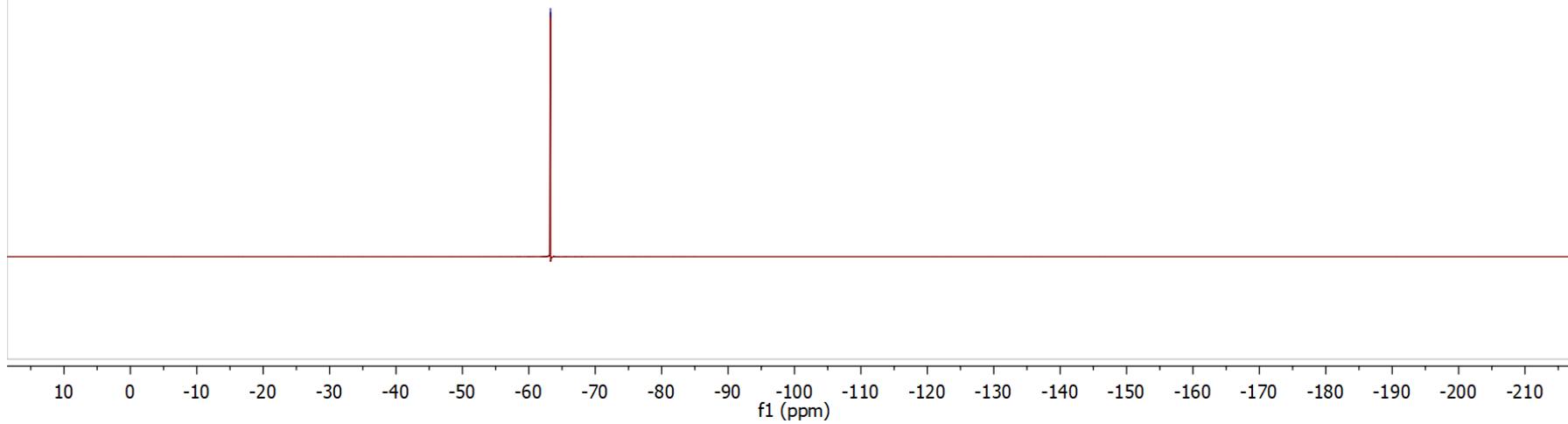
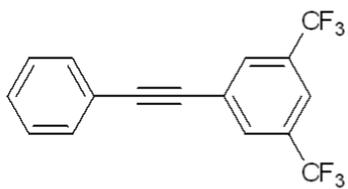
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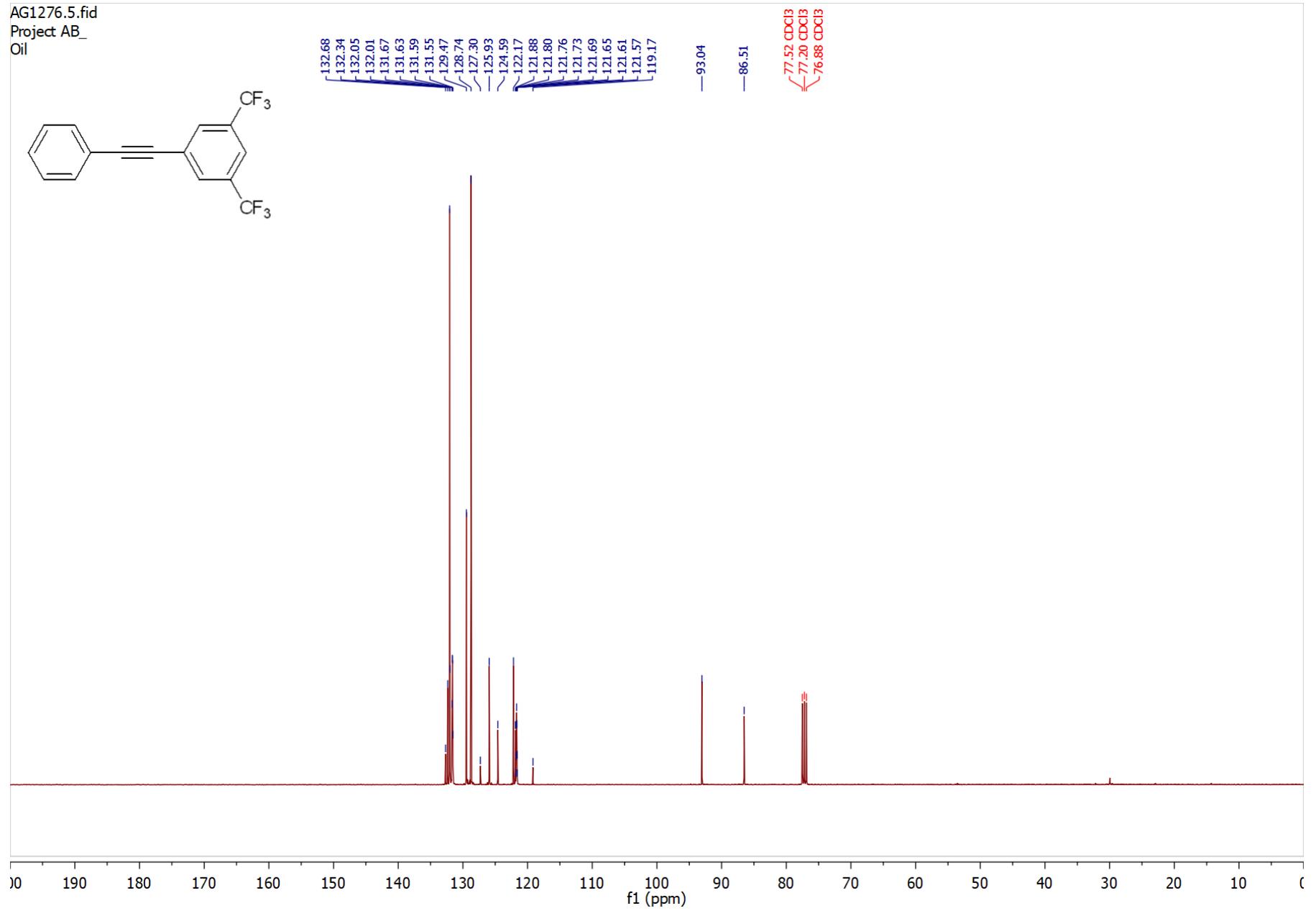
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—63.26



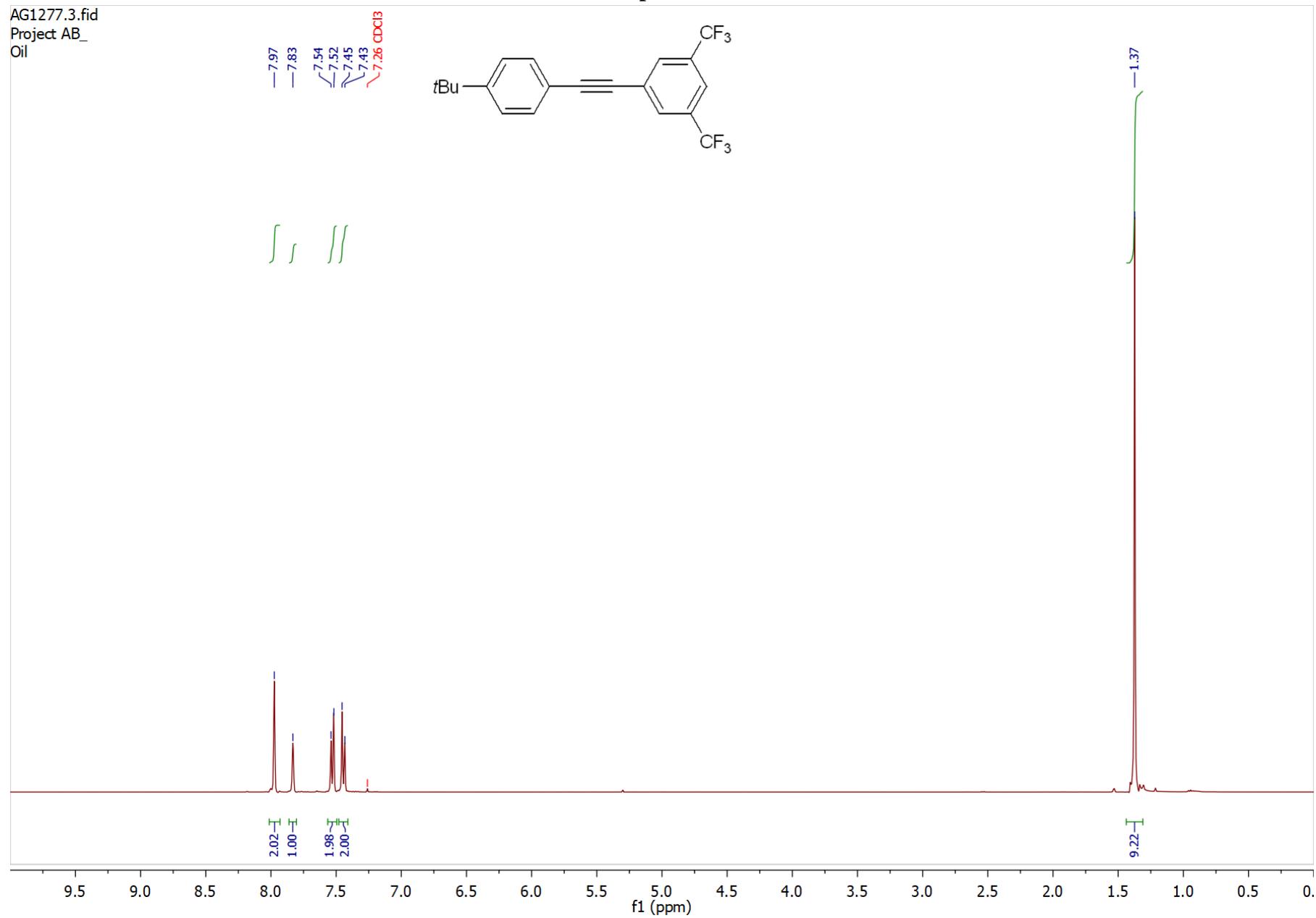
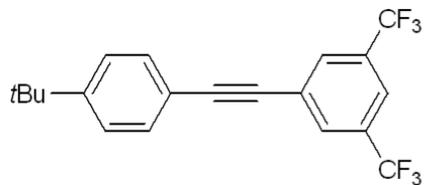
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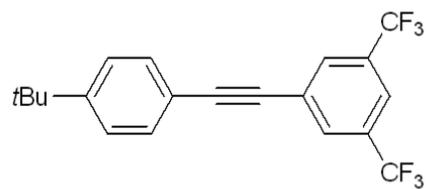
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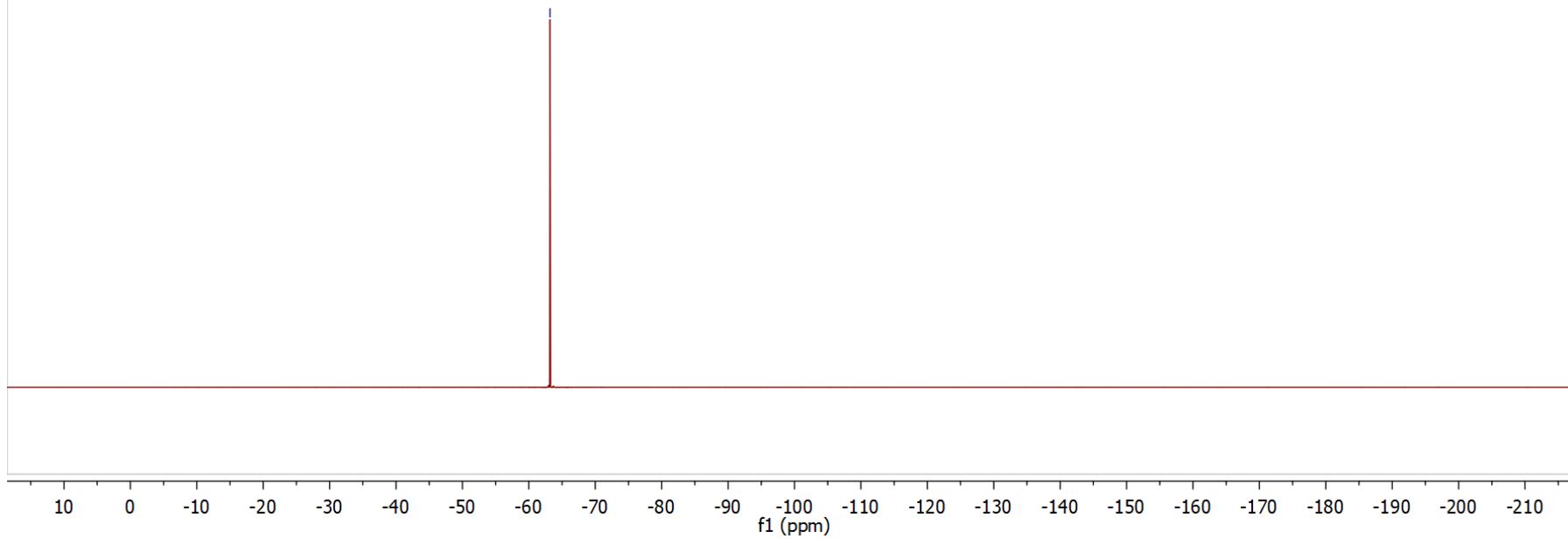


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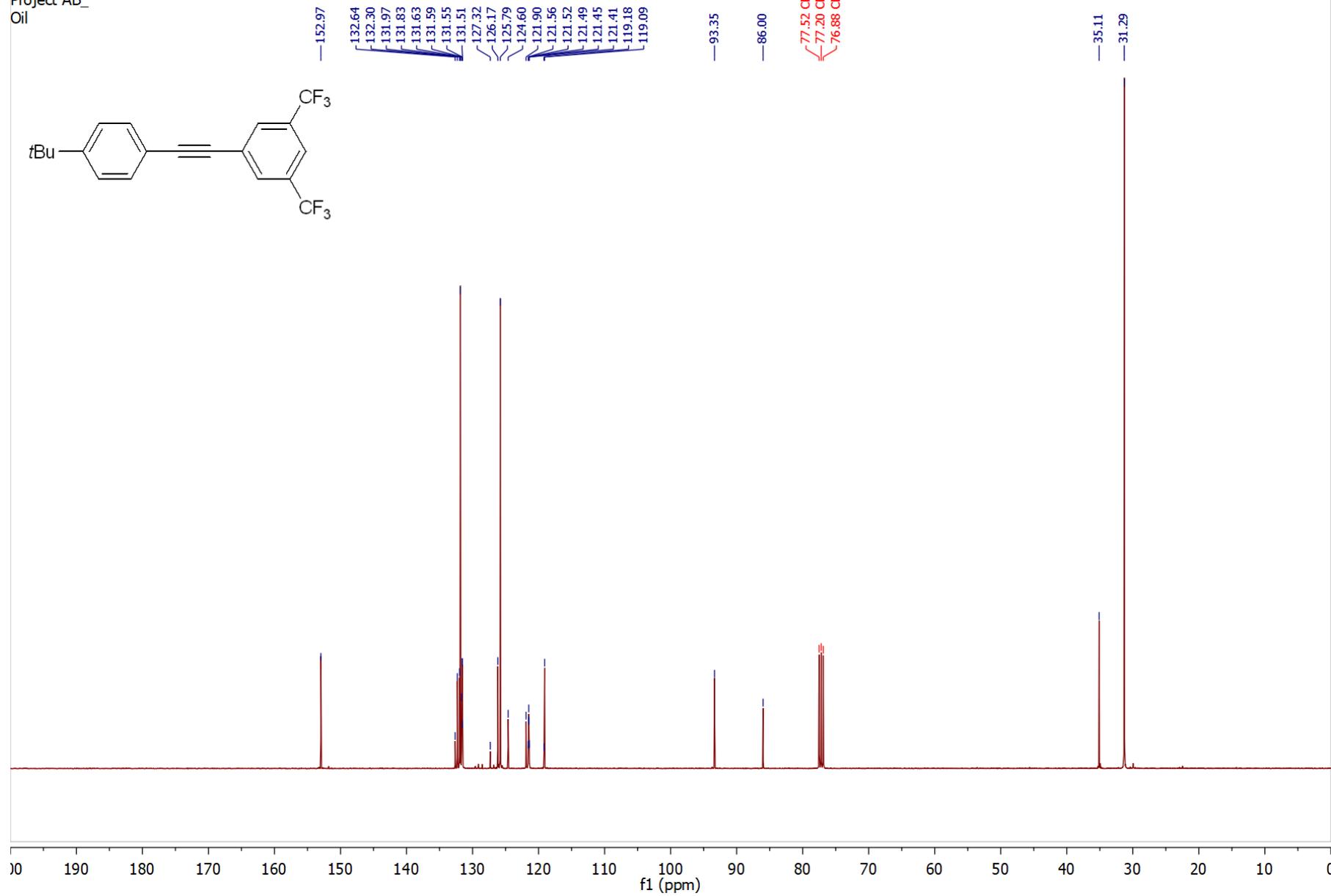


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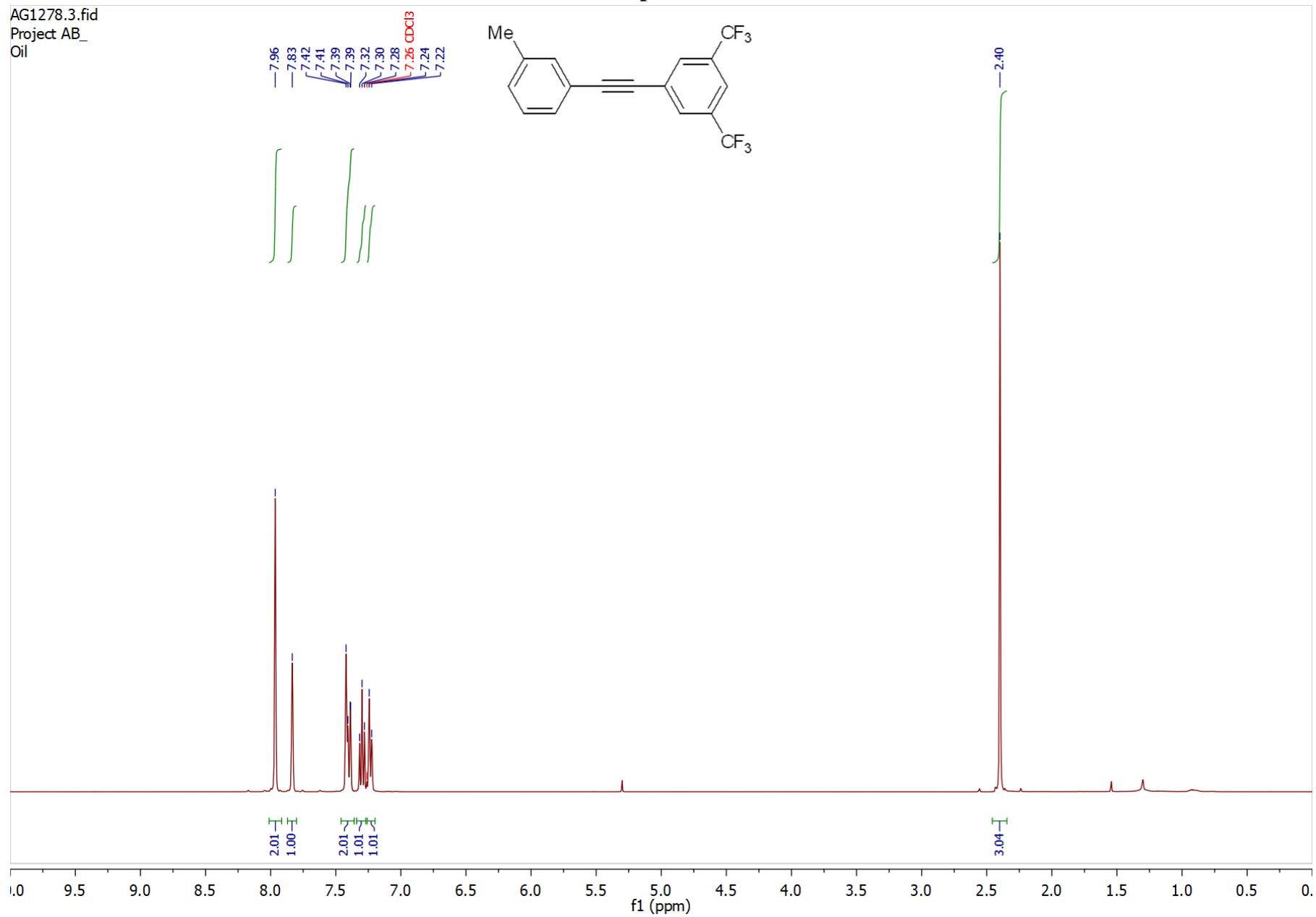
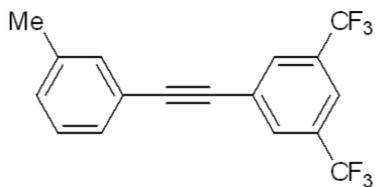
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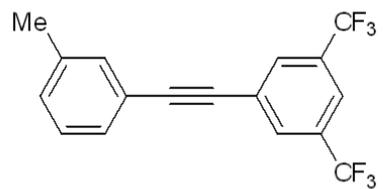
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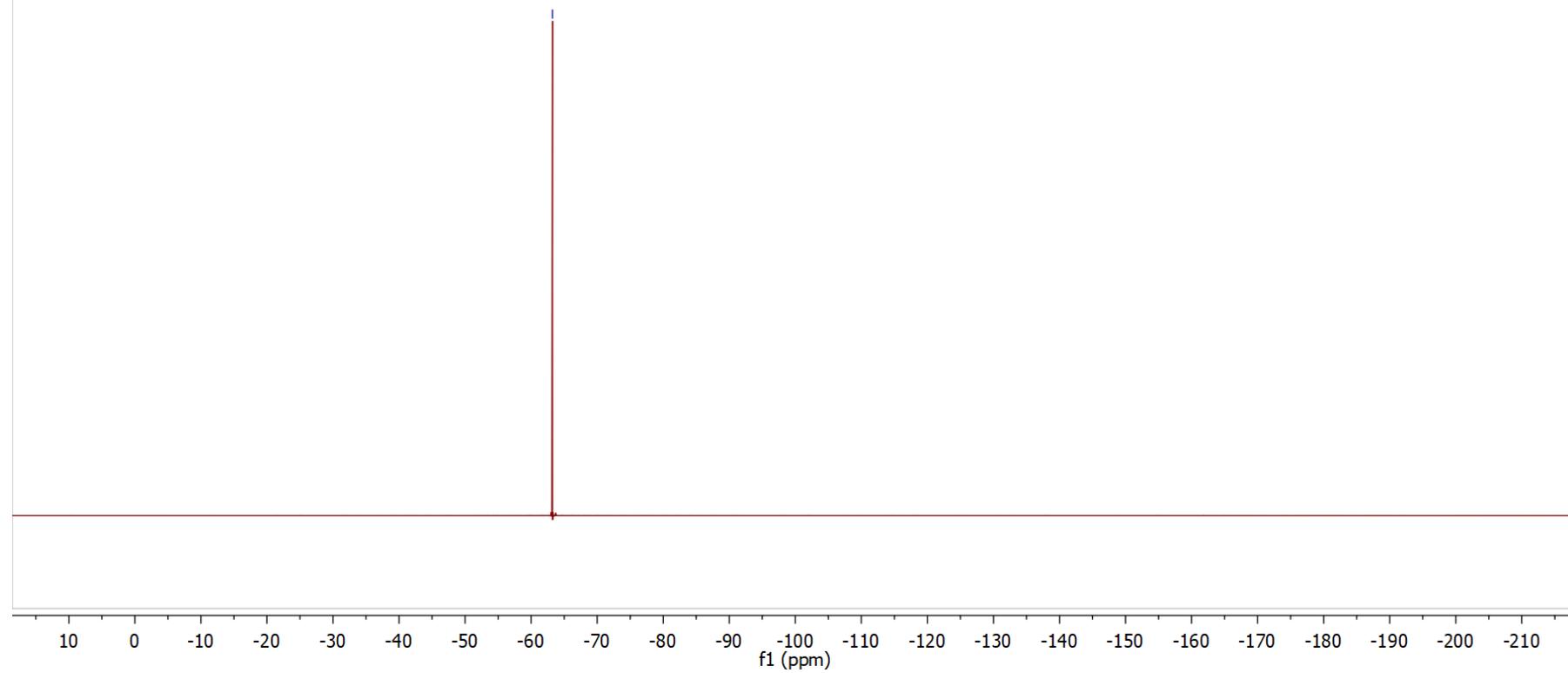


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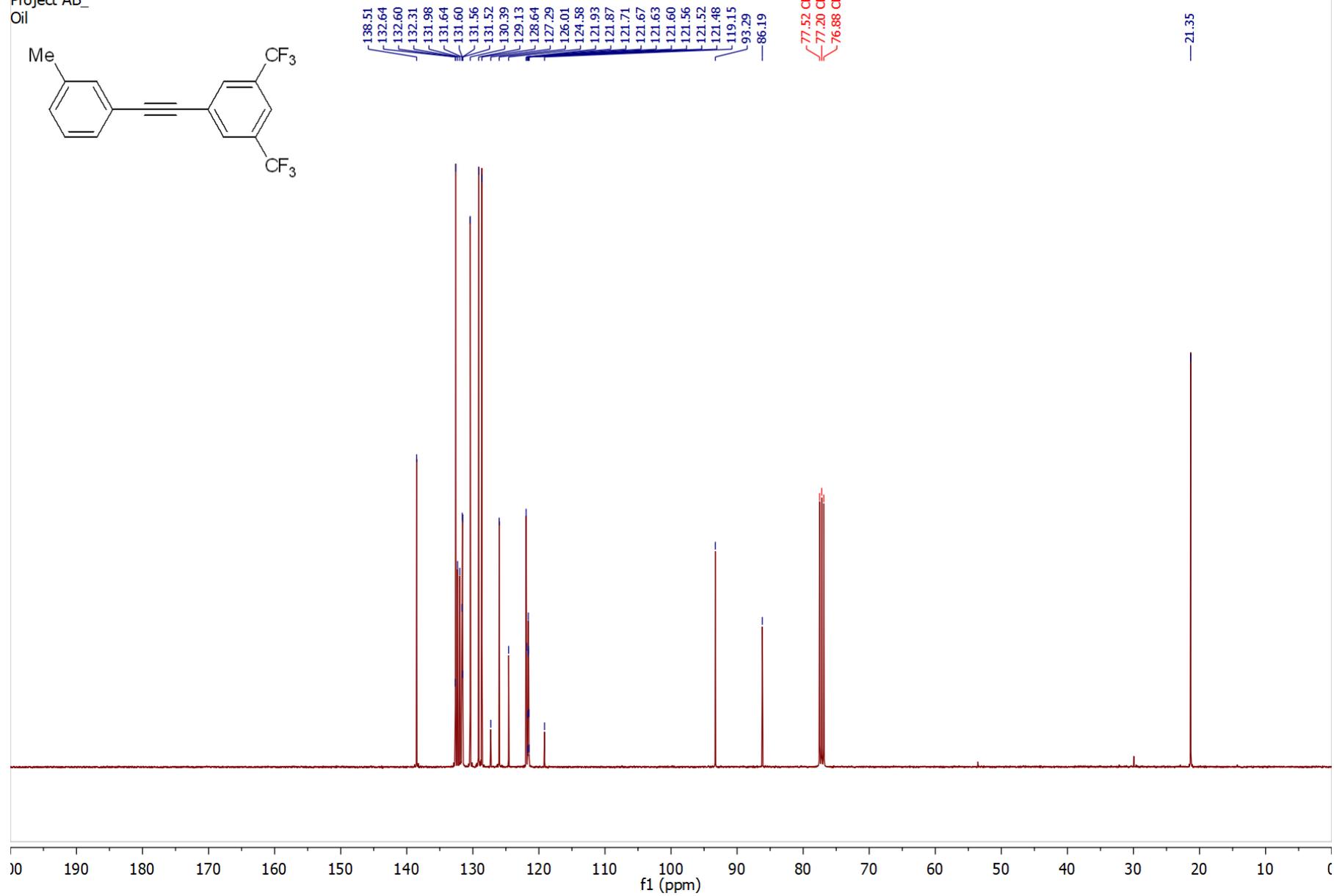
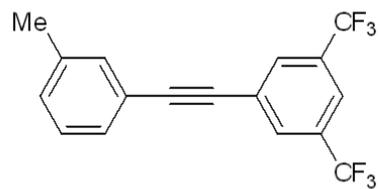


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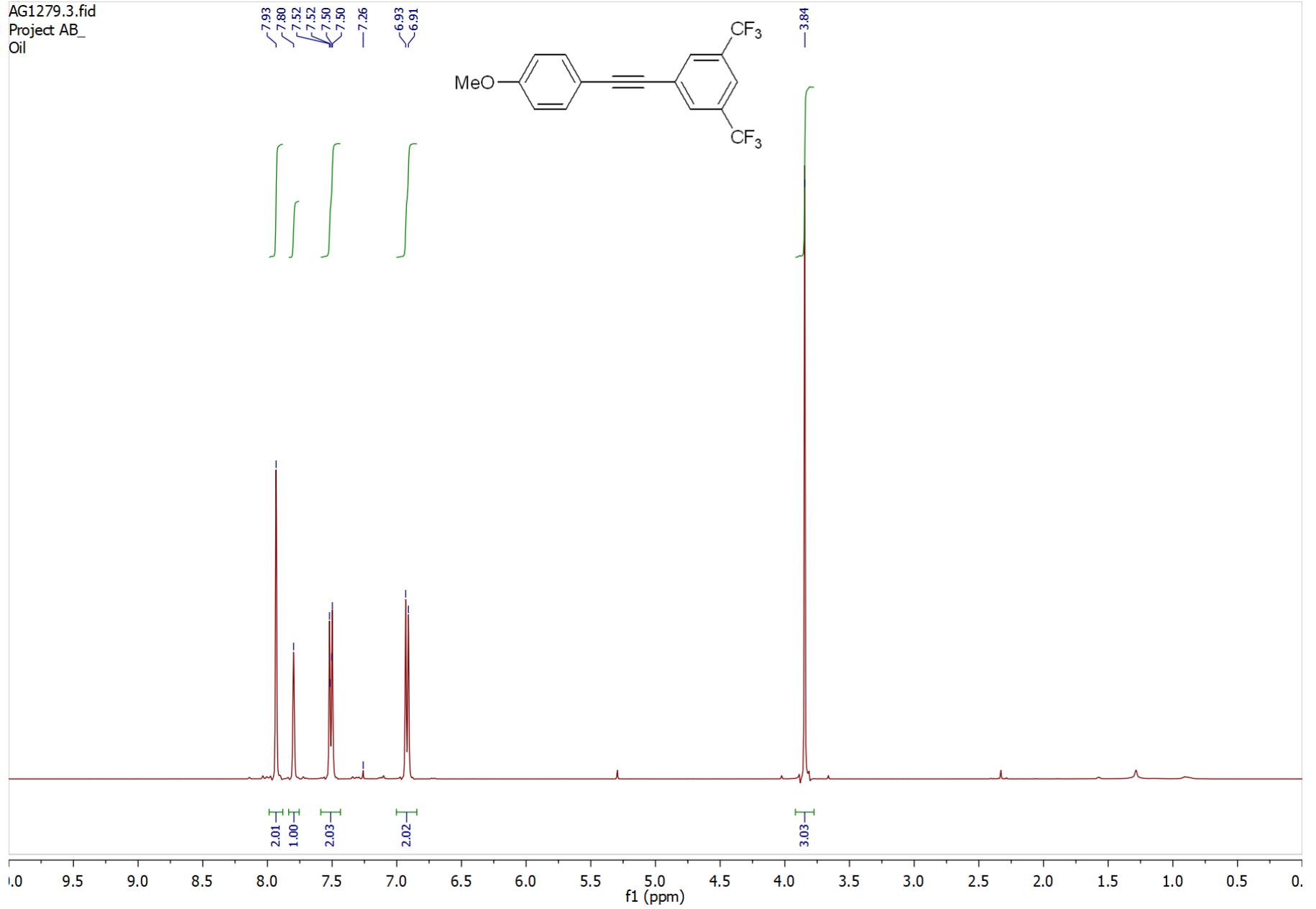
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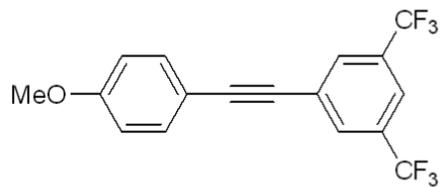
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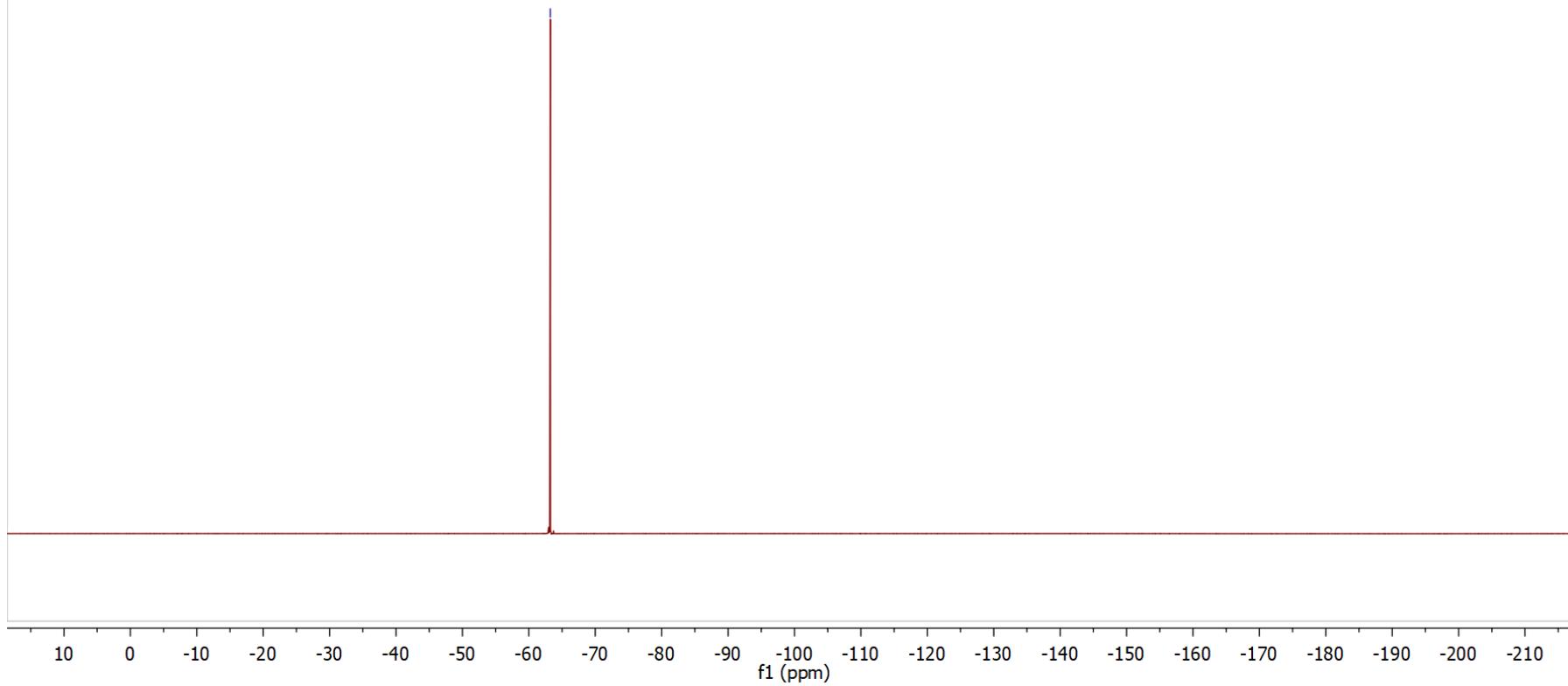


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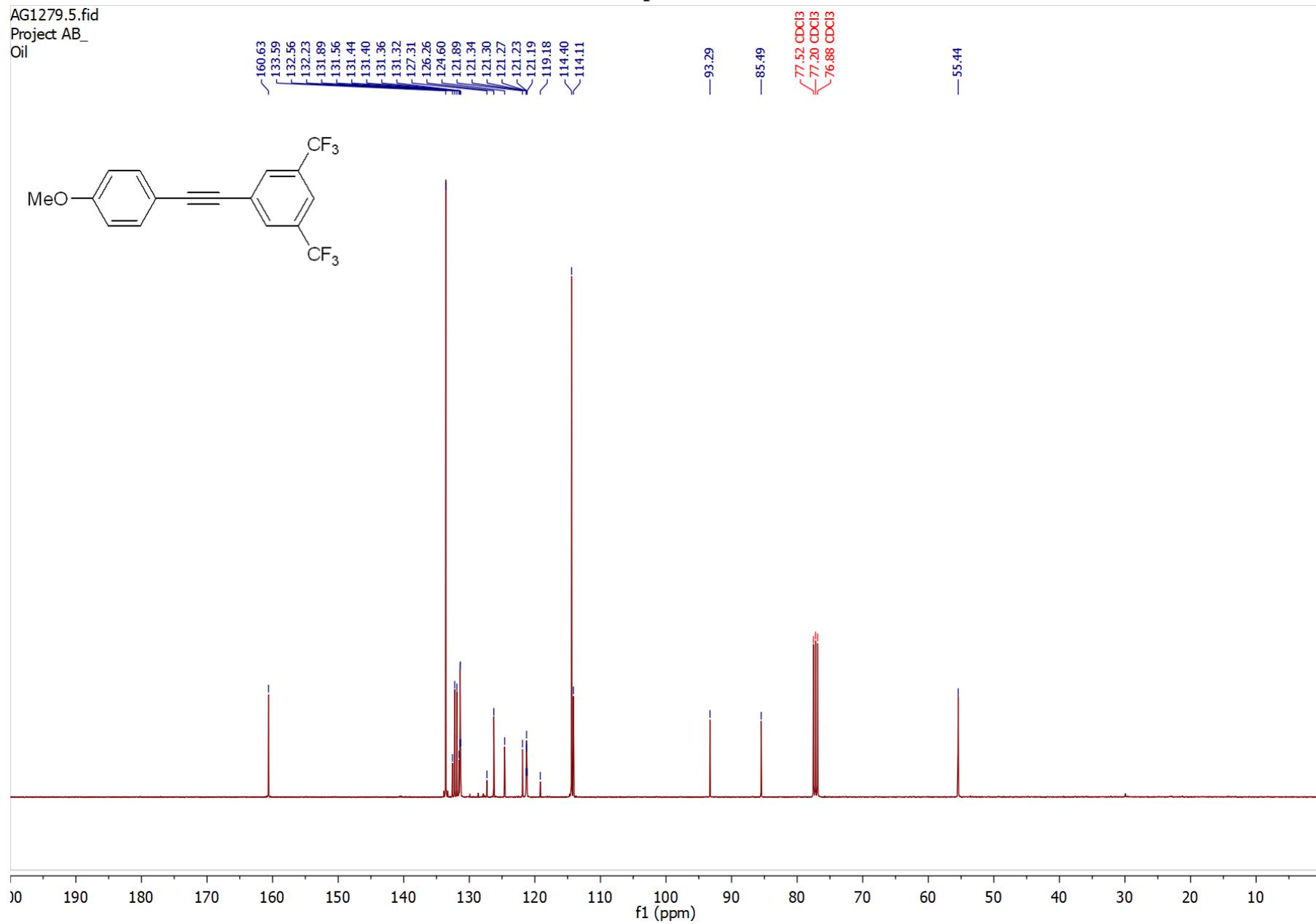


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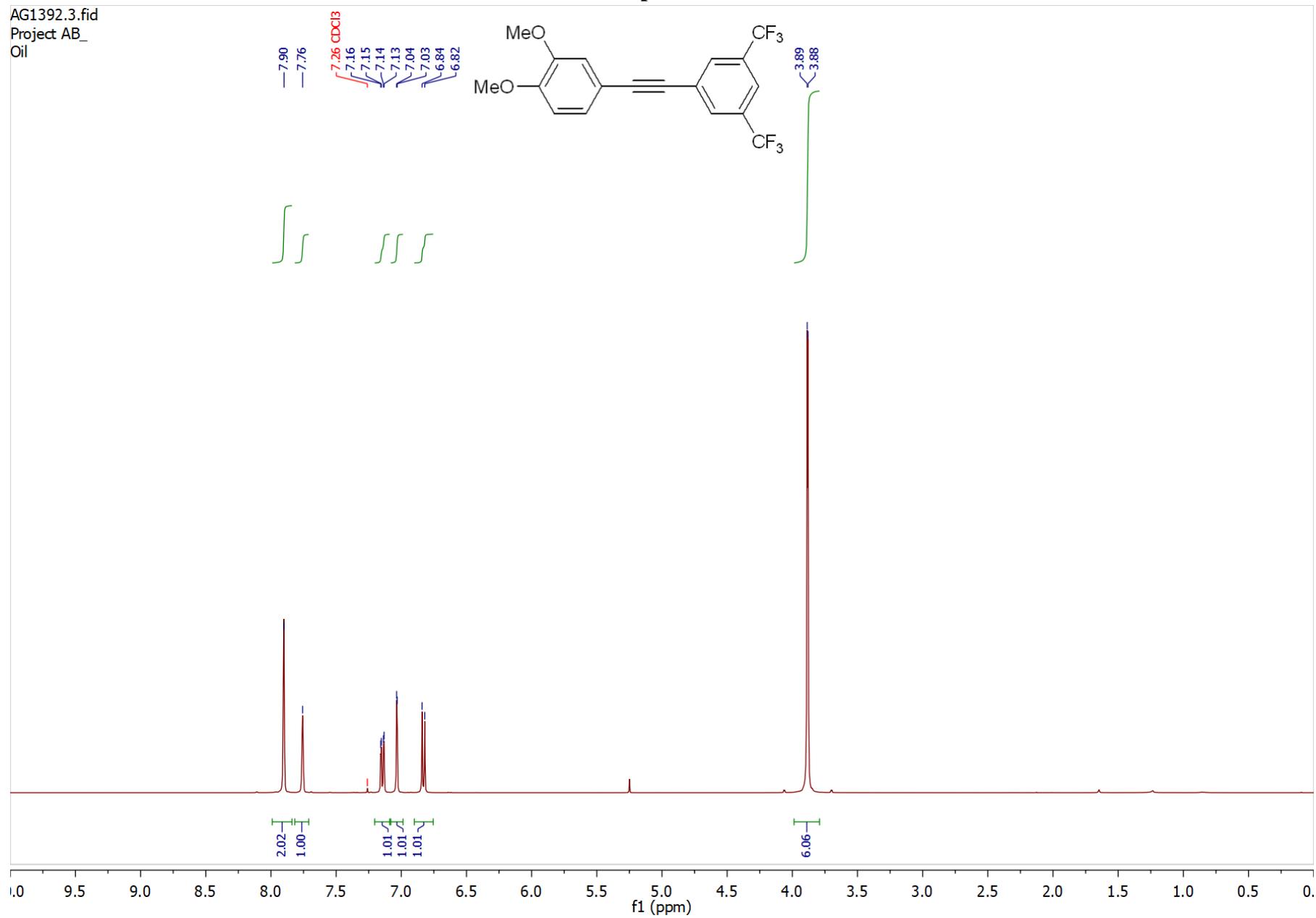
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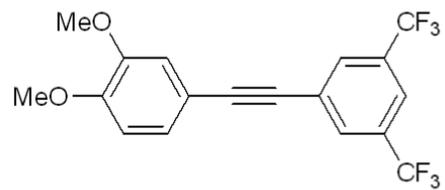
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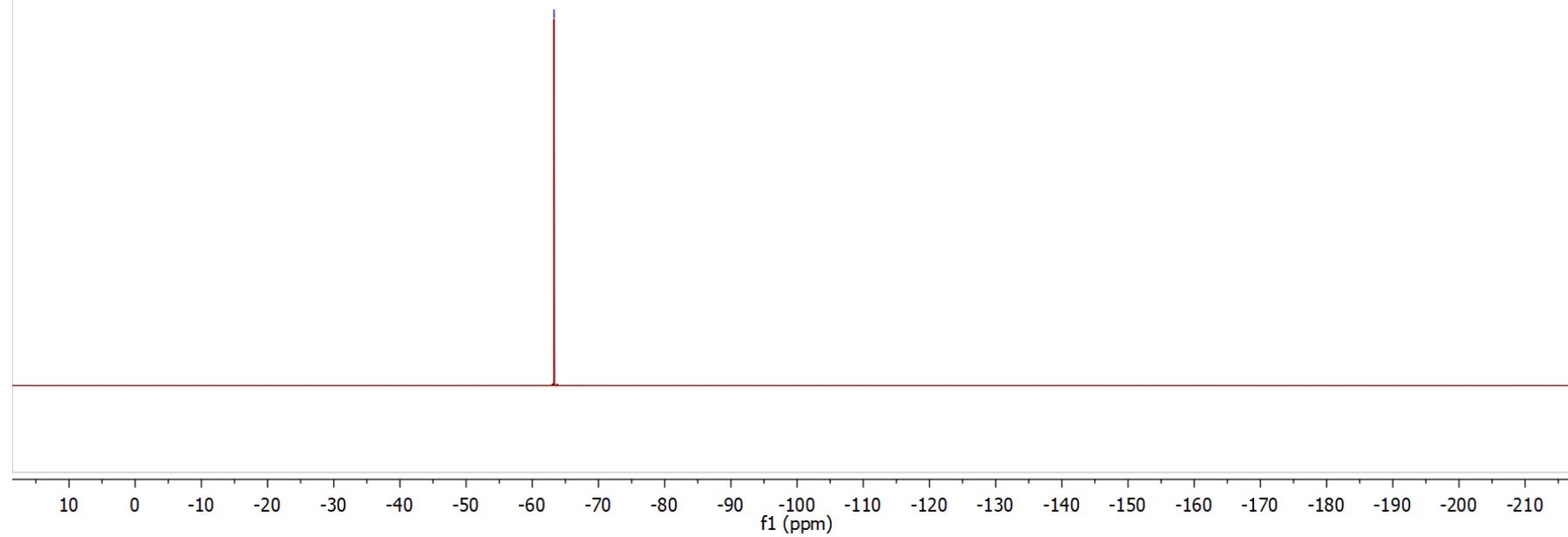


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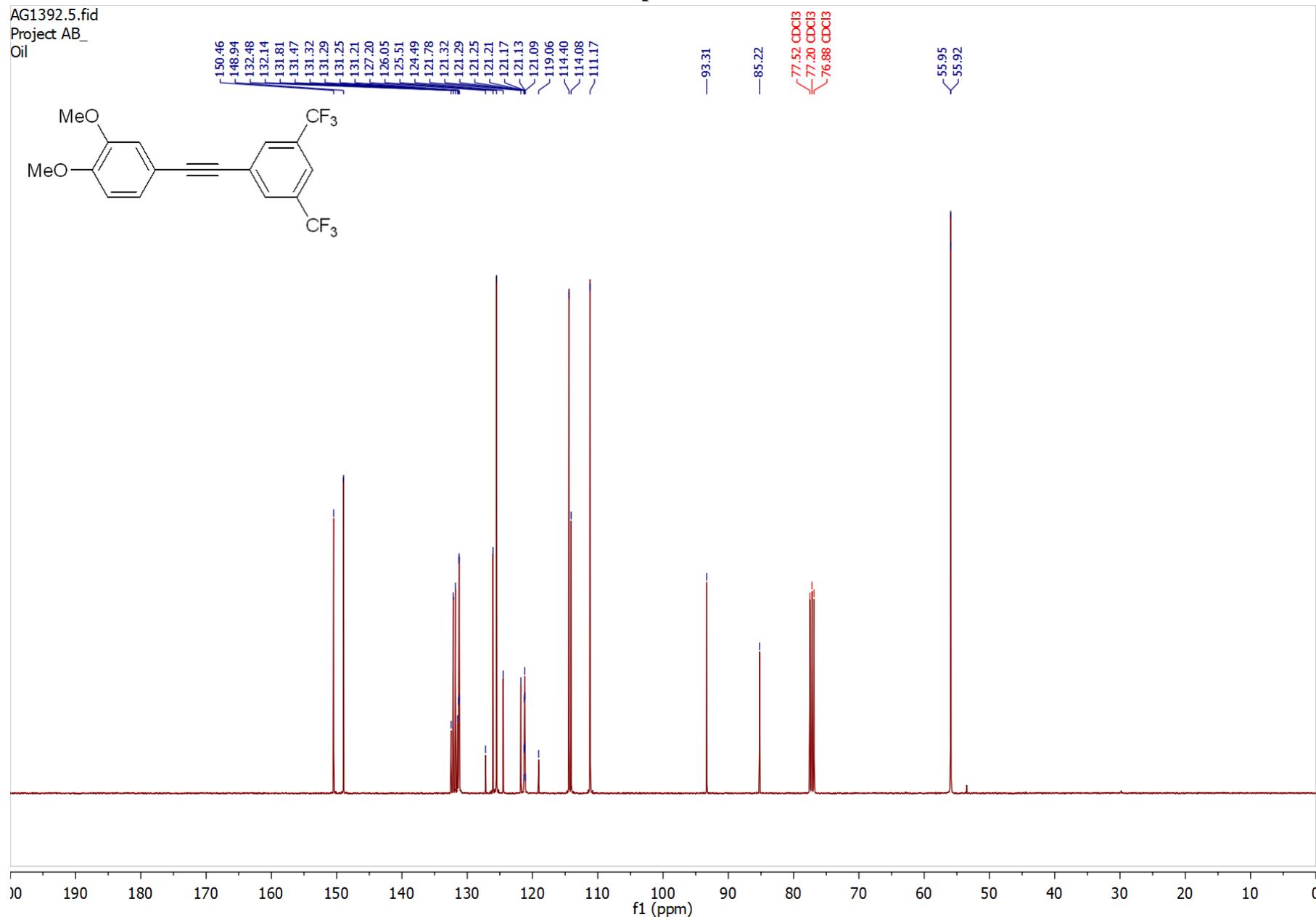


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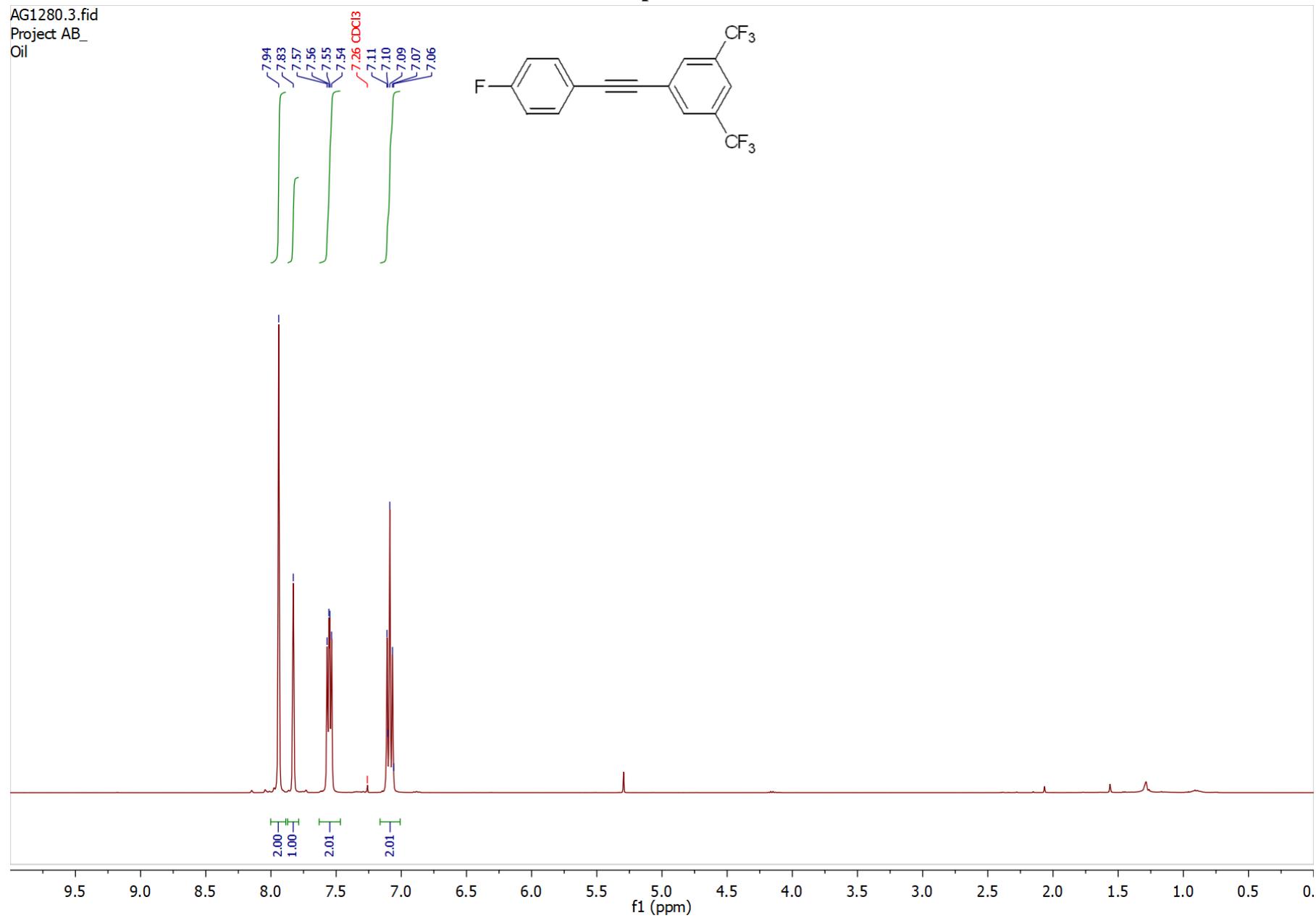
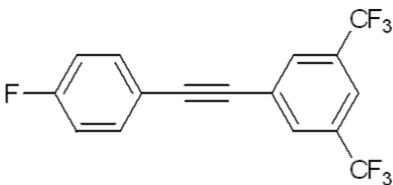
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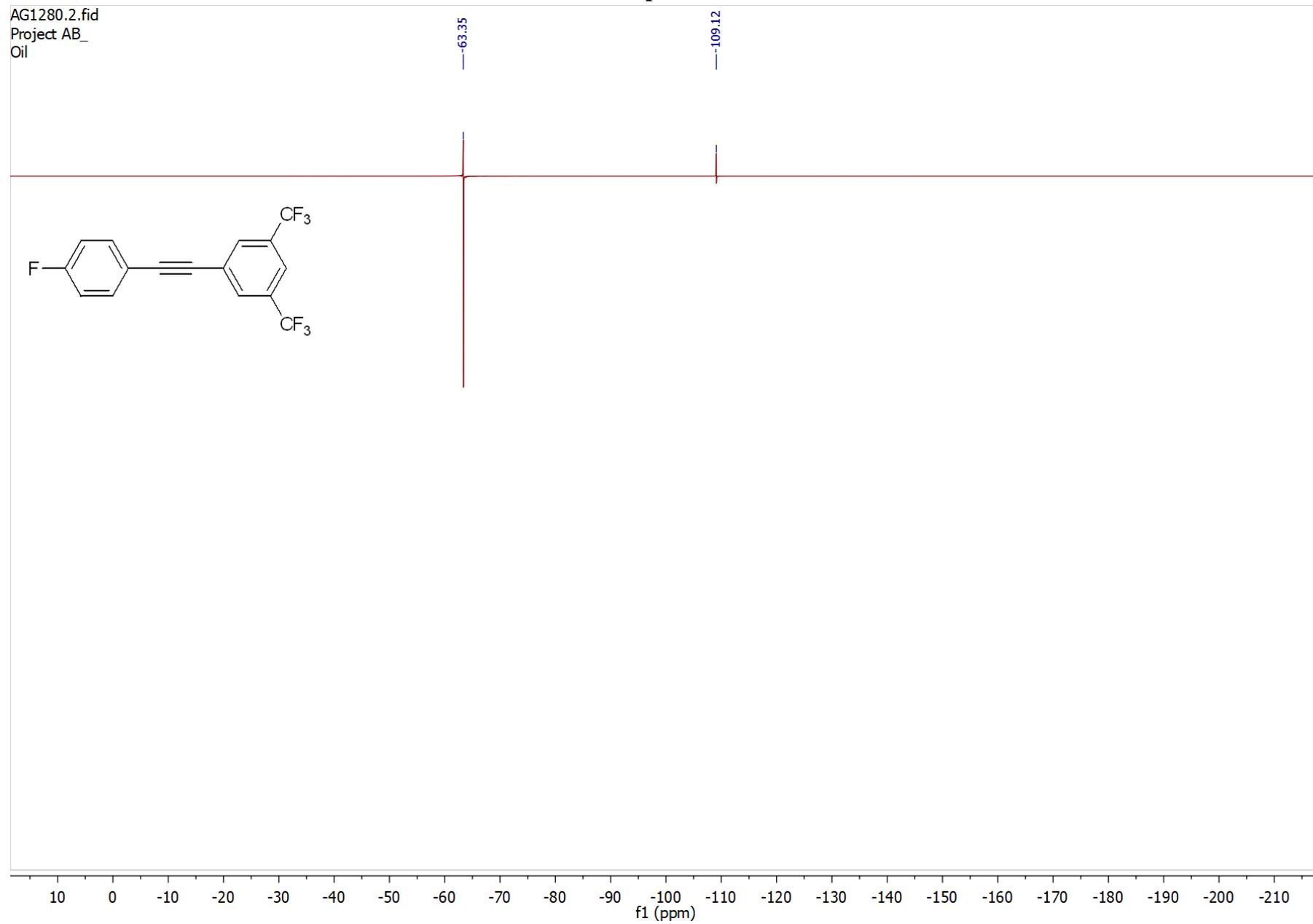
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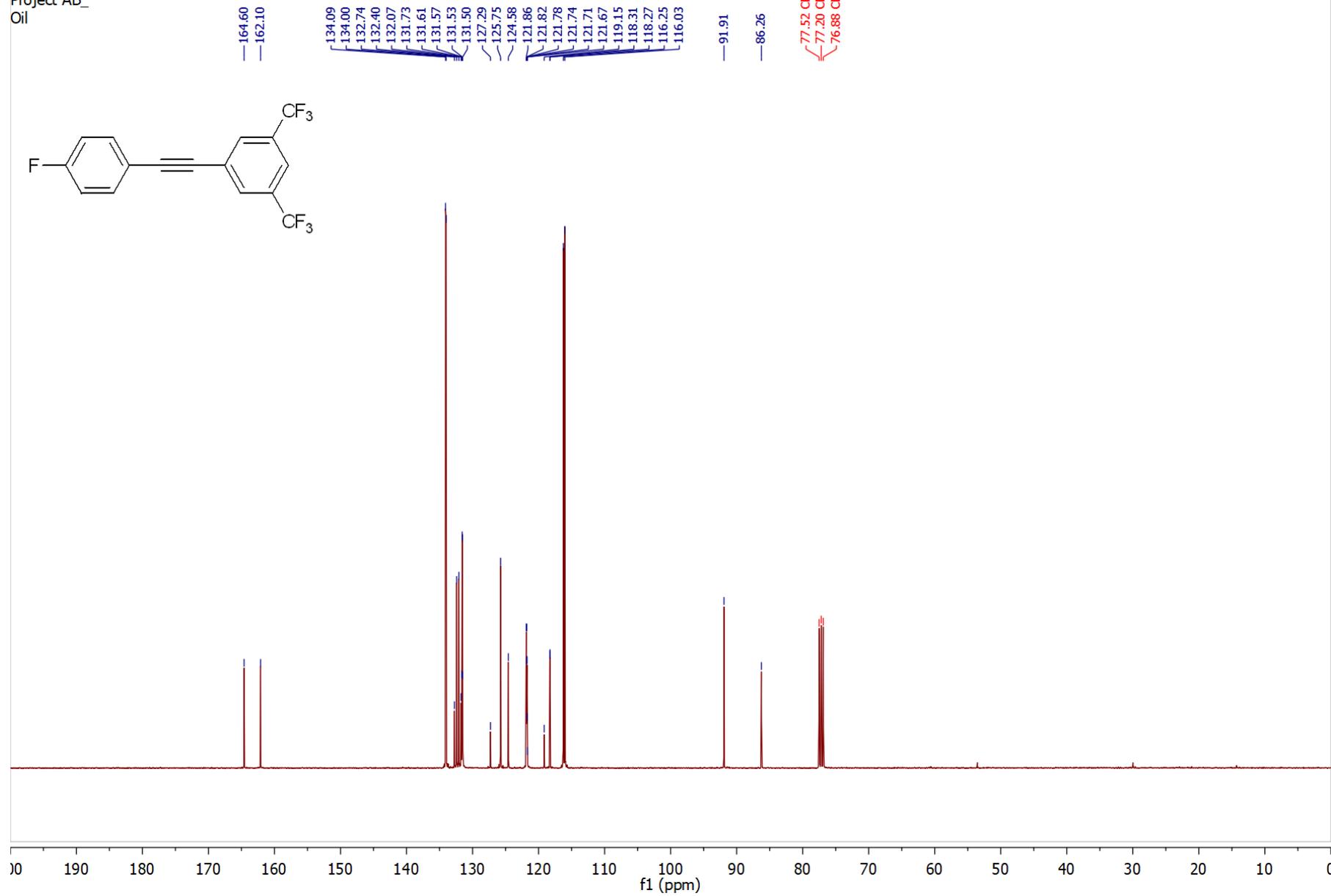
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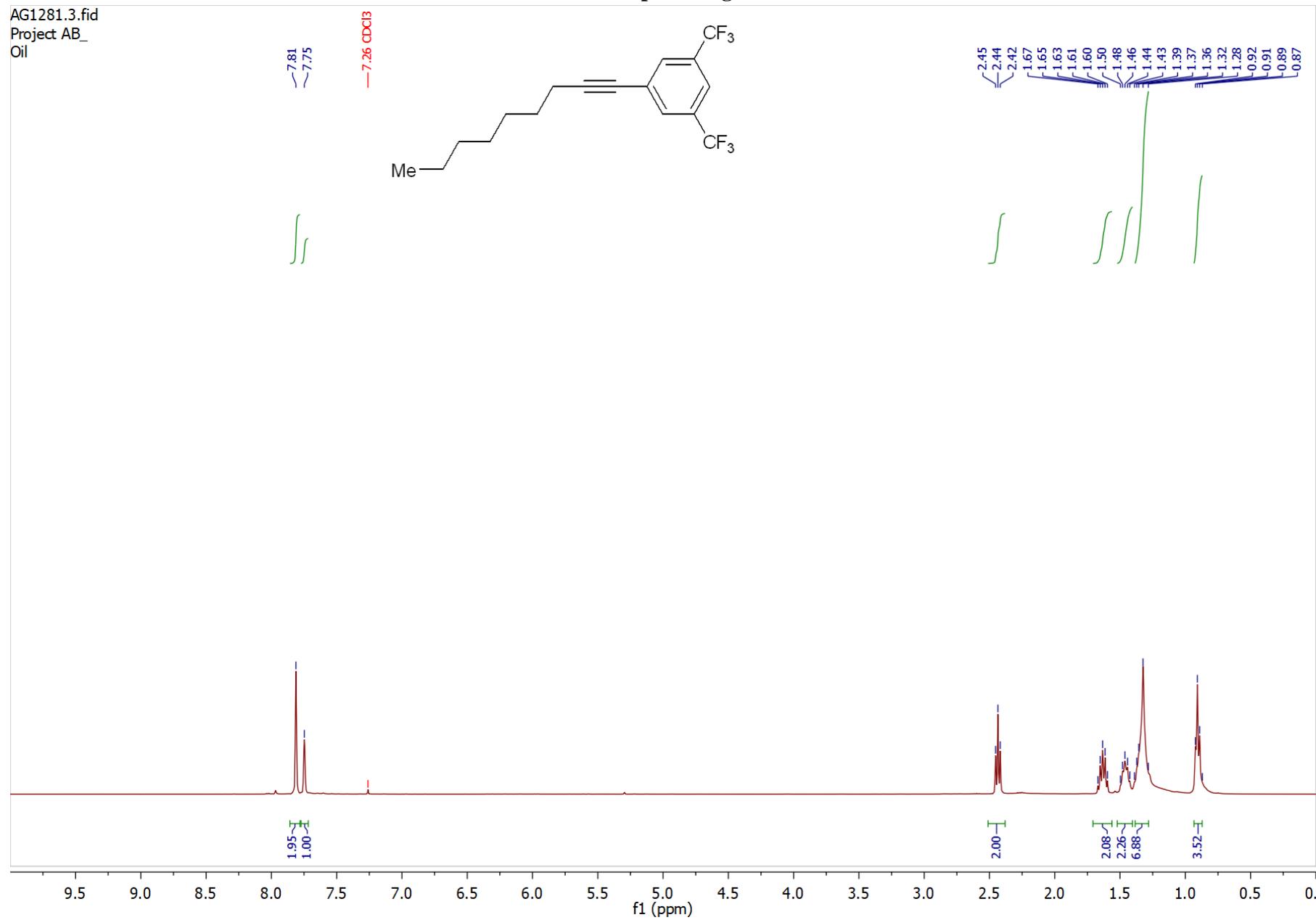
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Oil



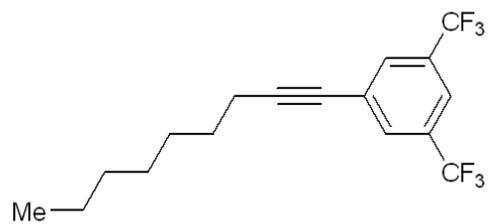
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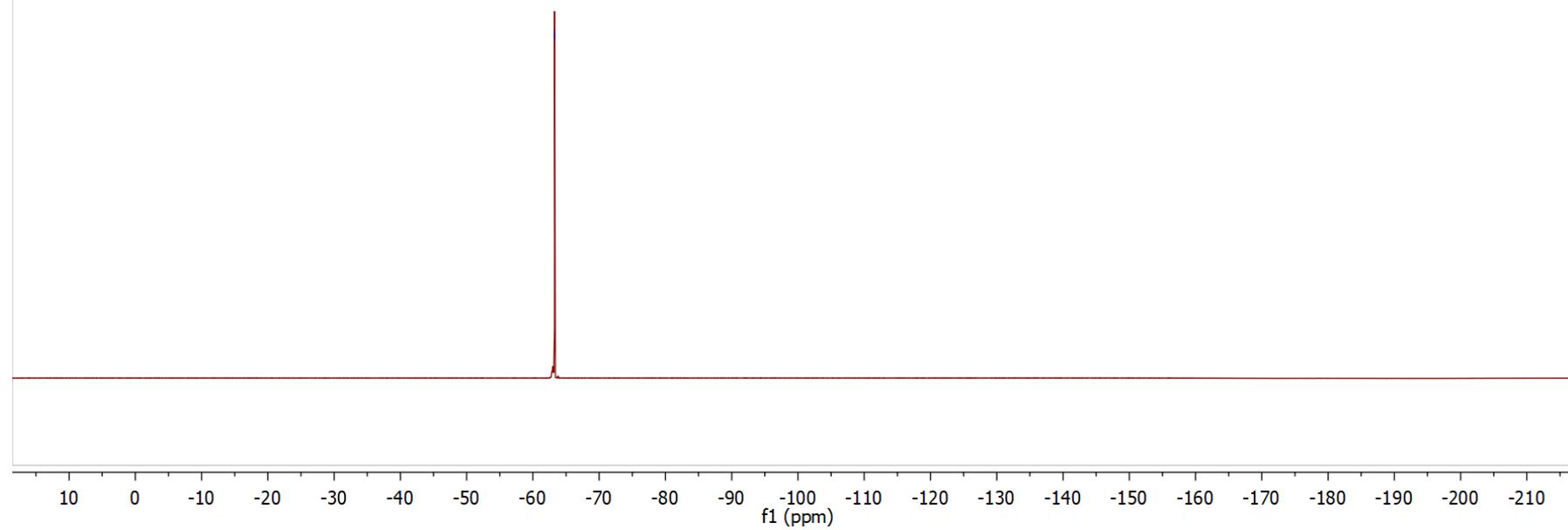


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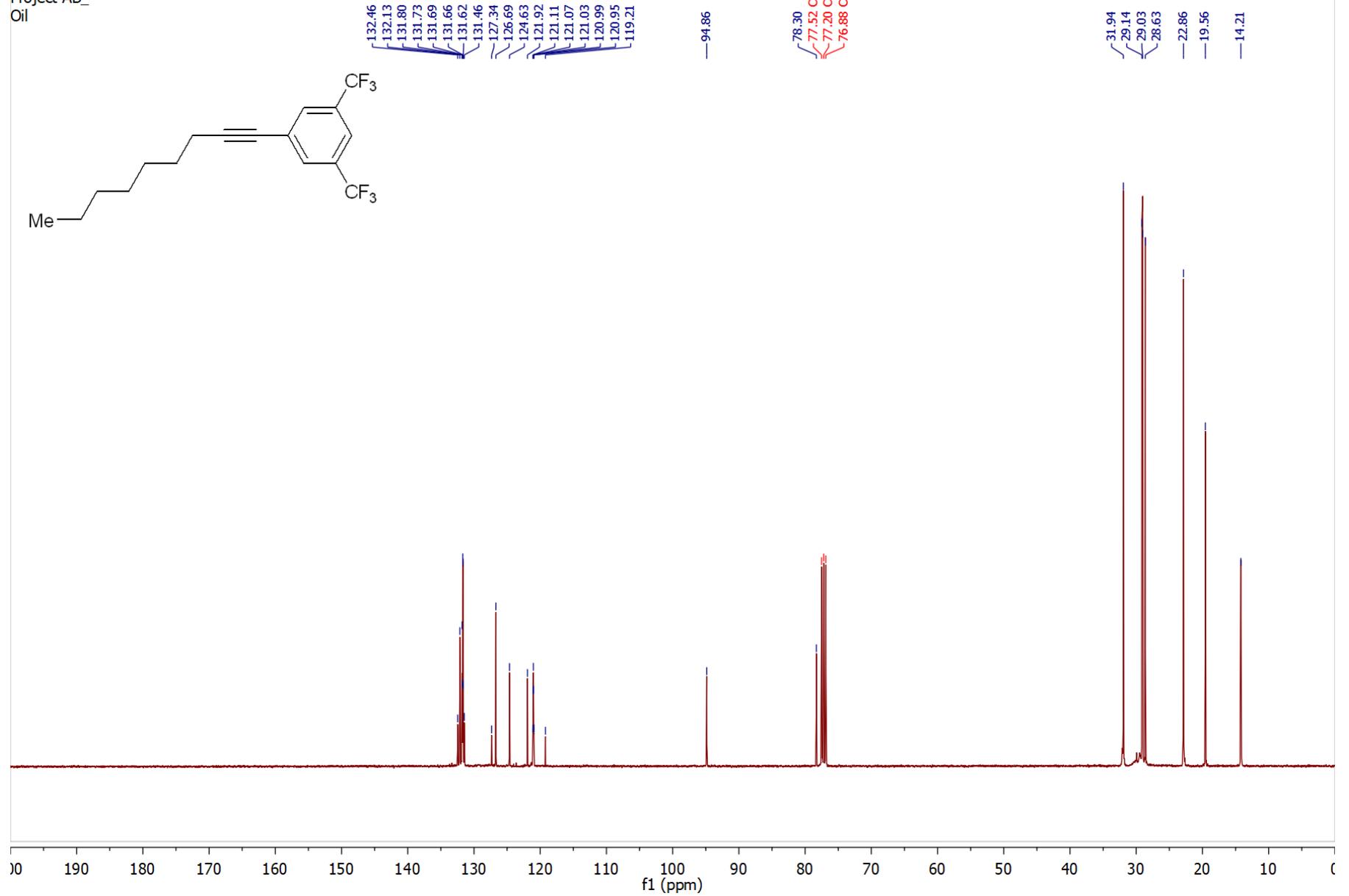


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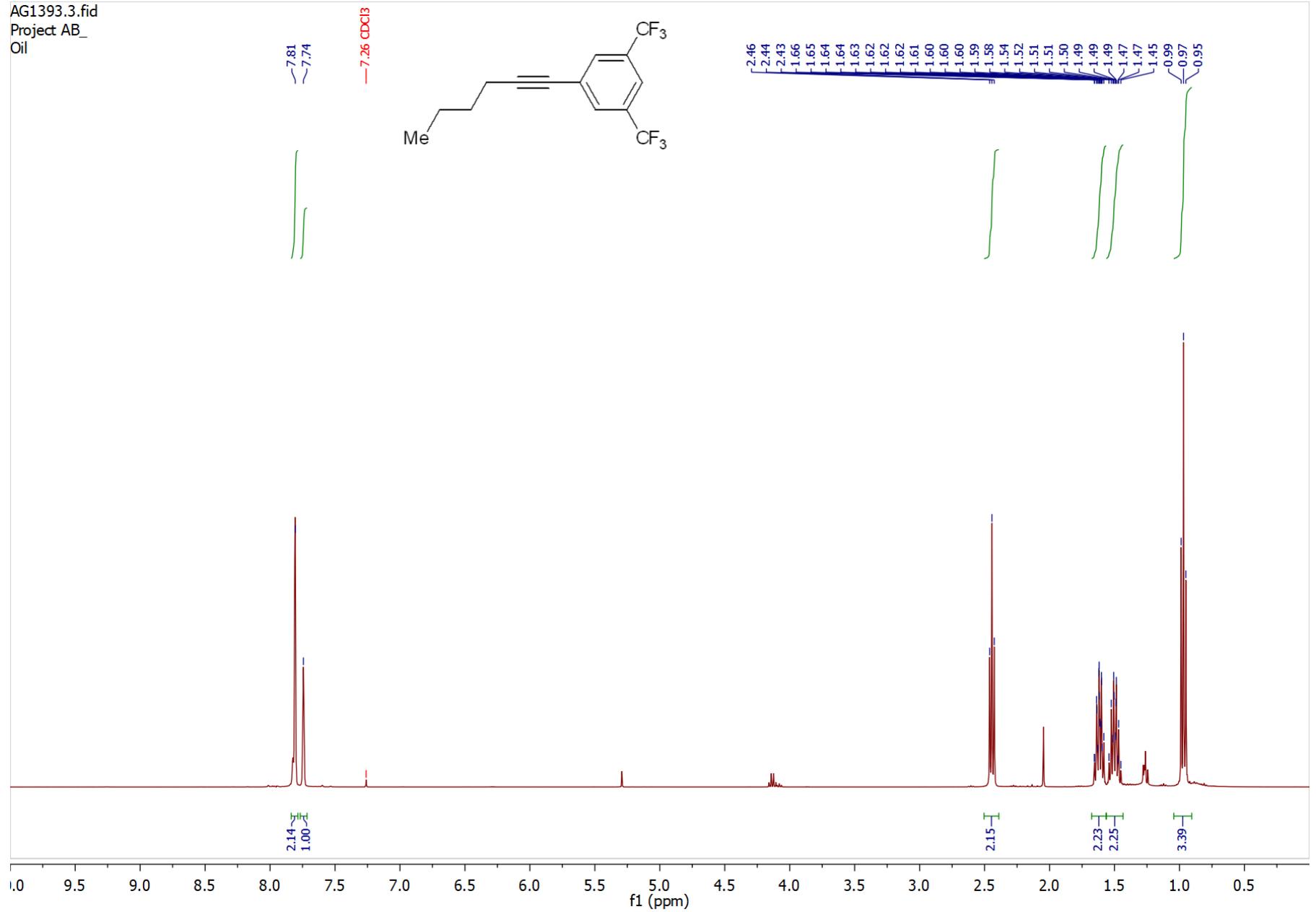
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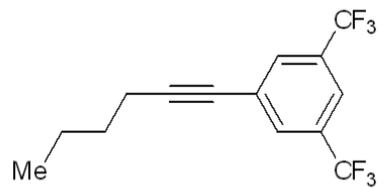
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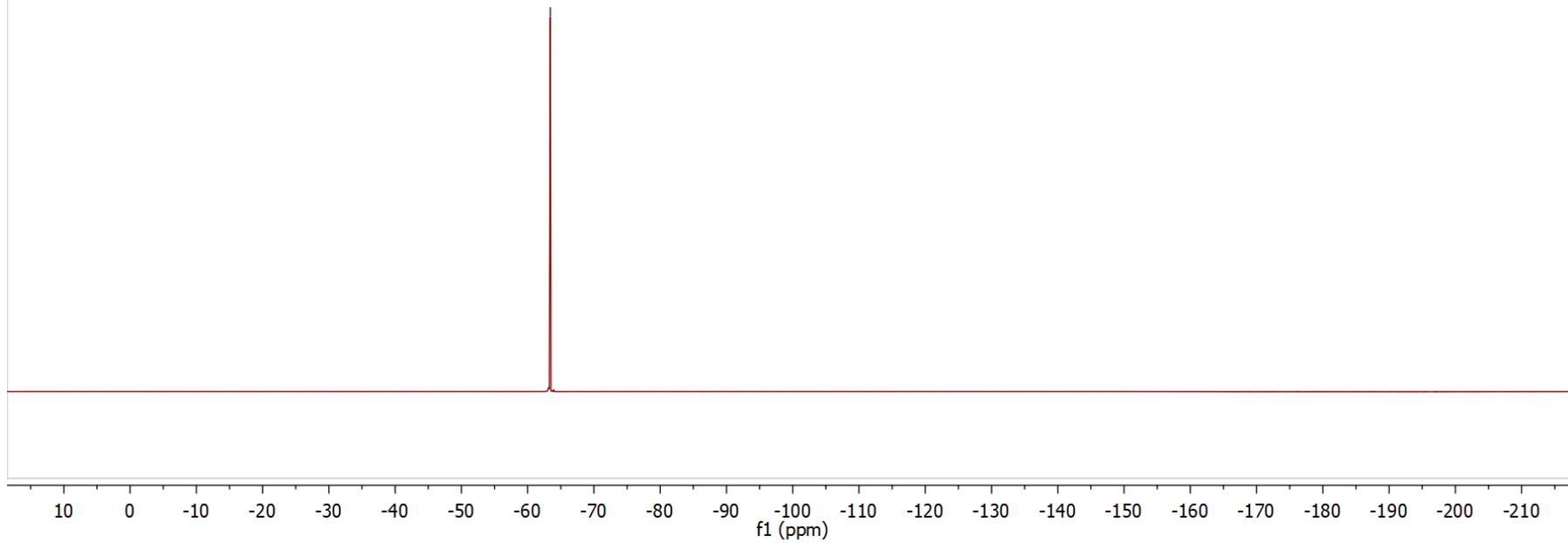


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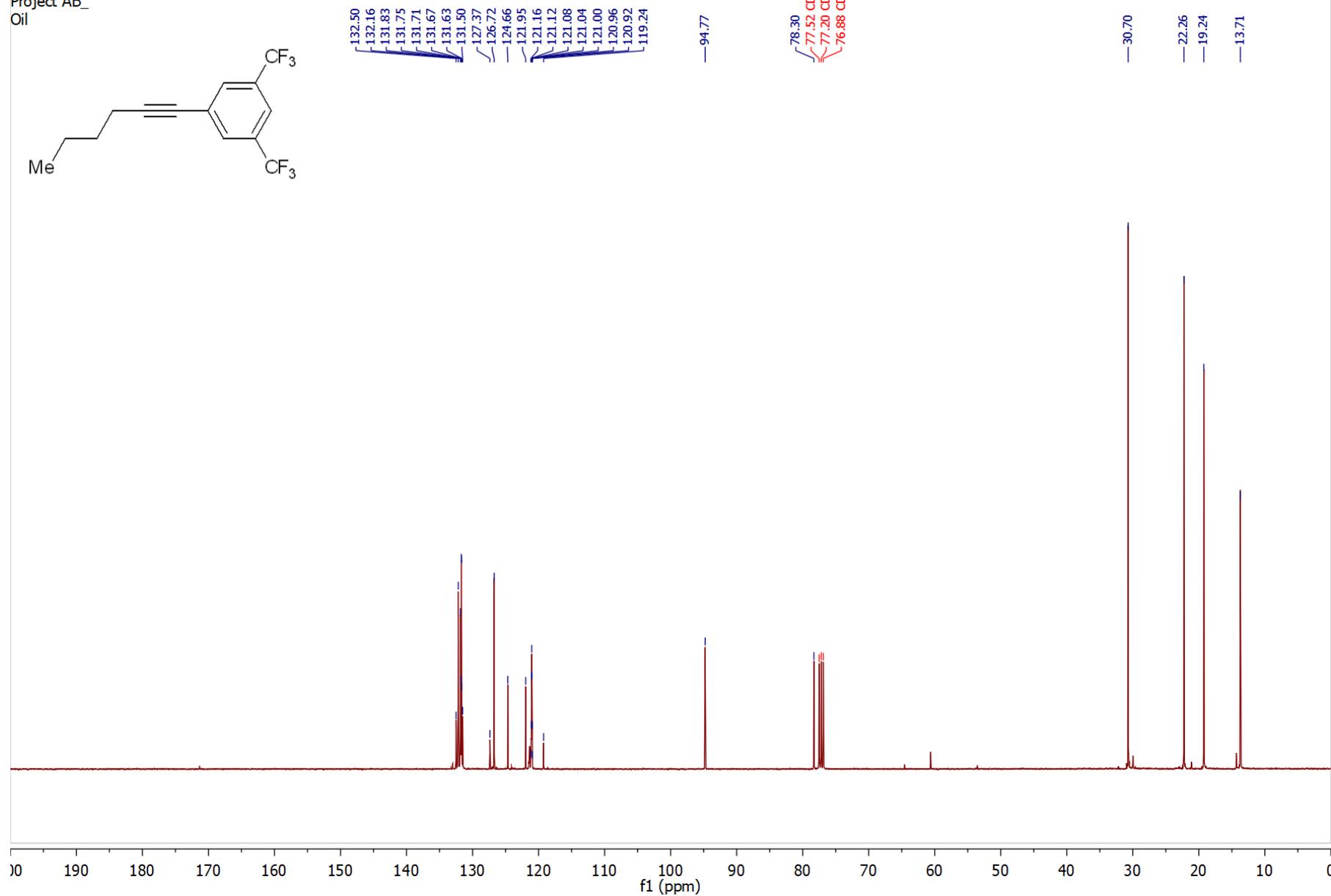
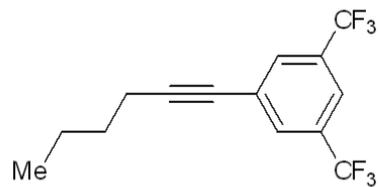


63.42



Compound 9h

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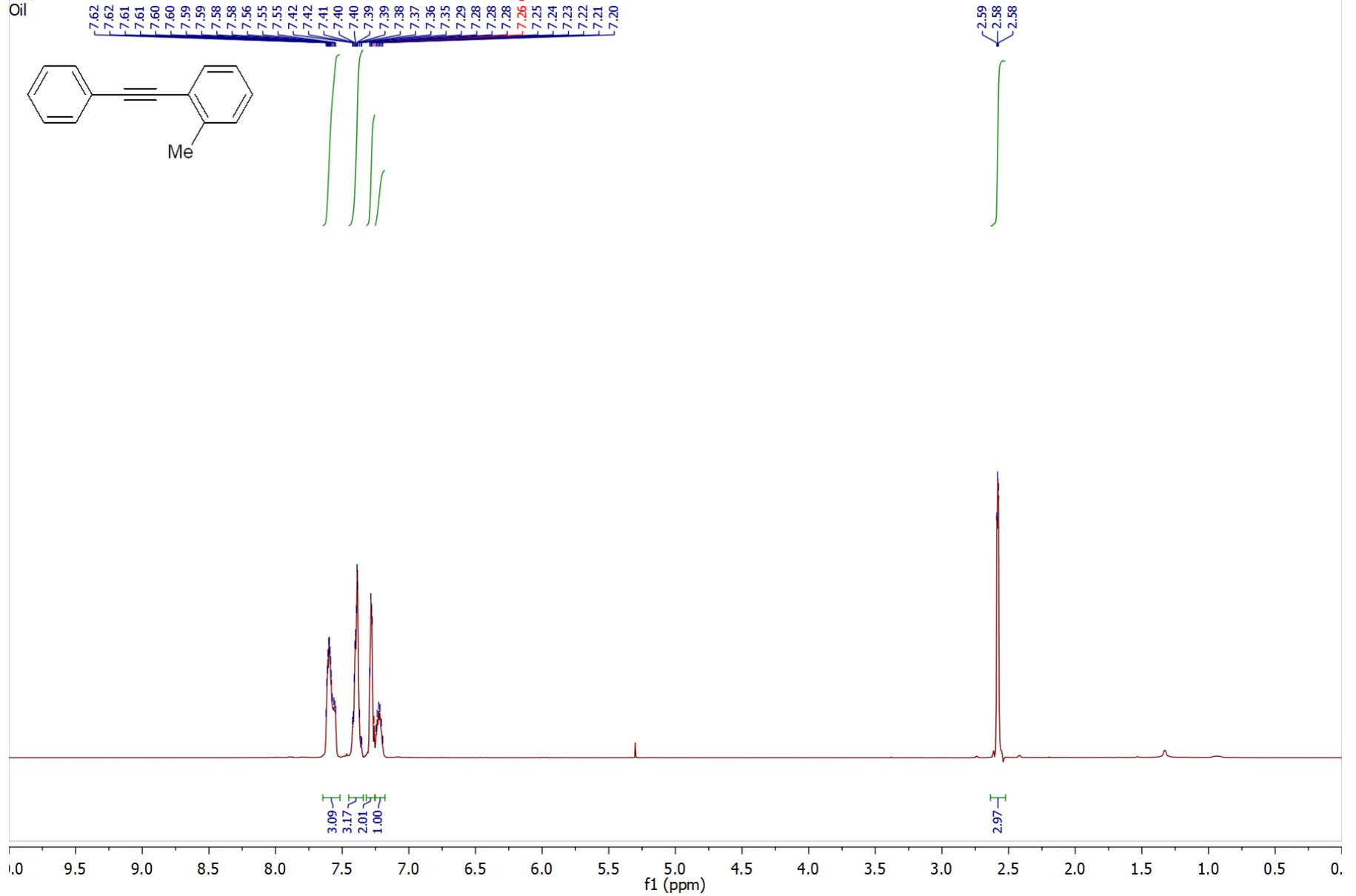


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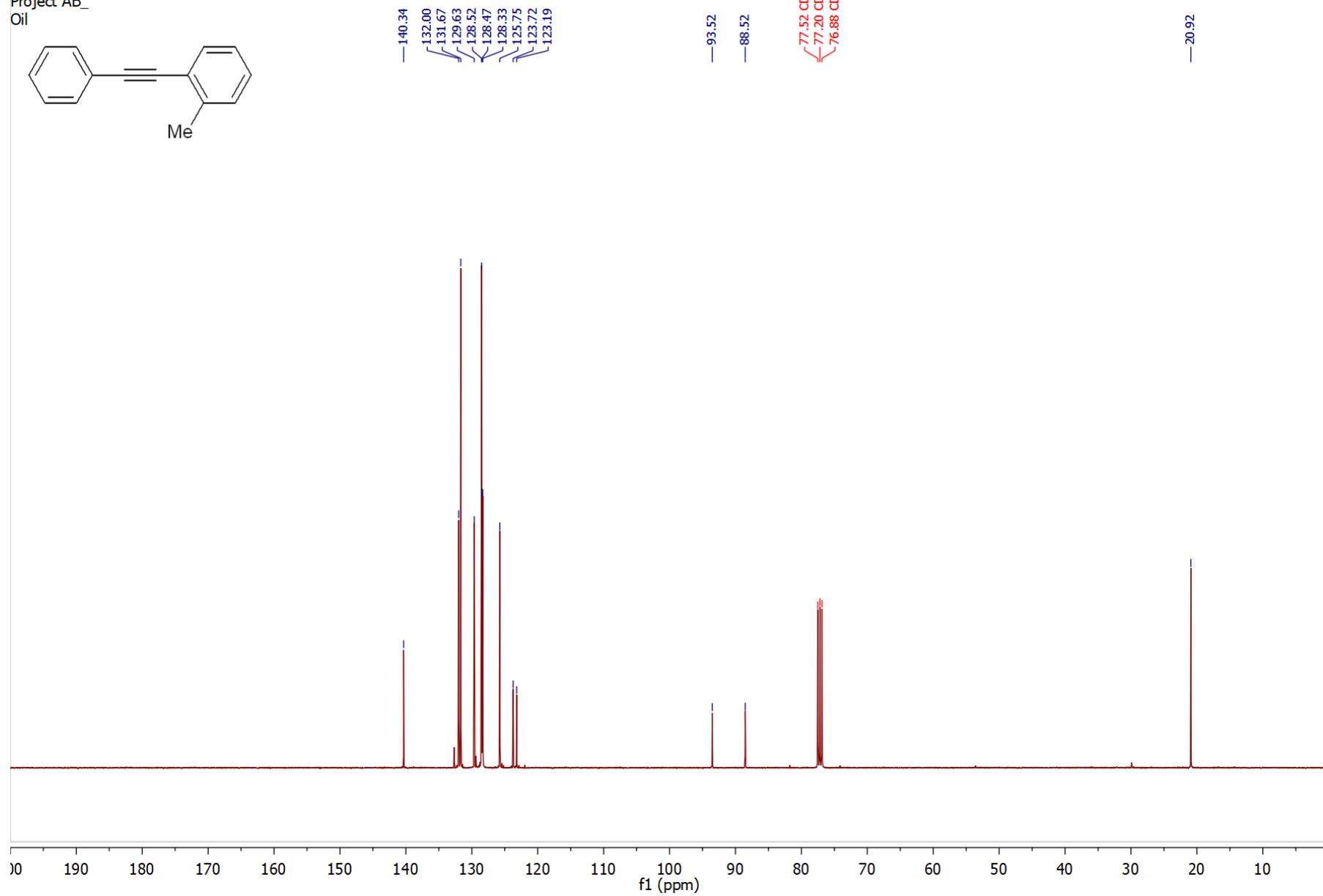
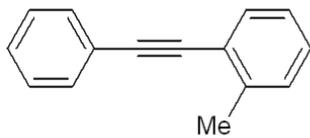
Project AB_

Oil



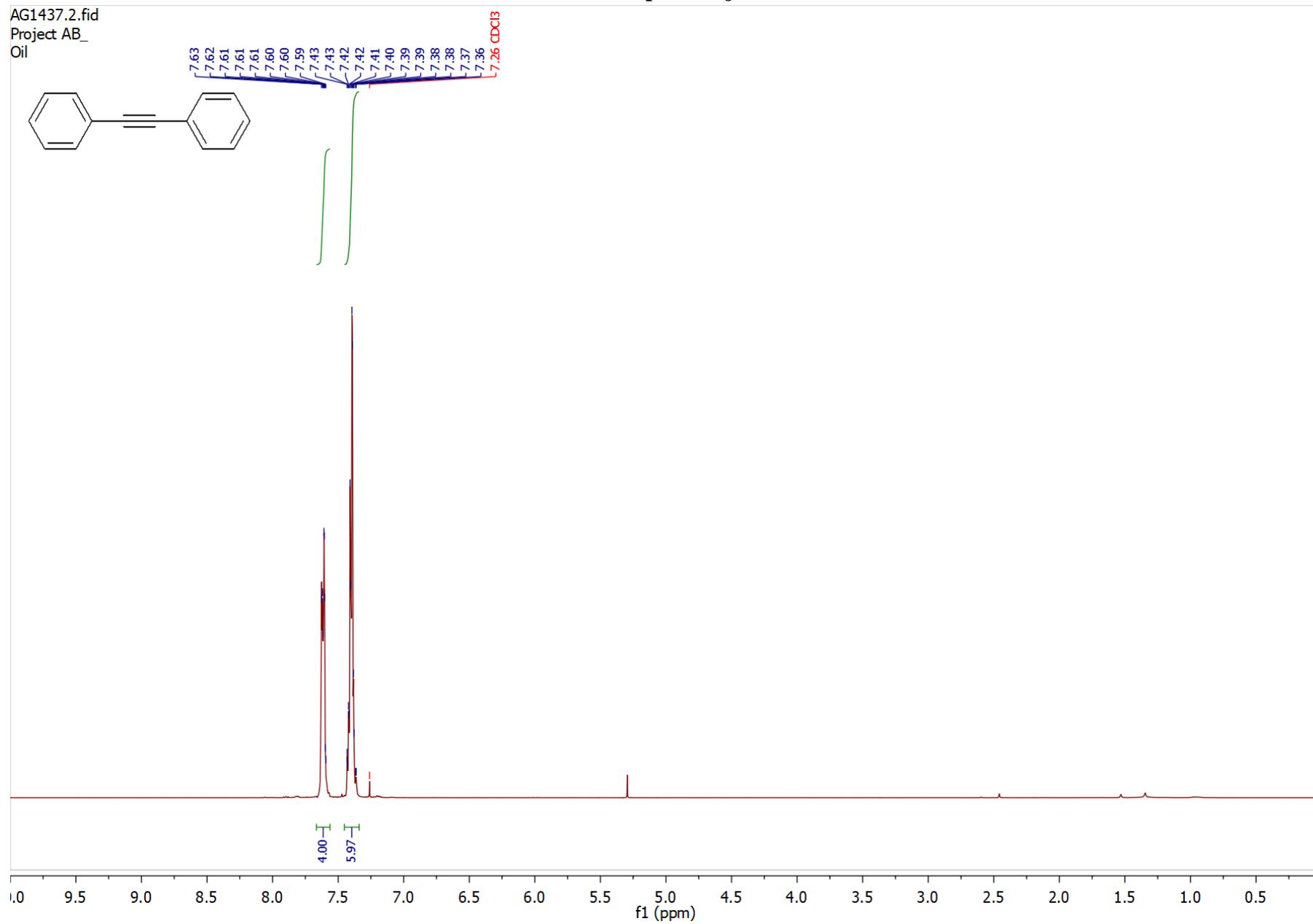
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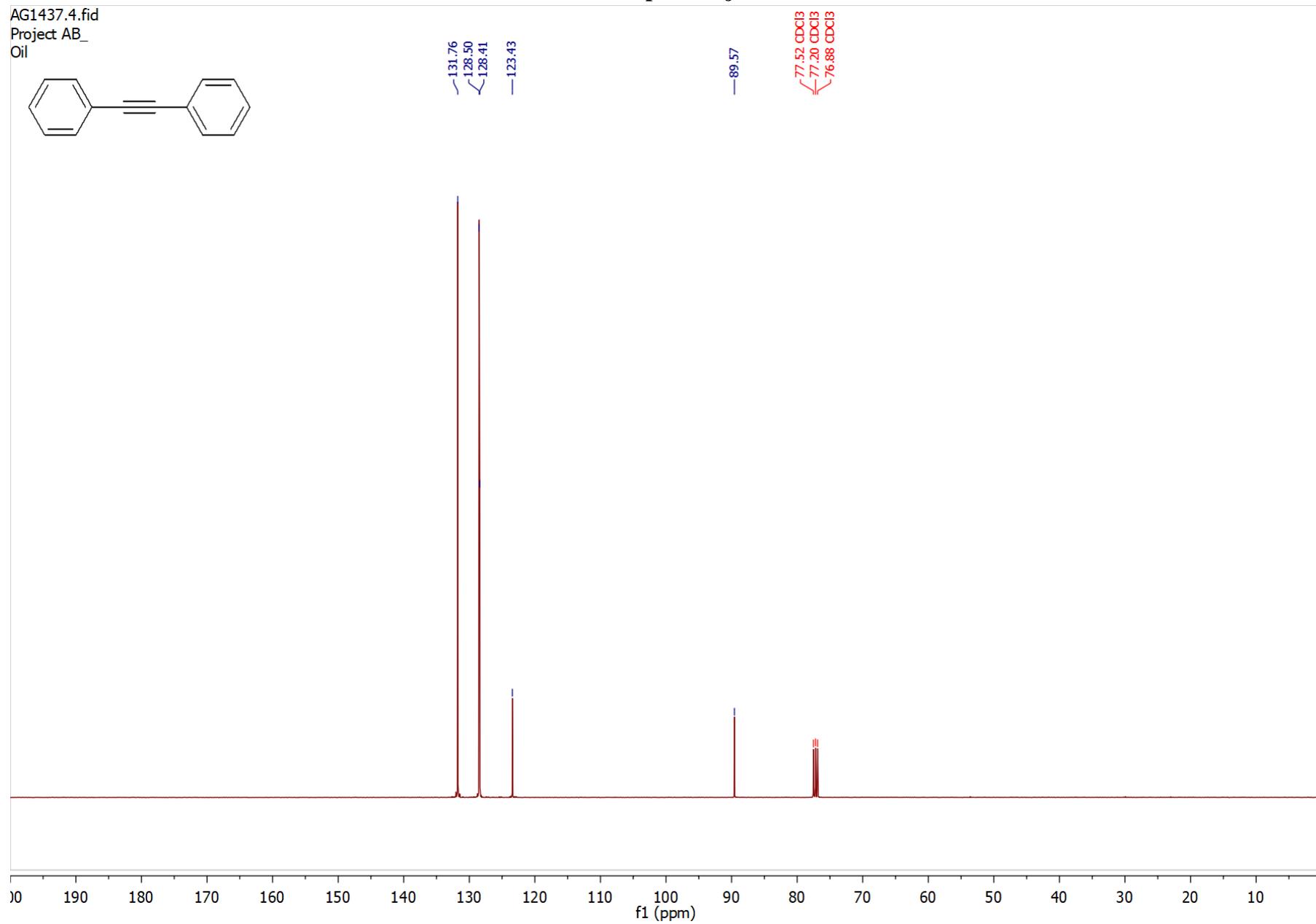
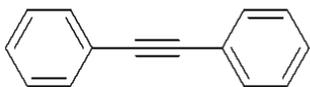
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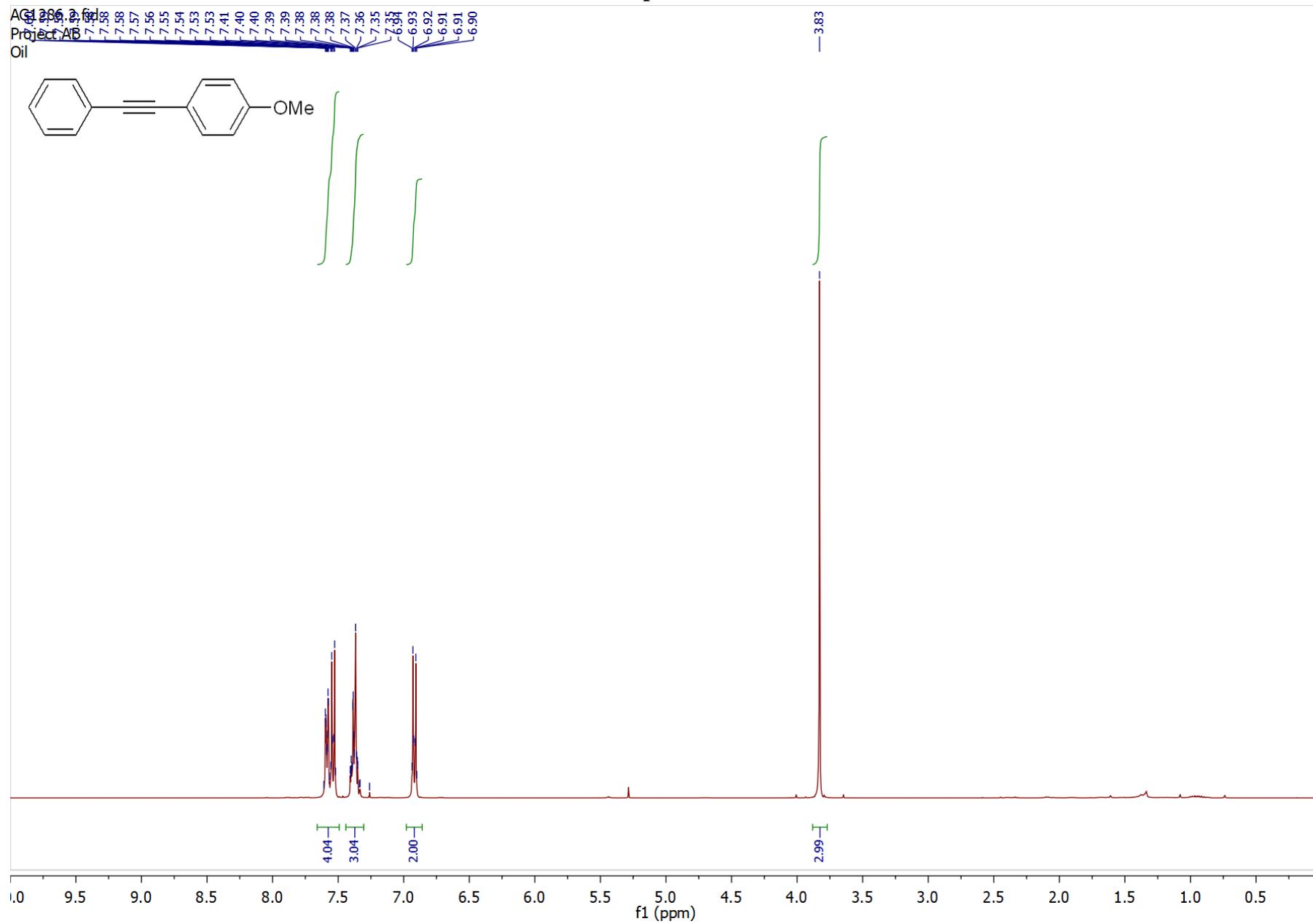


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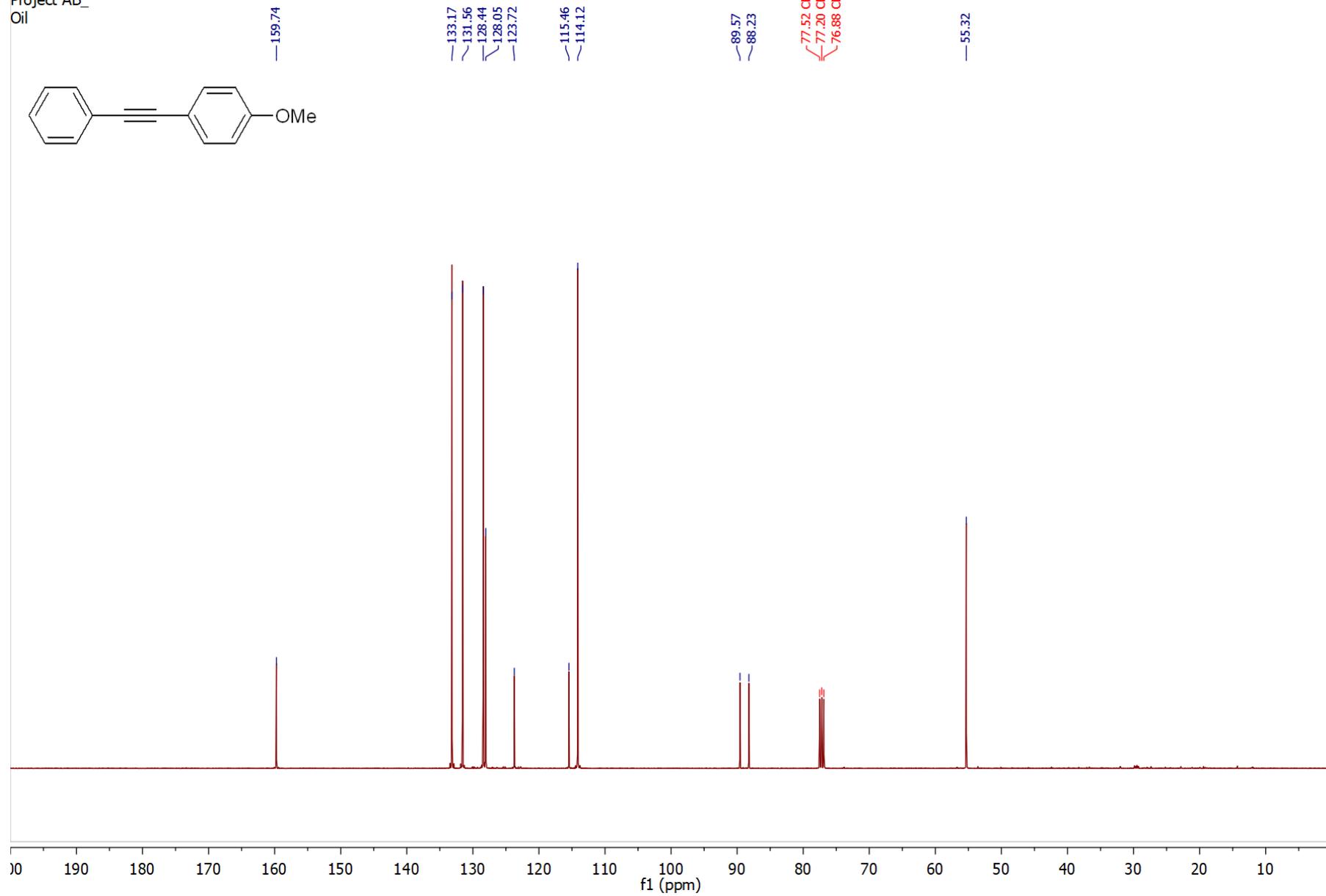


Compound 9k



Compound 9k

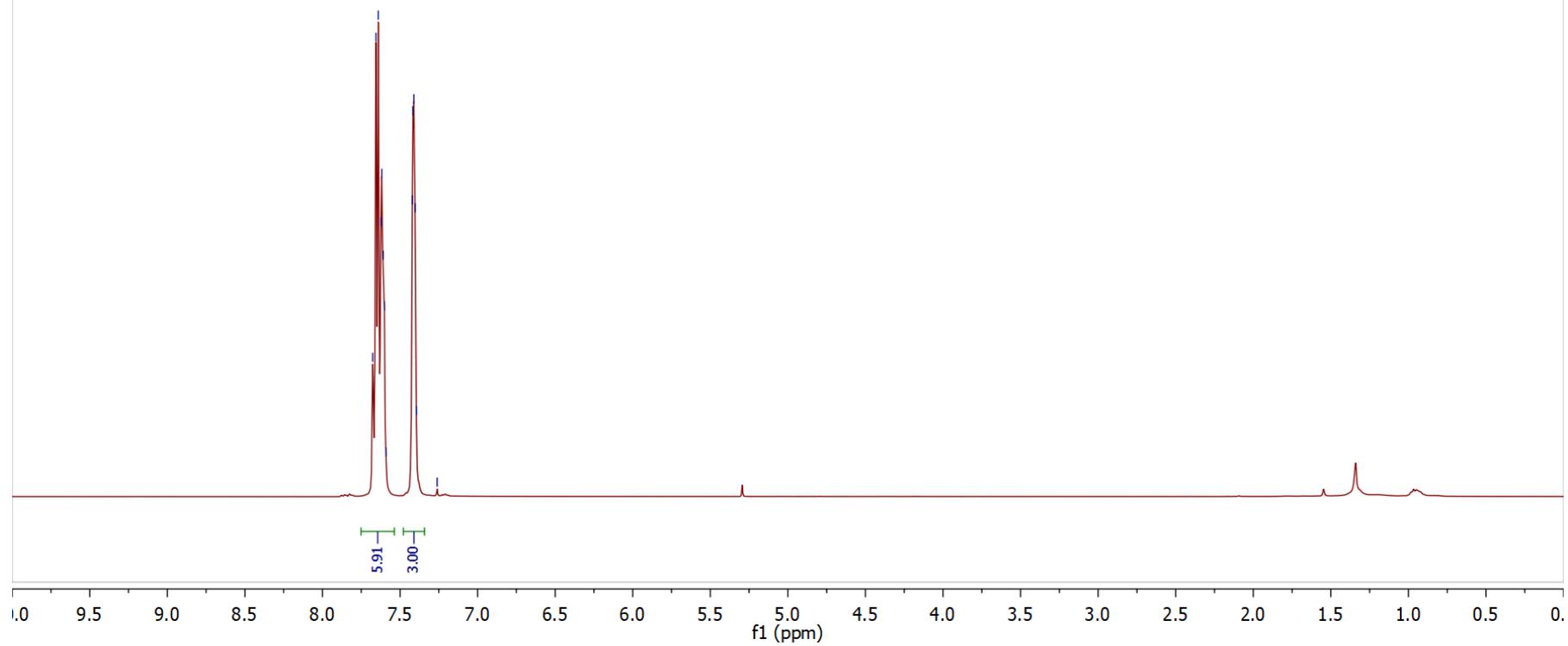
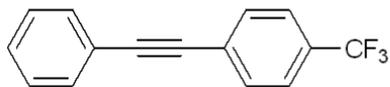
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Compound 91

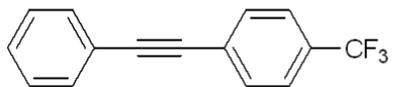
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7.26

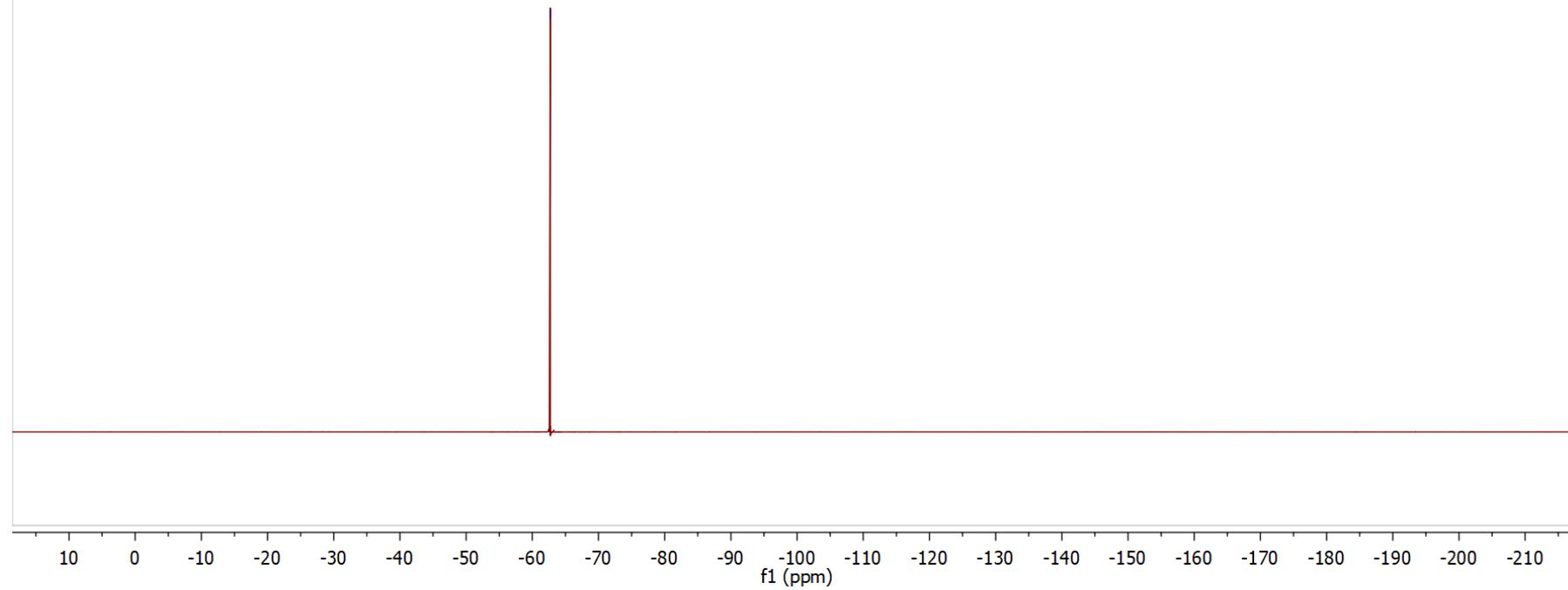


Compound 9I

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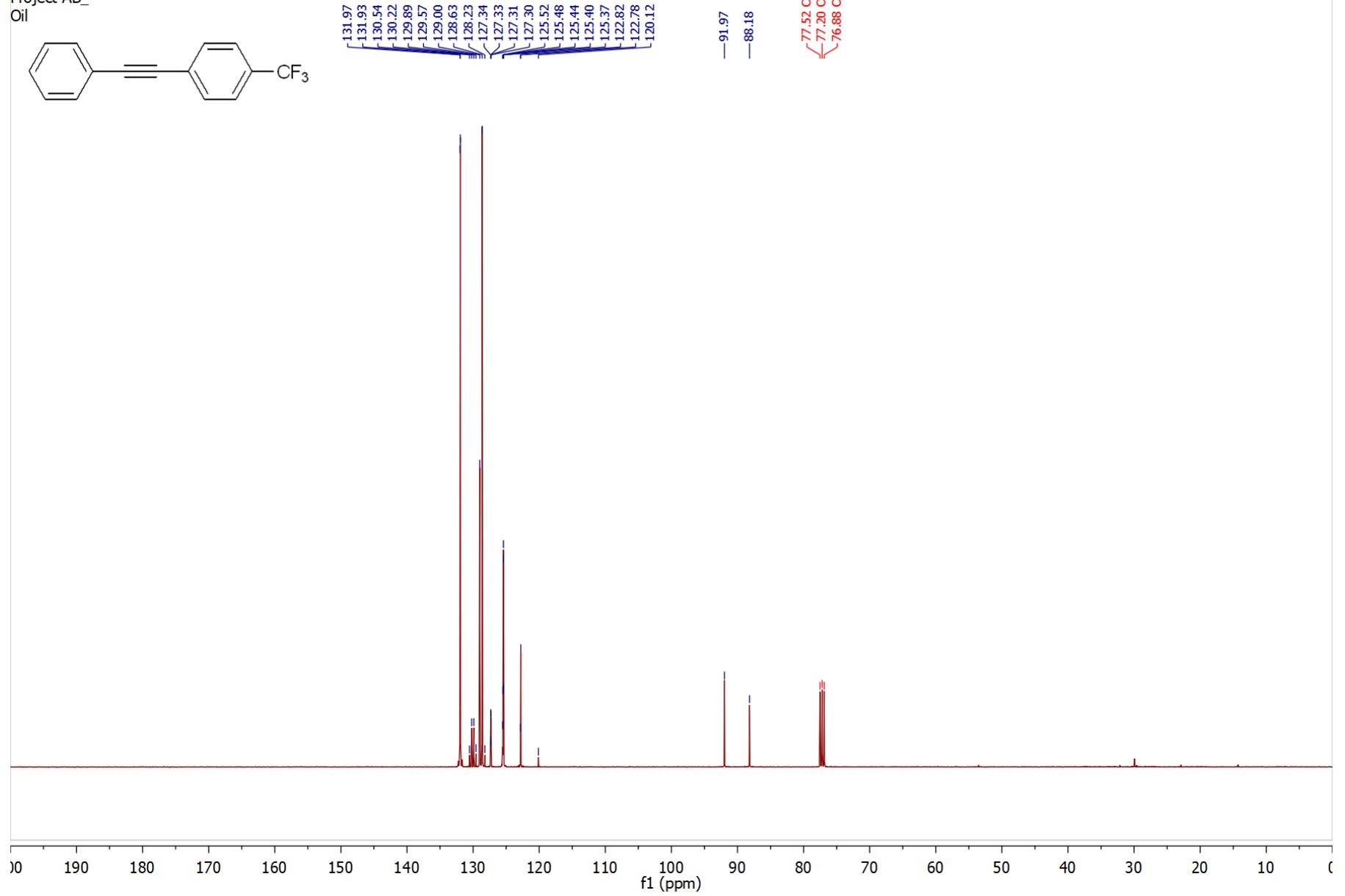
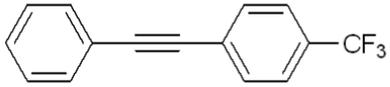


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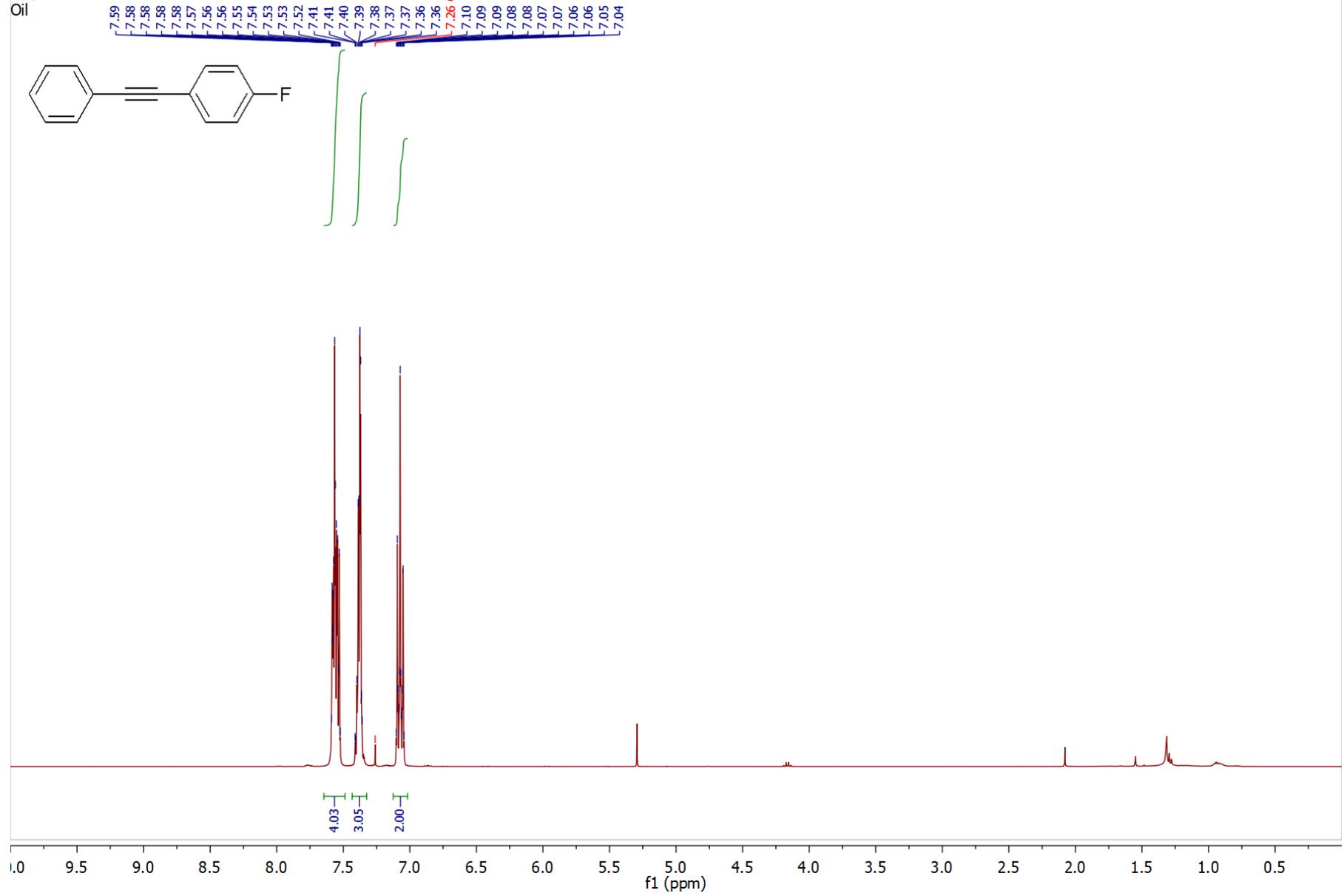
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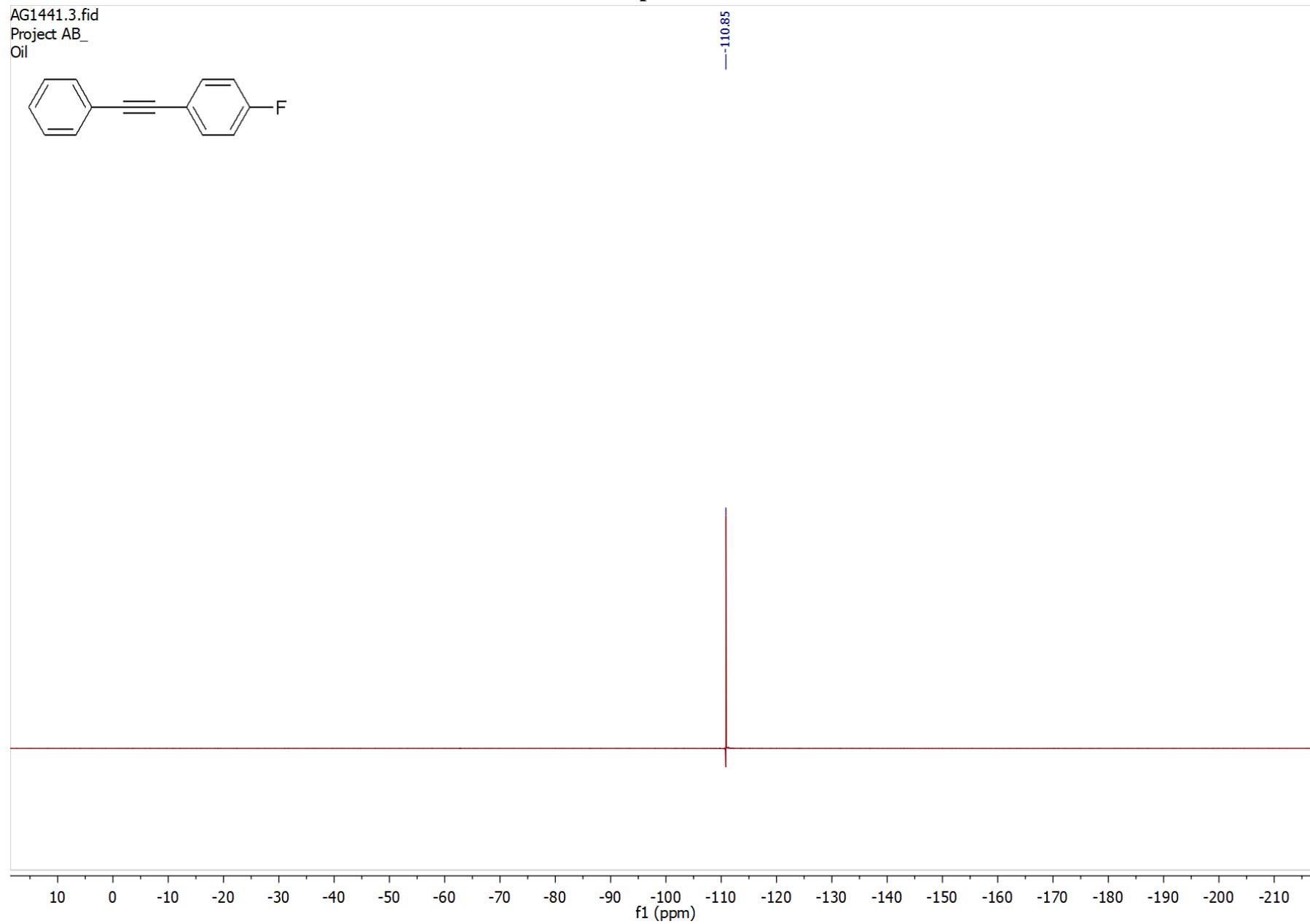
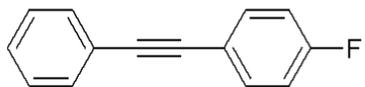
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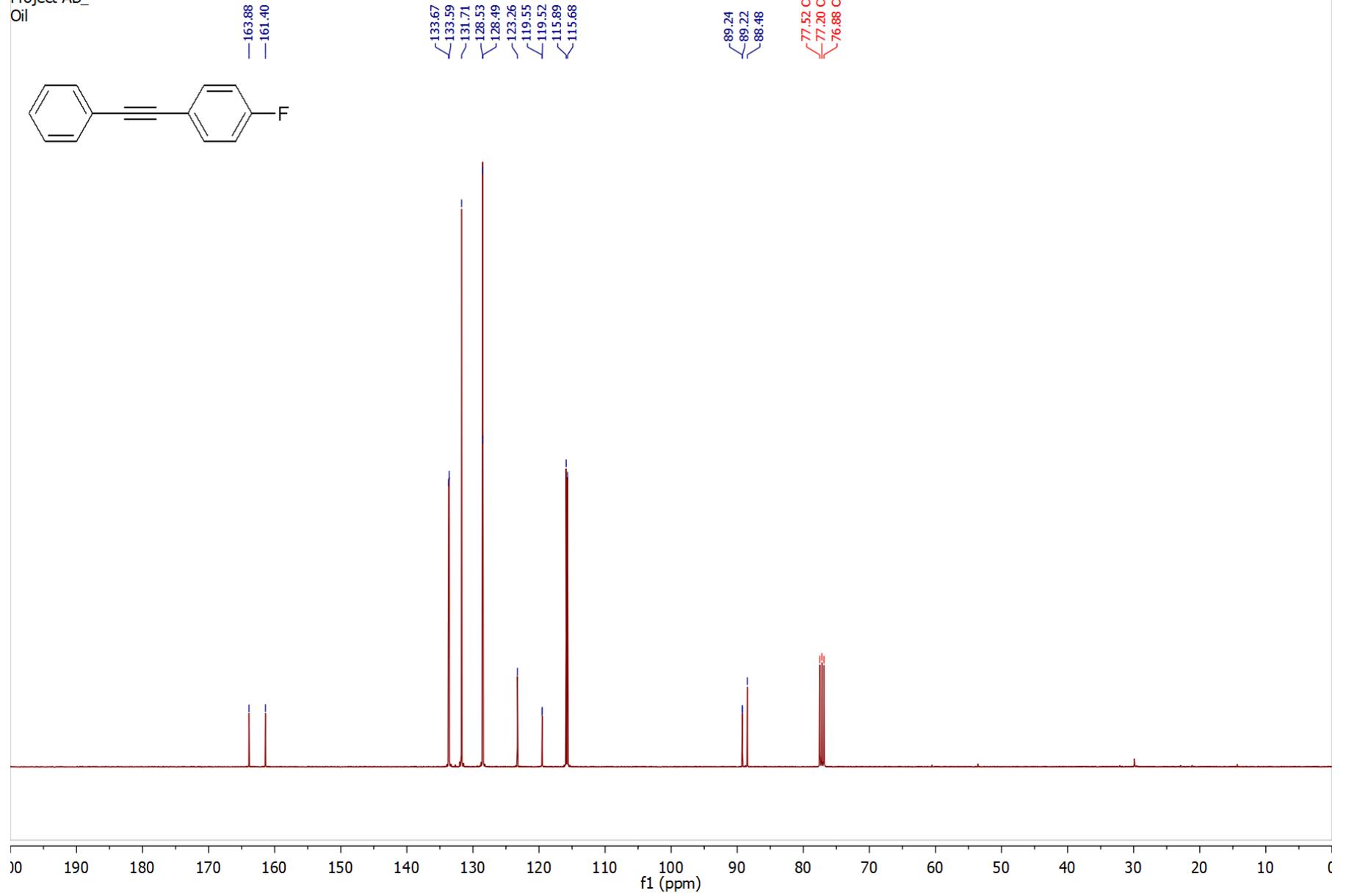
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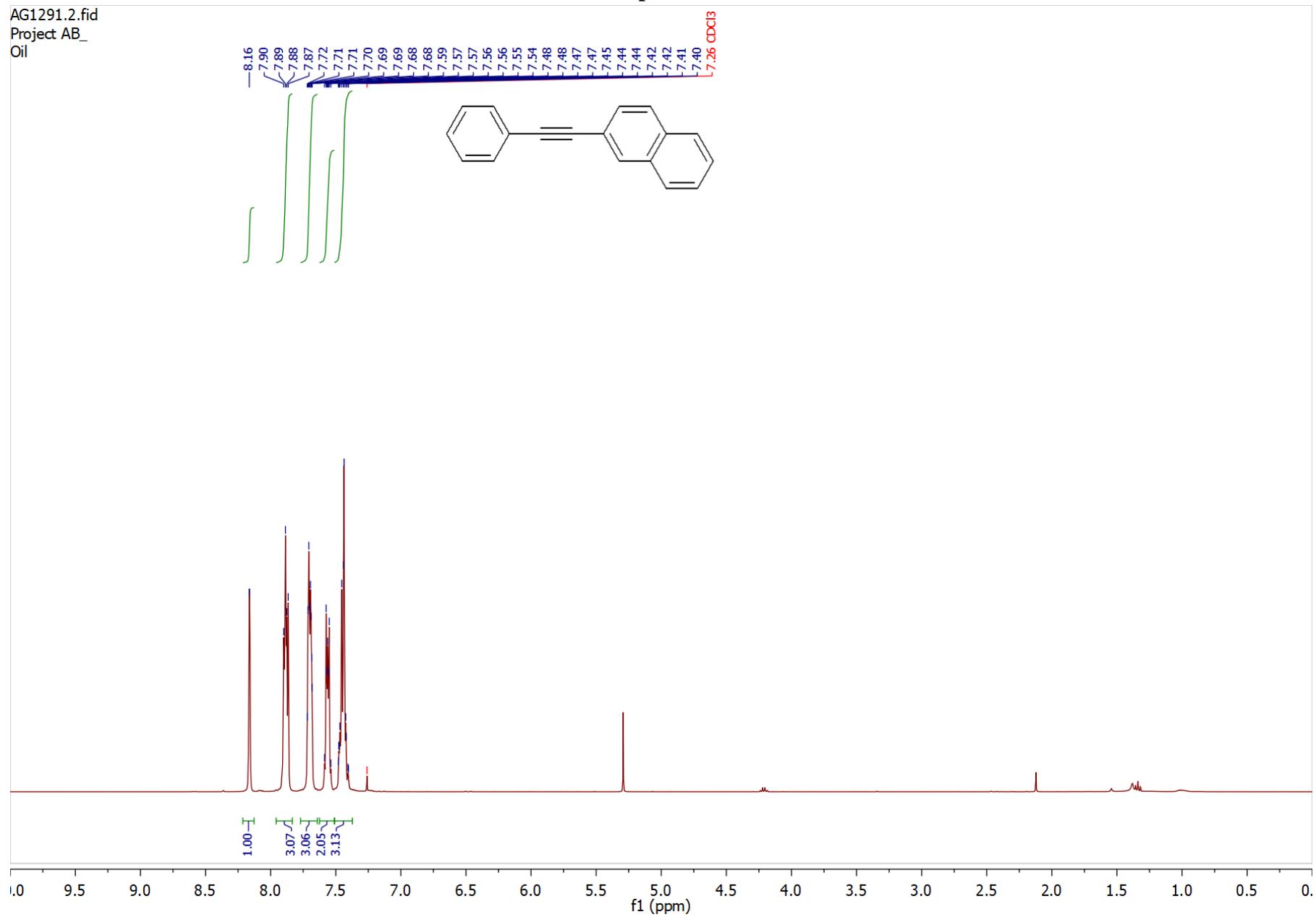
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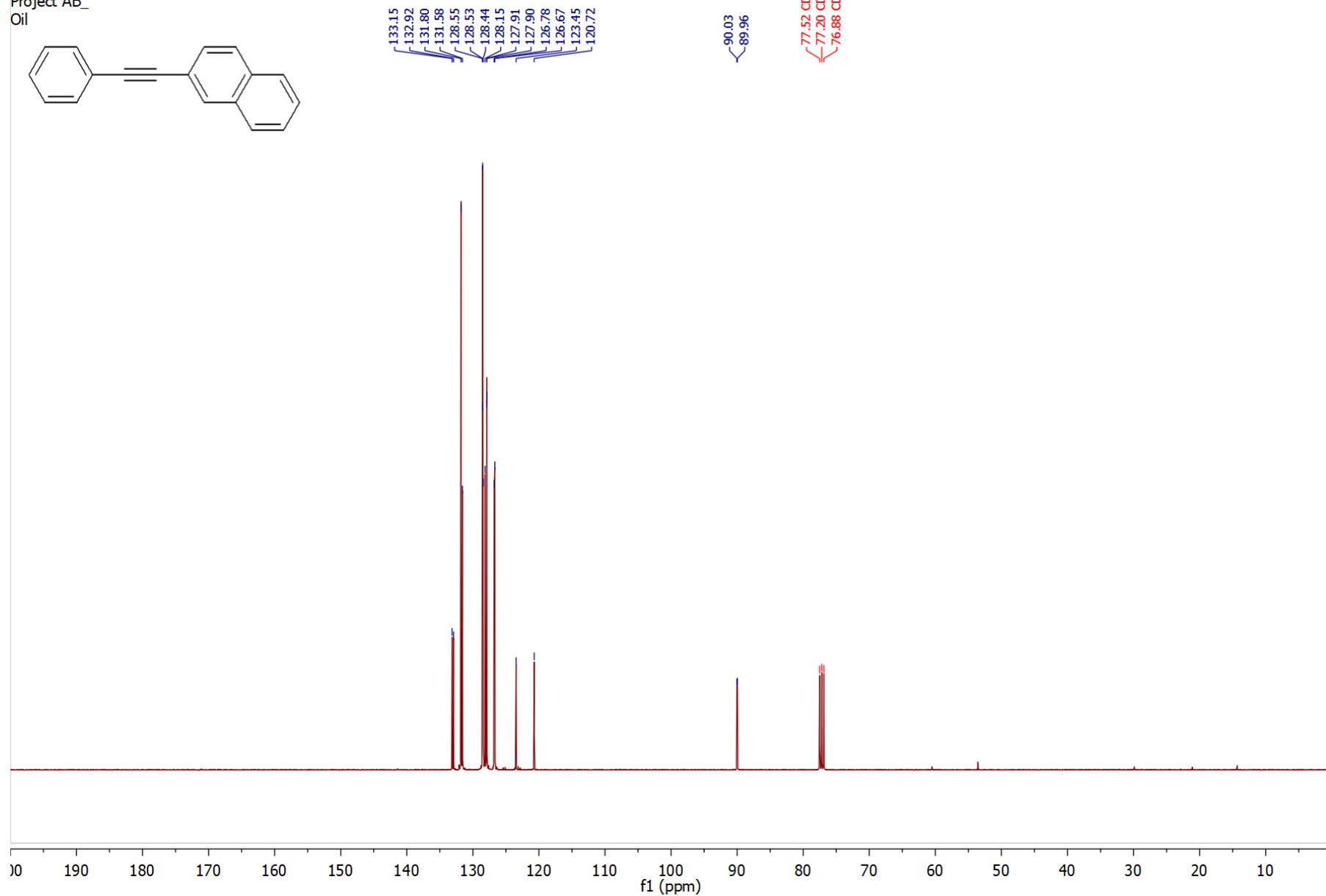
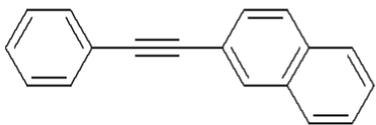
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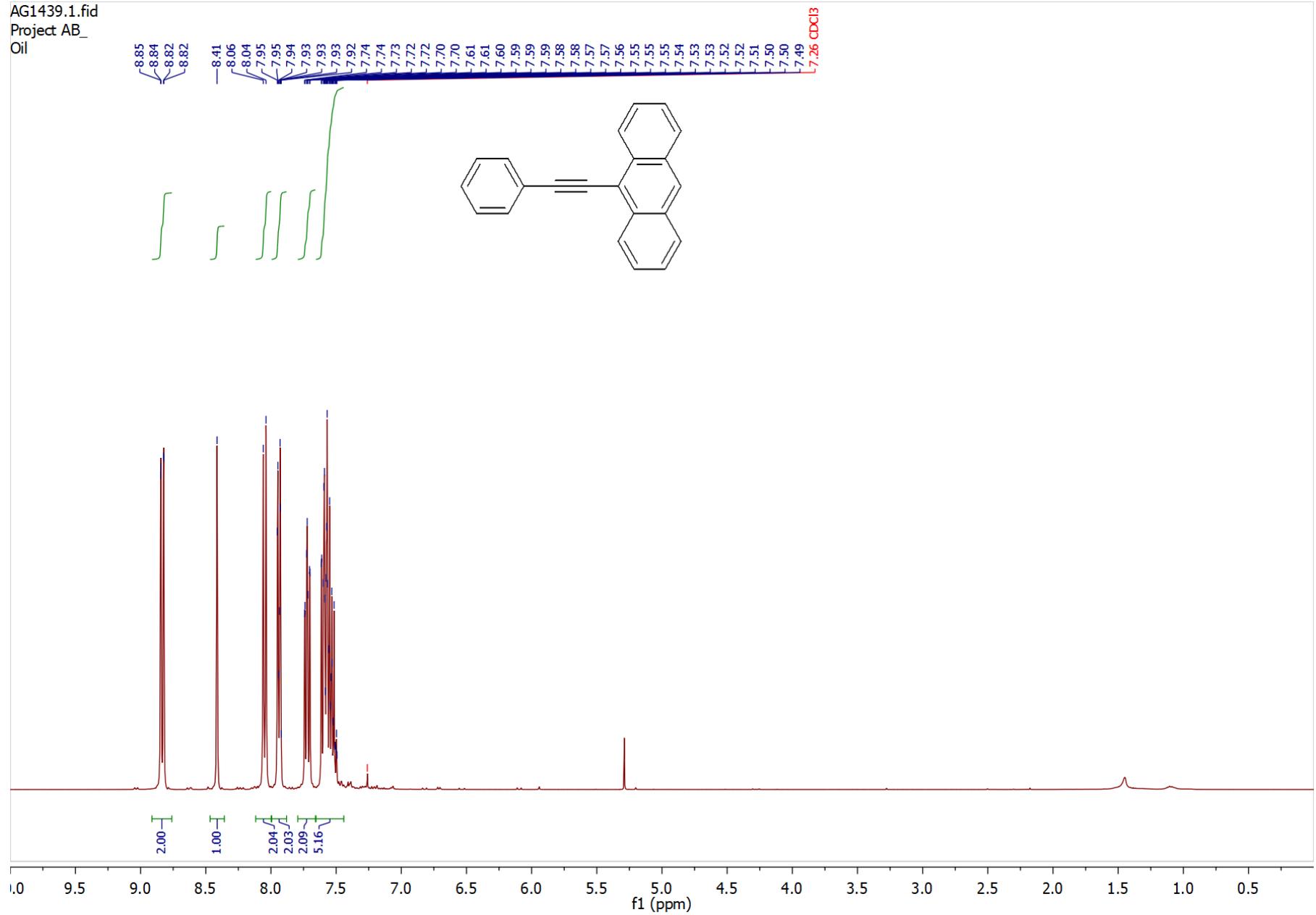
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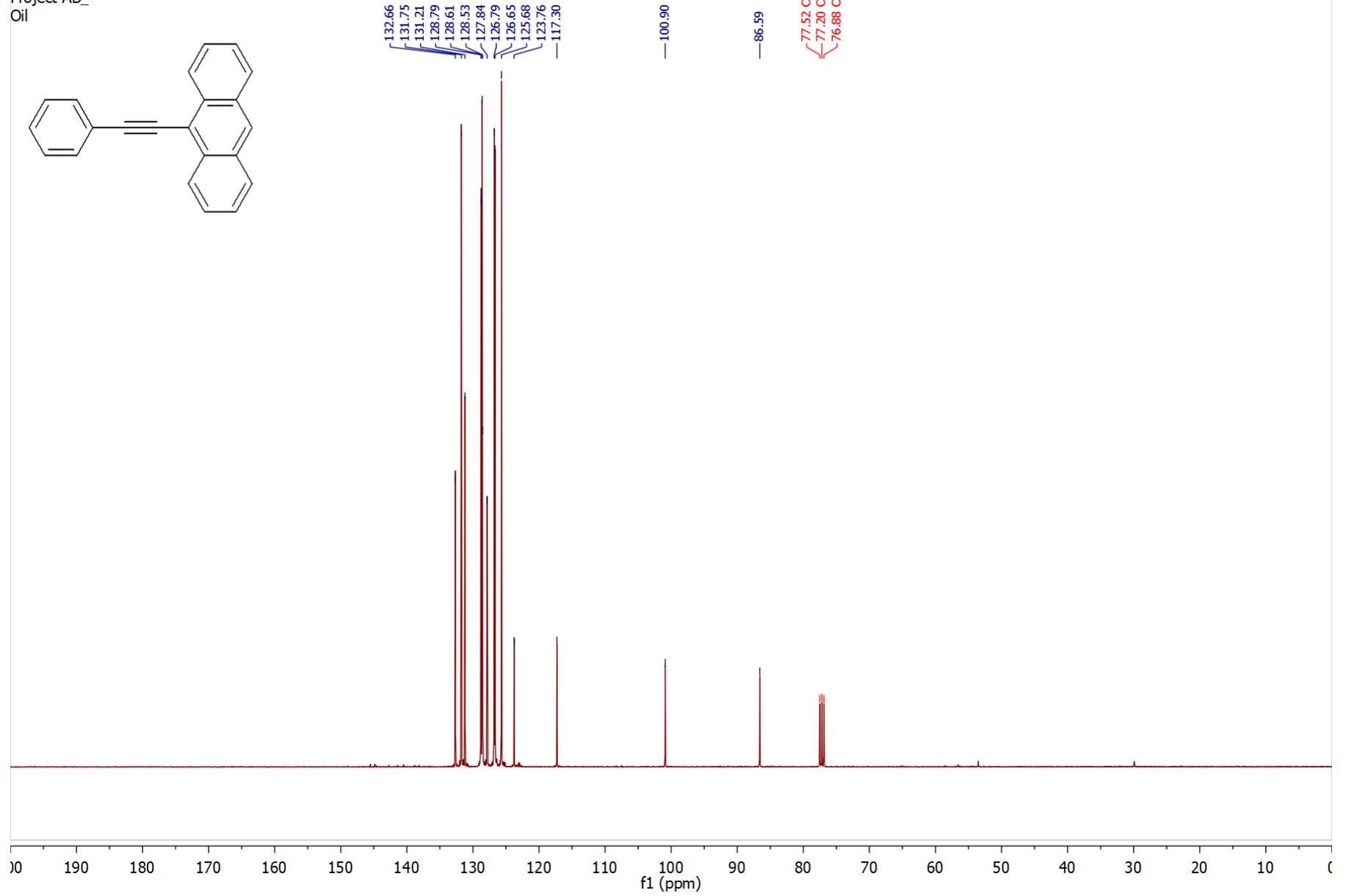
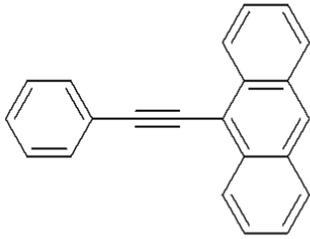
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Compound 9o

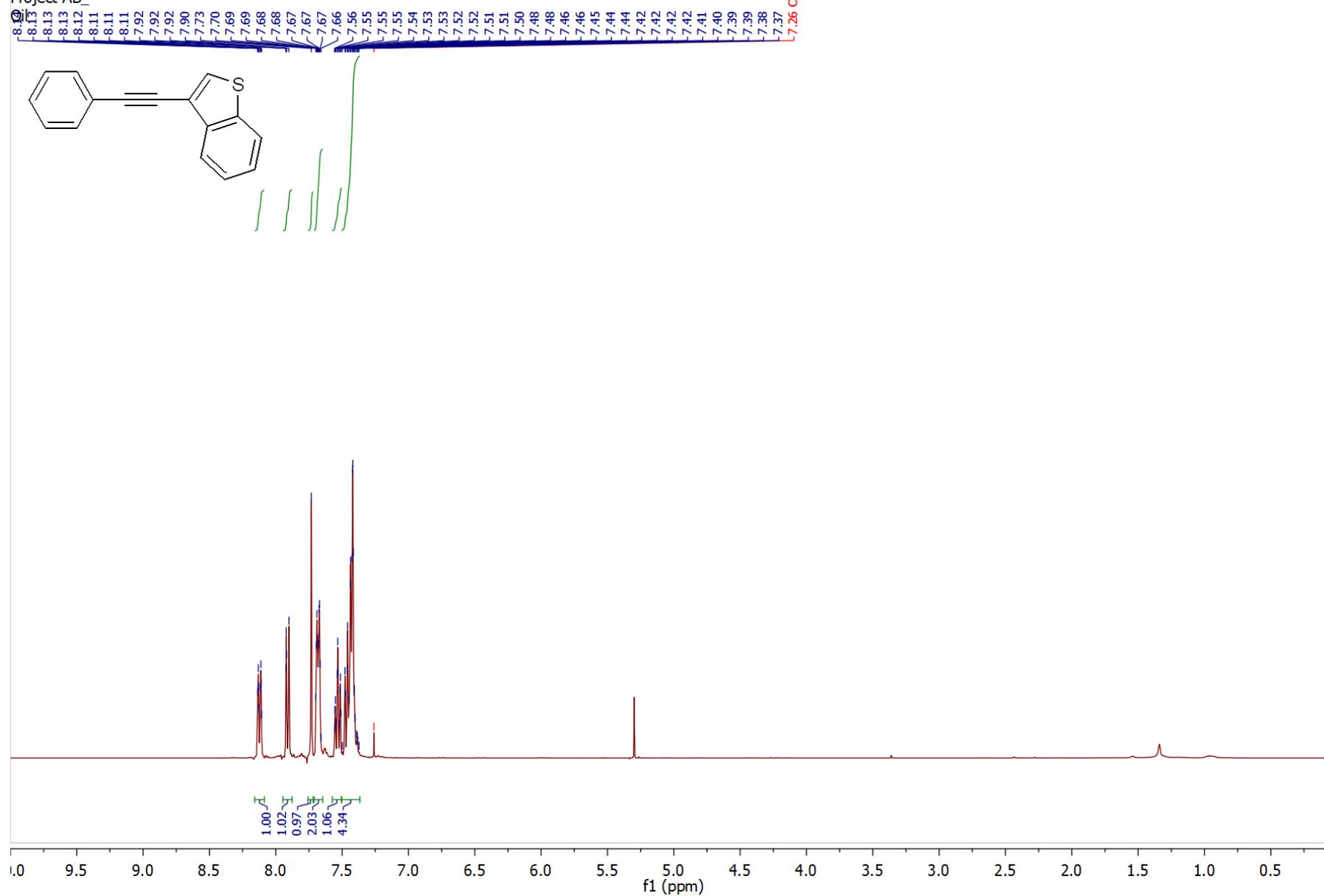
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Compound 9p

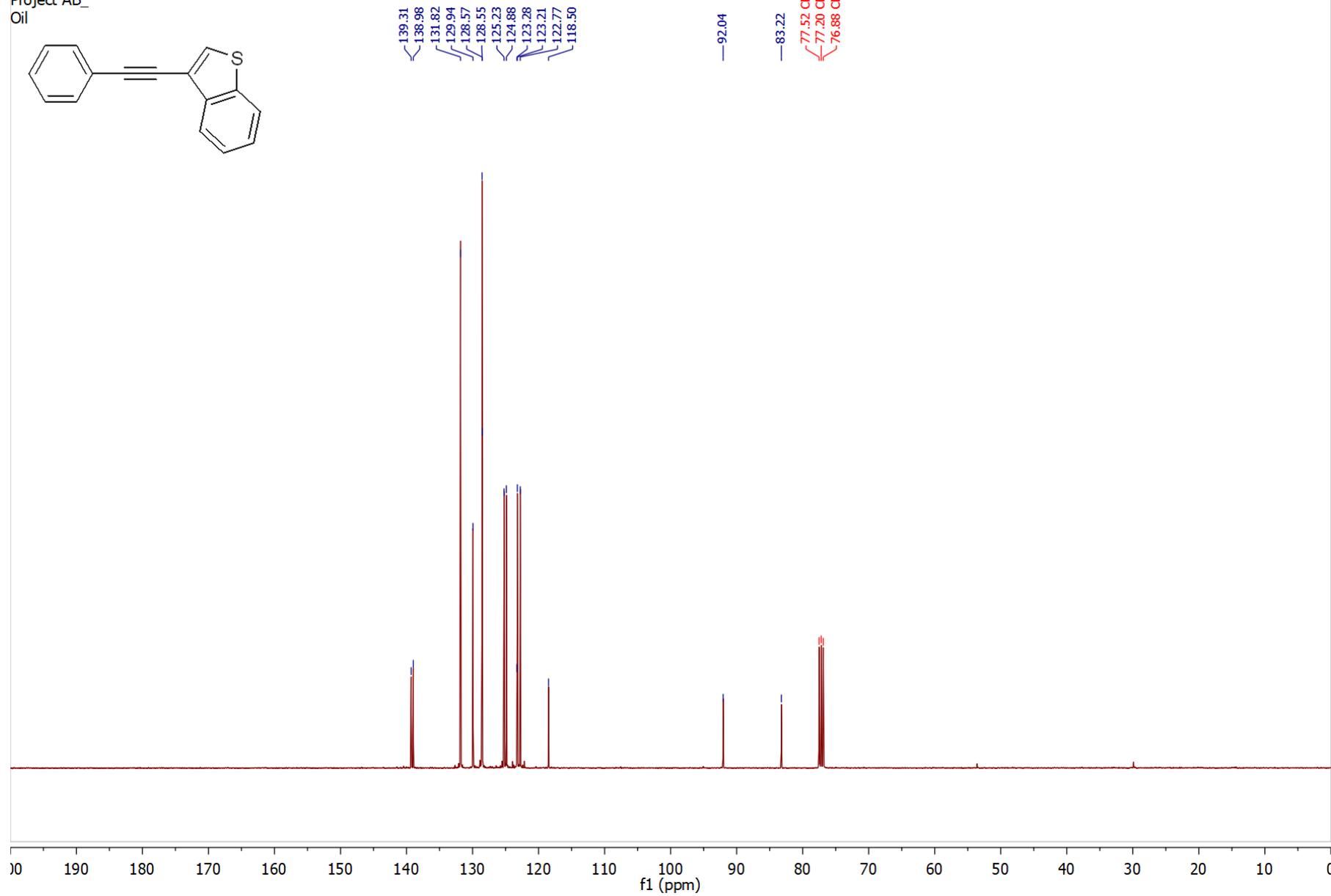
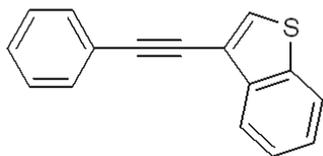
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Project AB



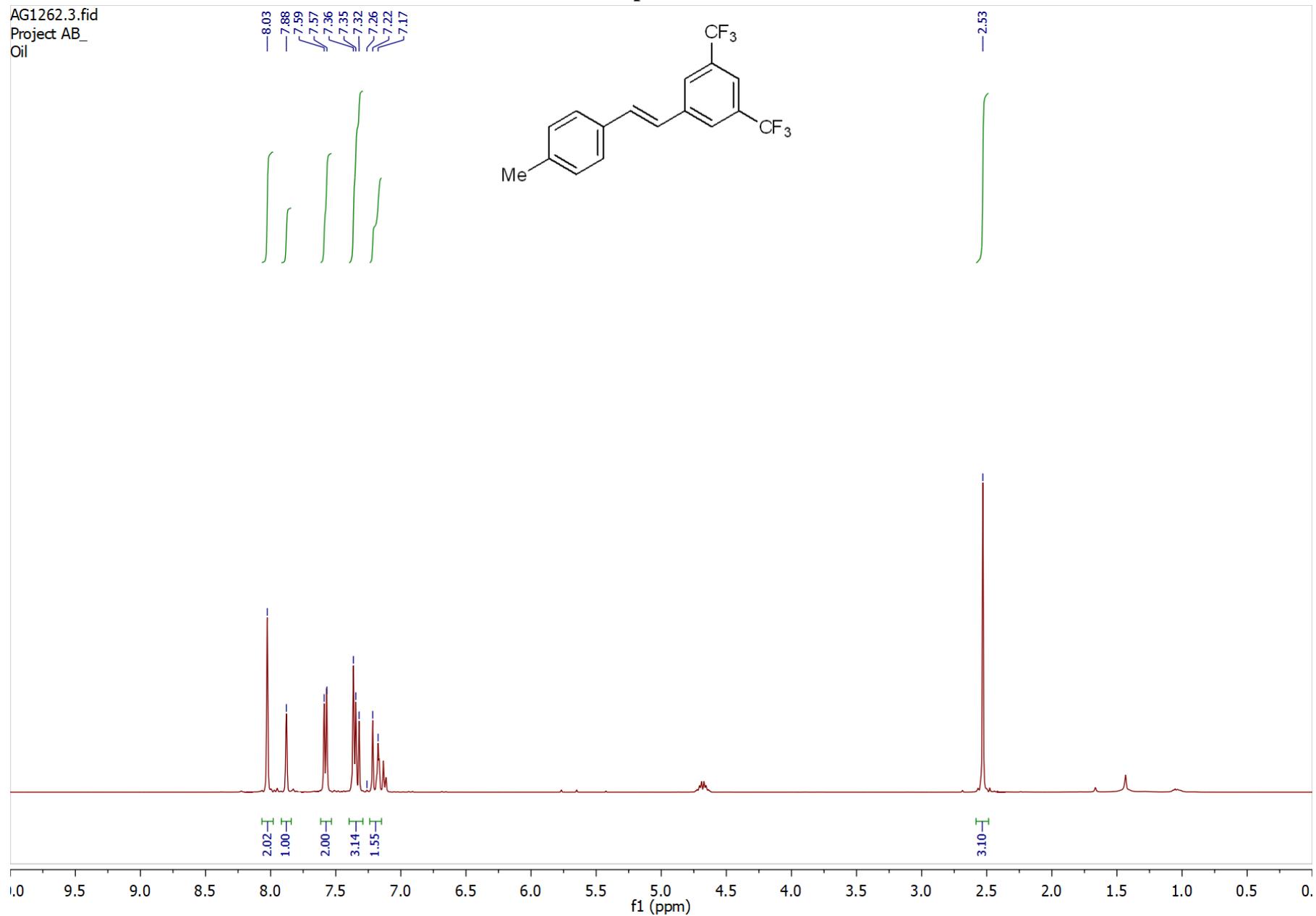
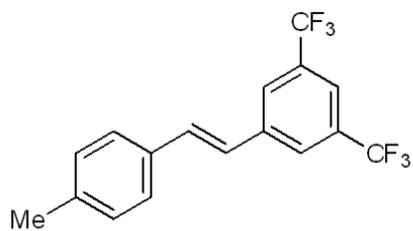
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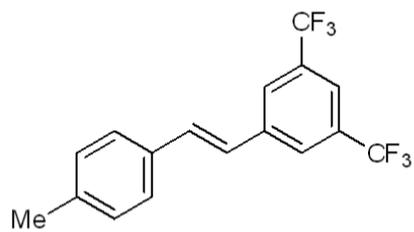
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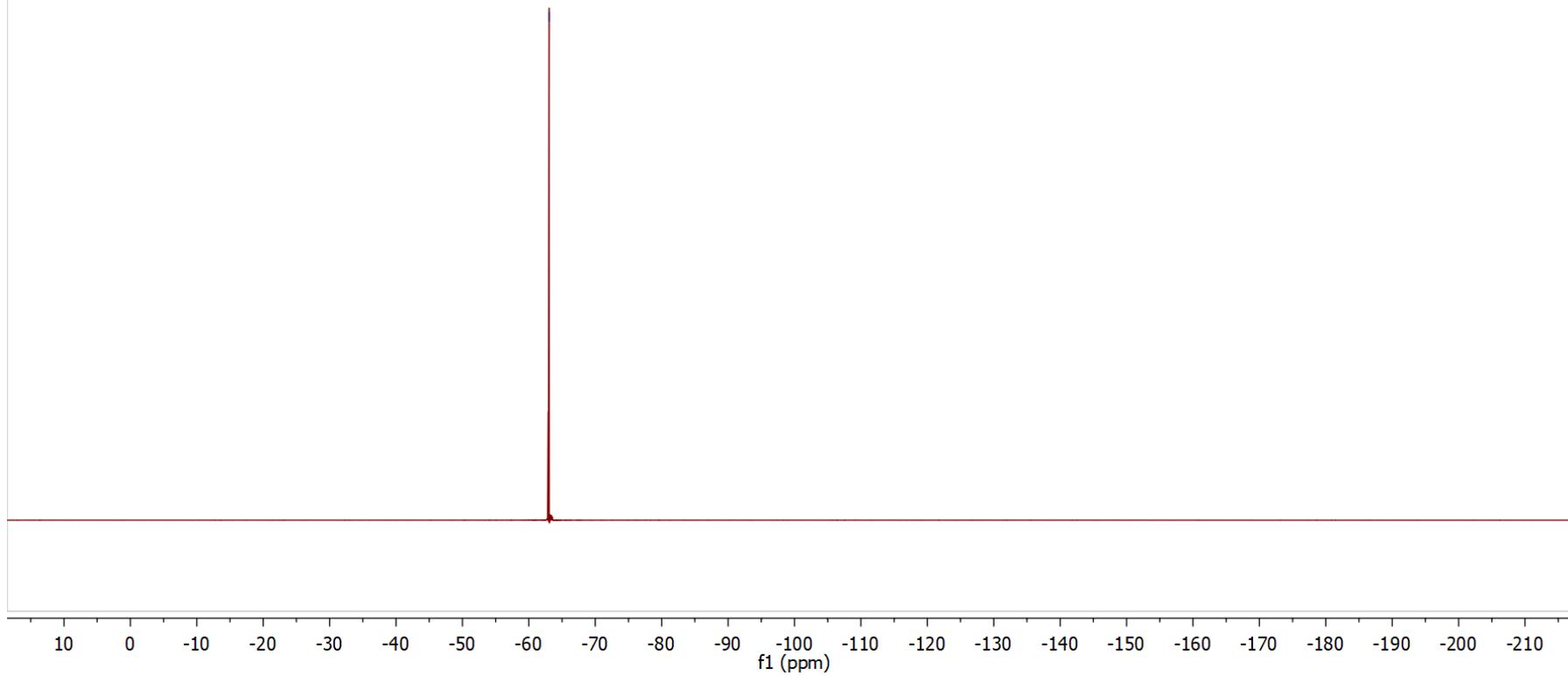


Compound 11a

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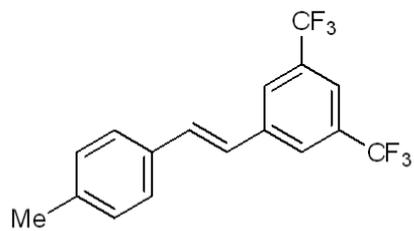


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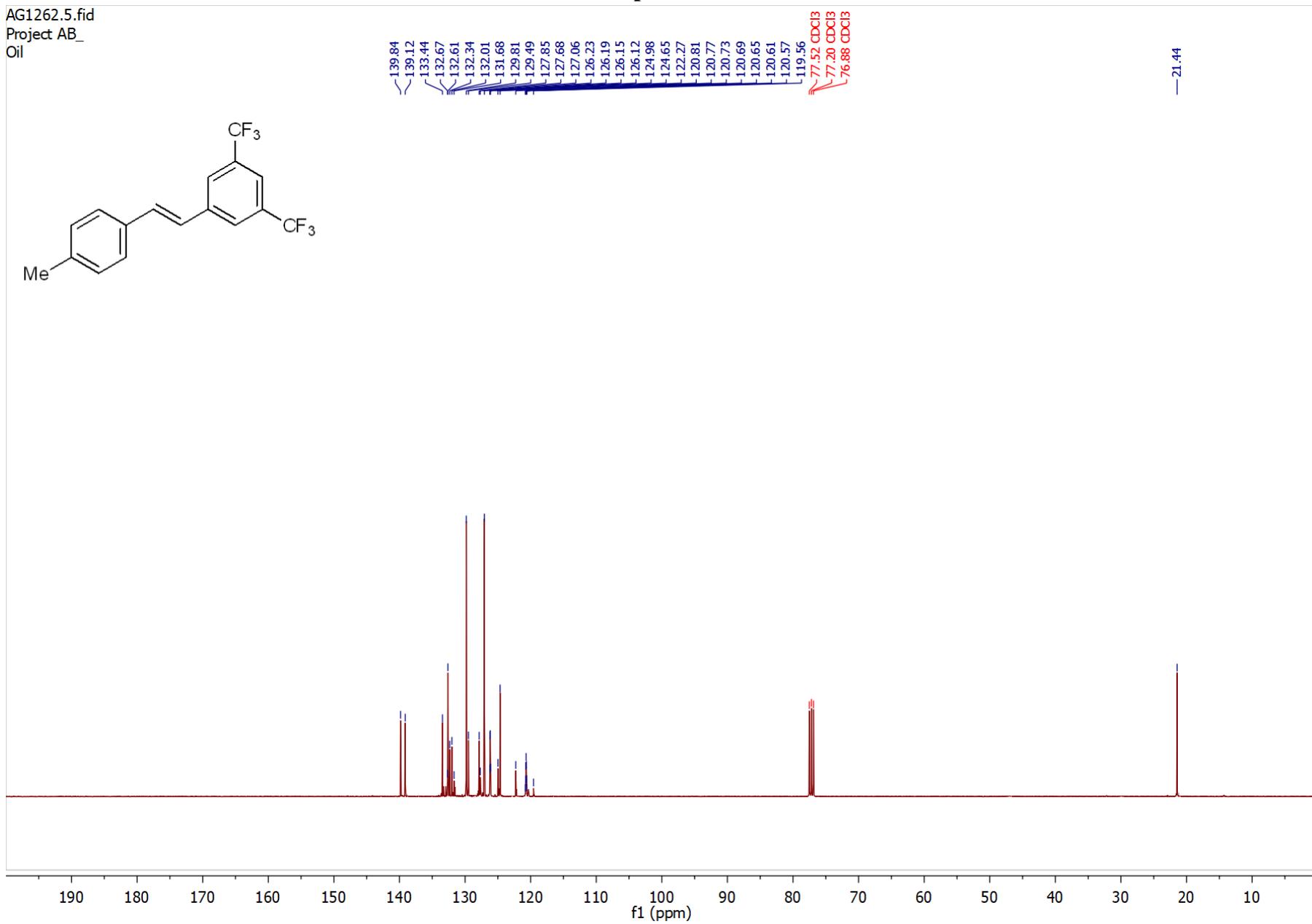
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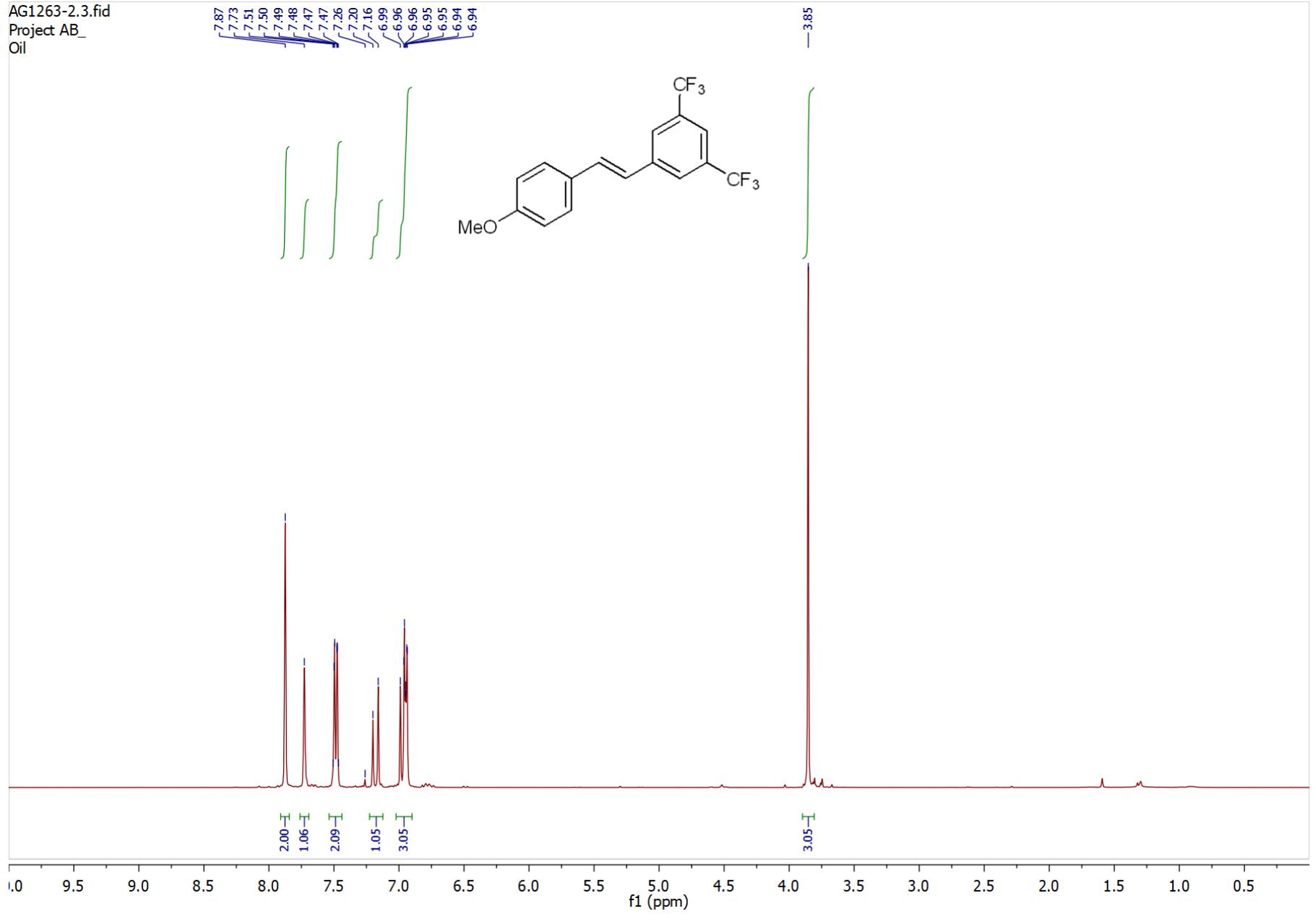
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21.44



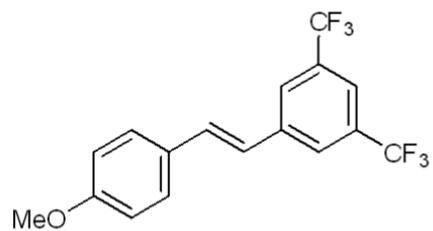
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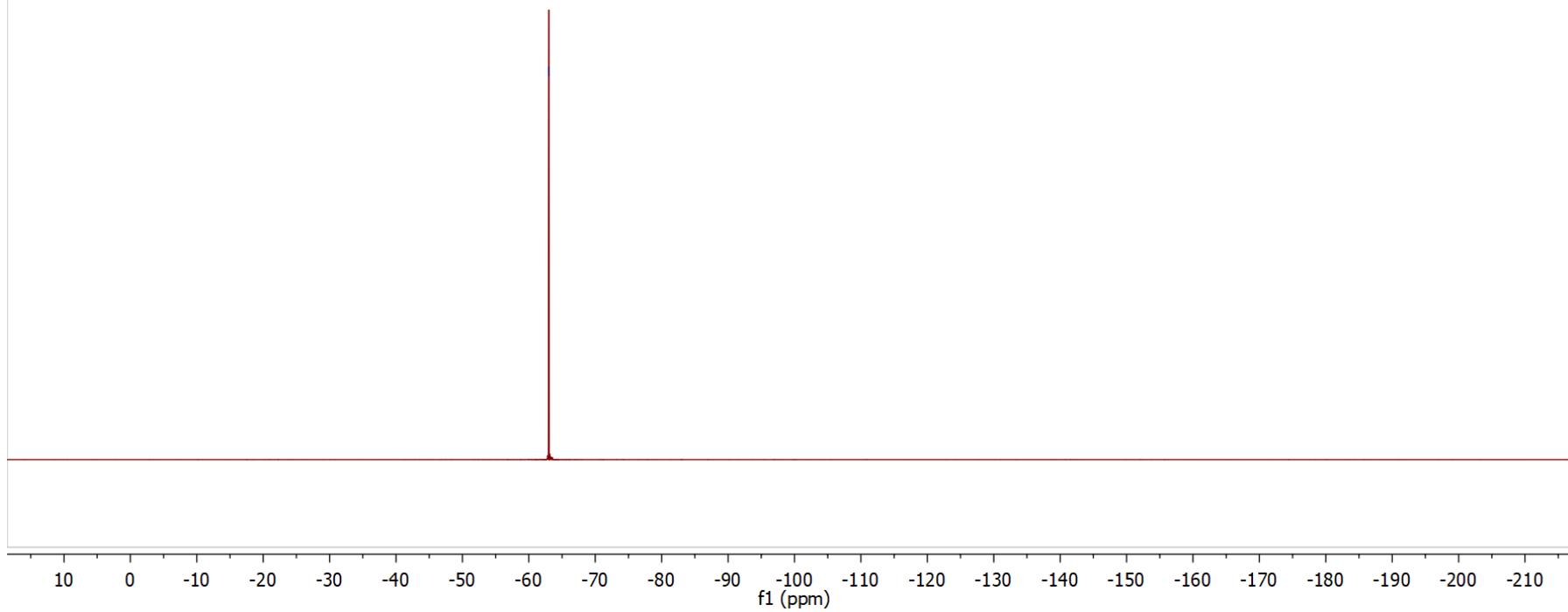


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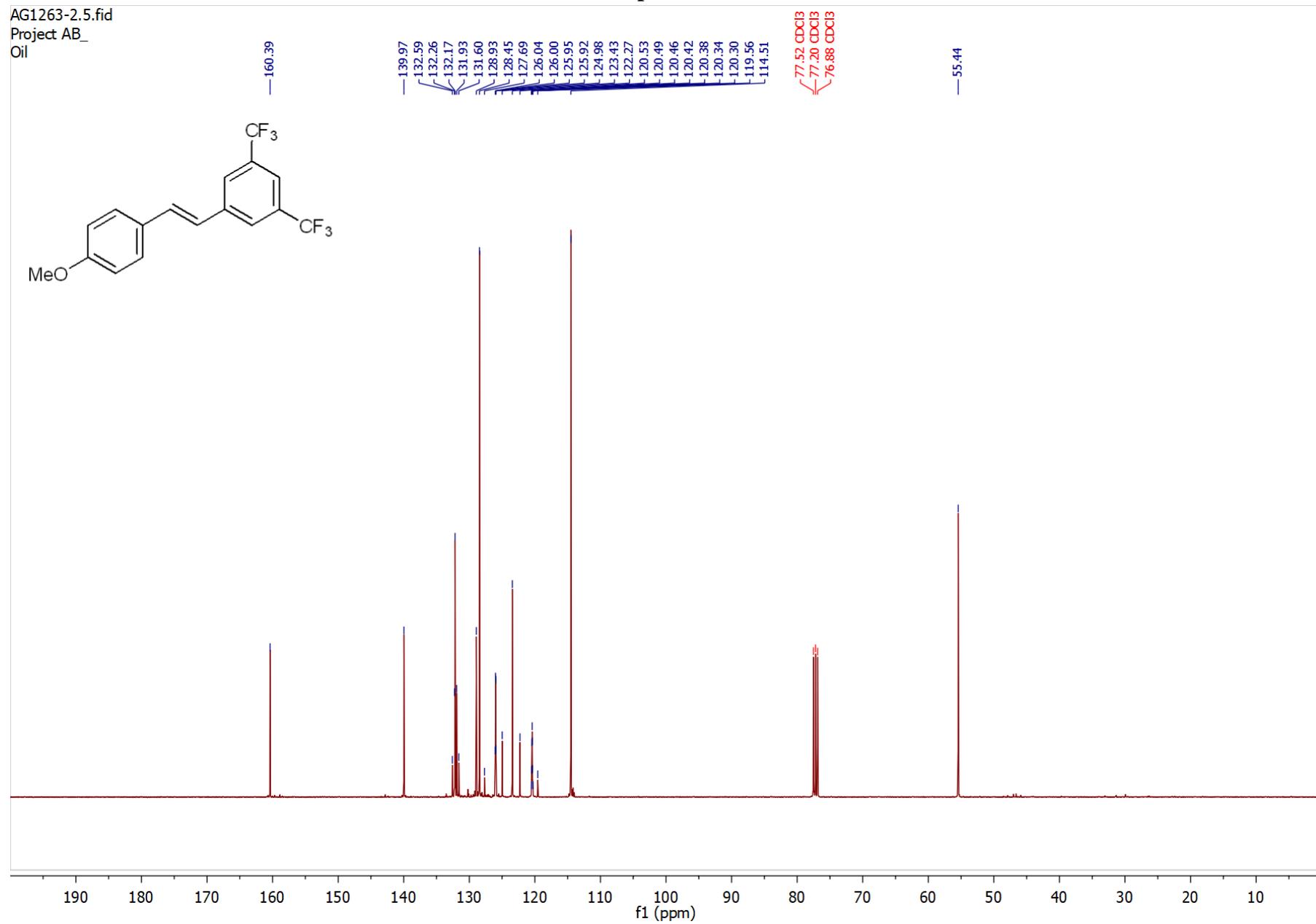


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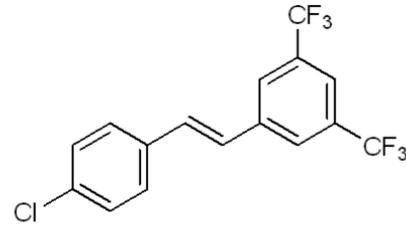
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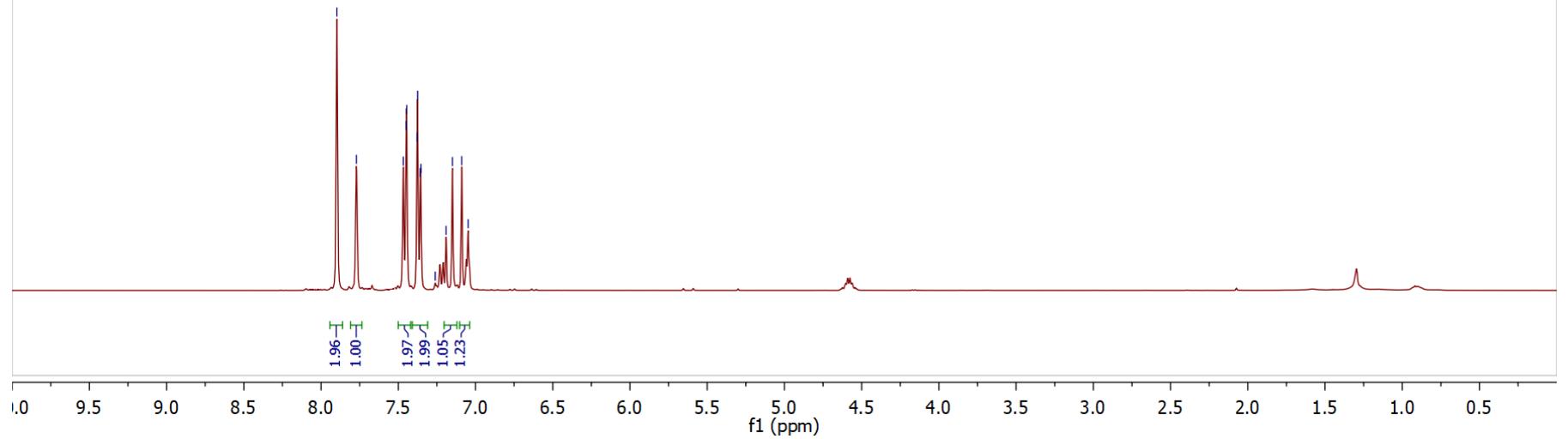
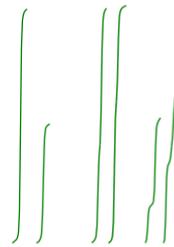


Compound 11c

AG1264.3.fid
Project AB_
Oil

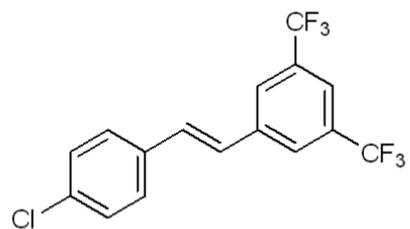


7.90
7.77
7.47
7.45
7.45
7.38
7.37
7.36
7.35
7.26
7.19
7.15
7.09
7.05

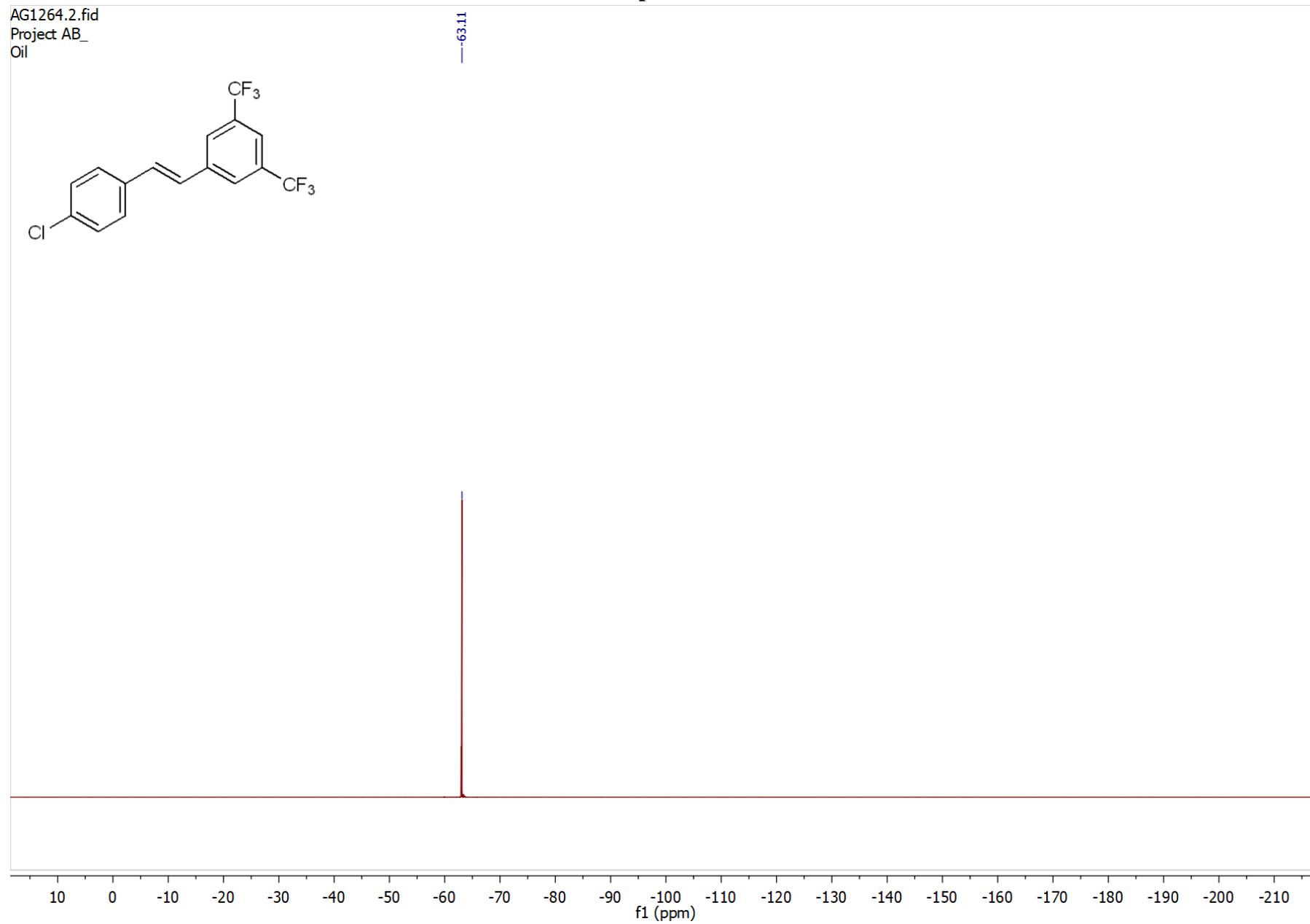


Compound 11c

AG1264.2.fid
Project AB_
Oil

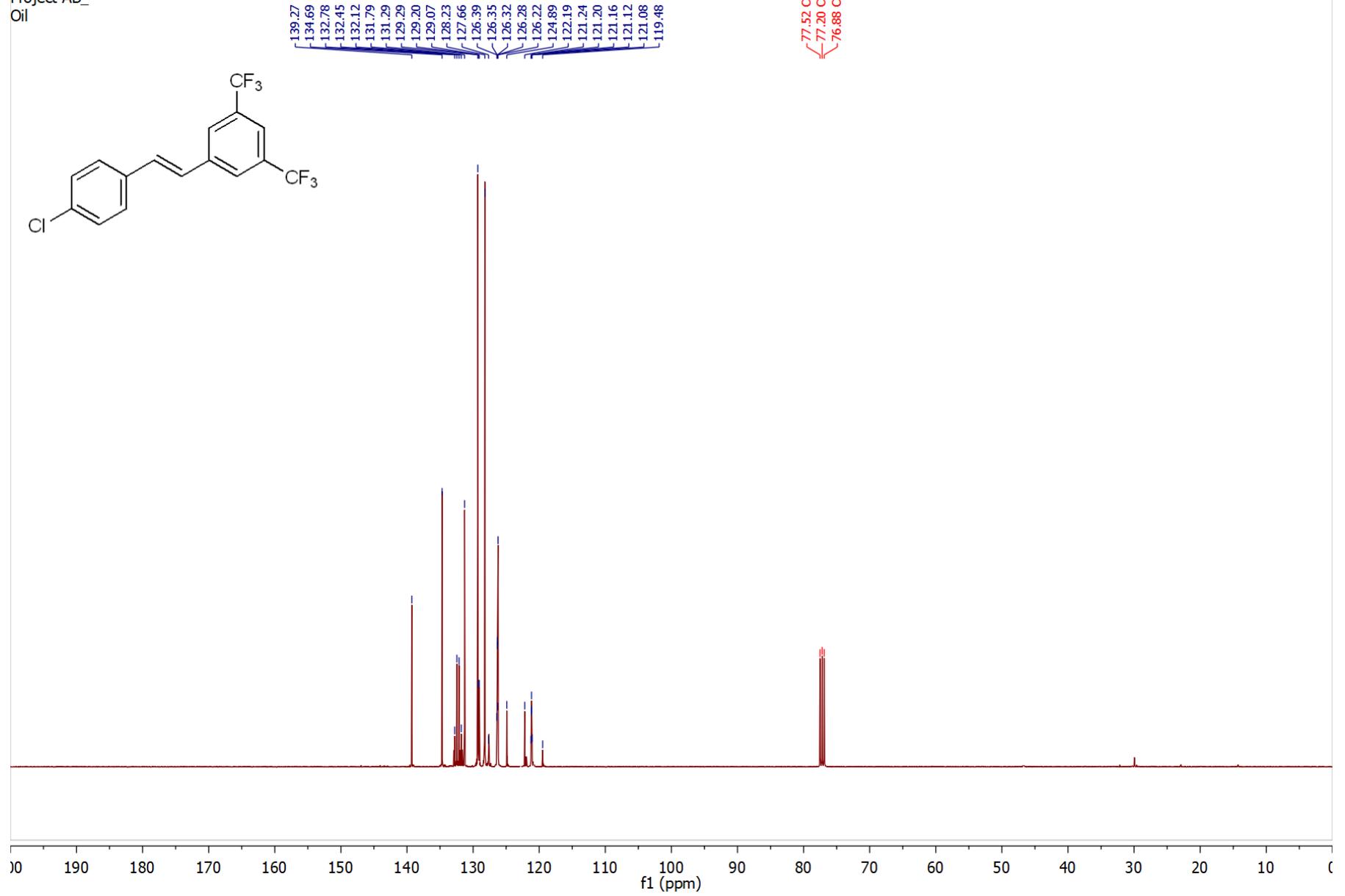


—63.11



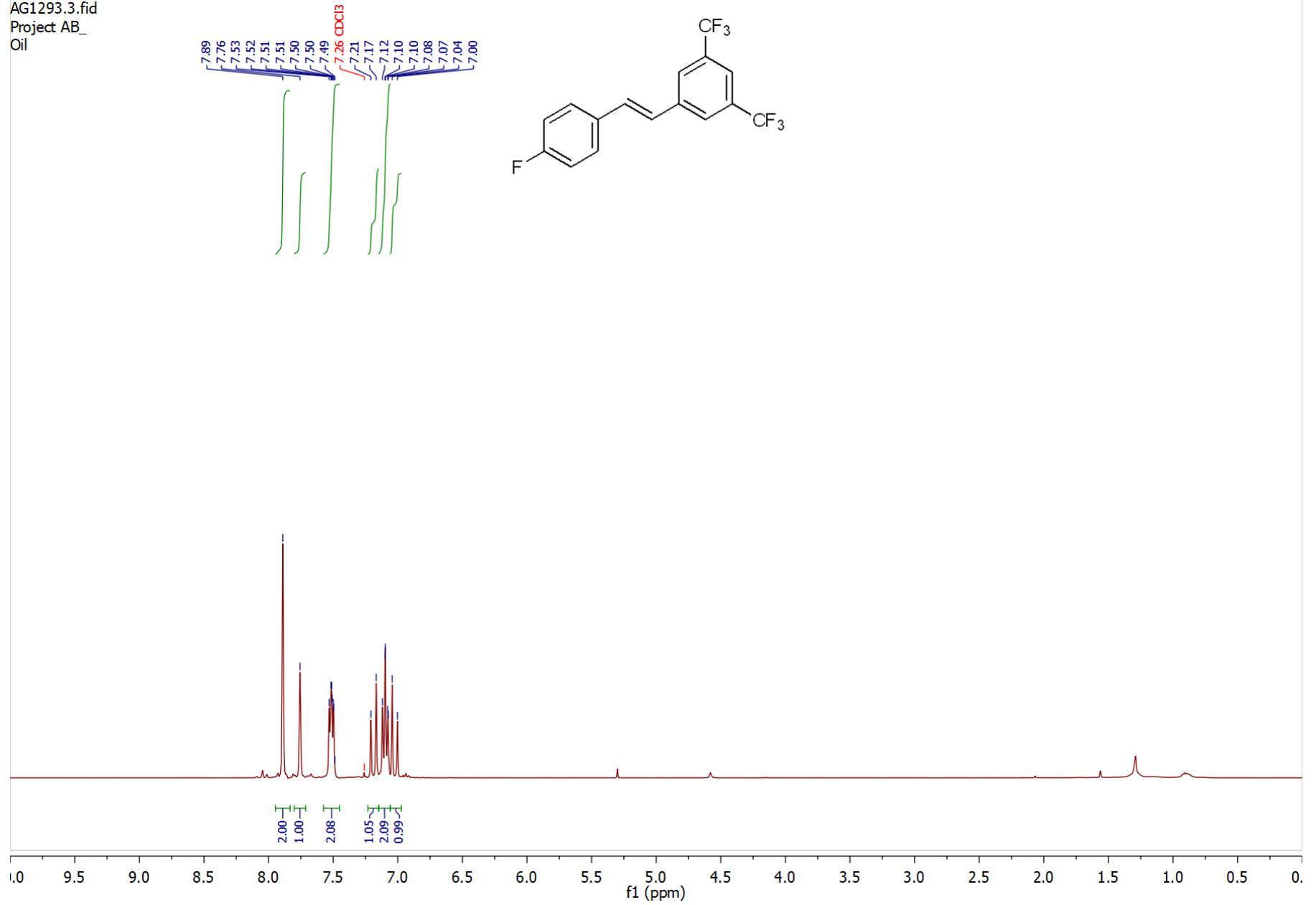
Compound 11c

AG1264.5.fid
Project AB_
Oil



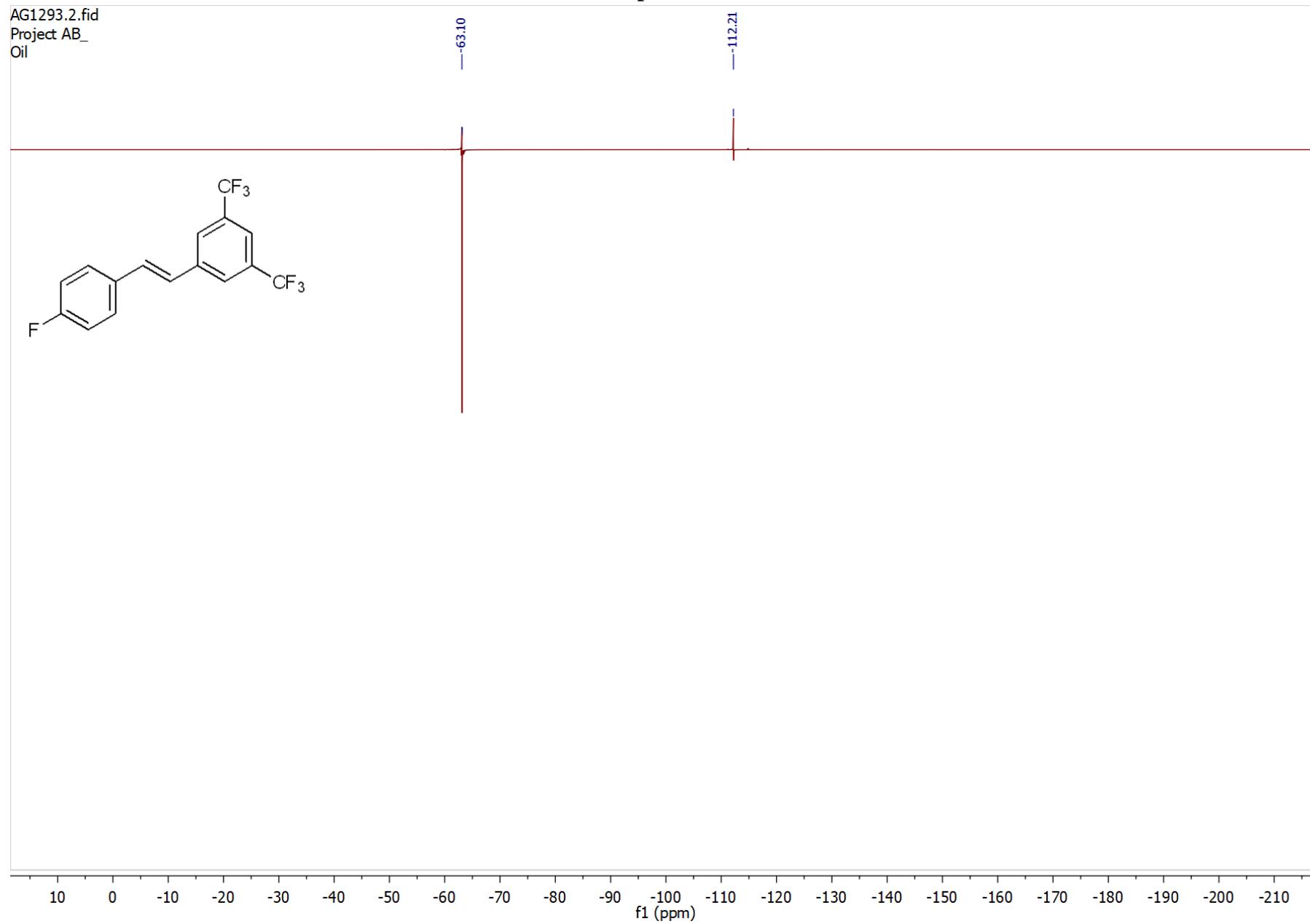
Compound 11d

AG1293.3.fid
Project AB_
Oil



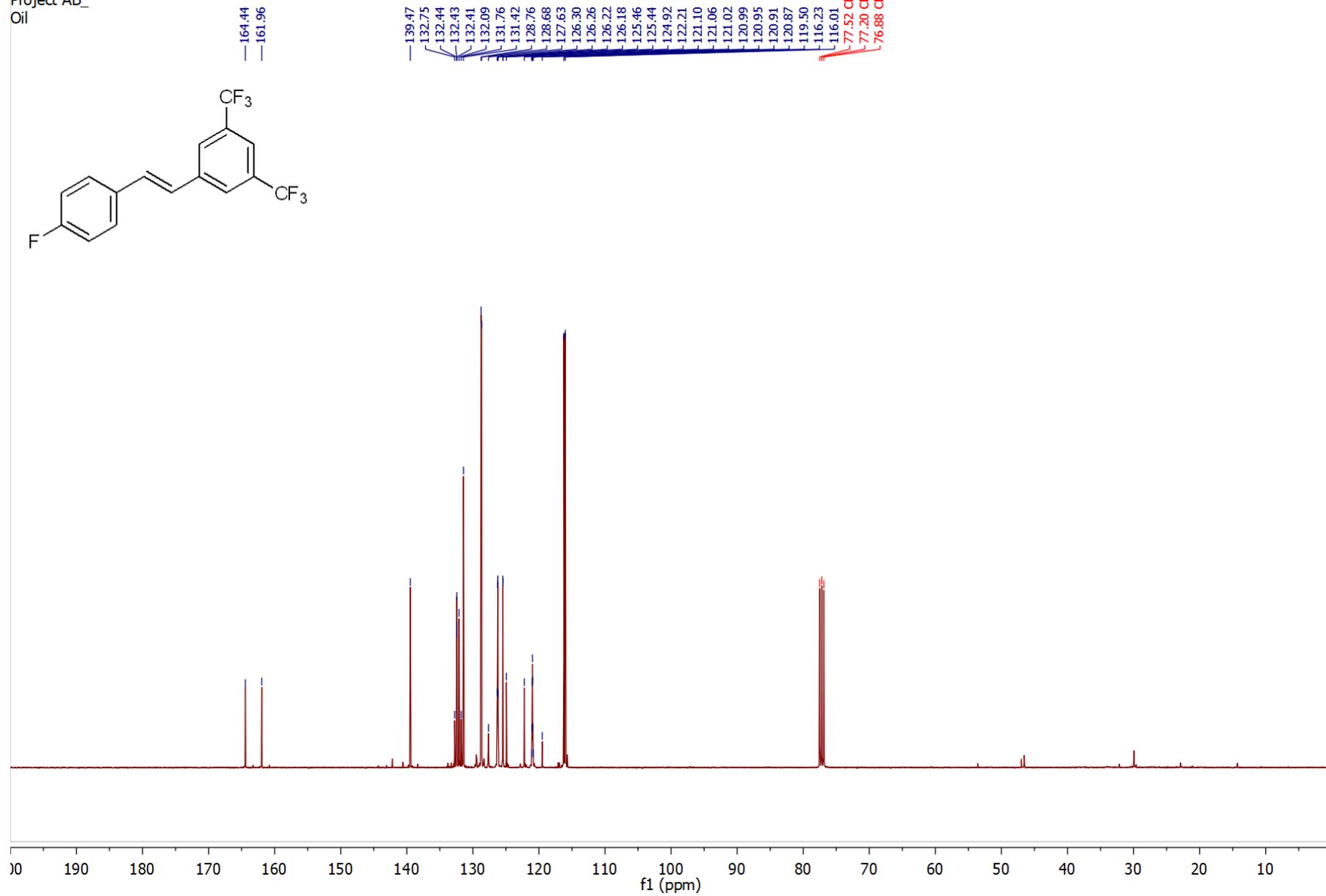
Compound 11d

AG1293.2.fid
Project AB_
Oil



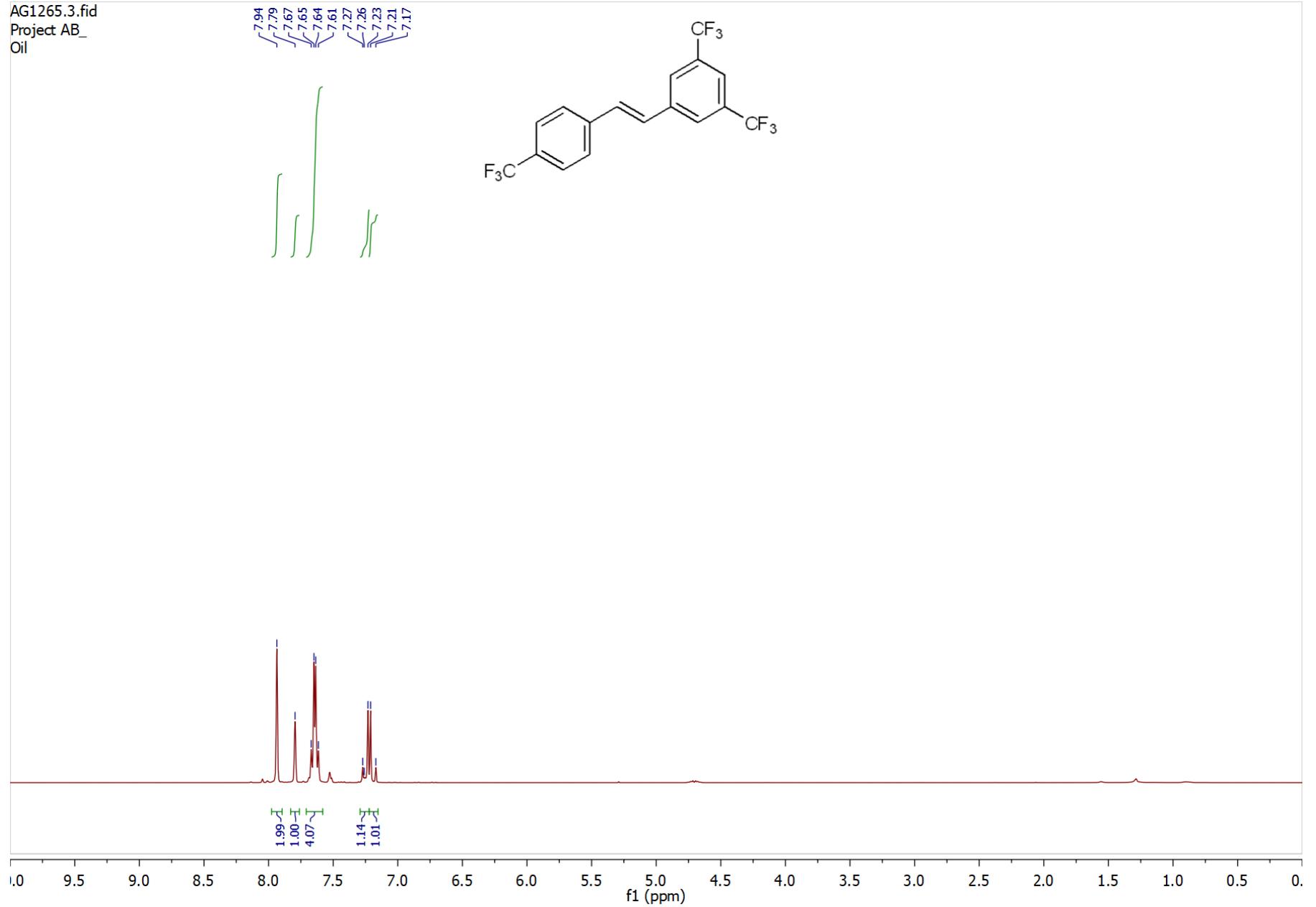
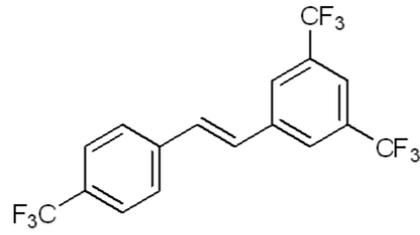
Compound 11d

AG1293.5.fid
Project AB_
Oil



Compound 11e

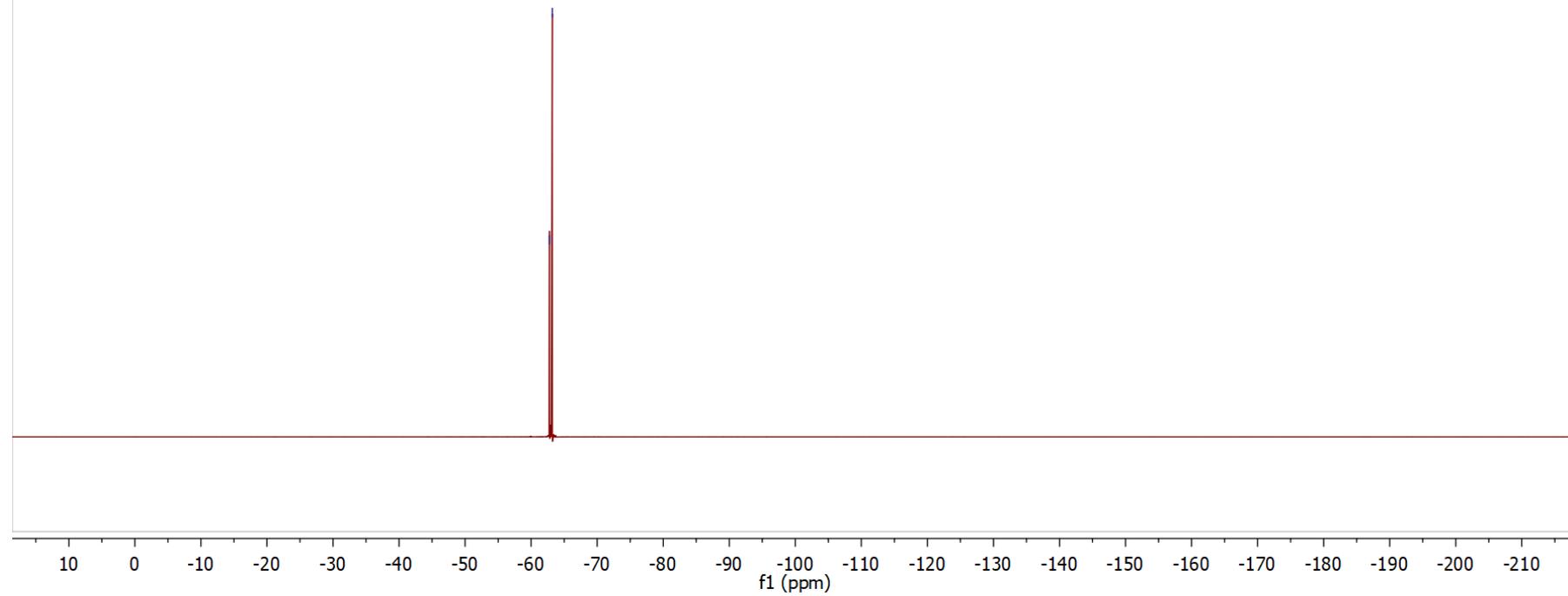
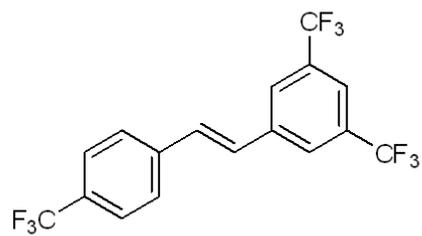
AG1265.3.fid
Project AB_
Oil



Compound 11e

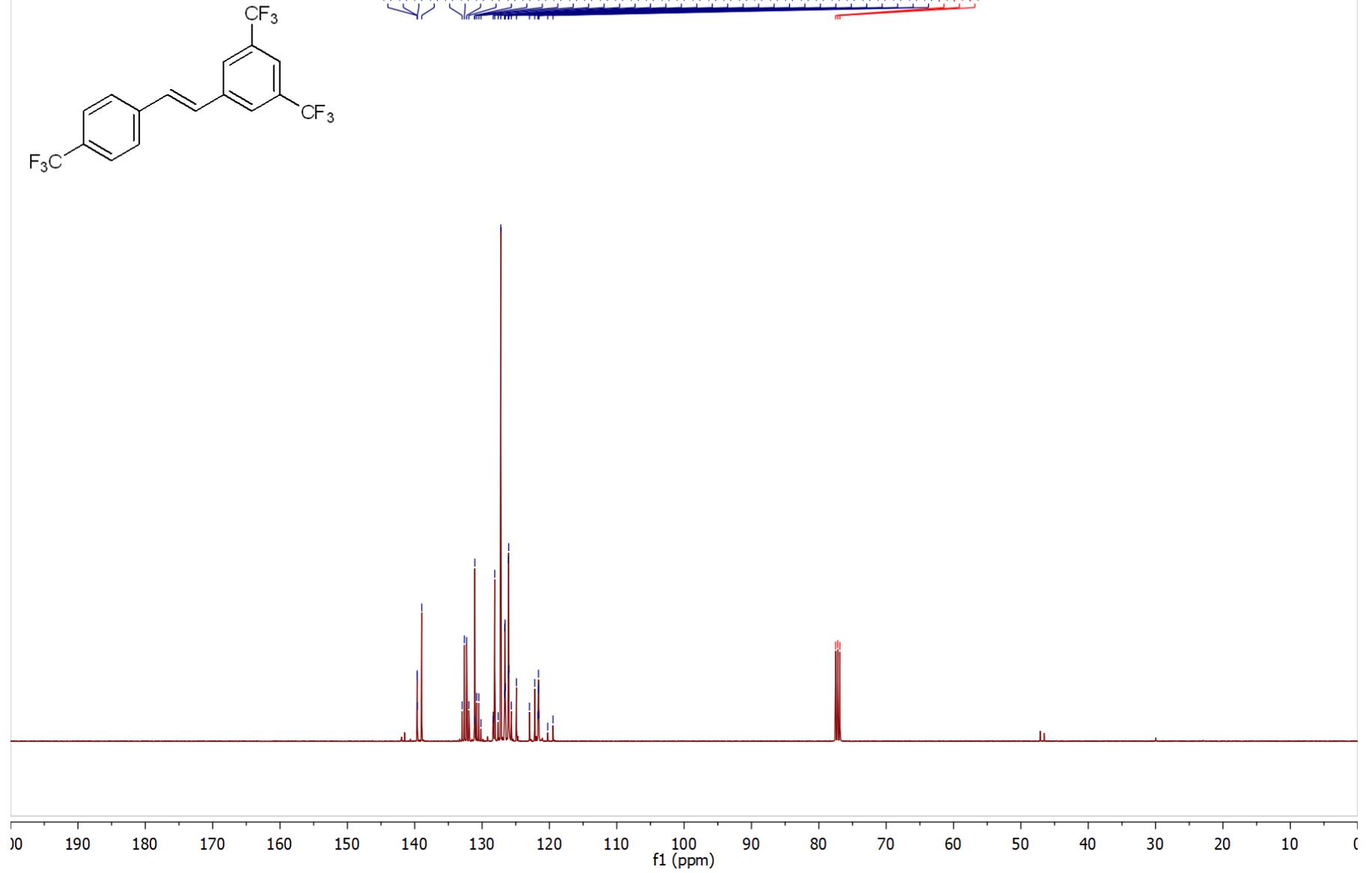
AG1265.2.fid
Project AB_
Oil

-62.79
-63.20



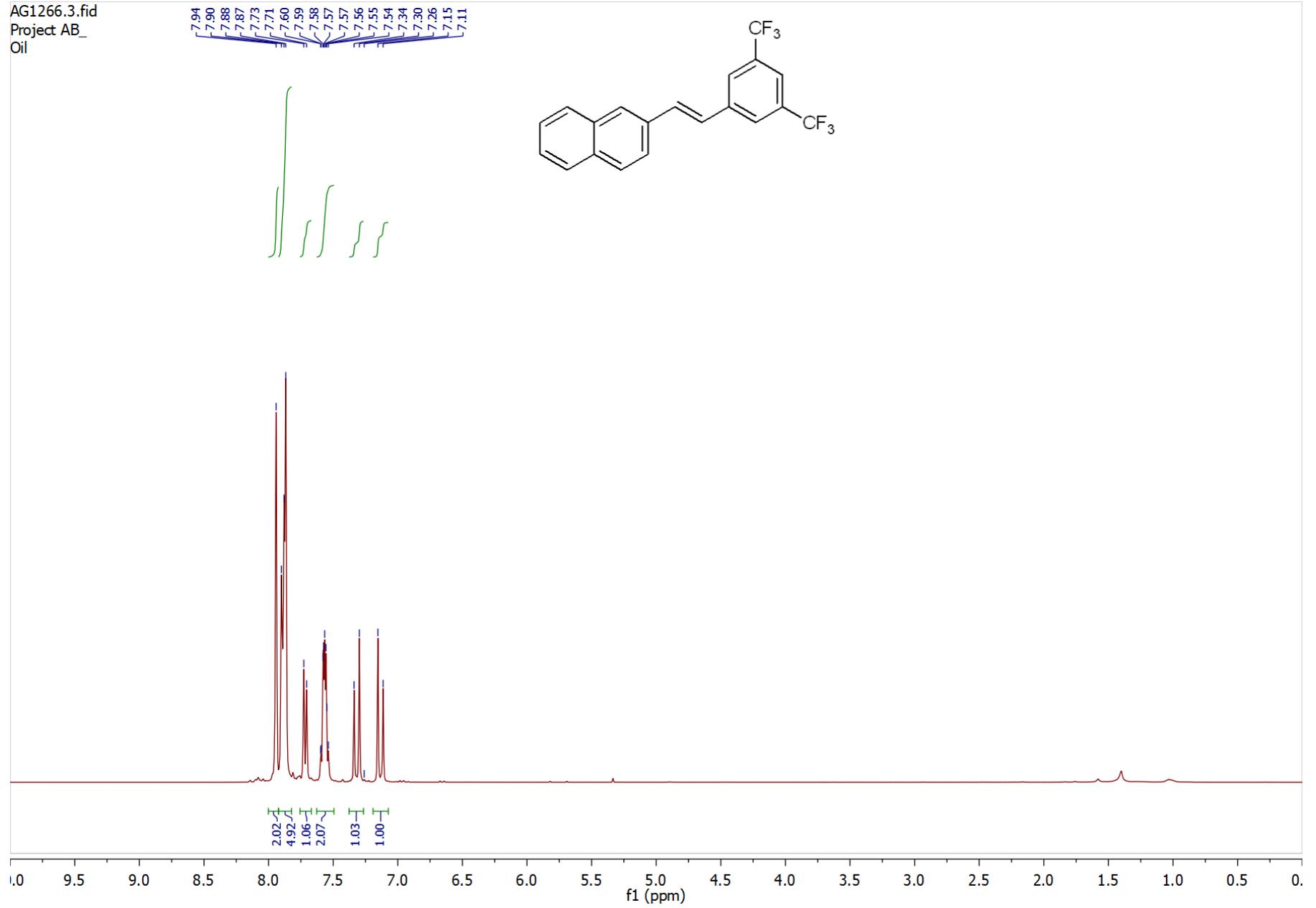
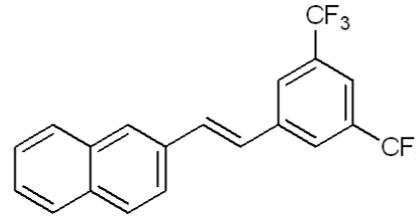
Compound 11e

AG1265.5.fid
Project AB_
Oil



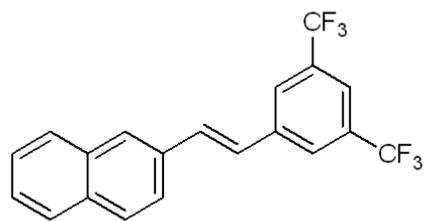
Compound 11f

AG1266.3.fid
Project AB_
Oil

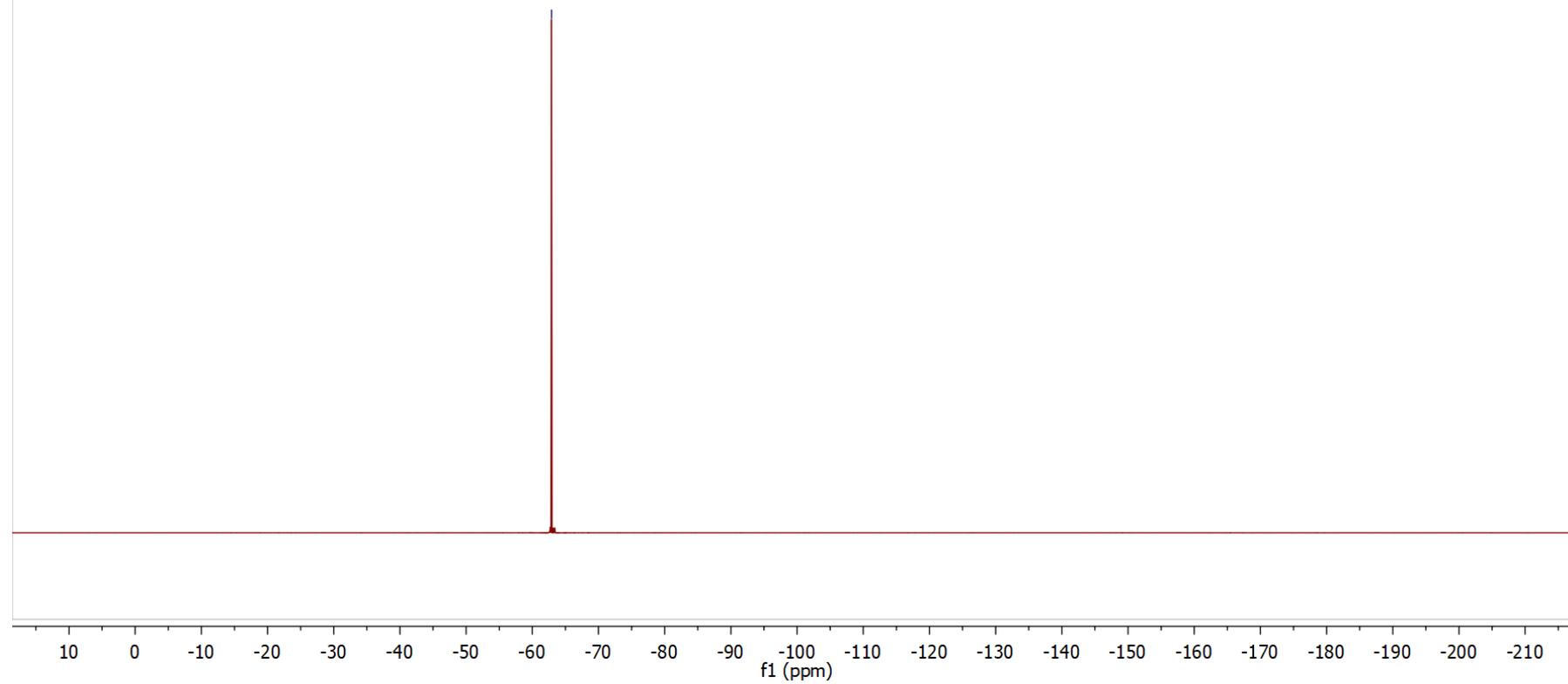


Compound 11f

AG1266.2.fid
Project AB_
Oil

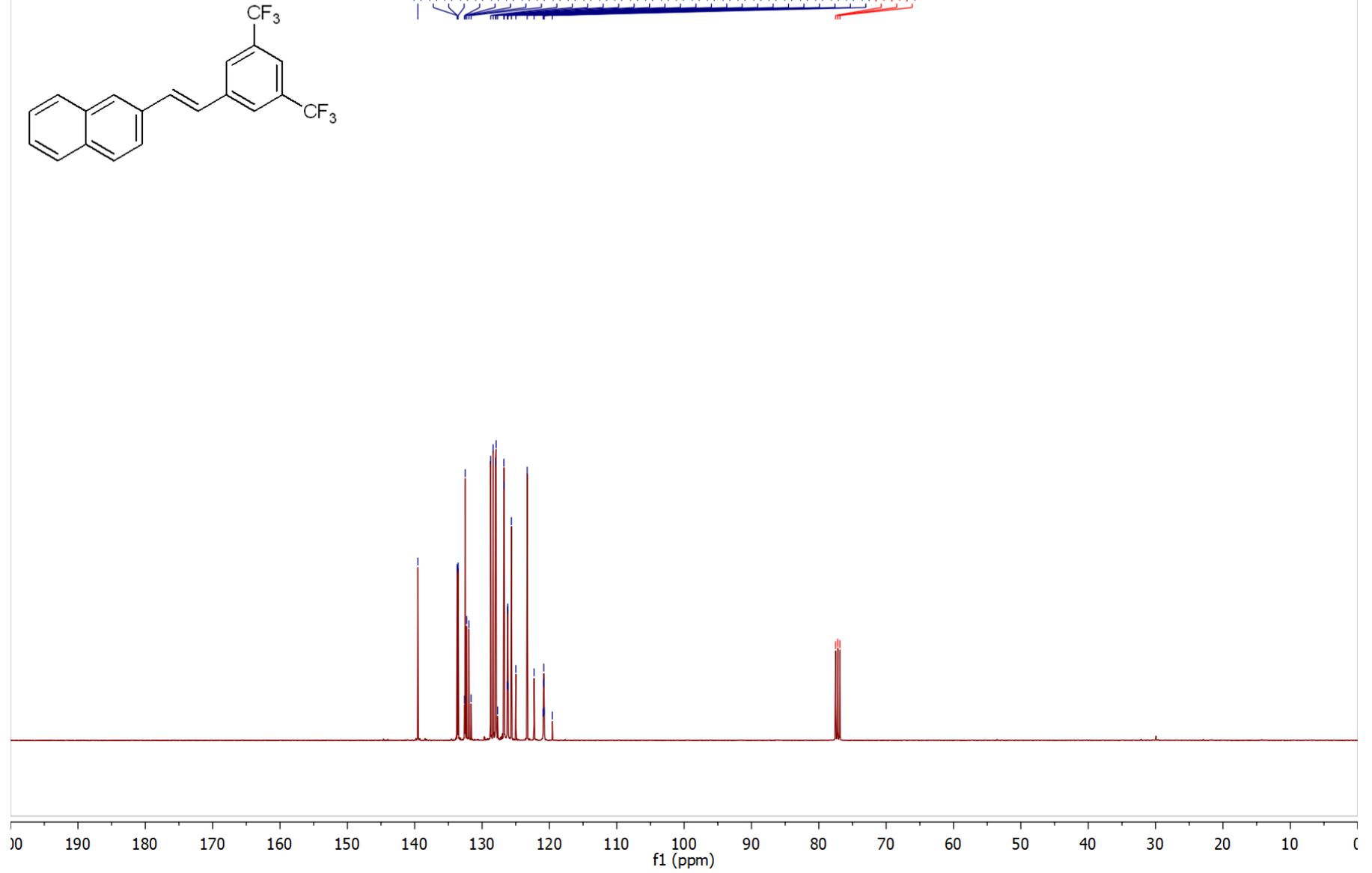


-62.91



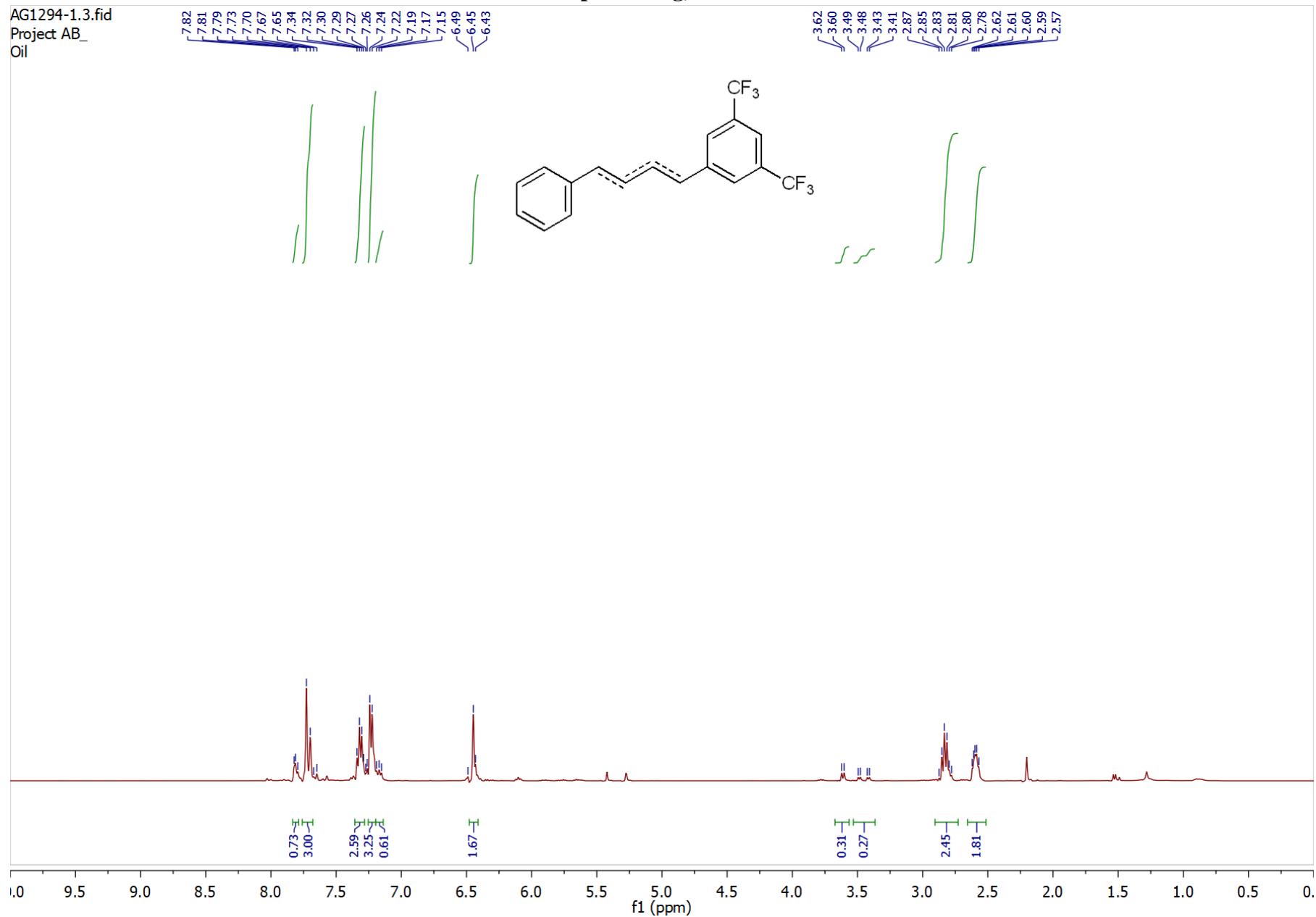
Compound 11f

AG1266.5.fid
Project AB_
Oil



Compound 11g, fraction 1

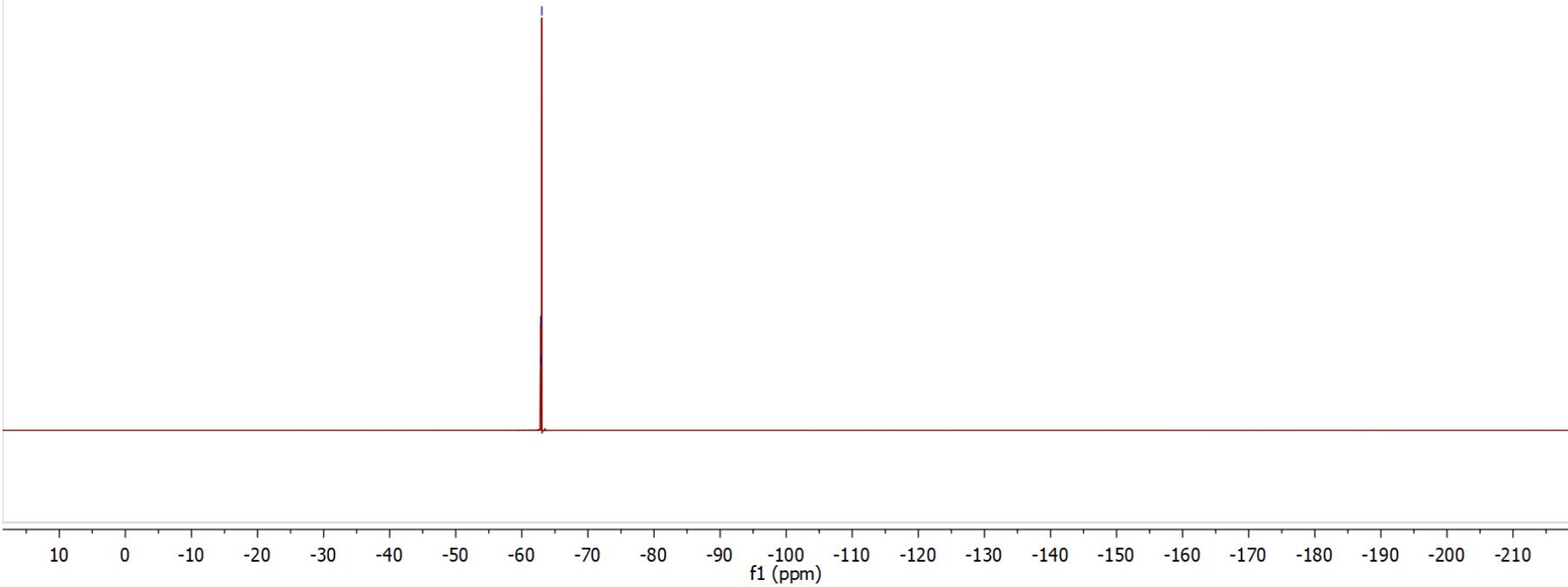
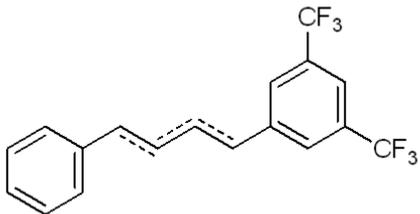
AG1294-1.3.fid
Project AB_
Oil



Compound 11g, fraction 1

AG1294-1.2.fid
Project AB_
Oil

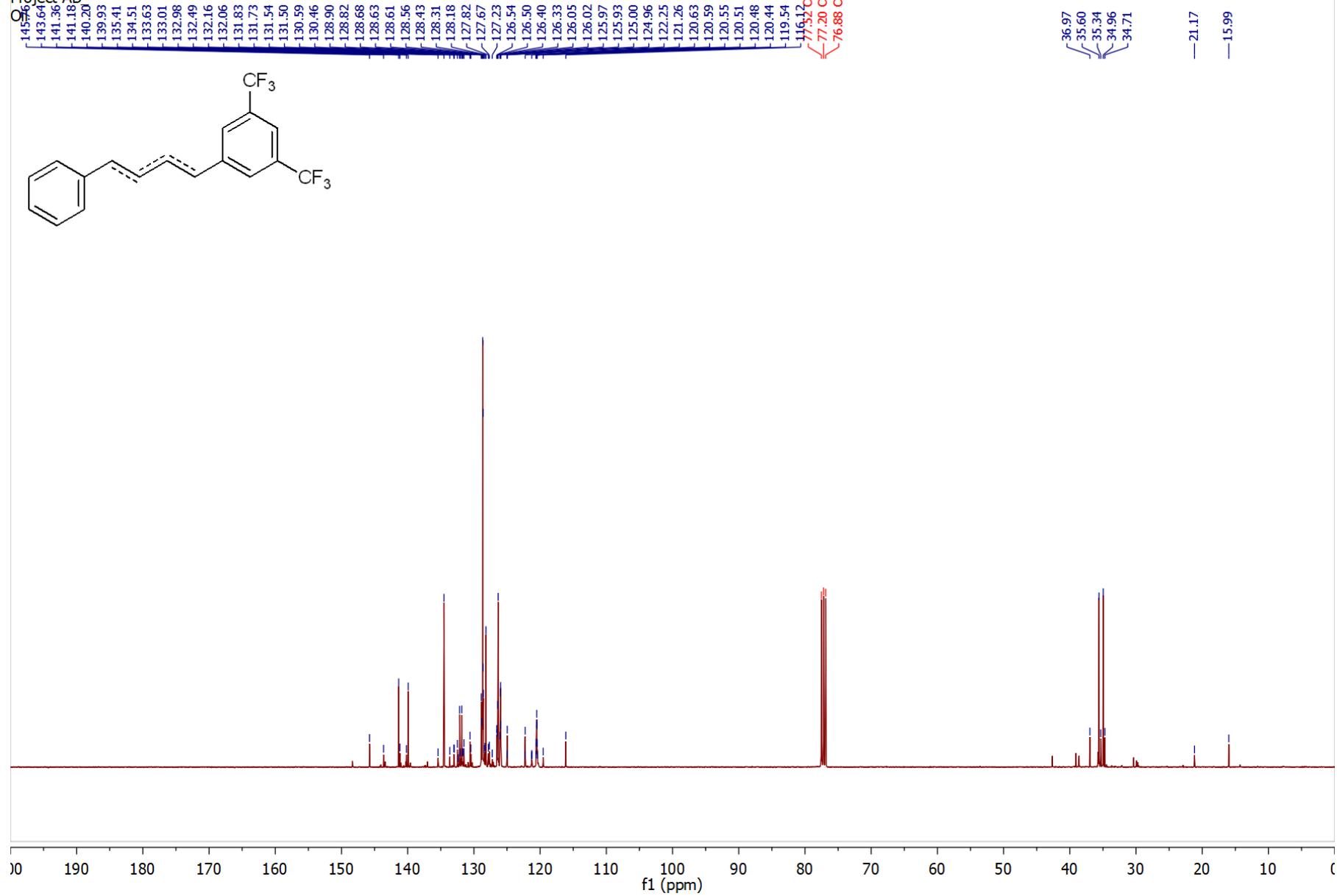
62.85
62.89
63.03



Compound 11g, fraction 1

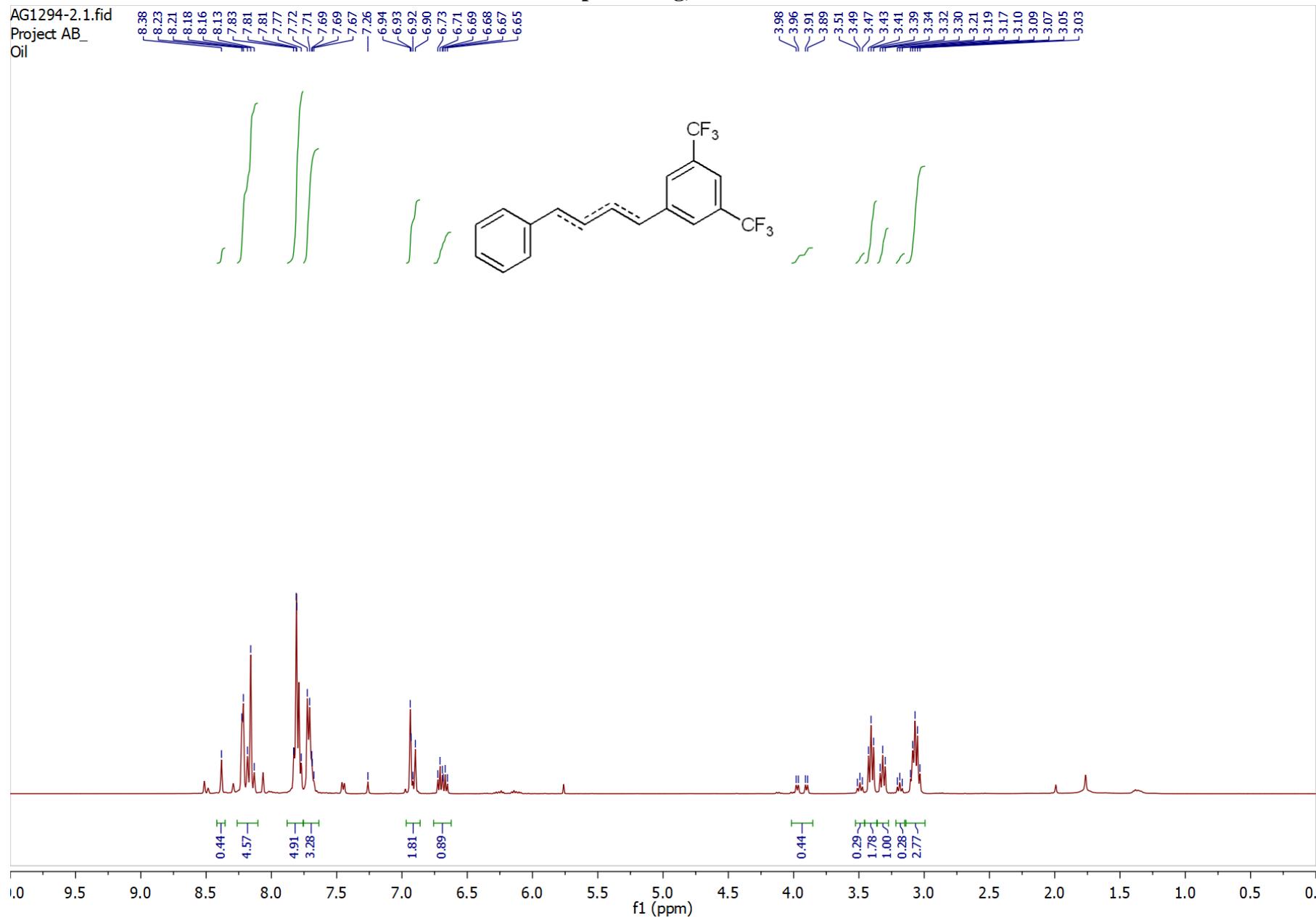
AG1294-1.5.fid

Project AB



Compound 11g, fraction 2

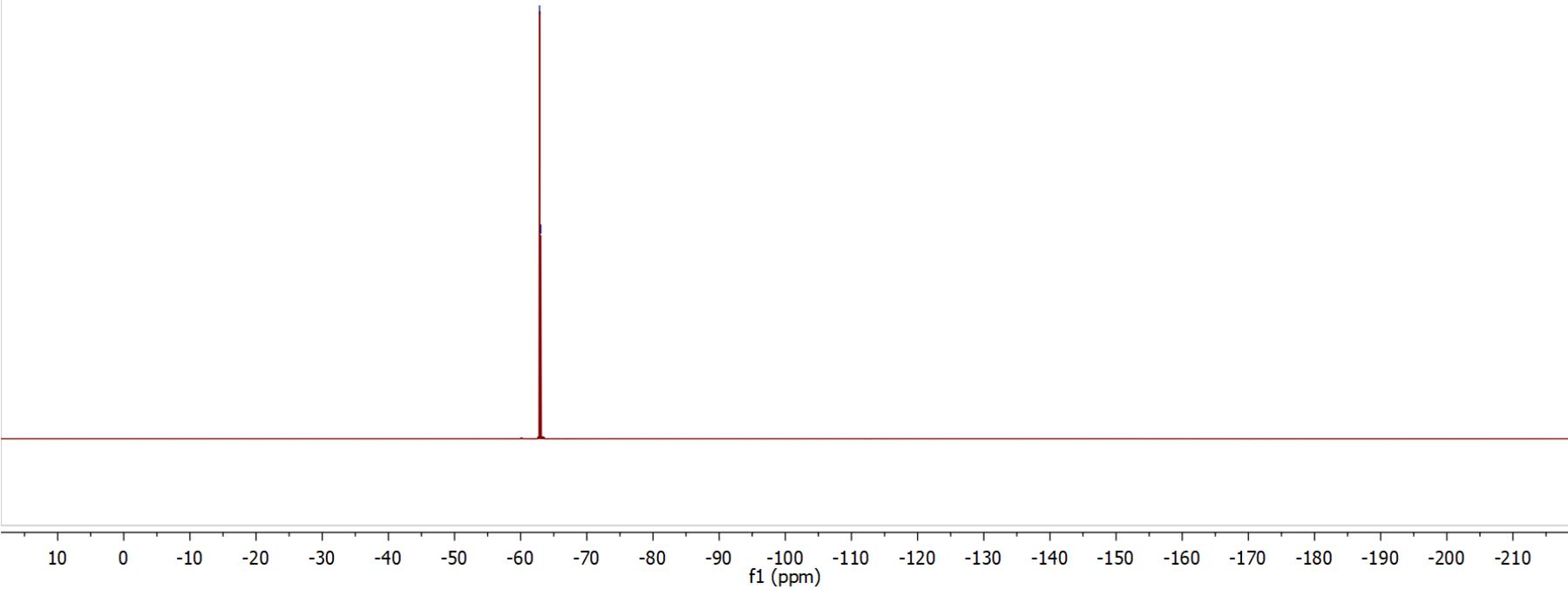
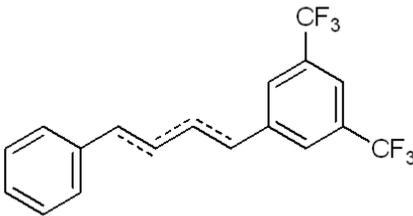
AG1294-2.1.fid
Project AB_
Oil



Compound 11g, fraction 2

AG1294-2.2.fid
Project AB_
Oil

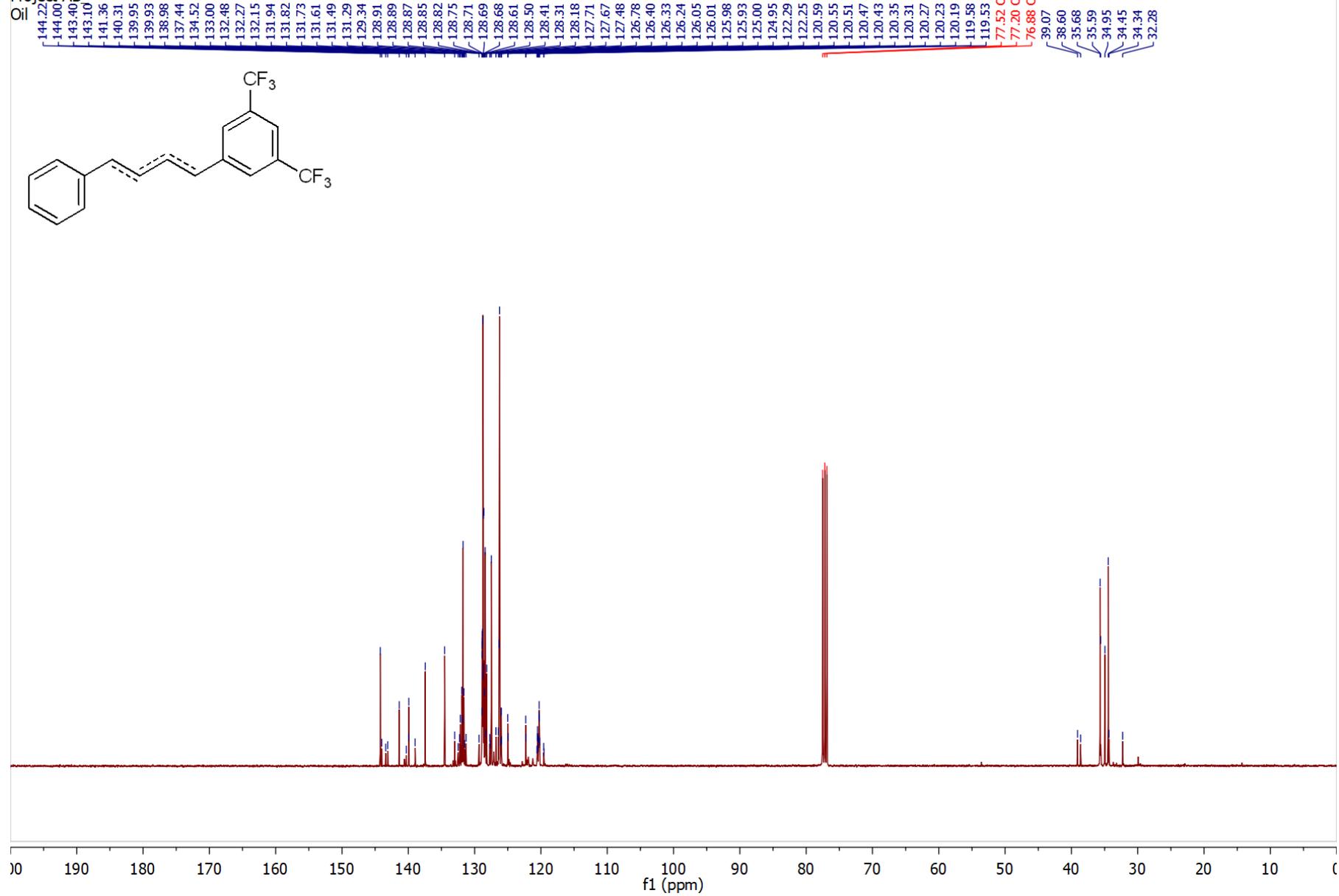
-62.85
-63.03



Compound 11g, fraction 2

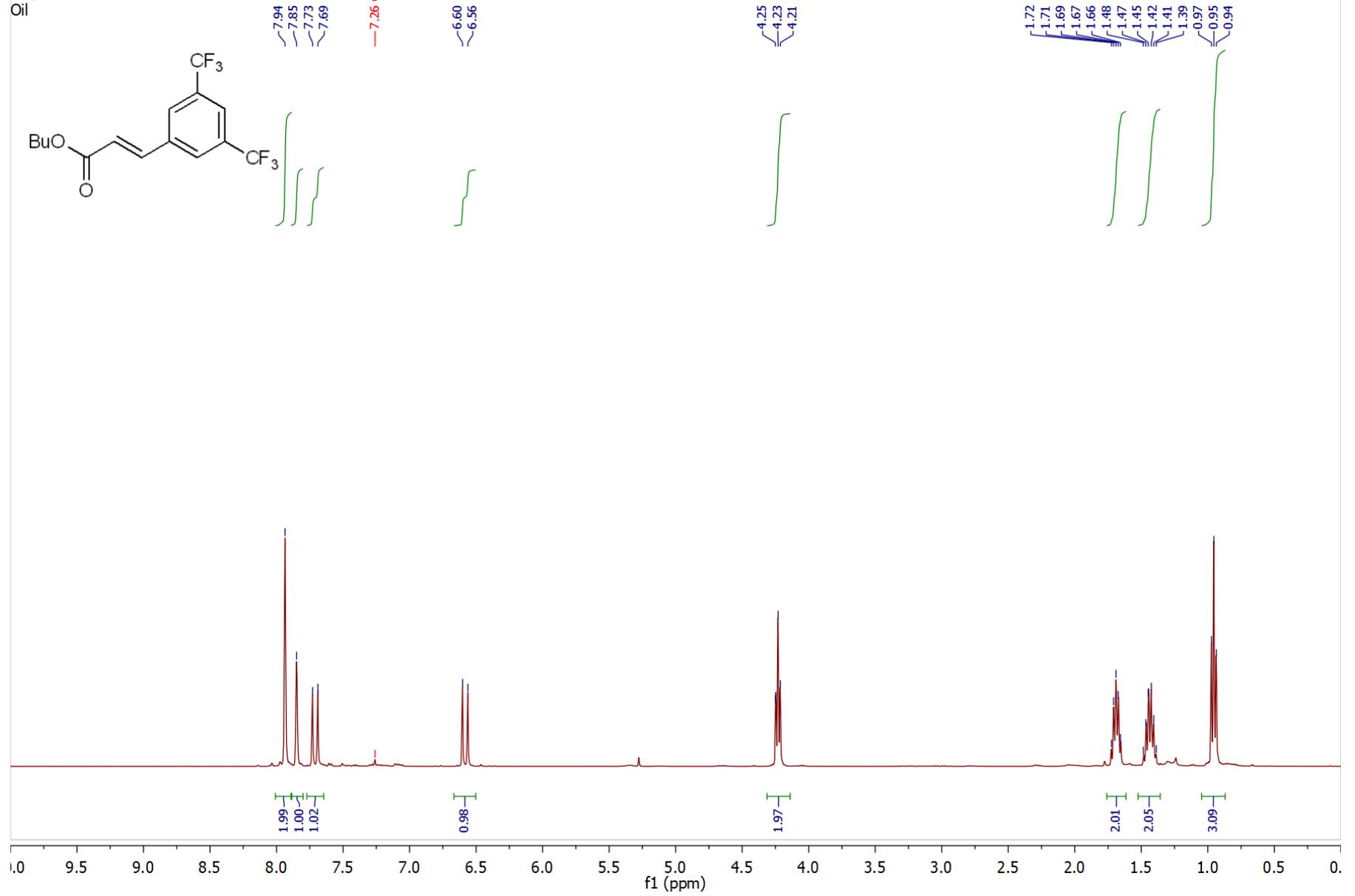
AG1294-2.5.fid

Project AB



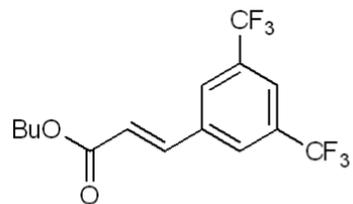
Compound 11h

AG1295.1.fid
Project AB_
Oil

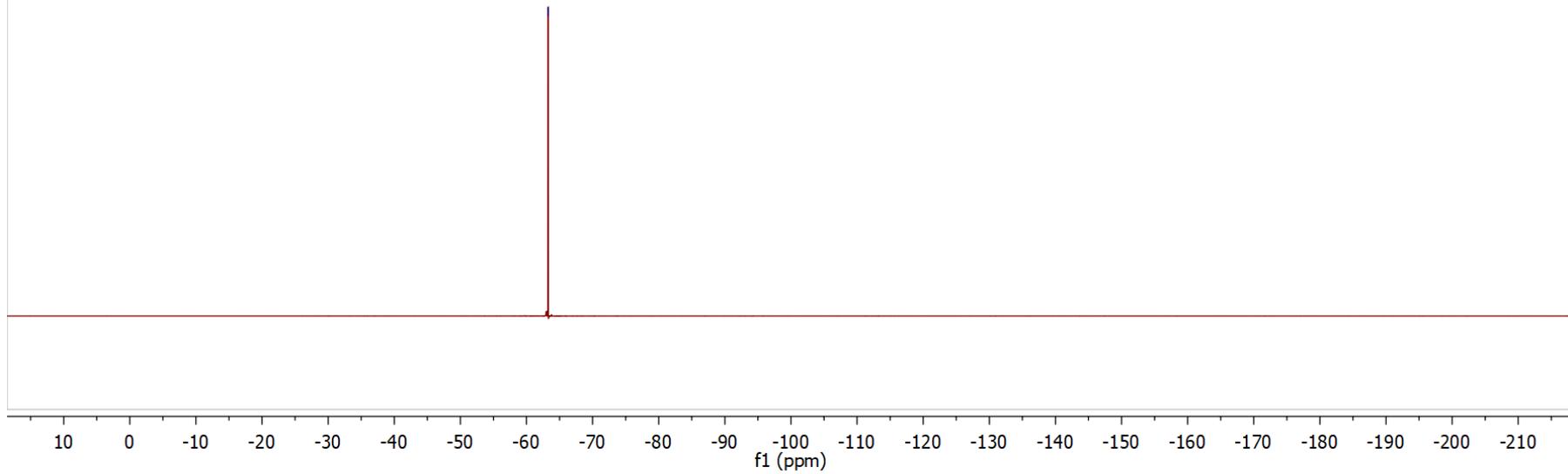


Compound 11h

AG1295.2.fid
Project AB_
Oil

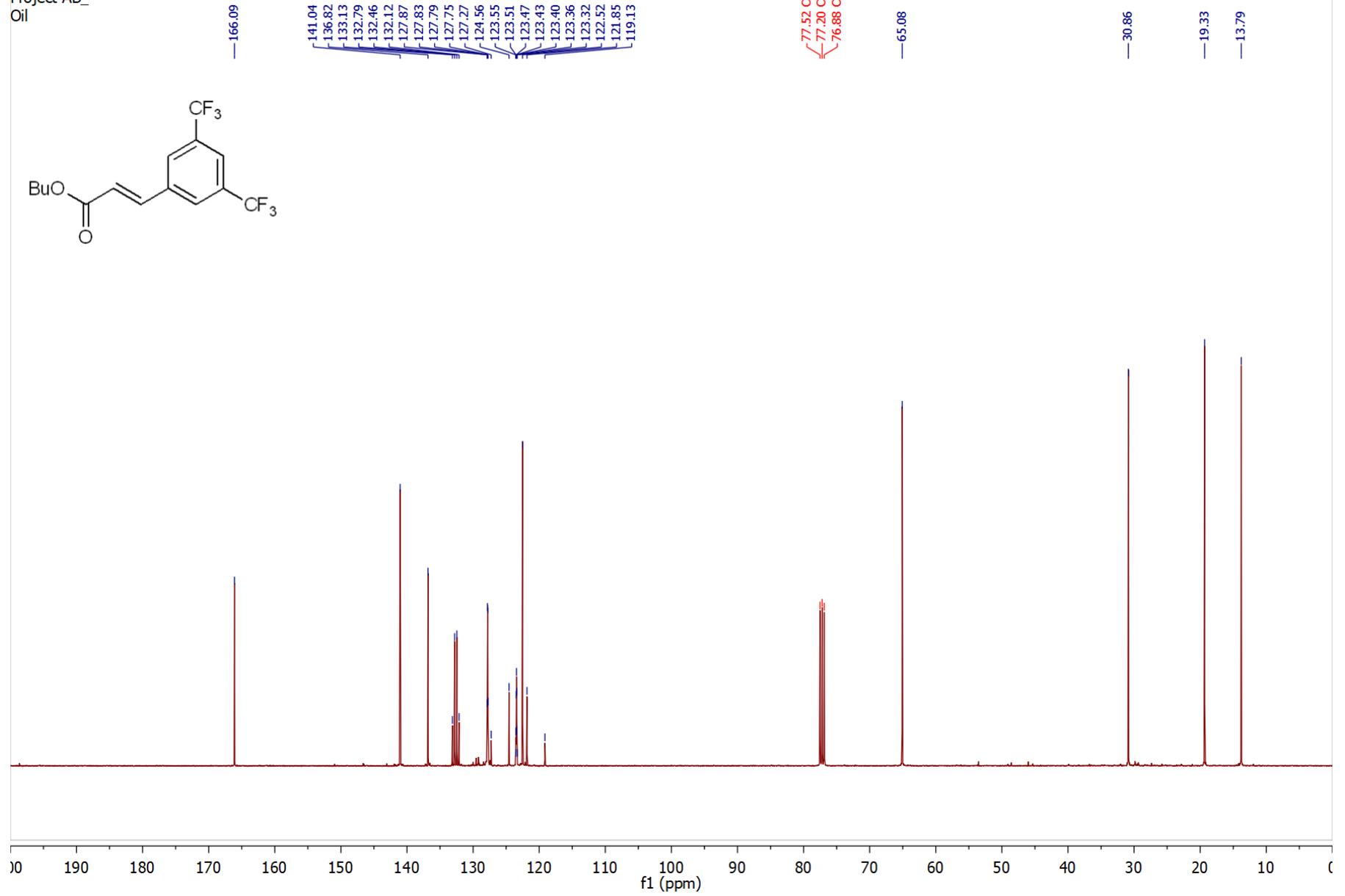
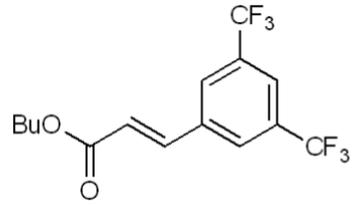


-63.29



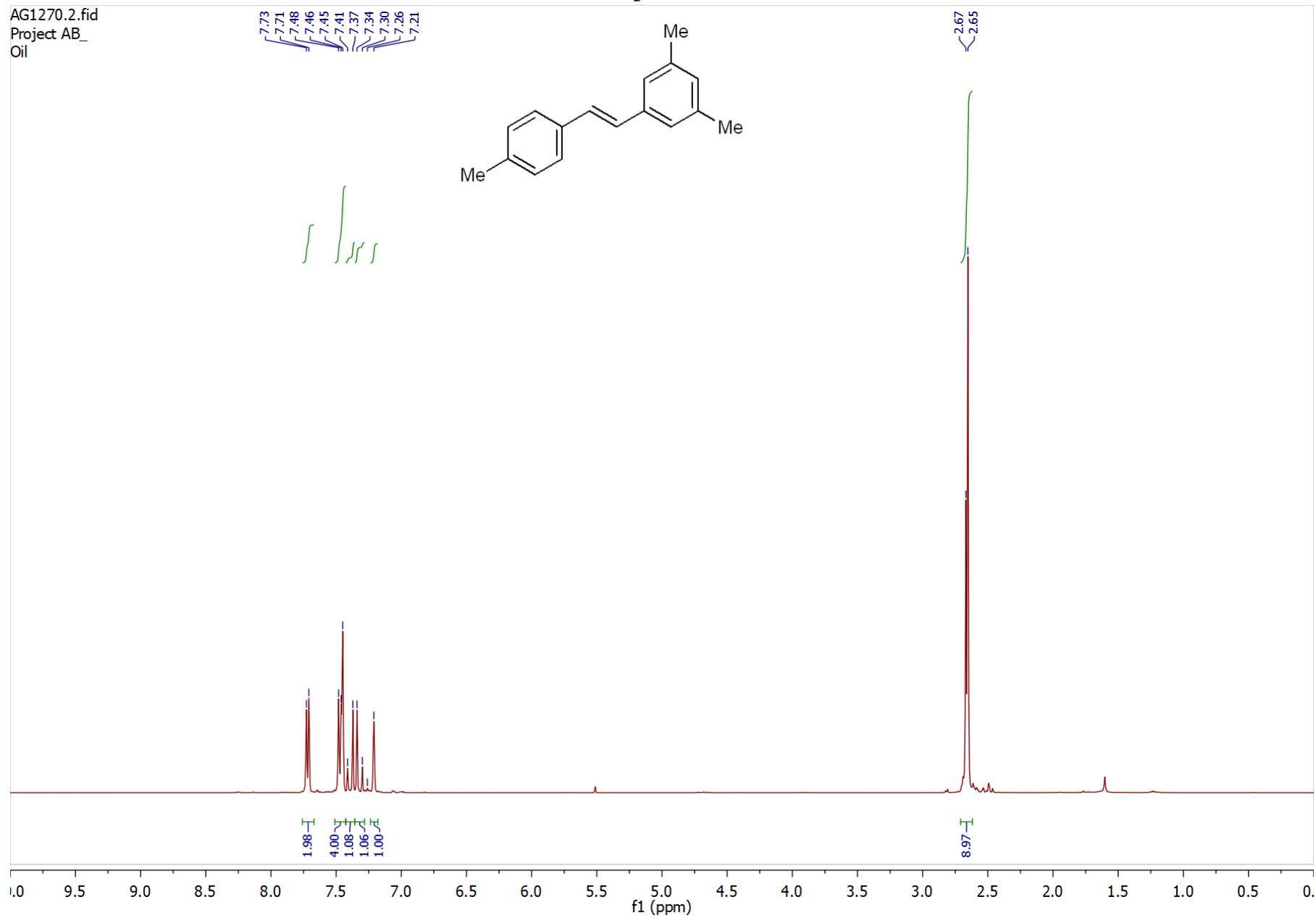
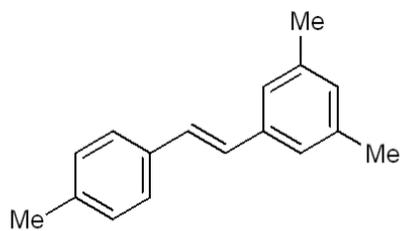
Compound 11h

AG1295.5.fid
Project AB_
Oil



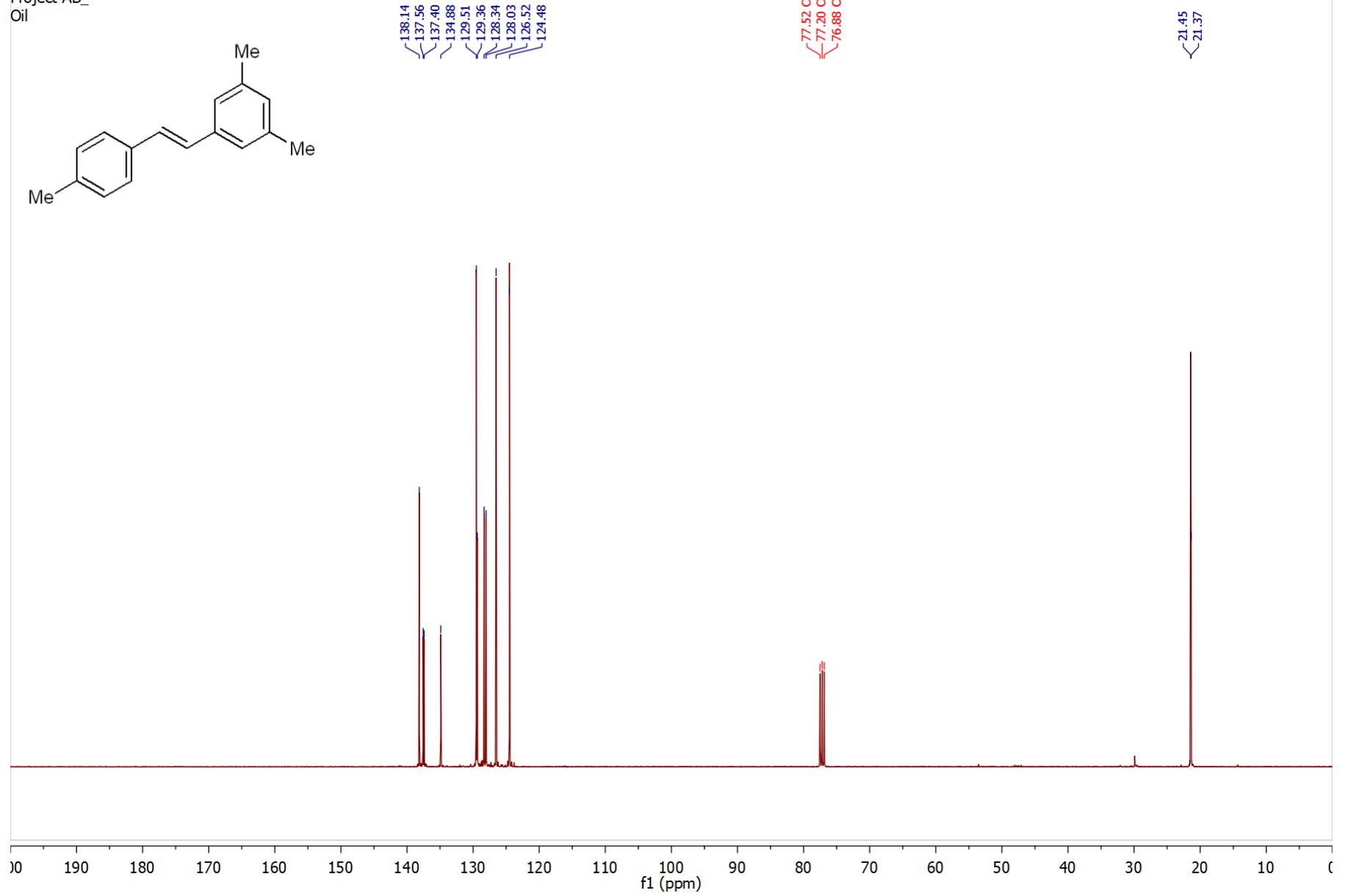
Compound 11k

AG1270.2.fid
Project AB_
Oil



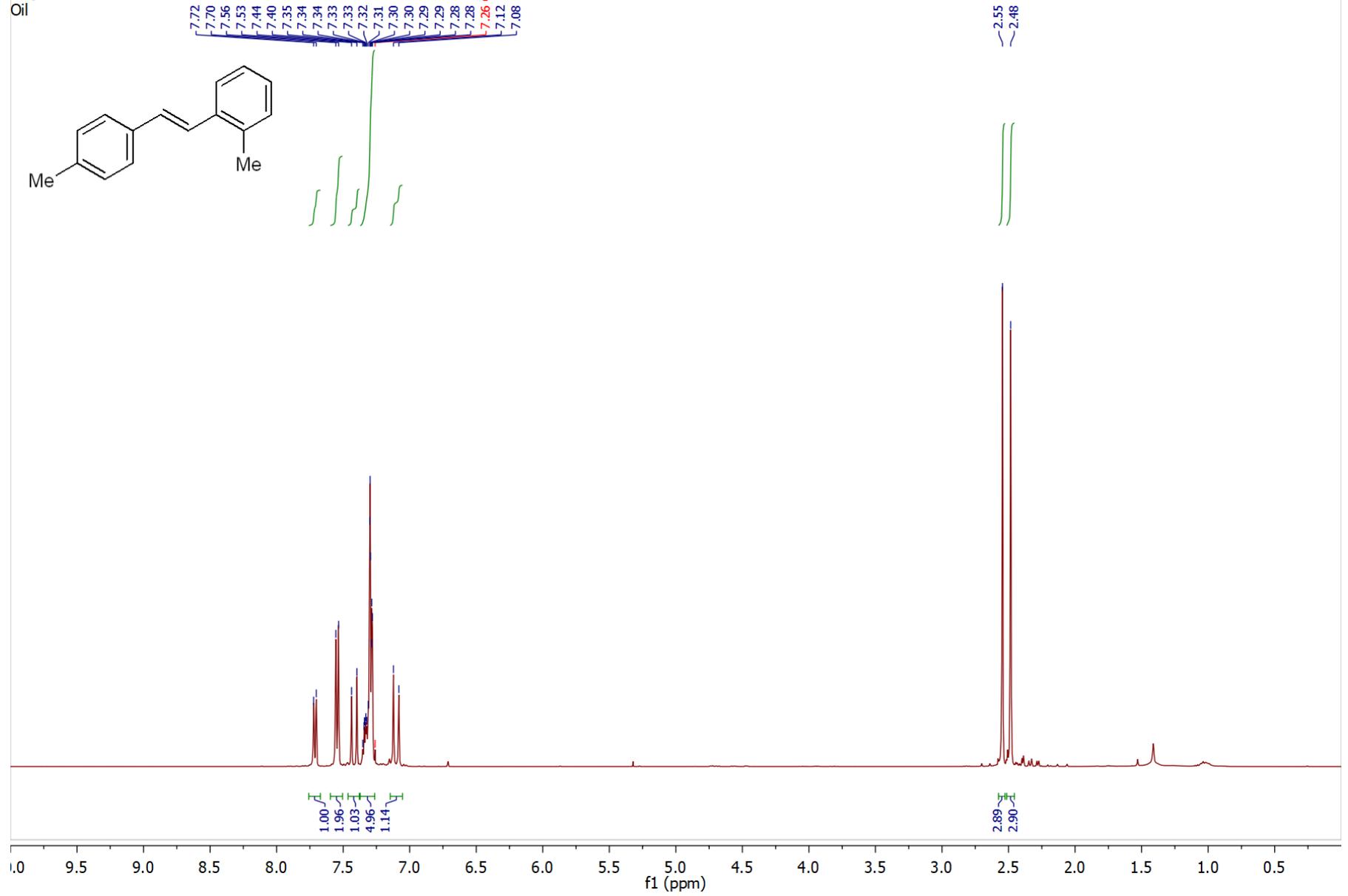
Compound 11k

AG1270.4.fid
Project AB_
Oil



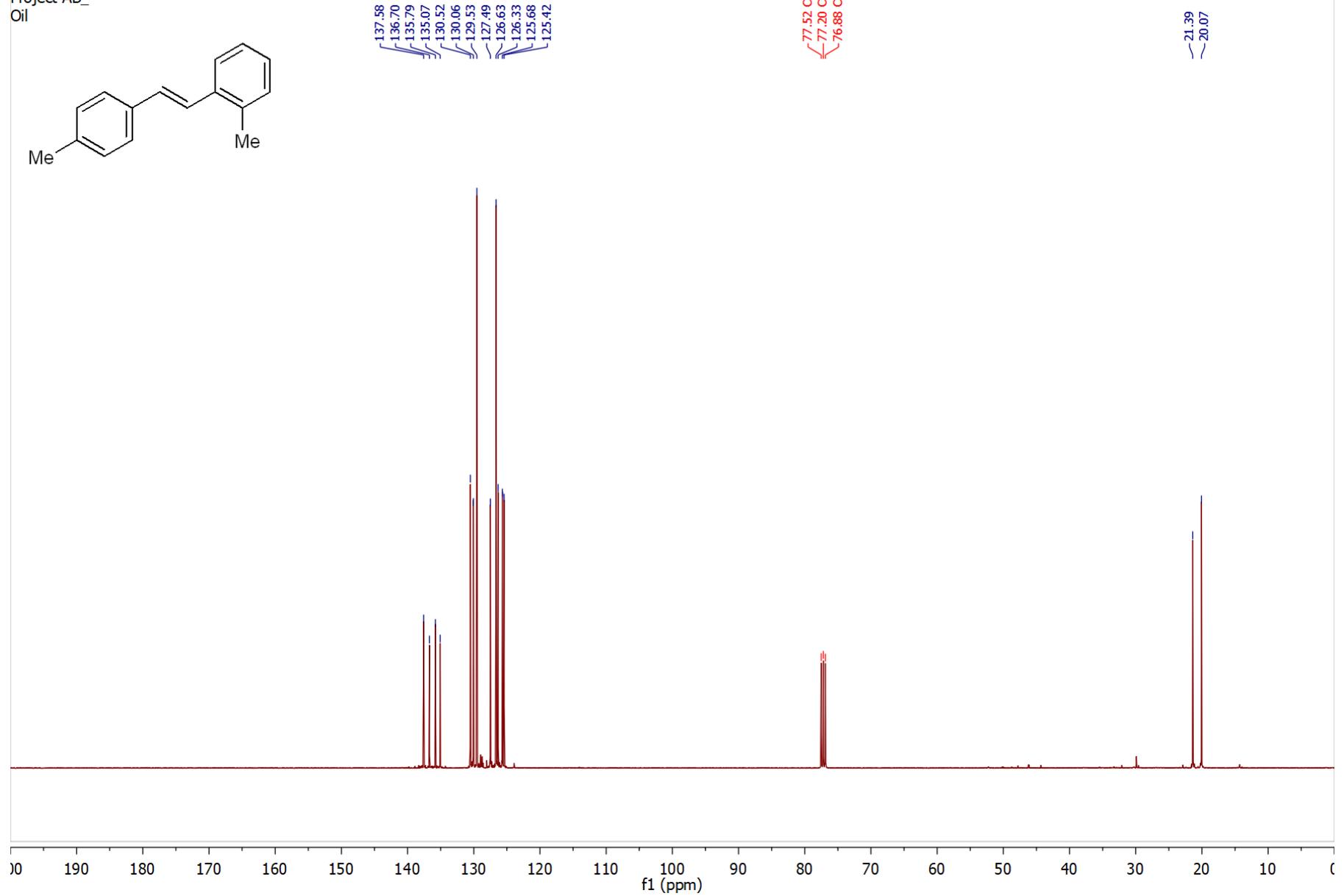
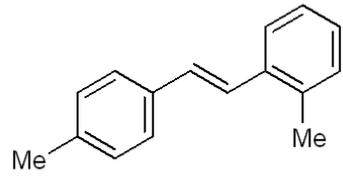
Compound 111

AG1303.2.fid
Project AB_
Oil



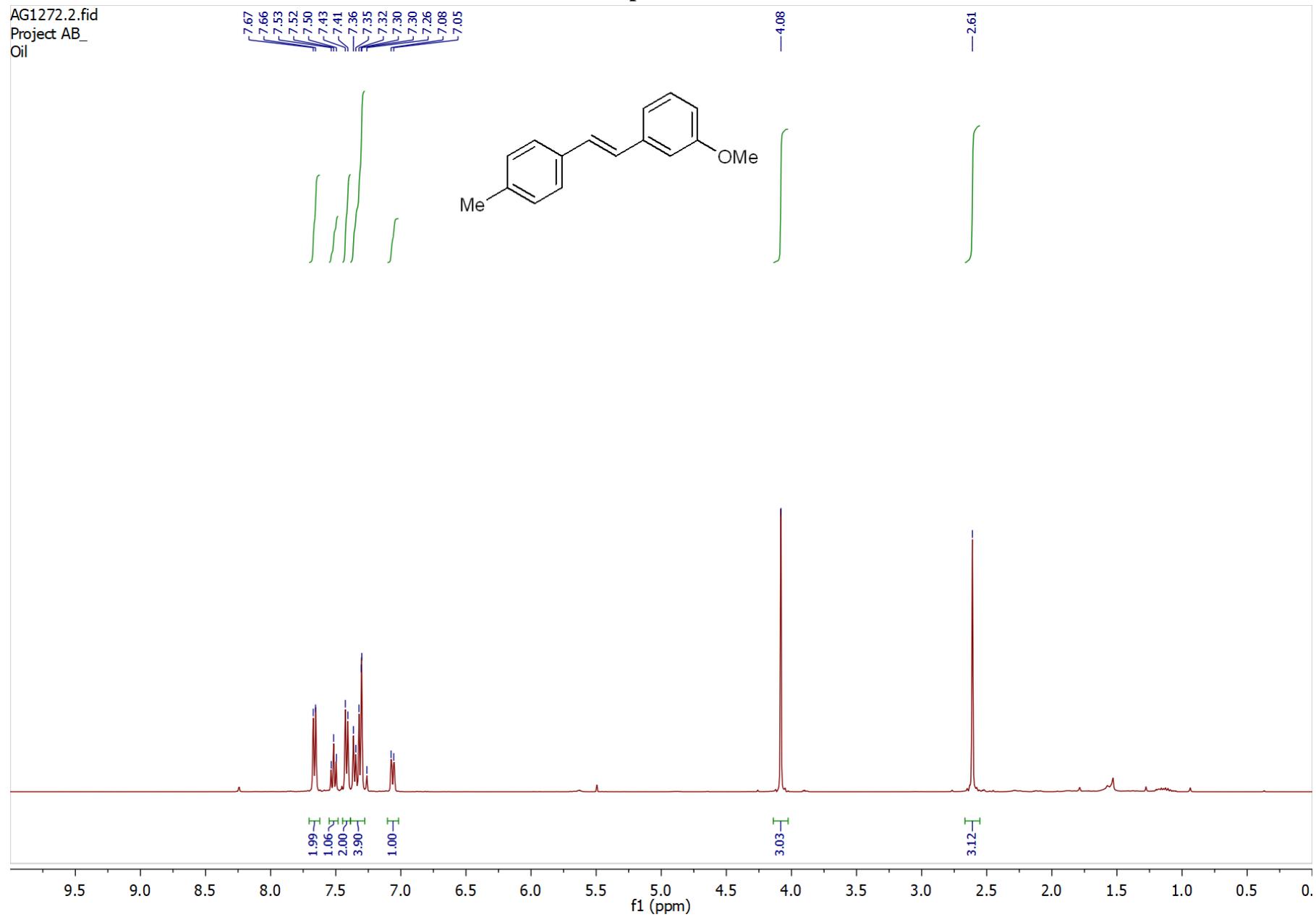
Compound 111

AG1303.4.fid
Project AB_
Oil



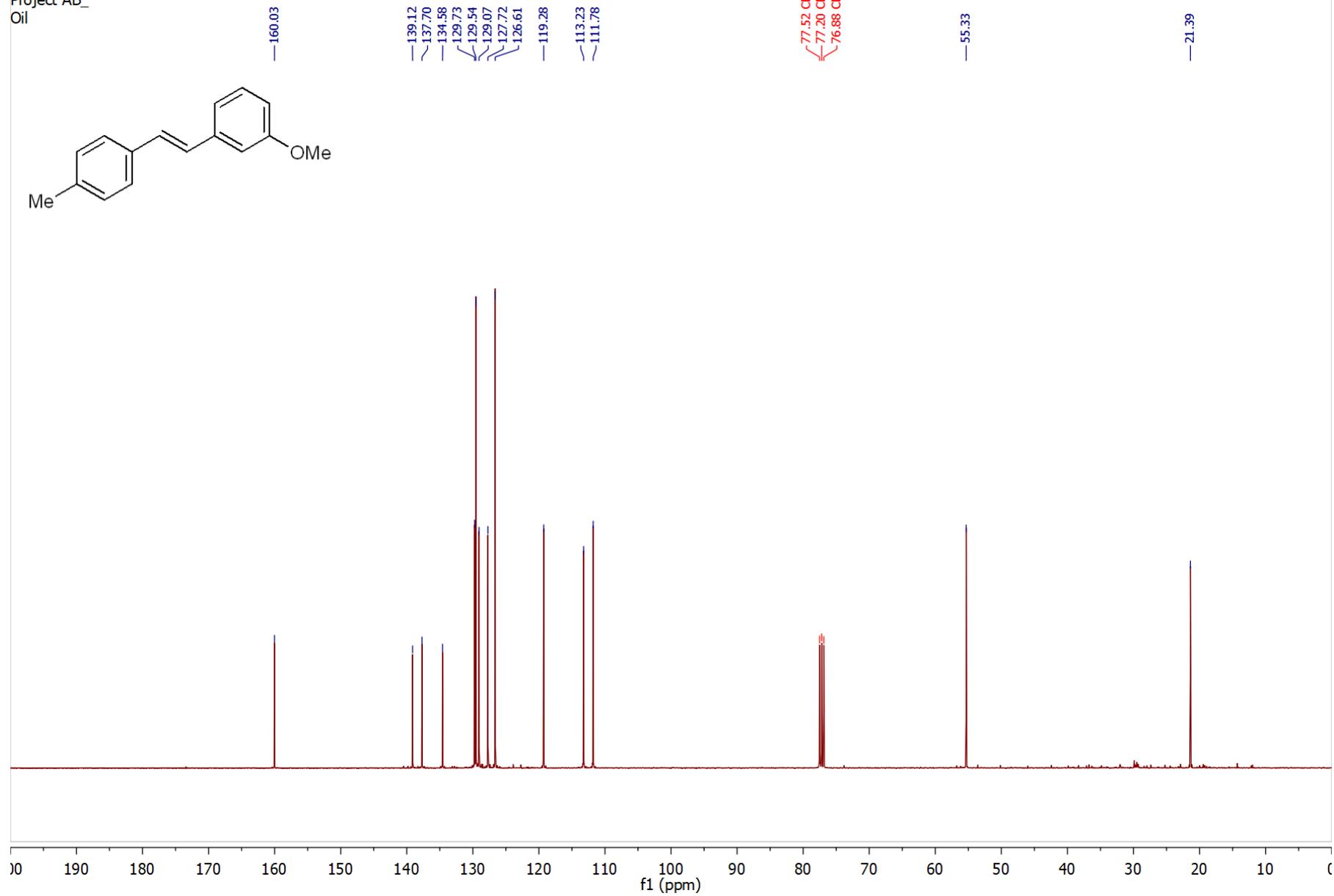
Compound 11m

AG1272.2.fid
Project AB_
Oil



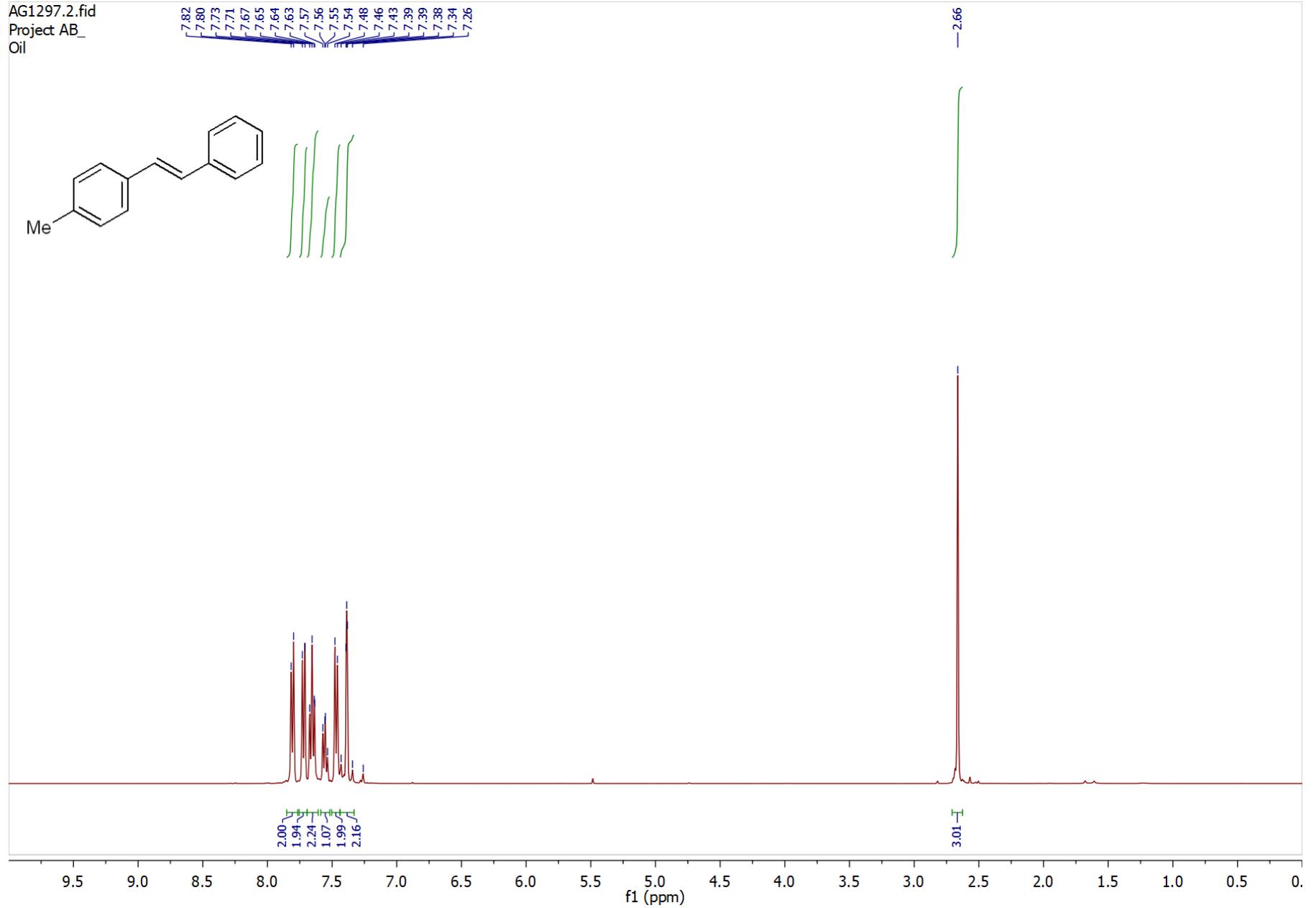
Compound 11m

AG1272.4.fid
Project AB_
Oil



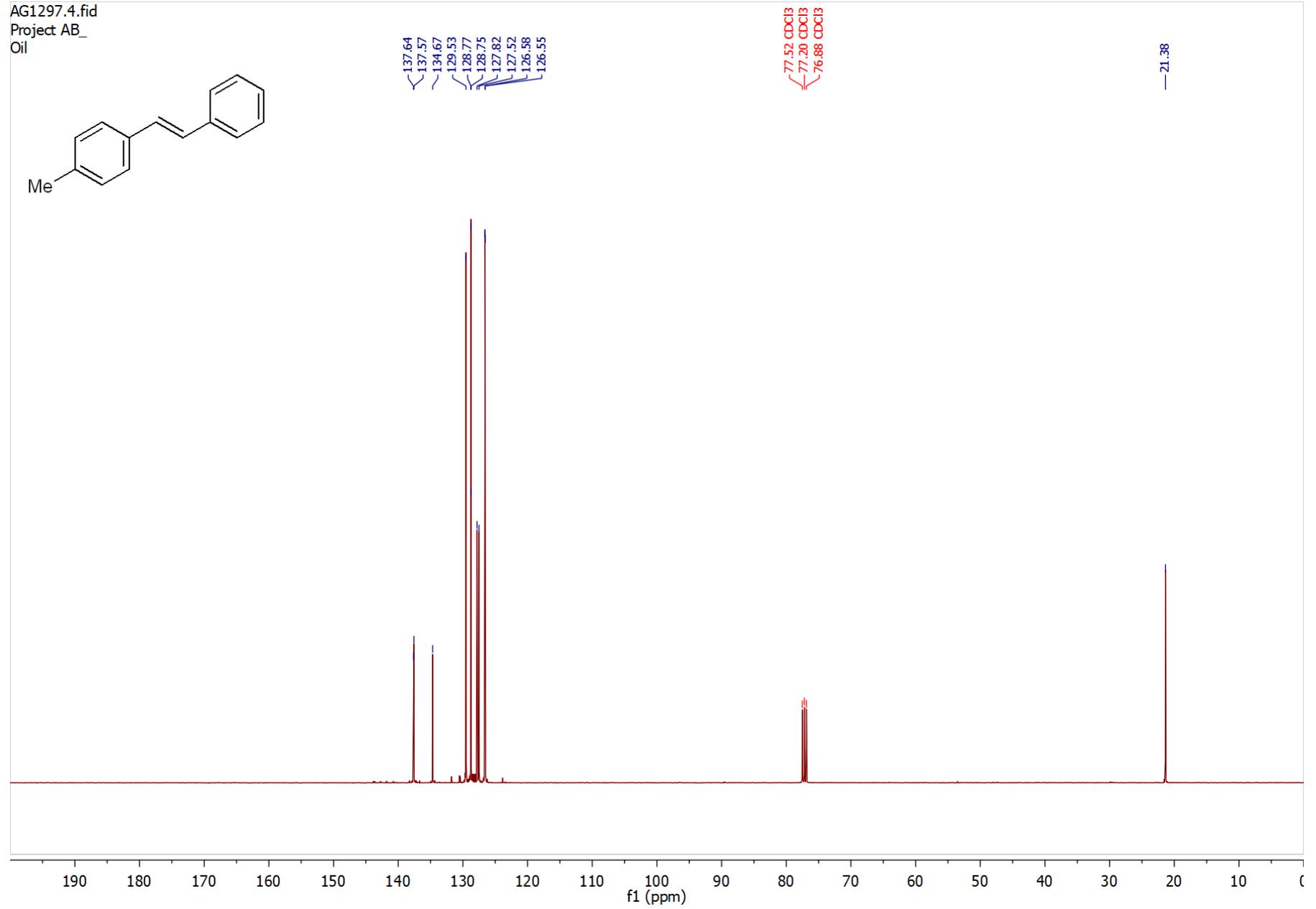
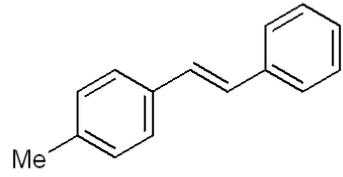
Compound 11n

AG1297.2.fid
Project AB_
Oil



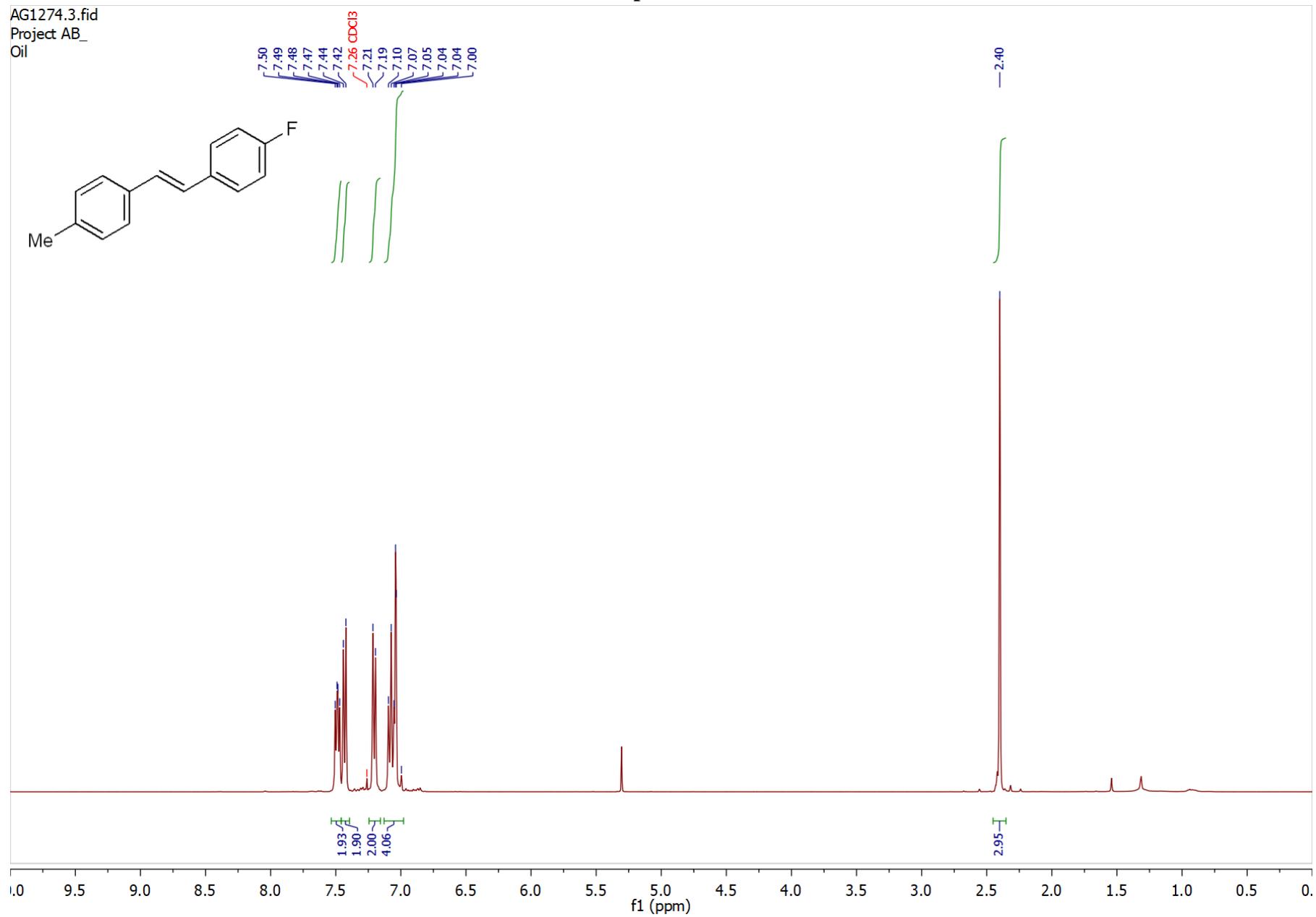
Compound 11n

AG1297.4.fid
Project AB_
Oil



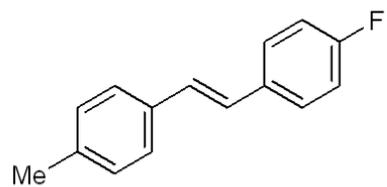
Compound 11o

AG1274.3.fid
Project AB_
Oil

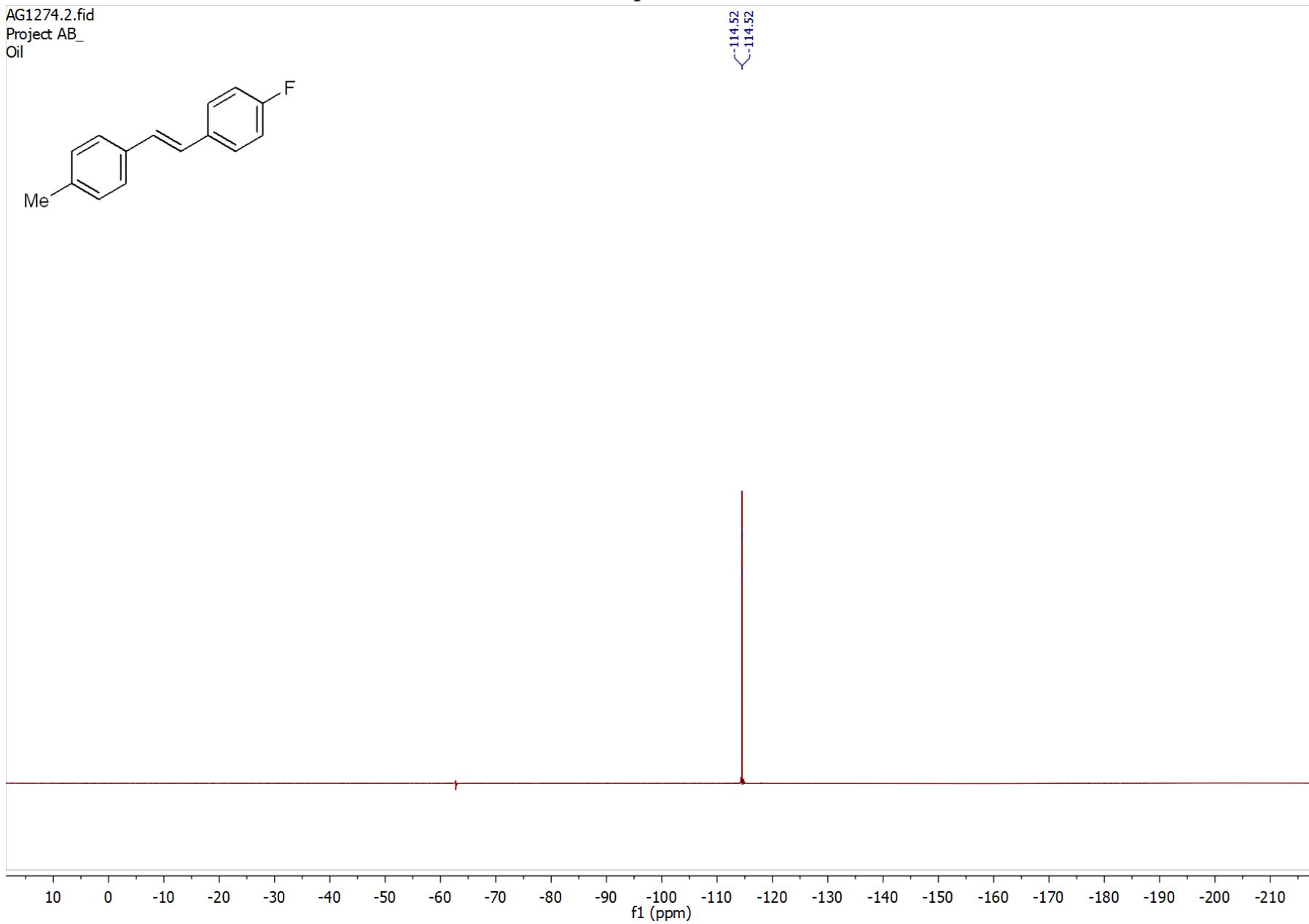


Compound 11o

AG1274.2.fid
Project AB_
Oil

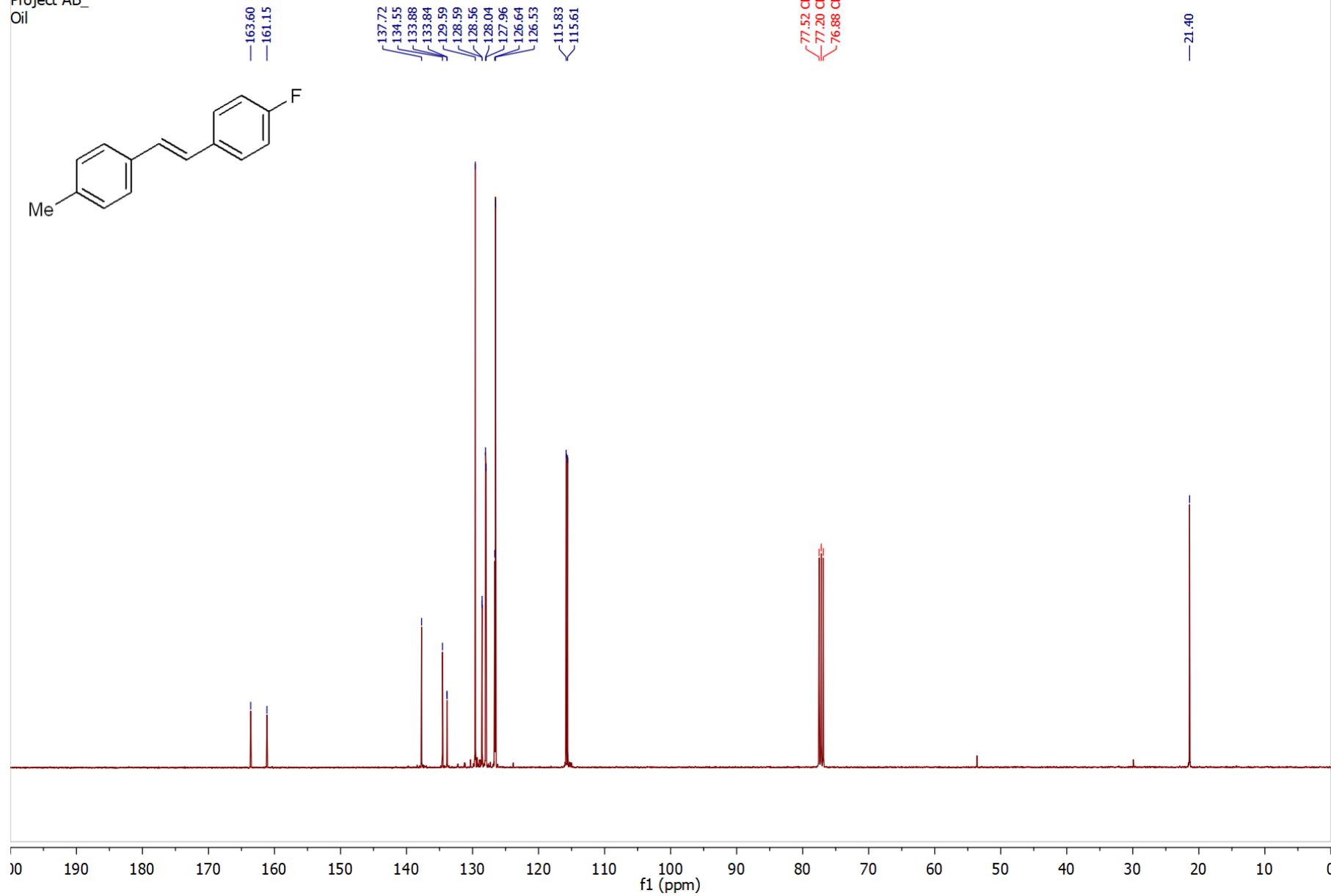


114.52
114.52



Compound 11o

AG1274.5.fid
Project AB_
Oil

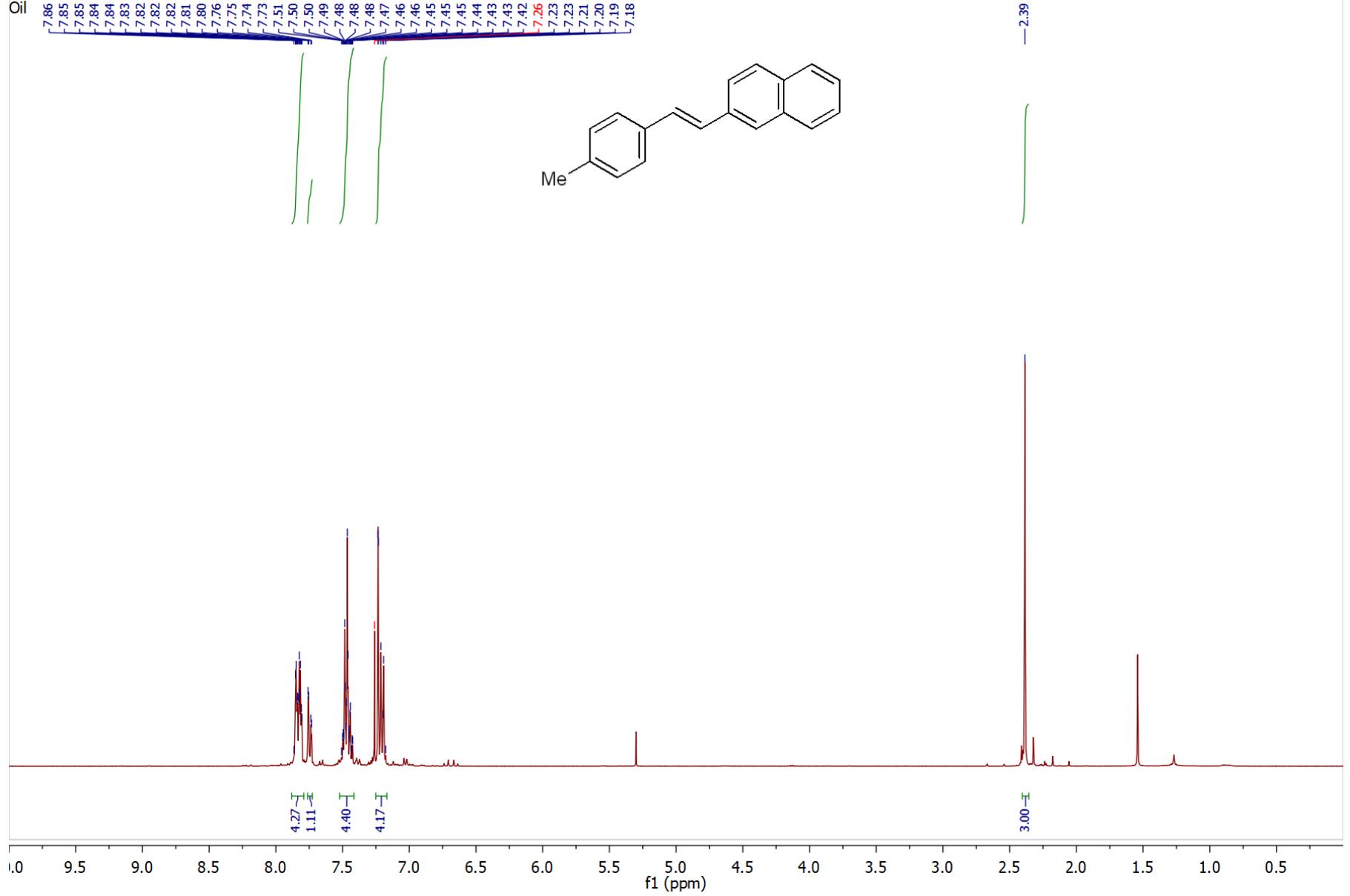


Compound 11p

AG1275.2.fid

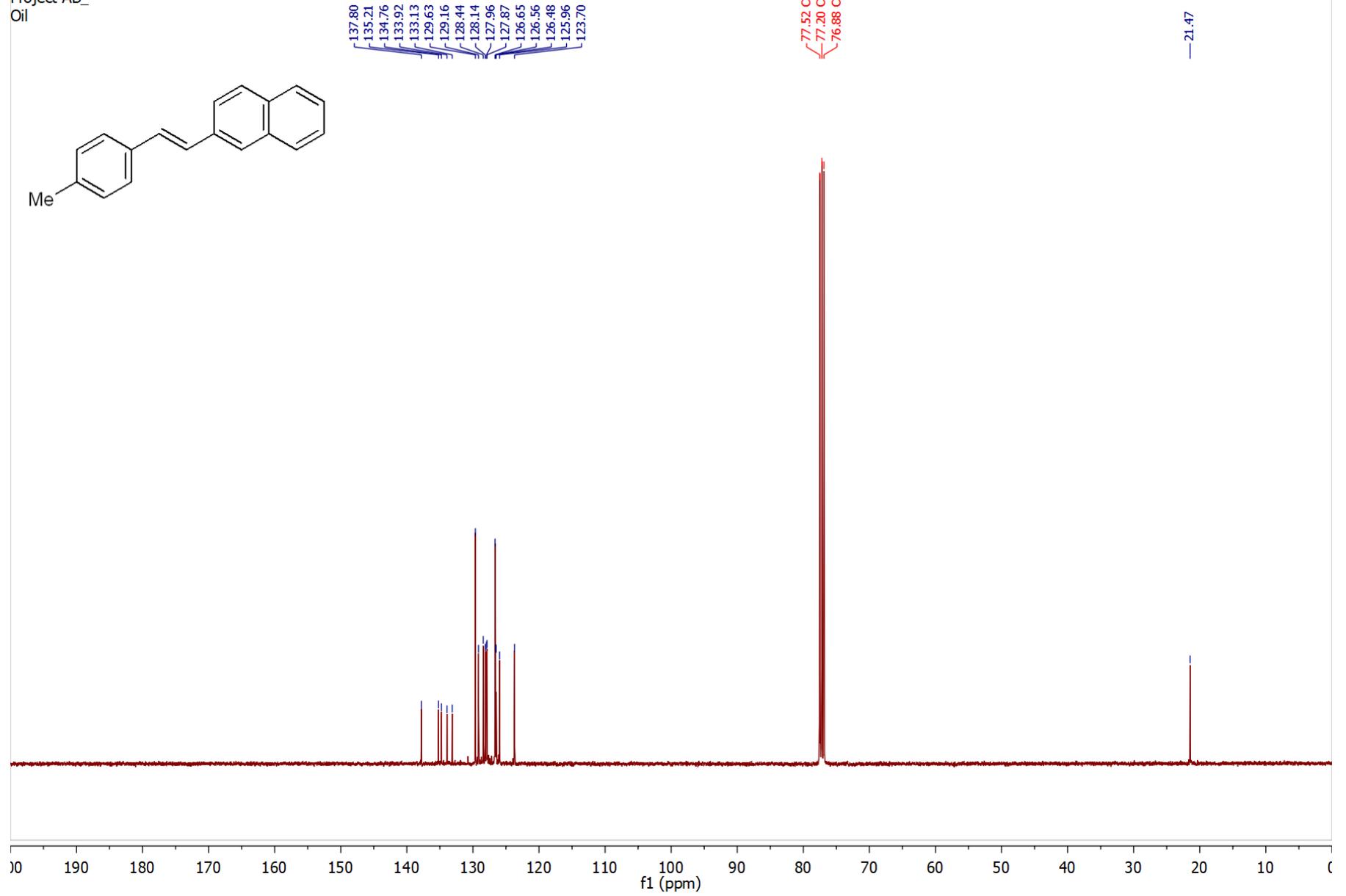
Project AB

Oil



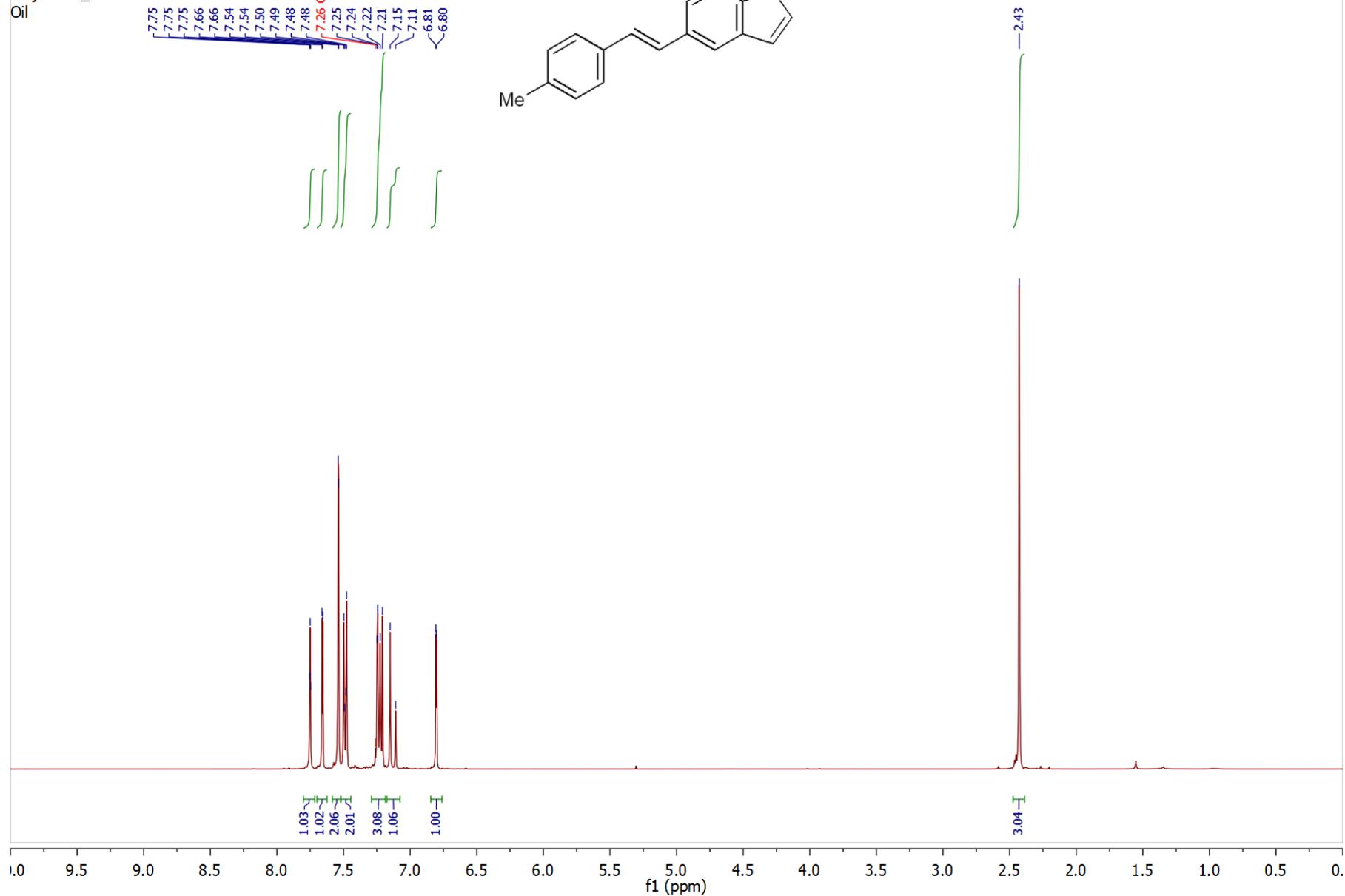
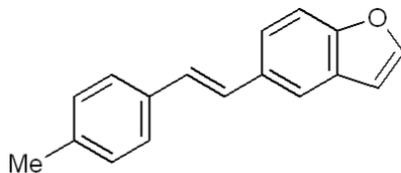
Compound 11p

AG1275.4.fid
Project AB_
Oil



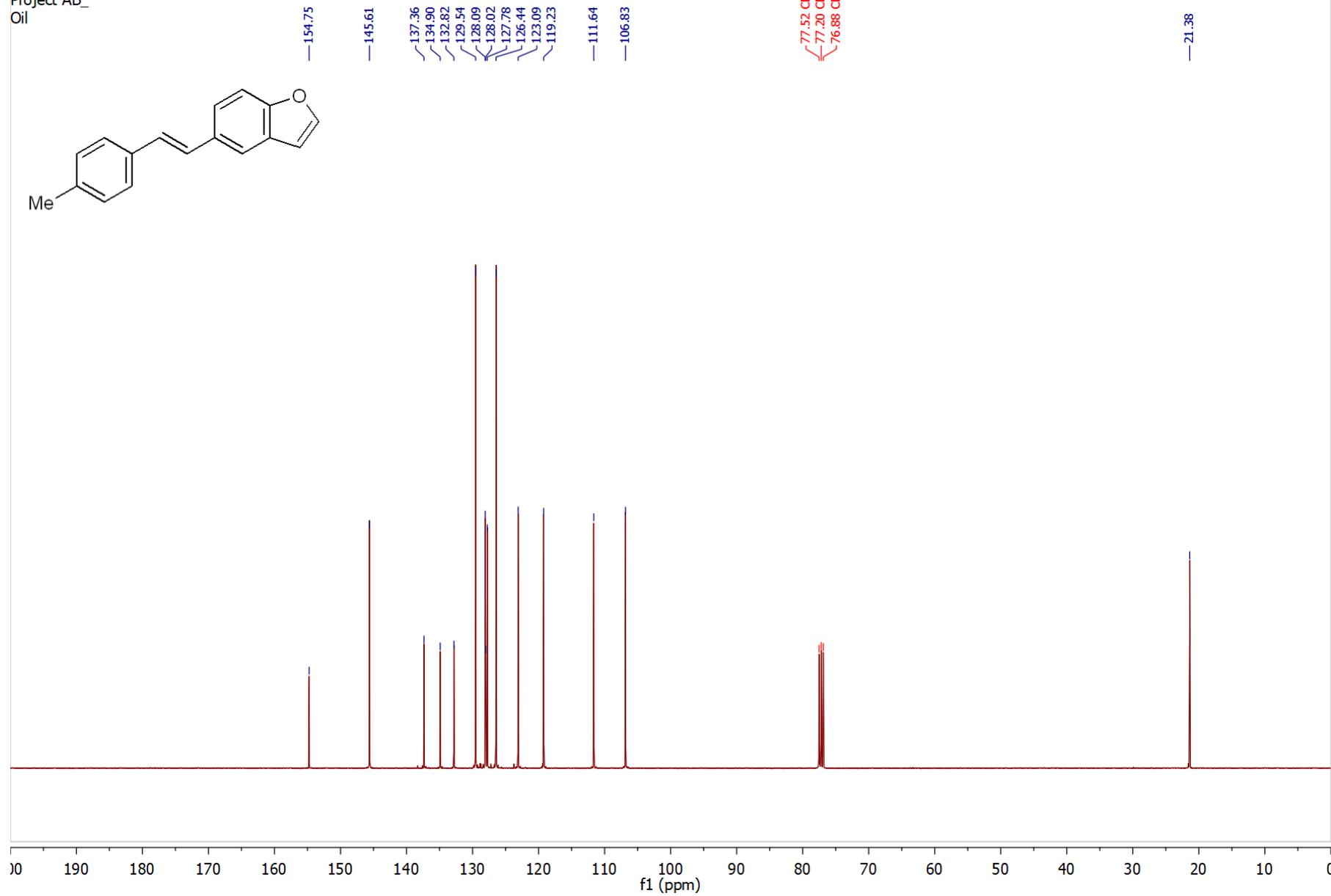
Compound 11q

AG1309.2.fid
Project AB_
Oil



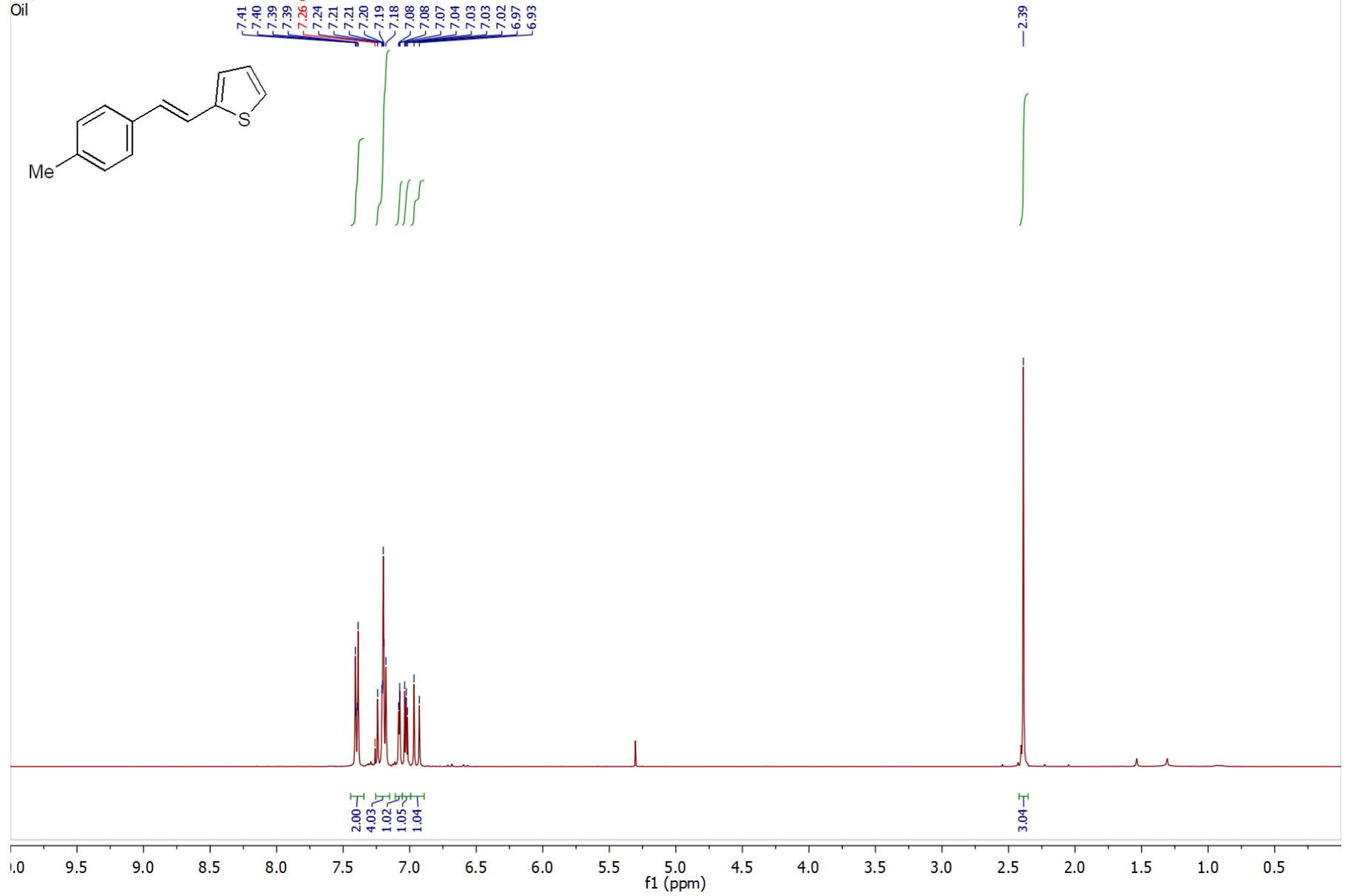
Compound 11q

AG1309.4.fid
Project AB_
Oil



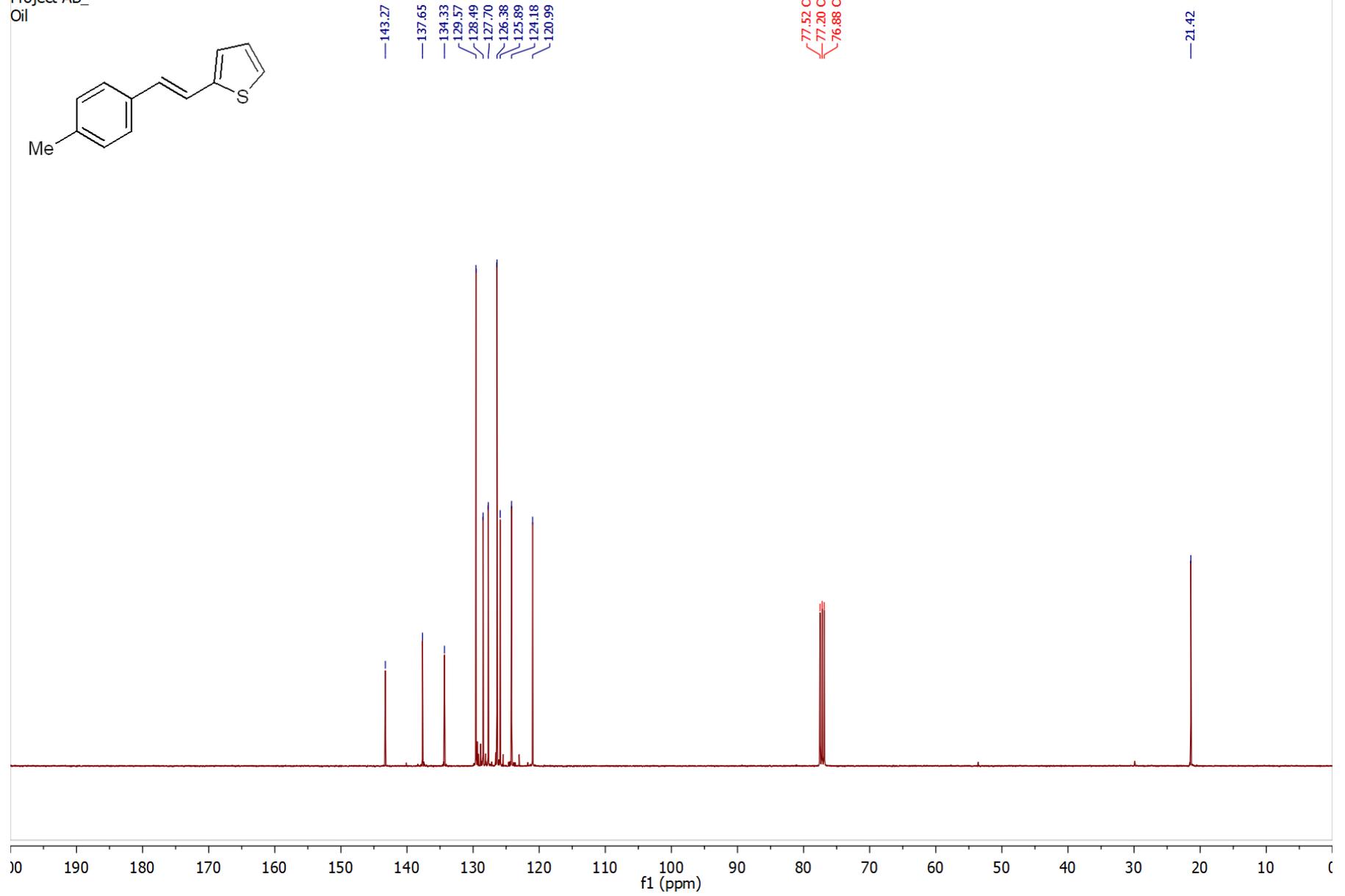
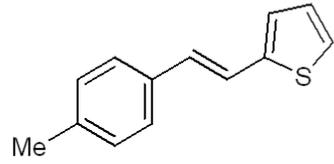
Compound 11r

AG1310.1.fid
Project AB_
Oil



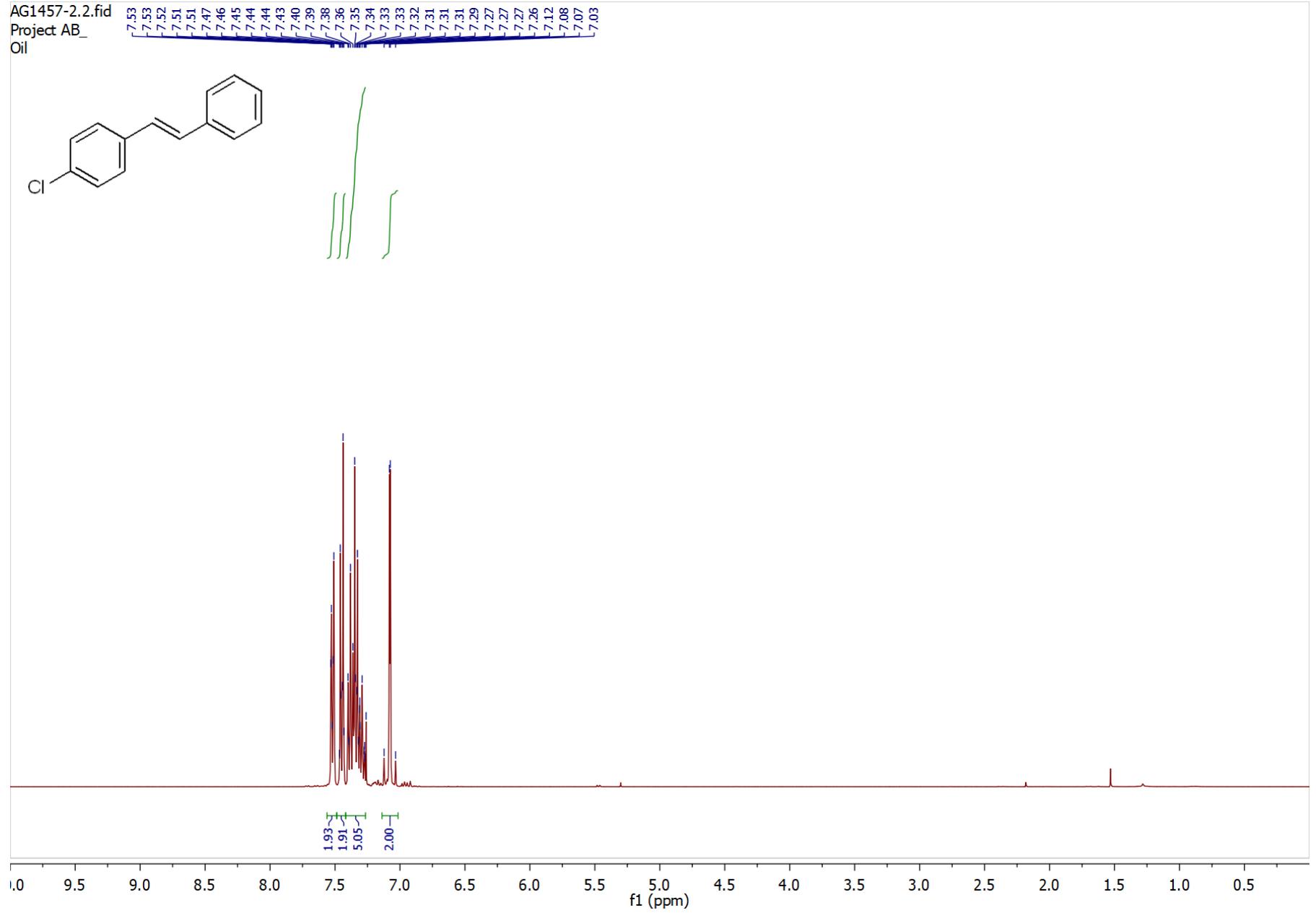
Compound 11r

AG1310.4.fid
Project AB_
Oil



Compound 11s

AG1457-2.2.fid
Project AB_
Oil



Compound 11s

AG1457-2.4.fid
Project AB_
Oil

