

Supporting Information

Fabrication of robust protein-based foams with multifunctionality by manipulating intermolecular interactions

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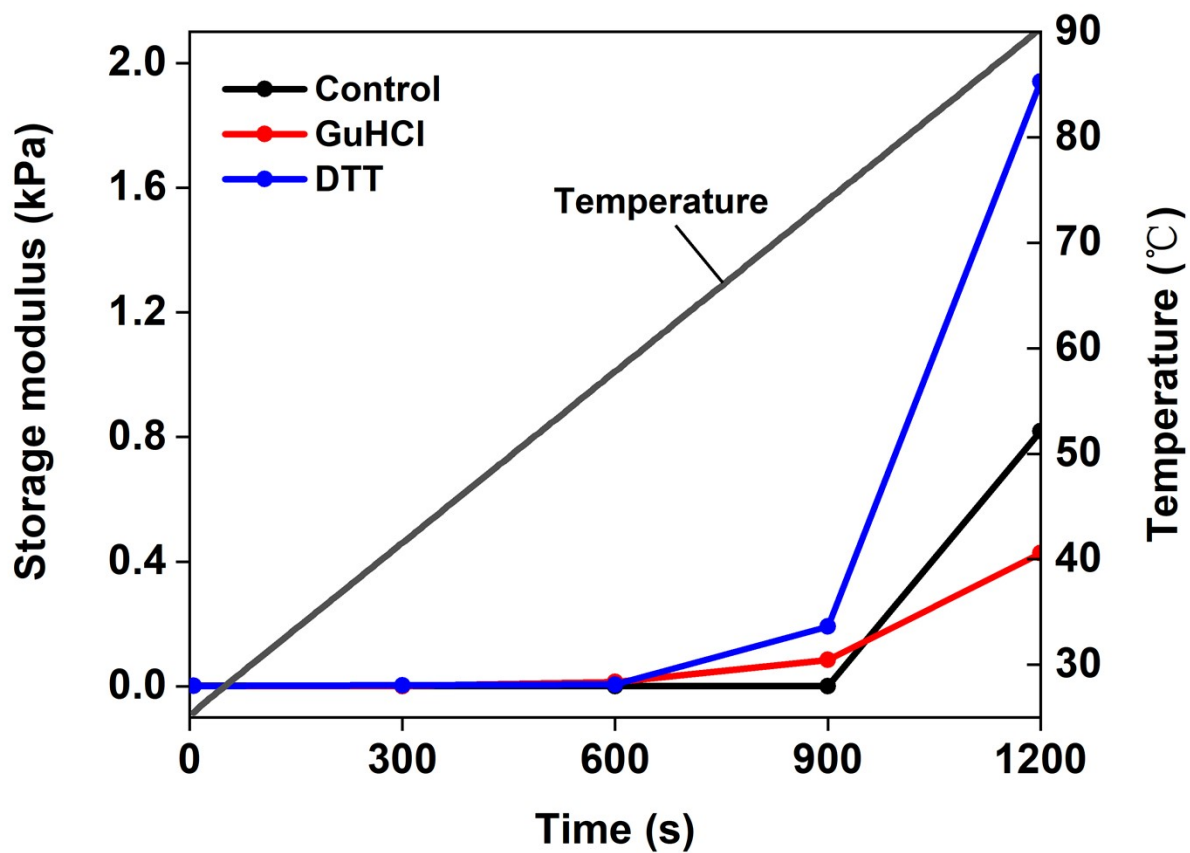


Fig. S1 Shear modulus as a function of time during the heating process (25–90 °C) of whey protein emulsions. The control, guanidinium hydrochloride (GuHCl), and dithiothreitol (DTT) samples contained 200 mM NaCl, 200 mM NaCl + 2 M GuHCl, and 200 mM NaCl + 10 mM DTT, respectively.

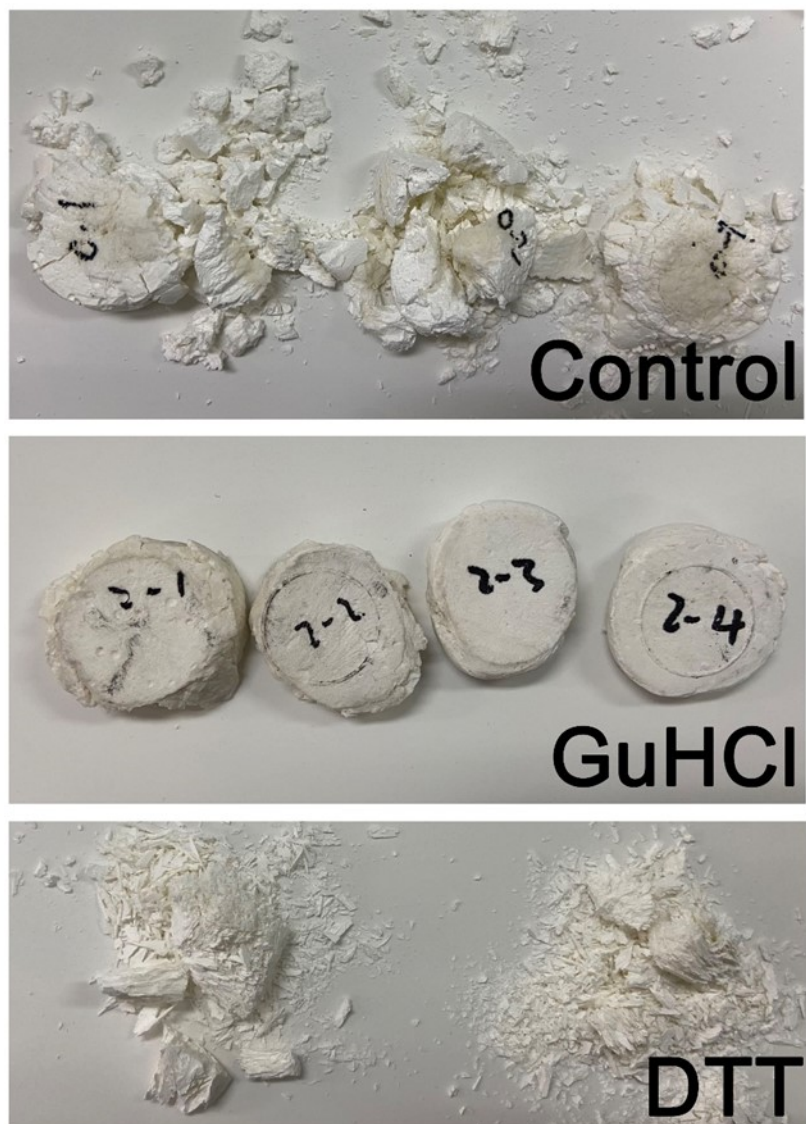


Fig. S2 Photographs of foams upon 50% deformation during the compression test. The control, guanidinium hydrochloride (GuHCl), and dithiothreitol (DTT) samples contained 200 mM NaCl, 200 mM NaCl + 2 M GuHCl, and 200 mM NaCl + 10 mM DTT, respectively.

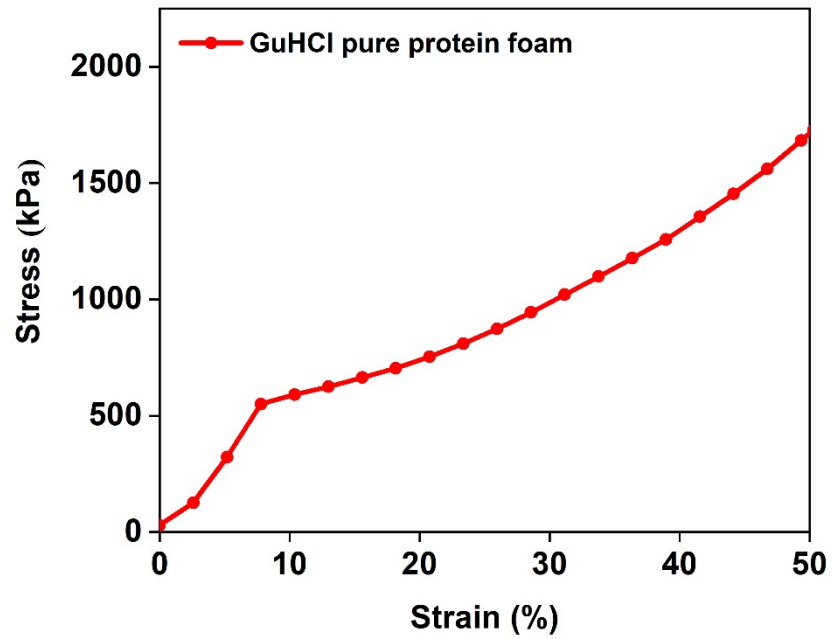


Fig. S3 Stress–strain curve of the guanidinium hydrochloride (GuHCl) pure protein foam during the compression test. The sample contained 200 mM NaCl + 2 M GuHCl.

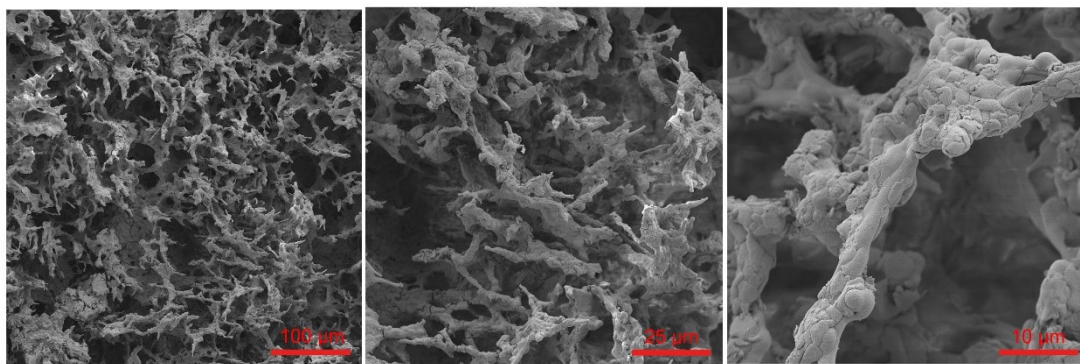


Fig. S4 Scanning electron micrographs of the guanidinium hydrochloride (GuHCl) pure protein foam at different magnifications.

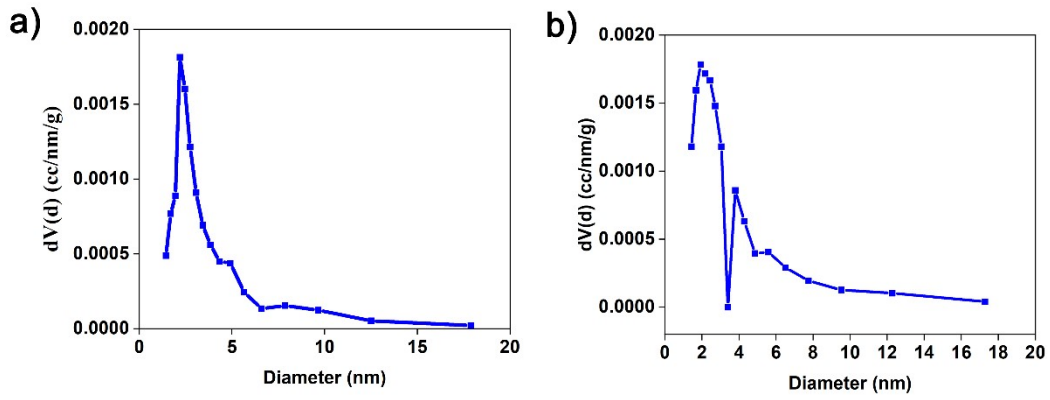


Fig. S5 Pore size distribution curves of the guanidinium hydrochloride (GuHCl) foam obtained from (a) the adsorption process and (b) the desorption process.



Fig. S6 Photographs of 2 mL of various dye solutions (100 mg/L) before and after filtration. MB, methylene blue; MG, methylene green; CV, crystal violet.

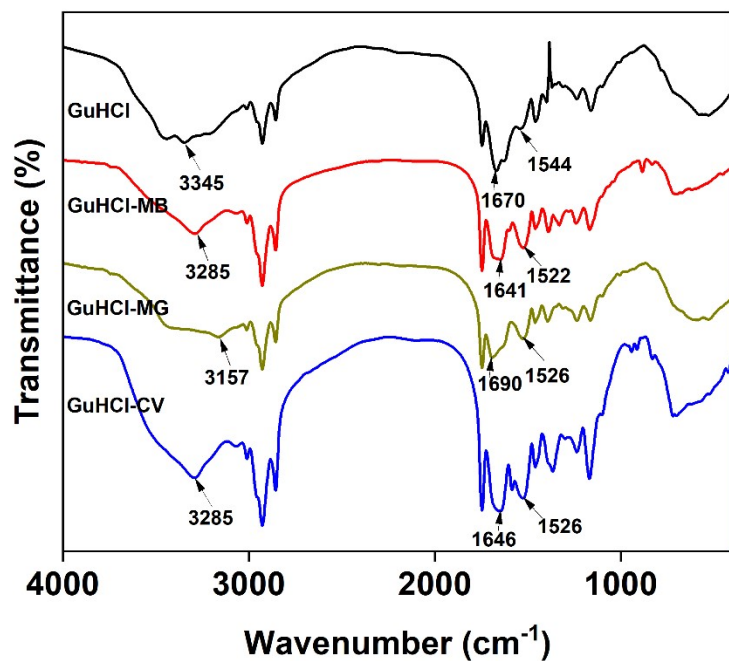


Fig. S7 Fourier transform infrared spectra of the guanidinium hydrochloride (GuHCl) foam before and after MB, MG, and CV adsorption, respectively. MB, methylene blue; MG, methylene green; CV, crystal violet.

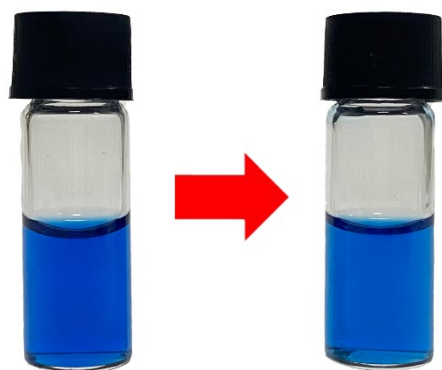


Fig. S8 Photographs of 1 mL of Indigo Carmine solution (100 mg/L) before and after filtration.

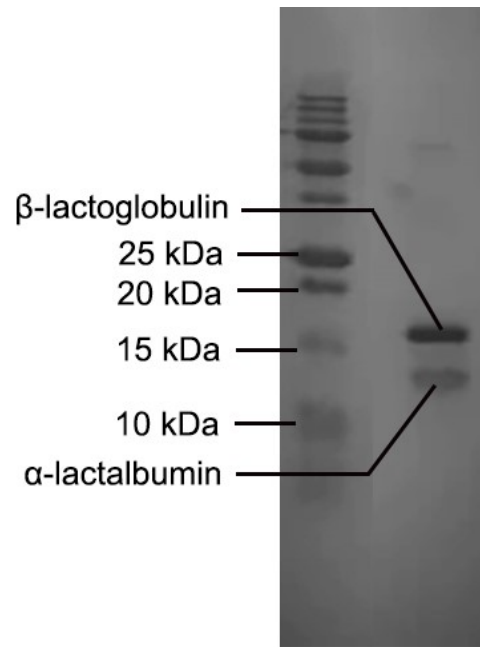


Fig. S9 Sodium dodecyl sulfate-polyacrylamide gel electrophoresis profiles of whey protein isolate. α -lactalbumin and β -lactoglobulin correspond to the bands at molecular weights of 14.4 and 18.1 kDa, respectively.

Table S1 Comparison of mechanical properties of various emulsion-filled protein gels

| Materials | Testing methods | Modulus (kPa) | Fracture stress (kPa) | Fracture strain (%) | Ref. |
|---|---------------------------|---------------|-----------------------|---------------------|-----------------------|
| 9 wt% whey protein, 25 wt% sunflower oil | Uniaxial compression test | 50 | 26 | 57 | [1] |
| 9 wt% casein micelle and sodium caseinate, 25 wt% sunflower oil | Uniaxial compression test | 20 | 7 | 39 | [1] |
| 13 w/v% soybean protein isolate, 10 v/v% soybean oil | Uniaxial compression test | Not measured | 14 | 70 | [2] |
| 6 w/v% soybean protein isolate, 20 v/v% soybean oil | Uniaxial compression test | 104 | 8.6 | 32 | [3] |
| 4.5 w/v% egg white protein, 0.72 w/v% glucono delta-lactone, 10 v/v% grape seed oil | Uniaxial compression test | Not measured | 18.5 | 72 | [4] |
| 10 wt% gelatin, 20 wt% medium-chain triglycerides oil | Uniaxial compression test | 43 | 38 | 58 | [5] |
| 10 wt% whey protein, 5 wt% soybean oil droplets | Tensile test | 42 | 55 | 147 | This work (GuHCl gel) |

Table S2 Mechanical properties of the emulsion-filled whey protein foam during the compression test

| Sample ¹ | Young's modulus (MPa) | Compressive strength ² (MPa) |
|---------------------|-----------------------|---|
| Control | 2.23 ± 0.39 | 0.53 ± 0.06 |
| GuHCl | 16.86 ± 3.62 | 1.84 ± 0.07 |
| DTT | 0.35 ± 0.04 | 0.51 ± 0.04 |

¹ The control, guanidinium hydrochloride (GuHCl), and dithiothreitol (DTT) samples contained 200 mM NaCl, 200 mM NaCl + 2 M GuHCl, and 200 mM NaCl + 10 mM DTT, respectively.

² The value is the stress at 50% strain.

Table S3 Comparison of preparation and mechanical properties of various protein-based aerogels/foams

| Materials | Hydrogel preparation complexity | Foam preparation method | Mechanical properties | Ref. |
|---|---------------------------------|---|---|------------------------|
| 10% (w/v) whey protein | Simple | Freeze drying | Modulus: 2.4 MPa; yield stress: 0.3 MPa; yield strain: 15% | [6] |
| 10% (w/v) whey protein, 2.5% (w/v) alginate, 2.5 wt% clay | Moderate | Freeze drying | Modulus: 2.5 MPa Yield stress: < 0.1 MPa; yield strain: < 5% | [6] |
| 4 w/v% silk fibroin, 2 w/v% hyaluronic acid, 1 w/v% heparin, EDC (13.6 mg/mL) ¹ , NHS (5 mg/mL) ² | Complicated | Freeze drying, annealing with ethanol vapor | Modulus: 13.1 kPa | [7] |
| 5.3% soybean protein, 2.7% nanofibrillar cellulose | Simple | Freeze drying | Modulus: 4.4 MPa | [8] |
| 10 w/w% protein extracts from canola seed meal | Simple | Freeze drying | Yield stress: < 0.1 MPa; yield strain: < 6% | [9] |
| 8 w/v% whey protein, 1 w/v% cellulose particles | Moderate | Freeze drying | Modulus: \approx 400 kPa; yield stress: 50 kPa; yield strain: < 25% | [10] |
| 10 wt% whey protein, 5 wt% soybean oil droplets | Simple | Freeze drying | Modulus: 16.9 MPa; yield stress: 1.4 MPa; yield strain: 16.5% | This work (GuHCl foam) |

¹ EDC, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride.

² NHS, N-hydroxysuccinimide.

Mercury porosimetry. Mercury intrusion porosimetry was conducted on an AutoPore V 9600 Mercury Porosimeter (Micromeritics Instruments, USA). A 0.12 g sample was put into measurement cells and was measured under the experimental conditions of 27 K, 0.1-61000 psia, and 130° contact angle of mercury on the foam.

Table S4 Pore characterization results of the GuHCl foam¹

| Sample | Surface area and pore size analyzer | | | Mercury porosimetry | | |
|--------------------|-------------------------------------|---|--|----------------------------|-----------------------------------|--------------|
| | Average pore size of skeleton (nm) | Specific surface area (m ² /g) | Total pore volume (cm ³ /g) | Average pore diameter (μm) | Bulk density (g/cm ³) | Porosity (%) |
| GuHCl ² | 6.04 | 2.60 | 3.90 × 10 ⁻³ | 16.05 | 0.31 | 69.53 |

¹The GuHCl samples contained 200 mM NaCl + 2 M GuHCl.

²GuHCl, guanidinium hydrochloride.

Table S5 Comparison of precursors, crosslinkers, and organic solvents used in various biopolymer-based aerogels/foams, and their preparation complexity

| Precursors | Crosslinkers | Organic solvents | Preparation complexity | Ref. |
|--|---|--------------------------------|------------------------|------------------------|
| Soy flour | Formaldehyde and tannin | Ethylene glycol, methanol | Complicated | [11] |
| Gelatin powders/TiO ₂ /branched polyethyleneimine | Glutaraldehyde | None | Moderate | [12] |
| Cellulose nanofibril | Methylene diphenyl diisocyanate and triethylamine | <i>tert</i> -butanol, acetone | Complicated | [13] |
| Chitosan | Formaldehyde | Methanol, acetone, acetic acid | Moderate | [14–16] |
| Periodate-oxidized cellulose | None | Octylamine | Complicated | [17] |
| Gelatin and attapulgit | Glutaraldehyde | Ethanol | Moderate | [18] |
| Cellulose | 3-(glycidyloxypropyl) trimethoxysilane and branched-polyethyleneimine | None | Simple | [19] |
| Nanocellulose | Polyethyleneimine and hexamethylenediamine | None | Complicated | [20] |
| Gelatin and cellulose nanofibers | Epichlorohydrin | None | Simple | [21] |
| Whey protein | Soybean oil droplets | None | Simple | This work (GuHCl foam) |

References

1. L. Oliver, E. Scholten and G. A. van Aken, *Food Hydrocoll.*, 2015, 43, 299–310.
2. K. H. Kim, J. M. S. Renkema and T. van Vliet, *Food Hydrocoll.*, 2001, 15, 295–302.
3. C. H. Tang, L. Chen and E. A. Foegeding, *J. Agric. Food Chem.*, 2011, 59, 4071–4077.
4. F. Alavi, Z. Emam-Djomeh, M. Salami and M. Mohammadian, *Int. J. Biol. Macromol.*, 2020, 153, 523–532.
5. G. Sala, T. van Vliet, M. Cohen Stuart, F. de Velde and G. A. van Aken, *Food Hydrocoll.*, 2009, 23, 1853–1863.
6. H.-B. Chen, Y.-Z. Wang and D. A. Schiraldi, *Eur. Polym. J.*, 2013, 49, 3387–3391.
7. M. Najberg, M. H. Mansor, T. Taillé, C. Bouré, R. Molina-Peña, F. Boury, J. L. Cenis, E. Garcion and C. Alvarez-Lorenzo, *Carbohydr. Polym.*, 2020, 237, 116107.
8. J. C. Arboleda, M. Hughes, L. A. Lucia, J. Laine, K. Ekman and O. J. Rojas, *Cellulose*, 2013, 20, 2417–2426.
9. S. E. Fitzpatrick, S. Deb-Choudhury, S. Ranford and M. P. Staiger, *J. Mater. Sci.*, 2020, 55, 4848–4863.
10. M. Ahmadi, A. Madadlou and A. A. Saboury, *Food Chem.*, 2016, 196, 1016–1022.
11. G. Amaral-Labat, L. Grishechko, A. Szczurek, V. Fierro, A. Pizzi, B. Kuznetsov and A. Celzard, *Green Chem.*, 2012, 14, 3099–3106.
12. J. Jiang, Q. Zhang, X. Zhan and F. Chen, *Chem. Eng. J.*, 2019, 358, 1539–1551.
13. F. Jiang and Y.-L. Hsieh, *ACS Appl. Mater. Interfaces*, 2017, 9, 2825–2834.

14. S. Takeshita, A. Sadeghpour, W. J. Malfait, A. Konishi, K. Otake and S. Yoda, *Biomacromolecules*, 2019, 20, 2051–2057.
15. S. Takeshita and S. Yoda, *Nanoscale*, 2017, 9, 12311–12315.
16. S. Takeshita and S. Yoda, *Chem. Mater.*, 2015, 27, 7569–7572.
17. N. T. Cervin, E. Johansson, P. A. Larsson and L. Wågberg, *ACS Appl Mater. Interfaces*, 2016, 8, 11682–11689.
18. J. Zhu, F. Zhao, R. Xiong, T. Peng, Y. Ma, J. Hu, L. Xie and C. Jiang, *Compos. A - Appl. Sci. Manuf.*, 2020, 138, 106040.
19. C. Tang, P. Brodie, Y. Li, N. J. Grishkewich, M. Brunsting and K. C. Tam, *Chem. Eng. J.*, 2020, 392, 124821.
20. L. Hossain, V. S. Raghuwanshi, J. Tanner, C.-M. Wu, O. Kleinerman, Y. Cohen and G. Garnier, *J. Colloid Interface Sci.*, 2020, 568, 234–244.
21. A. Mirtaghavi, A. Baldwin, N. Tanideh, M. Zarei, R. Muthuraj, Y. Cao, G. Zhao, J. Geng, H. Jin and J. Luo, *Int. J. Biol. Macromol.*, 2020, 164, 1949–1959.